Bacterial lipoxygenases are associated with host-microbe interactions and may provide cross-kingdom host jumps

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Abstract

In this bioinformatic research, we studied the association of bacterial lipoxygenases (LOXs) with pathogenic and symbiotic traits by text networks analysis, phylogenetic analysis, and statistical analysis of molecular structure. We found that bacterial lipoxygenases are associated with a broad host range — from coral to plants and humans. In humans, bacterial LOXs are associated with opportunistic and nosocomial infections as well as with affecting specific patient populations like cystic fibrosis patients. Moreover, bacterial LOXs are associated with plant-human (or human-plant) host jumps in emerging pathogens. We also inferred a possible mechanism of such host jumps working *via* a host's oxylipin signalling "spoofing".

Keywords: lipoxygenases, oxylipins, cross-kingdom pathogens, cross-kingdom host jumps, emerging pathogens, cystic fibrosis

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Background

Recently, the problem of host switch (also host shift) in pathogens has gained new relevance due to the growing threat posed by emerging pathogens [1]. The mechanism of host switching by the viruses responsible for the current (coronaviruses) or possible (henipaviruses, filoviruses) pandemics has been particularly actively studied. Host shifts typically involve two or more relatively closely related taxa with similar morphology and physiology. Coronaviruses and filoviruses pass to humans from bats, most often via intermediate hosts (palm civets, camels, primates), while new human-threatening influenza viruses often originate in waterfowl. In contrast, there are virtually no known examples of viruses capable of transferring from humans to plants and *vice versa*. The only

possible candidate for such a virus is the hepatitis D virus [2], which is similar to plant viroids, but its direct relatedness between them is controversial.

However, cross-kingdom host jump — a host switch in which the new host belongs to a different kingdom from the previous one — is common among bacteria. Most of the known cross-kingdom jumps occur exactly between bacteria and humans, and such bacteria have a specific clinical profile [3]. As a rule, they cause opportunistic infections (septicemia, pneumonia, surgical infections) without affecting healthy immunocompetent individuals. Less frequently, plant-animal host jumps have been recorded [4].

The question of mechanisms of such host jumps are intriguing. Such distant hosts have different molecular structures determining susceptibility to infection, and the biochemical basis of pathogen compatibility to different hosts is yet unclear although extensively studied. Proteases, phospholipases, components of quorum sensing systems, toxins, and secondary metabolism regulators have been identified as virulence factors common to plant- and animal-targeted virulence [3, 5].

In our recent paper [6], we had suggested the role of lipoxygenases and oxylipin signalling in cross-kingdom pathogenicity. Lipoxygenases are very conserved enzymes involved in the synthesis of oxylipins — signalling compounds regulation stress and immune responses in a vide range of multicellular eukaryotes, such as animals (including humans), plants, and different groups of algae. The main goal of our work was to trace evolutionary origins of oxylipin signalling using lipoxygenases as a bioinformatic proxy for oxylipin biosynthetic ability. We have found that in bacteria, lipoxygenases were primarily associated with multicellularity which reflects the possible role of oxylipins as ancient cell-to-cell signalling compounds.

However, some bacteria from our dataset did not possess any (albeit primitive) multicellularity. For some of these bacteria, ecophysiological data was not available. Regarding the fraction with described ecophysiology, it could be divided into three groups: (1) human- or vertebrate-associated bacteria, (2) plant-associated bacteria, (3) bacteria associated with marine organisms. Notably, the first two groups were significantly overlapped: at least two bacteria in our dataset (*Pseudomonas aeruginosa* and *Burkholderia gladioli*) were cross-kingdom pathogens capable of infecting both plants and humans. Continued updating of our dataset resulted in adding one more species (*Pantoea ananas*) to the list of lipoxygenase-positive cross-kingdom pathogens [7]. These facts lead to an assumption that lipoxygenases may facilitate cross-kingdom host jumps and serve as versatile virulence/symbiosis factors.

In our previous paper [6], we have noticed some additional traits common for lipoxygenase-positive bacteria. They were opportunistic pathogens (these bacteria are prone to affect immunocompromised people, especially patients with cystic fibrosis). The most concerning common trait we noticed is multiple drug resistance [7].

However, these traits were noticed during the non-systematic review of literature data on bacteria in which a lipoxygenase was detected. The list of these characteristic traits was not reinforced by thorough statistical analysis. The main focus of the previous papers was a link between LOXs and multicellularity, and conclusions on a link between LOXs and host-microbe interactions were preliminary.

In the current article, we continue the same research project and elaborate this association in more details. The aim of this bioinformatic research was to find additional evidence that lipoxygenases and oxylipins contribute to host colonization and invasion, as well as to cross-kingdom host jumps.

Materials and Methods

Collecting and updating the dataset

We started with the same dataset of bacterial lipoxygenases as used in our previous research [6]. However, we are updating it in an ongoing manner because LOX-like sequences are constantly added to protein databases, and some sequences are revised and deleted.

The method of updating the dataset is the same as for collecting the initial dataset (described in our previous paper [6]). The difference was that we used pathogen and symbiont LOX sequences form the "old" dataset as queries and downloaded all hits that belong to pathogens and symbionts (they were typically sorted at the top of hits list) along with some cyanobacterial and myxobacterial LOXs (according to our previous research, they are outgroups for all pathogen and symbiont LOXs). The methods for checking LOX-like hits were the same as described in [6].

The resulting list of LOX-carrying bacteria was used for the estimation of statistically reinforced ecological profile of bacterial LOXs and LOX-carrying bacteria. The criteria of inclusion into this list were:

- 1) the presence of a species in the updated LOX dataset;
- 2) availability of any ecological data for in in the literature;
- 3) pathogenic/symbiotic properties or other association with any host.

Ecological profile generation by network text analysis

We used a combination of systematic literature review and network text analysis to estimate the ecological profile of LOX-carrying pathogenic and symbiotic bacteria.

The species names form the list of LOX-carrying bacteria were used as the queries for PubMed database search (<u>https://pubmed.ncbi.nlm.nih.gov/</u>). The first ten results for each query were used for further text analysis. We used article abstracts, titles, and keywords for manual extraction of terms characterizing the ecology of the respective species (referred to below as "terms"). Totally, this research used an array of 137 papers for meta-analysis [8-144].

These terms were manually normalized to the root form and, in some cases, to the most frequent form (e.g., "emergent" to "emerging", "growth promotion" to "growth-promoting"). An important exclusion (that could potentially bias the statistics of the term) were any terms signing antimicrobial resistance: they all were normalized as "AMR", and any terms signing multidrug resistance were normalized as "MDR".

For each term, we counted the number of species in which it occurs to estimate the abundance of each term in the collective ecological profile of the LOX-carrying bacteria. Then we explored associations between the terms by building a graph where each node represents a term, and each edge represents an occurrence of two terms (represented by its nodes) in the same species. The weight of each edge represents the number of species where the respective terms occur together. We classified all terms to 6 groups:

- 1) "vertebrate-related" or "human-related" terms associated with affecting humans and pathogenicity to humans and/or vertebrates (i.e., "human", "lung", "abscess", cystic fibrosis" etc.);
- "plant-related" terms associated with plant pathogenicity or plant symbiosis (i.e., "plant", "rhizosphere", "root" etc.);
- 3) "insect-related" group included only two terms ("insect" and "larvae");
- 4) "marine-related" terms reflecting association with marine invertebrates or algae;
- 5) "public health threat" group (i.e. "emerging", "AMR", "MDR", "threat" etc.);
- 6) other terms not fitting in any of the above groups.

This analysis was performed with the aid of Microsoft Excel 2019, Python 3, Gephi 0.9 [145], and Cosmograph (<u>https://cosmograph.app/</u>) software.

Phylogenetics and conservation analysis

Phylogenetic analysis was performed with the same software and with the same protocols as described in our previous paper: MAFFT online v. 7 [146],

MEGA X [147], and iTOL [148]. Amino acid conservation analysis was performed with ConSurf server [149] in ConSeq mode [150].

Binding site statistical analysis

For the statistical analysis of binding site structures, we used amino acid residue volumes provided by Perkins (1986) [151]. Statistical analysis was performed with MS Excel 2019 and PAST [152].

The inclusion criteria for the statistical analysis were:

- 1) a LOX is represented on any phylogenetic tree;
- 2) (preferable) the bacteria, possessing this LOX, is described in terms of their ecophysiology.

Results

Collecting and updating the dataset

The full updated list of LOX-carrying bacteria is provided in **Table 1** with the color-coded ecological functions.

Order	Species
Burkholderiales	V. paradoxus, V. guangxiensis, V. gossypii, B. gladioli, B. singularis, B. thailandensis, B. stagnalis
Corynebacteriales	Nocardia seriolae, N. pseudobrasiliensis, N. brasiliensis, Mycobacteroides abscessus, Rhodococcus erythropolis, Rhodococcus sp. 66b
Enterobacterales	Pluralibacter gergoviae, Kosakonia sp. AG348, K. sacchari, E. hormaechei, Enterococcus faecium, Cedecea lapagei, Pantoea sp. OXWO6B1, P. ananas, Moellerella wisconsensis, Dickeya zeae
Holosporales	Candidatus Finniella inopinata
Nitrospinae/ Tectomicrobia group	Candidatus Entotheonella palauensis
Oceanospirillales	Gynuella sunshinyii, Endozoicomonas numazuensis

Oligoflexales	Pseudobacteriovorax antillogorgiicola
Pseudomonadales	Pseudomonas aeruginosa
Pseudonocardiales	Kutzneria sp. 744, Pseudonocardia acaciae, Lentzea kentuckyensis
Vibrionales	V. vulnificus, V. penaeicida, Enterovibrio norvegicus, E. coralii, E. calviensis, E. nigricans

Table 1. Full list of analyzed LOX-carrying species with color-coded ecological functions. Human/vertebrate-associated bacteria are depicted in **red**, plants-associated bacteria are depicted in **green**, insect-associated bacteria are depicted in **violet**, marine-associated bacteria are depicted in **blue**. If a bacterium has several host types, the corresponding colours are all represented in its name.

The ecological profile reveals intriguing traits of LOX-carrying bacteria

The most common terms (summarized in **Figure 1**) provide some insights into the most characteristic ecological traits of LOX-carrying pathogens and symbionts. The most concerning fact is that "public health threat" group of the terms are extremely prevalent among these bacteria. The leader of prevalence was the "AMR" term. The analysis of the most prevalent terms of the "human-related" and "plant-related" groups shed light to some ecological details of host-microbe interaction in LOX-carrying bacteria. For instance, plant-associated LOX carriers usually colonize roots or are endophytic. LOX carriers pathogenic for humans are prone to affect lungs and cause nosocomial/opportunictic infections.

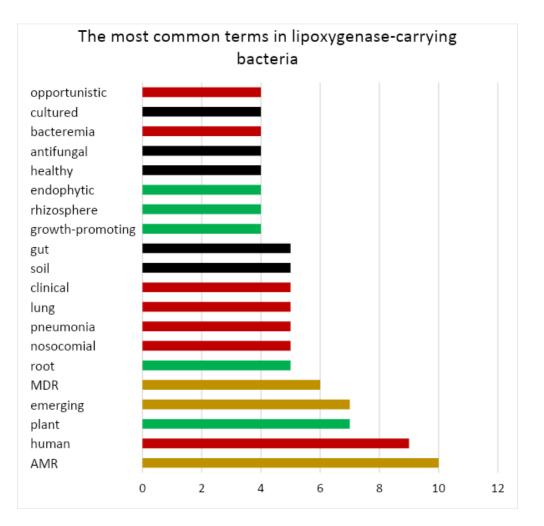


Figure 1. The diagram depicting the occurrence of each term (i.e., the number of species in which it occurred). This occurrence ranged from 2 to 10. Here, the most common terms are summarized, whose occurrence is not less than 4. The "human-related" and "vertebrate-related" terms are depicted in **red**, the "plant-related" terms are depicted in **green**, and the "public health threat" group of terms is depicted in **yellow**. *Created with MS Excel 2019*

The network analysis results (Figure 2 a, b) show that the entire network can be divided in some clusters of terms:

- 1) a "plant-related" cluster comprising "plant-related" terms;
- 2) a "human and public health threat" cluster comprising both "human related" and "public health threat" term groups;
- 3) a "marine-related" cluster comprising "marine-related" terms;

The "insect-related" terms do not form stable clusters which would persist in different graph layouts.

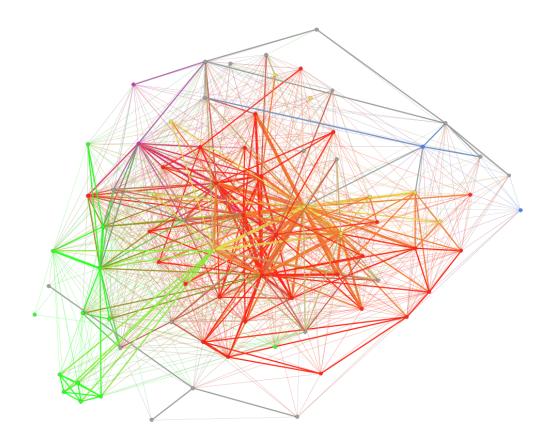


Figure 2a. The full network of all terms in our dataset.

The nodes depict terms, the edges depict their occurrence in the same bacterial species. The "human-related" and "vertebrate-related" terms are depicted in **red**, the "plant-related" terms are depicted in **green**, and the "public health threat" group of terms is depicted in **yellow**, the "insect-related" terms are **magenta**, and the "marine-related" terms are **blue**. *Created with Gephi 0.9*

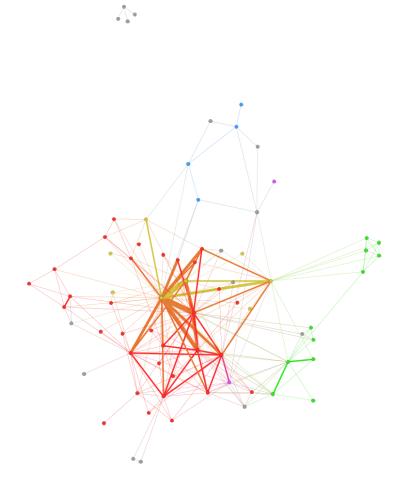


Figure 2b. The "backbone" of our term network depicting only edges with the weight not less than 2 and the corresponding nodes only (i.e., only the terms occurring in the same LOX-carrying species not less than two times). This graph better depicts the network-forming connections and hubs of the network. Notably, the yellow nodes (the "public health threat" group terms) are the network-forming hubs providing connectivity to the entire networks, as well as the "human-related" group terms".

The nodes depict terms, the edges depict their occurrence in the same bacterial species. The "human-related" and "vertebrate-related" terms are depicted in **red**, the "plant-related" terms are depicted in **green**, the "public health threat" term group is depicted in **yellow**, the "insect-related" terms are **magenta**, and the "marine-related" terms are **blue**. *Created with Gephi 0.9*

The "plant-related" cluster had many connections to the "human and public health threat" cluster presumably *via* the network-forming hubs. These hubs represented the same terms from the "human-related" and the "public health threat" groups that were included in **Figure 1**. Thus, the "AMR" term had direct connections to 83% of all nodes including more than 50% of "plant-related" terms. The "nosocomial" term had direct connections to 69% of all nodes including the same proportion of "plant-related" terms and is tightly clustered with the "human"

term. This fact corresponds to the most common term prevalence and confirms the fact that the human infections caused by LOX-carrying bacteria are predominantly nosocomial or opportunistic. Finally, the term "emerging" is a hub connected to 73% of all nodes including almost all "human-related" and "plant-related" clusters (but almost not connected to the "marine-related" cluster) and reflecting the fact that pathogenic LOX carriers are predominantly characterized in the literature as emerging pathogens.

When analyzing the more specific terms, it appeared that the "cystic fibrosis" term had connections to 55% of all nodes, including 42% of "plant-related" terms such as "endophytic" and "leaves". It is a very interesting association: the endophytic dwelling capacity in plants is associated with affecting cystic fibrosis patients in LOX-carrying bacteria. The organ-denominating term "lung" was also a hub having connections to 70% of all plant organ terms such as "leaves" and "root". So, the same bacterial LOXs are probably associated with affecting lungs in humans and affecting roots in plants.

The results of this analysis also show that the plant pathogenesis terms (such as "rot") and plant symbiosis terms (such as "growth-promoting") lie within the same "plant-related" cluster. This shows that bacterial LOXs are associated both with plant pathogenicity and plant symbionts. In contrast, we do not see such an association in the "human" cluster where all terms are pathogenesis-related.

We may also conclude that this network analysis strongly supports our previous hypothesis that cross-kingdom host jump ability is a common trait of the LOX-carrying bacteria. They are mainly represented by plant-human host jumps. Conversely, "marine cluster" occupied the peripheral position and had significantly lower number of connections to other clusters indicating that plant-marine or human marine host jumps are less common.

Phylogenetic analysis results correlate with the collective ecological profile of LOX-carrying bacteria

The most important finding was that the results of bacterial LOX phylogenetic analysis correlated not with the phylogeny of the bacterial species themselves rather with their ecological profile described above.

All LOXs analyzed in the current study can be divided into 4 clusters. Two clusters (**Figure 3**) comprised the LOXs from phylogenetically distinct species that are: (1) human/vertebrate pathogens; (2) plant pathogens; (3) both — are able to affect both plants and humans. They reflected two independent series of the horizontal transfer of the LOX genes between plant symbionts, versatile (plant/human) pathogens (including *Pseudomonas aeruginosa*), and human/vertebrate pathogens. It corresponds well to a large number of connections between "plant-related" and "human/vertebrate-related" term groups in the

common ecological profile of LOX-carrying bacteria. Notably, there is a fourth group in Cluster 1 and Cluster 2 represented by plant symbionts. This phylogenetic connection of plant pathogenesis and symbiosis in LOX carriers also corresponds to the similar associations on the ecological profile.

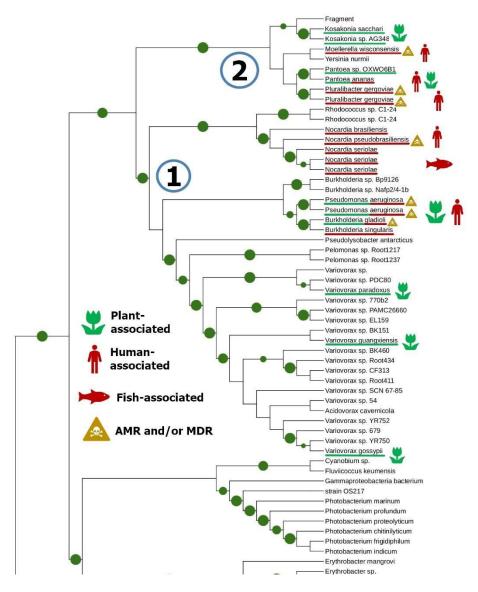


Figure 3. Clusters 1 and 2 of bacterial LOXs reflect two independent series of horizontal gene transfers between plant symbionts, versatile pathogens, and vertebrate/human pathogens. Here, only a fragment of the phylogenetic tree is represented. *Created with iTOL*

Conversely, the LOXs of bacteria associated with marine invertebrates were predominantly separated to the distinct cluster (we call it "Cluster 4", Figure 4) and a distinct subcluster within the "Cluster 3" along with plant and human/vertebrate pathogens. Notably, no versatile (cross-kingdom) pathogens were included in the Cluster 3 (Figure 5), and it is the only cluster where the possible "terrestrial-marine" LOX gene transfer was observed. So, this cluster is significantly dissimilar from the Clusters 1 and 2, although this phylogenetic

picture still corresponds to the collective ecological profile described above in terms of low connectivity between "marine-related" terms and "plant-related"/"human-related" term groups. Taken together, these data indicate that bacterial LOXs are associated with host-microbe interactions and — especially — with cross-kingdom host jumps.

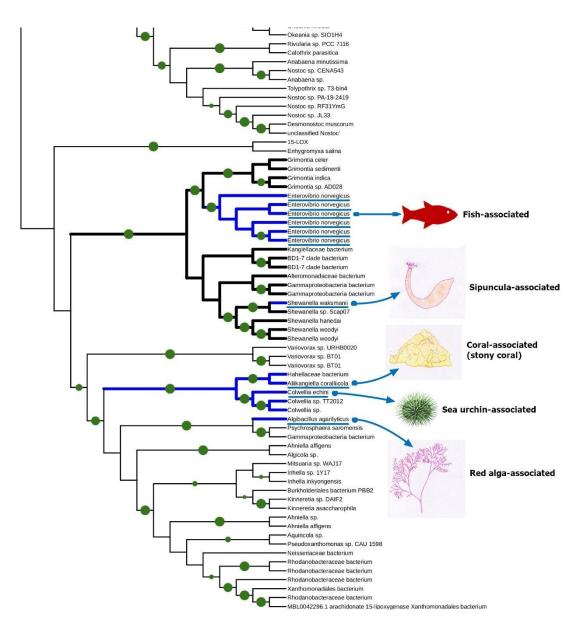


Figure 4. The fragment of phylogenetic tree depicting the "Cluster 4" of bacterial LOXs. This cluster encompasses bacteria isolated from different aquatic organisms: a red alga, a sea urchin, a stony coral, a sipunculid worm, and a fish. *Created with iTOL and BioRender.com*

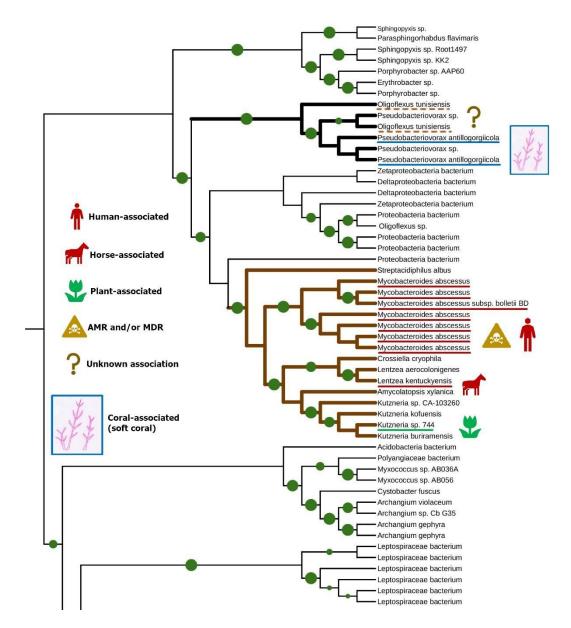


Figure 5. This fragment of a phylogenetic tree depicts the "Cluster 3" of bacteria LOXs — the only cluster, where the transfer between plant- and vertebrate associated bacteria and a coral-associated bacteria was recorded. *Created with iTOL*

Binding site analysis reveals that (ω-6)S-specificity of bacterial LOXs may contribute to plant-human host jumps

We have revealed that bacterial LOXs are associated with plant-human host jumps, while host jumps between marine and terrestrial organisms are less common. It could be explained by the environmental difference: both plants and humans live in the terrestrial environment and have much more chances to contact each other than to contact any organism from the aquatic environment. However, we decided to check if any biochemical properties of these LOXs determine and explain their ecology.

We used ConSurf and additional statistical analysis to find any differences in key residues determining the substrate insertion direction, stereospecificity, and regiospecificity. These residues had been already characterized in papers [153, 154]. We have not found any significant differences between studied bacterial lipoxygenases in the terms of insertion-determining residues (probably the "tail first" insertion in all lipoxygenases) and Coffa residue (Ala in all our LOXs, conservation score = 9, which means that all LOXs in our dataset have S-stereospecificity).

LOX regiospecificity is determined by the bottom triad of the substrate-binding site -3 amino acid residues forming the binding site bottom. We calculated the total volume of these residues (as the sum of residue volumes) and performed *t*-test for three groups of LOXs:

- 1) LOXs of human- and vertebrate-associated bacteria (n=12);
- 2) LOXs of plant-associated bacteria (n=10);
- LOXs of marine-associated bacteria (n=6) (the LOXs of cross-kingdom pathogens were included both in the "plant" and the "human-vertebrate" samples).

There was no statistically significant difference between the "human/vertebrate group" and the "plant group" (t=1.68). But there were statistically significant differences between each of the above groups and the "marine group" (t=4.5 and t=5.92, respectively) even when checking the null hypothesis against p=0.01 (it approximately corresponds to p=0.05 with Bonferroni correction).

Indeed, the LOXs of human-associated bacteria and the LOXs of plant-associated bacteria have on average almost the same total volumes of the binding site bottom triad (mean=475.24×10⁻³ nm³, σ =30.45×10⁻³ nm³ and mean=497.27×10⁻³ nm³, σ =30.82×10⁻³ nm³, respectively). The bottom volumes of the LOXs of marine-associated bacteria were much lower (mean=411.87×10⁻³ nm³, σ =21.72×10⁻³ nm³). This is graphically shown at the box plot (**Figure 6**).

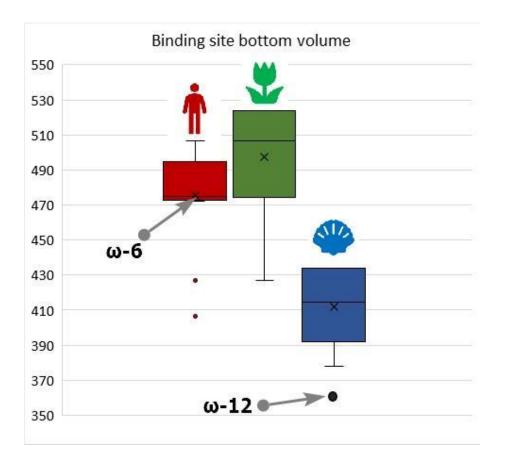


Figure 6. The box plot of total LOX ligand binding site bottom volumes for "human/vertebrate-associated" (**red**), "plant-associated" (**green**), and "marine-associated" (**blue**) groups. All volumes are given in 10^{-3} nm³ according to [151]. LOXs of "human/vertebrate-associated" and "plant-associated" groups have total bottom volumes at almost the same level; in contrast, LOXs of the "marine-associated" group have significantly lower bottom volumes (p<0.05 with Bonferroni correction). The bottom volumes of "human/vertebrate-associated" and "plant-associated" groups correspond to (ω -6)-LOX activity.

The box plot (**Figure 6**) also graphically shows the presence of two outliers in the "human-vertebrate" group with extremely low total bottom volume. But they fully correspond to the average values for the "marine" group and lie within 1σ from its mean. These LOXs phylogenetically belong to the Cluster 3 — the only cluster where the terrestrial-marine LOX gene transfer was recorded.

Thus, binding site bottom volume of bacterial LOXs strictly corresponds with the possibility of their horizontal transfer and ecological traits of their carriers.

Discussion

Possible mechanism of the LOX-mediated plant-human host jumps

We have for the first time confirmed that pathogen and symbiont LOXs are associated with broad host range and cross-kingdom host jump ability. The correlation of the LOX phylogeny with the host-microbe interactions also confirms

that these enzymes are involved in pathogenesis and symbiosis and probably interact with a host.

One of the pathogen LOXs in our dataset – *Pseudomonas aeruginosa* LoxA — was earlier experimentally characterized providing the pathophysiological insights into this interaction [155]. This LOX is represented in our dataset, our statistical analysis sample and in the Cluster1 of the phylogenetic tree and attributed to be a LOX of a cross-kingdom pathogen because of a broad host range of *Pseudomonas aeruginosa*. However, the available experimental data regard only the role of *Pseudomonas aeruginosa* LOX in interactions with a human organism.

When invading human tissues, this pathogen secretes a 15S-LOX which converts the host's arachidonic acid to 15S-HETE which is further metabolized by the human leukocytes to anti-inflammatory mediators such as lipoxin A_4 (**Figure** 7). Their action, in turn, leads to decrease of immune cell recruiting and immune response itself, and, finally, to the invasion enhancement [155].

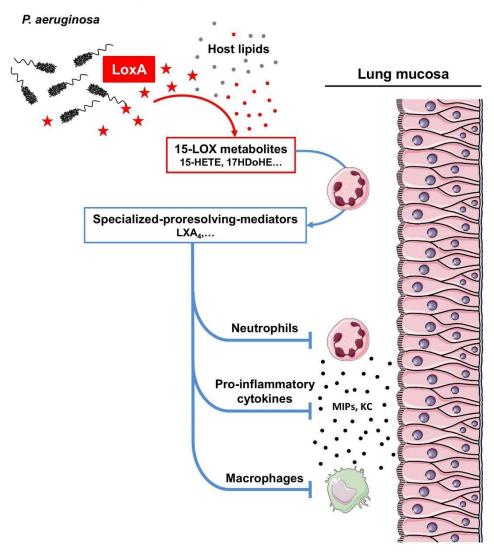


Figure 7. The graphical outline of Pseodomonas aeruginosa LOX action to facilitate a host's tissue invasion. *Image credit:* Morello, E. et al. (2019) *Frontiers in Microbiology* [155], CC BY 4.0

Our results suggest that such an interaction is not restricted to humans, and some bacteria use LOXs to interact with different hosts — from plants to corals — in the course of both parasitic and symbiotic relationships. Moreover, LOX genes can make horizontal transfers thus finding new ecological niche for themselves (like Richard Dawkins's "selfish genes" [156]) and also enable their carriers to make cross-kingdom host jumps. Such species are discussed above as versatile pathogens.

According to our data, versatile LOX-carrying bacteria affect plants, vertebrates, sometimes insects, but never affect marine invertebrates. Furthermore, plant-vertebrate LOX transfers are the most frequent events according to our phylogeny (at least 3 independent series of plant-human transfers), while vertebrate-marine or plant-marine LOX transfers are rarer (only one even according to our phylogeny). In our ecological networks, a small "marine-related" cluster is located apart from the "plant-related" and "human-related" clusters. It leads to an assumption that bacterial lipoxygenases can easily switch from a plant host to a vertebrate host (and vice versa), but the switch to a marine host is very uncommon. The simplest explanation could be the ecological isolation: both plants and humans live in the terrestrial environment and have much more tight contact between each other than with any aquatic organism. According to this hypothesis, if humans were aquatic mammals (like dolphins) and lived in a coral reef ecosystem, we could observe multiple "coral-human" host jumps. This hypothesis assumes full "compatibility" of pathogen/symbiont LOXs with any organism: vertebrate, insect, plant, or marine invertebrate.

However, we have found that LOXs themselves have structural differences in the ligand-binding site which correlate with the host type. The "plant-compatible" and the "vertebrate-compatible" LOXs appeared to have no statistically significant differences in the binding site structure in contrast to the "marine-compatible" LOXs. It leads to an assumption that bacterial LOXs face some biochemical "compatibility requirements" for successful host-microbe interactions. According to this hypothesis, bacterial LOXs are capable of cross-kingdom "jumps" because plant and animal hosts require the same regio- and stereospecificity.

The experimental data on *Pseudomonas aeruginosa* and *Burkholderia thailandensis* LOXs [155, 157, 158, 159] (both belonging to the "human/vertebrate" group in our statistics) enable us to infer the LOX specificity needed both for colonizing plants and humans. The both LOXs are (ω -6)S-LOXs and have total binding site bottom volumes (472.2×10⁻³ nm³ and 475×10⁻³ nm³, respectively) near to the mean of the "human-related" group. The mean of the "plant-related" group is even more. It must also correspond to the (ω -6)S-LOX

activity because it requires the minimally profound substrate penetration into the active site (and, thus, the maximal volume of the site bottom). So, we may conclude that (ω -6)S-LOX activity is required for plant-vertebrate host jumps.

In the case of human (and other vertebrates), any (ω -6)S-LOX of a pathogen will behave like *Pseudomonas aeruginosa* LOX: for arachidonic acid prevalent in the vertebrate PUFA pool, (ω -6)S-LOX activity mush mean 15S-LOX activity, which contributes to the lipoxin biosynthetic pathway and suppresses the host's immune response (which is fulfilled in the case of *Pseudomonas aeruginosa*).

But what about plants? In contrast to humans, we have no direct experimental data on the interaction of any bacterial LOX with them. However, we could biochemically infer this interaction. In the case of linolenic acid (abundant in plant cells), any (ω -6)S-LOX will produce 13S-HpOT, which is a normal precursor of jasmonates. So, if (ω -6)S-LOX enhances lipoxin production in the human tissues, they should enhance jasmonate production in the plant body.

Here, the "explanative gap" is fully closed because jasmonate signalling hijacking is a well-known trick of some plant pathogens. They increase a plant's susceptibility by inappropriate activation of jasmonate signalling pathway. However, almost all instances of such pathogenesis known up do date involve the use of toxin mimicking the natural jasmonate or acting as prohormone. The best-known example is *Pseudomonas syringae* that uses coronatine activating the jasmonate receptor JAZ-COI1 [160] to facilitate the invasion. A grapevine pathogen *Lasiodiplodia mediterranea* facilitates its invasion by the prohormone toxin lasiojasmonate A [161]. The current study provides computational evidence that bacteria can use lipoxygenase to "spoof" the plant immunity with natural jasmonates rather than jasmonate-mimicking toxin. By accident or by the convergent evolution, the same LOX activity appeared to be "compatible" with a human lipoxin pathway which enabled LOX-carrying bacteria to be versatile pathogens (**Figure 8**).

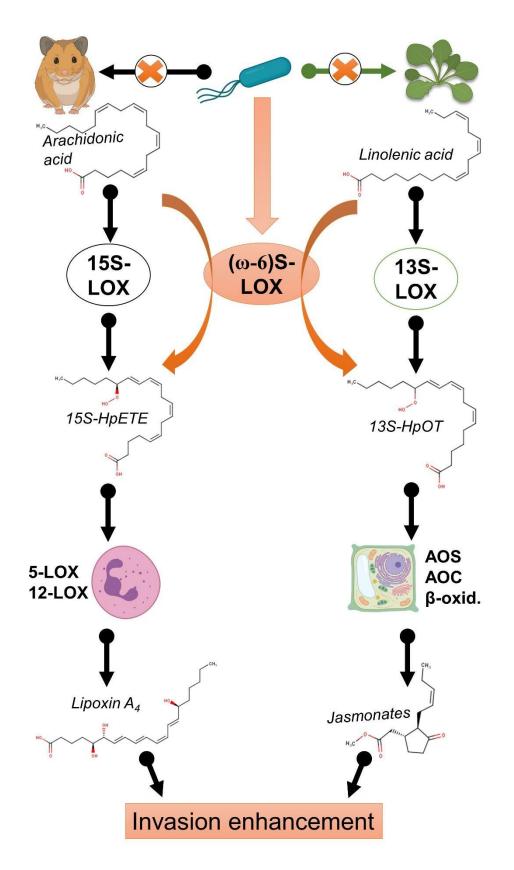


Figure 8. Hypothetical mechanism of plant-animal host jumps provided by bacteria LOXs. In humans and vertebrates, a secreted bacterial (ω -6)S-LOX oxidizes arachidonic acid to form 15S-HpETE, which is further converted into lipoxins in leukocytes by other LOX isozymes. Lipoxins suppress inflammation and immune response thus facilitating the invasion. *Created with BioRender.com*

The immune response suppression hypothesis partially explains why this feature is shared by plant pathogens and plant symbionts (as well as insect pathogens and symbionts): the both groups need to evade the host's immune response to survive in its body. However, this raises an additional question: why human symbionts do not use this strategy?

It is not the only specific thing of interaction between LOX-carrying bacteria and a human (or a vertebrate) organism. As discussed above, the ability of such pathogens to affect humans is extremely strongly associated with nosocomial or opportunistic traits. It means that these bacteria are prone to affect immunocompromised people (the terms "compromised" and "immunocompromised" are also present in the "human-related" cluster, but have relatively low connectivity compared to hubs described above). Conversely, in the case of plants and insects, we did not observe such associations: LOX carriers are capable of full-fledged pathogenesis and symbiosis.

These differences are tightly connected: if a pathogen oxylipin "spoofing" is insufficient to suppress the healthy human's immune response, there is no reason to expect that it will be sufficient for human symbionts to evade the immune response and survive. Evidently, there is something in the human (or, more broadly, vertebrate) immune system that makes it more resilient to oxylipin signalling hijacks. Here, we come on shaky grounds: we have insufficient data to explain what it could be. But it is reasonable to suggest that it is the presence of adaptive immunity. Indeed, one of the mechanisms used by human microbiota to evade immune response is immune tolerance — the acquired silencing of a host's adaptive immunity to the bacterium's antigens [162].

Public health risks and molecular epidemiology surveillance

The fact that the terms "emerging", "AMR", and "MDR" are prevalent in the collective ecological profile of LOX carriers and serve as hubs in the term representing this profile should network raise concerns. Moreover. antimicrobial-resistant and multi-drug resistant bacteria are represented in each of 3 phylogenetic clusters of the pathogen and symbiont LOXs. It means that bacterial LOXs are associated with emerging status, antimicrobial resistance and some other public health threats. Furthermore, multiple connections between these traits and plant or human pathogenicity entitle us to say that emerging pathogens use oxylipin signalling for plant-human (or human-plant) host jumps.

But this conclusion should be considered carefully. Firstly, we have mentioned above that the traits "AMR" and "MDR" could have overestimated weights due to normalization bias. We are sure that this possible bias has limited effect because the used term "AMR" corresponds to the real trait of studied bacteria. Moreover, the term "emerging" has also a very big weight, but cannot be subjected to normalization bias. But on the visual level, it could produce a formidable picture and lead to the emergence of news about "new deadly pathogens" or a "new global epidemic". We caution all science journalists reading this article against doomsaying.

Secondly, our data provide strong evidence that LOX-carrying pathogens have low epidemic and pandemic potential. As we discussed above, they are prone to affect immunocompromised people or people with specific diseases (e.g., cystic fibrosis), and cannot fully overcome the immune barrier of a healthy human despite multiple recorded host jump events. So, we find no reason to expect a full-fledged epidemic or pandemic caused by LOX-carrying pathogens.

However, all statistical signs of "public health threat" are connected with the extreme danger they pose to specific populations — immunocompromised people or people with comorbidities, especially with cystic fibrosis. Regarding these populations, we cannot rule out the possibility of limited outbreaks. Sporadic cases are also dangerous for these people and could lead to mortality and decrease in the quality of life. Thus, bacterial lipoxygenases could be a useful molecular epidemiology marker for oncology, hematology, transplantology, and cystic fibrosis medicine.

We continue updating our "pathogen blacklist" — the list of LOX-carriyng pathogens that can pose danger to humans — on the grounds of publicly available data. The last update before this article was published in English in the *Nature Portfolio Microbiology Community* blog [163] and in Russian in *Priroda* [7]. In this work, this list is incorporated into Table 1. But the full-fledged molecular epidemiology surveillance requires much more strain-dependent data. We are open for further collaboration for this purpose.

Possible implications for marine biology

In contrast to plants and humans, our knowledge about LOX-carrying bacteria in marine organisms is limited. In most cases, we suggested the association with marine organisms only on the basis of isolation of respective bacteria from a particular organism. So, the ecological functions of LOX-carrying bacteria (pathogen/symbiont) are unknown for all "marine-associated" bacteria in our dataset. But we can suggest that in the underwater world, more intriguing host jumps of LOX-carrying bacteria occur. They involve corals, sea urchins, algae, sipunculids, and fishes. We have found that these host jumps require other LOX regiospecificity, than for plants and humans. We cannot infer any mechanism of their actions because of scarce data on oxylipin signalling in marine invertebrates, but can suggest that bacterial LOXs could be a useful tool to study it.

Conclusions

In this article, we have proposed a mechanism of cross-kingdom host jumps by interacting with evolutionary distant immune systems. The analysis of available data shows that some bacteria may exploit the hosts' oxylipin signalling systems which are widespread across different kingdoms. The most prevalent are plant-human host jumps because of a biochemical "intercompatibility" of plant and human LOX activities needed for immune response suppression.

This finding could be interesting for basic scientists in a wide range of areas – from immunology to marine biology. But it could also be useful for medical microbiology and public health. Bacterial LOXs in a nosocomial environment could indicate an emerging pathogen which could be dangerous for some vulnerable groups of patients. Bacterial LOX carriers isolated from patients may require further consideration in terms of antimicrobial therapy and the particular role of this bacteria in the patient's condition pathogenesis (as is the case of cystic fibrosis). It is evident that pathogen and symbiont LOXs require further investigation in both basic and applied life sciences.

Acknowledgements

The author acknowledges the data visualization expert Anastasiya Kuznetsova (<u>https://nastengraph.medium.com/</u>) for her valuable advice in the course of this research.

Ethics statement

This research did not involve any experimental animals or humans as a research object.

Conflict of interests

The author declares no conflict of interests. This research did not have any special funding.

Data availability

All datasets, spreadsheets, interactive data and other forms of data generated for this research are available by request to the author's email.

References

- 1. Pulliam, J. R. (2008). Viral host jumps: moving toward a predictive framework. *EcoHealth*, *5*(1), 80-91.
- 2. Taylor, J., & Pelchat, M. (2010). Origin of hepatitis δ virus. *Future microbiology*, *5*(3), 393-402.
- 3. Kirzinger, M. W., Nadarasah, G., & Stavrinides, J. (2011). Insights into cross-kingdom plant pathogenic bacteria. *Genes*, 2(4), 980-997.
- 4. Van Baarlen, P., Van Belkum, A., Summerbell, R. C., Crous, P. W., & Thomma, B. P. (2007). Molecular mechanisms of pathogenicity: how do pathogenic microorganisms develop cross-kingdom host jumps?. *FEMS microbiology reviews*, *31*(3), 239-277.
- Rahme, L. G., Ausubel, F. M., Cao, H., Drenkard, E., Goumnerov, B. C., Lau, G. W., ... & Tompkins, R. G. (2000). Plants and animals share functionally common bacterial virulence factors. *Proceedings of the National Academy of Sciences*, 97(16), 8815-8821.
- 6. Kurakin, G. F., Samoukina, A. M., & Potapova, N. A. (2020). Bacterial and Protozoan Lipoxygenases Could be Involved in Cell-to-Cell Signaling and Immune Response Suppression. *Biochemistry (Moscow)*, *85*(9), 1048-1063.
- Kurakin., G. F. (2022). Oksilipiny bakterij: kljuch k mnogokletochnosti i bor'be s ustojchivost'ju k antibiotikam? [Bacterial oxylipins: a key to multicellularity and to combating antimicrobial resistance?] *Priroda*, 2, 26–32.
- Toukabri, W., Ferchichi, N., Hlel, D., Jadlaoui, M., Kheriji, O., Mhamdi, R., & Trabelsi, D. (2021). Response of intercropped barley and fenugreek to mono- and co-inoculation with Sinorhizobium meliloti F42 and Variovorax paradoxus F310 under contrasting agroclimatic regions. *Archives of microbiology*, 203(4), 1657–1670.
- 9. Garcia Teijeiro, R., Belimov, A. A., & Dodd, I. C. (2020). Microbial inoculum development for ameliorating crop drought stress: A case study of Variovorax paradoxus 5C-2. *New biotechnology*, *56*, 103–113.
- 10.Gao, J. L., Yuan, M., Wang, X. M., Qiu, T. L., Li, J. W., Liu, H. C., Li, X. A., Chen, J., & Sun, J. G. (2015). Variovorax guangxiensis sp. nov., an aerobic, 1-aminocyclopropane-1-carboxylate deaminase producing bacterium isolated from banana rhizosphere. *Antonie van Leeuwenhoek*, 107(1), 65–72.
- 11. Kämpfer, P., Busse, H. J., McInroy, J. A., & Glaeser, S. P. (2015). Variovorax gossypii sp. nov., isolated from Gossypium hirsutum. *International journal of systematic and evolutionary microbiology*, 65(12), 4335–4340.
- 12.Belli, G., Giovannini, M., Dolce, D., Terlizzi, V., Orioli, T., & Taccetti, G. (2021). Burkholderia gladioli infection in a pediatric patient with cystic

fibrosis: the clinical challenges of an emergent pathogen. *Minerva pediatrics*, 73(5), 468–470.

- 13.Cui, G., Yin, K., Lin, N., Liang, M., Huang, C., Chang, C., Xi, P., & Deng, Y. Z. (2020). *Burkholderia gladioli* CGB10: A Novel Strain Biocontrolling the Sugarcane Smut Disease. *Microorganisms*, 8(12), 1943.
- 14.Bedir Demirdag, T., Ozkaya Parlakay, A., Aygar, I. S., Gulhan, B., & Kanik Yuksek, S. (2020). Major Aspects of Burkholderia gladioli and Burkholderia cepacia Infections in Children. *The Pediatric infectious disease journal*, 39(5), 374–378.
- 15.Zanotti, C., Munari, S., Brescia, G., & Barion, U. (2019). Burkholderia gladioli sinonasal infection. *European annals of otorhinolaryngology, head and neck diseases*, *136*(1), 55–56.
- 16.Dursun, A., Zenciroglu, A., Karagol, B. S., Hakan, N., Okumus, N., Gol, N., & Tanir, G. (2012). Burkholderia gladioli sepsis in newborns. *European journal of pediatrics*, 171(10), 1503–1509.
- 17.Jones, C., Webster, G., Mullins, A. J., Jenner, M., Bull, M. J., Dashti, Y., Spilker, T., Parkhill, J., Connor, T. R., LiPuma, J. J., Challis, G. L., & Mahenthiralingam, E. (2021). Kill and cure: genomic phylogeny and bioactivity of *Burkholderia gladioli* bacteria capable of pathogenic and beneficial lifestyles. *Microbial genomics*, 7(1), mgen000515.
- 18.Marom, A., Miron, D., Wolach, B., Gavrieli, R., & Rottem, M. (2018). Burkholderia gladioli-associated facial pustulosis as a first sign of chronic granulomatous disease in a child - Case report and review. *Pediatric allergy and immunology : official publication of the European Society of Pediatric Allergy and Immunology*, 29(4), 451–453.
- 19.Ritterband, D., Shah, M., Cohen, K., Lawrence, J., & Seedor, J. (2002). Burkholderia gladioli keratitis associated with consecutive recurrent endophthalmitis. *Cornea*, *21*(6), 602–603.
- 20.Boyanton, B. L., Jr, Noroski, L. M., Reddy, H., Dishop, M. K., Hicks, M. J., Versalovic, J., & Moylett, E. H. (2005). Burkholderia gladioli osteomyelitis in association with chronic granulomatous disease: case report and review. *The Pediatric infectious disease journal*, 24(9), 837–839.
- 21.Zhou, F., Ning, H., Chen, F., Wu, W., Chen, A., & Zhang, J. (2015). Burkholderia gladioli infection isolated from the blood cultures of newborns in the neonatal intensive care unit. *European journal of clinical microbiology & infectious diseases: official publication of the European Society of Clinical Microbiology*, 34(8), 1533–1537.
- 22. Vandamme, P., Peeters, C., De Smet, B., Price, E. P., Sarovich, D. S., Henry, D. A., Hird, T. J., Zlosnik, J., Mayo, M., Warner, J., Baker, A., Currie, B. J., & Carlier, A. (2017). Comparative Genomics of *Burkholderia singularis* sp. nov., a Low G+C Content, Free-Living Bacterium That Defies Taxonomic Dissection of the Genus *Burkholderia*. *Frontiers in microbiology*, *8*, 1679.

- 23. Wang, Y., Hoffmann, J. P., Chou, C. W., Höner Zu Bentrup, K., Fuselier, J. A., Bitoun, J. P., Wimley, W. C., & Morici, L. A. (2020). Burkholderia thailandensis outer membrane vesicles exert antimicrobial activity against drug-resistant and competitor microbial species. *Journal of microbiology (Seoul, Korea)*, 58(7), 550–562.
- 24.Klaus, J. R., Majerczyk, C., Moon, S., Eppler, N. A., Smith, S., Tuma, E., Groleau, M. C., Asfahl, K. L., Smalley, N. E., Hayden, H. S., Piochon, M., Ball, P., Dandekar, A. A., Gauthier, C., Déziel, E., & Chandler, J. R. (2020). Burkholderia thailandensis Methylated Hydroxyalkylquinolines: Biosynthesis and Antimicrobial Activity in Cocultures. *Applied and environmental microbiology*, *86*(24), e01452-20.
- 25.Garcia E. C. (2017). Burkholderia thailandensis: Genetic Manipulation. *Current protocols in microbiology*, *45*, 4C.2.1–4C.2.15.
- 26. Vitale, A., Paszti, S., Takahashi, K., Toyofuku, M., Pessi, G., & Eberl, L. (2020). Mapping of the Denitrification Pathway in Burkholderia thailandensis by Genome-Wide Mutant Profiling. *Journal of bacteriology*, 202(23), e00304-20.
- 27.Place, D. E., Briard, B., Samir, P., Karki, R., Bhattacharya, A., Guy, C. S., Peters, J. L., Frase, S., Vogel, P., Neale, G., Yamamoto, M., & Kanneganti, T. D. (2020). Interferon inducible GBPs restrict Burkholderia thailandensis motility induced cell-cell fusion. *PLoS pathogens*, *16*(3), e1008364.
- 28. Pilátová, M., & Dionne, M. S. (2012). Burkholderia thailandensis is virulent in Drosophila melanogaster. *PloS one*, 7(11), e49745.
- 29.Garcia, E. C., & Cotter, P. A. (2016). Burkholderia thailandensis: Growth and Laboratory Maintenance. *Current protocols in microbiology*, *42*, 4C.1.1–4C.1.7.
- 30.Lei, X., Zhao, R., Geng, Y., Wang, K., Yang, P. O., Chen, D., Huang, X., Zuo, Z., He, C., Chen, Z., Huang, C., Guo, H., & Lai, W. (2020). Nocardia seriolae: a serious threat to the largemouth bass Micropterus salmoides industry in Southwest China. *Diseases of aquatic organisms*, 142, 13–21.
- 31.Han, H. J., Kwak, M. J., Ha, S. M., Yang, S. J., Kim, J. D., Cho, K. H., Kim, T. W., Cho, M. Y., Kim, B. Y., Jung, S. H., & Chun, J. (2019). Genomic characterization of Nocardia seriolae strains isolated from diseased fish. *MicrobiologyOpen*, 8(3), e00656.
- 32.Kim, J. D., Lee, N. S., Do, J. W., Kim, M. S., Seo, H. G., Cho, M., Jung, S. H., & Han, H. J. (2018). Nocardia seriolae infection in the cultured eel Anguilla japonica in Korea. *Journal of fish diseases*, *41*(11), 1745–1750.
- 33. Wang, P. C., Chen, S. D., Tsai, M. A., Weng, Y. J., Chu, S. Y., Chern, R. S., & Chen, S. C. (2009). Nocardia seriolae infection in the three striped tigerfish, Terapon jarbua (Forsskål). *Journal of fish diseases*, 32(4), 301–310.

- 34.Hou, S., Wang, W., Chen, G., Xia, L., Wang, Z., & Lu, Y. (2021). Identification of a secreted superoxide dismutase (SOD) from Nocardia seriolae which induces apoptosis in fathead minnow (FHM) cells. *Journal of fish diseases*, 44(1), 63–72.
- 35.Hou, S., Chen, G., Wang, W., Xia, L., Wang, Z., & Lu, Y. (2020). Identification of a cell-wall peptidase (NlpC/P60) from Nocardia seriolae which induces apoptosis in fathead minnow cells. *Journal of fish diseases*, 43(5), 571–581.
- 36. Yasuike, M., Nishiki, I., Iwasaki, Y., Nakamura, Y., Fujiwara, A., Shimahara, Y., Kamaishi, T., Yoshida, T., Nagai, S., Kobayashi, T., & Katoh, M. (2017). Analysis of the complete genome sequence of Nocardia seriolae UTF1, the causative agent of fish nocardiosis: The first reference genome sequence of the fish pathogenic Nocardia species. *PloS one*, *12*(3), e0173198.
- 37.Makadia, S., Patel, I., Soosaipillai, I., & Tarasiuk-Rusek, A. (2020). First Case of *Nocardia pseudobrasiliensis* Causing Primary Cutaneous Nocardiosis in an Immunocompetent Patient. *Journal of investigative medicine high impact case reports*, 8, 2324709620938228.
- 38.Ruimy, R., Riegel, P., Carlotti, A., Boiron, P., Bernardin, G., Monteil, H., Wallace, R. J., Jr, & Christen, R. (1996). Nocardia pseudobrasiliensis sp. nov., a new species of Nocardia which groups bacterial strains previously identified as Nocardia brasiliensis and associated with invasive diseases. *International journal of systematic bacteriology*, 46(1), 259–264.
- 39.Sakai, K., Komaki, H., & Gonoi, T. (2015). Identification and Functional Analysis of the Nocardithiocin Gene Cluster in Nocardia pseudobrasiliensis. *PloS one*, *10*(11), e0143264.
- 40.Harent, S., Vuotto, F., Wallet, F., Flateau, C., Chopin, M. C., Faure, K., & Guery, B. (2013). Pneumonie à Nocardia pseudobrasiliensis chez un patient transplanté cardiaque [Nocardia pseudobrasiliensis pneumonia in a heart transplant recipient]. *Medecine et maladies infectieuses*, *43*(2), 85–87.
- 41.Lebeaux, D., Lanternier, F., Degand, N., Catherinot, E., Podglajen, I., Rubio, M. T., Suarez, F., Lecuit, M., Mainardi, J. L., & Lortholary, O. (2010). Nocardia pseudobrasiliensis as an emerging cause of opportunistic infection after allogeneic hematopoietic stem cell transplantation. *Journal of clinical microbiology*, *48*(2), 656–659.
- 42. Veerappan Kandasamy, V., Nagabandi, A., Horowitz, E. A., & Vivekanandan, R. (2015). Multidrug-resistant Nocardia pseudobrasiliensis presenting as multiple muscle abscesses. *BMJ case reports*, 2015, bcr2014205262.
- 43.Seol, C. A., Sung, H., Kim, D. H., Ji, M., Chong, Y. P., & Kim, M. N. (2013). The first Korean case of disseminated mycetoma caused by Nocardia pseudobrasiliensis in a patient on long-term corticosteroid therapy for the

treatment of microscopic polyangiitis. *Annals of laboratory medicine*, *33*(3), 203–207.

- 44.Zhu, J. W., Zhou, H., Jia, W. Q., You, J., & Xu, R. X. (2020). A clinical case report of brain abscess caused by Nocardia brasiliensis in a non-immunocompromised patient and a relevant literature review. *BMC infectious diseases*, 20(1), 328.
- 45. Mangieri, N. A., Guevara Nuñez, D., Echavarría, G., Bertona, E., Castello, L., Benchetrit, G., & De Paulis, A. N. (2021). Nocardiosis esporotricoide por Nocardia brasiliensis [Sporotrichoid nocardiosis by Nocardia brasiliensis]. *Revista Argentina de microbiologia*, *53*(1), 43–47.
- 46. Verma, P., & Jha, A. (2019). Mycetoma: reviewing a neglected disease. *Clinical and experimental dermatology*, *44*(2), 123–129.
- 47.Salinas-Carmona M. C. (2000). Nocardia brasiliensis: from microbe to human and experimental infections. *Microbes and infection*, *2*(11), 1373–1381.
- 48.Johansen, M. D., Herrmann, J. L., & Kremer, L. (2020). Non-tuberculous mycobacteria and the rise of Mycobacterium abscessus. *Nature reviews*. *Microbiology*, 18(7), 392–407.
- 49.Meir, M., & Barkan, D. (2020). Alternative and Experimental Therapies of *Mycobacterium abscessus* Infections. *International journal of molecular sciences*, *21*(18), 6793.
- 50.Strnad, L., & Winthrop, K. L. (2018). Treatment of Mycobacterium abscessus Complex. *Seminars in respiratory and critical care medicine*, *39*(3), 362–376.
- 51. Yonekawa, A., Miyake, N., Minami, J., Murakami, D., Fukano, H., Hoshino, Y., Kubo, K., Chong, Y., Akashi, K., & Shimono, N. (2022). Parotitis caused by Mycobacteroides abscessus subspecies abscessus. *Auris, nasus, larynx*, 49(3), 525–528.
- 52.Degiacomi, G., Sammartino, J. C., Chiarelli, L. R., Riabova, O., Makarov, V., & Pasca, M. R. (2019). *Mycobacterium abscessus*, an Emerging and Worrisome Pathogen among Cystic Fibrosis Patients. *International journal* of molecular sciences, 20(23), 5868.
- 53. Yoshida, S., Morizumi, S., Sumitomo, K., & Shinohara, T. (2021). Tracheobronchopathia Osteochondroplastica Complicated with Mycobacteroides abscessus Pulmonary Disease. *Internal medicine (Tokyo, Japan)*, *60*(18), 3051–3052.
- 54.de Carvalho, C. C., & da Fonseca, M. M. (2005). The remarkable Rhodococcus erythropolis. *Applied microbiology and biotechnology*, *67*(6), 715–726.
- 55.Ma, X., Duan, D., Wang, X., Cao, J., Qiu, J., & Xie, B. (2021). Degradation of *Rhodococcus erythropolis* SY095 modified with functional magnetic Fe₃O₄ nanoparticles. *Royal Society open science*, *8*(12), 211172.

- 56.Korzhenkov, A. A., Bakhmutova, E. D., Izotova, A. O., Bavtushnyi, A. A., Sidoruk, K. V., Patrusheva, E. V., Patrushev, M. V., & Toshchakov, S. V. (2021). Draft Genome Sequence of Rhodococcus erythropolis VKPM Ac-1659, a Putative Oil-Degrading Strain Isolated from Polluted Soil in Siberia. *Microbiology resource announcements*, 10(29), e0053521.
- 57. Thatcher, L. F., Myers, C. A., O'Sullivan, C. A., & Roper, M. M. (2017). Draft Genome Sequence of *Rhodococcus* sp. Strain 66b. *Genome announcements*, 5(21), e00229-17.
- 58.Cunningham-Oakes, E., Pointon, T., Murphy, B., Connor, T. R., & Mahenthiralingam, E. (2020). Genome Sequence of Pluralibacter gergoviae ECO77, a Multireplicon Isolate of Industrial Origin. *Microbiology resource announcements*, 9(9), e01561-19.
- 59. Chan, K. G., Tee, K. K., Yin, W. F., & Tan, J. Y. (2014). Complete Genome Sequence of Pluralibacter gergoviae FB2, an N-Acyl Homoserine Lactone-Degrading Strain Isolated from Packed Fish Paste. *Genome announcements*, *2*(6), e01276-14.
- 60.Khashei, R., Edalati Sarvestani, F., Malekzadegan, Y., & Motamedifar, M. (2020). The first report of *Enterobacter gergoviae* carrying *bla*_{NDM-1} in Iran. *Iranian journal of basic medical sciences*, *23*(9), 1184–1190.
- 61.Périamé, M., Pagès, J. M., & Davin-Regli, A. (2015). Enterobacter gergoviae membrane modifications are involved in the adaptive response to preservatives used in cosmetic industry. *Journal of applied microbiology*, *118*(1), 49–61.
- 62.Périamé, M., Philippe, N., Condell, O., Fanning, S., Pagès, J. M., & Davin-Regli, A. (2015). Phenotypic changes contributing to Enterobacter gergoviae biocide resistance. *Letters in applied microbiology*, *61*(2), 121–129.
- 63.Kesieme, E. B., Kesieme, C. N., Akpede, G. O., Okonta, K. E., Dongo, A. E., Gbolagade, A. M., & Eluehike, S. U. (2012). Tension Pneumatocele due to Enterobacter gergoviae Pneumonia: A Case Report. *Case reports in medicine*, 2012, 808630.
- 64. Freire, M. P., de Oliveira Garcia, D., Cury, A. P., Spadão, F., Di Gioia, T. S., Francisco, G. R., Bueno, M. F., Tomaz, M., de Paula, F. J., de Faro, L. B., Piovesan, A. C., Rossi, F., Levin, A. S., David Neto, E., Nahas, W. C., & Pierrotti, L. C. (2016). Outbreak of IMP-producing carbapenem-resistant Enterobacter gergoviae among kidney transplant recipients. *The Journal of antimicrobial chemotherapy*, *71*(9), 2577–2585.
- 65.Shinjo, R., Uesaka, K., Ihara, K., Loshakova, K., Mizuno, Y., Yano, K., & Tanaka, A. (2016). Complete Genome Sequence of Kosakonia sacchari Strain BO-1, an Endophytic Diazotroph Isolated from a Sweet Potato. *Genome announcements*, *4*(5), e00868-16.

- 66.Mezzatesta, M. L., Gona, F., & Stefani, S. (2012). Enterobacter cloacae complex: clinical impact and emerging antibiotic resistance. *Future microbiology*, *7*(7), 887–902.
- 67.Ranawat, B., Mishra, S., & Singh, A. (2021). Enterobacter hormaechei (MF957335) enhanced yield, disease and salinity tolerance in tomato. *Archives of microbiology*, *203*(5), 2659–2667.
- 68.Ranawat, B., Bachani, P., Singh, A., & Mishra, S. (2021). Enterobacter hormaechei as Plant Growth-Promoting Bacteria for Improvement in Lycopersicum esculentum. *Current microbiology*, *78*(4), 1208–1217.
- 69.Zhang, Q., Wang, S., Zhang, X., Zhang, K., Liu, W., Zhang, R., & Zhang, Z. (2021). Enterobacter hormaechei in the intestines of housefly larvae promotes host growth by inhibiting harmful intestinal bacteria. *Parasites & vectors*, *14*(1), 598.
- 70.Gou, J. J., Liu, N., Guo, L. H., Xu, H., Lv, T., Yu, X., Chen, Y. B., Guo, X. B., Rao, Y. T., & Zheng, B. W. (2020). Carbapenem-Resistant *Enterobacter hormaechei* ST1103 with IMP-26 Carbapenemase and ESBL Gene *bla*_{SHV-178}. *Infection and drug resistance*, *13*, 597–605.
- 71.Monowar, T., Rahman, M. S., Bhore, S. J., & Sathasivam, K. V. (2021). Endophytic Bacteria *Enterobacter hormaechei* Fabricated Silver Nanoparticles and Their Antimicrobial Activity. *Pharmaceutics*, 13(4), 511.
- 72. Wang, Z., Duan, L., Liu, F., Hu, Y., Leng, C., Kan, Y., Yao, L., & Shi, H. (2020). First report of Enterobacter hormaechei with respiratory disease in calves. *BMC veterinary research*, *16*(1), 1.
- 73.Bonnin, R. A., Girlich, D., Jousset, A. B., Emeraud, C., Creton, E., Gauthier, L., Jové, T., Dortet, L., & Naas, T. (2021). Genomic analysis of VIM-2-producing Enterobacter hormaechei subsp. steigerwaltii. *International journal of antimicrobial agents*, 57(3), 106285.
- 74.Gao, W., Howden, B. P., & Stinear, T. P. (2018). Evolution of virulence in Enterococcus faecium, a hospital-adapted opportunistic pathogen. *Current opinion in microbiology*, *41*, 76–82.
- 75.Gök, Ş. M., Türk Dağı, H., Kara, F., Arslan, U., & Fındık, D. (2020). Klinik Örneklerden İzole Edilen Enterococcus faecium ve Enterococcus faecalis İzolatlarının Antibiyotik Direnci ve Virülans Faktörlerinin Araştırılması [Investigation of Antibiotic Resistance and Virulence Factors of Enterococcus faecium and Enterococcus faecalis Strains Isolated from Clinical Samples]. *Mikrobiyoloji bulteni*, 54(1), 26–39.
- 76.Freitas, A. R., Pereira, A. P., Novais, C., & Peixe, L. (2021).
 Multidrug-resistant high-risk Enterococcus faecium clones: can we really define them?. *International journal of antimicrobial agents*, *57*(1), 106227.
- 77.Freitas, A. R., Pereira, A. P., Novais, C., & Peixe, L. (2021).
 Multidrug-resistant high-risk Enterococcus faecium clones: can we really define them?. *International journal of antimicrobial agents*, 57(1), 106227.

- 78. Trościańczyk, A., Nowakiewicz, A., Gnat, S., Łagowski, D., Osińska, M., & Chudzik-Rząd, B. (2021). Comparative study of multidrug-resistant *Enterococcus faecium* obtained from different hosts. *Journal of medical microbiology*, 70(3), 10.1099/jmm.0.001340.
- 79. van Hal, S. J., Willems, R., Gouliouris, T., Ballard, S. A., Coque, T. M., Hammerum, A. M., Hegstad, K., Westh, H. T., Howden, B. P., Malhotra-Kumar, S., Werner, G., Yanagihara, K., Earl, A. M., Raven, K. E., Corander, J., Bowden, R., & Enterococcal Group (2021). The global dissemination of hospital clones of Enterococcus faecium. *Genome medicine*, *13*(1), 52.
- 80.Montironi, I. D., Moliva, M. V., Campra, N. A., Raviolo, J. M., Bagnis, G., Cariddi, L. N., & Reinoso, E. B. (2020). Characterization of an Enterococcus faecium strain in a murine mastitis model. *Journal of applied microbiology*, *128*(5), 1289–1300.
- 81.Yu, Z., Shi, D., Liu, W., Meng, Y., & Meng, F. (2020). Metabolome responses of Enterococcus faecium to acid shock and nitrite stress. *Biotechnology and bioengineering*, *117*(11), 3559–3571.
- 82. Duarte, B., Pereira, A. P., Freitas, A. R., Coque, T. M., Hammerum, A. M., Hasman, H., Antunes, P., Peixe, L., & Novais, C. (2019). 2CS-CHX^T Operon Signature of Chlorhexidine Tolerance among Enterococcus faecium Isolates. *Applied and environmental microbiology*, 85(23), e01589-19.
- 83.Dündar H. (2016). Bacteriocinogenic Potential of Enterococcus faecium Isolated from Wine. *Probiotics and antimicrobial proteins*, 8(3), 150–160.
- 84.Salazar, G., Almeida, A., & Gómez, M. (2013). Infección de herida traumática por Cedecea lapagei: Comunicación de un caso y revisión de la literatura [Cedecea lapagei traumatic wound infection: case report and literature review]. *Revista chilena de infectologia : organo oficial de la Sociedad Chilena de Infectologia*, 30(1), 86–89.
- 85.Ramaswamy, V. V., Gummadapu, S., & Suryanarayana, N. (2019). Nosocomial pneumonia and sepsis caused by a rare organism *Cedecea lapagei* in an infant and a review of literature. *BMJ case reports*, *12*(7), e229854.
- 86. Duperret M. E. (2020). Sinusitis caused by a rare organism, *Cedecea lapagei*. *BMJ case reports*, *13*(7), e235331.
- 87.Biswal, I., Hussain, N. A., & Grover, R. K. (2015). Cedecea lapagei in a patient with malignancy: Report of a rare case. *Journal of cancer research and therapeutics*, *11*(3), 646.
- 88.Hai, P. D., Dung, N. M., Tot, N. H., Chinh, N. X., Thuyet, B. T., Hoa, L., Son, P. N., & Thanh, L. V. (2020). First report of pneumonia and septic shock caused by *Cedecea lapagei* in Vietnam. *New microbes and new infections*, 36, 100698.

- 89. Ahmad, N., Ali, S. M., & Khan, A. U. (2017). First reported New Delhi metallo-β-lactamase-1-producing Cedecea lapagei. *International journal of antimicrobial agents*, *49*(1), 118–119.
- 90. Chavez Herrera, V. R., Rosas De Silva, M. F., Orendain Alcaraz, H., Ceja Espiritu, G., Carrazco Peña, K., & Melnikov, V. (2018). Death related to Cedecea lapagei in a soft tissue bullae infection: a case report. *Journal of medical case reports*, 12(1), 328.
- 91.Dalamaga, M., Karmaniolas, K., Arsenis, G., Pantelaki, M., Daskalopoulou, K., Papadavid, E., & Migdalis, I. (2008). Cedecea lapagei bacteremia following cement-related chemical burn injury. *Burns : journal of the International Society for Burn Injuries*, 34(8), 1205–1207.
- 92.Lopez, L. A., Ibarra, B. S., de la Garza, J. A., Rada, F., Nuñez, A. I., & López, M. G. (2013). First reported case of pneumonia caused by Cedecea lapagei in America. *The Brazilian journal of infectious diseases : an official publication of the Brazilian Society of Infectious Diseases*, 17(5), 626–628.
- 93.Mathews, S. L., Epps, M. J., Blackburn, R. K., Goshe, M. B., Grunden, A. M., & Dunn, R. R. (2019). Public questions spur the discovery of new bacterial species associated with lignin bioconversion of industrial waste. *Royal Society open science*, 6(3), 180748.
- 94. Town, J., Audy, P., Boyetchko, S. M., & Dumonceaux, T. J. (2016).
 High-Quality Draft Genome Sequence of Biocontrol Strain Pantoea sp. OXWO6B1. *Genome announcements*, 4(3), e00582-16.
- 95.Coutinho, T. A., & Venter, S. N. (2009). Pantoea ananatis: an unconventional plant pathogen. *Molecular plant pathology*, *10*(3), 325–335.
- 96.Xue, Y., Hu, M., Chen, S., Hu, A., Li, S., Han, H., Lu, G., Zeng, L., & Zhou, J. (2021). *Enterobacter asburiae* and *Pantoea ananatis* Causing Rice Bacterial Blight in China. *Plant disease*, 105(8), 2078–2088.
- 97.Asselin, J., Bonasera, J. M., Helmann, T. C., Beer, S. V., & Stodghill, P. V. (2021). Complete Genome Sequence Resources for the Onion Pathogen, *Pantoea ananatis* OC5a. *Phytopathology*, *111*(10), 1885–1888.
- 98. Choi, O., Kang, B., Lee, Y., Lee, Y., & Kim, J. (2021). Pantoea ananatis carotenoid production confers toxoflavin tolerance and is regulated by Hfq-controlled quorum sensing. *MicrobiologyOpen*, *10*(1), e1143.
- 99. Weller-Stuart, T., De Maayer, P., & Coutinho, T. (2017). Pantoea ananatis: genomic insights into a versatile pathogen. *Molecular plant pathology*, *18*(9), 1191–1198.
- Athanasakopoulou, Z., Sofia, M., Giannakopoulos, A., Papageorgiou, K., Chatzopoulos, D. C., Spyrou, V., Petridou, E., Petinaki, E., & Billinis, C. (2022). ESBL-Producing *Moellerella wisconsensis*-The Contribution of Wild Birds in the Dissemination of a Zoonotic Pathogen. *Animals : an open access journal from MDPI*, *12*(3), 340.

- 101. Hickman-Brenner, F. W., Huntley-Carter, G. P., Saitoh, Y., Steigerwalt, A. G., Farmer, J. J., 3rd, & Brenner, D. J. (1984). Moellerella wisconsensis, a new genus and species of Enterobacteriaceae found in human stool specimens. *Journal of clinical microbiology*, 19(4), 460–463.
- 102. Chilton, N. B., Dergousoff, S. J., Brzezowska, V., Trost, C. N., & Dunlop, D. R. (2020). American Dog Ticks (Dermacentor variabilis) as Biological Indicators of an Association between the Enteric Bacterium Moellerella wisconsensis and Striped Skunks (Mephitis mephitis) in Southwestern Manitoba, Canada. *Journal of wildlife diseases*, 56(4), 918–921.
- 103. Cardentey-Reyes, A., Jacobs, F., Struelens, M. J., & Rodriguez-Villalobos, H. (2009). First case of bacteremia caused by Moellerella wisconsensis: case report and a review of the literature. *Infection*, 37(6), 544–546.
- Casalinuovo, F., & Musarella, R. (2009). Isolation of Moellerella wisconsensis from the lung of a goat. *Veterinary microbiology*, *138*(3-4), 401–402.
- 105. Aller, A. I., Castro, C., Medina, M. J., González, M. T., Sevilla, P., Morilla, M. D., Corzo, J. E., & Martín-Mazuelos, E. (2009). Isolation of Moellerella wisconsensis from blood culture from a patient with acute cholecystitis. *Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases*, 15(12), 1193–1194.
- 106. Hugouvieux-Cotte-Pattat, N., & Van Gijsegem, F. (2021). Diversity within the *Dickeya zeae* complex, identification of *Dickeya zeae* and *Dickeya oryzae* members, proposal of the novel species *Dickeya parazeae* sp. nov. *International journal of systematic and evolutionary microbiology*, *71*(11), 10.1099/ijsem.0.005059.
- Huang, N., Pu, X., Zhang, J., Shen, H., Yang, Q., Wang, Z., & Lin, B. (2019). In Vitro Formation of Dickeya zeae MS1 Biofilm. *Current microbiology*, 76(1), 100–107.
- 108. Hu, M., Li, J., Chen, R., Li, W., Feng, L., Shi, L., Xue, Y., Feng, X., Zhang, L., & Zhou, J. (2018). Dickeya zeae strains isolated from rice, banana and clivia rot plants show great virulence differentials. *BMC microbiology*, 18(1), 136.
- Jiang, S., Zhang, J., Yang, Q., Sun, D., Pu, X., Shen, H., Li, Q., Wang, Z., & Lin, B. (2021). Antimicrobial Activity of Natural Plant Compound Carvacrol Against Soft Rot Disease Agent Dickeya zeae. *Current microbiology*, 78(9), 3453–3463.
- 110. Feng, L., Schaefer, A. L., Hu, M., Chen, R., Greenberg, E. P., & Zhou, J. (2019). Virulence Factor Identification in the Banana Pathogen Dickeya zeae MS2. *Applied and environmental microbiology*, *85*(23), e01611-19.

- 111. Boluk, G., Arizala, D., Dobhal, S., Zhang, J., Hu, J., Alvarez, A. M., & Arif, M. (2021). Genomic and Phenotypic Biology of Novel Strains of *Dickeya zeae* Isolated From Pineapple and Taro in Hawaii: Insights Into Genome Plasticity, Pathogenicity, and Virulence Determinants. *Frontiers in plant science*, 12, 663851.
- 112. Liang, Z., Huang, L., He, F., Zhou, X., Shi, Z., Zhou, J., Chen, Y., Lv, M., Chen, Y., & Zhang, L. H. (2019). A Substrate-Activated Efflux Pump, DesABC, Confers Zeamine Resistance to Dickeya zeae. *mBio*, 10(3), e00713-19.
- 113. Hess, S., Suthaus, A., & Melkonian, M. (2015). "Candidatus Finniella" (Rickettsiales, Alphaproteobacteria), Novel Endosymbionts of Viridiraptorid Amoeboflagellates (Cercozoa, Rhizaria). *Applied and environmental microbiology*, 82(2), 659–670.
- 114. Chung, E. J., Park, J. A., Jeon, C. O., & Chung, Y. R. (2015). Gynuella sunshinyii gen. nov., sp. nov., an antifungal rhizobacterium isolated from a halophyte, Carex scabrifolia Steud. *International journal of systematic and evolutionary microbiology*, 65(Pt 3), 1038–1043.
- 115. Nishijima, M., Adachi, K., Katsuta, A., Shizuri, Y., & Yamasato, K. (2013). Endozoicomonas numazuensis sp. nov., a gammaproteobacterium isolated from marine sponges, and emended description of the genus Endozoicomonas Kurahashi and Yokota 2007. *International journal of systematic and evolutionary microbiology*, 63(Pt 2), 709–714.
- 116. Kieffer, N., Poirel, L., Fournier, C., Haltli, B., Kerr, R., & Nordmann, P. (2019). Characterization of PAN-1, a Carbapenem-Hydrolyzing Class B β-Lactamase From the Environmental Gram-Negative *Pseudobacteriovorax antillogorgiicola*. *Frontiers in microbiology*, *10*, 1673.
- 117. McCauley, E. P., Haltli, B., & Kerr, R. G. (2015). Description of Pseudobacteriovorax antillogorgiicola gen. nov., sp. nov., a bacterium isolated from the gorgonian octocoral Antillogorgia elisabethae, belonging to the family Pseudobacteriovoracaceae fam. nov., within the order Bdellovibrionales. *International journal of systematic and evolutionary microbiology*, 65(Pt 2), 522–530.
- Mielko, K. A., Jabłoński, S. J., Milczewska, J., Sands, D., Łukaszewicz, M., & Młynarz, P. (2019). Metabolomic studies of Pseudomonas aeruginosa. *World journal of microbiology & biotechnology*, 35(11), 178.
- 119. Chevalier, S., Bouffartigues, E., Bodilis, J., Maillot, O., Lesouhaitier, O., Feuilloley, M., Orange, N., Dufour, A., & Cornelis, P. (2017). Structure, function and regulation of Pseudomonas aeruginosa porins. *FEMS microbiology reviews*, 41(5), 698–722.
- 120. Sharma, G., Rao, S., Bansal, A., Dang, S., Gupta, S., & Gabrani, R. (2014). Pseudomonas aeruginosa biofilm: potential therapeutic

targets. *Biologicals : journal of the International Association of Biological Standardization*, *42*(1), 1–7.

- 121. Jurado-Martín, I., Sainz-Mejías, M., & McClean, S. (2021). *Pseudomonas aeruginosa*: An Audacious Pathogen with an Adaptable Arsenal of Virulence Factors. *International journal of molecular sciences*, 22(6), 3128.
- 122. Kang, D., & Kirienko, N. V. (2018). Interdependence between iron acquisition and biofilm formation in Pseudomonas aeruginosa. *Journal of microbiology (Seoul, Korea)*, 56(7), 449–457.
- 123. Skariyachan, S., Sridhar, V. S., Packirisamy, S., Kumargowda, S. T., & Challapilli, S. B. (2018). Recent perspectives on the molecular basis of biofilm formation by Pseudomonas aeruginosa and approaches for treatment and biofilm dispersal. *Folia microbiologica*, 63(4), 413–432.
- 124. Broberg, A., Menkis, A., & Vasiliauskas, R. (2006). Kutznerides 1-4, depsipeptides from the actinomycete Kutzneria sp. 744 inhabiting mycorrhizal roots of Picea abies seedlings. *Journal of natural products*, 69(1), 97–102.
- 125. Jiang, W., Heemstra, J. R., Jr, Forseth, R. R., Neumann, C. S., Manaviazar, S., Schroeder, F. C., Hale, K. J., & Walsh, C. T. (2011). Biosynthetic chlorination of the piperazate residue in kutzneride biosynthesis by KthP. *Biochemistry*, 50(27), 6063–6072.
- 126. Setser, J. W., Heemstra, J. R., Jr, Walsh, C. T., & Drennan, C. L. (2014). Crystallographic evidence of drastic conformational changes in the active site of a flavin-dependent N-hydroxylase. *Biochemistry*, 53(38), 6063–6077.
- 127. Duangmal, K., Thamchaipenet, A., Matsumoto, A., & Takahashi, Y. (2009). Pseudonocardia acaciae sp. nov., isolated from roots of Acacia auriculiformis A. Cunn. ex Benth. *International journal of systematic and evolutionary microbiology*, 59(Pt 6), 1487–1491.
- Labeda, D. P., Donahue, J. M., Sells, S. F., & Kroppenstedt, R. M. (2007). Lentzea kentuckyensis sp. nov., of equine origin. *International journal of systematic and evolutionary microbiology*, 57(Pt 8), 1780–1783.
- 129. Elmahdi, S., DaSilva, L. V., & Parveen, S. (2016). Antibiotic resistance of Vibrio parahaemolyticus and Vibrio vulnificus in various countries: A review. *Food microbiology*, *57*, 128–134.
- Miyamoto, K., Kawano, H., Okai, N., Hiromoto, T., Miyano, N., Tomoo, K., Tsuchiya, T., Komano, J., Tanabe, T., Funahashi, T., & Tsujibo, H. (2021). Iron-Utilization System in *Vibrio vulnificus* M2799. *Marine drugs*, *19*(12), 710.
- Metelmann, C., Metelmann, B., Gründling, M., Hahnenkamp, K., Hauk, G., & Scheer, C. (2020). Vibrio vulnificus, eine zunehmende Sepsisgefahr in Deutschland? [Vibrio vulnificus, an increasing threat of sepsis in Germany?]. *Der Anaesthesist*, 69(9), 672–678.

- 132. Yuan, Y., Feng, Z., & Wang, J. (2020). *Vibrio vulnificus* Hemolysin: Biological Activity, Regulation of *vvhA* Expression, and Role in Pathogenesis. *Frontiers in immunology*, *11*, 599439.
- Baker-Austin, C., & Oliver, J. D. (2018). Vibrio vulnificus: new insights into a deadly opportunistic pathogen. *Environmental microbiology*, 20(2), 423–430.
- 134. Li, G., & Wang, M. Y. (2020). The role of Vibrio vulnificus virulence factors and regulators in its infection-induced sepsis. *Folia microbiologica*, *65*(2), 265–274.
- 135. Phillips, K. E., & Satchell, K. J. (2017). Vibrio vulnificus: From Oyster Colonist to Human Pathogen. *PLoS pathogens*, *13*(1), e1006053.
- 136. Fukami, K., Takagi, F., Shimizu, S., Ishigo, K., Takahashi, M., & Horikawa, T. (2021). Isolation of bacteria able to degrade poly-hydroxybutyrate-co-hydroxyhexanoate, and the inhibitory effects of the degradation products on shrimp pathogen Vibrio penaeicida. *Microbial pathogenesis*, *160*, 105167.
- 137. Fukami, K., Takagi, F., Sonoda, K., Okamoto, H., Kaneno, D., Horikawa, T., & Takita, M. (2021). Effects of the Monomeric Components of Poly-hydroxybutyrate-co-hydroxyhexanoate on the Growth of *Vibrio penaeicida* In Vitro and on the Survival of Infected Kuruma Shrimp (*Marsupenaeus japonicus*). *Animals : an open access journal from MDPI*, *11*(2), 567.
- 138. Kawato, S., Nozaki, R., Kondo, H., & Hirono, I. (2018). Draft Genome Sequence of Vibrio penaeicida Strain TUMSAT-NU1, Isolated from Diseased Shrimp in Japan. *Genome announcements*, *6*(11), e00153-18.
- 139. Goarant, C., & Merien, F. (2006). Quantification of Vibrio penaeicida, the etiological agent of Syndrome 93 in New Caledonian shrimp, by real-time PCR using SYBR Green I chemistry. *Journal of microbiological methods*, 67(1), 27–35.
- 140. Aguirre-Guzmán, G., Ascencio, F., & Saulnier, D. (2005). Pathogenicity of Vibrio penaeicida for white shrimp Litopenaeus vannamei: a cysteine protease-like exotoxin as a virulence factor. *Diseases of aquatic organisms*, 67(3), 201–207.
- 141. Kadowaki, T., Inagawa, H., Kohchi, C., Nishizawa, T., Takahashi, Y., & Soma, G. (2011). Anti-lipopolysaccharide factor evokes indirect killing of virulent bacteria in kuruma prawn. *In vivo (Athens, Greece)*, *25*(5), 741–744.
- 142. Thompson, F. L., Hoste, B., Thompson, C. C., Goris, J., Gomez-Gil, B., Huys, L., De Vos, P., & Swings, J. (2002). Enterovibrio norvegicus gen. nov., sp. nov., isolated from the gut of turbot (Scophthalmus maximus) larvae: a new member of the family Vibrionaceae. *International journal of systematic and evolutionary microbiology*, *52*(Pt 6), 2015–2022.

- 143. Thompson, F. L., Thompson, C. C., Naser, S., Hoste, B., Vandemeulebroecke, K., Munn, C., Bourne, D., & Swings, J. (2005). Photobacterium rosenbergii sp. nov. and Enterovibrio coralii sp. nov., vibrios associated with coral bleaching. *International journal of systematic and evolutionary microbiology*, 55(Pt 2), 913–917.
- 144. Pascual, J., Macián, M. C., Arahal, D. R., Garay, E., & Pujalte, M. J. (2009). Description of Enterovibrio nigricans sp. nov., reclassification of Vibrio calviensis as Enterovibrio calviensis comb. nov. and emended description of the genus Enterovibrio Thompson et al. 2002. *International journal of systematic and evolutionary microbiology*, 59(Pt 4), 698–704.
- 145. Bastian M., Heymann S., Jacomy M. (2009). Gephi: an open source software for exploring and manipulating networks. International AAAI Conference on Weblogs and Social Media.
- 146. Katoh, K., Rozewicki, J., & Yamada, K. D. (2019). MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Briefings in bioinformatics*, *20*(4), 1160-1166.
- 147. Xie, H. (2008). Activity assay of membrane transport proteins. *Acta biochimica et biophysica Sinica*, 40(4), 269-277.
- 148. Letunic, I., & Bork, P. (2021). Interactive Tree Of Life (iTOL) v5: an online tool for phylogenetic tree display and annotation. *Nucleic acids research*, *49*(W1), W293-W296.
- 149. Ashkenazy, H., Abadi, S., Martz, E., Chay, O., Mayrose, I., Pupko, T., & Ben-Tal, N. (2016). ConSurf 2016: an improved methodology to estimate and visualize evolutionary conservation in macromolecules. *Nucleic acids research*, *44*(W1), W344-W350.
- 150. Berezin, C., Glaser, F., Rosenberg, J., Paz, I., Pupko, T., Fariselli, P., ... & Ben-Tal, N. (2004). ConSeq: the identification of functionally and structurally important residues in protein sequences. *Bioinformatics*, 20(8), 1322-1324.
- 151. Perkins, S. J. (1986). Protein volumes and hydration effects: the calculations of partial specific volumes, neutron scattering matchpoints and 280-nm absorption coefficients for proteins and glycoproteins from amino acid sequences. *European Journal of Biochemistry*, *157*(1), 169-180.
- 152. Hammer, Ø., Harper, D. A., & Ryan, P. D. (2001). PAST: Paleontological statistics software package for education and data analysis. *Palaeontologia electronica*, *4*(1), 9.
- 153. Kim, S. E., Lee, J., An, J. U., Kim, T. H., Oh, C. W., Ko, Y. J., ... & Oh, D. K. (2022). Regioselectivity of an arachidonate 9S-lipoxygenase from Sphingopyxis macrogoltabida that biosynthesizes 9S, 15S-and 11S, 17S-dihydroxy fatty acids from C20 and C22 polyunsaturated fatty acids. *Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids*, 1867(3), 159091.

- 154. Ivanov, I., Heydeck, D., Hofheinz, K., Roffeis, J., O'Donnell, V. B., Kuhn, H., & Walther, M. (2010). Molecular enzymology of lipoxygenases. *Archives of biochemistry and biophysics*, 503(2), 161-174.
- 155. Morello, E., Pérez-Berezo, T., Boisseau, C., Baranek, T., Guillon, A., Brea, D., ... & Si-Tahar, M. (2019). Pseudomonas aeruginosa lipoxygenase LoxA contributes to lung infection by altering the host immune lipid signaling. *Frontiers in microbiology*, 10, 1826.
- 156. Dawkins, R. (1976) *The Selfish Gene*. New York: Oxford University Press.
- 157. Banthiya, S., Kalms, J., Yoga, E. G., Ivanov, I., Carpena, X., Hamberg, M., ... & Scheerer, P. (2016). Structural and functional basis of phospholipid oxygenase activity of bacterial lipoxygenase from Pseudomonas aeruginosa. *Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids*, 1861(11), 1681-1692.
- 158. Vidal-Mas, J., Busquets, M., & Manresa, A. (2005). Cloning and expression of a lipoxygenase from Pseudomonas aeruginosa 42A2. *Antonie van Leeuwenhoek*, 87(3), 245-251.
- 159. An, J. U., Kim, B. J., Hong, S. H., & Oh, D. K. (2015). Characterization of an omega-6 linoleate lipoxygenase from Burkholderia thailandensis and its application in the production of 13-hydroxyoctadecadienoic acid. *Applied microbiology and biotechnology*, *99*(13), 5487-5497.
- 160. Yan, C., & Xie, D. (2015). Jasmonate in plant defence: sentinel or double agent?. *Plant biotechnology journal*, *13*(9), 1233-1240.
- 161. Chini, A., Cimmino, A., Masi, M., Reveglia, P., Nocera, P., Solano, R., & Evidente, A. (2018). The fungal phytotoxin lasiojasmonate A activates the plant jasmonic acid pathway. *Journal of experimental botany*, 69(12), 3095-3102.
- 162. Zheng, D., Liwinski, T., & Elinav, E. (2020). Interaction between microbiota and immunity in health and disease. *Cell research*, *30*(6), 492-506.
- 163. Kurakin, G. (2021) Bacterial oxylipins: a key to multicellularity and to combating antimicrobial resistance?. Nature Portfolio Microbiology Community. URL: https://microbiology.community.pature.com/posts/bacterial-oxylipins-a-key

https://microbiologycommunity.nature.com/posts/bacterial-oxylipins-a-key-t o-multicellularity-and-to-combatting-the-antimicrobial-resistance. Accessed on 21.06.2022.