Modeling foot-and-mouth disease dissemination in Brazil and evaluating the effectiveness

of control measures

Nicolas C. Cardenas[1], Francisco P. N. Lopes[2], Gustavo Machado[1,*]

Running title: Foot-and-mouth disease dissemination across species.

[1] Department of Population Health and Pathobiology, College of Veterinary Medicine, North

Carolina State University, Raleigh, NC, USA

[2] Departamento de Defesa Agropecuária, Secretaria da Agricultura, Pecuária e Desenvolvimento

Rural, Porto Alegre, Brazil

Abstract

Foot-and-mouth disease (FMD) infects multiple food-animal species and disseminate among

ungulate species. Here, we present a multiscale compartmental stochastic model that considers

population dynamics, births, deaths, and species-specific transmission dynamics. The model

considers two between-farm modes of FMD transmission integrating disease dynamics at the

between-farm and within-farm levels. Our model outputs include the number of secondarily

infected animals and farms, spatial dissemination, the role of animal movement and between-

farm distances as transmission pathways, and the effectiveness of countermeasures. Here, we

demonstrated that after 20 days of FMD dissemination, the median number of infected farms was

8. Spatial proximity was the predominant route associated bovine infection while in swine were

linked with animal movements. Furthermore, the median distance between seeded and secondary

infections was 5.77 with the highest spatial dissemination reaching 695.40 kilometers. The

simulated control strategy results showed that depopulating 12 farms and vaccinating 15,000

farms per day after 20 days of silent FMD dissemination would contain 93.4% of epidemics with

a median of 9 infected farms within 54 days after implementing control actions. In conclusion,

the developed model highlights the need for developing multispecies FMD transmission models,

especially in regions where multiple species are raised on the same premises.

Keywords: dynamical models, infectious disease control, epidemiology, transmission, targeted

control.

1.   Introduction

Foot-and-mouth disease (FMD) is an infectious disease in cloven-hoofed animals that affects

multiple species, including pigs, sheep, goats, and wildlife[1]. During the 2001 FMD epidemic in

the U.K. and the Netherlands, more than 6.7 million animals were slaughtered, including those

from healthy (preemptive measures) and infected farms [2]. The infected animals in both outbreaks

were not solely from one species; for example, infection was also detected in goats on mixed

dairy-goat/veal-calf farms[2,3]. The agricultural sector loss was approximately 3.2 billion euros,

and there were additional costs to other sectors, such as tourism, with a total expenditure of

approximately 2.7 to 3.2 billion euros[4]. In addition, the distribution of FMD outbreaks in South

America has changed over the years, with the most recent outbreaks (in 2017 and 2018)

occurring in Colombia[5]; no epidemics have been reported in Latin America since then, despite

Venezuela's absence of official international status for FMD[6].

The official data from the World Organisation for Animal Health[7], consisting of more

than 57,000 outbreaks in 99 counties from 1996 to 2012, show that 68% of FMD cases were

associated with cattle, 22% with swine, and 21% with small ruminants and other species; buffalo

were not directly mentioned[8]. Despite strong evidence indicating that all susceptible species can

drive large FMD epidemics[9], such as the 2001 UK epidemic (traced back to an infected Essex

swine slaughterhouse[10]), control measures and surveillance programs have mostly targeted

dissemination within cattle populations, thus neglecting the role of other species in FMD

transmission[11].

Notably, FMD pathogenesis and transmission dynamics vary among species, given

differences in viral loads needed to cause infection, viral latency and infection duration[12–15]. For

example, swine shed more viral particles than cattle and sheep, with widespread epidemics

expected when swine are also infected[14]. Thus, it is pivotal to consider such heterogeneity in

transmission dynamics when modeling within- or between-farm FMD dissemination[11]. While

between-farm transmission primarily occurs through direct contact among susceptible and

infected animals[12,16], field observations and experimental trials have demonstrated the spread of

the virus via indirect contact with fomites and long-distance transport of aerosols, a process

known as spatial transmission[16].

Mathematical models have been widely used to investigate FMD epidemic propagation.

Although significant technical and computational advancements have been achieved, these

models are simplified because of computational costs and/or lack of details about herd structure

(e.g., number of individuals, number born alive) and animal movements. The most common

simplification is restricting the epidemiological models to a single species[4,17,18]. Despite the

different applications and efforts in modeling FMD, outstanding questions remain regarding

measurements of the epidemic trajectory and epidemic control strategies given heterogeneous

transmission dynamics among the different susceptible species coexisting on the same farm[11].

In this work, we propose a novel multihost, single-pathogen, multiscale model that

incorporates heterogeneous (species-specific) transmission dynamics. We present a quantitative

assessment of each species' epidemiological role in FMD dissemination, aiming to quantify

transmission routes and the spatial distance of spread. Our simulations utilized population data,

including birth and death information for the entire food-animal species population of Rio

Grande do Sul, Brazil. Additionally, we examine the effectiveness of control and elimination

actions according to the national FMD control plan[19], which includes i) vaccination; ii)

depopulation; iii) various restrictions regarding between-farm movement; and iv) surveillance

activities within control zones.

2.   Material and Methods

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2.1 Data sources

We collected information from 415,364 farms officially registered in Rio Grande do Sul,

Brazil[20]. The data contained information on the population (of cattle and buffalo, pigs, and sheep

and goats), number of farms, and daily between-farm movement and movements to

slaughterhouses from January 1st, 2018 to December 31st, 2020. Population and movement data

from cattle and buffalo farms were combined into one category: "bovines". Similarly, we

grouped sheep and goats into "small ruminants". Inspection of the population data revealed 5,709

(1.37%) farms with missing geolocation data; these farms were excluded from subsequent

analysis. Further information about the species population distributions is available in

Supplementary Figure S1. In Brazil, producers are required to report farming activities to the

state animal health office once a year; this report includes information about food-animal

production activities. We utilized producer-reported data combined with the 2020 data to classify

farms as "active" if at least one food animal was present. Farms that were declared inactive by

the producer or that had zero animals were considered inactive and were not included in this

study. Therefore, our final active population included 409,655 farms.

2.2 Birth and death data

Similar to the requirements regarding the total number of individuals, producers must report the

number of animals born alive and the number of animal deaths, including natural deaths. We

accessed such data through the state database[20]. Producers can report birth and death data at any

time throughout the year; however, reporting is mandatory at least once a year in a window from

January 1st to May 30th. We collected 256,504 individual records associated with birth and

death. The monthly births and deaths (by specie) is presented in Supplementary Figure S2. Farm

population data were computed based on animal movement, birth, and death records; of the

409,655 farms, 1.14% were found to have negative populations. Farms with a negative

population caused by animal movement or death were allocated a population value equal to their

most recent positive population.

2.3 Movement data

From January 1st, 2018 to December 31st, 2020, 1,634,940 unique between-farm and

slaughterhouse movements were registered in the state database. We inspected the movement

data and found 181 movements missing farm identification, 93,871 movements with the exact

identification of the origin and destination, 51,490 movements to or from other states, and

112,834 movements from or to farms with missing geolocation information. After removing

these movement data, the final database comprised 1,564,487 movements and 73,823,866

animals transported (Supplementary Figure S3 shows the movement data variations over time).

2.4 Model formulation

We implemented a multihost, single-pathogen, coupled multiscale model to simulate FMD

epidemic trajectories[21] with and without implementing control actions. This model led to the

development of a novel R package, "MHASpread: A animal spread stochastic multilevel model"

(version 0.1.0). MHASpread allows the explicit specification of species-specific transmission

probabilities and latent and infectious periods of a disease that infects multiple species. The

within-farm level includes the entry and exit of animals due to between-farm movements,

movements to slaughterhouses, births, and deaths for each species (Figure 1). Between-farm

movements and spatial proximity drive the between-farm dynamics.

2.5 Within-farm dynamics

The within-farm dynamics consist of mutually exclusive health states (i.e., an individual can only

be in one state per discrete time step) for animals of each taxa (bovines, swine, and small

ruminants). These health states (hereafter, "compartments") include susceptible (S), exposed

(E), infectious, (I), and recovered (R), defined as follows:

(i) Susceptible: animals that are not infected but susceptible to infection.

(ii) Exposed: animals that have been exposed to disease but not yet infected.

(iii) Infectious: infected animals that can successfully transmit the infection.

(iv) Recovered: animals that have recovered and are no longer susceptible.

Here, we also consider the birth and death data described in the section above. The total

population is calculated as $N = S + E + I + R$. The number of individuals within each

compartment transitions from $S\beta \rightarrow E, 1/\lambda_1 \rightarrow I, 1/\lambda_2 \rightarrow R$ according to the following

equations:

$$\frac{dS_i(t)}{dt} = u_i(t) - v_i(t) - \frac{\beta S_i(t) I_i(t)}{N_i} \tag{1}$$

$$\frac{dE_i(t)}{dt} = \frac{\beta S_i(t) I_i(t)}{N_i} - v_i(t) E_i(t) - \frac{1}{\lambda_1} E_i(t) \tag{2}$$

$$\frac{dI_i(t)}{dt} = \frac{1}{\lambda_1} E_i(t) - \frac{1}{\lambda_2} I_i(t) - v_i(t) I_i(t) \tag{3}$$

$$\frac{dR_i(t)}{dt} = \frac{1}{\lambda_2} I_i(t) - v_i(t) \tag{4}$$

In these equations, births are represented by the number of animals born alive ($u_{i,t}$) that enter the

S compartment on farm i at time t according to the day-to-day records; similarly, $v_{i,t}$ represents

the exit of the animals from any compartment due to death at time t (Figure 1). The transition

from E to I is driven by $1/\lambda_1$ and the transition from I to R is driven by $1/\lambda_2$; these values are

both drawn from the distribution generated from each specific taxa according to the literature

(Table 1). As infection progresses, animals transition into a recovered (R) state based on $1/\lambda_2$

(derived from the infectious period); both λ_1 and λ_2 are specific to the host taxa (Table 2).

Figure 1. Within-farm dynamics. The framework represents the transition between

compartments, $S\beta \rightarrow E, 1/\lambda_1 \rightarrow I, 1/\lambda_2 \rightarrow R$. Susceptible animals ($u$) are introduced in the S

compartment, whereas deaths (v) were removed from any compartment.

For within-farm dynamics, we assumed that populations were homogeneously distributed and

that species were homogeneously mixed. The frequency-dependent transmission depends on

both the infected and the susceptible host taxa, as reflected by the species-specific FMD

transmission coefficient β (Table 1, Supplementary Figure S4, and Figure 2A). Similarly, the

transition from E to I and from I to R occurs according to each taxon's parameters (listed in

Table 1).

Table 1. The distribution of each host-to-host transmission coefficient (β) per animal^{-1} day^{-1}.

Infected taxon	Susceptible taxon	Transmission coefficient (β), shape and distribution	Reference
Bovines	Bovines	PERT (0.018, 0.024, 0.056)	Calculated from the 2000-2001 FMD outbreaks in the state of Rio Grande do Sul[22]
Bovines	Swine	PERT (0.018, 0.024, 0.056)	Assumed
Bovines	Small ruminants	Triangular (0.020, 0.026, 0.031)	[23,24]
Swine	Bovines	PERT (0.014, 0.044, 0.033)	[25]
Swine	Swine	PERT (0.014, 0.044, 0.033)	[25,26]
Swine	Small ruminants	PERT (0.014, 0.044, 0.033)	[25]
Small ruminants	Bovines	PERT (0.012, 0.031 0.065)	[27]
Small ruminants	Swine	PERT (0.006, 0.024, 0.09)	[28]

Small ruminants	Small ruminants	PERT (0.018, 00.24, 0.056)	Assumed

158 **Table 2**. The within-farm distribution of latent and infectious FMD parameters for each species.

FMD parameter	Taxon	Distribution	Reference
Latent period, λ_1	Bovines	Weibull (a = 1.78, b = 3.97)	29
	Swine	Log-logistic (shape = 7.60; scale = 1.06)	14
	Small ruminants	PERT (m = 3.96, a = 0, b = 13.98)	29
Infectious period, λ_2	Bovines	Gamma (a = 3.97, b = 1.11)	29
	Swine	Weibull (shape = 7.16; scale = 11.04)	14
	Small ruminants	Pearson type 5 (a = 6.19, b = 17.19)	29

Note: The time unit is days.

2.6 Between-farm dynamics

For the farm-level dynamics, we considered two transmission pathways: i) between-farm

movements and ii) spatial transmission. We utilized the number of unique movements (batches)

and animals from each taxa to model between-farm transmission and considered the current

within-farm transmission dynamics (Figure 2). For each movement batch, we determined the

corresponding infection status of the available population at the farm of origin to determine the

proportions of animals in each compartment transferred to the destination farm (Figure 2B).

Animal movements to slaughterhouses were similarly considered; we drew the number of

animals to be removed from the current state of the system (Figure 2B).

2.7 Spatial transmission dynamics

Spatial transmission encompasses a variety of transmission processes, such as airborne

transmission, animal contact over fence lines and equipment sharing between farms. We also

used each farm's geolocation to group farms according to a grid hexagon of 0.76 km,

representing an area of 503,516 m^2 (Supplementary Figure S5A). Farms within the same

hexagon had the same spatial transmission probability (Figure 2C). Local spread was fitted using

a spatial transmission kernel, in which the likelihood of transmission decreased as a function of

the between-farm distance. The probability PE at time t describes the probability that a farm

becomes exposed and is calculated as follows:

$$PE_j(t) = 1 - \prod_i \left(1 - \frac{I_i(t)}{N_i} \varphi e^{-\alpha d_{ij}}\right) \tag{5}$$

where j represents the uninfected population and αd_{ij} represents the distance between farm j and

infected farm i, with a maximum of 40 km. Given the extensive literature on distance-based

FMD dissemination and a previous comprehensive mathematical simulation study[30], distances

above 40 km were not considered. Here, $1 - \frac{I_i}{N_i} \varphi e^{-\alpha d_{ij}}$ represents the probability of

transmission between farms i and j (scaled by infection prevalence on farm i, $\frac{I_i}{N_i}$) given the

distance between the farms in kilometers (Supplementary Figure S5B). The parameters φ and α

control the shape of the transmission kernel; $\varphi = 0.12$ is the probability of transmission when

$d_{ij} = 0$, and $\alpha = 0.6$ controls the steepness with which the probability declines with distance[31].

Figure 2. Schematic of state transitions during within-farm and between-farm dynamics.

A) Within-farm dynamics: green arrows indicate the introduction of animals (births) into the

susceptible (S) compartment at farm i at time t. Each circle indicates a farm with single or

multiple taxa with specific host-to-host transition parameters (β, λ_1, λ_2); dashed lines represent

interactions within and between host taxa. The red arrows represent the removal of animals

(deaths) regardless of infection status. B) Between-farm dynamics: the layer represents the

number of animals moved (batches; n) from the farm of origin (i) to a destination farm (j) at

time t (indicated by the black dashed arrows). Animals moved to the slaughterhouse were

removed from the simulation regardless of their infection status and are indicated by red dashed

arrows. C) Spatial transmission distances.

2.8 Initial model conditions

We used 2018 population data to represent the initial susceptible population. However,

the sheer number of farms (409,655) within Rio Grande do Sul constituted a computational

bottleneck; we therefore drew a representative sample to model the Rio Grande do Sul

population. Briefly, a multistage and stratified sample was collected according to the number of

farms containing each taxa per municipality[32]. The sample size was obtained by considering a

prevalence of 50% with a level of significance of 95% and a margin of error of 1%, resulting in a

total of 15,721 farms used to seed FMD infection for ten repeats. Each repeat was given a

random start date between January 1st, 2018 and December 31st, 2020, commencing with one

infected animal. In farms with more than one species, the infection seed priority order was as

follows (in descending priority): bovines, swine, and small ruminants. Of note, we assumed that

all animals were susceptible to FMD because the annual vaccination campaign has been

suspended in Rio Grande do Sul since May 2021[22].

2.9 Local sensitivity analysis

We used a combination of Latin hypercube sampling (LHS), developed by McKay[33], and the

partial rank correlation coefficient (PRCC) technique to perform a local sensitivity analysis. LHS

is a stratified Monte Carlo sampling method without replacement that provides unbiased

estimates of modeling output measures subject to combinations of varying parameters. The

PRCC approach can be used to classify how output measures are influenced by changes in a

specific parameter value while linearly accounting for the effects of other parameters[34]. As input

model parameters, we selected the parameters and distributions listed in Tables 1 and 2 with

population sizes of each host taxa (bovines, swine, and small ruminants) ranging from ten to

1,000. In total, 18 parameters were used to classify the monotone relation infection status with

our input variables to classify model sensitivity. The inputs include one farm where the initial

conditions varied across 10,000 simulations over the LHS space. A positive PRCC indicates a

positive relationship with the number of infected animals, whereas a negative PRCC indicates an

inverse relationship with the number of infected animals; however, the magnitude of PRCC does

not necessarily indicate the importance of a parameter[35].

3.1 Evaluation of control and eradication scenarios

We simulated FMD epidemic trajectories for up to 20 days after the initial outbreaks were

seeded. The decision to implement control actions at a maximum of 20 days was based on

historic FMD outbreak data in the region and consultation with local animal health officials.

Consequently, our simulated control actions were implemented after ten and 20 days of silent

FMD dissemination. Thus, the initial numbers of infected farms at ten and 20 days under a

predefined baseline control scenario based on the national FMD response plan[36] were compared

with the numbers of infected farms after implementing the three alternative control scenarios

described below.

Baseline control scenario: This scenario constituted the national FMD control and

eradication plan[36], which includes the following measures: i) depopulation of infected farms, ii)

vaccination of uninfected farms, iii) cessation of animal movement for 30 days, and iv)

establishment of three distinct control zones around infected farms at the following distances: 3

km (infected zone), 7 km (buffer zone), and 15 km (surveillance zone) (Supplementary Figure

S6). In the infected zones, infected farms are depopulated as FMD is detected, and vaccination is

implemented first in uninfected farms within these infected zones. If vaccine doses are still

available after vaccinating the farms in the infected zones, then farms in buffer zones will be

vaccinated. Surveillance activities are defined as clinical examinations. In our simulation, all

farms within control zones were visited every three days. Contact tracing was used to identify

farms with direct links to infected farms within the past 30 days; these farms underwent

surveillance. If positive cases were detected in these farms, animals were moved into the infected

compartment. We also incorporated the state budget as follows: i) the maximum number of

farms depopulated per day was set to four, and ii) the maximum number of animals vaccinated

per day was set to 5,000; both assumptions are based on the state's maximum personnel and

equipment capacity (personal communication, Animal Health Official Working Group). We also

included a delay in the implementation of vaccination equal to seven days after the first case was

detected; vaccination was assumed to be effective immediately, and thus vaccinated animals

were moved into the recovered compartment. In addition, we assumed that i) surveillance was

100% sensitive to FMD and ii) farms exhibited 100% compliance with the 30-day animal

movement cessation.

In addition to the baseline control scenario, three alternative scenarios were considered:

Scenario 1: Vaccination & depopulation. This scenario differed from the baseline control

scenario by increasing the depopulation and vaccination of farms, with depopulation of infected

farms set to eight (vs. four) farms per day and vaccination set to 10,000 (vs. 5,000) animals per

day.

Scenario 2: Depopulation only. This scenario differed from the baseline control scenario

by increasing the depopulation of infected farms to eight (vs. four) per day.

Scenario 3: Enhanced vaccination & depopulation. This scenario differed from the

baseline control scenario by extensively increasing the depopulation and vaccination of farms,

with depopulation of infected farms set to 12 (vs. four) per day and vaccination set to 15,000 (vs.

5,000) animals per day.

3.2 Model outputs

Our simulations tracked the number of animals and farms in each health compartment at each

daily time step. We also utilized the epidemic trajectories to calculate the geodesic distances

between the seeded infections and secondary cases. In addition, we determined the probability of

distance-dependent transmission by calculating the cumulative empirical spatial distribution. By

tracing the average number of secondarily infected animals resulting from each transmission

route, we estimated the contribution of between-farm movement and spatial transmission to

infection in each taxa; here, we present the average contribution of each route along with

credible intervals estimated by using an equal-tailed interval method[37]. We also determined the

numbers of animals and farms within control zones at 10 and 20 days post-FMD dissemination.

We used 10 and 20 days of silent FMD spread as the initial model conditions. We first

removed simulations with more than 400 infected farms because such large epidemics exceeded

the maximum capacity of the State of Rio Grande do Sul (State Animal Health Official, personal

communication, 2022). We then implemented control actions in the remaining simulations

(99.51%). Simulations with control actions were then classified according to the success of FMD

elimination. Specifically, simulations were categorized as having unsuccessful FMD elimination

if the number of infected farms exceeded 450 or the control action duration exceeded 100 days;

otherwise, the simulations were considered successful. Finally, we tracked the number of days

that control actions were applied.

The software used to develop MHASpread, graphics, and maps was R v. 4.1.1[38] in

RStudio with the following packages: sampler [39], tidyverse[40] (Wickham et al., 2019), sf [41],

brazilmaps[42], doParallel[43], and lubridate[44].

3. Results

3.1 Farm- and animal-level FMD temporal dynamics

Of 15,721 seeded infections, 10,473 simulations started in bovine farms, 5,179 in swine farms,

and 69 in small-ruminant farms; 4,702 farms (29%) had multiple taxa on the same premises.

Examining the epidemic trajectories at 20 days after infection seeding, we estimated a median of

15 (IQR: 5-41) infected animals, of which 12 (IQR: 4-32) were bovines, 5 (IQR: 2-12) were

swine, and 1 (IQR: 1-2) was a small ruminant (Supplementary Table S1 and Figure S7). The

compartment distribution at 10 and 20 days after each animal-level infection was seeded is

shown in Supplementary Table S1.

The FMD distribution at the farm level after 20 days was highly variable; the median

number of infected farms was 8 (IQR: 3-22), of which 7 (IQR: 3-18) were bovine farms, 3 (IQR:

1-6) were swine farms, and 1 (IQR: 1-1) was a small-ruminant farm (Figure 3). We observed

large-scale epidemics with up to 2,197 infected farms; however, these rare events were outside

the 95% credibility interval (CI) (Figure 3). The farm-level infection distribution at 10 and 20

days after infection seeding is available in Supplementary Table S2.

Figure 3. Distribution of the daily number of infected farms. The boxplots with outliers show

the number of infected farms. The colored background area represents the 95% credible interval

for the daily number of infected farms. The y-axis is on a $\log_{10}$ scale.

3.2 Contribution of each transmission route

We evaluated the contributions of the between-farm and spatial-transmission routes. Overall,

during the first 17 days of FMD dissemination, the main transmission route was animal

movements, which led to a daily average of 4.9 (CI: 1-23) newly infected animals; however, at

20 days after FMD dissemination, spatial transmission was by far the dominant route, with a

daily average of 11 (CI: 1-95.6) newly infected animals. We then examined transmission

contributions by taxa. At 10 days, bovine infection shifted from spreading via animal movement

to spreading via spatial transmission. At 20 days, new bovine infections occurred predominantly

by spatial transmission, with an estimated average of 10.7 (CI: 1-57) new infections per day,

while animal movement resulted in an average of 4.5 (CI: 1-32) new infections. For swine,

animal movement was the most critical route throughout the simulation period; at 20 days,

animal movement generated an average of 11.4 (CI: 1-123) newly infected pigs, while spatial

transmission produced an average of 3.1 (CI: 1-90) newly infected pigs. Finally, for small

ruminants, both routes appear to have a similar contribution (Figure 4). The summary statistics of

the contribution of each route at 10 and 20 days are available in Supplementary Table S3.

Figure 4. FMD transmission due to between-farm movement or spatial transmission. The x-

axis represents the number of days after the FMD infection was seeded. The y-axis (represented

on a $\log_{10}$ scale) shows the number of newly infected animals, the solid line represents the

average number of infected individuals, and the colored background represents the 95% credible

interval.

3.3 Spatial transmission dynamics

Of the 15,721 seeded infections, 386,043 simulations generated secondary infections. The

median distance between seeded and secondary infections was 5.77 km (IQR: 1-10.29 km). Our

simulations also captured long-distance spread at 100 km and 500 km; the maximum dispersal

distance was 695.40 km (Figure 5A). As noted in Figure 5B, the probability of infection

decreased by 11% at 20 km compared with one km.

Figure 5. Distribution of the cumulative epidemic distances. A) Frequency histogram of

secondary infections at varying distances from the seeded infection. B) Empirical cumulative

distribution function (1-ECDF) probability of infection according to distances from the initial

outbreak. Both x-axes are a $\log_{10}$ scale.

3.4 Sensitivity analysis

We evaluated the sensitivity of 18 model parameters with weights ranging from -0.17 to 0.24;

this sensitivity indicated a limited influence of model parameters on the number of simulated

secondary infections. Specifically, λ_1 of each host taxa had a negative influence on the number

of secondary infections; in contrast, β for small ruminants to swine, small ruminants to bovines,

and small ruminants to small ruminants as well as λ_2 for small ruminants positively influenced

the number of secondary infections. The complete sensitivity analysis results are depicted in

Supplementary Figure S10.

3.5 Effectiveness of control strategies

On day 20 of silent FMD dissemination, infected zones contained a median of 8,660 (IQR: 308 -

53, max: 783) animals and 1,785 (IQR: 583 - 4,804) farms; in buffer zones, the median was

77,679 (IQR: 2,408 - 384,439) animals, and 2,303 (IQR: 4,357 - 30,900) farms; and within

surveillance zones, the median was 369,541 (IQR: 9,599 - 1,460,202) animals and 50,148 (IQR:

19,295 - 105,943) farms (Figure 6 and Supplementary Figure S8, for the animal-level

distributions). Results at 10 days at the animal level are shown in Supplementary Table S4 and

Supplementary Figure S9; at the farm level, they are shown in Supplementary Figure S11 and

Table S5.

Figure 6. Distribution of farms within control zones at 20 days after the initial infection.

The y-axis shows the number of animals within infected zones, buffer zones, and surveillance

zones on a log$_{10}$ scale. The box plots (left) include outliers and the density distribution (right).

As expected, the number of days necessary to eliminate most epidemics was dependent on the

number of secondary infections at 20 days and the duration of control action implementation

(Figure 7). The baseline control scenario handled 93.4% of the epidemics with a median of 9

(IQR: 3 - 23, max: 221) infected farms; scenario one, which has twice the depopulation and

vaccination rates of the baseline scenario, eliminated 97.8% of the simulated epidemics with a

median of 10 (IQR: 4 - 26, max: 223) infected farms. Scenario two (depopulation only)

eliminated 78.4% of simulated epidemics, stopping mostly small-scale epidemics with a median

of 7 (IQR: 3 - 14, max: 74) infected farms. Scenario three, which involved depopulation and

vaccination rates three times those of the baseline control scenario, could control 98.3% of

epidemics with a median of 10 (IQR: 4 - 26, max: 248) initially infected farms (Figure 7 and

Supplementary Table S6). Regarding epidemics in which control actions were unsuccessful,

scenario two (depopulation only) was the least effective scenario; it was unable to contain

epidemics with a median of 61 (IQR: 43 - 93, max: $\geq$400) initially infected farms. The baseline

control scenario was unsuccessful in eliminating simulated epidemics with a median of 116

(IQR: 82-161, max: $\geq$400) infected farms, while scenarios one and three were unable to eliminate

epidemics with medians of 183 (IQR: 152 - 244, max: $\geq$400) and 203 (IQR: 170 - 264, max:

$\geq$400) initially infected farms, respectively (Figure 7).

Figure 7. Distribution of successfully and unsuccessfully halted FMD epidemics under four control and eradication scenarios of 20 days of silent spread. The box plot (y-axis: $\log_{10}$ scale) shows the distribution of secondarily infected farms before the implementation of control actions. The background scatter plot represents each stochastic simulation, in which the color represents the number of days since control and eradication actions were implemented. The x-axis represents the control action scenarios implemented to halt the epidemic.

Of the successfully halted epidemics shown in the left panel of Figure 7, the baseline control scenario and scenario two eliminated all epidemics within 99 days after implementing control actions (Figure 8). In contrast, scenarios one and three took only 84 days and 54 days, respectively, to eliminate the simulated epidemics (Figure 8).

Figure 8. Average number of days to successfully eliminate FMD epidemics. The vertical

dashed line marks the 20th day of silent FMD spread and the implementation of control actions.

The solid black line represents the average daily number of infected farms. In addition, we

simulated control actions starting 10 days after seeding. The initial infection is described in

greater detail in the Supplementary Material ("Effectiveness of control actions started 10 days

after initial simulated infection").

4. Discussion

We developed a multiscale and multispecies stochastic model that explicitly incorporated

species-specific transmission coefficients. However, even after including interactions among

different taxa, bovines remained the most infected taxa in the simulated epidemics, followed by

swine and small ruminants. Animal movement was the most effective transmission route at the

beginning of the epidemic, after which bovine infection developed predominantly by spatial

transmission, while swine infection developed mainly through between-farm movement for the

first 20 days. Our results also demonstrated that most (74.7%) secondary cases resulted from

spread to farms within 10 km of the initially seeded infection. We also demonstrated that the

baseline control scenario and alternative scenarios were able to eradicate most epidemics within

100 days from the start of the simulated outbreaks.

By incorporating population characteristics and between-farm animal movements in the

State of Rio Grande do Sul, we demonstrated that FMD is likely to disseminate between different

taxa, e.g., bovines to swine and vice versa, regardless of which taxa was infected first

(Supplementary Figure S14). The preferred route of infection was not consistent across taxa, a

phenomenon that has been demonstrated previously[45]. Specifically, until day 17, between-farm

movement was the predominant dissemination route of swine and small ruminants, whereas for

bovines, spatial transmission became the preferred transmission route at day 10. This dominance

of spatial transmission later in the epidemic may be explained, at least to some degree, by

climactic conditions (i.e., cool temperatures and high humidity) in some regions that favor the

spread of FMD[4,30,46], areas with a high density of farms and spatial clustering. More importantly,

given the high connectivity of bovine farms in some municipalities[47], the initial peak in bovine

infection could be explained by animal movement, while the shift toward spatial transmission

could have been facilitated when secondary cases spread into densely populated areas with up to

12.23 farms per km^2 [47]. These densely populated areas coincide with areas in our simulations in

which control actions were not sufficient to eliminate large-scale epidemics (Supplementary

Figure S15). Even though we considered two main transmission routes, data regarding other

important indirect transmission routes, such as fomites and transportation vehicles, were not

considered[1,48]. Importantly, when we evaluated swine infection, animal movements were by far

the most relevant transmission pathway, consistent with other findings [49,50]. Regarding small-

ruminant dynamics, we did not determine any dominant transmission route. Even though small

ruminants did not significantly contribute to FMD dissemination, their contribution was not

negligible. Thus, we suggest prioritizing control interventions by improving animal traceability

early in an FMD epidemic, especially within the first 10 days; if FMD is detected after 10 days

of dissemination, enhancing surveillance activities within control zones should be prioritized.

Assuming that animal health official veterinarians can vaccine 1,000 animals per day

(personal communication, animal health officials), simulations in which FMD was identified at

20 days after the initial infection would require 160 veterinarians to oversee an average of 4,222

farms within infected zones and administer approximately 53,434 vaccine doses (if all taxa are

vaccinated) or 21,640 vaccine doses (without vaccinating swine). If vaccination is also deployed

in buffer zones, 320,223 vaccine doses and 961 veterinarians (all taxa) or 137,046 vaccine doses

and 411 veterinarians (without vaccinating swine) would be needed. Similar modeling work in

Denmark has predicted an average of 22 epidemics that last 34 days would need 123

veterinarians[31]. Importantly, the estimated number of veterinarians and vaccine doses in our

study was similar to those recorded by the UK and Argentina during their past large-scale

outbreaks that included up to 398 secondarily infected farms[51,52]. If such a large-scale epidemic

was to occur in Rio Grande do Sul, there would not be sufficient animal health officials to

implement the various disease control activities nor sufficient vaccine doses. Hence, future work

should focus on estimating not only the necessary number of animal health officials but also

administrative, direct, and indirect costs, including those resulting from large-scale epidemics. In

addition, the revised transmission model should include longer delays in disease detection (e.g.,

30 days or 40 days), the necessary biosecurity downtime between farm visits, and other relevant

logistical roadblocks linked to controlling and eliminating large-scale FMD epidemics [51].

Several modeling studies have demonstrated the effectiveness of combining vaccination

and depopulation to achieve FMD eradication[53–55], with some exceptions in high-density

livestock areas, such as in the Netherlands, where vaccination combined with ring culling was

insufficient[31]. In our study, scenarios that combined vaccination and depopulation were

consistently more effective than depopulation alone (Figures 7 and 8). One potential explanation

is related to the predominance of spatial transmission, since most simulated FMD cases occurred

within a median of seven km from infected farms, which falls within the infected zones. Even

though the use of depopulation alone exhibited the lowest rate of successful FMD elimination,

this strategy was still able to eliminate small-scale epidemics (with maximum of 74 infected

farms). One advantage of eliminating FMD without vaccination is the speed at which the state

could reclaim an FMD-free status[56]. Regarding our alternative control scenarios, scenario three

was the most aggressive and most effective; this scenario assumed that the state would be able to

depopulate 12 farms and vaccinate 15,000 farms per day. Unfortunately, this strategy was still

unable to eliminate large-scale epidemics (with more than 400 infected farms detected after 20

days of silent FMD dissemination). Ultimately, to eliminate all simulated epidemics, we

estimated that 120 farms would need to be depopulated and 25,000 animals would need to be

vaccinated within seven days; these recommendations are extremely unrealistic given the current

staffing and available resources in the State of Rio Grande do Sul, Brazil. Thus, we reached the

same conclusion as the modeling work in Austria: the number of animal health officials needs to

be doubled to eliminate large-scale epidemics[31,53,57]. Even though large-scale FMD epidemics are

expected to challenge veterinary services worldwide, our simulations in Rio Grande do Sul

showed that secondary infections at three to 23 infected farms would be eliminated within 100

days, even using the baseline control scenario. While we simulated 100 days of control action

implementation, most epidemics could be eliminated within 50 days if there were enough

veterinarians to depopulate 12 farms and vaccinate 15,000 animals per day (i.e., scenario three).

It is also important to consider the profile of infected farms, such as location and the size of the

farm's contact network[47], along with the results of these simulations, to reach more informed

decisions. Finally, as described earlier, we assumed full compliance with all associated

surveillance actions and movement restrictions; thus, the results should be interpreted in this

light.

5. Limitations and Further Remarks

We have identified a number of limitations, mostly related to assumptions made in the control

scenarios. We utilized the FMD outbreak in 2000-2001 as well as published data to select

transmission parameters that could impact the final simulations. Our sensitivity analysis of the

population characteristics, latent and infection periods, and β of each taxa did not indicate that

these parameters significantly affected the number of infected animals. Thus, our model has an

acceptable level of robustness. Additionally, other important between-farm transmission routes,

such as transportation vehicles and farm-staff movements, which have been previously

associated with FMD dissemination[58,59], were not included in our model. If such indirect contact

networks are considered, the results would likely change, and model realism would be

improved[53].

We assumed 100% compliance with the restriction of between-farm movement from

infected farms and farms directly linked to infected farms as well as the restriction of movement

into and from control zones; we also assumed that the disposal of depopulated animals

eliminated any possibility of further dissemination. Nevertheless, real-world compliance with the

control actions is not perfect, and future work could consider partial compliance with movement

restriction and delays in the time and effectiveness of depopulation to increase model realism.

Additionally, given the volume of vaccines necessary to vaccinate animals in infected and buffer

zones, further work should consider restricting vaccination to each taxa, thus allowing

benchmarking and better estimation of vaccine stockpiles[54,55,60]. Another limitation of the present

study is that we assumed vaccination provided instantaneous efficacy, which is an obvious model

simplification; in a future version of this model, vaccination waning should be considered.

Notably, FMD virulence, infectivity, and transmission can also vary among strains[1,13].

Even though the most recent outbreaks in the State of Rio Grande do Sul were caused by

serotypes O and A [22], we cannot rule out the possibility that other strains were introduced and

exhibited different dissemination patterns. Future work could include transmission scenarios

with strains circulating in neighboring countries.

To our knowledge, this is the first FMD transmission model developed for Latin

American countries. While previous FMD transmission models have included multiple species in

their transmission dynamics [30,31,55,60], to our knowledge, none have explicitly considered the

heterogeneity of transmission rates among species. Another unique feature of our model is the

estimation of successful and unsuccessful epidemic management strategies while also assessing

the maximum number of outbreaks and the number of days needed to complete FMD

elimination. Thus, we provided rules of thumb for the four main control and eradication

strategies by estimating the number of secondary infections that the simulated scenarios could

eliminate. These results thus offer scientific support to decision-makers.

In addition, rather than selecting a limited number of farms to seed infection, our

modeling approach was designed to generate a fair representation of the entire at-risk population.

While our approach was more computationally expensive, our results provide policy-makers

with an overview of all possible introduction scenarios, which is important because it is nearly

impossible to predict where FMD may resurface. Finally, our model can also provide a

distribution of expected FMD epidemics for any current or future control actions listed in the

Brazilian control and elimination plan. Nevertheless, because our results are based on population

data and between-farm movement data from Rio Grande do Sul, the interpretation of our

findings should not be extrapolated to other Brazilian states. However, since the MHASpread

model infrastructure is highly flexible, if the necessary population data and animal-movement

data is made available, MHASpread can be easily extended to other Brazilian states and,

possibly, other Latin American countries.

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6. Conclusion

After 20 days of silent FMD dissemination, we estimated that the number of secondarily infected

farms ranged from three to 22. Early detection was pivotal to the success of FMD elimination,

and we demonstrated that the deployment of depopulation alone was effective in simulations

with up to 22 infected farms. However, large-scale epidemics required the depopulation of 100

farms and vaccination of 25,000 animals per day to eliminate 100% of simulated outbreaks. We

showed that during the first 17 days of an FMD epidemic, animal movement was the

predominant transmission route among the taxa; however, there were significant differences in

the dissemination of bovine and swine infections. Regardless of which taxon FMD was first

introduced in, the median spread distance fell within the control zones (from one to 10 km). We

also showed that the baseline control scenario, which was based on the Brazilian response plan,

was sufficient to eliminate 93.83% of epidemics within 100 days of deploying the

countermeasures. Our model projections, along with the necessary software, are available to

local animal health officials. Thus, our model can be used as a policy tool not only for future

responses to FMD epidemics through computer-based preparedness drills and capacity building

but also during an emergency response to an FMD epidemic, by providing rules of thumb

generated from simulated control scenarios.

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Authors' contributions

NCC and GM conceived the study. NCC and GM participated in the study design. FPNL

coordinated the data collection. NCC performed data processing and cleaning, designed the model

and simulated scenarios with the assistance of GM. NCC and GM planned the computational

analysis. NCC conducted the formal coding. GM and NCC wrote and edited the manuscript. All

authors discussed the results and critically reviewed the manuscript. GM secured the funding.

Conflict of interest

The authors declare that they have no conflicts of interest.

Ethical statement

The authors confirm that this study conformed to the ethical policies of the journal, as provided on the

journal's author guidelines page. Since this work did not involve animal sampling or the collection of

questionnaire data by researchers, no ethics permits were needed.

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Data availability statement

The datasets analyzed in the current study are not publicly available due to data protection but may be

available from the corresponding author on reasonable request.

References

1. Paton, D. J., Gubbins, S. & King, D. P. Understanding the transmission of foot-and-mouth

disease virus at different scales. *Curr. Opin. Virol.* **28**, 85–91 (2018).

2. Bouma, A. *et al.* The foot-and-mouth disease epidemic in The Netherlands in 2001. *Prev. Vet.*

Med. **57**, 155–166 (2003).

3. Thompson, D. K. *et al.* Economic costs of the foot and mouth disease outbreak in the United

Kingdom in 2001: -EN- -FR- -ES-. *Rev. Sci. Tech. OIE* **21**, 675–687 (2002).

4. Chanchaidechachai, T., de Jong, M. C. M. & Fischer, E. A. J. Spatial model of foot-and-

mouth disease outbreak in an endemic area of Thailand. *Prev. Vet. Med.* **195**, 105468 (2021).

5. Gomez, F., Prieto, J., Galvis, J., Moreno, F. & Vargas, J. Identification of Super-Spreaders of

Foot-and-Mouth Disease in the cattle transportation network: The 2018 outbreak case in Cesar

(Colombia). in *2019 4th World Conference on Complex Systems (WCCS)* 1–6 (IEEE, 2019).

doi:10.1109/ICoCS.2019.8930765.

6. PANAFTOSA-OPS/OMS. Informe de Situación de los Programas de Erradicación de la

Fiebre Aftosa en Sudamérica y Panamá, año 2019. (2019).

7. WOAH. Terrestrial Code Online Access. *WOAH - World Organisation for Animal Health*

https://www.woah.org/en/what-we-do/standards/codes-and-manuals/terrestrial-code-online-

access/.

8. Santos, D. V. dos *et al.* Identification of foot and mouth disease risk areas using a multi-

criteria analysis approach. *PLOS ONE* **12**, e0178464 (2017).

9. Bertram, M. R., Yadav, S., Stenfeldt, C., Delgado, A. & Arzt, J. Extinction Dynamics of the

Foot-and-Mouth Disease Virus Carrier State Under Natural Conditions. *Front. Vet. Sci.* **7**, 276

(2020).

10.   Davies, G. The foot and mouth disease (FMD) epidemic in the United Kingdom 2001.

Comp. Immunol. Microbiol. Infect. Dis. **25**, 331–343 (2002).

11.   Seymour, R. G., Kypraios, T. & O'Neill, P. D. Bayesian nonparametric inference for

heterogeneously mixing infectious disease models. *Proc. Natl. Acad. Sci.* **119**, e2118425119

(2022).

12.   Alexandersen, S., Zhang, Z., Donaldson, A. I. & Garland, A. J. M. The Pathogenesis and

Diagnosis of Foot-and-Mouth Disease. *J. Comp. Pathol.* **129**, 1–36 (2003).

13.   Arzt, J. *et al.* Quantitative impacts of incubation phase transmission of foot-and-mouth

disease virus. *Sci. Rep.* **9**, 2707 (2019).

14.   Moreno-Torres, K. I. *et al.* Parameterization of the durations of phases of foot-and-mouth

disease in pigs. *Prev. Vet. Med.* **202**, 105615 (2022).

15.   Stenfeldt, C., Diaz-San Segundo, F., de los Santos, T., Rodriguez, L. L. & Arzt, J. The

Pathogenesis of Foot-and-Mouth Disease in Pigs. *Front. Vet. Sci.* **3**, (2016).

16.   van Andel, M., Tildesley, M. J. & Gates, M. C. Challenges and opportunities for using

national animal datasets to support foot-and-mouth disease control. *Transbound. Emerg. Dis.*

68, 1800–1813 (2021).

17.   Sseguya, I., Mugisha, J. Y. T. & Nannyonga, B. Outbreak and control of Foot and Mouth

Disease within and across adjacent districts—A mathematical perspective. *Results Control*

Optim. **6**, 100074 (2022).

18.   Beck-Johnson, L. M. *et al.* An exploration of within-herd dynamics of a transboundary

livestock disease: A foot and mouth disease case study. *Epidemics* **42**, 100668 (2023).

19.   MAPA. Febre Aftosa. *Ministério da Agricultura, Pecuária e Abastecimento*

https://www.gov.br/agricultura/pt-br/assuntos/sanidade-animal-e-vegetal/saude-

animal/programas-de-saude-animal/febre-aftosa/febre-aftosa (2020).

20.   SEAPI. *Secretaria da Agricultura Pecuaria e Irrigação.* (2021).

21.   Garira, W. A primer on multiscale modelling of infectious disease systems. *Infect. Dis.*

Model. **3**, 176–191 (2018).

22.   da Costa, J. M. N., Cobellini, L. G., Cardenas, N. C., Groff, F. H. S. & Machado, G.

Assessing epidemiological parameters and dissemination characteristics of the 2000 and 2001

foot-and-mouth disease outbreaks in Rio Grande do Sul, Brazil. *bioRxiv* 2022.05.22.492961

(2022).

23.   Cabezas, A. H., Sanderson, M. W. & Volkova, V. V. Modeling Intervention Scenarios

During Potential Foot-and-Mouth Disease Outbreaks Within U.S. Beef Feedlots. *Front. Vet.*

Sci. **8**, (2021).

24.   Chis Ster, I., Dodd, P. J. & Ferguson, N. M. Within-farm transmission dynamics of foot

and mouth disease as revealed by the 2001 epidemic in Great Britain. *Epidemics* **4**, 158–169

(2012).

25.   van Roermund, H. J. W., Eblé, P. L., de Jong, M. C. M. & Dekker, A. Mckay. *Vaccine*

28, 4452–4461 (2010).

26.   Kinsley, A. C., Patterson, G., VanderWaal, K. L., Craft, M. E. & Perez, A. M. Parameter

Values for Epidemiological Models of Foot-and-Mouth Disease in Swine. *Front. Vet. Sci.* **3**,

44 (2016).

27.   Bravo de Rueda, C., de Jong, M. C., Eblé, P. L. & Dekker, A. Quantification of

transmission of foot-and-mouth disease virus caused by an environment contaminated with

secretions and excretions from infected calves. *Vet. Res.* **46**, 43 (2015).

28.   Goris, N. E., Eblé, P. L., Jong, M. C. M. de & Clercq, K. D. Quantifying foot-and-mouth

disease virus transmission rates using published data. *ALTEX - Altern. Anim. Exp.* **26**, 52–54

(2009).

29.   Mardones, F., Perez, A., Sanchez, J., Alkhamis, M. & Carpenter, T. Parameterization of

the duration of infection stages of serotype O foot-and-mouth disease virus: an analytical

review and meta-analysis with application to simulation models. *Vet. Res.* **41**, 45 (2010).

30.   Björnham, O., Sigg, R. & Burman, J. Multilevel model for airborne transmission of foot-

and-mouth disease applied to Swedish livestock. *PLOS ONE* **15**, e0232489 (2020).

31.   Boklund, A., Halasa, T., Christiansen, L. E. & Enøe, C. Comparing control strategies

against foot-and-mouth disease: Will vaccination be cost-effective in Denmark? *Prev. Vet.*

Med. **111**, 206–219 (2013).

32.   Ziegel, E. R. & Lohr, S. L. Sampling: Design and Analysis. *Technometrics* **42**, 223

(2000).

33.   Mckay, M. D., Beckman, R. J. & Conover, W. J. A Comparison of Three Methods for

Selecting Values of Input Variables in the Analysis of Output From a Computer Code.

Technometrics **42**, 55–61 (2000).

34.   Marino, S., Hogue, I. B., Ray, C. J. & Kirschner, D. E. A methodology for performing

global uncertainty and sensitivity analysis in systems biology. *J. Theor. Biol.* **254**, 178–196

(2008).

35.   *Infectious Diseases and Our Planet*. vol. 7 (Springer International Publishing, 2021).

36.   MAPA. Plano Estratégico do PNEFA 2017-2026 — Português (Brasil). *Plano*

Estratégico do PNEFA https://www.gov.br/agricultura/pt-br/assuntos/sanidade-animal-e-

vegetal/saude-animal/programas-de-saude-animal/febre-aftosa/plano-estrategico-pnefa-2017-

2026 (2022).

37.   Lecoutre, B. The Bayesian Approach to Experimental Data Analysis. in *Essential*

Statistical Methods for Medical Statistics 308–344 (Elsevier, 2011). doi:10.1016/B978-0-444-

53737-9.50014-1.

38.   R Core Team. *R: A Language and Environment for Statistical Computing*. (R Foundation

for Statistical Computing, 2021).

39.   Baldassaro, M. sampler: Sample Design, Drawing & Data Analysis Using Data Frames.

(2019).

40.   Wickham, H. *et al.* Welcome to the Tidyverse. *J. Open Source Softw.* **4**, 1686 (2019).

41.   Pebesma, E. {Simple Features for R: Standardized Support for Spatial Vector Data}. *R J.*

10, 439--446 (2018).

42.   Prado Siqueira, R. brazilmaps: Brazilian Maps from Different Geographic Levels. (2021).

43.   Corporation, M. & Weston, S. doParallel: Foreach Parallel Adaptor for the 'parallel'

Package. (2022).

44.   Grolemund, G. & Wickham, H. Dates and Times Made Easy with {lubridate}. *J. Stat.*

Softw. **40**, 1--25 (2011).

45. Probert, W. J. M. *et al.* Real-time decision-making during emergency disease outbreaks.

PLOS Comput. Biol. **14**, e1006202 (2018).

46. Green, D. M., Kiss, I. Z. & Kao, R. R. Modelling the initial spread of foot-and-mouth

disease through animal movements. *Proc. R. Soc. B Biol. Sci.* **273**, 2729–2735 (2006).

47. Cardenas, N. C., Sykes, A. L., Lopes, F. P. N. & Machado, G. Multiple species animal

movements: network properties, disease dynamics and the impact of targeted control actions.

Vet. Res. **53**, 14 (2022).

48. Rossi, G., Smith, R. L., Pongolini, S. & Bolzoni, L. Modelling farm-to-farm disease

transmission through personnel movements: from visits to contacts, and back. *Sci. Rep.* **7**,

2375 (2017).

49. Machado, G. *et al.* Quantifying the dynamics of pig movements improves targeted

disease surveillance and control plans. *Transbound. Emerg. Dis.* **68**, 1663–1675 (2021).

50. Shimizu, Y. *et al.* Epidemiology of Classical Swine Fever in Japan—A Descriptive

Analysis of the Outbreaks in 2018–2019. *Front. Vet. Sci.* **7**, 573480 (2020).

51. Anderson, I. Foot and mouth disease: Lessons to be learned inquiry report HC888. *Lond.*

Station. Off. (2002).

52. Perez, A. M., Ward, M. P. & Carpenter, T. E. Epidemiological investigations of the 2001

foot-and-mouth disease outbreak in Argentina. *Vet. Rec.* **154**, 777–782 (2004).

53. Sanson, R. *et al.* Evaluating the benefits of vaccination when used in combination with

stamping-out measures against hypothetical introductions of foot-and-mouth disease into New

Zealand: a simulation study. *N. Z. Vet. J.* **65**, 124–133 (2017).

54. Capon, T. R. *et al.* A Simulation Study of the Use of Vaccination to Control Foot-and-

Mouth Disease Outbreaks Across Australia. *Front. Vet. Sci.* **8**, 648003 (2021).

55.   Dürr, S. *et al.* Evaluation of the benefit of emergency vaccination in a foot-and-mouth

disease free country with low livestock density. *Prev. Vet. Med.* **113**, 34–46 (2014).

56.   Geale, D. W., Barnett, P. V., Clarke, G. W., Davis, J. & Kasari, T. R. A Review of OIE

Country Status Recovery Using Vaccinate-to-Live Versus Vaccinate-to-Die Foot-and-Mouth

Disease Response Policies II: Waiting Periods After Emergency Vaccination in FMD Free

Countries. *Transbound. Emerg. Dis.* **62**, 388–406 (2015).

57.   Marschik, T. *et al.* What Are the Human Resources Required to Control a Foot-and-

Mouth Disease Outbreak in Austria? *Front. Vet. Sci.* **8**, 727209 (2021).

58.   Muroga, N. *et al.* Risk factors for the transmission of foot-and-mouth disease during the

2010 outbreak in Japan: a case–control study. *BMC Vet. Res.* **9**, 150 (2013).

59.   Yang, Q. *et al.* Impact of truck contamination and information sharing on foot-and-mouth

disease spreading in beef cattle production systems. *PLOS ONE* **15**, e0240819 (2020).

60.   Roche, S. E. *et al.* Evaluating vaccination strategies to control foot-and-mouth disease: a

model comparison study. *Epidemiol. Infect.* **143**, 1256–1275 (2015).

Supplementary Material

Modeling foot-and-mouth disease dissemination in Brazil and evaluating the effectiveness of control measures

Nicolas C. Cardenas[1], Francisco P. N. Lopes[2], Gustavo Machado[1,*]

Running title: Foot-and-mouth disease dissemination across species.

[1] Department of Population Health and Pathobiology, College of Veterinary Medicine, North Carolina State University, Raleigh, NC, USA

[2] Departamento de Defesa Agropecuária, Secretaria da Agricultura, Pecuária e Desenvolvimento Rural, Porto Alegre, Brazil

Supplementary Figure S1. The 2018 distribution of farm-level population for each species in the state of Rio Grande do Sul, Brazil. The y-axis is the number of farms and the x-axis is the number of animals, both on a log10 scale.

Supplementary Figure S2. The monthly distribution of animal births and deaths by species,

from January 1st, 2018 to December 31st, 2020.

Supplementary Figure S3. The weekly between-farm animal movements, from January 1st,

2018 to December 31st, 2020.

Supplementary Figure S4. Representation of the different population interactions at the within-farm level. For this example, we considered farms with all three species. In A and B, the dashed line represents the interaction within and between species, and the color denotes a β for each host-to-host interaction (Table 2). The solid line between panel A and B represents the interaction among the farms with different infection probabilities modulated by the spatial proximity (spatial transmission). PE_{ij} denotes the transmission probability given a distance between farm i to farm j.

Supplementary Figure S5. A) Representation of grid-hexagons of size 0.76 km (503,516 m^2).

Each hexagon can host one or more farm locations. The color of the hexagon represents the

probability of PE exposure assuming ten infected animals in the center (black hexagon). B) PE

distance in km.

Supplementary Figure S6. **Control zones mapping.** The infected control zones of 3 km are represented as red dots, the 7 km buffer zone is represented as yellow dots, and the 15 km surveillance zone is represented as green dots. Of note, control zones with overlapping are merged.

Supplementary Figure S7. Distribution of animals per compartment, exposed (E), infected (I), and recovered (R) by species. Because the number of susceptible animals is very large (19,267,676), the S compartment is not shown. The solid line represents the mean and the dots are outliers. The colored background area represents the 95% credible interval. The y-axis is on a log10 scale.

Supplementary Figure S8. The distribution of animals within control zones at 20 days from the initial infection. The y-axis shows the number of animals within infected zones, buffer zones, and surveillance on a log10 scale. The left side boxplots include outliers and the right size density distribution.

Supplementary Figure S9. The distribution of animals within control zones at 10 days from the initial infection. The y-axis shows the number of animals within infected zones, buffer zones, and surveillance on a log10 scale. The left side boxplots include outliers and the right size density distribution.

Supplementary Figure S10. Sensitivity of the number of infectious animals to changes over the

parameters using LHS-PRCC.

Supplementary Figure S11. The distribution of farms within control zones at 10 days from the initial infection. The y-axis shows the number of animals within infected zones, buffer zones, and surveillance on a log10 scale. The left side boxplots include outliers and the right size density distribution.

Effectiveness of control actions started 10 days after initial simulated infection

At 10 days of FMD dissemination, the baseline control scenario fade out 99.90% of epidemics with a median of 2 (IQR: 1-4, max: 162) infected farms, while scenario one eliminated 99.96% of simulated epidemics with a median of 2 (IQR: 1-4, max: 162). Scenario two (depopulation only) eliminated 99.5% of simulated epidemics with a median of 2 (IQR: 1-5, max: 45) infected farms. Scenario three which has three times the depopulation and vaccination of the baseline control scenario was enough to fade out 99.97% of epidemics with a median of 2 (IQR: 1-4, max: 164) initially infected farms (Supplementary Material Figure S12). Similar to the control actions simulations at 20 days scenario two (depopulation only) failed more often and was not able to control epidemics with a median of 30 (IQR: 19-41, max: 248) initially infected farms. In addition, for simulation that was not able to eliminate simulated epidemics, the baseline control scenario failed in eliminating simulations with a median of 69 (IQR: 40-157, max: 258) infected farms, while scenarios one and three, epidemics with a median of 164 (IQR: 164-202, max: 258) and median of 183 (IQR: 162-216, max: 258), respectively (Supplementary Material Figure S12).

Supplementary Figure S12. The distribution of FMD epidemics successfully and unsuccessfully halted under four control and eradication scenarios after 10 days from the initial seeded infection. The boxplot shows the distribution of secondarily infected farms at 10 days, the background scattered plot shows individual stochastic simulations in which color represents the number of days control and eradication measures were applied.

For the baseline control scenario from the initial deployment of control actions, the maximum average number of infected farms was 51.5, for scenario one 139 farms, 15 farms in scenario two, and 189 in scenario three. Of the successfully halted epidemic, the baseline control scenario took 63 days to fade out epidemics. In comparison with the baseline scenario, scenario two took 53, scenario one 45, and scenario three 28 days (Supplementary Material Figure S13).

Supplementary Figure S13. The average number of days to stamp out FMD epidemics (of simulation that successfully eliminated outbreaks). The vertical dashed line marks the 10 days of

silent spread and the start of control actions. The solid black line represents the average of

infected farms per day. The box plot shows the distribution of secondarily infected farms.

Supplementary Figure S14. Distribution of animals per compartment, exposed (E), infected

(I), and recovered (R) by species and by initial farm type where the simulation started. The

right y-axis represents the type of farm where the simulation model started. Because the number

of susceptible animals is very large (19,267,676), the S compartment is not shown. The solid line

represents the mean, and the dots are outliers. The colored background area represents the 95%

credible interval. The y-axis is on a log10 scale.

Supplementary Figure S15. Farms density of the Rio Grande do Sul state when considering

farms that were not successfully controlled, overall control action simulations across all

scenarios. The color background represents the farm density value when considering 0.05 units

of the latitude and longitude projection. The solid black line represents the municipalities'

boundaries of the state of Rio Grande do Sul.

Supplementary Table S1. Estimated animal-level distribution of at 10 and 20 days after initial infection.

Category	State	Quantile 0.25	Mean	Median	Quantile 0.75	Maximum	Days after initial infection
All species	Susceptible	19,459,735	19,495,401	19,490,270	19,525,934	19,717,989	10
All species	Exposed	2	10	5	10	1,783	10
All species	Infected	1	6	3	6	2,735	10
All species	Recovered	2	5	3	5	2,741	10
Bovine	Susceptible	14,330,643	14,330,468	14,330,608	14,330,506	14,306,163	10
Bovine	Exposed	2	8	4	9	983	10
Bovine	Infected	1	4	2	5	518	10
Bovine	Recovered	2	4	3	5	1,129	10
Swine	Susceptible	4,886,604	4,886,448	4,886,600	4,886,591	4,824,419	10
Swine	Exposed	1	5	2	3	1,318	10

Swine	Infected	1	7	2	3	2,586	10
Swine	Recovered	1	7	2	3	2,579	10
Small ruminants	Susceptible	50,408	50,390	50,403	50,391	49,761	10
Small ruminants	Exposed	1	1	1	2	51	10
Small ruminants	Infected	1	1	1	2	25	10
Small ruminants	Recovered	1	1	1	2	10	10
All species	Susceptible	19,675,945	19,719,056	19,708,463	19,751,901	19,958,581	20
All species	Exposed	8	74	25	72	7,215	20
All species	Infected	5	49	15	41	9,765	20
All species	Recovered	8	39	15	34	12,250	20
Bovine	Susceptible	14,330,578	14,329,352	14,330,292	14,329,318	14,218,361	20
Bovine	Exposed	7	56	21	61	4,149	20
Bovine	Infected	4	29	12	32	2,636	20

Bovine	Recovered	7	27	14	30	2,768	20
Swine	Susceptible	4,886,599	4,885,767	4,886,577	4,886,129	4,740,943	20
Swine	Exposed	2	23	5	14	4,098	20
Swine	Infected	2	28	5	12	8,082	20
Swine	Recovered	2	22	4	7	10,234	20
Small ruminants	Susceptible	50,407	50,384	50,401	50,384	49,669	20
Small ruminants	Exposed	1	2	1	2	154	20
Small ruminants	Infected	1	2	1	2	166	20
Small ruminants	Recovered	1	2	1	2	132	20

Supplementary Table S2. Estimated number of infected farms at 10 and 20 days after initial infection.

Category	Quantile 0.25	Mean	Median	Quantile 0.75	Maximum	Days after initial infection
All species	1	3.37	2	4	470	10
Bovine	1	3.02	2	4	389	10
Swine	1	1.57	1	2	90	10
Small ruminants	1	1.02	1	1	4	10
All species	3	20.75	8	22	2,197	20
Bovine	3	17.62	7	18	1,862	20
Swine	1	5.45	3	6	450	20
Small ruminants	1	1.18	1	1	11	20

Supplementary Table S3. Summary statistics for animal movement and spatial transmission.

We show the distribution of newly infected animals after 10 and 20 days of FMD dissemination

Route of transmission	Species	Quantile 0.25	Mean	Median	Quantile 0.75	Maxi mum	Days after initial infection
Animal movement	Bovine	1	2.4	1	16	188	10
Animal movement	Swine	1	7.5	1	58.975	1120	10
Animal movement	Small ruminants	1	1.1	1	2	5	10
Animal movement	All species	1	4.9	1	23	1120	10
Spatial transmission	Bovine	1	2.5	1	9	206	10
Spatial transmission	Swine	1	1.4	1	4	38	10
Spatial transmission	Small ruminants	1	1.0	1	1	2	10
Spatial transmission	All species	1	2.7	2	10	245	10
Animal movement	Bovine	1	4.5	1	32	467	20
Animal movement	Swine	1	11.4	1	123	1617	20
Animal movement	Small ruminants	1	1.4	1	3	105	20
Animal movement	All species	1	11.0	1	95.6	1639	20
Spatial transmission	Bovine	1	10.7	5	57	529	20
Spatial transmission	Swine	1	3.1	2	13	90	20

Spatial transmission	Small ruminants	1	1.1	1	2	4	20
Spatial transmission	All species	1	12.1	5	65	617	20

Supplementary Table S4. Statistics summary of the number of animals in each control zones after 10 and 20 days of FMD dissemination.

Zone	Species	Quantile 0.25	Mean	Median	Quantile 0.75	Maximum	Days after initial infection
Infected	All host	47	8,748.78	1,833	10,078	1,081,512	10
Infected	Bovine	3,560	13,997.84	7,935	16,649	1,031,244	10
Infected	Swine	496	12,184.92	4,028	15,336	1,081,512	10
Infected	Small ruminants	2	63.56	18	62	4,529	10
Buffer	All host	435	66,309.93	20,151	85,476	4,100,452	10
Buffer	Bovine	30,453	103,109.40	62,935	125,653	4,100,452	10
Buffer	Swine	8,047.25	95,344.95	47,321.5	128,585	2,839,570	10
Buffer	Small ruminants	76	475.44	217	541	23,352	10
Surveillance	All host	2,112	278,247.19	117,367	393,387	6,090,216	10
Surveillance	Bovine	152,906	449,516.51	299,849	573,640	6,090,216	10
Surveillance	Swine	6,4614.25	383,209.77	252,319	548,438	3,019,033	10
Surveillance	Small ruminants	452	2,015.28	1,098	2,466	27,256	10
Infected	All host	308	53,434.84	8,660	53,783	3,595,388	20

Infected	Bovine	11,169	84,844.08	35,665	97,155	3,595,388	20
Infected	Swine	6,007	7,5074.99	28,463	84,868	2,853,501	20
Infected	Small ruminants	27	385.46	129	450	17,036	20
Buffer	All Species	2,408	320,223.14	77,679	384,439	6,504,836	20
Buffer	Bovine	87,028	501,054.35	250,295	631,415	6,504,836	20
Buffer	Swine	66,341	457,269.51	230,509	611,856	3,162,750	20
Buffer	Small ruminants	297	2,345.56	1,059	3,046	28,802	20
Surveillance	All host	9,599	885,913.31	369,541	1,460,202	6,546,421	20
Surveillance	Bovine	406,570	1,554,385.98	1,065,027	2,337,789	6,546,421	20
Surveillance	Swine	332,548	1,096,357.88	936,872	1,835,041	3,190,801	20
Surveillance	Small ruminants	1,475	6,996.05	4,581	10,959	30,615	20

Supplementary Table S5. Statistics summary of the number of farms in each control zones after 10 and 20 days of spread disease simulation

Area	Specie	Quantile 0.25	Mean	Median	Quantile 0.75	Maximum	Days after initial infection
Infected	Total	529	4,163	1,736	4,632	102,232	10
Infected	Bovine	395	3,129	1,305	3,481	77,273	10
Infected	Swine	152	1,265	507	1,387	31,567	10
Infected	Small ruminants	3	26	11	30	608	10
Buffer	Total	4,058	23,950	12,032	29,995	202,441	10
Buffer	Bovine	3,022	17,869	8,988	22,397	149,288	10
Buffer	Swine	1,206	7,443	3,711	9,462	56,669	10
Buffer	Small ruminants	26	157	79	199	1,431	10
Surveillance	Total	18,034	65,564	49,450	104,070	200,581	10
Surveillance	Bovine	13,334	47,883	36,420	76,477	144,105	10

Surveillance	Swine	5,499	19,232	15,042	31,569	54,601	10
Surveillance	Small ruminants	120	445	332	710	1,447	10
Infected	Total	583	4,222	1,785	4,804	164,089	20
Infected	Bovine	431	3,176	1,333	3,610	123,200	20
Infected	Swine	165	1,282	521	1,454	50,188	20
Infected	Small ruminants	3	27	11	31	1,114	20
Buffer	Total	4,357	24,247	12,303	30,900	216,163	20
Buffer	Bovine	3,240	18,082	9,177	23,106	156,541	20
Buffer	Swine	1,299	7,517	3,791	9,707	59,156	20
Buffer	Small ruminants	28	159	82	204	1,548	20
Surveillance	Total	19,295	66,457	50,148	105,943	203,462	20
Surveillance	Bovine	14,187	48,514	36,858	77,700	147,013	20

| Surveillance | Swine | 5,817 | 19,456 | 15,346 | 31,937 | 55,281 | 20 |
| Surveillance | Small ruminants | 128 | 453 | 340 | 720 | 1,498 | 20 |

Supplementary Table S6. Results for the predicted number of epidemics fade out for each control and eradication strategy after 20 days of initial simulated infection.

Scenario	FMD fade out	Number of simulations (%)
Baseline control scenario: Vaccination of 5,000 animals/day + depopulation of 4 farms/day	Successfully control	104,598 (93.39%)
Baseline control scenario: Vaccination of 5,000 animals/day + depopulation of 4 farms/day	Unsuccessfully control	7,407 (6.61%)
Scenario one: Vaccination of 10,000 animals/day) + depopulation of 8 farms/day	Successfully control	109,593 (97.85%)
Scenario one: Vaccination of 10,000 animals/day) + depopulation of 8 farms/day	Unsuccessfully control	2,412 (2.15%)
Scenario two: Depopulation of 8 farms/day	Successfully control	87,759 (78.35%)

Scenario two: Depopulation of 8 farms/day	Unsuccessfully control	24,246 (21.65%)
Scenario three: Vaccination of 15,000 animals/day + depopulation of 12 farms/day)	Successfully control	110,115 (98.31%)
Scenario three: Vaccination of 15,000 animals/day + depopulation of 12 farms/day))	Unsuccessfully control	1,890 (1.69%)

Supplementary Equation 1.

The following simple formula was used for calculating the adequate sample size in prevalence study

$$n = \frac{Z^2 P(1 - P)}{d^2}$$

Where n is the sample size, z is the statistic corresponding to level of 95% confidence, P is expected prevalence is this case 0.5 or 50%.