The development of intracytoplasmic membranes in alphaproteobacteria involves the conserved mitochondrial crista-developing Mic60 protein

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Abstract

Mitochondrial cristae expand the surface area of respiratory membranes and ultimately allow for the evolutionary scaling of respiration with cell volume across eukaryotes. The discovery of Mic60 homologs among alphaproteobacteria, the closest extant relatives of mitochondria, suggested that cristae might have evolved from bacterial intracytoplasmic membranes (ICMs). Here, we investigated the predicted structure and function of alphaproteobacterial Mic60, and a protein encoded by an adjacent gene Orf52, in two distantly related purple alphaproteobacteria, *Rhodobacter sphaeroides* and *Rhodopseudomonas palustris*. In addition, we assessed the potential physical interactors of Mic60 and Orf52 in *R. sphaeroides*. We show that the three α-helices of mitochondrial Mic60's mitofilin domain, as well as its adjacent membrane-binding amphipathic helix, are present in alphaproteobacterial Mic60. The disruption of Mic60 and Orf52 caused photoheterotrophic growth defects, which are most severe under low light conditions, and both their disruption and overexpression lead to enlarged ICMs in both studied alphaproteobacteria. We also found that alphaproteobacterial Mic60 physically interacts with BamA, the homolog of Sam50, one of the main physical interactors of eukaryotic Mic60. This interaction, responsible for making contact sites at mitochondrial envelopes, has been conserved in modern alphaproteobacteria despite more than a billion years of evolutionary divergence. Our results suggest a role for Mic60 in photosynthetic ICM development and contact site formation at alphaproteobacterial envelopes. Overall, we provide support for the hypothesis that mitochondrial cristae evolved from alphaproteobacterial ICMs, and therefore have improved our understanding of the nature of the mitochondrial ancestor.

Keywords: *Cereibacter*, *Rhodobacter*, *Rhodopseudomonas*, chromatophores, eukaryote, endosymbosis, MICOS.

Introduction

Mitochondria are organelles inferred to have been present in the last common ancestor of all eukaryotes (reviewed in Roger et al. 2017). Unlike most other organelles (e.g., the endoplasmic reticulum, nucleus, cytoskeleton, etc.), the mitochondrion evolved from an endosymbiont most closely related to extant alphaproteobacteria (Muñoz-Gómez et al. 2022; Martijn et al. 2018; Fan et al. 2020). In aerobic eukaryotes, mitochondria produce most of the ATP of the cell through aerobic respiration, i.e., the harnessing of energy through the coupling of electron transport to chemiosmosis with oxygen as a terminal electron acceptor, in addition to compartmentalizing other metabolic pathways (Roger et al. 2017; Hammond et al. 2022). Because aerobic respiration occurs at internalized membranes that

can expand greatly, mitochondria allow for the proportional increase (or linear scaling) of respiration with cell volume across eukaryotes (Schavemaker and Muñoz-Gómez 2022). Mitochondria are thus one of the innovations that likely allowed many eukaryotes to achieve larger cell volumes coupled to relatively fast growth rates, and ultimately opened new evolutionary trajectories. Elucidating the origin of mitochondria and their respiratory membranes may shed light on the origin of eukaryotic cells.

The specialization of mitochondria as respiratory organelles is most clearly reflected in their internal structure. The mitochondrial inner membrane (MIM) invaginates into specialized sub-compartments called cristae, the structural hallmarks of the organelle (Daems and Wisse 1966; Mannella et al. 1994; Perkins et al. 1997). The MICOS (Mitochondrial Contact Site and Cristae Organizing System) complex and oligomers of the F_1F_O-ATP synthase are two of the most evolutionarily conserved factors responsible for the development and shape of cristae (Zick et al. 2009; Wideman and Muñoz-Gómez 2016). Whereas ATP synthase oligomers bend crista membranes at their rims to produce diverse crista shapes (Kühlbrandt 2019; Pánek et al. 2020), the MICOS complex creates both crista junctions (CJs) and contact sites (CSs) that compartmentalize, stably anchor, and maintain cristae at the mitochondrial envelope (Rampelt et al. 2017). These functions of the MICOS complex appear to be largely conserved across phylogenetically disparate eukaryotes, such as in the animal *Homo sapiens*, the fungus *Saccharomyces cerevisiae*, the land plant *Arabidopsis thaliana*, and the parasitic protist *Trypanosoma brucei* (Kozjak-Pavlovic 2017; Wollweber et al. 2017; Michaud et al. 2016; Kaurov et al. 2018).

Studies on the evolutionary history of MICOS revealed that this multi-protein complex is ancestrally present in all eukaryotes and predates the origin of mitochondria (Muñoz-Gómez et al. 2015; Huynen et al. 2016). The central and scaffolding subunit of the MICOS complex, the Mic60 protein, traces back to the *Alphaproteobacteria*, the group from which mitochondria descended. Indeed, Mic60 serves as a phylogenetic marker that is uniquely present in the *Alphaproteobacteria* and the mitochondrial lineage (Muñoz-Gómez et al. 2022). While only the C-terminal signature mitofilin domain of Mic60 is sufficiently conserved at the sequence level, the overall predicted secondary structure of Mic60 has also been conserved in both mitochondrial and alphaproteobacterial homologs (Muñoz-Gómez et al. 2015). In addition to having homologues of Mic60, many alphaproteobacteria also develop either lamellar or vesicular intracytoplasmic membranes (ICMs) that house diverse electron transport chains involved in methanotrophy, nitrification, and anoxygenic photosynthesis (Drews 1991; Niederman 2006; Pinevich 1997; LaSarre et al. 2018; Sener et al. 2016). This raises the possibility that Mic60 is involved in the development and/or stability of ICMs in alphaproteobacteria, and that mitochondrial cristae evolved from ancestral alphaproteobacterial ICMs (Muñoz-Gómez et al. 2017). Support for the functional conservation of alphaproteobacterial Mic60 comes from its capacity to bind and bend membranes *in vitro* and heterologously in the gammaproteobacterium *Escherichia coli* (Tarasenko et al. 2017). However, the precise role of Mic60 has not yet been directly studied in alphaproteobacteria, and thus the evolutionary relationship between cristae and ICMs remains unknown.

Though early ideas linked cristae to ICMs based simply on morphological resemblance, the iconography of the field (i.e., the aggregate of scientific diagrams) has mostly depicted cristae as post-endosymbiotic adaptations of mitochondria (Muñoz-Gómez et al. 2017). To better understand the function of Mic60 in alphaproteobacteria and the origin of mitochondrial cristae, we investigated the role of *mic60*, and its adjacent gene *orf52*, in two distantly related purple alphaproteobacteria: the vesicular ICM-developing *Rhodobacter* (*Cereibacter*) *sphaeroides* (*Rhodobacterales*) and lamellar ICM-developing *Rhodopseudomonas palustris* (*Rhizobiales*). We first explored the sequence and structural conservation of the alphaproteobacterial Mic60 homologue. We then experimentally investigated the effects of the disruption and overexpression of Mic60 and Orf52 in photoheterotrophic growth and ICM development. Finally, we assessed the higher-order assembly and physical interactors of alphaproteobacterial Mic60 and Orf52.

Results

<u>The predicted secondary and tertiary structure of alphaproteobacterial Mic60 is conserved</u>

Previous studies have shown that the overall predicted secondary structure of Mic60 is conserved both in eukaryotes and alphaproteobacteria (Muñoz-Gómez et al. 2015; Huynen et al. 2016). In the former, Mic60 consists of an N-terminal pre-sequence (a targeting signal to the mitochondrion), followed by a transmembrane segment, a central region with coiled-coils, and a C-terminal signature mitofilin domain. The same motifs and domains are found in the same order in the alphaproteobacterial Mic60 homolog, except for the N-terminal pre-sequence which is missing, as expected. Recently, mitochondrial Mic60 has been shown to deform membranes, and thus likely to introduce curvature at CJs (Hessenberger et al. 2017; Tarasenko et al. 2017). This membrane-deforming capability depends on a lipid-binding site (LBS) that is found in between the central coiled-coils and the C-terminal mitofilin domain. This LBS comprises two α-helices (LBS1 and LBS2), the first of which is amphipathic and presumably inserts itself into the MIM (Hessenberger et al. 2017). LBS1 is extremely important for the function of Mic60 as its removal or mutation leads to the loss of membrane binding and deformation, and also to phenotypes quite similar to those obtained when the entire *MIC60* gene is deleted in *S. cerevisiae* (Hessenberger et al. 2017).

To investigate whether the membrane-bending α-helices of eukaryotic Mic60 are present in its alphaproteobacterial homologs, we first performed profile Hidden Markov Model (pHMM)-sequence searches against the UniProtKB database. We also predicted the secondary structure of both eukaryotic and alphaproteobacterial Mic60 homologs with JPred4 (Drozdetskiy et al. 2015). A detailed inspection of both the alignments and predicted secondary structure revealed that alphaproteobacterial Mic60 has retained the two α-helices that comprise the LBS of eukaryotic Mic60 (Fig. 1A). Helical wheel projections further show that LBS1 is amphipathic in both *R. sphaeroides* and *R. palustris* (Fig.1B, C). In addition, the functionally critical amino acid position Phe573 in the yeast *Chaetomium thermophilum* (Hessenberger et al. 2017) is largely conserved among alphaproteobacteria (e.g., Phe327 in *R. sphaeroides*; see Fig. 1A). Next, we predicted the tertiary structure of the Mic60 homologs of the yeast *Lachancea thermotolerans*, whose crystal structure was recently partially solved (Bock-Bierbaum et al. 2022), and the alphaproteobacteria *R. sphaeroides* and *R. palustris* using AlphaFold2 (Fig. 1D-G). The predicted structures confirm the presence of the two α-helices (LBS) in the linker region between the central coiled-coils and the C-terminal mitofilin domain (Fig. 1D, E). Furthermore, the predicted structures show that the three conserved α-helices that comprise the mitofilin domain of eukaryotic Mic60 (α1-3M), as well as the last two small α-helices of the central coiled-coil region (α2-3C), are also present in alphaproteobacterial Mic60 (Fig. 1D-G). Structural alignments reveal that the C-terminal region of alphaproteobacterial Mic60 (i.e., LBS+mitofilin) largely overlap with that of its eukaryotic homolog (Fig. 1F, G). The major structural differences between eukaryotic and alphaproteobacterial Mic60 homologs are that the former is, on average, a longer protein with a larger segment of central coiled coils, and has a transmembrane segment much closer to the N-terminus of the protein (Fig. 1D, E). These observations indicate that alphaproteobacteria Mic60 has a largely conserved secondary and tertiary structure, and that it may bind and bend membranes through a mechanism similar to that of its eukaryotic ortholog.

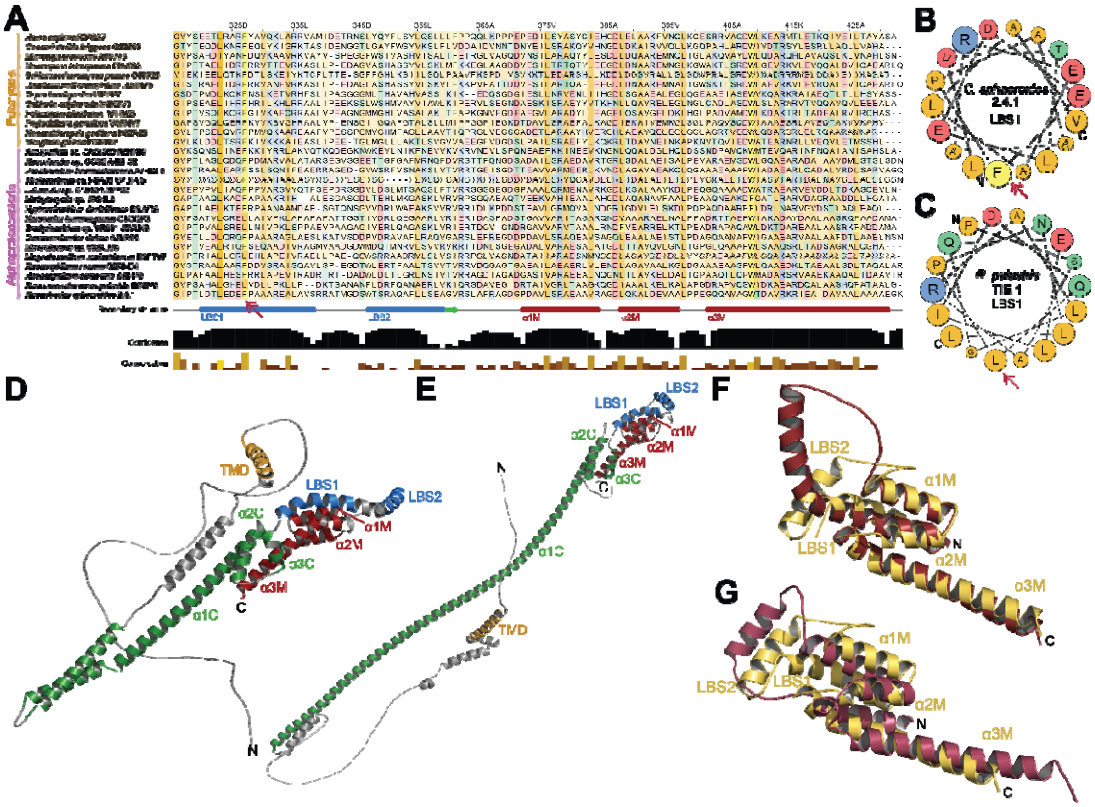

Figure 1. Evolutionary conservation of the secondary and tertiary structure of mitochondrial and alphaproteobacterial Mic60. A. Alignment of C-terminal signature mitofilin domain and its adjacent N-terminal region of representative alphaproteobacteria and eukaryotes. This alignment has the Mic60 amino acid sequence of *R. sphaeroides* as a reference and was obtained from JPred4 (Drozdetskiy et al. 2015). The amino acid colors follow a coloring scheme based on physicochemical properties, and the intensity of the color reflect evolutionary conservation of the site in the alignment. The red arrow points to the conserved Phe327 in the *R. sphaeroides* Mic60 homolog. Common gaps are hidden from the alignment as per JPred4 output. **B.** Helical wheel projection of the amphipathic helix that comprises LBS1 in *R. sphaeroides* as predicted by the HELIQUEST web server (Gautier et al. 2008). **C.** Helical wheel projection of the amphipathic helix that comprises LBS1 in *R. palustris* predicted as in B. **D.** Predicted tertiary structures of the Mic60 homolog of the alphaproteobacterium *R. sphaeroides*. **E.** Predicted tertiary structures of the Mic60 homolog of the yeast *L. thermotolerans*. α-helices are colored according to domain (mitofilin in red, LBS1 and LBS2 in blue, middle coiled coils in green, and transmembrane segment in orange) and follow the coordinates predicted by JPred4. **F.** Structural alignment of the C-terminal region of Mic60 homologs from the eukaryote *L. thermotolerans* (yellow) and the bacterium *R. sphaeroides* (red). **G.** Structural alignment of the C-terminal region of Mic60 homologs from the eukaryote *L. thermotolerans* (yellow) and the bacterium *R. palustris* (pink).

<u>Alphaproteobacterial *mic60* is clustered with a syntenic neighboring gene, *orf52*</u>

A survey of the genomic context of *mic60* in several alphaproteobacterial species revealed that *mic60* is genetically linked to genes involved in the heme biosynthesis pathway (Huynen et al. 2016). In most alphaproteobacterial genomes, *mic60* is downstream of *hemC* (hydroxymethylbilane synthase; HMBS) and *hemD* (uroporphyrinogen-III synthase; UROS), and upstream of a hypothetical protein-coding gene sometimes (mis)annotated as *hemY*. All four genes have the same orientation and are usually tightly clustered with little intergenic space in between them, which may suggest that they are functionally related or co-transcribed as part of the same operon. This is consistent with the regulatory requirements of Mic60 according to its hypothesized function in ICM development, which requires heme biosynthesis for the proper assembly of cytochromes (Muñoz-Gómez et al. 2017). Indeed, HemD is occasionally fused to Mic60 in members of the *Rhodospirilales, Kiloniellales*, and *Rhizobiales* (Fig. 2A) (Huynen et al. 2016). In the yeast *S. cerevisiae*, the MICOS complex has been reported to interact with the enzyme ferrochelatase (HemH in alphaproteobacteria) which catalyzes

the insertion of ferrous iron into protoporphyrin IX, the eighth and final step in heme biosynthesis (Dietz et al. 2021).

Although, *hemC* and *hemD* are genuine enzymes of the heme biosynthesis pathway, the gene downstream of *mic60* clearly does not encode a heme biosynthetic enzyme. This protein is usually misannotated as HemY because it contains a conserved hemY_N domain (PF07219) at its N-terminus. Confusingly, this domain is unrelated to genuine *hemY* (protoporphyrinogen IX oxidase), which is rather uncommon among alphaproteobacteria. Instead of *hemY*, most alphaproteobacteria use the product of *hemJ* to synthesize protoporphyrin IX (very few alphaproteobacteria have a genuine *hemY* gene homologous to that of *E. coli* and *Bacillus subtilis*) (Kobayashi et al. 2014). Here, the protein encoded by the gene downstream of *mic60* will be referred to as Orf52 based on its predicted size of 52 kDa in *R. sphaeroides* 2.4.1. Like alphaproteobacterial Mic60, Orf52 is an integral membrane protein, but it possesses two transmembrane segments instead of one at its N-terminus, and seems to expose its bulk to the periplasmic space. Moreover, Orf52 contains several tetratricopeptide repeat motifs, which are usually involved in protein-protein interactions and found in proteins that are part of multiprotein complexes. In *R. sphaeroides*, the *mic60□orf52* gene pair is co-transcribed as indicated by a transcription start site (TSS) in the intergenic region between *hemD* and *mic60* (Myers et al. 2020) (Fig 2A). In the magnetosome gene island (MAI) of *Magnetospirillum gryphiswaldense*, *mic60* and *orf52* are also co-transcribed separately from their neighboring genes (Dziuba et al. 2021). The conserved motif order and composition, as well as transcriptional coupling, of Mic60 and Orf52 indicate that these proteins have structural roles and may physically interact with each other at alphaproteobacterial envelopes.

<u>Mic60 and Orf52 have broad phylogenetic distributions that overlap with the distribution of ICMs in the *Alphaproteobacteria*</u>

Prokaryotic homologs of Mic60 have been previously shown to be restricted to alphaproteobacterial species, thus serving as a synapomorphy for the clade that comprises both mitochondria and the *Alphaproteobacteria* (Muñoz-Gómez et al. 2015; Huynen et al. 2016; Muñoz-Gómez et al. 2022). Moreover, phylogenetically diverse alphaproteobacteria are known to develop extensive bioenergetic ICMs that house electron transport chains associated with physiological processes such as methanotrophy, nitrification (nitrite oxidation), and (aerobic or anaerobic) anoxygenic photosynthesis (Drews 1991; Niederman 2006; Pinevich 1997; Muñoz-Gómez et al. 2017; LaSarre et al. 2018). However, it is unclear to what extent the presence of Mic60, which has been hypothesized to be involved in ICM development (Muñoz-Gómez et al. 2017), correlates with the occurrence of ICMs. To better understand the evolutionary relationship between Mic60 and ICMs, we investigated their large-scale phylogenetic distributions in the *Alphaproteobacteria*. To do this, we searched the GTDB R207 database that currently comprises more than 7,684 alphaproteobacterial genomes, with pHMMs for Mic60, Orf52, and markers for ICM-associated physiologies (Fig. 2B).

These analyses show that both Mic60 and Orf52 have a broad and dense phylogenetic distribution that encompasses the much more sporadic distribution of reported ICMs in the *Alphaproteobacteria* (Fig. 2B). Methanotrophy and nitrite oxidation are restricted to a very few genera (e.g., *Methylocella*, *Methylosinus*, and *Methylocystis*, and *Nitrobacter*), whereas photoautotrophy (or the capacity to harvest light's energy to fix carbon dioxide) is phylogenetically widespread (Fig. 2B). The presence of these ICM-associated physiologies largely overlaps with those species reported to develop extensive ICMs (Fig. 2B). Furthermore, the prediction of a photoautotrophic physiology in several species not yet studied ultrastructurally suggests that these may also develop extensive ICMs. On the other hand, phototrophs (represented by aerobic anoxygenic photoheterotrophs) have a much broader phylogenetic distribution (Imhoff et al. 2018) (Fig. 2B), though they are not often associated with the presence of extensive ICMs. It is known, however, that aerobic anoxygenic phototrophs can develop less conspicuous ICMs under some environmental conditions (Iba et al. 1988). Altogether, these phylogenetic patterns suggests that both Mic60 and Orf52 are ancestrally present in the *Alphaproteobacteria* and are required by extant species that either have or lack the capacity to develop ICMs.

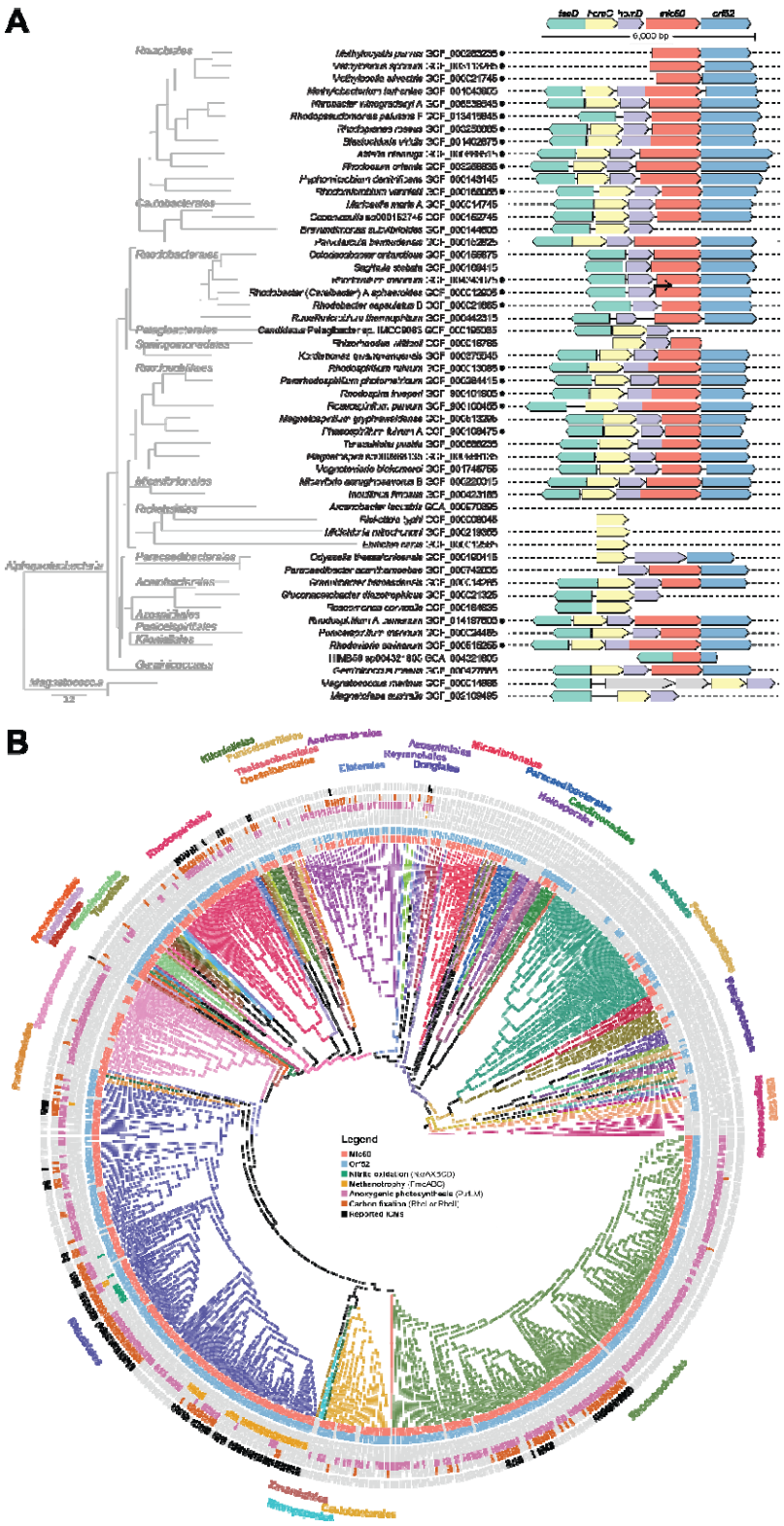

Figure 2. Genomic context of *mic60* and phylogenetic distribution of Mic60, Orf52, and ICMs across the *Alphaproteobacteria*. A. The genomic context for *mic60* in representative ICM-developing alphaproteobacteria and their relatives. Black circles to right of species identifiers denote alphaproteobacteria that develop extensive ICMs. The bent arrow denotes the TSS in *R. sphaeroides*. Representative taxa were selected manually and a supermatrix of 117 single-copy marker genes was assembled with GToTree (Lee 2019). The compositional

heterogeneity of the supermatrix was decreased with ∟trimmer and a phylogenetic tree was inferred with IQ-TREE and the LG+C60+F+G4 model (Nguyen et al. 2015). **B.** A comprehensive phylogenetic tree of the *Alphaproteobacteria* that displays the distributions of Mic60, Orf52, markers for ICM-associated physiologies, and reported ICMs across a maximally diverse set of taxa down sampled from the GTDB R207 database (Parks et al. 2022). A supermatrix of single-copy marker genes was assembled with GTDB-Tk, a phylogenetic tree was inferred with IQ-TREE (-fast mode), and the longest-branching taxa were identified and removed with TreeShrink (Mai and Mirarab 2018). The down sampling was performed with Treemmer (Menardo et al. 2018) and constrained in such a way that all taxa with ICMs and ICM-associated physiologies were retained. TreeViewer was used to display the phylogenetic distributions of traits on the tree. Protein searches were performed with hmmsearch of the HMMER suite using both Pfam and custom pHMMs (Mistry et al. 2021).

Knock out of *mic60* and *orf52* significantly affects photoheterotrophic growth in *R. sphaeroides* and *R. palustris*

Purple alphaproteobacteria develop extensive intracellular membranes (ICM) in the presence of light and the absence of oxygen (Drews 1991; Drews and Golecki 2004; Niederman 2006). These ICMs house the photosynthetic apparatus and electron transport chain, which is generally composed of light-harvesting complexes 1 and 2 (LH1 and LH2), a type II reaction center (RC), a cytochrome bc_1, a periplasmic cytochrome c_2, and an ATP synthase (Greening and Lithgow 2020). By means of cytochrome bc_1, which is also shared with the respiratory chain, the photosynthetic chain creates a proton motive force across the ICM that is harvested by the ATP synthase to produce ATP (Niederman 2006). ICMs are often continuous with, but sometimes detached from (Noble et al. 2018), the cytoplasmic membrane (CM) (Scheuring et al. 2014), just as cristae are continuous with the MIM. In *S. cerevisiae* mitochondria, the disruption of Mic60 leads to functional defects such as decreased growth rate under respiratory conditions (i.e., non-fermentable media) and increased production of oxygen radicals (John et al. 2005; Warnsmann et al. 2022). If Mic60 is involved in the development of photosynthetic ICMs, then its disruption should lead to growth defects in the absence of oxygen and the presence of light (i.e., photosynthetic conditions).

To assess whether Mic60 and Orf52 have an impact on photoheterotrophic growth and ICM development, we first knocked out these genes in two phylogenetically distant purple alphaproteobacteria amenable to reverse genetics, *R. sphaeroides* and *R. palustris* (Jaschke et al. 2011; Jiao and Newman 2007). To create knock-out strains, suicide plasmids containing the knock-out gene construct flanked by homologous stretches were transferred into both species via conjugation using a suitable *E. coli* strain. After selection and counter-selection for a first and second recombination to insert the suicide plasmid into the host genome, and excise it, respectively, these knock-out strains (Δ*mic60* and Δ*orf52*) were confirmed by PCR assays using several sets of internal and external primers that flank the knock-out gene construct junctions (Fig. S1A-D).

Purple alphaproteobacteria develop a moderate amount of ICMs under anoxia and high light intensity. The Δ*mic60* strains had a slower photoheterotrophic growth rate relative to $mic60^{+}orf52^{+}$ wild type (WT) in both *R. sphaeroides* and *R. palustris* (Fig. 3A, 3C). Under these conditions, the Δ*orf52* decreased photoheterotrophic growth significantly in *R. palustris* but not in *R. sphaeroides*. Furthermore, purple alphaproteobacteria develop larger amounts of ICMs and upregulate the LH2 complex to increase light capture under anoxia and low light intensity. As expected, both Δ*mic60* and Δ*orf52* strains displayed even slower photoheterotrophic growth rates in both *R. sphaeroides* and *R. palustris* in low light (Fig. 3B, 3D. Interestingly, Δ*orf52* displayed much more severe decreases in growth rate, and even lower yield at stationary phase, than Δ*mic60* in *R. palustris* (Fig. 3B). This is opposite from what was observed in *R. sphaeroides* (Fig. 3D) and indicates that these proteins might contribute differently to photoheterotrophic growth in these two distantly related species. In summary, the slower photoheterotrophic growth rate of the knock-out strains, especially under low light when larger amounts of ICMs are required, suggests that both Mic60 and Orf52 affect the development of photosynthetic ICMs.

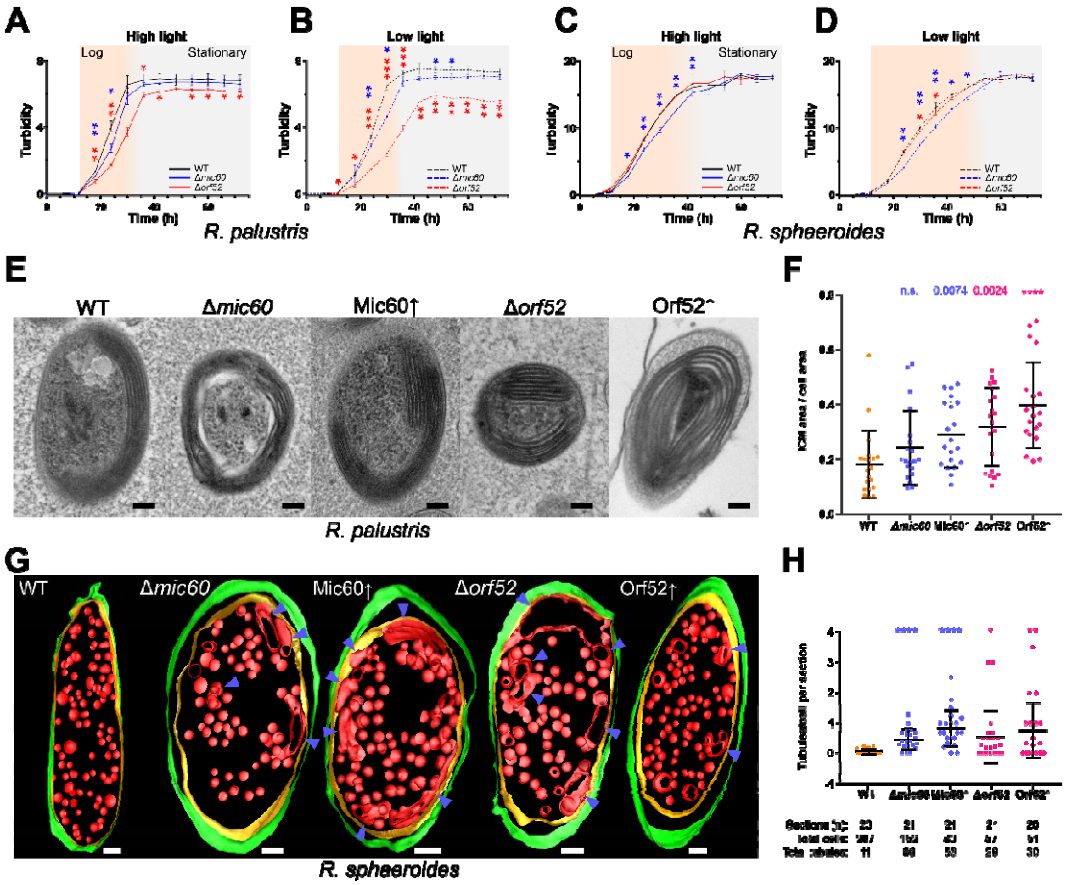

Figure 3. The disruption of the *mic60* and *orf52* genes causes defects in photoheterotrophic growth and ICM development. A-D. Growth curves of *R. palustris* WT and Δ*mic60* and Δ*orf52* strains under high (**A**) and low (**B**) light and *R. sphaeroides* under high (**C**) and low (**D**) light. Turbidity was measured at 565 nm and expressed in arbitrary units given on the *y*-axis. Time points are expressed in hours on the *x*-axis. Growth stages are color-coded according to the labels at the top. Colored asterisks show statistical significance of differences at each time point between WT and either Δ*mic60* (blue) or Δ*orf52* (red). **E.** Exemplar TEM micrographs of each *R. palustris* strain grown in low light. **F.** Column scatter plot of the ratio of *R. palustris* ICM area/whole cell area (*y*-axis) measured from 20 cells imaged using TEM in *R. palustris* WT, knock-out (Δ*mic60* and Δ*orf52*) and overexpression (Mic60↑ and Orf52↑) strains. Mean and standard deviation shown by middle bar and whiskers, respectively. Values and asterisks above each column represent statistical significance of the difference of each population in comparison to WT. **G.** Exemplar electron tomograms of each *R. sphaeroides* strain grown in low light. Blue arrows point at aberrant ICMs. **H.** Column scatter plot of tubules/cell per section (*y*-axis) for *R. sphaeroides* WT, knock-out (Δ*mic60* and Δ*orf52*) and overexpression (Mic60↑ and Orf52↑) strains. The number of sections, cells and tubules for each cell lines are given in the table below the *x*-axis. Mean and standard deviation shown by middle bar and whiskers, respectively. Asterisks above each column represent statistical significance of the difference of each mutant population in comparison to WT. Related to Movie S1-S4. Scale bars in (**D**) and (**F**), 100 nm. Statistical significance: *, P<0.05; **, P<0.01; ***, P<0.001; ****, P<0.0001. See also Fig. S1.

Mic60 and Orf52 are involved in the formation of lamellar and vesicular ICMs

The disruption of mitochondrial Mic60 leads to structural defects such as the loss of CJs, the detachment of cristae from the mitochondrial envelope, and elongated crista membranes (Rabl et al. 2009; Kaurov et al. 2018). Hypotheses about the role of Mic60 in ICM development postulate that this protein might be responsible for creating ICM junctions (ICMJs) and CSs that compartmentalize ICMs and anchor them to the bacterial envelope (Muñoz-Gómez et al. 2017). The loss of Mic60 and Orf52 from ICM-developing alphaproteobacteria might thus lead to the loss of ICMJs or detached and elongated ICMs. Their overexpression, in contrast, might lead to a larger number of ICMJs or branching and larger ICMs.

To directly address whether the gene products of *mic60* and *orf52* are involved in ICM formation, we generated *R. palustris* and *R. sphaeroides* strains capable of overexpressing Mic60 and Orf52 (Mic60↑ and Orf52↑, respectively). These strains were verified by reverse transcription quantitative PCR (RT-qPCR) upon induction for six hours with 1 mM isopropyl-β-D-thiogalactoside (IPTG). Both Mic60↑ and Orf52↑ strains had an increase in gene expression of ~7□9-fold in *R. sphaeroides*, and of about two-fold in *R. palustris*, relative to ex-conjugants grown in the absence of IPTG (Fig. S1C). The knock-out and WT strains were grown under photoheterotrophic conditions and low light and IPTG-induced for six hours (in case of overexpression ex-conjugants) to maximize ICM development. After harvesting, the strains were cryopreserved, contrasted, and imaged using transmission electron microscopy (TEM). Randomized TEM micrographs were blindly scored to quantitatively evaluate phenotypes in accord with the cytoplasmic membrane-adjacent stacked lamellar ICMs of *R. palustris* and the uniformly distributed vesicular ICMs of *R. sphaeroides* (LaSarre et al. 2018) (Fig. 3E-H, Movie S1-4).

In *R. palustris*, the area occupied by the lamellar ICMs increased relative to the whole cell area in both knock-out (Δ*mic60* and Δ*orf52*) and overexpression (Mic60↑ and Orf52↑) strains, as compared to the WT (ratio ICM area/cell area=0.18±0.03) (Fig. 3E-F). This increase was statistically significant in Mic60↑ (0.29±0.03) but not in Δ*mic60* (0.24±0.03). Both Δ*orf52* (0.32±0.03) and Orf52↑ (0.40±0.04) demonstrated higher and statistically significant increases in ICM:cell area ratios when compared to their Mic60 counterparts. Wavy ICM membranes and/or wide gaps between lamellae were often observed in the knock-out strains (Fig 3E). In *R. sphaeroides*, we scored for the appearance of elongated ICMs (i.e., tubules) per cell as these were rarely observed in the WT strain (tubules/cell/section=0.04±0.02) (Fig 3G-H). Both Δ*mic60* (0.46±0.07) and Mic60↑ (0.82±0.13) showed a statistically highly significant increase in the number of tubules, whereas these structures were observed to a lesser extent in Δ*orf52* (0.54±0.19) and Orf52↑ (0.74±0.20); the most conspicuous outliers (≥3 tubules/cell/section) were observed in the latter cells. Electron tomograms were used to render representative images for *R. sphaeroides* cells whose ICM membranes were not well contrasted (Fig 3G, Movie S1-4). This revealed the presence of elongated ICMs of various lengths and volumes in all the mutant strains assayed, indicating an increase in ICM membrane area. Furthermore, the periplasm of the mutants appears to be expanded compared to the WT.

The ultrastructural defects of the Δ*mic60* and Δ*orf52* strains of *R. sphaeroides* and *R. palustris* are consistent with their lower photoheterotrophic growth dynamics. Namely, the Δ*orf52* strain exhibits more pronounced phenotypes than the Δ*mic60* strains in *R. palustris*, whereas the opposite is true in *R. sphaeroides*. We also observed that both the knock-out and overexpression strains led to ICM area expansion. It is possible that both the disruption and overproduction of Mic60 lead to unregulated membrane growth that, under the image analyses employed here, produce seemingly similar phenotypes. The sporadic wide gaps observed between the ICM lamellae in *R. palustris*, and the expanded periplasm in *R. sphaeroides*, may be a consequence of the loss of ICMJs between lamellae and the loss of CSs with the outer membrane, respectively. However, improved sample preservation and larger-scale volumetric electron microscopy will be required to quantitatively assess changes in the shape and number of ICMJs, which potentially differ between knock-out and overexpression strains. The expansion of photosynthetic ICMs is similar to the enlarged respiratory cristae observed in *S. cerevisiae* and *T. brucei* mitochondria defective of Mic60, and to the enlarged magnetosomes reported in the magnetotactic alphaproteobacterium *M. gryphiswaldense* upon deletion of the *mic60* paralog found within the MAI (see Discussion). Together, the ultrastructural defects displayed by both Δ*mic60* and Δ*orf52* suggest that these genes are involved in the development of photosynthetic ICMs in purple alphaproteobacteria.

<u>Mic60 and Orf52 are assembled into a 250 kDa complex in the ICMs of *R. sphaeroides*</u>

Eukaryotic MICOS comprises 6-9 subunits in *H. sapiens*, *S. cerevisiae*, and *T. brucei*. Four of them, namely Mic60, Mic10, Mic19 and Mic12, are inferred to have been ancestral to eukaryotes (Muñoz-Gómez et al. 2015; Huynen et al. 2016). Furthermore, the MICOS complex has been suggested to engage in interactions with a myriad of other proteins, such as the outer membrane β-barrel insertase Sam50, the outer membrane translocase Tom40, and the mitochondrial intermembrane space assembly protein Mia40 (Pánek et al. 2020; Muñoz-Gómez et al. 2015b). It is thus possible that

alphaproteobacterial Mic60 and Orf52 are part of a larger multi-protein complex at alphaproteobacterial envelopes.

To investigate whether Mic60 and Orf52 form part of a multi-protein complex, *R. sphaeroides* was conjugated with a suitable *E. coli* strain to transfer plasmids for the IPTG-inducible expression of either Mic60 or Orf52 bearing a C-terminal hexa-histidine (6xHis) tag. The expression of Mic60-6xHis and Orf52-6xHis was verified by Western blot analysis using anti-6xHis antibodies (Fig. 4A). Mic60-6xHis was found to migrate as a ~62 kDa protein, in contrast to its theoretical 43.8 kDa molecular weight; this is likely explained by the anomalous mobility of acidic proteins in SDS-PAGE (Guan et al. 2015). The two overexpression strains, i.e., Mic60-6xHis↑ and Orf52-6xHis↑, alongside the $Mic60^{+}Orf52^{+}$ (WT) control strain, were disrupted by high pressure homogenization to allow for ICM isolation (i.e., chromatophores) by differential centrifugation. The absorption spectra of the isolated ICMs from each strain were essentially identical, which indicates that the overexpression of Mic60-6xHis and Orf52-6xHis does not affect the RC-LH1:LH2 protein composition ratio of ICMs (Fig. 4B).

The isolated ICMs were resolved in blue native PAGE (BN-PAGE) gels and transferred onto a membrane for probing with an anti-His antibody (Fig. 4C). Both Mic60-6xHis and Orf52-6xHis incorporate into a ~250 kDa multi-protein complex, with the antibody signal absent from the WT lane. Notably, the immunoreactive ~250 kDa-size band seems to correspond to a faint and similarly sized Coomassie-stained band in the BN-PAGE gel. The proteins from four excised ~250 kDa bands were eluted and analyzed by liquid chromatography-tandem mass spectroscopy (LC-MS/MS) to identify proteins migrating in this region of the BN-PAGE gel (Fig. S2A). Mic60 and Orf52 were among the 175 top proteins identified with a mean intensity score >23 in all four independent biological replicates and thus considered high-confidence interactors (Dataset 1). The top-three hits in the ~250 kDa band corresponded to the RC complex subunits H, M, and L, which were previously shown to co-migrate with Mic60 in the BN-PAGE gels (D'Amici et al. 2010). This is consistent with both Mic60 and Orf52 previously being detected in isolated ICM and/or in its developmental precursor, i.e., 'upper pigmented bands' (UPB) (D'Amici et al. 2010; Jackson et al. 2012). These experiments thus show that both Mic60 and Orf52 are part of a higher-order assembly complex of ~250 kDa present in photosynthetic ICMs.

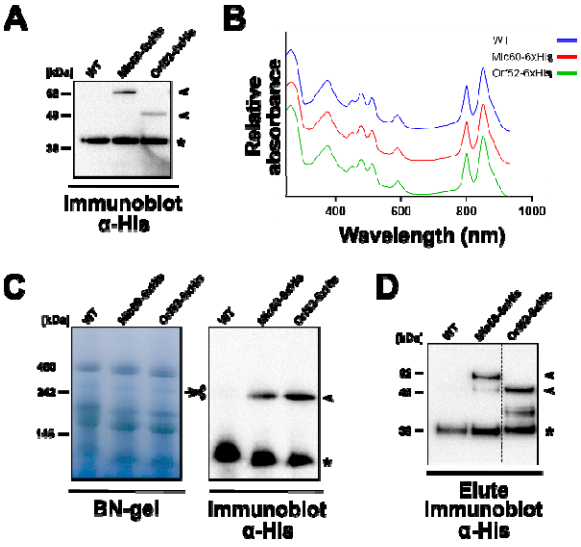

Figure 4. Mic60 and Orf52 assemble into a 250 kDa protein complex in *R. sphaeroides*. A. Immunoblot verifying the expression of Mic60-6xHis and Orf52-6xHis. **B.** Absorption spectra (~280-950 nm scan, *x*-axis) of ICMs isolated from Mic60-6xHis, Orf52-6xHis, and WT control strains. Relative absorbance given on *y*-axis and spectra are normalized to the bacteriochlorophyll *a* Qx peak at 590 nm. Spectra are stacked on top of each other and color-coded according to the legend on the upper right-hand corner. **C.** Protein complexes from detergent-solubilized isolated ICMs as resolved by BN-PAGE. Scissor symbol indicates the ~250 kDa band excised for MS analysis on left gel (see also Fig. S3A and Dataset 1). Right gel shows immunoblot demonstrating that Mic60-

6xHis and Orf52-6xHis assemble into a ~250 kDa band. **D.** Immunoblot of Mic60-6xHis and Orf52-6xHis AP eluates showing that the 6xHis-tagged bait proteins were successfully purified from detergent-solubilized isolated ICMs. For **A**, **C**, and **D**, molecular weight markers shown on left. All immunoblots use arrows to point at specific antibody signals from Mic60-6xHis and Orf52-6xHis (absent for WT) controls whereas asterisks denoted non-specific band used as loading control.

Mic60 and Orf52 interact with the outer membrane β-barrel insertase BamA

Among several interactions reported for the eukaryotic MICOS complex (e.g., with Tom40, Sam50, Mia40), it was suggested that the Mic60-Sam50 interaction may have predated the origin of mitochondria (Muñoz-Gómez et al. 2015b). This interaction is required for making CSs at the mitochondrial envelope of phylogenetically disparate eukaryotes such as *H. sapiens*, *S. cerevisiae*, and *T. brucei* (Kozjak-Pavlovic 2017; Wollweber et al. 2017; Kaurov et al. 2018), and the bacterial homolog of Sam50, BamA, is a ubiquitous protein of bacteria surrounded by two membranes (diderms). In these bacteria (e.g., *Proteobacteria*), BamA required for the assembly of β-barrels in the outer membrane (Höhr et al. 2018; Doyle and Bernstein 2019; Voulhoux et al. 2003). Furthermore, *mic60* is genetically linked to *orf52* and co-transcribed as part of the same operon (Myers et al. 2020). These observations and inferences prompted us to next investigate potential physical interactors of alphaproteobacterial Mic60 and Orf52.

We isolated ICMs from *R. sphaeroides* and performed affinity purifications (AP) of Mic60-6xHis and Orf52-6xHis. The ICMs were solubilized with a mixture of digitonin (1.5% w/v) and Triton X-100 (0.1% v/v), and the cleared lysate was incubated with cobalt-coated Dynabeads to capture the 6xHis-tagged baits. After extensive washing, the successful capture of Mic60-6xHis and Orf52-6xHis was confirmed on a portion of beads taken for Western blot analysis with an anti-His antibody (Fig. 4D)—a mock AP was done on *R. sphaeroides* WT as a control for non-specific binding. The remaining beads from triplicate APs of Mic60-6xHis, Orf52-6xHis and the mock controls were trypsinized and analyzed by LC-MS/MS. Protein enrichment in bait-APs in comparison to the mock control was quantified using label-free quantification as previously described (Cadena et al. 2021). In each AP, 497 high-confidence proteins were found (Dataset 2) based on mean Andromeda confidence scores >100 in three biological replicates (Cox et al. 2011). Among these, proteins were considered to have true physical interaction if they (1) had Log_2-transformed fold-enrichment and p-values larger than three (Fig. 4A-B), (2) were part of the higher-order ~250 kDa complex (see above; Dataset 1), (3) were previously reported in ICM and/or upper pigmented bands (or UPBs, which are ICM precursors; Fig. 5C-D), and (4) were predicted to have transmembrane segments or a β-barrel structure (Fig. 5C-D).

The data show that both Mic60-6xHis and Orf52-6xHis interact with BamA (Fig. 5, S2), the β-barrel insertase subunit of the β-barrel assembly machine (BAM) complex (Voulhoux et al. 2003). Somewhat unexpectedly, Mic60-6xHis and Orf52-6xHis do not interact with each other, as neither protein was found to co-purify with either bait protein (Fig. 5A-B). We cannot rule out that the C-terminal 6x-His tag may have interfered with their interaction, although this is unlikely due to its small size. Mic60 was also not found to interact with the enzyme HemD, although these proteins have fused in some alphaproteobacteria (Fig 2A) (Huynen et al. 2016). This was not unexpected as the HemD-Mic60 fusion protein is predicted to be membrane-anchored at its midpoint, having the HemD fragment exposed to the cytoplasm and Mic60 fragment on the periplasm. Moreover, the high degree of enrichment of Mic60 relative to BamA (Fig 5B, 5D) is consistent with the idea that this protein engages in homotypic interactions and forms homooligomers as its eukaryotic counterparts (Bock-Bierbaum et al. 2022; Hessenberger et al. 2017; John et al. 2005).

In contrast to Mic60, which exhibits a strong interaction solely with BamA, Orf52 also interacts with a β-barrel protein annotated as Outer Membrane Efflux (OM Efflux) and a very small 4.8 kDa protein with an N-terminal TMD that is encoded by the RSP_7517 locus (Fig. 5A, 5C; Fig. S3D). The former belongs to the TolC protein family and was found to be more abundant in ICM than the UBP precursor (Jackson et al. 2012), whereas the latter is among the most enriched proteins in the Orf52 AP. The enrichment of RSP_7515 is surprising as it was not previously detected in ICMs or UPBs (Jackson et al., 2012). It was however, detected in the ~250 kDa BN-PAGE band, albeit below our defined threshold by a single peptide (Dataset 1). This may be explained by the fact that this protein is highly hydrophobic and would only yield two peptides by trypsinization, making it less amenable to detection

by shotgun LC-MS/MS, as employed for analyzing the protein composition of the ~250 kDa BN-PAGE band (Barrera and Robinson 2011) (Fig. S3D). In conclusion, the Sam50 homolog BamA appears to be the only physical interactor of alphaproteobacterial Mic60 (Fig. 5E), and this interaction has been conserved despite ~2 billion years of evolutionary divergence of modern alphaproteobacteria from mitochondria.

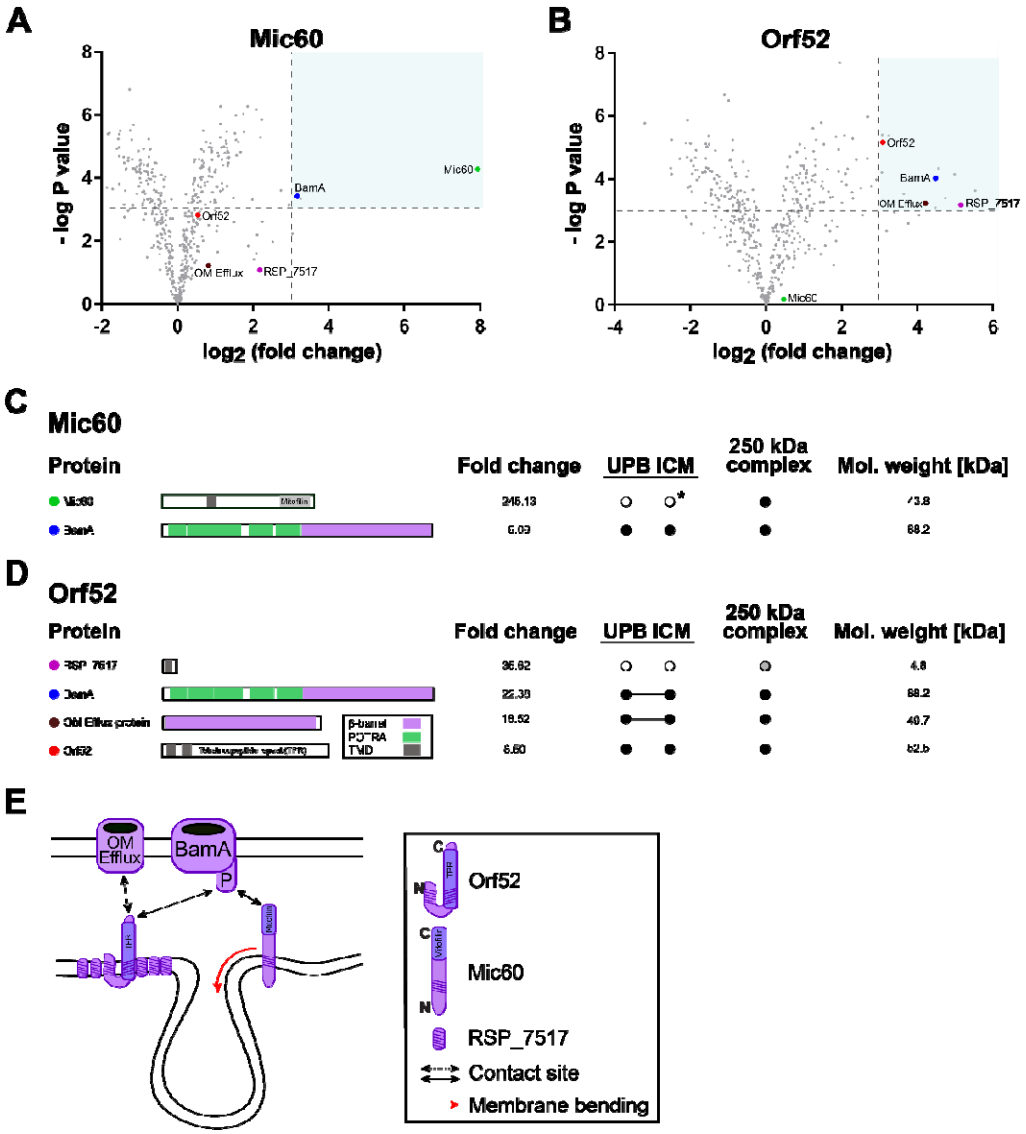

Figure 5. The interactomes of Mic60 and Orf52 demonstrate that both proteins interact with the outer membrane β-barrel protein BamA in *R. sphaeroides*. A-B. Volcano plots of proteins identified by MS in AP eluates of Mic60-6xHis (**A**) and Orf52-6xHis (**B**), with -Log P-values for each protein (*y*-axis) plotted against Log_2-tranformed fold-enrichment over WT negative control (*x*-axis); thresholds for each value for a protein considered enriched interactors are indicated by dotted lines, demarking the shaded area of each plot. Baits and enriched interactors are indicated by labelled and colored large dots, whereas other proteins are demarked by small, grey dots. **C-D.** Summary of enriched interactors found in Mic60-6xHis (**C**) and Orf52-6xHis (**D**) APs showing the same color scheme as in **A** and **B**. From the left of protein names: schemas depicting the domain architecture of each interacting protein; its fold change in APs compared to WT negative controls; presence in upper pigmented band (UPB) and ICM proteomes (Jackson et al., 2012); presence in ~250 kDa band proteome (Dataset 1); theoretical molecular (Mol.) weight. Black dots indicate presence in the proteome; grey dot indicates presence below our defined threshold; *, presence in ICM according to MS analysis of D'Amici *et al.* (2010). **E.** Schema of

Mic60 and Orf52 interactions in *R. sphaeroides*. Boxed contains a key to depicted proteins, with N- and C-termini shown in red. Related to Fig. S2 and Dataset 2.

Discussion

The overarching aim of this study was to investigate the structure and function of alphaproteobacterial Mic60, thereby testing the hypothesis that ICMs were transformed into cristae during the early evolution of mitochondria (Muñoz-Gómez et al. 2017). To this end, we focused on two distantly related purple alphaproteobacteria, *R. palustris* and *R. sphaeroides*. We showed that alphaproteobacterial Mic60 has (1) a conserved tertiary structure and membrane-bending amphipathic helix, (2) affects photoheterotrophic growth and ICM development and shape, (3) is part of a higher-order 250 kDa multi-protein complex, and (4) physically interacts with the core BamA subunit of the outer membrane BAM complex. Furthermore, we also showed that Orf52, which is encoded by *mic60*'s neighboring gene, also affects photoheterotrophic growth, ICM development and shape, and, although it does not interact with Mic60, it also physically interacts with the outer membrane protein BamA.

These findings are consistent with previous observations and experiments that also support a functional conservation of Mic60 and its involvement in the development of both vesicular and lamellar ICMs. First, the expression profile of Mic60 follows the development of photosynthetic ICMs in the absence of oxygen and presence of light (Callister et al. 2006). Second, Mic60 localizes to photosynthetic ICMs in three phylogenetically disparate alphaproteobacteria, *R. sphaeroides (Rhodobacterales*), *Rhodospirillum rubrum* (*Rhodospirillales*), and *R. palustris* (*Rhizobiales*) (D'Amici et al. 2010; Jackson et al. 2012; Selao et al. 2011; Fejes et al. 2003), as indicated by the proteomes of isolated ICMs. Third, *R. sphaeroides*' Mic60 is capable of binding and tubulating membranes *in vitro*, and its heterologous overexpression induces ICM-like structures in *E. coli* (Tarasenko et al. 2017). Fourth, both *mic60* and *orf52* have paralogs in the MAI of the alphaproteobacterial genus *Magnetospirillum* of the order *Rhodospirillales* (Muñoz-Gómez et al. 2015) whose disruption leads to fewer and larger magnetosomes (Lohße et al. 2011; Lohße et al. 2014). Multiple sources of evidence thus support the notion that the function of Mic60 has been conserved in alphaproteobacteria.

What is the precise role of Mic60 in ICM development? The capacity of Mic60 to bend membranes (Tarasenko et al. 2017), presumably through its conserved amphipathic helix (see Fig. 1), suggests that Mic60 induces curvature at ICMJs. Moreover, alphaproteobacterial Mic60 interacts with BamA, the central subunit of the BAM complex and homolog of Sam50. Mitochondrial Sam50 interacts with Mic60 through its intermembrane space-protruding POTRA domains, and it is likely that the same type of interaction occurs in the periplasm of alphaproteobacteria. This ancient protein-protein interaction suggests that, in addition to aiding the formation of ICMJs, alphaproteobacterial Mic60 is also involved in the formation of CJs that anchor ICMs to the alphaproteobacterial envelope. Thus, it appears that both the formation of CSs and CJs/ICMJs have been conserved in alphaproteobacteria and mitochondria. The formation of mitochondrial CJs and CS by Mic60 are aided by Mic19 (Hessenberger et al. 2017; Bock-Bierbaum et al. 2022), but there is no evidence for a Mic19-like protein in alphaproteobacteria; our pull-downs did not reveal proteins with Coiled-Coil-Helix-Coiled-Coil-Helix (CHCH) motifs. It is plausible that CSs stabilize ICMs by providing an anchor to the OM at points of high membrane curvature, i.e., ICMJs (Tang et al. 2020). This may contribute to the biogenesis of ICMs by ensuring their continuity with the CM where protein complex subunits may first be inserted and assembled, giving rise to the UPB. Moreover, the larger macromolecular complex that Mic60 is part of in *R. sphaeroides* is reminiscent of the extended interaction network, ERMIONE, that in *S. cerevisiae* mitochondria involves Mic60 (Wideman and Muñoz-Gómez 2016). It is possible that BamA, through its interactions with Mic60, Orf52, OM Efflux, and RSP_7515, serves as the hub of a larger protein-interaction network, that, for example, facilitates lipid transfer and protein export to the outer membrane. Future studies are required to determine whether alphaproteobacterial Mic60 forms homodimers and homotetramers as recently reported for fungal Mic60 (Bock-Bierbaum et al. 2022), and elucidate the molecular mechanisms by which alphaproteobacterial Mic60 interacts with BamA.

The evidence for the structural and functional conservation of alphaproteobacterial Mic60 relative to its mitochondrial homolog is most compatible with an evolutionary scenario in which ICMs and cristae are homologous. This view implies that cristae most likely evolved from the ICMs developed by the

last common ancestor of mitochondria and its sister group, the *Alphaproteobacteria*; cristae thus have a pre-endosymbiotic origin. If this hypothesis turns out to be correct, bioenergetic ICMs might have pre-adapted the first mitochondrial ancestor to become an efficient bioenergetic or respiratory organelle (Muñoz-Gómez et al. 2017). The widespread but sporadic phylogenetic distribution of ICMs across the *Alphaproteobacteria* is then most likely explained by multiple independent losses. This is conceivable as both cristae and ICMs are known to have been lost repeatedly as a result of physiological specialization to different environments (e.g., switches to anaerobiosis in mitochondria or to heterotrophy in photosynthetic bacteria). However, some alphaproteobacteria that have Mic60 homologs are not known to develop ICMs. In these, Mic60 might play a more general function, e.g., CS formation by interacting with BamA for lipid transfer or protein export, that is still required in the absence of ICMs. This more general function might have been ancestral to Mic60 and agrees with the observation that a distant homologue of Mic60 that lacks the conserved C-terminal mitofilin domain, namely HemX, is restricted to and widespread in the *Gammaproteobacteria* (Huynen et al. 2016). It is thus conceivable that the signature mitofilin domain first evolved in a HemX-like protein and that this coincided with the origin of ICMs in a common ancestor of alphaproteobacteria and mitochondria. Future endeavors focused on phylogenetically disparate alphaproteobacteria with and without ICMs will shed light on the functions and mechanisms of Mic60 in prokaryotes. Altogether, the structural and functional conservation of alphaproteobacterial Mic60 shown here, suggests a role of this protein in curving membranes at ICMJs and making CSs at envelopes. Therefore, it is probable that the mitochondrial ancestor was an ICM-bearing alphaproteobacterium.

MATERIALS AND METHODS

Bacterial strains, media and growth conditions

R. palustris TIE-1 and *E. coli* BW29427-λpir-RP4 were kindly provided by Dianne K. Newman (California Institute of Technology) (Jiao et al. 2005). *R. sphaeroides* 2.4.1, *E. coli* S17-1- λpir, and *E. coli* DH5α-λpir were kindly provided by Jeanette Johnson-Beatty (University of British Columbia). *R. palustris* TIE-1 strain was grown both chemo- and photo-heterotrophically at 30 °C in YPS rich medium (Jiao et al., 2005) unless otherwise noted. *R. palustris* TIE-1 was grown photoheterotrophically at 30 °C on FEM minimal medium (Ehrenreich and Widdel 1994). *R. sphaeroides* 2.4.1 strain was grown chemoheterotrophically at 30 °C in LB or RLB rich media (Jun et al. 2014), unless otherwise noted, and photoheterotrophically at 30 °C on RCVBN minimal medium (Beatty and Gest 1981). Table S1 lists all strains used this study.

For growth analysis *R. sphaeroides* and *R. palustris* mutants and WT strains were grown in front to three incandescent 40 W lightbulbs at a temperature of 29 °C, with the distances of the culture tubes from the light sources adjusted to allow the appropriate light intensities and temperature. The light intensities were measured to be ~200 µmol photons m^{-2} s^{-1} for high light and ~10 µmol photons m^{-2} s^{-1} for low light. The latter was achieved by placing the two layers of approximately ~ 50 % neutral density filter sheeting between the light source and culture tubes. Prior to growth analysis, cells were grown in in rich media in triplicate (*R. sphaeroides*, RLB without antibiotics; *R. palustris*, YPS no antibiotic for WT, 400 µg/ml kanamycin for mutants) until mid-log phase. At mid-log phase equivalent amounts of cells (turbidity x volume = 20, i.e., if the turbidity value was equal to 5.10, then 3.92 ml of cells were decanted) were removed from the growth tubes into 15 ml Falcon tubes and pelleted by centrifugation. The pellet was washed once in minimal media and resuspended in 1 ml of the required minimal media (*R. sphaeroides* 10 ml RCVBN + 8 ml RLB and *R. palustris* FEM). A 150 µl volume was of each cell line suspension was inoculated in triplicate; this was time point 0. Turbidity measurements were performed using a McFarland Densitometer DEN-1B that measures at λ = 565 ±15 nm. Turbidity measurements were then taken every 6 h for a total experiment length of 72 h.

DNA methods and plasmid construction

Total genomic DNA (gDNA) was extracted from *R. sphaeroides* and *R. palustris* strains using the ZR Bacterial DNA Miniprep Kit (Zymo Research) with a BIO101/Savant FastPrep FP120 high-speed bead beater and a 30 min incubation at 60°C with 20 µl of proteinase K (20 mg/mL), or the Epicentre MasterPure DNA Purification Kit (Epicentre Biotechnologies). All plasmid constructs were built with the Gibson Assembly Master Mix or the NEB Builder HiFi DNA Assembly (both New England

Biolabs). Plasmids were isolated using the AxyPrep Plasmid Miniprep Kit (Axygen). All plasmid constructs were confirmed by Sanger sequencing using both forward and reverse primers. Table S2 lists all plasmids used. Table S3 lists all primers used in this study.

Construction of *R. sphaeroides* and *R. palustris* knock-out strains

To knock out genes in *R. sphaeroides*, a knock-out construct was assembled into the suicide plasmid vector pZDJ (Brimacombe et al. 2013) with either the Gibson Assembly Cloning Kit or the NEBuilder HiFi DNA Assembly Master Mix (both New England Biolabs). The antibiotic resistance cassette flanked by flippase recognition target (FRT) sites used to interrupt the genes to be knocked out corresponds to that used in the Keio collection (Baba et al. 2006). The resulting suicide vector with the knock-out construct was cloned into *E. coli* S17-1 λ-pir-RP4, which can replicate the suicide plasmid. This strain was then conjugated with *R. sphaeroides 2.4.1*. Briefly, donor and recipient cells grown to stationary phase were mixed in a 1:2 volume ratio and pelleted by centrifugation for 1 min at 4,500-6,000 × g, and then washed twice with antibiotic-free RCVBN minimal medium. After the last wash, the pellet was resuspended in 50 µl of RCVBN, and 10 µl aliquots were spotted onto antibiotic-free RCVBN solid medium. The plates were incubated at 30° C overnight to allow conjugation to take place. Afterwards, an emulsion was made from the several inoculation spots and streaked onto antibiotic-containing RCVBN+Gm solid medium. *R. sphaeroides* exconjugants were then successively streaked onto new LB+Gm solid medium until no *E. coli* S17-1 λ-pir-RP4 contamination remained. The resulting pure *R. sphaeroides* exconjugants contained the pZDJ plasmid, with the knock-out gene construct, integrated in the chromosome by a first crossing over (recombination) event. To induce a second crossing over to excise the pZDJ suicide plasmid from the host chromosome, colonies were picked and grown on liquid LB with no antibiotic selection until late stationary phase (about 2-3 days). The cultures were then streaked on LB+10% sucrose solid medium, which allows for counter-selecting of those colonies that have lost the integrated pZDJ plasmid. The counter-selection relies on the *sacB* gene carried by the suicide plasmid (Jaschke et al. 2011). In order to induce a third crossing over between the FRT sites of the kanamycin cassette, cells were grown in liquid LB without any antibiotic selection until late stationary phase and then streaked on LB solid medium. Resultant colonies were then screened through PCR assays.

To make knock-out strains for *R. palustris*, the same general protocol used for *R. sphaeroides* was followed (Welander et al. 2012). The suicide plasmid used for R. palustris was pJQ200SK (Quandt and Hynes 1993) and the host strain was *E. coli* BW29427-λpir-RP4, which requires 300 µM diaminopimelic acid for growth. This auxotrophy allows to easily remove the plasmid-donor bacterium from the medium after conjugation.

Construction of *R. sphaeroides* and *R. palustris* overexpression strains

To create Mic60- and Orf52-overexpressing strains, the inducible expression plasmids for *R. sphaeroides* pIND4 -Km and pIND4-Gm were kindly provided by Judith P. Armitage (University of Oxford), and Alexander Westbye (University of British Columbia), respectively. The inducible expression plasmid pSRK was used in *R. palustris*. The coding sequences of the *mic60* and *orf52* genes of *R. sphaeroides* and *R. palustris* were amplified by PCR and assembled into the expression vectors pIND4 and pSRK, respectively, with the NEBuilder HiFi DNA Assembly Master Mix (New England Biolabs). The *mic60* and *orf52* homologs were cloned into kanamycin resistance-conferring plasmids and gentamycin resistance-conferring plasmids, respectively. To insert the C-terminal 6xHis tag as part of the coding sequences of *mic60* and *orf52* the BamHI and BglII restriction sites in pIND4 were used, or the tag was incorporated as part of the primer used to amplify the targets in *R. palustris*. The assembled plasmids were transformed into suitable conjugative *E. coli* hosts, and conjugation assays with recipient *R. sphaeroides* and *R. palustris* strains were done as described above. After conjugation, the exconjugants were repeatedly streaked onto antibiotic-containing plates to remove the plasmid-donor bacterium. The resulting overexpression strains were further verified by RT qPCR.

RT qPCR

R. sphaeroides and *R. palustris* strains were grown in triplicate until mid-log phase as described for growth analysis, after which 1 mM IPTG was added, and the cells allowed to grow for 7 h 20 min for *R. sphaeroides* and 4 h 30 min for *R. palustris*; cultures were grown in parallel in the absence of IPTG. Then 1 ml of cell suspension was centrifuged and the pellet resuspended in 1 ml of PGTX nucleic acid extraction buffer (Pinto et al. 2009). The samples were then flash frozen and stored at -20 °C until the PGTX-mediated nucleic acid extraction procedure, essentially following the classical phenol/chloroform extraction method (Pinto et al., 2009). Briefly, isolated total nucleic acid concentration was measured using a NanoDrop DeNovix DS-11 (Thermo Fisher) and diluted to 100 ng/µL in 50 µl DNase/RNase-free H_2O. The digestion of gDNA was performed by addition of DNase I (Qiagen) and the concentration of total RNA was measured using the Qubit 2.0 Fluorometer (Thermo Fisher). A total of 100 ng RNA was subsequently reverse transcribed using the Transcriptor First Strand cDNASynthesis Kit (Roche) at 55 °C for cDNA synthesis. Quantitative PCR (qPCR) was performed in triplicates in a CFX96 qPCR cycler (Bio-Rad) in 20 µl reactions containing 1x PowerUp Sybr Green master mix (Applied Biosystems, USA), 8 pmol of each primer, and 10 µg cDNA with the following program: initial denaturation 10 min at 95 °C; 45 cycles of 15 s at 95 °C and 30 s at 60 °C. Primers annealing to *mic60*, *orf52*, and the *rpoZ* cDNAs used for qPCR are listed in Table S3. Cycle threshold values were automatically computed with the CFX Maestro software (Bio-Rad). Additionally, non-reverse transcribed RNA sample was used as a control to verify complete degradation of genomic DNA. Relative abundance of transcripts in cells grown in the presence of IPTG compared to those grown without the inducing agent was calculated using the $2^{-\Delta\Delta CT}$ method (Pfaffl 2001); the unaffected *rpoZ* housekeeping gene cDNA was used to normalize the *mic60* and *orf52* values.

Transmission electron microscopy

Both classical TEM and electron tomography on 80 nm-thick sections were done as previously described (Cadena et al., 2021; Kaurov et al., 2018). Scoring of *R. palustris* ICM area/TEM area and *R. sphaeroides* tubules/cell per section were performed blinded on images obtained using JEOL 1010 TEM operating at an accelerating voltage of 80 kV and equipped with a MegaView III CCD camera (SIS). Measurement of *R. palustris* ICM and total cell areas was done using Image J software (Schneider et al. 2012) by tracing along the outermost electron dense membranes of the ICM network and the outer membrane, respectively. The occurrence of tubule-like ICMs in *R. sphaeroides* was counted on 20-21 images that were taken at 40,000x magnification (2.56 nm/pixel) at random parts of the grid as before (Kaurov et al., 2018). These images were mixed and randomized prior to blind scoring.

Electron tomograms were collected at a range of ±65º with tilt 1º steps using the JEOL 2100F TEM working at 200 kV, equipped with Gatan camera K2 Summit and controlled with SerialEM automated acquisition software (Mastronarde 2005). IMOD software (Mastronarde and Held 2017) was used for tomogram reconstruction and generating 3D models by segmentation.

SDS PAGE and Western blotting

Bacterial lysates were separated on a Bolt 4-12% Bis-Tris Plus gel (Invitrogen), blotted onto a PVDF membrane (Amersham), blocked in 5% low-fat, powdered milk (w/v) in phosphate buffered saline with 0.1% Tween 20 (v/v) (PBS-T), and probed with 6x-His Tag Monoclonal Antibody (HIS.H8) (Thermo Fisher Scientific #MA1-21315) diluted in 5% milk in PBS-T (1:500). This was followed by incubation with secondary HRP-conjugated anti-mouse antibody (1:2000; Bio-Rad). Proteins were visualized using the Pierce ECL system (Genetica/Bio-Rad) on a ChemiDoc imager (Bio-Rad).

Isolation of ICMs via high-pressure homogenization and light spectroscopy

R. sphaeroides strains were cultured identically in C-succinate media (Cohen-Bazire et al. 1957) in flat, glass bottles at 28-30°C under anaerobic conditions. Illumination was provided by one incandescent 40W bulb providing ~10 µmol photons m^{-2} s^{-1}. The cells were fully adapted to the incident light-intensity and harvested by centrifugation once they had reached mid-log phase. Cell pellets were washed in 20 mM MES, 100 mM KCl, pH 6.8, then flash frozen and stored at −80 °C until required

The cell pellets were re-suspended in 20 mM Tris.Cl, pH 8.0 and homogenised thoroughly with a few grains of DNAse I (Qiagen) and a few mg of $MgCl_2$. The cells were broken by passage three times through an Emulsiflex-CS cell disrupter (Avestin). The ruptured cell solution was first subjected to a low-speed centrifugation step (10 min, 10,000 × *g*, 4°C) to remove any unbroken cells. The decanted supernatant was ultra-centrifuged (120 min, 180,000 × *g*, 4°C), after which the supernatant was discarded. The resulting chromatophore pellet was gently re-suspended in 20 mM Tris-HCl pH 8.0 before the optical density was adjusted to 10 cm^{-1} at the Qx absorption maximum (~590 nm) using a Shimadzu UV-Vis-NIR UV2600 spectrophotometer equipped with an integrating sphere.

Blue Native PAGE

BN-PAGE of isolated ICMs was adapted from published protocols (Cadena et al., 2021). Briefly, 1 mg of total protein was resuspended in 100 µl NativePAGE sample buffer (Invitrogen), lysed with 1.5% digitonin (v/v) and 0.1% Triton X-100 (v/v) for 1 h on ice then cleared by centrifugation (22,000 × g, 20 min, 4 °C). Subsequently, 5% Coomassie brilliant blue G-250 was added before loading ~100 µg on a 3-12% Bis-Tris BNE gel (Invitrogen). After electrophoresis (2.5 hours, 150 V, 4 °C), the gel was blotted onto a PVDF membrane (Amersham) and probed as described above.

Affinity purification

Affinity purification (AP) of tagged proteins from 1 mg isolated ICMs were solubilized in IPP50 buffer (50 mM KCl, 20 mM Tris-HCl pH 7.7, 3 mM $MgCl_2$, 10% glycerol, 1 mM phenylmethanesulfonyl fluoride, complete EDTA free protease inhibitor cocktail (Roche) supplemented with 1.5% digitonin (v/v) and 0.1% Triton X-100 (v/v) for 1 h on ice. After centrifugation (22,000 ×g, 20 min, 4 °C) the supernatant was added to 2.0 mg of cobalt conjugated Dynabeads (Thermo Fisher) to capture the His-tag. The Dynabeads were pre-washed in 300 µl of IPP50 + 1.5% digitonin for 5 min at RT. The solubilized ICMs were rotated with beads for 90 min at 4 °C. After removal of the flow-through, the beads were washed three times in IPP50 + 1.5% digitonin. Prior to eluting, the beads were transferred into a new tube. Elution was done with 300 mM imidazole in IPP50 for 15 min at RT and shaking at 1000 rpm. The elutes were further processed for $LC\text{-}MS^2$ analysis or resolved by SDS-PAGE. APs were performed in triplicate.

Protein preparation and mass spectroscopy

Individual bands containing proteins of interest were excised from Coomasie stained SDS PAGE gel using a razor blade and cut into small pieces (~1 mm^3). Bands were destained by sonication for 30 min in 50% acetonitrile (ACN) and 50 mM ammonium bicarbonate (ABC). After destaining, gels were dried in ACN. Disulfide bonds were reduced using 10mm DTT in 100mM ABC at 60°C for 30 min. After that, samples were again dried with ACN and free cysteine residues were blocked using 55 mM iodoacetamide in 100 mM ABC for 10 min at room temperature in the dark. Samples were dried thoroughly and then digestion buffer (10% ACN, 40 mM ABC and 13 ng/µl trypsin) was added to cover gel pieces. Proteins were digested at 37 °C overnight. After digestion, 150 µl of 50% ACN with 0,5% formic acid was added and sonicated for 30 min. Supernatant containing peptides was transferred to a new microcentrifuge tube and another 150 µl of elution solution was added and sonicated for 30 min. This solution was removed, combined with the previous solution and dried by SpeedVac. Dried peptides were reconstitued in 2% ACN with 0,1% TFA and injected into Ultimate 3000 Nano LC coupled to Orbitrap Fusion.

Eluates of co-AP proteins and thin-sliced BN-PAGE gels were processed for MS analysis as described elsewhere (Cadena et al., 2021). In brief, eluate samples were resuspended in 100 mM tetraethylammonium bromide containing 2% sodium deoxycholate. Cysteines were reduced with 10 mM tris(2-carboxyethyl)phosphine and subsequently cleaved with 1 µg trypsin overnight at 37 °C. After digestion, 1% trifluoroacetic acid (TFA) was added to wash twice and eluates were resuspended in 20 µl TFA per µg of protein. A nano reversed-phased column (EASY-Spray column, 50 cm x 75 µm inner diameter, PepMap C18, 2 µm particles, 100 Å pore size) was used for LC/MS analysis. Mobile phase buffer A consisted of water and 0.1% formic acid. Mobile phase D consisted of acetonitrile and 0.1% formic acid. Samples were loaded onto the trap column (Acclaim PepMap300, C18, 5 µm, 300 Å pore size, 300 µm x 5 mm) at a flow rate of 15 µl/min. The loading buffer consisted of water, 2%

acetonitrile, and 0.1% TFA. Peptides were eluted using a Mobile phase B gradient from 2% to 40% over 60 min at a flow rate of 300 ml/min. The peptide cations eluted were converted to gas-phase ions via electrospray ionization and analyzed on a Thermo Orbitrap Fusion (Q-OT- qIT, Thermo Fisher). Full MS spectra were acquired in the Orbitrap with a mass range of 350-1, 400 *m/z*, at a resolution of 120,000 at 200 *m/z* and with a maximum injection time of 50 ms. Tandem MS was performed by isolation at 1,5 Th with the quadrupole, high-energy collisional dissociation (HCS) fragmentation with normalized collision energy of 30, and rapid scan MS analysis in the ion trap. The MS/MS ion count target was set to 10^4 and the max infection time at 35 ms. Only those precursors with a charge state of 2-6 were sampled. The dynamic exclusion duration was set to 45 s with a 10 ppm tolerance around the selected precursor and its isotopes. Monoisotopic precursor selection was on with a top speed mode of 2 s cycles.

Analysis of mass spectroscopy peptides

Label-free quantification of the data were analyzed using the MaxQuant software (version 1.6.2.1) (Cox et al. 2014). The false discovery rates for peptides and for proteins was set to 1% with a specified minimum peptide length of seven amino acids. The Andromeda search engine was used for the MS/MS spectra against the *R. sphaeroides* 2.4.1 predicted proteome (ASM1290v2) downloaded from NCBI GenBank on October 2020. Enzyme specificity was set to C-terminal Arg and Lys, alongside for cleavage at proline bonds with a maximum of two missed cleavages. Dithiomethylation of cysteine was selected as a fixed modification with N-terminal protein acetylation and methionine oxidation as variable modifications. The 'match between runs' feature in MaxQuant was used to transfer identification to other LC-MS/MS runs based on mass and retention time with a maximum deviation of 0.7 min. Quantifications were performed using a label-free algorithm as previously described (Cox et al., 2014). Data analysis was performed using Perseus software (version 1.6.1.3). Eluate co-AP proteins identified with a mean Log2 ratio ($protein_{His}$/WT) >3-fold change and having a Andromeda confidence score >100 (Cox et al., 2011) in three independent biological replicates were analyzed, while gel-slice proteins identified with a mean Log2 transformed LFQ score >23 and present in four biological replicates were analyzed.

Data Availability

The LC-MS/MS data have been deposited to the ProteomeXchange Consortium (http://www.proteomexchange.org) via the PRIDE partner repository with the data set identifier PXD032747.

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Supplemental Material

Figure S1. Verification of *mic60* and *orf52* knock-out strains and IPTG-induced overexpression of Mic60 and Orf52**. (A & B)** PCR assays confirm the disruption of the *mic60* and *orf52* genes in *R. palustris* **(A)** and *R. sphaeroides* **(B)**. Genomic contexts of the relevant loci for the WT, Δ*mic60* and Δ*orf52* strains shown on top, with primer pairs and their expected amplicon sizes shown below each schematic gene arrangement. Lower panel shows each PCR amplicon from each strain (labelled above gel) after agarose gel electrophoresis. Size markers shown either to the left or right of the gel. **(C)** Real time PCR showing relative abundancies of Mic60 and Orf52 mRNAs in *R. palustris* (left) and *R. sphaeroides* (right) strains grown in the presence of the expression induction agent IPTG relative to the same strains grown without IPTG. Error bars show standard deviation from 3 replicates of assayed induced and non-induced cells.

Figure S2. Proteomic analysis of isolated ICMs plus Mic60 and Orf52 interactomes. **(A)** Blue native gel resolved detergent-solubilized ICMs from *R. sphaeroides* WT in which the ~250 kDa band (boxed) was excised in quadruplet for subsequent MS analysis. To right of gel is a histogram of the Coomassie-stained band intensities (*I*) *along* the vertical axis of the run. The scissors denote the ~250 kDa band intensity signal. **(B & C)** A list of all proteins, including excluded contaminants, found within the enriched protein area of the volcano plots in in Figure 4A-B for Mic60 **(B)** and Orf52 **(C)**. Columns as described in legend of Figure 5C-D. Note the presence of likely contaminant found in both APs, PAS-fold containing histidine kinase (PAS), which is a large protein amenable to LC-MS/MS in contrast to RSP_7517 and not found in any of our requisite proteomes. **(D)** Kyle and Doolittle hydropathy plot of RSP_7517, whose amino acid sequence is given below. Predicted transmembrane domain shaded and a potential oligomerization motif AxxxA underlined. Lysine (K) residues recognized by trypsin protease are in bold.

Dataset 1. List of proteins found within Blue Native gel slices as indicated in Figures 3C and S4A in quadruplicates.

Dataset 2. List of all proteins found in the Orf52 and Mic60 interactomes in comparison to WT negative controls by MS of AP eluates. Related to Figure 4A-B.

Table S1. List of strains used in this study.

Table S2. List of plasmids used in this study.

Table S3. List of primers and their sequences used in this study.

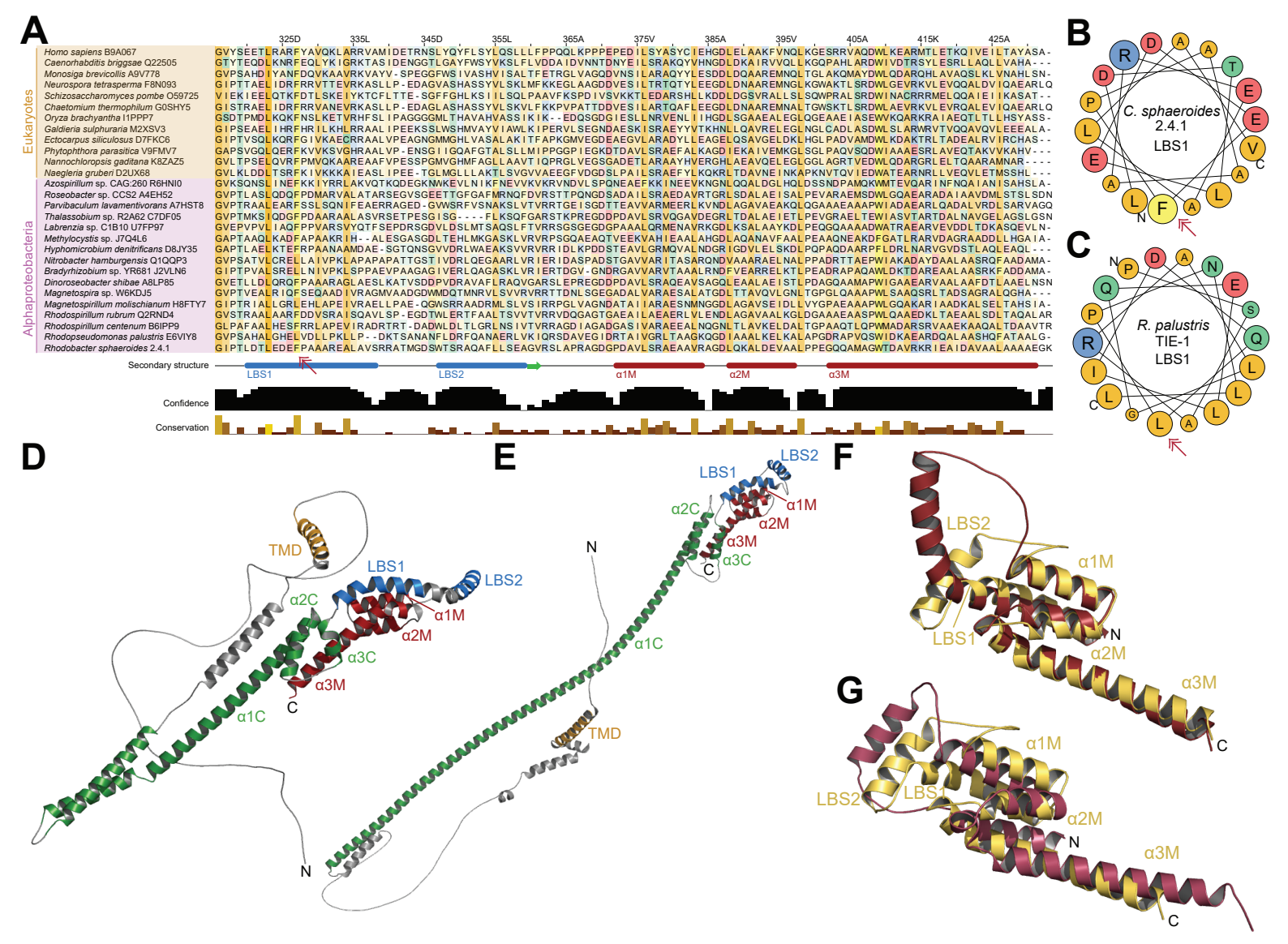
Movie S1. 3D reconstruction model of *R. sphaeroides* WT as rendered from electron tomograms shown at the start of the video. Red, ICM membranes; yellow, cytoplasmic membrane; green, outer membrane. Lower right corner, 200 nm scale bar.

Movie S2. 3D reconstruction model of *R. sphaeroides* Δ*mic60* as rendered from electron tomograms shown at the start of the video. Red, ICM membranes; yellow, cytoplasmic membrane; green, outer membrane. Lower right corner, 200 nm scale bar.

Movie S3. 3D reconstruction model of *R. sphaeroides* Mic60↑ as rendered from electron tomograms shown at the start of the video. Red, ICM membranes; yellow, cytoplasmic membrane; green, outer membrane. Lower right corner, 200 nm scale bar.

Movie S4. 3D reconstruction model of *R. sphaeroides* Δ*orf52* as rendered from electron tomograms shown at the start of the video. Red, ICM membranes; yellow, cytoplasmic membrane; green, outer membrane. Lower right corner, 200 nm scale bar.

Movie S5. 3D reconstruction model of *R. sphaeroides* Orf52↑ as rendered from electron tomograms shown at the start of the video. Red, ICM membranes; yellow, cytoplasmic membrane; green, outer membrane. Lower right corner, 200 nm scale bar.


A
325D
335L
345D
355L
365A
375V
385A
395V
405A
415K
425A
Eukaryotes
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Caenorhabditis briggsae Q22505
Monosiga brevicollis A9V778
Neurospora tetrasperma F8N093
Schizosaccharomyces pombe O59725
Chaetomium thermophilum G0SHY5
Oryza brachyantha I1PPP7
Galdieria sulphuraria M2XSV3
Ectocarpus siliculosus D7FKC6
Phytophthora parasitica V9FMV7
Nannochloropsis gaditana K8ZAZ5
Naegleria gruberi D2UX68
Alphaproteobacteria
Azospirillum sp. CAG:260 R6HNI0
Roseobacter sp. CCS2 A4EH52
Parvibaculum lavamentivorans A7HST8
Thalassobium sp. R2A62 C7DF05
Labrenzia sp. C1B10 U7FP97
Methylocystis sp. J7Q4L6
Hyphomicrobium denitrificans D8JY35
Nitrobacter hamburgensis Q1QQP3
Bradyrhizobium sp. YR681 J2VLN6
Dinoroseobacter shibae A8LP85
Magnetospira sp. W6KDJ5
Magnetospirillum molischianum H8FTY7
Rhodospirillum rubrum Q2RND4
Rhodospirillum centenum B6IPP9
Rhodopseudomonas palustris E6VIY8
Rhodobacter sphaeroides 2.4.1
Secondary structure
LBS1
LBS2
α1M
α2M
α3M
Confidence
Conservation
B
C. sphaeroides
2.4.1
LBS1
C
R. palustris
TIE-1
LBS1
D
TMD
LBS1
LBS2
α2C
α1M
α2M
α3C
α3M
α1C
C
N
E
LBS2
LBS1
α2C
α1M
α2M
α3M
α3C
C
N
α1C
TMD
N
F
LBS2
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C
G
α1M
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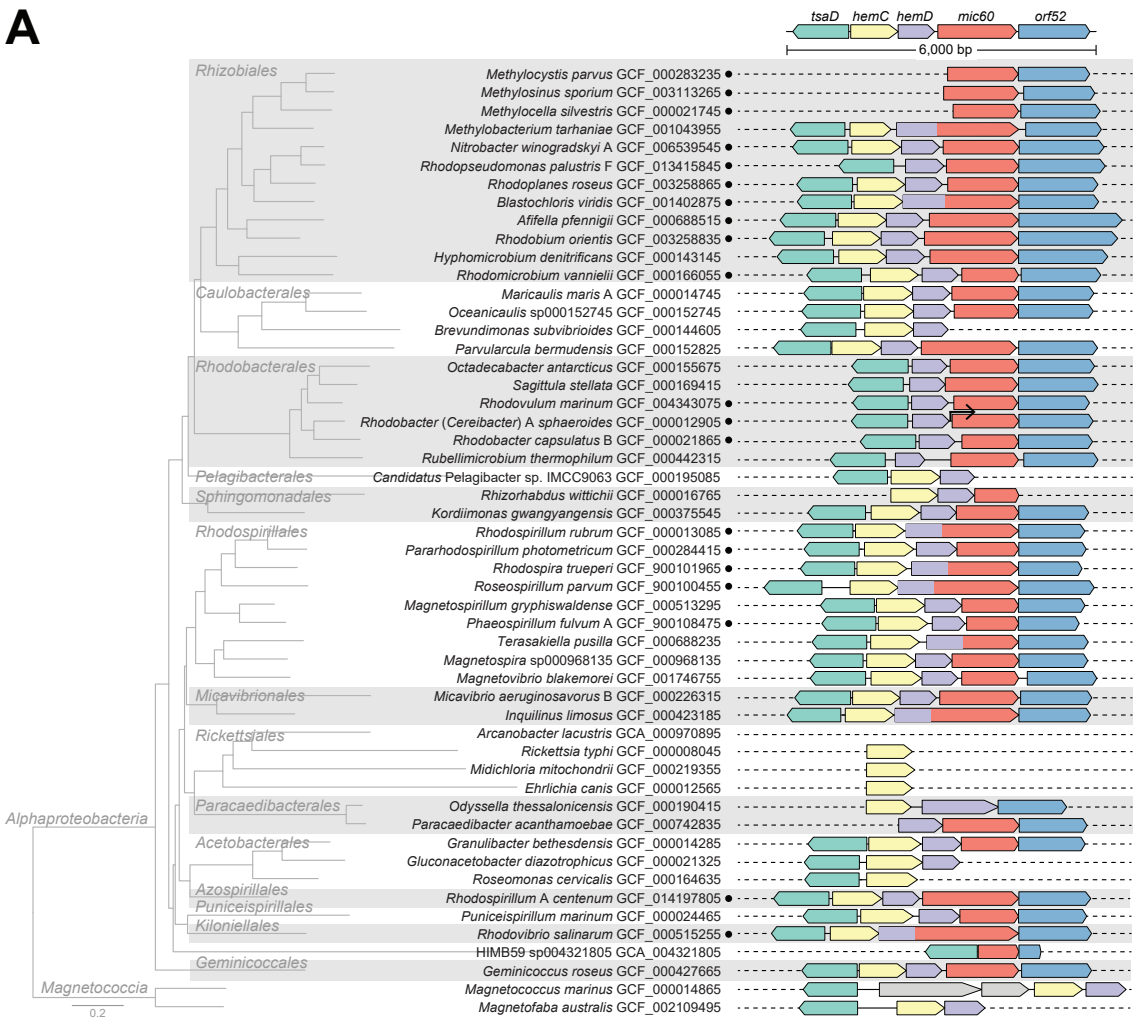
LBS1
α2M
N
α3M
C


A
tsaD
hemC
hemD
mic60
orf52
6,000 bp
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Methylocella silvestris GCF_000021745
Methylobacterium tarhaniae GCF_001043955
Nitrobacter winogradskyi A GCF_006539545
Rhodopseudomonas palustris F GCF_013415845
Rhodoplanes roseus GCF_003258865
Blastochloris viridis GCF_001402875
Afifella pfennigii GCF_000688515
Rhodobium orientis GCF_003258835
Hyphomicrobium denitrificans GCF_000143145
Rhodomicrobium vannielii GCF_000166055
Caulobacterales
Maricaulis maris A GCF_000014745
Oceanicaulis sp000152745 GCF_000152745
Brevundimonas subvibrioides GCF_000144605
Parvularcula bermudensis GCF_000152825
Rhodobacterales
Octadecabacter antarcticus GCF_000155675
Sagittula stellata GCF_000169415
Rhodovulum marinum GCF_004343075
Rhodobacter (Cereibacter) A sphaeroides GCF_000012905
Rhodobacter capsulatus B GCF_000021865
Rubellimicrobium thermophilum GCF_000442315
Pelagibacterales
Candidatus Pelagibacter sp. IMCC9063 GCF_000195085
Sphingomonadales
Rhizorhabdus wittichii GCF_000016765
Kordiimonas gwangyangensis GCF_000375545
Rhodospirillales
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Terasakiella pusilla GCF_000688235
Magnetospira sp000968135 GCF_000968135
Magnetovibrio blakemorei GCF_001746755
Micavibrionales
Micavibrio aeruginosavorus B GCF_000226315
Inquilinus limosus GCF_000423185
Rickettsiales
Arcanobacter lacustris GCA_000970895
Rickettsia typhi GCF_000008045
Midichloria mitochondrii GCF_000219355
Ehrlichia canis GCF_000012565
Paracaedibacterales
Odyssella thessalonicensis GCF_000190415
Paracaedibacter acanthamoebae GCF_000742835
Alphaproteobacteria
Acetobacterales
Granulibacter bethesdensis GCF_000014285
Gluconacetobacter diazotrophicus GCF_000021325
Roseomonas cervicalis GCF_000164635
Azospirillales
Rhodospirillum A centenum GCF_014197805
Puniceispirillales
Puniceispirillum marinum GCF_000024465
Kiloniellales
Rhodovibrio salinarum GCF_000515255
HIMB59 sp004321805 GCA_004321805
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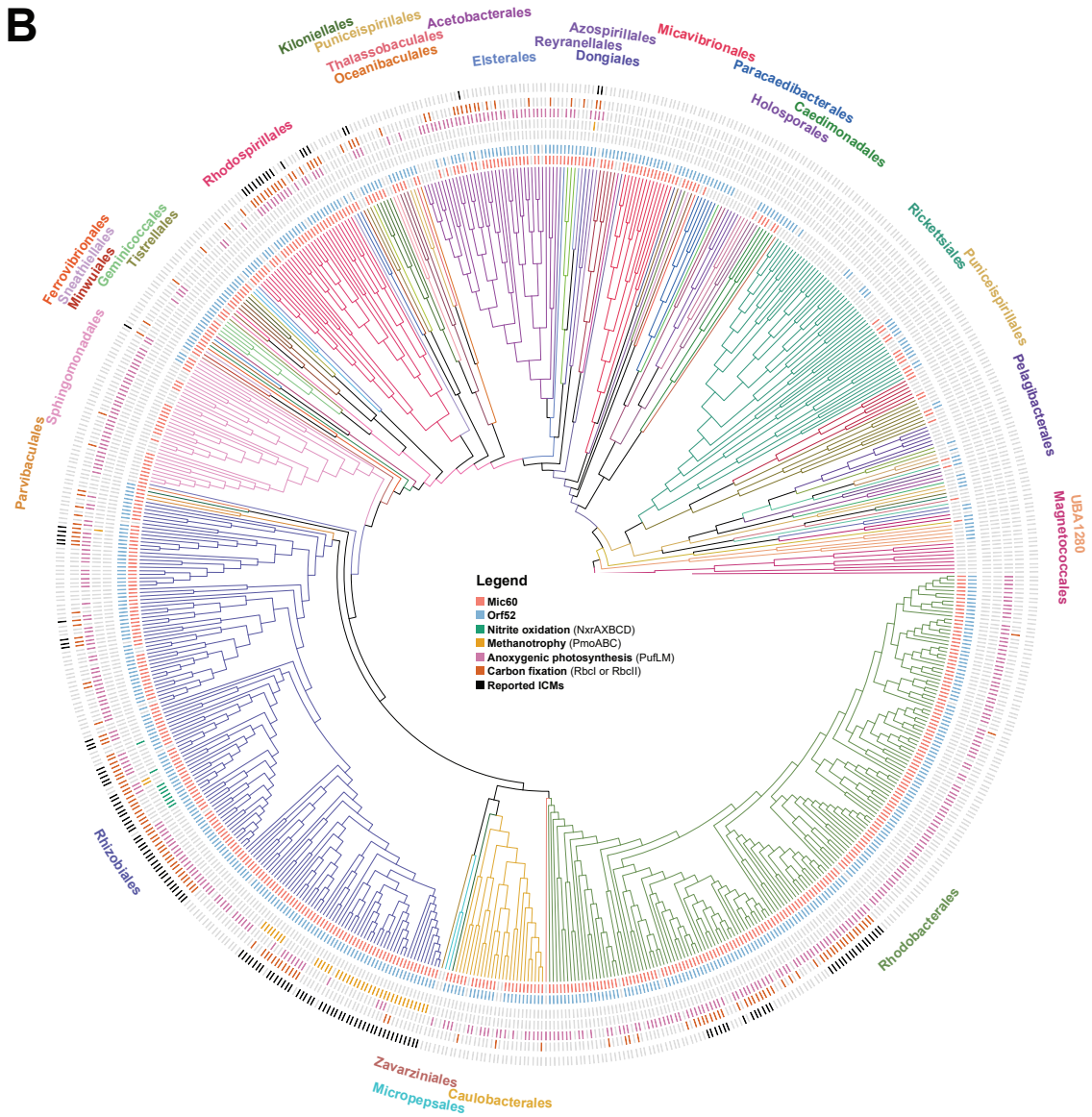
B
Kiloniellales
Puniceispirillales
Acetobacterales
Thalassobaculales
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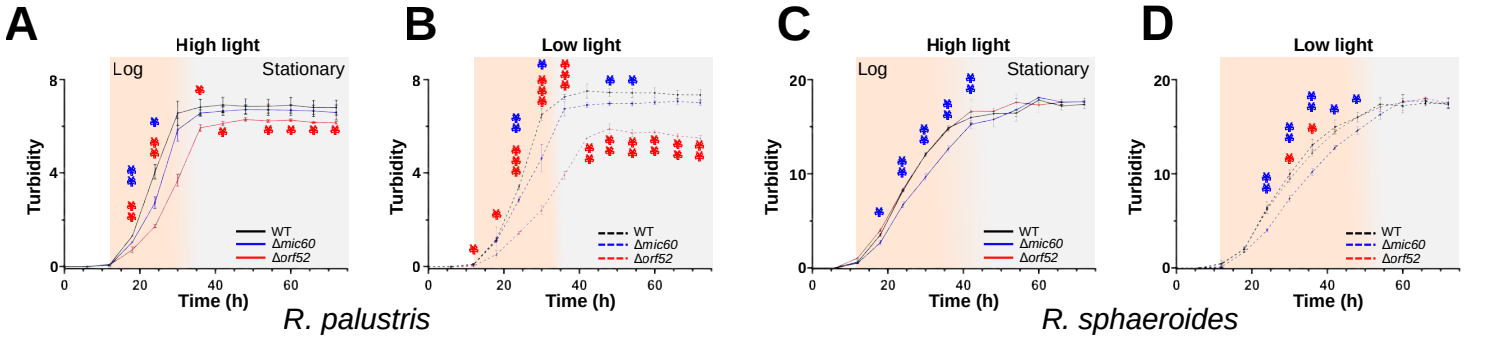
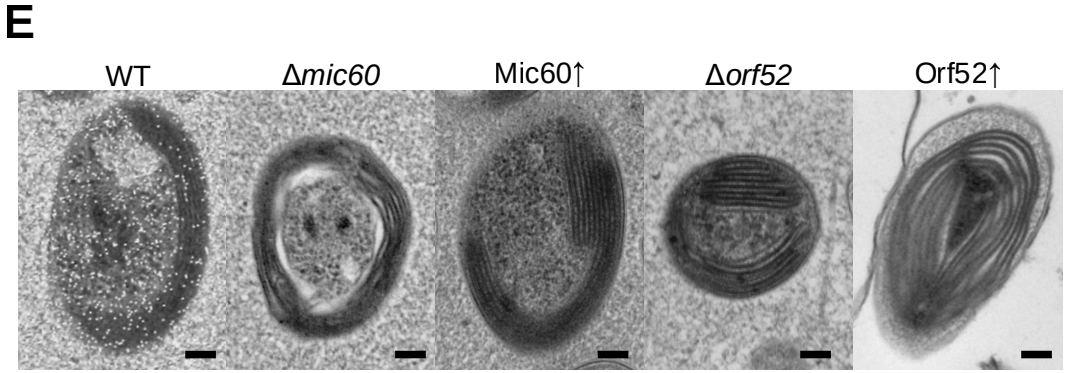
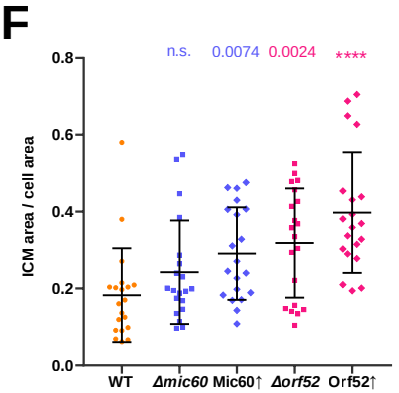
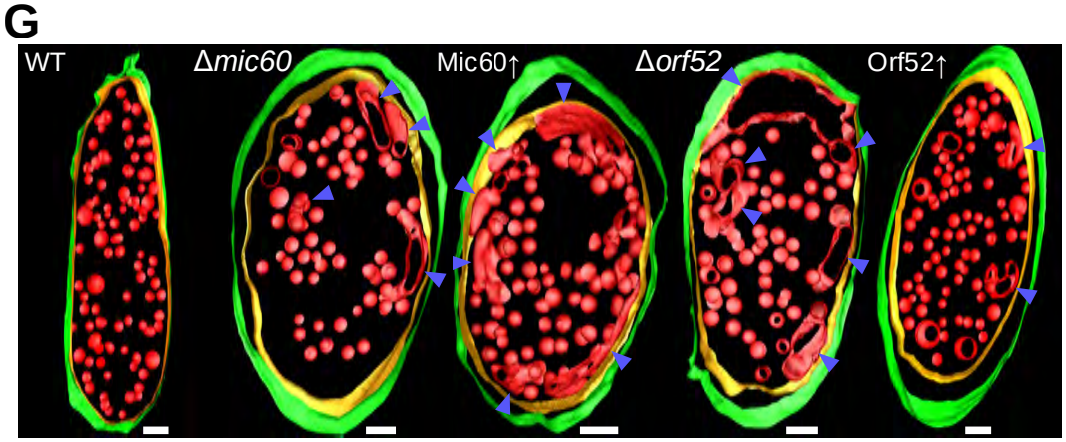
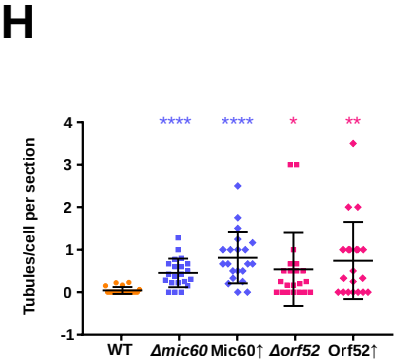
Elsterales
Azospirillales
Reyranellales
Dongiales
Micavibrionales
Paracaedibacterales
Caedimonadales
Holosporales
Rickettsiales
Puniceispirillales
Pelagibacterales
UBA1280
Magnetococcales
Rhodobacterales
Rhizobiales
Zavarziniales
Caulobacterales
Micropepsales
Parvibaculales
Sphingomonadales
Ferrovibrionales
Sphaerobacterales
Minwuales
Geminicoccales
Tistrellales
Rhodospirillales
Legend
Mic60
Orf52
Nitrite oxidation (NxrAXBCD)
Methanotrophy (PmoABC)
Anoxygenic photosynthesis (PufLM)
Carbon fixation (RbcI or RbcII)
Reported ICMs



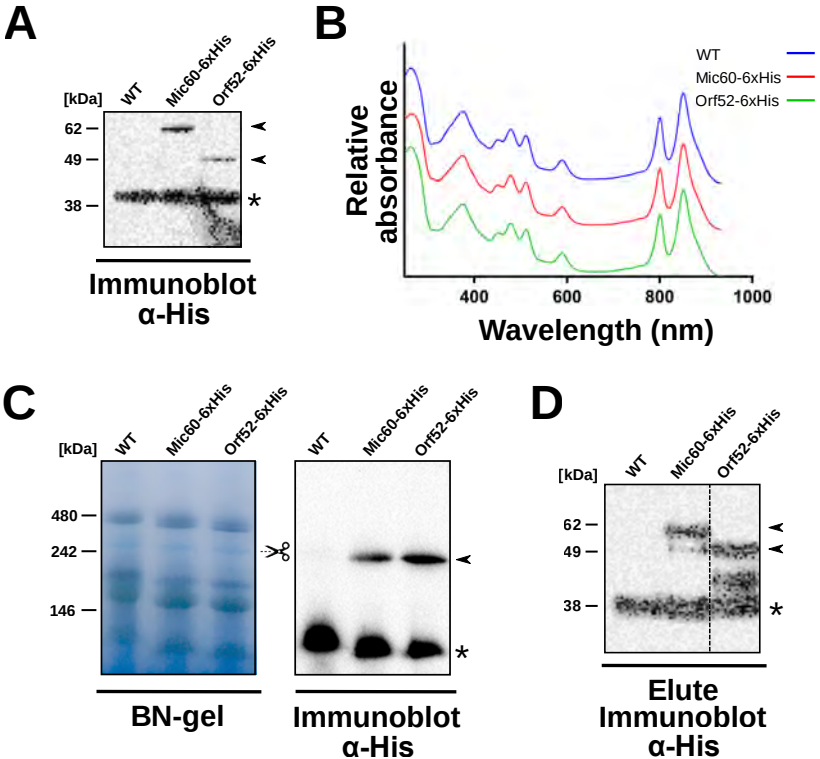


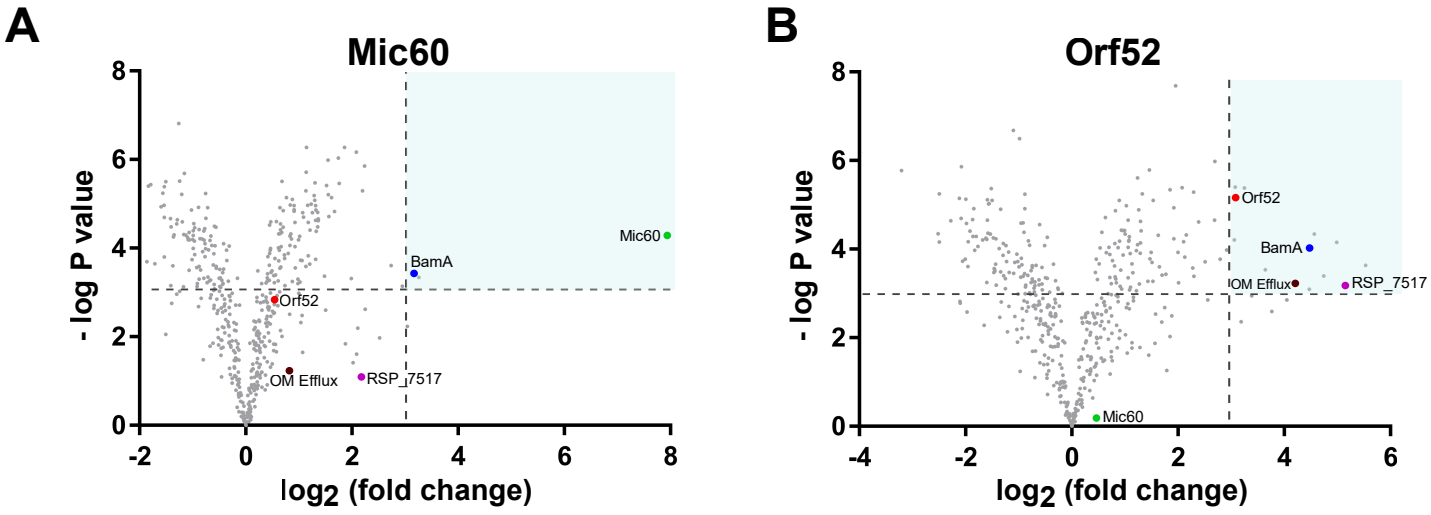






	WT	Δmic60	Mic60↑	Δorf52	Orf52↑
Sections (n):	20	21	21	21	20
Total cells:	267	152	80	87	51
Total tubules:	11	66	56	29	30


A
[kDa]
WT
Mic60-6xHis
Orf52-6xHis
62
49
38
Immunoblot
α-His
B
Relative absorbance
WT
Mic60-6xHis
Orf52-6xHis
400
600
800
1000
Wavelength (nm)
C
[kDa]
WT
Mic60-6xHis
Orf52-6xHis
480
242
146
BN-gel
Immunoblot
α-His
D
[kDa]
WT
Mic60-6xHis
Orf52-6xHis
62
49
38
Elute
Immunoblot
α-His



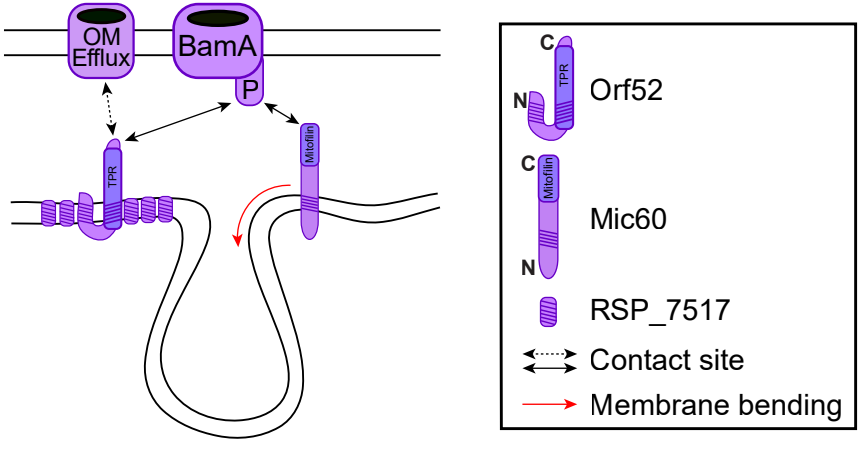
C

Mic60

Protein	Fold change	UPB ICM	250 kDa complex	Mol. weight [kDa]
Mic60	245.13	○ ○*	●	43.8
BamA	9.09	●—●	●	88.2

D

Orf52

Protein	Fold change	UPB ICM	250 kDa complex	Mol. weight [kDa]
RSP_7517	35.62	○ ○	◉	4.8
BamA	22.38	●—●	●	88.2
OM Efflux protein	18.52	●—●	●	49.7
Orf52	8.50	●—●	●	52.5

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