A wheat chromosome segment substitution line series supports characterisation and use of progenitor genetic variation

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1 Abstract

Genome-wide introgression and substitution lines have been developed in many plant species. 2 3 enhancing mapping precision, gene discovery and the identification and exploitation of 4 variation from wild relatives. Created over multiple generations of crossing and/or backcrossing accompanied by marker-assisted selection, the resulting introgression lines are a 5 6 fixed genetic resource. In this study we report the development of spring wheat chromosome segment substitution lines generated to systematically capture genetic variation from tetraploid 7 (Triticum turgidum ssp dicoccoides) and diploid (Aegilops tauschii) progenitor species. 8 Generated in a common genetic background over four generations of backcrossing, the material 9 is a base resource for the mapping and characterisation of wheat progenitor variation. To 10 facilitate further exploitation the final population was genetically characterised using a high-11 density genotyping array and a range of agronomic and grain traits assessed to demonstrate the 12 the potential use of the populations for trait localisation in wheat. 13

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15 Keywords: introgression, CSSL, Aegilops tauschii, Triticum aestivum

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18 Introduction

Hexaploid wheat (*Triticum aestivum*) arose through natural hybridization between the tetraploid grass species *T. turgidum* and the diploid species *Aegilops tauschii* (Fu & Somers, 2009). The infrequency of hybridisation resulted in limited genetic diversity (Cox, 1997) and progenitor species are a valuable source of variation (Zhang *et al.*, 2015). Numerous strategies have been proposed and deployed to capture this for application in genetic studies and wheat breeding (Leigh *et al.*, 2022).

A wealth of genomic and germplasm resources is now available for functional genetic 25 26 characterisation in polyploid wheat (Adamski et al., 2020). This includes mapping and gene discovery resources for wild relatives (e.g. stable wild wheat introgression lines developed by 27 Grewal et al., 2020). These build on early cytogenetic resources including monosomic and 28 nullisomic chromosome substitution lines series (Sears, 1954; Unrau et al., 1956) which 29 supported the foundations of wheat genetics (Kuspira and Unrau, 1957; Law, 1966). The 30 original monosomics took over 25 years to develop (Berke et al., 1992) and were used to 31 characterise a range of traits (Zemetra et al., 1986) as well as to create reciprocal series of 32 chromosome substitution lines, allowing genetic trait mapping (Berke et al., 1992). 33

Substitution line series serve many purposes. Law (1966) described their benefits as 34 35 three-fold: (1) localisation of genetic effects enabling targeted use in breeding, (2) revealing genetic trait architecture and relating it to patterns of descent and (3) validating predictions of 36 magnitude of genetic effects. Their direct use in breeding has been demonstrated as base 37 populations for trait mapping and line selection (Eshed et al., 1992), identification (Basava, 38 39 2019) and transfer of wild species variation (Eshed & Zamir, 1994; Nie et al., 2015) and pyramiding of target regions or introgression segments (Ali et al., 2010). In understanding trait 40 41 architecture, they support precise QTL mapping (Basava, 2019) and fine mapping (Fulop et al., 2016), gene discovery and cloning (Eshed & Zamir, 1994) along with marker development 42 43 (Qiao et al., 2016). Substitution lines typically show little variation for agronomic (growth and development) traits and can be used to confirm predicted magnitude of genetic effect, thereby 44 increasing homogeneity between experiments (Keurentjes et al., 2007) as well as eliminating 45 the interference of genetic background (Zhai et al., 2016). They are permanent populations 46 47 (Zhai *et al.*, 2016) that can be maintained as immortal seed stocks allowing assessment across environments and traits (Keurentjes et al., 2007), thereby increasing statistical power to detect 48 small effects QTLs and accurately determine the magnitude of genotype x environment 49 interactions (Rae et al, 1999; Keurentjes et al., 2007). 50

Chromosome segment substitution lines (CSSLs) are a form of substitution series and 51 capture genome-wide diversity from a donor species in a fixed genetic background. First 52 described in tomato as a series of interspecific introgression lines capturing small, overlapping 53 chromosome segments of the wild species Lycopersicum pennellii into cultivated tomato (L. 54 esculentum; Eshed et al., 1992; Eshed & Zamir, 1994) the resulting lines were used to map 55 quantitative traits including yield (Eshed & Zamir, 1995; Fridman et al., 2004) and leaf 56 characters (Holtan & Hake, 2003). The introgression lines have been subsequently used to 57 detect thousands of QTLs for adaptation, morphological characters, yield, metabolism and fruit 58 59 quality traits (reviewed in Lippman et al., 2007) and cellular features underlying leaf traits 60 (Chitwood *et al.*, 2013).

Further application has been demonstrated in the model species *Arabidopsis thaliana* (Keurentjes *et al.*, 2007; Fletcher *et al.*, 2013) as well as in several cultivated crops including rice (reviewed by Ali *et al.*, 2010), brassica (Howell *et al.*, 1996; Ramsay *et al.*, 1996; Rae *et al.*, 1999; Li *et al.*, 2015), peanut (Fonceka *et al.*, 2012), durum wheat (Blanco *et al.*, 2006) and pearl millet (Kumari *et al.*, 2014; Basava *et al.*, 2019). A recent review of resources by Balakrishnan *et al.* (2019) detailed populations available over sixteen cultivated crop species.

Despite the importance of wheat in global food security and its limited diversity due to 67 68 evolutionary history (Stebbins, 1950; Gross & Olsen, 2010) there are few substitution resources available beyond the foundation monosomic and nullisomic series. In hexaploid wheat Zemetra 69 70 et al. (1986) used the Nebraska reciprocal CSSL series to map heading date. More recently Gu et al. (2015) developed four sets of hexaploid introgression lines capturing the endemic 71 72 Chinese wheat subspecies T. aestivum yunnanense, tibetanum and petropavlovskyi along with a synthetic hexaploid wheat. The lines were used to map QTLs for height, spike length and 73 74 grain number per spike and several lines were identified for use in breeding to incorporate favourable introgression segments. In tetraploid wheat, Joppa and Cantrell (1990) created a 75 76 'Langdon'(durum)-T. dicoccoides substitution series via crossing between a high grain protein content (GPC) T. dicoccoides donor and a series of 'Langdon' D-genome disomic substitution 77 78 lines. This allowed the identification of lines with superior GPC, subsequently leading to its mapping on chromosome 6B (QGpc.ndsu-6Bb; Joppa et al., 1997). Further refinement of 79 80 mapping to the 6B short arm (Olmos et al., 2003) provided the basis for the cloning of the GRAIN PROTEIN CONTENT-B1 (GPC-B1) gene (Uauy et al., 2006a; Uauy et al., 2006b). 81 This demonstrates the utility of precision introgression resources for trait mapping and gene 82 cloning. In durum wheat, Blanco et al. (2006) also developed a set of 92 backcross 83 introgression lines using T. turgidum ssp. dicoccoides. Developed to capture high GPC from 84

the wild species, several generations of backcrossing allowed extraction of a single favourable
line. This was used to derive a population with the original recurrent parent and to extract high
and low protein bulks from the derived F₃ population for marker discovery and QTL detection.

Selection of CSSLs within a series aims to capture segments supporting genetic 88 dissection and that optimise donor coverage. Balakrishnan et al. (2019) reviewed the breadth 89 90 and characteristics of existing CSSL resources reporting that sets were composed of 35-200 lines, typically capturing 90% of donor genome. Tian et al. (2006) reported 67% coverage for 91 wild to cultivated rice compared to near complete (99%) coverage in other rice population (Jie 92 93 et al., 2006). In supporting genetic dissection, substitution line phenotypic effects that differ between an introgression line and the recurrent parent are ascribed to the substituted region 94 (Rae et al., 1999). To optimise this Wu et al. (2006) proposed an additive-dominance model 95 for accurately determining and ascribing substituted segment effects. 96

In this study we describe the development and characterization of a CSSL series 97 capturing the genomes of hexaploid wheat's primary progenitor species: tetraploid wild emmer 98 (T. turgidum ssp. dicoccoides; AB genomes) and diploid goat grass (Ae. tauschii; D genome). 99 100 We genetically characterize the resource using a high-density marker platform to provide accurate resolution of introgression boundaries. We demonstrate that the series can be used to 101 102 associate trait variation from progenitor species to specific genetic regions. The series is publicly available to provide characterised introgression lines (and associated markers) to 103 support further trait research and progression into breeding. 104

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106 Materials and Methods

107 Selection of parental material

Two independent backcross-derived CSSL populations were generated to capture the genomes 108 of the progenitor species T. turgidum ssp. dicoccoides (denoted NIAB_AB) and Ae. tauschii 109 (denoted NIAB D). The hexaploid spring wheat 'Paragon' was used as the recurrent parent for 110 both populations as it has been widely used for genetic studies and the creation of genetic 111 stocks. For the NIAB_AB population, the Israeli wild emmer wheat accession TTD-140 was 112 selected as the donor parent. For the NIAB_D population, the Armenian Ae. tauschii accession 113 ENT-336 was selected and the synthetic hexaploid line NIAB-SHW041 (created via 114 resynthesis from a Hoh-501 (AB) x ENT-336 (D) cross) used as the introgression donor. 115

Comparative genetic analysis of 'Paragon', TTD-140 and NIAB-SHW041 used single
nucleotide polymorphism (SNP) marker data generated using the Axiom® Wheat Breeder's
Genotyping Array (Allen *et al.*, 2017). Two datasets were used, the first with A and B genome

mapped markers and 'Paragon', 24 T. dicoccoides (including TTD-140), 12 T. dicoccum, 12 T. 119 durum accessions and the second using only D genome markers and 'Paragon' and 51 primary 120 synthetic hexaploid wheats (including NIAB-SHW041). Markers were thinned using pairwise 121 comparisons between SNPs using a Pearson correlation test and a single marker was removed 122 in comparisons that yielded an absolute value of the correlation coefficient (r) greater than 123 124 0.75. The final marker number in the A/B and D genome datasets 2,748 and 217, respectively. The relationships between individuals in each set were compared using principal coordinate 125 analysis (PCoA) of Euclidean genetic distance matrices computed from the marker data, 126 127 implemented in the R package ape (v5.3, Paradis & Schliep 2018).

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129 *Construction of populations*

For NIAB_AB, pollen from the wild emmer wheat donor TTD-140 was used to fertilise 130 'Paragon' to create a pentaploid F₁ generation. Six F₁ plants were used as the maternal parents 131 in the creation of the BC1 generation, where 'Paragon' was used to pollinate F1 plants, 132 continuing to BC₄. For NIAB_D the donor ENT-336 was first crossed to the durum wheat Hoh-133 134 501 to produce a fertile hexaploid synthetic wheat (NIAB-SHW041). The synthetic wheat was crossed to 'Paragon' and the F₁ progeny then backcrossed four times to 'Paragon' to derive 135 136 BC₄ plants. At each BC generation genomic DNA was extracted from 2-week-old seedlings of all progenies using a modified Tanksley extraction protocol (Fulton et al., 1995). Marker-137 assisted selection (MAS) was used to select individuals carrying heterozygous introgressions 138 in the F_1 to BC₄ generations and homozygous introgressions in the BC₄F₂ generation. A 139 schematic of the crossing scheme is given in **Supplementary Figure S1**. The number of lines 140 generated, genotyped, and advanced at each generation is summarised in Supplementary 141 142 Table S1.

MAS at each BC generation used co-dominant KASPTM markers developed to cover the TTD-140 and ENT-336 genomes and to allow for differentiation from 'Paragon'. In addition to foreground selection, evenly spaced markers on the non-target genomes were used to allow background selection against the donor. At BC₄F₄ additional genome-wide SNP genotyping (using the Axiom® Wheat Breeder's Genotyping Array; Allen *et al.*, 2017) was used to improve the accuracy of final selection.

For the NIAB_AB population, 191 KASPTM markers (98 A genome; 93 B genome)
distributed at approximately 20-30 cM intervals were selected to screen the BC generations
(Supplementary Table S2). An additonal set of 50 co-dominant D-genome mapped markers
(1-2 per chromosome arm) was used to facilitate selection against tetraploids and nullisomic

aneuploids and amplified a product in D-genome diploids and hexaploids, but not in ABgenome tetraploids, and gave expected results across nulli-tetrasomic series (Supplementary
Table S3).

Based on graphical genotypes derived from KASPTM marker positions, subsets of BC₁ 156 lines were manually created. The BC₁ lines were selected to carry multiple but complementary 157 158 heterozygous introgressions across the TTD-140 genome. Lines that failed to amplify at loci targeted by D genome specific KASPTM markers were discarded. For each subsequent cycle of 159 backcrossing, the same selection criteria were applied. A summary of the number of individuals 160 161 created, genotyped, and selected at each crossing generation is given in **Supplementary Table** S1. From each of the 138 selected BC_4F_1 individuals, $8 BC_4F_2$ seeds were grown and genotyped 162 (using the above KASPTM marker set; Supplementary Table S2) and one individual was 163 selected based on homozygosity for the marker at the target introgression but minimising 164 background introgressions (using markers in Supplementary Table S3). Eight BC₄F₃ seeds 165 from each selected homozygous BC₄F₂ line were subsequently sown and genotyped with the 166 KASPTM marker set to identify the target introgression and to ensure progression of 167 homozygous lines. 168

At the BC₄F₄ generation, the lines were genotyped using the Axiom[®] Wheat Breeder's 169 170 Genotyping Array (Allen et al., 2017). To derive the final NIAB_AB population we used 1,444 A-genome and 1,707 B-genome SNPs which were verified to share the same genetic and 171 physical genome chromosome. Genetic positions were taken from the existing consensus map 172 (Allen et al., 2017) and physical positions determined using the Basic Local Alignment Search 173 174 Tool (BLAST+) command-line applications on the RefSeq v1.0 genome assembly with default parameters (Camacho et al., 2009, International Wheat Genome Sequencing Consortium 175 176 (IWGSC) 2018). Only markers with a single A or B genome physical hit were used. Introgression selections were made using physical positions and graphical visualisation of 177 allele variation different from 'Paragon' using Flapjack (v-1.18.06.29, Milne et al., 2010). 178 Introgressions were visually selected on: (a) coverage across the target genome chromosome, 179 (b) homozygosity within each introgression and (c) minimal off-target introgressions. 180

For the NIAB_D population, 60 D-genome KASPTM markers were selected based on distribution across the D-genome, co-dominance and polymorphism between the recurrent parent 'Paragon', the tetraploid component of NIAB-SHW041, Hoh-501 and the *Ae. tauschii* donor ENT-336 (**Supplementary Table S4**). In addition, 66 A- (33) and B-genome (33) markers were used for background selection against Hoh-501 (**Supplementary Table S5**). The 186 CSSLs were developed using MAS as described for the NIAB_AB population but with
'Paragon' used as the maternal parent, as summarized in Supplementary Table S1.

At the BC_4F_4 generation, lines were genotyped using the Axiom[®] Wheat Breeder's 188 Genotyping Array. Using the same approach as for the NIAB_AB population, 644 SNPs which 189 shared a genetic and physical D genome chromosome (where markers had only a single 190 191 physical D genome hit) were used for the final line selection. Introgression line selections were based on coverage across each chromosome, maximising homozygosity and minimising off-192 target introgressions. Seed of both the NIAB_AB and NIAB_D populations is available from 193 194 the Germplasm Resources Unit at the John Innes Centre (entries WCSSL0001 to WCSSL0112; available via https://www.seedstor.ac.uk/) and all genotypic data is available from 195 www.cerealsdb.uk.net/cerealgenomics/CerealsDB/array info.php. 196

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198 Phenotypic characterisation

The NIAB_AB population was grown under glasshouse conditions to assess grain yield per plant and grain characteristics. Two or three replicate plants of each line were grown in an incomplete block design, surrounded by a 'Paragon' border to minimise edge effects. Each plant was grown in a 1L pot with 16-hour daylength at 22°C and 15°C overnight. At maturity, all plants plus one ear per plant were photographed, and all plant images are available from:

204 <u>https://opendata.earlham.ac.uk/wheat/under_license/toronto/Horsnell,Leigh,Bentley,Wright_</u>

205 <u>2020 NIAB_CSSL D genome_yield trial H2020/</u>. Seed from each plant was hand threshed
 206 and weighed to estimate grain yield per plant. Grains were analysed using a MARVIN seed
 207 analyser (MARViTECH GmbH) to obtain grain dimensions and thousand grain weight (TGW).

To assess phenotypic variance in the NIAB_D CSSLs, three replicates of each line 208 along with six replicates of the recurrent parent 'Paragon' were sown in a randomised yield 209 (2x6 m harvested area) trial in Hinxton, Cambridge in 2020. The trial was laid out in an 210 incomplete block design, with 6 plots per block and three complete replicates. Traits assessed 211 in the field included: grain yield (kg/plot at 85% dry matter content), early ground coverage 212 using normalised difference vegetation index (NDVI^{early}; an average of two measurements 213 taken between 11:00 and 14:00 GMT on the 15th and 21st of April 2020 at 2m above the canopy 214 using a handheld RapidScan CS-45 (Holland Scientific)), plant height (cm from base to tip 215 (including scurs or awns) of three random plants per plot at maturity), flowering time (days to 216 growth stage (GS) 61 (start of anthesis) recorded when 50% of the plot was at GS61; Zadoks 217 et al., 1974). All field trial data is available from Grassroots (Bian et al., 2017): 218 https://grassroots.tools/fieldtrial/study/61faaf25c68884365e7bcc34. Images of each plot were 219

taken to assess glaucosity using a RGB camera (Olympus TG4) at a height of 320cm from the 220 ground. Images were cropped to eliminate edge distortion with the final 8 Mpix image covering 221 85% plot. Post-processed images for each plot are available 222 of the here: https://opendata.earlham.ac.uk/wheat/under_license/toronto/Horsnell,Leigh,Bentley,Wright_ 223 2020_NIAB_CSSL_D_genome_yield_trial_H2020/. Post-harvest yield component traits were 224 225 assessed on ten replicate ears per plot (collected at random) including ear length (cm from base to tip of the ear, not including scurs or awns) and spikelet number. Each of the 10 ears was 226 weighed to determine ear weight, threshed individually and grains analysed using a MARVIN 227 228 seed analyser (MARViTECH GmbH) to determine seed number, thousand grain weight 229 (TGW) and grain dimensions (length, width).

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231 *Phenotypic analysis*

For both the NIAB_AB glasshouse and NIAB_D field trial, analysis was conducted in R v4.0.3 232 233 (R Core Team, 2020). Each trial included the final CSSLs selected to form the graphical genotype as well as a proportion of CSSLs that were not selected in the final population but 234 235 carried introgressions. The NIAB AB glasshouse trial analysis included 146 replicated CSSL individuals. In the NIAB D field trial analysis there were 33 replicated CSSLs. For each trait, 236 237 variance between replicates and population distributions were plotted and visually inspected to remove outliers. In the NIAB_D field trial, 10 ears were harvested per plot as technical 238 replicates for measurements on ear and seed characteristics. The ears were measured separately 239 and variance within a plot was then inspected to aid in outlier removal. The technical replicates 240 were then averaged to form plot means for the analysis. 241

Mixed linear effect models were constructed with restricted maximum likelihood 242 (REML) using the R packages lme4 (Bates et al., 2015) and lmerTest (Kuznetsova et al., 2017). 243 Genotype was treated as a fixed factor and the trial design components including 'replicate' 244 and 'block' were tested within each model as random factors. Due to visual patterns in model 245 246 residuals, in addition to the direction of phenotypic assessment and application of agronomic inputs, the trial layout components 'column' in the NIAB_D and 'row' in the NIAB_AB 247 glasshouse trial were included as random factors. Final model selection was based on Akaike 248 249 Information Criterion (AIC) and (exact) restricted likelihood ratio tests implemented through the R package RLRsim (Scheipl et al., 2008). In each model, the significance of the genotypic 250 effect was estimated using ANOVA with Satterthwaite's degrees of freedom method, 251 implemented through lmerTest. Generalised heritability (H^2) was estimated using genotypic 252 best linear unbiased predictors (BLUPs) and the Cullis et al. (2006) model, using the R script 253

developed by Schmidt *et al.* (2019). Ranked bar plots, including only the final selected CSSLs,
were created for grain yield (NIAB_D trial), seed width (NIAB_D trial) and seed length
(NIAB_AB trial). The package emmeans (Lenth, 2020) was used to complete pairwise
comparisons of all BLUEs for these traits and a false discovery rate (FDR) correction was used
to adjust *P* values for multiple testing. Finally, the package predictmeans (Luo *et al.*, 2020)
was used to inspect the residuals from final models and to extract the BLUEs for examining
variation across all traits.

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262 Introgression-trait association

Categorical traits were assessed in each CSSL population to demonstrate the use of the resource 263 for detecting introgression-trait association. In NIAB_AB this used a major known locus linked 264 to awn presence and in NIAB_D to a putative marker association with flag leaf glaucousness. 265 Awn presence was scored in the NIAB_AB glasshouse trial and graphical genotypes were 266 267 inspected to detect introgressions from the awned donor TTD-140 overlapping with regions where major awn inhibitors have previously been mapped. Huang et al. (2020) characterised 268 269 and identified the B1 locus as a C2H2 zinc finger encoding gene that corresponds to a coding region on chromosome 5A (698,528,636 to 698,529,001bp; IWGSC 2018). This region was 270 271 taken as the physical location of the gene for the introgression-trait association.

To demonstrate that CSSLs can also be used to dissect the genetic basis of a less well 272 characterised association, ear, stem, and leaf glaucousness was scored using visual assessment 273 and RGB images in the NIAB_D field trial. This identified a subset of individuals that shared 274 the non-glaucous phenotype, and these were assessed for common introgression regions. A 275 previous study (Würschum et al., 2020a) detected a putative marker-trait association linked to 276 277 flag leaf glaucousness mapped to 2D (~2.9 Mb) and the ability of the series to validate this interval was assessed via introgression-trait association. Off target regions were also inspected 278 279 to confirm that the subset shared no other common introgressions. Once each introgressiontrait association was identified, schematics using graphical genotype selections were formed 280 for each association and a SNP cluster plot was extracted from Axiom Analysis Suite (Thermo 281 Fisher Scientific) that confirmed the CSSL region segregating within the introgressions of 282 283 interest. For both populations, markers that were excluded or dropped during the QC were inspected visually to improve resolution of introgression boundaries. 284

285

286 **Results**

287 *Genetic relationships between parents*

The genetic relationships between 48 tetraploid wheat accessions and 51 D genome donors (as 288 novel SHWs) were compared based on Euclidean genetic distance and plotted through PCoA 289 (Supplementary Figure S2). TTD-140, the tetraploid donor of the NIAB AB population 290 clustered with most of the T. dicoccoides lines, though the T. dicoccoides accessions were 291 shown to contain significant diverse (Supplementary Figure S2A). The main cluster of T. 292 dicoccoides was separate from the cultivated wheats by PCoA1, which explained 12% of the 293 variation. The domesticated wheats, including accessions of T. durum, T. dicoccum and 294 'Paragon', all clustered above zero on PCoA1. These cultivated groups were separated by the 295 296 second axis which explained 9% of the variation.

The synthetic donor used for the NIAB_D population (NIAB-SHW041) clustered closely with recurrent parent 'Paragon' on the first PCoA axis (**Supplementary Figure S2B**), where there was a distinct group of primary synthetics that clustered separately from the rest of the material towards the right of the plot (slightly above 5 on PCoA1). PCoA1 accounted for 20% of the variation compared to 12% PCoA2. However, within the lineage shared by NIAB-SHW041 and 'Paragon', the two lines do not appear closely related, which is supported by their separation on PCoA2 in **Supplementary Figure S2B**.

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305 Selection of introgression lines based on graphical genotypes

Final lines were selected based on graphical genotypes from the BC₄F₄ (Supplementary Table 306 307 S1). Two different sets of SNP markers were used for line selection in NIAB AB: 1,444 markers with physical locations on the A genome and 1,707 markers with physical positions 308 309 on the B genome. From these data sets, 44 genotypes were identified that had A genome introgressions from TTD-140 (Figure 1A). Using the B genome data set, 33 genotypes were 310 identified with introgressions covering the B genome (Figure 1B). These 77 selections 311 originated from 62 unique NIAB_AB individuals with some genotypes selected for 312 introgressions at multiple loci. For example, NIAB_AB 1A.04 and 5A.05 represent the same 313 unique individual but are included twice to represent distinct target introgressions. Using the 314 NIAB_D population and a D genome marker set of 644 SNPs, 32 genotypes were selected with 315 introgressions from NIAB-SHW041 (Figure 1D). These selections originated from 25 unique 316 NIAB_D lines. 317

Individuals were typically selected for a single introgression and genotypes with multiple introgressions were selected against. However, the majority of BC₄F₄ individuals carried small off-target introgressions (**Figure 1**). Within each genome, each selected genotype contained on average two introgressions (**Figure 1**; **Supplementary Table S6**). The average

length of the introgression ranged from an average of 102 Mb for the B genome to 129 Mb for 322 the D genome. Off-target introgressions were also present across the different genomes for 323 each selected line (Supplementary Figures S3-S5). Therefore, as an estimate of recurrent 324 parent background recovery, the percentage of markers with a 'Paragon' allele was quantified 325 for each genotype, using markers with mapped genetic positions on the consensus map (Allen 326 et al., 2017) and genotypes had 96% average recurrent parent recovery (Supplementary Table 327 S6). Several NIAB_AB lines appeared to have large chromosomal introgressions from TTD-328 140 across the D genome (Supplementary Figures S3, S4). As the tetraploid donor lacks a D 329 330 genome, these lines were likely D-genome aneuploids and alternative (not 'Paragon') alleles were called for markers in that region. The extent of amplification of homeologous loci is not 331 characterised for the markers described herein, although D genome amplification in tetraploid 332 accessions indicates that for some markers, priming sites are sufficiently conserved across 333 genomes to allow amplification. 334

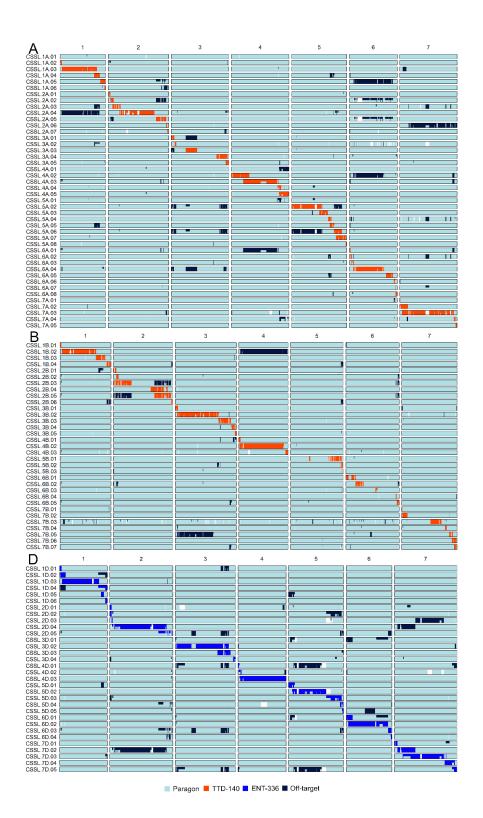
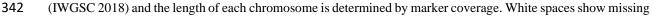




Figure 1. Graphical genotype selections for the A, B and D genome, using the NIAB_AB and NIAB_D
populations. A selection of 77 lines was used to form the A and B graphical genotype, originating from 62 unique
NIAB_AB CSSLs. The light blue colour represents a 'Paragon' like genotype, whereas orange represents a TTD140 introgression. Off-target introgressions are shown by navy blue. For the D graphical genotype, 32 lines were
selected using 25 unique NIAB_D CSSLs. The medium blue represents an introgression from the NIAB-SHW041
donor, which was formed using the *Ae. tauschii* ENT-336. The chromosomes are scaled on physical position



data and heterozygous regions are represented by divided colours. The figure was created using the R package
SelectionTools (v-19.1, www.uni-giessen.de).

345

346 *Phenotypic variation*

Variation was observed for every trait measured in comparison to the recurrent parent 347 'Paragon'. Trait summary statistics from the NIAB_AB glasshouse and NIAB_D field trial are 348 shown in **Table 1**. For grain length in both trials, and plant height in the NIAB_D field trial, 349 the mean of the CSSL lines was on average higher than 'Paragon' (Table 1). However, most 350 351 trait means for the CSSLs were lower than the recurrent parent, although individual CSSL lines had higher means than 'Paragon' for all traits. In the NIAB_AB glasshouse trial there was a 352 353 43.4% increase in grain yield per plant in the highest yielding CSSL line (compared to 'Paragon') and in NIAB_D field trial the highest TGW for a CSSL line gave a 32.5% increase 354 over 'Paragon'. For other traits, such as spikelet number and flowering time in NIAB D, there 355 were more moderate increases in the maximum CSSL mean compared to 'Paragon' (Table 1). 356 A significant genotypic effect (P < 0.001) was observed for all traits (**Table 1**). In 357 NIAB_D, moderate or high generalised heritability (H^2) was observed. The highest H^2 was 358 found for grain length ($H^2 = 0.91$) and flowering time, ear length and seed area also showed 359 high H^2 (**Table 1**). Estimates of H^2 were lowest for grain yield in the NIAB AB glasshouse 360 assessment ($H^2 = 0.39$), likely due to measurement on a per plant basis. 361

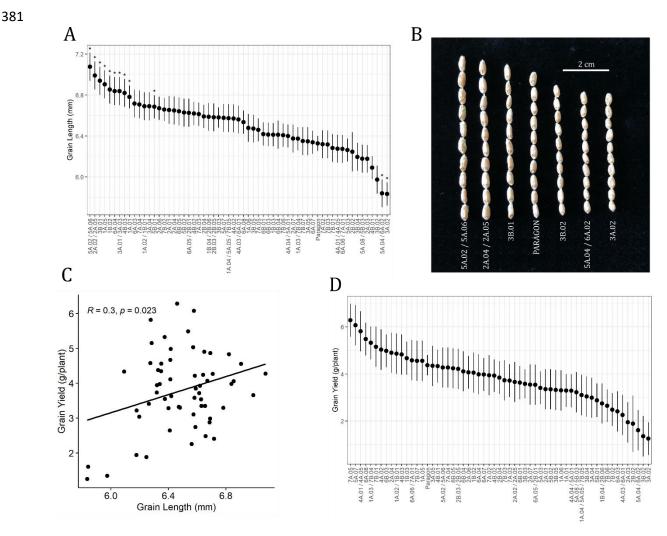
362

363Table 1. Summary statistics and results from two replicated trials: NIAB_AB glasshouse and NIAB_D field trial.**364**Results are taken from all introgressed lines included in both trials and the recurrent parent 'Paragon'. The best**365**linear unbiased estimates (BLUEs) are shown for 'Paragon' and the CSSLs that had the maximum and minimum**366**BLUE for each trait. The genotypic effect for each trait is shown through ANOVA, using the Satterthwaite's**367**degrees of freedom method. Generalized heritability (H^2) is shown for every trait. For the NIAB_D field trial, ear**368**and seed characteristics (including seed weight) are shown on a per ear basis, averaged from 10 technical replicates**369**per plot.

Trait NIAB_AB glassh	Paragon	CSSL min	CSSL mean	CSSL max	Num df	Den df	F	Р	H^2
Grain yield (g/plant)	4.38	1.06	3.74	6.28	143	161.36	1.67	< 0.001	0.39
Grain length (mm)	6.32	5.83	6.53	7.14	144	204.02	3.56	< 0.001	0.70
NIAB_D fiel	d trial								
Grain yield (85% DMC kg/plot)	6.13	4.90	5.89	6.73	33	55.85	3.07	< 0.001	0.68

Flowering time (Days from drilling to	189.13	183.99	189.05	191.75	33	54.33	8.34	< 0.001	0.88
GS65) Plant height (cm)	97.65	92.31	97.84	104.68	33	57.62	4.17	< 0.001	0.76
(index)	0.64	0.52	0.59	0.67	33	51.97	2.94	< 0.001	0.65
Spikelet no. (count)	22.29	20.22	21.57	22.93	33	59.68	6.51	< 0.001	0.83
Ear length (cm)	10.57	9.51	10.33	11.83	33	59.10	8.76	< 0.001	0.88
Ear weight (g)	2.95	2.38	2.77	3.51	33	56.52	2.81	< 0.001	0.64
Seed no. (no. ear ⁻¹)	58.62	42.81	56.43	63.64	33	57.46	5.88	< 0.001	0.82
Seed weight (g ear ⁻¹)	2.39	1.91	2.21	2.89	33	56.65	3.20	< 0.001	0.68
TGW (g)	40.56	31.69	39.10	53.74	33	59.21	7.66	<0.001	0.87
Grain length (mm)	6.63	6.35	6.70	7.33	33	64.46	11.22	<0.001	0.91
Grain width (mm)	3.43	3.13	3.35	3.65	33	59.68	5.09	< 0.001	0.80
Grain area (mm ²)	16.80	14.66	16.55	20.12	33	60.15	8.65	<0.001	0.88

In the NIAB AB glasshouse trial, significant variation was observed for grain length 370 (Table 1, Figure 2A). The majority of the CSSLs had a higher mean than 'Paragon'; compared 371 to the recurrent parent there was a 13.0% increase in the CSSL with the maximum grain length 372 (NIAB AB 5A.02/5A.06). For the NIAB AB selections, post hoc tests indicated several lines 373 with grain length significantly different to 'Paragon' (Figure 2A) and variation for a subset of 374 lines is shown in Figure 2B, clearly demonstrating visual variation. Grain length showed a 375 positive weak correlation with grain yield per plant (r = 0.3, P = 0.02; Figure 2C). Overall, 376 there was a significant genotypic effect observed for grain yield per plant (Table 1), although 377 the post hoc tests did not identify any CSSLs that differed significantly to 'Paragon' and 378 measurement on a per plant basis in the glasshouse is the likely cause of the large standard 379 error associated with the BLUEs (Figure 2D). 380



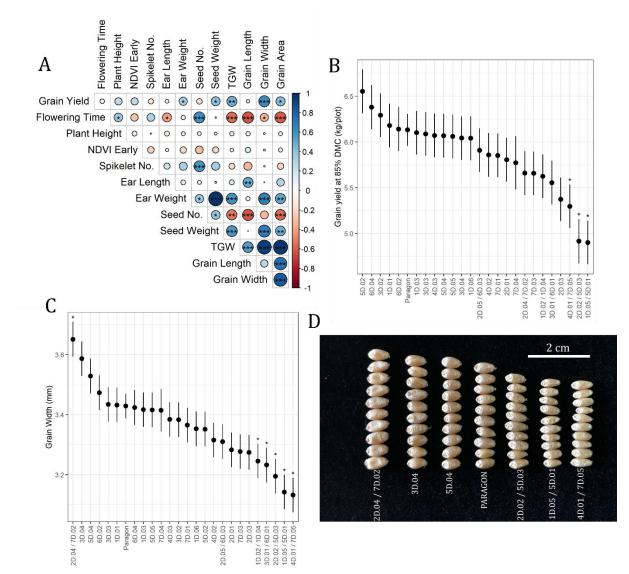
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Figure 2. Variation for grain length and yield per plant in the NIAB_AB glasshouse trial: A) Ranked best linear unbiased estimates (BLUEs) for seed length, with the NIAB_AB lines selected for the graphical genotype and the recurrent parent 'Paragon' shown. The asterisks above genotypes indicate where CSSLs were significantly different to 'Paragon' (P < 0.05). B) Variation in seed length observed with CSSLs with the longest and shortest seeds included, and 'Paragon' shown for comparison. C) The relationship between grain length and yield, with the Pearson correlation coefficient test statistics overlayed on the plot. D) NIAB_AB CSSLs and 'Paragon' ranked BLUEs for grain yield per plant. Error bars in A and D represent standard error of the BLUEs.

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Several traits showed a positive correlation with grain yield in the NIAB_D trial (Figure 3A), of which the positive correlation with grain width was the most significant (r = 0.61, P < 0.001; Figure 3A). Several significant negative trade-offs were also observed with flowering time having the most negative correlations with other traits (Figure 3A). Earlier flowering was negatively associated with ear length, TGW and grain length, width, and area. A significant negative relationship was also observed between seed number (per ear) and other grain size related traits (including TGW, seed length and area).

Grain yield was reduced in the majority of selected CSSLs compared to 'Paragon', 398 except in five CSSLs (Figure 3B). Lines selected for multiple introgressions across the D 399 genome gave some of the lowest yield performance. Post-hoc pairwise comparisons showed 400 that three of these were significantly lower than 'Paragon' (4D.01/7D.05, 2D.02/5D.03 and 401 1D.05/5D.01). None of the CSSLs had significantly higher grain yield than 'Paragon'. Several 402 NIAB D CSSLs showed significantly different means for grain width compared to 'Paragon': 403 five CSSLs had a lower predicted mean and a single CSSL had a higher mean (2D.04/7D.02, 404 Figure 3C). There were clear visual differences in grain width observed across the NIAB_D 405 406 CSSLs (Figure 3D). However, despite wide grain width CSSL.2D.04/7D.02 did not have a 407 higher grain yield than 'Paragon' (Figure 3B).



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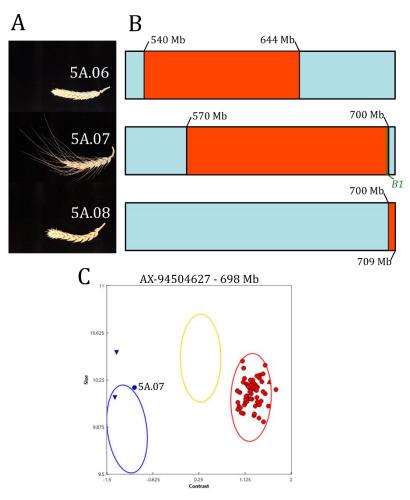
Figure 3. Trait correlations and variation for grain yield and grain width in the NIAB_D field trial: A) a Pearson's
correlation matrix calculated using the best linear unbiased estimates (BLUEs) from models fitted for each trait.

The heat scale bar shows the Pearson correlation coefficient (r). The asterisks within the matrix show the 412 significance threshold taken from the *P*-value for each comparison: *** = P < 0.001, ** = P < 0.01 and * = P < 0.05413 created using R package corrplot (Wei and Simko, 2017). B) Ranked BLUEs for grain yield and C) grain width 414 415 from a mixed linear model fitted via lme4 (Bates et al., 2015). Post-hoc pairwise comparisons of the BLUEs used 416 the package emmeans (Lenth 2020). The asterisks indicate significant difference to 'Paragon' (P < 0.05). All error 417 bars shown represent standard error of the predicted means. D) A photograph showing diversity in grain width using samples taken from plots in the NIAB_D field trial. The CSSLs with the three highest and lowest grain 418 419 widths are shown, with 'Paragon' included for comparison.

420

421 Introgression-trait association and validation

For the introgression-trait analysis, markers that were excluded during the earlier stages of QC 422 were included to provide further resolution on introgression boundaries. The presence of awns 423 was detected in a single NIAB AB line (5A.07; Figure 4A). The CSSLs with neighbouring 424 introgressions (5A.06 and 5A.08) were non-awned (Figure 4A). The awn inhibitor B1 has been 425 previously identified (TraesCS5A02G542800, 5A; 698,528,636 to 698,529,001bp; Huang et 426 al., 2020). The introgression from TTD-140 in 5A.07 overlapped this gene (570 Mb to 700 427 Mb), whereas in 5A.06 and 5A.08 the introgressions were in adjacent regions (Figure 4B). The 428 SNP marker AX-94504627 had a physical location (698,003,176 bp) close to the B1 locus. The 429 SNP cluster plot extracted from the Axiom Analysis Suite (Thermo Fisher Scientific) showed 430 that the only CSSL to carry the TTD-140 allele at this location was 5A.07. As shown in Figure 431 1, NIAB AB 5A.07 exhibits few off target introgressions on other chromosomes. No 432 introgressions were observed in the regions harbouring the other reported awn inhibitors 433 (Hooded locus (Hd)) on 4A and the awn inhibitor (Tipped 2) B2 on 6B (Mcintosh et al., 2013). 434 435



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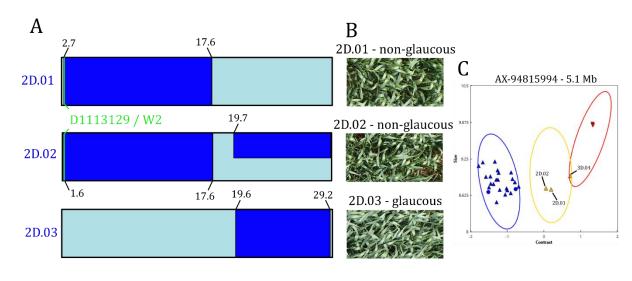
437 Figure 4. Introgression-trait association using the NIAB_AB population. A) Ears from the NIAB_AB glasshouse 438 trial, showing the single awned CSSL (5A.07). B) The awned CSSL had an introgression from TTD-140 on 5A 439 that extended over the B1 awn inhibitor locus (Huang et al., 2020) and awnless CSSLs 5A.06 and 5A.08 did not 440 have an introgression in this region. A TTD-140 introgression is represented by the colour orange, light blue 441 represents a 'Paragon' genotype. C) A genotyping plot taken from Axiom Analysis Suite (Thermo Fisher 442 Scientific) for a SNP close to the gene (AX-94504627; 698,003,176 bp) showing CSSLs from the AB graphical 443 genotype (circular points), two TTD-140 replicates (inverted triangle points) and 'Paragon' (a single triangle 444 point). The only line to share the TTD-140 allele (blue) is CSSL.5A.07. Physical positions are in mega base pairs 445 (Mb) and were taken from IWGSC (2018).

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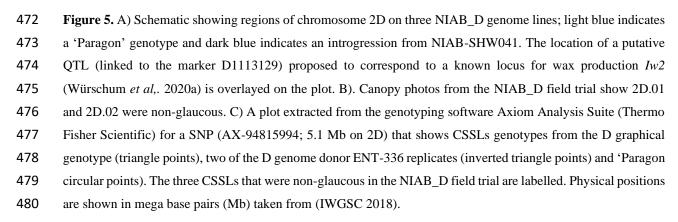
Three NIAB_D CSSLs displayed non-glaucous ear, stem and leaf phenotypes based on visual and RGB image assessment: 2D.01, 2D.02 and 3D.04. The CSSLs 2D.01 and 2D.02 were selected for NIAB-SHW041 introgressions at the start of 2D (**Figure 5A**), while 3D.04 was selected for a 3D introgression. However, 3D.04 also carried an off-target introgression at the start of 2D, from 1.6 to 13.7 Mb (**Figure 1**). The CSSL lines 2D.01 and 2D.02 carried an introgression on 2D from 2.7 to 17.6 Mb and 1.6 to 17.6 Mb, respectively. Additionally, 2D.02 had a downstream heterozygous introgression starting at around 19.7 Mb (**Figure 5A**). The

CSSL 2D.03 did not share the non-glaucous phenotype, which is visible in the RGB canopy 454 photographs of field plots for these genotypes (**Figure 5B**). The line 2D.03 had a homozygous 455 introgression from 19.6 to 29.2 Mb. The previously identified putative marker-trait association 456 with flag leaf glaucousness was based on a marker located at 2.9 Mb on 2D and hypothesized 457 to be *Iw2* (Würschum et al., 2020a), the D-genome paralog of *INHIBITOR of WAX1* (*Iw1*; 458 identified by Huang et al., 2017). All three NIAB D lines that were non-glaucous in the 459 graphical genotype carried an introgression overlapping this region, which can be observed for 460 2D.01 and 2D.02 in Figure 5A. A SNP marker located at 5.1 Mb also confirmed that the three 461 462 non-glaucous CSSLs did not possess the 'Paragon' allele close to the marker-trait association (Figure 5C). The introgressions in these lines at the start of 2D were typically homozygous 463 (Figure 1). However, the genotype calls at AX-94815994 are heterozygous (Figure 5C). It is 464 likely that the calls for the lines were homozygous for the ENT-336 (D-genome) allele but 465 differences in ploidy impacting SNP calling between the CSSLs and the Ae. tauschii donor has 466 467 separated the cluster. Whole genome graphical genotypes were inspected visually and the CSSLs 2D.01, 2D.02 and 3D.04 did not share any other overlapping introgressions. 468









481 Discussion

We report the creation of a genetic resource for detecting and exploiting wheat progenitor 482 variation. Publicly available pure seed, phenotyping and genotyping data support use as 483 introgression donors and/or near-isogenic lines. The introgressions are genetically tractable, of 484 reasonable size for forward use and have low off-target segments. This builds on early work to 485 develop wheat resources for gene localisation and characterisation (Sears 1953 & 1954) and 486 genetic mapping (Zemetra et al., 1986). Since this time numerous forward and reverse wheat 487 genetic resources have been developed and the reported wheat CSSL series adds a further 488 489 precision resource for exploiting progenitor diversity.

The NIAB_AB and NIAB_D series were created using backcrossing and MAS 490 followed by high-density genotyping to facilitate progenitor segment introgression with low 491 off-target effects. This approach is relatively time and cost intensive. Different methods of 492 population construction have been proposed, including those with a lower marker requirement 493 494 (e.g. Bian *et al.* (2010) used 8 informative markers at BC_3F_4 followed by pooled genotyping to extract individuals with small donor segments). This reduced genotyping approach is an 495 496 alternative, low-cost approach to support CSSL development although larger numbers of individuals need to be progressed through backcrossing. Future development of CSSL series is 497 498 likely to benefit from the reducing costs of genotyping, allowing more intensive screening and enhancing both accuracy of donor introgression and background recovery. The reducing cost 499 500 of whole genome skim sequencing is also likely to allow higher resolution of introgression 501 segments.

502 In both the NIAB AB and NIAB D series, we detected significant trait variation and transgressive segregation compared to 'Paragon'. Grain size and shape are largely independent 503 504 traits and we observed strong variation for grain length and width. Grain length of ancestral wheat has been shown to be generally higher than in elite bread wheat (Gegas et al., 2010) and 505 506 selection of alleles that confer increased grain length could increase grain size. Brinton et al. (2017) identified a significant QTL for TGW on chromosome 5A (*Qtgw-cb.5A*) demonstrating 507 that the increase in grain area (driving TGW) was predominantly conferred by an increase in 508 grain length. Yang et al. (2019) also identified a novel allele associated with increased grain 509 510 length on 5AL (cloning TaGL3-5A-G). This allele, found in ancestral wheats, represents a breeding target, particularly if it could be pyramided in combination with favourable alleles for 511 grain width (e.g. TaGW2; Simmonds et al., 2016) and grain size (TaGS5-3A; Ma et al., 2015). 512 In the NIAB_AB population, there was a significant positive effect of grain yield per plant 513 correlated with grain length. This identified individual CSSL lines with introgressions 514

associated with increased grain length which are effectively near-isogenic lines for the TTD-515 140 segment. The introgression in the lines with the greatest increase in grain length is in the 516 same region as TaGL3-5A (Yang et al., 2019) and future work could establish whether it 517 contains the favourable allele described by Yang et al (2019). These lines could be field tested 518 to validate the effect, and the segments transferred to additional elite backgrounds to test 519 520 stability. In addition, the extreme positive and negative (compared to 'Paragon') lines could be used in a bulk-segregant genotyping approach (e.g. based on exome sequencing; Martinez et 521 al., 2020 or RNA-seq; Zhu et al., 2020) to determine the specific grain length controllers. 522

523 For the NIAB_D population, significant trait variation and high trait heritabilities were observed. The significant genotypic effects, despite the high recovery of 'Paragon' 524 background, indicate effective capture (and contribution) of functional variation from the Ae. 525 tauschii donor ENT-336. Variation was observed for grain yield although was only significant 526 for CSSLs with yields lower than 'Paragon'. Although not statistically significant, five CSSLs 527 yielded higher than 'Paragon'. Further testing through multi-site trials could establish if there 528 is a significant yield effect associated with these CSSLs. Grain width was highly correlated 529 530 with yield. This allowed identification of CSSLs with positive and negative effects for grain width which are available as near-isogenic lines, introgression segment donors and for further 531 532 genetic characterisation, as above. Grain width has been well characterised in hexaploid wheat (Simmonds et al., 2016) and alternative alleles for TaGW2 can increase grain width and 533 therefore grain weight, though phenotypes may be subtle due to allele dosage effects. Previous 534 GWAS analysis of Ae. tauschii collections have identified numerous QTL for grain 535 characteristics conferred by genetic effects on multiple chromosomes (Arora et al., 2017; Zhao 536 et al., 2021). The further development of markers for the regions underlying these QTLs will 537 facilitate the validation of allelic effects and potential use for breeding. 538

In addition to identification of lines with positive and negative introgression effects, 539 specific trait-introgression effects could be assessed based on physical locations of the 540 segments. Although the series can't be used directly for mapping *per se* due to the low power 541 of detection it can be used to associate introgressions with phenotypes of interest. This is useful 542 to validate previously observed effects as well as being a starting point for fine mapping. Here, 543 544 our definition of introgression segment boundaries is limited to the coverage of Axiom® Wheat Breeder's Genotyping Array (Allen et al., 2017) but further genotyping would allow higher 545 resolution and definition of boundaries and allow full automation of the process (array data 546 requires manual curation below software quality thresholds). However, the current resolution 547 is likely to be sufficient for transfer of segments via MAS. 548

The recurrent parent 'Paragon' carries the B1 awn inhibition locus. Using the 549 NIAB_AB population, and through replacement of the region with the wild type (recessive) 550 progenitor allele via introgression we confirmed the location of the locus. This was near the 551 edge of the introgression boundary but could be reliably detected and linked to the phenotype. 552 The *Tipped 1* (B1) mutant is one of the three major loci linked to awn presence in wheat 553 (Watkins and Ellerton, 1940; Sourdille et al., 2002; Yoshioka et al., 2017) and has been 554 identified as an important determinant of awn development (Mackay et al., 2014; Würschum 555 et al., 2020b). The B1 locus has been fine mapped in multiple studies to 5AL (DeWitt et al., 556 557 2020; Huang et al., 2020; Niu et al., 2020; Wang et al., 2020; Würschum et al., 2020b). Awn presence and the B1 locus have been recently linked to an offset in the negative association 558 between grain yield and protein content (Scott et al., 2021). The detection of the awn inhibitor 559 segment demonstrates the use of the resource for categorical traits and could be used to further 560 understand awn and linked-awn trait haplotype contributions from T. diccocoides and expedite 561 their transfer to breeding. CSSLs with and without awns could also be used to further 562 understand the mechanisms of the recently reported trade-offs between awns, quality, and 563 564 nutritional traits (Scott et al., 2021).

In the NIAB D population we demonstrate the use of trait-introgression association to 565 566 validate a putative association with a categorical trait contributed by the progenitor Ae. tauschii. This used a step-by-step statistical approach enabling validation of a marker-trait association 567 for glaucousness which fell slightly below the significance threshold in Würschum et al. 568 (2020a). Our approach aligned the glaucous phenotype of the CSSL introgression segment with 569 570 the physical position of the peak marker reported by Würschum et al. (2020a). In addition to confirming the genetic effect, this provides a 35K Axiom marker to facilitate MAS and tracking 571 572 in breeding, adding to the previously identified DArTseq peak marker from Würschum et al. (2020a). This is likely to be advantageous given the relative ease of conversion of array probes 573 compared to sequence-based probe conversion (Burridge et al., 2018). 574

Here we report the development, characterization, and initial testing of a CSSL wheat 575 series as a tractable genetic resource adding to the existing portfolio of functional genomics 576 resources for wheat improvement. The use of 'Paragon' as a genetic background complements 577 578 other available wheat resources including the A.E. Watkins hexaploid wheat nested association mapping panel, and EMS and γ mutant populations (Wingen *et al.*, 2017), QTL datasets 579 (Przewieslik-Allen et al., 2021), and publicly available synthetic hexaploid wheat populations 580 (Adamski et al., 2020). Further work is required to reduce off-target introgressions and to split 581 582 multiple introgression lines into discreet single-introgression lines. The application of higher density markers and/or sequencing will support further precision in defining introgression
boundaries. The extreme phenotypic lines can be further validated as near isogenic lines and
used as donor lines for transfer of introgressions to additional elite parents for use in breeding.
The NIAB_AB and NIAB_D lines can also be intercrossed to create favourable introgression
complements (e.g. grain length and grain width) for further testing and development of targeted
multi-introgression lines for specific traits of interest. Overall, the resource reported here
increases access to wheat genomic resources for research and breeding improvements.

590

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598

599 Conflict of interest

600 The authors declare no conflict of interest.

601

602 Author contributions

P.H., C.U., K.J.E. and A.R.B. conceived the project; R.H., F.J.L, T.I.C.W., A.R.B designed
experiments; R.H., F.J.L, A.J.B., A.L., A.M.P.-A. performed the experiments; F.J.L, T.I.C.W.,
P.H. and A.R.B analysed experiments, R.H., F.J.L, T.I.C.W. and A.R.B wrote the manuscript
with inputs from all other authors.

607

608 Data availability

Seed of the germplasm reported here is directly available from the Germplasm Resources Unit
at the John Innes Centre (via <u>https://www.seedstor.ac.uk/</u>), including the parental line
'Paragon' (WCSSL0001), donor lines TTD-140 (WCSSL0002) and NIAB-SHW041
(WCSSL0003) and for the NIAB_AB (WCSSL0004 to WCSSL0080) and NIAB_D
(WCSSL0081 to WCSSL0112) populations.

614 The Axiom® Wheat Breeder's Genotyping Array data is available for direct download for each615 of the three genomes here:

616	https://www.cerealsdb.uk.net/cerealgenomics/CerealsDB/genotyping_data/NIAB_CSSL_Age
617	nome.csv
618	https://www.cerealsdb.uk.net/cerealgenomics/CerealsDB/genotyping_data/NIAB_CSSL_Bge
619	nome.csv
620	https://www.cerealsdb.uk.net/cerealgenomics/CerealsDB/genotyping_data/NIAB_CSSL_Dge
621	nome.csv
622	Plant images from the NIAB_AB glasshouse trial are available from:
623	https://opendata.earlham.ac.uk/wheat/under_license/toronto/Horsnell,Leigh,Bentley,Wright_
624	2020_NIAB_CSSL_D_genome_yield_trial_H2020/
625	Plot images from the NIAB_D field trial are available from:
626	https://opendata.earlham.ac.uk/wheat/under_license/toronto/Horsnell,Leigh,Bentley,Wright_
627	2020_NIAB_CSSL_D_genome_yield_trial_H2020/
628	Data from the NIAB_D field trial are available from:
629	https://grassroots.tools/fieldtrial/study/61faaf25c68884365e7bcc34
630	
631	References
632	Adamski NM, Borrill P, Brinton J, et al, (2020) A roadmap for gene functional characterisation
633	in crops with large genomes: Lessons from polyploid wheat. eLife 9:e55646
634	doi:10.7554/eLife.55646
635	
636	Ali ML, Sanchez PL, Yu S et al. (2010) Chromosome Segment Substitution Lines: a powerful
637	tool for the introgression of valuable genes from Oryza wild species into cultivated rice (O.
638	sativa). Rice 3: 218–234. https://doi.org/10.1007/s12284-010-9058-3
639	
640	Allen AM, Winfield MO, Burridge AJ, et al (2017) Characterization of a Wheat Breeders'
641	Array suitable for high-throughput SNP genotyping of global accessions of hexaploid bread
642	wheat (Triticum aestivum). Plant Biotechnology Journal 15: 390-401.
643	https://doi.org/10.1111/pbi.12635
644	
645	Arora S, Singh N, Kaur S, Bains NS, Uauy C, Poland J, Chhuneja P (2017) Genome-wide
646	association study of grain architecture in wild wheat Aegilops tauschii. Frontiers in Plant
647	<i>Science</i> 8: 886.

648

649	Balakrishnan D, Surapaneni M, Mesapogu S, Neelamraju S (2019) Development and use of
650	chromosome segment substitution lines as a genetic resource for crop improvement.
651	Theoretical and Applied Genetics 132: 1-25. doi:10.1007/s00122-018-3219-y
652	
653	Basava RK, Hash CT, Mahendrakar MD, Kishor KPB, Satyavathi CT, Kumar S, Singh RB,
654	Yadav RS, Gupta R, Srivastava RK (2019) Discerning combining ability loci for divergent
655	environments using chromosome segment substitution lines (CSSLs) in pearl millet. PLoS
656	ONE 14: e0218916. https://doi.org/ 10.1371/journal.pone.0218916
657	
658	Bates D, Mächler M, Bolker B, Walker S (2015) Fitting linear mixed-effects models using
659	lme4. J Stat Softw 67 https://doi.org/10.18637/jss.v067.i01
660	
661	Berke TG, Baenziger PS, Morris WR (1992) Chromosomal location of wheat quantitative trait
662	loci affecting agronomic performance, using reciprocal chromosome substitutions. Crop
663	Science 32: 621–627.
664	
665	Bian JM, Jiang L, Liu LL, Wei XJ, Xiao YH, Zhang LJ, et al. (2010) Construction of a new set
666	of rice chromosome segment substitution lines and identification of grain weight and related
667	traits QTLs. Breeding Science 60: 305–13.
668	
669	Bian X, Tyrrell S, Davey RP (2017) The Grassroots life science data infrastructure.

- 670 <u>https://grassroots.tools</u>
- 671
- Blanco A, Simeone R, Gadaleta A (2006) Detection of QTLs for grain protein content in durum
 wheat. *Theoretical and Applied Genetics* 112: 1195–1204.
- 674
- Brinton J, Simmonds J, Minter F, Leverington-Waite M, Snape J, Uauy C (2017) Increased
- 676 pericarp cell length underlies a major quantitative trait locus for grain weight in hexaploid
- 677 wheat. *New Phytologist* 215: 1026-1038.
- 678
- 679 Burridge A, Wilkinson P, Winfield M, Barker G, Przewieslik-Allen S, Coghill J, Waterfall C,

Edwards K (2018) Conversion of array-based single nucleotide polymorphic markers for use

- 681 in targeted genotyping by sequencing in hexaploid wheat (Triticum aestivum). Plant
- 682 Biotechnology Journal 16: 867-876.

683	
684	Camacho C, Coulouris G, Avagyan V, Ma N, Papdopoloulos J, Bealer K, Madden TL (2009)
685	BLAST+: architecture and applications. BMC Bioinformatics 10, 421.
686	https://doi.org/10.1186/1471-2105-10-421
687	
688	Chitwood DH, Kumar R, Headland LR, Ranjan A, Covington MF et al. (2013) A quantitative
689	genetic basis for leaf morphology in a set of precisely defined tomato introgression lines. Plant
690	<i>Cell</i> 25: 2465–2481.
691	
692	Cox TS (1997) Deepening the wheat gene pool. Journal of Crop Production 1: 1-25. doi:
693	10.1300/J144v01n01_01.
694	
695	Cullis BR, Smith AB, Coombes NE (2006) On the design of early generation variety trials with
696	correlated data. Journal of Agricultural Biology and Environmental Statistics 11: 381-393.
697	https://doi.org/10.1198/108571106X15443.
698	
699	DeWitt, N., Guedira, M., Lauer, E., Sarinelli, M., Tyagi, P., Fu, D., et al. (2020) Sequence-
700	based mapping identifies a candidate transcription repressor underlying awn suppression at the
701	B1 locus in wheat. New Phytologist, 225, 326–339.
702	
703	Eshed Y, Abu-Abied M, Saranga Y Zamir D (1992) Lycopersicon esculentum lines containing
704	small overlapping introgressions from L. pennellii. Theoretical and Applied Genetics 83:
705	1027–1034.
706	
707	Eshed Y, Zamir D (1994) A genomic library of Lycopersicon pennellii in L. esculentum: a tool
708	for fine mapping of genes. Euphytica 79: 175–79.
709	
710	Eshed Y Zamir D (1995) An introgression line population of Lycopersicon pennellii in the
711	cultivated tomato enables the identification and fine mapping of yield-associated QTL.
712	Genetics 141: 1147–1162.
713	
714	Evans LT, Bingham J, Jackson P, Sutherland J (1972) Effect of awns and drought on the supply
715	of photosynthate and its distribution within wheat ears. Annals of Applied Biology, 70, 67–76.
716	

- 717 Falke KC, Miedaner T, Frisch M (2009) Selection strategies for the development of rye
- introgression libraries. *Theoretial and Applied Genetics* 119: 595–603.
- 719
- Feldman M, Millet E (1995) Methodologies for identification, allocation and transfer of
- quantitative genes from wild emmer into cultivated wheat. In: Li ZS, Xin ZY (eds) Proceedings
- of the 8th international wheat genetics symposium, Beijing, China, pp 19–27.
- 723
- Fletcher RS, Mullen JL, Yoder S *et al.* (2013) Development of a next-generation NIL library
 in *Arabidopsis thaliana* for dissecting complex traits. *BMC Genomics.* 14: 655.
 doi:10.1186/1471-2164-14-655.
- 727
- 728 Fonceka D, Tossim HA, Rivallan R, et al. (2012) Construction of chromosome segment
- substitution lines in peanut (*Arachis hypogaea* L.) using a wild synthetic and QTL mapping for
- plant morphology. *PLoS One* 7: e48642. doi:10.1371/journal.pone.0048642
- 731
- Fridman E, Carrari F, Liu Y-S, Fernie AR, Zamir D (2004) Zooming in on a quantitative trait
 for tomato yield using interspecific introgressions. *Science* 17: 1786-9.
- 734
- Fu YB, Somers DJ (2009) Genome-wide reduction of genetic diversity in wheat breeding. *Crop Science* 49: 61–168.
- 737
- Fulop D, Ranjan A, Ofner I, Covington MF, Chitwood DH, West D, Ichihashi Y, Headland L,
 Zamir D, Maloof JN, Sinha NR (2016) A new advanced backcross tomato population enables
 high resolution leaf QTL mapping and gene identification. *G3: Genes, Genomes, Genetics* 6:
 3169-3184.
- 742
- Fulton TM, Chunwongse J, Tanksley SD (1995) Microprep protocol for extraction of DNA
 from tomato and other herbaceous plants. *Plant Molecular Biology Reporter* 13: 207–209.
- 745
- Gaurav K, Arora S, Silva P *et al.* (2021) Population genomic analysis of *Aegilops tauschii*identifies targets for bread wheat improvement. *Nature Biotechnology*https://doi.org/10.1038/s41587-021-01058-4
- 749

- Gegas VC, Nazari A, Griffiths S, Simmonds J, Fish L, Orford S, Sayers L, Doonan JH, Snape
 JW (2010) A genetic framework for grain size and shape variation in wheat. *The Plant Cell* 22:
 1046–1056,
- 753
- Grewal S, Hubbart-Edwards S, Yang C, et al. (2020) Rapid identification of homozygosity and
 site of wild relative introgressions in wheat through chromosome-specific KASP genotyping
 assays. *Plant Biotechnology Journal* 18: 743-755. doi:10.1111/pbi.13241
- 757
- Gross BL, Olsen KM (2010) Genetic perspectives on crop domestication. *Trends in Plant Science* 15: 529–537.
- 760
- Gu L, Wei B, Fan R, Jia X, Wang X, Zhang X (2015) Development, identification and
 utilization of introgression lines using Chinese endemic and synthetic wheat as donors. *Journal*
- 763 *of Integrative Plant Biology* 8: 688–697
- 764
- Holtan HEE, Hake S (2003) Quantitative trait locus analysis of leaf dissection in tomato using *Lycopersicon pennellii* segmental introgression lines. *Genetics* 165: 1541-1550.
- 767
- Howell PM, Lydiate DJ, Marshall DF (1996). Towards developing intervarietal substitution
 lines in *Brassica napus* using marker-assisted selection. *Genome* 39: 348-358.
- 770
- Huang D, Feurtado A, Smith MA, Flatman LK, Koh C, Cutler AJ (2017) Long noncoding
 miRNA gene represses wheat β-diketone waxes. *Proceedings of the National Academy of Sciences* 114: E3149-E3158.
- 774
- Huang D, Zheng Q, Melchkart T, Bekkaoui Y, Konkin DJF, Kagale S et al. (2020) Dominant
 inhibition of awn development by a putative zinc-finger transcriptional repressor expressed at
 the *B1* locus in wheat. *New Phytologist*, 225, 340–355.
- 778
- International Wheat Genome Sequencing Consortium (IWGSC) (2018) Shifting the limits in
 wheat research and breeding using a fully annotated reference genome. *Science*. 17: 361
 (6403): eaar7191. doi: 10.1126/science.aar7191.
- 782

783	Jie C, Bughio H, Chen DZ, Liu G, Zheng K, Zhuang J (2006) Development of chromosomal
784	segment substitution lines from a backcross recombinant inbred population of interspecific rice
785	cross. Rice Science 13: 15–21.
786	
787	Joppa LR, Cantrell RG (1990) Chromosomal location of genes for grain protein content of wild
788	tetraploid wheat. Crop Science 30: 1059-1064.
789	
790	Joppa LR, Du C, Hart GE, Hareland GA (1997) Mapping gene(s) for grain protein in tetraploid
791	wheat (Triticum turgidum L.) using a population of recombinant inbred chromosome lines.
792	Crop Science 37: 1586-1589.
793	
794	Keurentjes JJB, Bentsink L, Alonso-Blanco C, Hanhart CJ, Vries HB-D, et al. (2007)
795	Development of a near-isogenic line population of Arabidopsis thaliana and comparison of
796	mapping power with a recombinant inbred line population. Genetics 175: 891–905.
797	
798	Kishii M (2019) An update of recent use of Aegilops species in wheat breeding. Frontiers in
799	Plant Science 10:585. doi: 10.3389/fpls.2019.00585
800	
801	Kumari BR, Kolesnikova-Allen MA, Hash CT, Senthilvel S, Nepolean T, KaviKishor PB,
802	Riera-Lizarazu O, Witcombe JR, Srivastava RK (2014) Development of a set of chromosome
803	segment substitution lines in pearl millet [Pennisetum glaucum (L.) R. Br.]. Crop Science 54:
804	2175–2182.
805	
806	Kuspira J, Unrau J (1957) Genetic analysis of certain characters in common wheat using whole
807	chromosome substitution lines. Canadian Journal of Plant Sciences 37: 300-326.
808	
809	Kuznetsova A, Brockhoff PB, Christensen RHB (2017) lmerTest package: tests in linear mixed
810	effects models. Journal of Statistics Software 82: https://doi.org/10.18637/jss.v082.i13
811	
812	Law CN (1966) Biometrical analysis using chromosome substitutions within a species. P59-85
813	In R. Riley and KR Lewis (ed.) Chromosome manipulations in plant genetics. Plenum Press,
814	New York.
815	

- Leigh FJ, Wright TIC, Horsnell RA, Dyer S, Bentley AR (2022) Progenitor species hold untapped diversity for potential climate-responsive traits for use in wheat breeding and crop improvement. *Heredity*
- 819
- Lenth RV (2020) emmeans: estimated marginal means, aka least-squares means. R package
 version 1.5.3. https://CRAN.R-project.org/package=emmeans.
- 822
- Martinez SA, Shorinola O, Conselman S. *et al.* (2020). Exome sequencing of bulked segregants
 identified a novel TaMKK3-A allele linked to the wheat ERA8 ABA-hypersensitive
 germination phenotype. *Theoretical and Applied Genetics* 133: 719–736.
- 826
- Li X, Wang W, Wang Z, Li K, Lim YP, Piao Z (2015). Construction of chromosome segment substitution lines enables QTL mapping for flowering and morphological traits in *Brassica*
- *rapa. Frontiers in Plant Science* 6:432. doi:10.3389/fpls.2015.00432
- 830
- Luo D, Ganesh S, Koolaard J (2020). Predictmeans: calculate predicted means for linear
 models. R package version 1.0.4. https://CRAN.R-project.org/package=predictmeans.
- 833
- Lippman ZB, Semel Y, Zamir D (2007) An integrated view of quantitative trait variation using
 tomato interspecific introgression lines. *Current Opinions in Genetics and Development* 17:
 545-552.

837

- Mcintosh R, Dubcovsky J, Rogers WJ, Morris *et al.* (2013) Catalogue of gene symbols for
 wheat. In 12th International Wheat Genetics Symposium, Yokohama, Japan.
- 840
- Ma L, Li T, Hao C, Wang Y, Chen X, Zhang X (2016) *TaGS5-3A*, a grain size gene selected
 during wheat improvement for larger kernel and yield. *Plant Biotechnology Journal*. 14: 1269-
- 843 1280. https://doi.org/10.1111/pbi.12492
- 844
- 845 Mackay IJ, Bansept-Basler P, Barber T, Bentley AR, Cockram J, Gosman N, Greenland AJ,
- Horsnell R, Howells R, O'Sullivan D, Rose GA, Howell P (2014) An eight-parent Multiparent
- 847 Advanced Generation Inter-Cross population for winter-sown wheat: creation, properties and
- validation. *G3: Genes, Genomes, Genetics* 4:1603-1610.
- 849

- 850 Millet E, Rong J-K, Qualset C, Mcguire PE, Bernard A, Sourdille P, Feldman M (2014) Grain
- yield and grain protein percentage of common wheat lines with wild emmer chromosome-arm
- substitutions. *Euphytica*. 195. 10.1007/s10681-013-0975-2.
- 853
- Milne I, Shaw P, Stephen G, et al. (2010) Flapjack-graphical genotype visualization.
- *Bioinformatics* 26: 3133–3134. https://doi.org/10.1093/bioinformatics/btq580
- 856
- Nevo E (2014) Evolution of wild emmer wheat and crop improvement. *Journal of Systematics and Evolution* 52: 673-696.
- 859
- Nie X, Tu J, Wang B, Zhou X, Lin Z (2015) A BIL population derived from *G. hirsutum* and
- *G. barbadense* provides a resource for cotton genetics and breeding. *PLoS ONE* 10: e0141064.
- 862 https://doi.org/10.1371/journal.pone.0141064
- 863
- Olmos S, Distelfeld A, Chicaiza O, Schlatter AR, Fahima T, Echenique V, Dubcovsky J (2003)
 Precise mapping of a locus affecting grain protein content in durum wheat. *Theoretical and Applied Genetics* 107: 1243-1251.
- 867
- Paradis E, Schliep K (2018) Ape 5.0: an environment for modern phylogenetics and
 evolutionary analyses in R. *Bioinformatics* 35: 526-528.
- 870
- Przewieslik-Allen A, Wilkinson PA, Burridge A, Winfield M, Dai X, Beaumont M, King J,
 Yang C, Griffiths S, Wingen L, Horsnell R, Bentley AR, Shewry P, Barker GLA, Edwards KJ
 (2021) The role of gene flow and chromosomal instability in shaping the bread wheat genome. *Nature Plants* 7: 172-183.
- 875
- Qiao W, Qi L, Cheng Z, Su L, Li J, Yan S, Ren J, Zheng Z, Yang Q (2016) Development and
 characterization of chromosome segment substitution lines derived from *Oryza rufipogon* in
 the genetic background of *O. sativa* spp. *indica* cultivar 9311. *BMC Genomics* 17: 580.
 https://doi.org/10.1186/s12864-016-2987-5.
- 880
- R Core Team (2020). R: A language and environment for statistical computing. R Foundation
 for Statistical Computing, Vienna, Austria. URL https://www.R-project.org/.
- 883

- Rae AM, Howell EC, Kearsey MJ (1999) More QTL for flowering time revealed by 884 substitution lines in Brassica oleracea. Heredity 83: 586-596. doi:10.1038/sj.hdy.6886050 885 886 Ramsay LD, Jennings DE, Bohuon EJR, Arthur AE, Lydiate DJ et al. (1996) The construction 887 of a substitution library of recombinant backcross lines in Brassica oleracea for the precision 888 889 mapping of quantitative trait loci. Genome. 39: 558-567. 890 Rodriguez-Alvarez MX, Boer MP, van Eeuwijk FA, Eilers PHC (2018) Correcting for spatial 891 892 heterogeneity in plant breeding experiments with P-splines. Spatial Statistics 23:52-71 https://doi.org/10.1016/j.spasta.2017.10.003 893 894 Scheipl F, Greven S, Kuechenhoff H (2008) Size and power of tests for a zero random effect 895 variance or polynomial regression in additive and linear mixed models. Computational 896 897 Statistics and Data Analysis, 52:3283-3299. 898 Scott MF, Ladejobi O, Amer S et al. (2020) Multi-parent populations in crops: a toolbox 899 integrating genomics and genetic mapping with breeding. *Heredity* 125: 396-416. 900 901 Scott MF, Fradgley N, Bentley AR, Brabbs T, Corke F, Gardner KA et al. (2021) Limited 902 903 haplotype diversity underlies polygenic trait architecture across 70 years of wheat breeding. Genome Biology, 22, 137. 904 905 906 Sears ER (1953) Nullisomic analysis in common wheat. American Naturalist 87: 245-252. 907 Sears ER (1954) The aneuploids of common wheat. Research Bulletin Missouri Agricultural 908 909 Experiment Station 572: 1-59. 910 Simmonds J, Scott P, Brinton J, Mestre TC, Bush M, Del Blanco A, Dubcovsky J, Uauy C 911 (2016) A splice acceptor site mutation in TaGW2-A1 increases thousand grain weight in 912 913 tetraploid and hexaploid wheat through wider and longer grains. Theoretical and Applied Genetics 129:1099-1112. 914 915 Sourdille P, Cadalen T, Gay G, Gill B, Bernard M. (2002) Molecular and physical mapping of 916
- genes affecting awning in wheat. *Plant Breeding* 121, 320–324.

918	
919	Stebbins GL (1950) Variation and Evolution in Plants. Columbia University Press, New York,
920	USA.
921	
922	Tian F, Li DJ, Fu Q, Zhu ZF, Fu YC, Wang XK, Sun CQ (2006) Construction of introgression
923	lines carrying wild rice (Oryza rufipogon Griff.) segments in cultivated rice (Oryza sativa L.)
924	background and characterization of introgressed segments associated with yield-related traits.
925	Theoretical and Applied Genetics 112: 570–580.
926	
927	Uauy C, Brevis JC, Dubcovsky J (2006a) The high grain protein content gene Gpc-B1
928	accelerates senescence and has pleiotropic effects on protein content in wheat. Journal of
929	Experimental Botany 57: 2785-94.
930	
931	Uauy C, Distelfeld A, Fahima T, Blechl A, Dubcovsky J (2006b) A NAC gene regulating
932	senescence improves grain protein, zinc, and iron content in wheat. Science 314:1298-301.
933	
934	Unrau J, Person C, Kuspira J (1956) Chromosome substitution in hexaploid wheat. Canadian
935	Journal of Botany 34: 629–640.
936	
937	Urrutia M, Bonet J, Arús P, Monfort A (2015) A near-isogenic line (NIL) collection in diploid
938	strawberry and its use in the genetic analysis of morphologic, phenotypic and nutritional
939	characters. Theoretical and Applied Genetics 128:1261-1275. doi:10.1007/s00122-015-2503-
940	3.
941	
942	Walkowiak S, Gao L, Monat C et al. (2020) Multiple wheat genomes reveal global variation
943	in modern breeding. <i>Nature</i> 588 : 277–283 https://doi.org/10.1038/s41586-020-2961-x
944	
945	Wang D, Yu K, Jin D, Sun L, Chu J, Wu W et al. (2020) Natural variations in the promoter of
946	Awn Length Inhibitor 1 (ALI-1) are associated with awn elongation and grain length in common
947	wheat. The Plant Journal 101; 1075–1090.
948	
949	Watkins AE, Ellerton S (1940) Variation and genetics of the awn in <i>Triticum</i> . Journal of
950	<i>Genetics</i> 40: 243–270.
951	

- Wei T, Simko, V (2017) R Package "Corrplot": visualization of a correlation matrix. Available
 online: https://github.com/taiyun/corrplot
- 954
- Wilkinson PA, Winfield MO, Barker GLA, Tyrell S, Bian X *et al.* (2016) CerealsDB 3.0:
 expansion of resources and data integration. *BMC Bioinformatics doi*.org/10.1186/s128590161139-x
- 958
- Wingen LU, West C, Leverington-Waite M, Collier S, Orford S, Goram R, Yang CY, King J,
 Allen AM, Burridge A, Edwards KJ, Griffiths S (2017) Wheat landrace genome diversity. *Genetics*. 205: 1657-1676.
- 962
- Wu J, Jenkins JN, McCarty JC, Saha S, Stelly DM (2006) An additive-dominance model to
 determine chromosomal effects in chromosome substitution lines and other gemplasms. *Theoretical and Applied Genetics* 112: 391–399.
- 966
- Würschum T, Langer SM, Longin CFH, Tucker MR, Leiser WL (2020a) Refining the genetic
 architecture of flag leaf glaucousness in wheat. *Theoretical and Applied Genetics*, 133, 981–
 969 991.
- 970

Würschum T, Jähne F, Phillips AL, Langer SM, Longin CFH, Tucker MR, Leiser WL (2020b)
Misexpression of a transcriptional repressor candidate provides a molecular mechanism for the
suppression of awns by *Tipped 1* in wheat. *Journal of Experimental Botany*, 71, 3428–3436.

974

- 978
- Yang J, Zhou Y, Wu Q. *et al.* (2019) Molecular characterization of a novel *TaGL3-5A* allele
 and its association with grain length in wheat (*Triticum aestivum* L.). *Theoretical and Applied Genetics* 132:1799–1814. https://doi.org/10.1007/s00122-019-03316-1
- 982

Yoshioka M, Iehisa JCM, Ohno R, Kimura T, Enoki H, Nishimura S *et al.* (2017) Three
dominant awnless genes in common wheat: Fine mapping, interaction and contribution to
diversity in awn shape and length. *PLoS ONE*, 12: 1–21.

<sup>Yu J, Holland JB, McMullen MD, Buckler ES (2008) Genetic design and statistical power of
nested association mapping in maize.</sup> *Genetics* 178: 539-551.
doi:10.1534/genetics.107.074245

986	

987	Zemetra RS, Morris WR, Schmidt JW (1986) Gene locations for heading date using reciprocal
988	chromosome substitutions in winter wheat. Crop Science 26:531-533.
989	
990	Zhai H, Gong W, Tan Y, Liu A, Song W, Li J, Deng Z, Kong L, Gong J, Shang H, Chen T, Ge
991	Q, Shi Y, Yuan Y (2016) Identification of chromosome segment substitution lines of
992	Gossypium barbadense introgressed in G. hirsutum and quantitative trait locus mapping for

- fiber quality and yield traits. *PLoS One* 11: e0159101. doi: 10.1371/journal.pone.0159101.
- 994
- 295 Zhang P, Dundas IS, McIntosh RA, Xu SS, Park RF, Gill BS, et al. (2015) Wheat–Aegilops
- 996 Introgressions. In: Alien Introgression in Wheat: Cytogenetics, Molecular Biology, and
- 997 Genomics (Molnár-Láng M, Ceoloni C, Doležel J, eds) pp 221-243. New York: Springer. doi:
- 998 10.1007/978-3-319-23494-6
- 999
- Zhao X, Lv L, Li J, Ma F, Bai S, Zhou Y, Zhang D, Li S, Song CP (2021) Genome-wide
 association study of grain shapes in *Aegilops tauschii*. *Euphytica*, 217: 1-14.
- 1002
- 1003
- 1004