

1 **First record of monogenean fish parasites in the Upper Lufira basin**
2 **(Democratic Republic of Congo): dactylogyrids and gyrodactylids infecting**
3 ***Oreochromis mweruensis*, *Coptodon rendalli* and *Serranochromis***
4 ***macrocephalus* (Teleostei: Cichlidae)**

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45 **Abstract**

46 **Background:** Monogenean parasites have never been formally reported on fish from the Lufira
47 basin. Then it is hypothesised that multiple monogenean species are to be recorded that are new
48 to the region. This study aimed to record the gill monogenean parasite fauna of three cichlid fish
49 species in the Upper Lufira basin by inventorying their diversity (species composition) and
50 analysing their infection parameters (prevalence, mean intensity and abundance).

51 **Methods:** *Oreochromis mweruensis*, *Coptodon rendalli*, and *Serranochromis macrocephalus*
52 were selected for the study, given their economic value and their abundance in the Upper Lufira
53 basin. Monogeneans were isolated from the gills and stomach, mounted on glass slides with
54 either Hoyer's medium or ammonium picrate-glycerin for further identification under a
55 stereomicroscope, based on morphological analysis of genital and haptor hard parts. Indices of
56 diversity and infections parameters were calculated.

57 **Results:** A total of thirteen gill monogenean parasite species (*Cichlidogyrus dossoui*, *C. halli*, *C.*
58 *karibae*, *C. mbirizei*, *C. papernastrema*, *C. quaestio*, *C. sclerosus*, *C. tiberianus*, *C. tilapiae*, *C.*
59 *zambezensis*, *Scutogyrus gravivaginus*, *S. cf. bailloni* and *Gyrodactylus nyanzae*) and one
60 stomach monogenean (*Enterogyrus malmbergi*) were reported. A species richness of S= 10 for

61 *O. mweruensis*, S= 6 for *C. rendalli* and S= 2 for *S. macrocephalus* were recorded. Five parasite
62 species were reported to be common amongst *O. mweruensis* and *C. rendalli*. The most prevalent
63 parasite species were *C. halli* (P= 80.9%) on *O. mweruensis*, *C. dossoui* (P= 92.9%) on *C.*
64 *rendalli* and *C. karibae* and *C. zambezensis* (both of which P = 9.1%) on *S. macrocephalus* with
65 a respective mean infection intensity of 7.9 on *O. mweruensis*, 9.8 on *C. rendalli* and 5 and 15,
66 respectively, on *S. macrocephalus*. Results of this study reported new host ranges for five
67 parasites species (*C. quaestio*, *S. cf. bailloni*, *E. malmbergi* on *O. mweruensis*, *C. halli* on *C.*
68 *rendalli* and *C. karibae* on *S. macrocephalus*) as well as new geographical records for three of
69 them (*S. cf. bailloni*, *E. malmbergi*, *C. karibae*).

70 **Conclusions:** This study highlights the richness of monogenean communities in the Upper Lufira
71 basin and is a starting point for future helminthological studies, e.g. on the use of fish parasites as
72 indicators of anthropogenic impacts.

73 **Keywords:** Lake Tshangalele, Haut-Katanga, *Cichlidogyrus*, *Enterogyrus*, *Gyrodactylus*,
74 *Scutogyrus*

75 **Background**

76 Across the African continent, the Congo basin harbours the greatest species richness of fish [1-
77 2]. The Congo basin covers 3,747 320 km², and drains most of the Democratic Republic of
78 Congo and parts of some of its bordering countries (Angola, Zambia, Tanzania, Burundi,
79 Rwanda, Central African Republic and Republic of Congo) and a small part of Cameroon [3].
80 The Congo basin includes different types of habitats and is subdivided into sections: Upper
81 Congo (called Lualaba), Middle Congo, and Lower Congo [2,4-5]. One of the major tributaries
82 in the Upper Congo drainage is the Lufira River [6]. The Lufira River is subdivided into three

83 sections: the Upper Lufira (from the source of the river to Lake Koni), the Middle Lufira (from
84 downstream Lake Koni to the Kyubo Falls), and the Lower Lufira (from downstream the Kyubo
85 Falls to the Kamalondo Depression, at the junction with the Lualaba River) [5,7]. In order to
86 provide hydroelectric power, two successive dams were built in the Upper Lufira River; this
87 created two artificial Lakes, Tshangalele (1930) and Koni (1949) [8-10]. Lake Tshangalele,
88 located about 35 km east of the town of Likasi, holds a variety of fish, and it is also an UNESCO
89 Man and the Biosphere Reserve, rich in birdlife [11-12]. In the Lufira River, most studies
90 undertaken on biodiversity focused on vertebrates such as fish and birds [13-16]. Vast and
91 speciose communities, which are often dominated by less sizeable animals such as flatworms or
92 various parasite taxa, remain understudied, as is the case all over the world [17-18]. In view of
93 the high biodiversity of potential host species in the tropics, it can be expected that
94 parasitological surveys there would lead to the recording of many parasite species, including
95 species new to science [19-20]. This study focuses on monogenean fish parasites due to their
96 diversity, wide distribution, high host-specificity and single-host lifecycle, rendering them
97 interesting models for studying the extent of parasite biodiversity and the underlying
98 diversification mechanisms [21]. Monogeneans are common parasitic flatworms
99 (Platyhelminthes) mostly infecting fish, and sporadically aquatic invertebrates, amphibians,
100 reptiles and a single species of mammal (the hippopotamus) [22-27]. Infection sites of
101 monogeneans on fish are typically gills, fins and/or skin [28], however they are also found rarely
102 in the stomach, urinary bladder, intestine, oral or nasal cavity, eyes and heart [29-30]. Because of
103 their one-host lifecycle and their close relationship with their host species, many monogeneans
104 are specialists, infesting only a single host species (oioxenous specificity), though others are
105 generalists, infesting two or more host species (stenoxenous specificity) [31-33]. Mendlová and

106 Šimková [34] used a more extensive number of categories of host specificity on the basis of the
107 phylogenetic relationships among (cichlid) host species. Parasites can be: (1) strict specialists
108 when infecting only one host species; (2) intermediate specialists when infecting two or more
109 congeneric host species; (3) intermediate generalists when infecting noncongeneric cichlid
110 species belonging to the same tribe; and finally (4) generalists, when infecting noncongeneric
111 cichlid species of at least two different tribes. African cichlids (taking also into account the
112 Levant) are known to harbour monogenean parasites belonging to six genera: *Enterogyrus*
113 Paperna, 1963; *Urogyrus* Bilong Bilong, Birgi & Euzet, 1994; *Onchobdella* Paperna, 1968;
114 *Scutogyrus* Pariselle & Euzet, 1995; *Cichlidogyrus* Paperna, 1960 (Dactylogyridea) and
115 *Gyrodactylus* von Nordmann, 1832 (Gyrodactylidea). The latter four are ectoparasitic genera,
116 and among them, *Cichlidogyrus* is the most species-rich group with more than 138 nominal
117 species described to date [35-37]. This study aims to record the monogenean parasite fauna of
118 three cichlid fishes in the Upper Lufira basin; these parasites were never formally reported from
119 this region. Objectives include: (i) inventorying the diversity of gill monogenean communities,
120 and (ii) analyzing infection parameters of these monogenean parasites.

121 **Methods**

122 **Study area**

123 This study was conducted in the Upper Lufira basin (Figure 1), which is localized across
124 the mining hinterland area in the west of the Haut-Katanga province (in the south of the
125 former Katanga province). The climate is of type AW6 following the classification of
126 Köppen [38], a rainy tropical climate with a rainy season extending from November to
127 April [39]. Most precipitation falls from December to March [40]. Fishing is done

128 essentially for *Coptodon rendalli* (Boulenger, 1896), *Oreochromis mweruensis*
129 Trewavas, 1983, *Serranochromis macrocephalus* Boulenger, 1899, *Clarias gariepinus*
130 (Burchell, 1822) and *Clarias ngamensis* (Castelnau, 1861) [12, 41]. Captured fish are
131 intended for human consumption, for a small part by the local population around the
132 Upper Lufira basin, and for most part in bigger towns such as Likasi and Lubumbashi.

133 **Fish sampling**

134 Three fish species, *Oreochromis mweruensis*, *Coptodon rendalli* and *Serranochromis*
135 *macrocephalus* were selected for the study, given their economic value and their abundance in
136 the Upper Lufira basin [12, 41]. Fish were collected using nets or were bought from fishermen
137 along the shores of the Lufira River, Lake Tshangalele and Lake Koni (Figure 1) between
138 September 2015 and August 2018. Fish were kept alive in an aerated tank, and transported to a
139 field laboratory. Fish were identified up to the species level following the keys by Skelton [42]
140 and Lamboj [43]. Fish were killed by severing the spinal cord just posterior to the cranium,
141 immediately prior to examination, following Olivier *et al.* [44]. Fish were processed as the total
142 length (TL) and the standard length (SL) were measured to the nearest centimetre, and the weight
143 was taken in gram for each fish.

144 **Parasite sampling**

145 To collect monogenean parasites, fish were dissected and the right gill arches removed by dorso-
146 ventral section. One fish amongst all the fishes sampled was randomly dissected and inspected
147 for monogenean parasites in its stomach. Gill arches and the stomach were placed in a Petri-dish
148 containing water for examination using a stereomicroscope Optica 4.0.0. Parasites were
149 dislodged from the gill filaments using entomological needles and fixed between a slide and

150 cover slip into a drop of either Hoyer's medium or ammonium picrate-glycerin (a preparation
151 described by Malmberg, 1957) according to Nack *et al.* [45]. Twenty-four hours later, coverslips
152 were sealed using nail varnish. Parasites were deposited in the invertebrate collection of the
153 Royal Museum of Central Africa (RMCA) under accession numbers XXX.

154 **Monogenean community composition, indices of diversity and infection parameters**

155 Morphological identifications of the retrieved parasite specimens were conducted based on the
156 sclerotized parts of the haptor, the male copulatory organ (MCO) and the vagina, using an Optica
157 BA310 and a phase-contrast Olympus BX50 microscope. Parasite identification up to species
158 level, and comparison with known congeners was based on García-Vásquez *et al.* [46-47],
159 Přikrylová *et al.* [48-49], Gillardin *et al.* [50], Muterezi *et al.* [51], Pariselle and Euzet [35,52],
160 and Fannes *et al.* [53]. Parasite diversity was summarized by the species richness index (S),
161 indices of Shannon (H) and Equitability of Pielou (J). Infection parameters: prevalence (P), mean
162 intensity (MI) and mean abundance (MA) were provided following definitions given by Margolis
163 *et al.* [54] and Bush *et al.* [55]. Statistical analysis was performed using Past 3.1 software.

164 **Results**

165 Fish processed for the study had different size and weight range. For *Oreochromis mweruensis*
166 (n=47) the mean TL was 18.2 ± 4.1 cm and 14.6 ± 3.2 cm for the mean SL, and the mean weight
167 was 72.7 ± 38.8 g. For *Coptodon rendalli* (n = 28) the mean TL was 15.1 ± 2.8 cm and $12.0 \pm$
168 2.4 cm for the mean SL, and the mean weight = 72.7 ± 38.8 g. For *Serranochromis*
169 *macrocephalus* (n = 11) the mean TL was 16.9 ± 3.4 cm and 14.0 ± 2.8 cm for the mean SL, and
170 the mean weight was 81.9 ± 51.5 g.

171 **Monogenean community composition and indices of diversity in the Upper Lufira basin**

172 Representatives of four genera of monogeneans, *Cichlidogyrus*, *Gyrodactylus* and *Scutogyrus*
173 (on the gills) and *Enterogyrus* (in the stomach), were collected (Table 1). Among them were ten
174 known species of *Cichlidogyrus*, one species of *Gyrodactylus*, two species of *Scutogyrus* and one
175 species of *Enterogyrus*. Parasite diversity indices were reported to be 10, 6 and 2 for S; 1.5, 1.2
176 and 0.6 for H; and 0.6, 0.8 and 0.8 for J respectively for *O. mweruensis*, *C. rendalli* and *S.*
177 *macrocephalus*. The distribution of monogeneans per sampling period or per season is shown in
178 Table 2.

179 **Infection parameters of monogenean parasites in the Upper Lufira basin**

180 Prevalence, mean intensity and mean abundance presented in this section take into account hosts
181 grouped without seasonal subdivision.

182 The highest prevalences recorded was 80.9% for *C. halli* on *O. mweruensis*, 92.3% for *C.*
183 *dossoui* on *C. rendalli*, and 9.1% for both *C. zambezensis* and *C. karibae* on *S. macrocephalus*. A
184 low prevalence of 2.1% was recorded for *C. tiberianus*, *S. cf. bailloni* for *O. mweruensis*, and
185 3.8% for *G. nyanzae* from *C. rendalli* (Figure 2).

186 For *G. nyanzae* the highest MI = 8.7 ± 9.9 was recorded from *O. mweruensis* and a low of MI= 1
187 ± 0 from *C. rendalli*. Conversely *C. papernastrema* obtained a MI of 17.1 ± 24 when examining
188 the latter fish host. For *S. macrocephalus*, *C. karibae* was the parasite with the highest mean
189 intensity (MI= 15) and *C. zambezensis* the lowest (MI= 5) (Figure 3).

190 The results regarding the mean abundance reveal that on *O. mweruensis*, *C. halli* (MA= $6.4 \pm$
191 7.7) is the most abundant species; on the gills of *C. rendalli*, *C. dossoui* (9.7 ± 15.6) is the most

192 abundant species; and the highest abundance of monogeneans on *S. macrocephalus* is 1.4 ± 4.5
193 per examined fish for *C. karibae* (Figure 4).

194 **Discussion**

195 This research was conducted to explore the monogenean parasite fauna of three economically
196 important and abundant cichlid species in the Upper Lufira basin, a part of the Upper Congo
197 basin. In this study thirteen gill and one stomach monogenean species were recorded. Parasite
198 species were already reported from fish belonging to the genera *Oreochromis*, *Coptodon* and
199 *Serranochromis* [35,51, 56]. Although few studies on monogenean parasites from the Congo
200 basin have been conducted in the Lake Tanganyika, Bangweulu-Mweru, Upper Lualaba, Kasai,
201 Lower Congo and Pool Malebo Ecoregions (*sensu* Thieme *et al.* [57]) (e.g. Vanhove *et al.* [58];
202 Gillardin *et al.*, [50]; Muterezi *et al.* [51]; Jorissen *et al.* [56, 59-60]; Geraerts *et al.* [61]), this
203 study is the first to record monogenean parasites in the Lufira basin.

204 The known host range of five parasite species is extended in this study. *Cichlidogyrus quaestio*,
205 *S. cf. bailloni* and *E. malmbergi* were recorded for the first time from *O. mweruensis*; *C. halli*
206 from *C. rendalli*; and *C. karibae* from *S. macrocephalus*. *Cichlidogyrus karibae* was described
207 by Douëllou [62] on *Sargochromis codringtonii* (Boulenger, 1908) in Lake Kariba (Zambezi
208 basin, Zimbabwe). *Enterogyrus malmbergi* was described by Bilong Bilong [63] from the
209 stomach of *Oreochromis niloticus* (Linnaeus, 1758) in the Sanaga River (Cameroon). *Scutogyrus*
210 *bailloni* was formally described by Pariselle and Euzet [52] on *Sarotherodon galilaeus* (L, 1758)
211 in the Mékrou River (Niger basin, Niger, West Africa). Since only a single similar parasite
212 specimen was retrieved in this study on the gills of *O. mweruensis*, it cannot be assigned to *S.*
213 *bailloni* with certainty. Nevertheless these (putative in case of *S. bailloni*) records substantially

214 expand the known geographical distribution of these three monogenean species. Considering
215 species richness, our results are similar to previous reports of monogenean gill parasites for these
216 fishes in the Congo basin. In this study, ten monogenean species were found on *O. mweruensis*,
217 while Jorissen *et al.* [56, 59] collected nine parasite species in the Bangweulu-Mweru ecoregion
218 on *O. mweruensis* (of which seven are shared, except for *Cichlidogyrus mbirizei*, *C. quaestio* and
219 *S. cf. bailloni* on *O. mweruensis* from the Lufira river system, and *C. cirratus* and *C.*
220 *papernastrema* on *O. mweruensis* from the Bangweulu-Mweru ecoregion). Six monogenean
221 species were found on *C. rendalli* in this study, while Jorissen *et al.* [59] collected five parasite
222 species (all but *C. halli* corresponding to those found in this study) in the Bangweulu-Mweru
223 ecoregion. On *S. macrocephalus*, two monogenean species (*C. karibae* and *C. zambezensis*) were
224 found in this study while Jorissen *et al.* [59] reported only the last species, on fewer host fish.
225 In terms of infection parameters, on *O. mweruensis*, one parasite species had a prevalence higher
226 than 50% in the Upper Lufira basin (*C. halli*, P= 80.9%) against two monogenean species in the
227 Bangweulu-Mweru reported by Jorissen *et al.* [59] (P= 57.1% for *C. dossoui* and *S.*
228 *gravivaginus*). On *C. rendalli*, *C. dossoui* (P= 92.3%) in the Upper Lufira basin, and *C. dossoui*,
229 *C. quaestio* and *C. tiberianus* in the Bangweulu-Mweru, have P>50% following comparison with
230 Jorissen *et al.* [59]. On *S. macrocephalus*, no parasite species had a prevalence higher than 50%
231 in the Upper Lufira basin, while *C. zambezensis* reaches a prevalence of 100% in the
232 Bangweulu-Mweru. Regarding the infection intensity (Table 1), on *O. mweruensis*, in the Upper
233 Lufira basin, the most infected fish harbour up to 30 specimens of *C. halli*, followed by 25
234 specimens of *G. nyanzae*, against 37 parasite specimens of *G. nyanzae* and 21 parasite specimens
235 of *C. cirratus* in Bangweulu-Mweru (reported by Jorissen *et al.* [59]). On *C. rendalli* in the
236 Upper Lufira basin, the most infected fish harboured up to 84 specimens of *C. papernastrema*,

237 followed by *C. dossoui* with 68 monogenean specimens against respectively 29 and 20
238 specimens of *C. dossoui* and *C. quaestio* in the Bangweulu-Mweru Ecoregion. Finally, on *S.*
239 *macrocephalus* in the Upper Lufira, the most infected fish contain up to 15 and 5 parasite
240 specimens of *C. karibae* and *C. zambezensis* respectively while Jorissen *et al.* [59] reported 21
241 parasite specimens of *C. zambezensis* in the Bangweulu Mweru. These differences in infection
242 parameters may be due to sample size, season, biogeographical distribution or other
243 environmental parameters, as communities of cichlid-infecting monogeneans have been observed
244 to fluctuate e.g. seasonally and between habitat types, and parasite species composition may
245 change between areas and basins [64-66].

246 **Conclusion**

247 We reported stomach and gill monogenean species richness and infection parameters from three
248 cichlid species in the Upper Lufira basin. A total of 13 monogenean species were recovered from
249 *O. macrochir*, *C. rendalli* and *S. macrocephalus*. These findings are the first record of
250 monogeneans in the Upper Lufira basin. For future sampling, it will also be interesting to study
251 other groups of fish parasites other than monogenean parasites, as well as other fish species or
252 families, to record the diversity of parasites [56, 59]. In addition, parasites can also be used as
253 bioindicators of water quality [67-69] in this ecosystem where there is a substantial
254 anthropogenic threat, especially from mine pollution [70-71]. The use of parasites as
255 bioindicators of environmental conditions has been applied previously on African cichlids [72].
256 This study can serve as a baseline whereby future studies conducted on fish from the Upper
257 Lufira basin can be compared to this study so as to establish if there has been a change in
258 parasite composition and parasite load over time.

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278 **Availability of data and materials**

279 Slides of monogenean parasites are available in the invertebrate collection of the Royal Museum
280 of Central Africa, Tervuren, Belgium.

281 **Authors' contributions**

282 ACM, JS and MPMV designed and supervised this study. ACM, EA, EJV contributed to
283 sampling, the collection and identification of fish. FMB, WJLP, WS, JRS and MPMV helped
284 with the collection and preparation of the gill parasites. AP, MWPJ, MPMV helped with the
285 morphological identification of parasites species. MPMV helped with the writing of the paper,
286 analysis of the data, interpretation and discussion of results and provided scientific background
287 in the field of monogenean research. All the authors critically read and edited the manuscript,
288 and approved the final manuscript.

289 **Ethics approval and consent to participate**

290 Fish were collected using nets or were bought from fishermen. In the absence of relevant animal
291 welfare regulations in the DRC, we had used the guidelines and authorization in accordance with
292 the Unité de Recherche en Biodiversité et Exploitation durable des Zones Humides (BEZHU) of
293 the Université de Lubumbashi

294 **Consent for publication**

295 Not applicable

296 **Competing interests**

297 The authors declare that they have no known competing financial interests or personal
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299 **References**

- 300 1. Shumway C, Musibono D, Ifuta S, Sullivan J, Schelly R, Punga J, et al. Biodiversity Survey:
301 Systematics, Ecology, and Conservation along the Congo River. Congo River Environment and
302 Development Project (CREDP), 2003; 127p.
- 303 2. Teugels GG, Thieme ML. Biological distinctiveness of African Ecoregions: Freshwater Fish
304 Biodiversity in the Congo basin. In: Thieme ML, Abell R, Skelton P, Lenher B, Teugels GG,
305 Dinerstein E et al. editors. Freshwater ecoregions of Africa and Madagascar. Island Press; 2005.
306 p. 35-70.
- 307 3. Runge J. The Congo River, Central Africa. In: Gupta A, editor. Large Rivers: Geomorphology
308 and Management; 2007; 293-309.
- 309 4. Roberts TR, Stewart DJ. An ecological and systematic survey of fishes in the rapids of the
310 Lower Zaïre or Congo River. *Bulletin Museum of Comparative Zoology*. 1976; 147(Suppl.
311 6):239-317.
- 312 5. Brummett R, Stiassny M, Harrison I. Background. In: Brooks EGE, Allen DJ, Darwall WRT,
313 editors. The Status and Distribution of Freshwater Biodiversity in Central Africa. Gland,
314 Switzerland and Cambridge, UK: IUCN; 2011.
- 315 6. Stiassny MLJ, Brummett RE, Harrison IJ, Monsembula R, Mamonekene V. The status and
316 distribution of freshwater fishes in central Africa. In: Brooks EGE, Allen DJ, Darwall WRT,

- 317 editors. The Status and Distribution of Freshwater Biodiversity in Central Africa. Gland,
318 Switzerland and Cambridge, UK : IUCN ; 2011.
- 319 7. Poll M. Poissons recueillis au Katanga par H.J. Bredo. Bulletin du Musée royal d'Histoire
320 naturelle de Belgique, Bruxelles, Tome XXIV, n° 21 ; 1948.
- 321 8. Damas H, Magis N, Nassogne A. Contribution à l'étude hydrobiologique des lacs
322 Mwadingusha, Koni & N'zilo. Université de Liège. Fondation de l'université de Liège pour les
323 recherches scientifiques au Congo et au Ruanda-Urundi 50p ; 1959.
- 324 9. Magis N. Nouvelle contribution à l'étude hydrobiologique des lacs de Mwadingusha, Koni et
325 N'zilo. Université de Liège. Fondation de l'université de Liège pour les recherches scientifiques
326 au Congo et au Ruanda-Urundi, 25p ; 1961.
- 327 10. Wilmet J. La répartition de la population dans la dépression des rivières Mufuvya et Lufira
328 (Haut-Katanga). Université de Liège. Fondation de l'université de Liège pour les recherches
329 scientifiques au Congo et au Ruanda-Urundi. Académie des sciences d'outre-mer, tome XIV,
330 fasc. 2 ; 1963.
- 331 11. Doumenge C, Palla F, Scholte P, Hiol Hiol F, Larzillière A. Aires protégées d'Afrique
332 centrale - État 2015. OFAC, Kinshasa, République Démocratique du Congo et Yaoundé,
333 Cameroun : 256 p ; 2015.
- 334 12. Squadrone S, Burioli E, Monaco G, Koya MK, Prearo M, Gennero S, et al. Human exposure
335 to metals due to consumption of fish from an artificial lake basin close to an active mining area
336 in Katanga (D.R. Congo). *Sci Total Environ.* 2016; 568:679-684.

- 337 13. Louette M, Hasson M. Rediscovery of the Lake Lufira Weaver *Ploceus ruweti*. *Bulletin ABC*
338 2009; 16(Suppl. 2): 168-173.
- 339 14. Craig AJFK, Hasson M, Jordaens K, Breman F, Louette M. Range extension of the Lufira
340 Masked Weaver *Ploceus ruweti*, endemic to Katanga province, Democratic Republic of Congo.
341 *OSTRICH* 2011; 82(Suppl. 1):77-78.
- 342 15. Ilunga MK, Abwe E, Decru E, Manda AC, Vreven E. Description of a new small-sized
343 *Synodontis* species (Siluriformes: Mochokidae) that is important for local subsistence fisheries in
344 the middle Lufira (upper Congo River, DR Congo). *J Fish Biol.* 2019;1-18.
- 345 16. Mulelenu CM, Manda BK, Decru E, Manda AC, Vreven E. The *Cyphomyrus* Myers 1960
346 (Osteoglossiformes: Mormyridae) of the Lufira basin (Upper Lualaba: DR Congo): A generic
347 reassignment and the description of a new species. *J Fish Biol.* 2020; 1-19.
- 348 17. Fonseca VG, Carvalho GR, Sung W, Johnson HF, Power DM, Neill SP, et al. Second-
349 generation environmental sequencing unmask marine metazoan biodiversity. *Nat Commun.*
350 2010; 1:98.
- 351 18. Vanhove MPM, Tessens B, Schoelinck C, Jondelius U, Littlewood DTJ, Artois T, et al..
352 Problematic barcoding in flatworms: A case-study on monogeneans and rhabdocoels
353 (Platyhelminthes). *ZooKeys.* 2013; 365:355-379.
- 354 19. Whittington ID. Diversity “down under”: monogeneans in the antipodes (Australia) with a
355 prediction of monogenean biodiversity worldwide. *Int J Parasitol.* 1998; 28:1481-1493.
- 356 20. Vanhove MPM, Snoeks J, Volckaert FAM, Huyse T. First description of monogenean
357 parasites in Lake Tanganyika: the cichlid *Simochromis diagramma* (Teleostei, Cichlidae)

- 358 harbours a high diversity of *Gyrodactylus* species (Platyhelminthes, Monogenea). Parasitology.
359 2011; 138:364-380 (Erratum in 138: 403).
- 360 21. Pariselle A, Morand S, Deveney MR, Pouyau L. Parasite species richness of closely related
361 hosts: historical scenario and “genetic” hypothesis, p. 147-166. In: Combes C, Jourdan J, editors.
362 Hommage à Louis Euzet - Taxonomie, écologie et évolution des métazoaires parasites.
363 Taxonomy, ecology and evolution of metazoan parasites. Perpignan, Les Presses Universitaires
364 de Perpignan, 2003. P. 380+396.
- 365 22. Thurston JP. The larva of *Oculotrema hippopotami* (Monogenea: Polystomatidae. J Zool.
366 1968; 154:475-480.
- 367 23. Thurston JP. The frequency distribution of *Oculotrema hippopotami* (Monogenea:
368 Polystomatidae) on *Hippopotamus amphibus*. J. Zool. 1968 ;154:481-485.
- 369 24. Silan P, Langlais M, Latu G. Dynamique des populations de monogènes, ectoparasites de
370 téléostéens : Stratégies démographiques et implications mathématiques. *Ecologie*, t. 1999
371 ;30(Suppl. 4) :1.
- 372 25. Silan P, Caltran H, Latu G. Ecologie et dynamique des populations de monogènes,
373 ectoparasites de téléostéens marins : approche et contribution montpelliéraines. Taxonomie,
374 écologie et évolution des métazoaires parasites. In: Combes C, Jourdan J, editors. PUP, France,
375 Perpignan Tome II. 2003;212-235.
- 376 26. Whittington ID, Cribb BW, Hamwood TE, Halliday JA. Host-specificity of monogenean
377 (platyhelminth) parasites: a role for anterior adhesive areas? Int J Parasitol. 2000; 30:305-320.

- 378 27. Ozturk T, Ozer A. Monogenean Fish Parasites, Their Host Preferences and Seasonal
379 Distributions in the Lower Kizilirmak Delta (Turkey). Turk J Fish Aquat Sc. 2014; 14:367-378.
- 380 28. Bagge AM. Factors affecting the development and structure of monogenean communities on
381 Cyprinid fish. Dissertation, University of Jyvaskyla 25p; 2005.
- 382 29. Llewellyn J. Amphidellid (monogenean) parasites of electric rays (Torpedinidae). J Mar
383 Biol Assoc UK. 1960; 39:561-589.
- 384 30. Euzet L, Combes C. The selection of habitats among the monogenean. Int J Parasitol. 1998;
385 28:1645-1652.
- 386 31. Jarkovský J, Morand S, Šimková A, Gelnar M. Reproductive barriers between congeneric
387 monogenean parasites (Dactylogyrus: Monogenea): attachment apparatus morphology or
388 copulatory organ incompatibility? Parasitol Res. 2004; 92:95-105.
- 389 32. Šimková A, Verneau O, Gelnar M, Morand S. Specificity and specialization of congeneric
390 monogeneans parasitizing cyprinid fish. Evolution. 2006;60(Suppl. 5):1023-1037.
- 391 33. Řehulková E, Mendlova M, Šimková A., Two new species of *Cichlidogyrus* (Monogenea:
392 Dactylogyridae) parasitizing the gills of African cichlid fishes (Perciformes) from Senegal:
393 morphometric and molecular characterization. Parasitol Res. 2013; 112:1399-1410.
- 394 34. Mendlová M, Šimková A. Evolution of host specificity in monogeneans parasitizing African
395 cichlid fish. Parasite Vector. 2014; 7:69.
- 396 35. Pariselle A, Euzet L. Systematic revision of dactylogyridean parasites (Monogenea) from
397 cichlid fishes in Africa, the Levant and Madagascar. Zoosystema. 2009;31(Suppl. 4):849-898.

- 398 36. Řehulková E, Seifertová M, Přikrylová I, Francová K. Monogenea. In: Scholz T, Vanhove
399 MPM, Smit N, Jayasundera Z, Gelnar M, editors. *Abc Taxa: A Guide to the Parasites of African*
400 *Freshwater Fishes*. 2018; Volume 18.
- 401 37. Cruz-Laufer AJ, Artois T, Smeets K, Pariselle A, Vanhove MPM. The cichlid–Cichlidogyrus
402 network: a blueprint for a model system of parasite evolution. *Hydrobiologia*. 2021; 848:3847-
403 3863.
- 404 38. Kottek M, Grieser J, Beck C, Rudolf B, Rubel F. World Map of the Köppen-Geiger climate
405 classification updated. *Meteorol Z*. 2006 ; 15(Suppl. 3) :259-263.
- 406 39. Katemo MB. Evaluation de la contamination de la chaîne trophique par les métaux lourds
407 dans le bassin de la Lufira supérieure (Katanga/RD Congo). DEA en Biologie Végétale et
408 Environnement. Université de Lubumbashi. République Démocratique du Congo, 44p ; 2009.
- 409 40. SNC-LAVALIN International. Etude sur la restauration des mines de cuivre et de cobalt en
410 République Démocratique du Congo, Rapport préliminaire M-6708 (603082), Montréal, p222 ;
411 2003.
- 412 41. Goortz A, Margis N, Wilmet J. Les aspects biologiques, humains et économiques de la pêche
413 dans le lac de barrage de la Lufira. Université de Liège. Fondation de l'université de Liège pour
414 les recherches scientifiques au Congo et au Ruanda-Urundi, 127p ; 1961.
- 415 42. Skelton P. *Freshwater fishes of Southern Africa*. Struik Publishers, Cape Town, 395p; 2001.
- 416 43. Lamboj A. *The Cichlid fishes of western Africa*. Verlag B.S. editor. Germany 256p; 2004.

- 417 44. Olivier PAS, Luus-Powell WJ, Saayman JE. Report on some monogenean and clinostomid
418 infestations of freshwater fish and waterbird hosts in Middle Letaba Dam, Limpopo Province,
419 South Africa. Onderstepoort J Vet Res. 2009; 76:187-199.
- 420 45. Nack J, Bitja Nyom AR, Pariselle A, Bilong Bilong CF. New evidence of a lateral transfer of
421 monogenean parasite between distant fish hosts in Lake Ossa, South Cameroon: the case of
422 *Quadriacanthus euzeti* n. sp. J Helminthol.2015;90:455-459.
- 423 46. García-Vásquez A, Hansen H, Shinn AP. A revised description of *Gyrodactylus cichlidarum*
424 Paperna, 1968 (Gyrodactylidae) from the Nile tilapia, *Oreochromis niloticus niloticus*
425 (Cichlidae), and its synonymy with *G. niloticus* Cone, Arthur et Bondad-Reantaso, 1995. Folia
426 Parasit. 2007; 54:129-140.
- 427 47. García-Vásquez A, Hansen H, Christison KW, Bron JE, Shinn AP. Description of three new
428 species of *Gyrodactylus* von Nordmann, 1832 (Monogenea) parasitizing *Oreochromis niloticus*
429 *niloticus* (L.) and *O. mossambicus* (Peters) (Cichlidae). Acta Parasitol. 2011;56(Suppl. 1):20-33.
- 430 48. Přikrylová I, Matějusková I, Musilová N, Gelnar M. *Gyrodactylus* species (Monogenea:
431 Gyrodactylidae) on the cichlid fishes of Senegal, with the description of *Gyrodactylus ergensi* n.
432 sp. from Mango tilapia, *Sarotherodon galilaeus* L. (Teleostei: Cichlidae). Parasitol Res. 2009;
433 106:1-6.
- 434 49. Přikrylová I, Blazek R, Vanhove MPM. An overview of the *Gyrodactylus* (Monogenea:
435 Gyrodactylidae) species parasitizing African catfishes, and their morphological and molecular
436 diversity. Parasitol Res. 2012; 110:1185-1200.

- 437 50. Gillardin C, Vanhove MPM, Pariselle A, Huyse T, Volckaert FAM. Ancyrocephalidae
438 (Monogenea) of Lake Tanganyika: II: description of the first *Cichlidogyrus* spp. Parasites from
439 Tropheini fish hosts (Teleostei, Cichlidae). Parasitol Res. 2012; 110:305-313.
- 440 51. Muterezi BF, Vanhove MPM, Van Steenberge M, Pariselle A. Ancyrocephalidae
441 (Monogenea) of Lake Tanganyika: III: *Cichlidogyrus* infecting the world's biggest cichlid and
442 the non-endemic tribes Haplochromini, Oreochromini and Tylochromini (Teleostei, Cichlidae).
443 Parasitol Res. 2012; 111:2049-2061.
- 444 52. Pariselle A, Euzet L. *Scutogyrus* gen. n. (Monogenea: Ancyrocephalidae) for *Cichlidogyrus*
445 *longicornis minus* Dossou, 1982, *C. l. longicornis*, and *C. l. gravivaginus* Paperna and Thurston,
446 1969, with description of three new species parasitic on African Cichlids. Journal of
447 helminthological society of Washington 1995;62(Suppl. 2):157-173.
- 448 53. Fannes W, Vanhove MPM, Huyse T. Redescription of *Cichlidogyrus tiberianus* Paperna,
449 1960 and *C. dossoui* Douëllou, 1993 (Monogenea: Ancyrocephalidae), with special reference to
450 the male copulatory organ. Syst Parasitol. 2017; 94:133-144.
- 451 54. Margolis L, Esch GW, Holmes JC, Kuris AM, Schad GA. The use of ecological terms in
452 parasitology (Report of an ad hoc committee of the American society of parasitologists). J
453 Parasitol. 1982;68(Suppl. 1):131-133.
- 454 55. Bush AO, Lafferty KD, Lotz JM, Shostak AW. Parasitology meets ecology on its own terms:
455 Margolis *et al.* revisited. J Parasitol. 1997 ;83(Suppl. 4):575-583.

- 456 56. Jorissen MWP, Huyse T, Pariselle A, Wamuini Lunkayilakio S, Muterezi Bukinga F, Chocha
457 Manda A, et al. Historical museum collections help detect parasite species jumps after tilapia
458 introductions in the Congo basin. *Biol Invasions*. 2020. [https://doi.org/10.1007/s10530-020-](https://doi.org/10.1007/s10530-020-02288-4)
459 02288-4.
- 460 57. Thieme ML, Abell R, Stiassny MLJ, Skelton P, Lehner B, Teugels GG, et al. *Freshwater*
461 *Ecoregions of Africa and Madagascar. A Conservation Assessment*. Island Press, Washington,
462 483p; 2005.
- 463 58. Vanhove MPM, Volckaert FAM, Pariselle A. Ancyrocephalidae (Monogenea) of Lake
464 Tanganyika: I: Four new species of *Cichlidogyrus* from *Ophthalmotilapia ventralis* (Teleostei:
465 Cichlidae), the first record of this parasite family in the basin. *Zoologia-Curitiba*. 2011
466 ;28(Suppl. 2) :253-263.
- 467 59. Jorissen MWP, Pariselle A, Huyse T, Vreven EJ, Snoeks J, Volckaert FAM, et al. Diversity
468 and host specificity of monogenean gill parasites (Platyhelminthes) of cichlid fishes in the
469 Bangweulu-Mweru ecoregion. *J Helminthol*. 2018 ;92(Suppl. 4):417-437.
- 470 60. Jorissen MWP, Pariselle A, Huyse T, Vreven EJ, Snoeks J, Decru E, et al. Six new
471 dactylogyrid species (Platyhelminthes, Monogenea) from the gills of cichlids (Teleostei,
472 Cichliformes) from the Lower Congo basin. *Parasite*. 2018 ;25(Suppl. 64).
- 473 61. Geraerts M, Muterezi Bukinga F, Vanhove MPM, Pariselle A, Chocha Manda A, Vreven E,
474 et al. Six new species of *Cichlidogyrus* Paperna, 1960 (Platyhelminthes: Monogenea) from the
475 gills of cichlids (Teleostei: Cichliformes) from the Lomami River basin (DRC: Middle Congo).
476 *Parasite Vector*. 2020; 13:187.

- 477 62. Douëllou L. Monogeneans of the genus *Cichlidogyrus* Paperna, 1960 (Dactylogyridae:
478 Ancyrocephalinae) from cichlid fishes of Lake Kariba (Zimbabwe) with descriptions of five new
479 species. Syst Parasitol. 1993; 25:159-185.
- 480 64. Morand S, Krasnov BR. The Biogeography of Host-parasite interactions. Oxford, UK:
481 University Press; 2010.
- 482 65. Akoll P, Fioravanti ML, Konecny R, Schiemet F. Infection dynamics of *Cichlidogyrus*
483 *tilapiae* and *C. sclerosus* (Monogenea, Ancyrocephalinae) in Nile tilapia (*Oreochromis niloticus*
484 L.) from Uganda. J Helminthol. 2012; 86:302-310.
- 485 66. Igeh PC, Gilbert BM, Avenant-Oldewage A. Seasonal variance in water quality, trace metals
486 and infection variables of *Cichlidogyrus philander* Douëllou, 1993 (Monogenea,
487 Ancyrocephalidae) infecting the gills of *Pseudocrenilabrus philander* (Weber, 1897) in the
488 Padda Dam, South Africa. Afr J Aquat Sci. 2020;1-12.
- 489 67. Sures B, Taraschewski H, Rydlo M. Intestinal fish parasites as heavy metal bioindicators: A
490 comparison between *Acanthocephalus lucii* (Palaeacanthocephala) and the Zebra Mussel,
491 *Dreissena polymorpha*. Bulletin of Environmental Contamination and Toxicology. 1997; 59:14-
492 21.
- 493 68. Thielen F, Zimmermann S, Baska F, Taraschewski H, Sures B. The intestinal parasite
494 *Pomphorhynchus laevis* (Acanthocephala) from barbel as a bioindicator for metal pollution in the
495 Danube River near Budapest, Hungary. Environ Pollut. 2004;129 :421-429.
- 496 69. Sanchez-Ramirez C, Vidal-Martinez VM, Aguirre-Macedo ML, Rodriguez-Canul RP, Gold-
497 Bouchot G, Sures B. *Cichlidogyrus sclerosus* (Monogenea: Ancyrocephalinae) and its host, the

- 498 Nile Tilapia (*Oreochromis niloticus*), as bioindicators of chemical pollution. *J Parasitol.* 2007
499 ;93(Suppl. 5) :1097-1106.
- 500 70. Katemo MB, Colinet G, André L, Chocha MA, Marquet JP, Micha JC. Evaluation de la
501 contamination de la chaîne trophique par les éléments traces (Cu, Co, Zn, Pb, Cd, U, V et As)
502 dans le bassin de la Lufira supérieure (Katanga/RD Congo). *Tropicultura.* 2010 ;28(Suppl.
503 4) :246-252.
- 504 71. Mees F, Masalehdani MNN, De Putter T, D'Hollander C, Van Biezen E, Mujinya BB, et al.
505 Concentrations and forms of heavy metals around two ore processing sites in Katanga,
506 Democratic Republic of Congo. *J Afr Earth Sci.* 2013; 77:22-30.
- 507 72. Madanire-Moyo GN, Luus-Powell WJ, Olivier PA. Diversity of metazoan parasites of the
508 Mozambique tilapia, *Oreochromis mossambicus* (Peters, 1852), as indicators of pollution in the
509 Limpopo and Olifants River systems. *Onderstepoort J Vet.* 2012;79(Suppl. 1) 362 [http://](http://dx.doi.org/10.4102/ojvr.v79i1.362)
510 dx.doi.org/10.4102/ojvr.v79i1.362.

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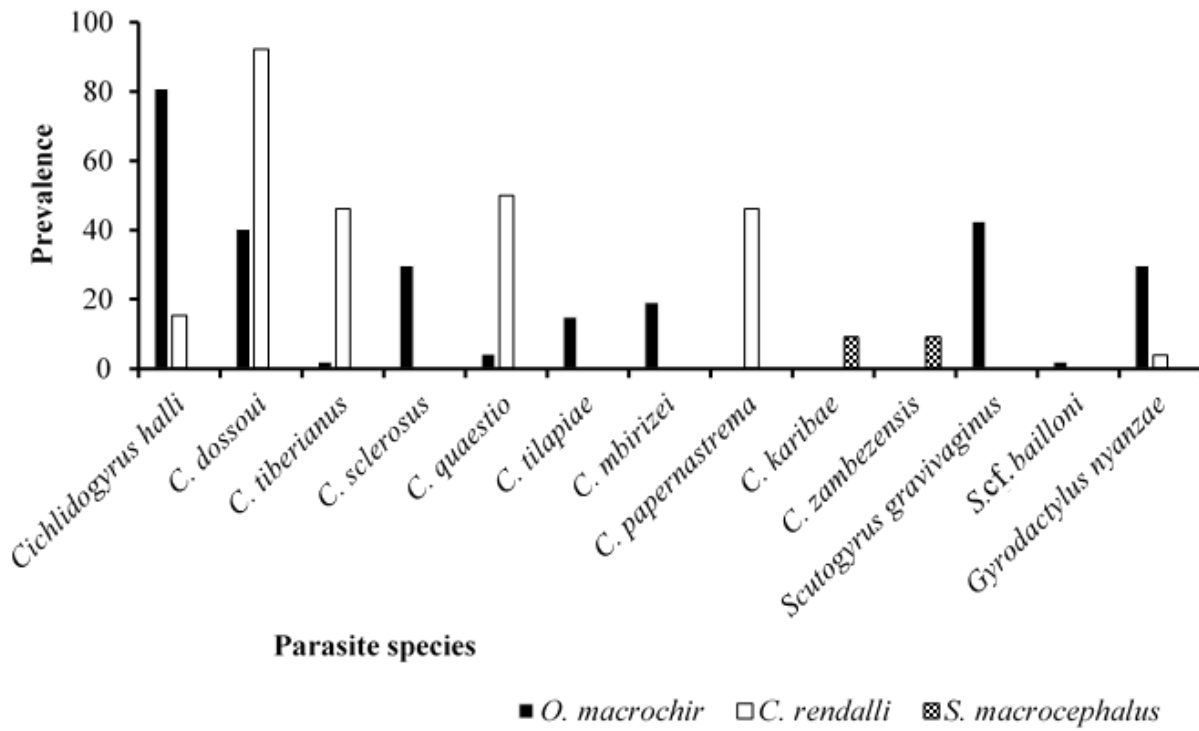
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525 **Figure 1** : Map of sampling sites in the Upper Lufira basin: Lufira River (Kaboko 11°4'31.60"S; 26°55'2.40"E and Buta

526 11°2'21.60"S; 26°57'23.10"E); Lake Tshangalele (Kisunka 10°50'52.10"S; 26°57'50.60"E, Kapolowe Mission 10°54'59.50"S; 26°58'17.70"E,

527 Yuka 10°56'25.30"S; 26°58'53.40"E and Mulandi 10°57'36.64"S; 27°6'44.88"E) and Lake Koni (Koni 10°43'3.65"S; 27°17'3.24"E)

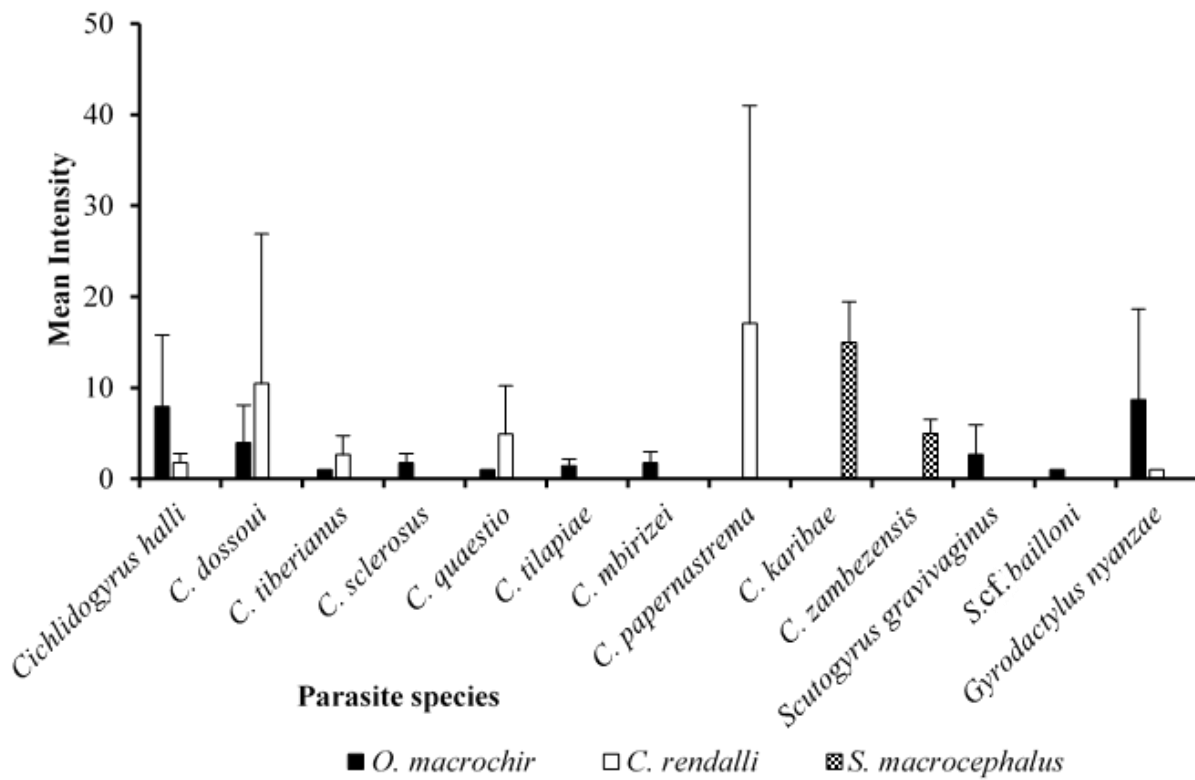


528

529 **Figure 2 :** Parasite prevalence (%) per monogenean species recovered on the gills of

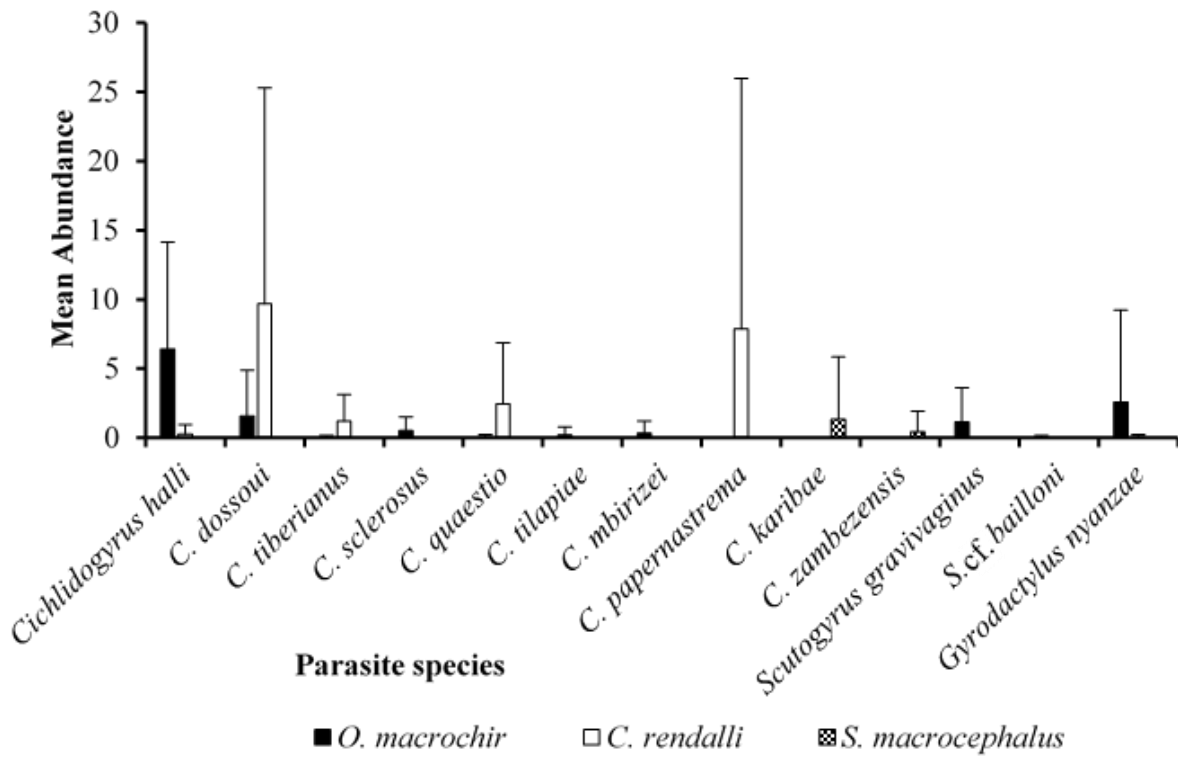
530 *Oreochromis mweruensis*, *Coptodon rendalli* and *Serranochromis macrocephalus* in the Upper

531 Lufira basin



532

533 **Figure 3** : Mean intensity of each monogenean species recovered on the gills of *Oreochromis*
534 *mweruensis*, *Coptodon rendalli* and *Serranochromis macrocephalus* in the Upper Lufira basin,
535 with bars about the mean indicating the standard deviation



536

537 **Figure 4 :** Mean abundance of each monogenean species recovered on the gills of *Oreochromis*

538 *mweruensis*, *Coptodon rendalli* and *Serranochromis macrocephalus* in the Upper Lufira basin,

539 with standard deviation

540

541 **Table 1** : The monogenean parasite species recovered from *Oreochromis mweruensis*, *Coptodon*
 542 *rendalli* and *Serranochromis macrocephalus* in the Upper Lufira basin

Parasite order	Parasite genus	Parasite species	Host species	# hosts examined	# hosts infected
Dactylogyridea	<i>Cichlidogyrus</i>	<i>C. halli</i> (Price & Kirk, 1937)	<i>O. mweruensis</i>	45	39
			<i>C. rendalli</i>	29	4
		<i>C. dossoui</i> Douëllou, 1993	<i>O. mweruensis</i>	45	19
			<i>C. rendalli</i>	29	25
		<i>C. sclerosus</i> Paperna & Thurston, 1969	<i>O. mweruensis</i>	45	14
		<i>C. tiberianus</i> Paperna, 1960	<i>O. mweruensis</i>	45	1
			<i>C. rendalli</i>	29	12

Parasite order	Parasite genus	Parasite species	Host species	# hosts examined	# hosts infected
		<i>C. quaestio</i> Douëllou, 1993	<i>O. mweruensis</i>	45	2
			<i>C. rendalli</i>	29	13
		<i>C. mbirizei</i> Muterezi Bukinga, Vanhove, Van Steenberge & Pariselle, 2012	<i>O. mweruensis</i>	45	9
		<i>C. tilapiae</i> Paperna, 1960	<i>O. mweruensis</i>	45	7
		<i>C. papernastrema</i> Price, Peebles & Bamford, 1969	<i>C. rendalli</i>	29	15
		<i>C. karibae</i> Douëllou, 1993	<i>S. macrocephalus</i>	11	1
		<i>C. zambezensis</i>	<i>S. macrocephalus</i>	11	1

Parasite order	Parasite genus	Parasite species	Host species	# hosts examined	# hosts infected
		Douëllou, 1993			
	<i>Enterogyrus</i>	<i>E. malmbergi</i> Bilong Bilong, 1988	<i>O. mweruensis</i>	1	1
	Paperna, 1963				
	<i>Scutogyrus</i>	<i>S. gravivaginus</i> (Paperna & Thurston, 1969)	<i>O. mweruensis</i>	45	20
	Pariselle and Euzet, 1995				
		<i>S. cf. bailloni</i> Pariselle & Euzet, 1995	<i>O. mweruensis</i>	45	1
Gyrodactylidea	<i>Gyrodactylus</i> Von	<i>G. nyanzae</i> Paperna, 1973	<i>O. mweruensis</i>	45	12
	Nordmann, 1832				
	Bychowsky, 1937		<i>C. rendalli</i>	29	1

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544

545 **Table 2** : X/Y: Number of specimens of a given parasite species, out of the number of infected
 546 fish per host according to sampling period [August, September (Sept.): dry season; March, April:
 547 rainy season]

Oreochromis mweruensis

Monogenean species	Sampling date				
	Sept. 2015	March 2016	April 2016	August 2016	Sept. 2017
<i>Cichlidogyrus dossoui</i>	7/5	17/3	22/7	13/1	16/3
<i>C. halli</i>	40/11	51/5	61/10	3/2	150/11
<i>C. mbirizei</i>			2/2		14/7
<i>C. quaestio</i>	1/1		1/1		
<i>C. sclerosus</i>	2/1	1/1	9/7		13/5
<i>C. tiberianus</i>			1/1		
<i>C. tilapiae</i>	1/1	2/1	3/3		2/2
<i>Gyrodactylus nyanzae</i>	26/4		23/1		67/7
<i>Scutogyrus gravivaginus</i>	5/4	5/3	28/6	7/2	9/5
<i>S. cf. bailloni</i>			1/1		

Total number of monogeneans, all species included	82	76	151	23	271
Number of examined fish	12	5	13	2	13
<i>Coptodon rendalli</i>					
Sampling date	Sept.	March	April	August	Sept.
Monogenean species	2015	2016	2016	2016	2017
<i>Cichlidogyrus dossoui</i>	41/8	44/6	170/8	33/2	2/1
<i>C. halli</i>	1/1		6/3		
<i>C. papernastrema</i>	7/6		149/5	50/2	5/2
<i>C. quaestio</i>	38/7	21/3	22/2		1/1
<i>C. tiberianus</i>	4/3	3/2	22/5	3/2	
<i>Gyrodactylus nyanzae</i>	1/1				
Total number of monogeneans, all species included	92	68	369	86	8

Number of examined fish	10	6	8	2	3
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Serranochromis macrocephalus

Sampling date	Sept.	August
	2017	2018

Monogenean
species

Cichlidogyrus karibae 15/1

C. zambenzensis 5/1

Total number of 20

monogeneans, all species

include

Number of examined fish	1	10
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