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### Increased levels of circulating neurotoxic metabolites in patients with mild Covid19

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#### 38 Abstract

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40 SARS-CoV-2 corona virus causes a multi-faceted and poorly defined clinical and 41 pathological phenotype involving hyperinflammation, cytokine release, and long-term 42 cognitive deficits, with an undefined neuropathological mechanism. Inflammation 43 increases the activity of the kynurenine pathway, which is linked to neurodegenerative 44 and psychiatric disorders. We sought to determine whether the kynurenine pathway is 45 impacted in patients with mild COVID-19, leading to elevated neurotoxic metabolites in 46 blood, and whether such changes are associated with pro-inflammatory cytokines. Serum 47 samples were taken from 150 patients and analyzed by ELISA and ultra-high 48 performance liquid chromatography (UHPLC). The data were analyzed using multiple 49 linear regression models adjusted for age and sex. We found increased levels of 50 kynurenine, guinolinic acid and 3-hydroxykynurenine in serum from patients with mild 51 COVID-19, together with increased levels of IL-6, ICAM-1, VCAM-1 and neopterin. The 52 levels of neurotoxic metabolites were significantly associated with key inflammatory 53 cytokines including IL-6 and TNF $\alpha$ . The COVID-19 risk-factor hypertension was 54 associated with the highest levels of neurotoxic metabolites in plasma. These neuroactive 55 metabolites could be part of the pathological mechanisms underlying cognitive 56 impairment during and post-COVID and should be explored as potential biomarkers for 57 long-COVID symptoms.

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59 Keywords: Mild COVID-19, Tryptophan metabolism, 3-HK, quinolinic acid

#### 60 Introduction

61 COVID-19, caused by SARS-CoV-2, can lead to systemic disease, including pneumonia, 62 acute respiratory distress syndrome, and impaired consciousness (1). COVID-19 also 63 leads to an activation of innate and adaptive immune responses which result in a 64 substantial inflammatory response (2). In a certain percentage of patients, symptoms 65 persist over time with the potential of symptoms lasting months and up to years after initial illness. In these patients, there is a predominance for those effected by the illness to 66 present neuropsychiatric symptoms (3). Recent studies have shown an increased risk for 67 68 acute and long-term sequalae after COVID-19 in both vaccinated and unvaccinated 69 patients (4). However, there is currently a lack of understanding for the factors that drive 70 the neuropsychiatric symptoms present during acute and long-term COVID-19. Thus, 71 there is an urgent need to identify biomarkers that can indicate the disease progression 72 during long-COVID, as well as contribute to the understanding of its underlying pathology. 73

74 The kynurenine pathway is the major route for tryptophan (TRP) metabolism, and it 75 contributes to several fundamental biological processes, all converging in energy 76 metabolism. Infections and inflammatory conditions can alter the activity of the enzymes 77 in this pathway (5). Several neuroactive metabolites of the kynurenine pathway that bind 78 neuronal receptors have an underlying role in both neurological and psychiatric symptoms 79 (5). As such, quinolinic acid (QUIN) is an agonist of the glutamatergic N-methyl-D-80 aspartate (NMDA) receptor and kynurenic acid (KYNA) is an antagonist of the same 81 receptor (6). In high concentrations, QUIN induces excitotoxic neuronal death by allowing 82 excessive amounts of calcium to enter the cell (6). While KYNA blocks the cholinergic  $\alpha$ 7

nicotinic receptor, it also antagonizes the glycine site of the NMDA-receptor, thus
preventing calcium influx (7, 8).

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86 Another kynurenine pathway metabolite, 3-hydroxykynurenine (3-HK) is also neurotoxic 87 and pro-inflammatory (9). 3-HK promotes reactive oxygen species (ROS) generation 88 through several oxidative conversions (10), and 3-HK can accelerate endothelial cell 89 apoptosis (11). The neurotoxic effects of QUIN and 3-HK are additive (5). Entry of QUIN 90 and KYNA into the central nervous system (CNS) is likely partially prohibited by an intact 91 blood brain barrier (BBB). KYN (the metabolite produced from TRP metabolism) and 3-92 HK, can freely pass the BBB (12). Within the brain, KYN is metabolized to KYNA by 93 astrocytes (5, 13), or to 3-HK and then further to QUIN by microglia and macrophages (5, 94 14). SARS-CoV-2 infection is known to affect the KYN levels by inducing the production 95 of the pro-inflammatory cytokine, interferon-gamma (IFN-x) that stimulates the rate-96 limiting enzyme in the kynurenine pathway, indoleamine-2,3-dioxygenase (IDO), thus 97 affecting KYN levels (15).

98 As previously mentioned, SARS-CoV-2 can lead to a substantial inflammatory response. 99 which affects the levels of different metabolites from the kynurenine pathway. However, 100 there are also other proteins involved in the inflammatory response, such as the 101 intercellular cell adhesion molecule-1 (ICAM-1) and the vascular cell adhesion protein-1 102 (VCAM-1), in which we were interested. Both ICAM-1 and VCAM-1 are cell surface 103 glycoproteins that govern immune cell migration to sites of inflammation and T-cell-104 mediated immunity in tissues (16-18). They are expressed in vascular endothelial cells 105 and in response to inflammation, their expression is induced in epithelial and immune 106 cells (19, 20). ICAM-1 and VCAM-1 play a central role in leukocyte trafficking, lymphocyte 107 activation and several immune responses, and the upregulation of ICAM-1 is a signature event during inflammation (21). A recent study showed that monocyte-derived 108 109 macrophages drive the inflammatory response to SARS-CoV-2 and long-term changes 110 in inflammatory response of monocytes can be detected in convalescent SARS-CoV-2 111 patients following mild infection (22). Therefore, it is of interest to determine whether the 112 levels of ICAM-1 and VCAM-1 are affected in patients with mild COVID-19, and whether 113 they are linked to kynurenine pathway activation.

114 In the current study, we analyzed serum samples from 150 patients, 44 of whom tested 115 positive for COVID-19 but exhibited mild disease and were non-hospitalized. We sought 116 to determine whether the production of neuroactive and neurotoxic metabolites along the 117 kynurenine pathway, as well as several proteins involved in inflammation, is altered in 118 patients with mild COVID-19, as this could be associated to the underlying 119 neuropsychiatry symptoms. Kynurenine pathway metabolites are correlated with the 120 severity and predicted negative outcomes of symptoms in COVID-19 patients; and could 121 potentially serve as biomarkers or predictors of neuropsychiatric long-covid symptoms.

#### 123 Results

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#### 125 **Demographics of study participants**

126 Serum samples were taken from 150 individuals, 44 were diagnosed with mild COVID-19 127 (defined for this purpose as positive, but not requiring hospitalization or treatment) and 128 106 were controls who tested negative for COVID-19. Demographics of study participants 129 are shown in Table 1. The average age of SARS-CoV-2 positive individuals was 44.2 + 130 13.1 years and controls were 45.6 + 13.4 years. 47 females (44.3%) were included as 131 controls and 25 females (56.8%) were enrolled with mild COVID-19. Of all participants, 8 132 controls (7.5%) and 7 COVID +ve (15.9%) were Asian, 5 controls (4.7%) and 0 COVID 133 +ve (0%) were Black/African, 84 controls (79.2%) and 35 COVID +ve (79.5%) were 134 White/Caucasian, 4 controls (3.8%) and 2 COVID +ve (4.5%) were Other, and 5 control 135 (4.7%) and 0 COVID +ve (0%) opted to not answer.

#### 136 Higher levels of IL-6 present in patients with mild COVID-19

Individuals with mild COVID-19 demonstrated significantly higher levels of IL-6 when compared to negative controls (ANOVA F: 5.260, p=  $0.0028^{**}$ ) (**Figure 1a**). TNF- $\alpha$  levels were not altered when both groups were compared (ANOVA F: 2.347, p= 0.075 ns) (**Figure 1b**). No significant differences were observed for the other inflammatory markers including IL-13 (F: 1.847, p= 0.141 ns), IL-8 (F: 1.223, p= 0.304 ns), IFN- $\gamma$  (F: 2.054, p= 0.109 ns, data not shown). All data was corrected for age and sex.

# Higher levels of several proteins related to inflammation present in patients with mild COVID-19

The level of ICAM-1 was increased in patients who tested positive for COVID-19 when compared to the ones that did not (data corrected for sex and age, ANOVA test F: 5.823,  $p<0.001^{***}$ ) (**Figure 2a**). Similarly, patients positive for COVID-19 also showed increased levels of VCAM-1 when compared to the PCR-negative patients (data adjusted for sex and age, ANOVA F: 3.307, p= 0.022 \*). The inflammatory related protein, neopterin was also increased in COVID-19 positive patients (data adjusted for age and sex<sub>7</sub> ANOVA test F: 3.309, p= 0.022 \*) (**Figure 2b and 2c**)

## Metabolites of the kynurenine pathway are increased in patients with mild COVID154 19

155 Previous research demonstrated alterations in kynurenine pathway activity following 156 Therefore, we investigated alterations in kynurenine pathway infection (23-25). 157 metabolite levels. Kynurenine (KYN), the first metabolite of the pathway, was significantly 158 increased in patients with mild COVID-19 when compared to patients who tested negative 159 for SARS-CoV-2 (ANOVA test F: 11.195, p= 0<0.001 \*\*\*, Figure 3). Additionally, 3-HK and QUIN. were increased in patients who tested positive for COVID-19 when compared 160 to patients who did not (ANOVA test F: 3.990, p= 0.009 \*\*; F: 8.492, p<0.001 \*\*\*, 161 162 respectively, Figure 3). Further, picolinic acid (PIC) levels in COVID-19 positive patients 163 were significantly decreased when compared to COVID negative patients (ANOVA test 164 F: 4.399, p= 0.005 \*\*). Finally, anthranilic acid (AA) was also significantly increased in 165 patients with mild COVID-19 when compared to the patients who tested negative for the

virus (ANOVA test F: 4.024, p= 0.009 \*\*, Figure 2d). All data was corrected for age and
sex.

168 The ratio of several metabolites was used to further investigate the induction of the 169 kynurenine pathway. The KYN/TRP ratio was quantified to determine the induction of the 170 kynurenine pathway. When both groups were compared, COVID-19 patients showed 171 significantly increased levels of the induction of the kynurenine pathway (ANOVA test F: 172 6.377, p< 0.001 \*\*\*, Figure 3). The QUIN/TRP ratio, which has been previously identified 173 as a biomarker for neurological diseases (26), was also analyzed. COVID-19 positive 174 cases showed a statistically significant increase in this ratio when compared to the non-175 COVID-19 patients (ANOVA test F: 5.837, p<0.001 \*\*\*, Figure 3). Another neurotoxic 176 ratio that was measured was QUIN/KYNA. In this study, COVID-19 positive patients 177 presented significantly higher ratios when compared to the control group (ANOVA test F: 178 2.847, p= 0.040 \*, Figure 3). All data was corrected for age and sex.

## 179 Correlation between neurotoxic metabolites of the kynurenine pathway and pro-

#### 180 inflammatory cytokines

Previous research has shown the relationship between inflammation and the kynurenine pathway activity. Therefore, we subsequently investigated whether the levels of kynurenine metabolites and inflammatory cytokines were correlated. As shown in **Figure** 4, there was a positive correlation between the levels of TNF- $\alpha$  and KYN (Pearson correlation: 0.453; p=0.002\*\*), 3-HK (Pearson correlation: 0.527; p<0.001\*\*\*) and QUIN (Pearson correlation: 0.482; p<0.001\*\*\*). Furthermore, a positive correlation between IL-6 and both 3-HK (Pearson correlation: 0.328; p=0.03\*) and QUIN (data adjusted for age and sex, Pearson correlation: 0.418; p=0.005\*\*) was also found. All data was corrected
for age and sex.

## 190 Increased levels of neurotoxic metabolites in COVID-19 patients with hypertension

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192 We then analyzed patients who presented with hypertension, a well-known risk factor for 193 severe COVID-19, and compared them with the patients that were not hypertensive in 194 both COVID-19 positive and negative patients. Regression analysis adjusted for age and 195 sex revealed that COVID-19 hypertensive patients presented evidence of higher levels when compared to non-hypertensive COVID-19 patients in the following proteins: IL-2 196 197 (fold change 2.66, p=0.003<sup>\*\*</sup>), IL-6 (fold change 1.45, p-value=0.014<sup>\*</sup>), TNF- $\alpha$  (fold 198 change 1.38, p=0.012\*), 3-HK (fold change 1.26, p=0.08), and the QUIN/TRP ratio (fold 199 change 1.14, p = 0.089) (Figure 5). 200

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#### 204 **Discussion**

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In the current study, we investigated the inflammatory and kynurenine pathway metabolite 206 207 signatures between mild COVID-19 cases and controls. We found increases in several 208 kynurenine pathway metabolites, such as KYN, QUIN, 3-HK and PIC, together with higher 209 levels of IL-6. Additionally, the increased levels in QUIN/TRP, QUIN/KYN, QUIN/KYNA 210 and KYN/TRP, further support a recent published study by Cihan and colleagues (23). 211 Pro-inflammatory pathway proteins, along with metabolites from the kynurenine pathway 212 were significantly increased in patients that presented with both COVID-19 and 213 hypertension when compared to COVID-19 patients that were not hypertensive. Those 214 proteins included IL-2, IL-6, TNFa, ICAM-1, VCAM-1, 3-HK, QUIN, QUIN:TRP and 215 KYN:TRP.

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217 Kynurenine pathway metabolites are correlated with severity and predicted negative 218 outcomes of symptoms in COVID-19 patients; therefore, it is important to understand the 219 role of kynurenine metabolites in mild COVID-19 patients and long-haulers, in particular 220 those with neuropsychiatric symptoms (15). In this study, we found the levels of ICAM-1 221 and VCAM-1 to be significantly increased in patients with mild SARS-CoV-2, in particular 222 those with hypertension. Increased levels of ICAM-1 and VCAM-1 in mild and severe 223 cases of COVID-19 infection has already been reported in a small study (27), where it 224 was observed that the severity of COVID-19 disease was associated with increased 225 levels of ICAM-1 and VCAM-1. The main caveat with that study was the small number of 226 patients analyzed. Our new findings support the prior observations and link increases in 227 endothelial cell adhesion molecules to kynurenine pathway metabolites, such as IDO.

IDO is expressed in endothelial cells of vessel walls, and under pathological states, a decrease in IDO leads to an increase in VCAM-1 (28). Therefore, understanding how the kynurenine pathway is involved in inflammatory diseases and how it can alter the levels of ICAM-1 and VCAM-1 will be important to understand, especially in the context of COVID-19.

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234 Lionetto et al., measured the levels of KYN and TRP in the serum of healthy patients, 235 SARS-CoV-2 negative patients, and SARS-CoV-2 positive patients (29). In SARS-CoV-2 236 positive patients, the KYN/TRP ratio was higher when compared to the negative and the 237 healthy controls (29). We also found an increase in the KYN/TRP ratio in SARS-CoV-2 238 positive patients when compared to those who tested negative. The KYN/TRP ratio is 239 usually used as an indirect measure of the activity of the IDO enzyme (30); which 240 catalyzes tryptophan. Previous research has demonstrated that IDO is regulated by IFN-241 y (31, 32). We observed no differences in the levels of serum IFN-y in mild COVID-19 242 samples. However, IDO activation and gene expression has also been shown to be 243 altered by noncanonical pathways in addition to IFN-y (33). This potentially explains the 244 increased kynurenine pathway activation that we observed in our cohort. The activation 245 of the kynurenine pathway may cause long-lasting inflammation rather than the acute 246 inflammation observed early in COVID-19. Therefore, it will be of interest to follow the 247 kynurenine metabolites as markers in patients with symptoms of long-COVID.

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Together with the KYN/TRP ratio, the QUIN/TRP ratio was increased in patients with COVID-19 when compared to controls. In a previous study, Drewes and colleagues

suggest that cerebrospinal fluid (CSF) QUIN/TRP ratio could be an early, predictive 251 252 marker of CNS disease (26). In simian-immunodeficient virus (SIV)-infected macagues, 253 the ratio of QUIN/TRP was significantly increased in cases that led to severe encephalitis 254 (26). Encephalitis is one of the many potential consequences of COVID-19 patients (34); 255 and the QUIN/TRP ratio could be used to determine the outcome of patients. Our data 256 show a significant increase in this ratio in COVID-19 patients, when compared to those 257 that tested negative for the virus. We also found that the ratio between QUIN/TRP was 258 also increased in hypertensive COVID-19 patients, suggesting that patients with hypertension might be prone to a worse outcome. 259

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261 Cihan and colleagues analyzed kynurenine pathway metabolites and inflammatory 262 cytokines and found a positive correlation between IL-6 and various metabolites from the 263 kynurenine pathway (23). Our study supports a positive correlation between IL-6 and QUIN as well as between IL-6 and 3-HK. Since Cihan's study analyzed severe and ICU 264 265 cases of COVID-19 patients, it is possible that the lack of correlation between IL-6 and 266 other kynurenine metabolites in our study is because only mild COVID-19 cases were 267 analyzed. The correlation of IL-6 and kynurenine metabolites could also potentially be 268 used as a biomarker of disease severity.

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270 The correlation between IL-6, IFN- $\gamma$ , and TNF- $\alpha$  and kynurenine metabolites supports the 271 link between inflammation, SARS-CoV-2 and the kynurenine pathway. We found a 272 positive correlation between TNF- $\alpha$  and three metabolites: 3-HK, KYN, and QUIN. TNF-273  $\alpha$  affects the kynurenine pathway in patients with schizophrenia, as a positive correlation

274 between the levels of TNF- $\alpha$  and KYN has been reported (35). TNF- $\alpha$  may accelerate the 275 formation of KYN from the catabolism of TRP (35). We did not observe differences in 276 TNF-α levels between negative and positive SARS-CoV-2 patients, however, we did find 277 a correlation between TNF- $\alpha$  and KYN levels, similar to what has been reported for 278 schizophrenia (35). In this study, QUIN, 3-HK and KYN were significantly increased in 279 patients with mild COVID-19. Both QUIN and 3-HK are neurotoxic metabolites and were 280 positively correlated with TNF- $\alpha$  and IL-6 in the case of QUIN, and TNF- $\alpha$  in the case of 3-HK, it is possible that the inflammation observed in COVID-19 patients further 281 282 contributes to the neuronal damage caused by the neurotoxic metabolites.

283 To summarize, we found an increase in neurotoxic metabolites of the kynurenine pathway 284 in patients with mild COVID-19. Furthermore, the neurotoxic metabolites were correlated 285 with inflammatory markers and vascular injury markers, such as TNF- $\alpha$ , IL-6, VCAM-1 286 and ICAM-1. We hypothesize that the activation of the neurotoxic branch of the 287 kynurenine pathway might contribute to neurological, cognitive, and psychiatric 288 symptoms experienced in COVID-19 and its aftermath. We suggest that these 289 metabolites should be studied further for their potential as biomarkers of long COVID and 290 as potential contributors to the disease mechanisms underlying long COVID.

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#### 297 Materials and Methods

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299 Blood samples

300 Blood was drawn by venipuncture of the right or left antecubital vein. Blood was allowed

301 to clot at room temperature prior to centrifugation. Serum was aliquoted into cryovials and

302 immediately transferred to -80°C for storage until biological assays.

303

#### 304 Detection of tryptophan, serotonin, and kynurenine metabolites

Serum samples were mixed with extraction solution, briefly vortexed and centrifuged. The supernatant, which contains the metabolites of interest was removed and dried under reduced pressure conditions for ninety minutes in a GeneVac EZ-2 Plus speedvac (SP Scientific, Warminster,PA). Dried down extracts were then resuspended in 0.1% formic acid in Milli-Q water, once resuspended samples were centrifuged through a COSTAR Spin-X 0.22-um filter tube and transferred to an amber vial containing a glass insert.

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#### 312 Quantification of kynurenine metabolites

Kynurenine pathway metabolites (KYN, KYNA, 3-HK, QUIN, PIC, NTA, NIC, AA), TRP and serotonin were quantified using reverse phase ultra-high-performance liquid chromatography (UHPLC) coupled to a triple quadrupole mass spectrometer (1290 Infinity II LC System, 6470 Triple quadrupole, Agilent Technologies, Santa Clara, CA). Five-microliters of samples were injected onto a Vanguard HSS T3 Pre-column that was connected to an Acquity HSS T3 analytical column. Elution conditions used a combination of Solvent A (0.1% formic acid in LC/MS grade Water) and Solvent B (0.1% formic acid

- 320 in 90% LC/MS grade acetonitrile) at a flow rate of 0.4mL/min. Agilent Masshunter
- 321 Quantitative Analysis Software (v9.0, Agilent) was used to analyze and export data.
- 322 Intra-assay coefficients of variability (CV) for plasma analytes: TRP 4.1%, KYN 2.5%,
- 323 KYNA 1.6%, 3-HK 5.0%, QUIN 2.8%, PIC 3.3%, NTA 1.6%, NIC 5.4%, AA 8.4% and 5-
- 324 HT 2.2%.
- 325 Inter-assay CVs: TRP 4.8%, KYN 1.8%, KYNA 1.5%, 3-HK 4.9%, QUIN 1.8%, PIC 1.9%,
- 326 NTA 1.8%, NIC 6.5%, AA 7.2% and 5-HT 4.9%.
- 327 Lower limits of detection (LLOD) were found to be as follows: TRP 36.6 nM, KYN 2.2 nM,
- 328 KYNA 0.16 nM, 3-HK 0.29 nM, QUIN 4.15 nM, PIC 0.63 nM, NTA 0.98 nM, NIC 0.07 nM,
- 329 AA 0.98 nM and 5-HT 0.73 nM.
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- 331 Quantification of cytokines and alpha-synuclein

332 Three Meso Scale Discovery (MSD) multiplex kits (Meso Scale Diagnostics LLC, 333 Rockville, MD) were read using a MESO QuickPlex SQ 120 plate reader. Samples were 334 run in duplicate according to the manufacturer's instructions and sample concentrations 335 were generated through MSD Discovery Workbench 4.0 software. Samples below the 336 average LLOD were denoted as the average LLOD across plates. For  $\alpha$ -synuclein, the 337 U-PLEX human  $\alpha$ -synuclein kit was used. Samples were diluted 1:8 with sample diluent 338 and generated an inter-plate CV of 3%, an average intra-assay CV of 5.2%, and the LLOD 339 of 0.876 pg/mL.

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Using the MSD V-PLEX human vascular injury II kit, we quantified C-reactive protein (CRP), serum amyloid A (SAA), ICAM-1, and VCAM-1. Samples were diluted 1:1000 with

343	manufacturer's diluent reagent. The inter-plate CV was 10.3% (CRP: 8.3%, SAA: 9.7%,
344	ICAM-1: 15.2%, VCAM-1: 8.1 %), average intra-assay CV 5.6% (CRP 4.2%, SAA 6.5%,
345	ICAM-1 6.6%, and VCAM-1 5.2%), the LLOD were 1.077pg/mL CRP, 12.839 pg/mL SAA,
346	1.037 pg/mL ICAM-1, and 6.618 pg/mL VCAM-1.
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348	The MSD V-PLEX human proinflammatory I kit was used to determine the level of IFN-y,
349	interleukin (IL)-10, IL-12p70, IL-13, IL-1β, IL-2, IL-4, IL-6, IL-8, and tumor necrosis factor
350	alpha (TNF- $\alpha$ ). Samples were diluted 1:1 with sample diluent. Inter-plate CV were
351	calculated (see Supplementary Table 1), and the LLOD was IFN- $\gamma$ : 0.087 pg/mL, IL-10:
352	0.025 pg/mL, IL-12p70: 0.0385 pg/mL, IL-13: 0.209 pg/mL, IL-1 $\beta$ : 0.032 pg/mL, IL-2:
353	0.024 pg/mL, IL-4: 0.004 pg/mL, IL-6: 0.045 pg/mL, IL-8: 0.017 pg/mL, and TNF-α: 0.049
354	pg/mL.

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#### 356 Quantification of Neopterin and S100B

357 Neopterin ELISA kits were purchased from IBL America (Immuno-Biological Laboratories 358 Inc., Minneapolis, MN). Undiluted serum samples were used following the manufacturer's 359 protocol. S100B ELISA kits were purchased from Millipore (EMD Millipore, St. Louis, MO) 360 and samples were diluted 1:1. Plates were read using a Tecan Infinite M200 Pro plate 361 reader (Tecan Group Ltd, Männedorf, Switzerland). Sample concentrations were generated using a 4 Parameter Logistic Curve Calculator (AAT Bioguest, "Quest Graph 362 363 Four Parameter Logistic (4PL) Curve Calculator" 15 Jul. 2021, 364 https://www.aatbio.com/tools/four-parameter-logistic-4pl-curve-regression-online-

365 <u>calculator</u>). The average inter-plate CV for Neopterin was 12.1%, the average intra-assay

366 CV was 2.2%, and the LLOD given by the manufacturer is 0.177 ng/mL. For S100B the 367 average inter-plate CV was 5.0%, the average intra-assay CV was 1.9%, and the LLOD 368 was 2.7 pg/mL as specified by the manufacturer.

- 369
- 370 Statistical analysis

Models were adjusted for age and sex in all instances. Robust linear regressions to assess differences between PCR+ individuals with and without hypertension were analyzed via R v 4.1.0 (<u>https://cran.r-project.org/</u>). Correlation analyses and ANOVAs were done with SPSS Statistics (version 28.0.1.0). Graphs were generated using GraphPad Prism version 9.0374 (GraphPad Software, La Jolla, CA). For all tests, statistical significance was considered as p<0.05 and weak evidence as 0.05 .

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#### 378 Study approval

379 This study utilized a subset of samples from the Beaumont Health Large-Scale 380 Automated Serologic Testing for COVID-19 study (36) and was approved by the 381 Institutional Review Board (IRB) at the Beaumont Research Institute Detroit, Michigan, 382 USA (2021-110). The final cohort consisted of 150 individuals (44 Mild COVID-19 cases 383 and 106 controls) randomly selected from the registry. Individuals testing positive for 384 SARS-CoV-2 in a qPCR test at the time of sampling were assigned "Covid19 positive" 385 whereas individuals testing negative in a qPCR test for C SARS-CoV-2 at the time of 386 blood sample were assigned "Covid19 negative" for the purpose of this study.

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#### **390** Author Contributions

Conceptualization, LB SFG, PB; data curation, ESM, ARB, CDC, CF, KH; formal analysis,
ESM, LB, SFG, PB, regression analysis with R, ZM; methodology, LB; visualization, ESM,
ARB; writing—original draft, ESM, ARB; writing—review and editing, SG, MXH, JAP, PB,
SFG, LB.

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#### 410 List of abbreviations

- 411 3-HK: 3-HydroxyKynurenine
- 412 AA: Anthranilic Acid
- 413 BBB: Blood Brain Barrier
- 414 COVID-19: Coronavirus Disease (2019)
- 415 CI: Confidence Interval
- 416 CSF: Cerebrospinal Fluid
- 417 Glu: Glutamate
- 418 ICAM-1: Intercellular Adhesion Molecule 1
- 419 ICU: Intensive Care Unit
- 420 IDO: Indoleamine 2,3-Dioxygenase
- 421 IFN-y: Interferon Gamma
- 422 IL: Interleukins
- 423 KYN: Kynurenine
- 424 KYNA: Kynurenic Acid
- 425 PA: Picolinic Acid
- 426 PCR: Polymerase Chain Reaction
- 427 ROS: Reactive Oxygen Species
- 428 S100B: S100- Calcium-Binding Protein B
- 429 SAA: Serum Amyloid A
- 430 SARS-CoV-2: Severe Acute Respiratory Syndrome Coronavirus 2
- 431 SIV: Simian Immunodeficient Virus
- 432 TNF-α: Tumor Necrosis Alpha

- 433 TRP: Tryptophan
- 434 QUIN: Quinolinic acid

- **Data availability**
- 437 Data generated is available upon request.

#### **Competing interests**

P.B. has received support as a consultant from Calico Life Sciences, CureSen, Enterin Inc, Idorsia Pharmaceuticals, Lundbeck A/S, AbbVie, Fujifilm-Cellular Dynamics International, and Axial Therapeutics. He has received commercial support for research from Lundbeck A/S and F. Hoffman-La Roche. He has ownership interests in Acousort AB, Axial Therapeutics, Enterin Inc and RYNE Biotechnology. During the time that this paper was written he became an employee of F. Hoffman-La Roche, although none of the data were generated by this company. 

#### 457 **References**

458

Mao L, Jin H, Wang M, Hu Y, Chen S, He Q, et al. Neurologic Manifestations of
Hospitalized Patients With Coronavirus Disease 2019 in Wuhan, China. JAMA Neurol.
2020;77(6):683-90. doi: 10.1001/jamaneurol.2020.1127. PubMed PMID: 32275288; PubMed
Central PMCID: PMC7149362.

Anka AU, Tahir MI, Abubakar SD, Alsabbagh M, Zian Z, Hamedifar H, et al. Coronavirus
disease 2019 (COVID-19): An overview of the immunopathology, serological diagnosis and
management. Scand J Immunol. 2021;93(4):e12998. Epub 20201203. doi: 10.1111/sji.12998.
PubMed PMID: 33190302; PubMed Central PMCID: PMC7744910.

- 3. Roy D, Ghosh R, Dubey S, Dubey MJ, Benito-León J, Kanti Ray B. Neurological and
  Neuropsychiatric Impacts of COVID-19 Pandemic. Can J Neurol Sci. 2021;48(1):9-24. Epub
  20200805. doi: 10.1017/cjn.2020.173. PubMed PMID: 32753076; PubMed Central PMCID:
  PMC7533477.
- 471 4. Al-Aly Z, Bowe B, Xie Y. Long COVID after breakthrough SARS-CoV-2 infection. Nat
  472 Med. 2022. Epub 20220525. doi: 10.1038/s41591-022-01840-0. PubMed PMID: 35614233.
- 5. Schwarcz R, Bruno JP, Muchowski PJ, Wu HQ. Kynurenines in the mammalian brain:
  when physiology meets pathology. Nat Rev Neurosci. 2012;13(7):465-77. doi: 10.1038/nrn3257.
  PubMed PMID: 22678511; PubMed Central PMCID: PMC3681811.
- Lugo-Huitrón R, Blanco-Ayala T, Ugalde-Muñiz P, Carrillo-Mora P, Pedraza-Chaverrí J,
  Silva-Adaya D, et al. On the antioxidant properties of kynurenic acid: free radical scavenging
  activity and inhibition of oxidative stress. Neurotoxicol Teratol. 2011;33(5):538-47. Epub
  20110713. doi: 10.1016/j.ntt.2011.07.002. PubMed PMID: 21763768.
- Hilmas C, Pereira EF, Alkondon M, Rassoulpour A, Schwarcz R, Albuquerque EX. The
  brain metabolite kynurenic acid inhibits alpha7 nicotinic receptor activity and increases nonalpha7 nicotinic receptor expression: physiopathological implications. J Neurosci.
  2001;21(19):7463-73. doi: 10.1523/jneurosci.21-19-07463.2001. PubMed PMID: 11567036;
  PubMed Central PMCID: PMC6762893.
- 485 8. Stone TW. Neuropharmacology of quinolinic and kynurenic acids. Pharmacol Rev.
  486 1993;45(3):309-79. PubMed PMID: 8248282.
- 487 9. Wang Q, Liu D, Song P, Zou MH. Tryptophan-kynurenine pathway is dysregulated in
  488 inflammation, and immune activation. Front Biosci (Landmark Ed). 2015;20(7):1116-43. Epub
  489 20150601. doi: 10.2741/4363. PubMed PMID: 25961549; PubMed Central PMCID:
  490 PMC4911177.
- Ishii T, Iwahashi H, Sugata R, Kido R. Formation of hydroxanthommatin-derived radical
  in the oxidation of 3-hydroxykynurenine. Arch Biochem Biophys. 1992;294(2):616-22. doi:
  10.1016/0003-9861(92)90733-d. PubMed PMID: 1314547.
- 494 Wang Q, Zhang M, Ding Y, Wang Q, Zhang W, Song P, et al. Activation of NAD(P)H 11. 495 oxidase by tryptophan-derived 3-hydroxykynurenine accelerates endothelial apoptosis and 496 dysfunction in Circ Res. 2014;114(3):480-92. Epub 20131126. doi: vivo. 497 10.1161/circresaha.114.302113. PubMed PMID: 24281189; PubMed Central PMCID: 498 PMC4104160.
- 499 12. Skorobogatov K, De Picker L, Verkerk R, Coppens V, Leboyer M, Müller N, et al. Brain
  500 Versus Blood: A Systematic Review on the Concordance Between Peripheral and Central
- 501 Kynurenine Pathway Measures in Psychiatric Disorders. Front Immunol. 2021;12:716980. Epub

502 20210923. doi: 10.3389/fimmu.2021.716980. PubMed PMID: 34630391; PubMed Central
 503 PMCID: PMC8495160.

504 13. Guillemin GJ, Kerr SJ, Smythe GA, Smith DG, Kapoor V, Armati PJ, et al. Kynurenine
505 pathway metabolism in human astrocytes: a paradox for neuronal protection. J Neurochem.
506 2001;78(4):842-53. doi: 10.1046/j.1471-4159.2001.00498.x. PubMed PMID: 11520905.

Agudelo LZ, Femenía T, Orhan F, Porsmyr-Palmertz M, Goiny M, Martinez-Redondo V,
et al. Skeletal muscle PGC-1α1 modulates kynurenine metabolism and mediates resilience to
stress-induced depression. Cell. 2014;159(1):33-45. doi: 10.1016/j.cell.2014.07.051. PubMed
PMID: 25259918.

- 511 15. Oxenkrug G, Summergrad P. Peripheral kynurenines as biomarkers and targets for
  512 prevention and treatment of psychiatric conditions associated with SARS-CoV-2 infection.
  513 Personalized Medicine in Psychiatry. 2021;29-30:100088. doi:
  514 https://doi.org/10.1016/j.pmip.2021.100088.
- 515 16. Lembas A, Zawartko K, Sapuła M, Mikuła T, Kozłowska J, Wiercińska-Drapało A.
- 516 VCAM-1 as a Biomarker of Endothelial Function among HIV-Infected Patients Receiving and
- 517 Not Receiving Antiretroviral Therapy. Viruses. 2022;14(3). Epub 20220311. doi: 10.3390/v14030578. PubMed PMID: 35336985; PubMed Central PMCID: PMC8955345.
- 519 Chen YH, Lightman S, Eskandarpour M, Calder VL. Adhesion Molecule Targeted Therapy 17. 520 for Non-Infectious Uveitis. Int J Mol Sci. 2022;23(1). Epub 20220103. doi: 521 10.3390/ijms23010503. PubMed PMID: 35008929; PubMed Central PMCID: PMC8745221.
- 522 18. Kang L, Kim M, Lee YM. Expression of ICAM-1 in Blood Vascular Endothelium and
  523 Tissues in Human Premalignant Lesion and Gastric/Hepatocellular Carcinomas. Korean J
  524 Gastroenterol. 2022;79(4):170-6. doi: 10.4166/kjg.2022.008. PubMed PMID: 35473775.
- Bui TM, Wiesolek HL, Sumagin R. ICAM-1: A master regulator of cellular responses in
  inflammation, injury resolution, and tumorigenesis. J Leukoc Biol. 2020;108(3):787-99. Epub
  20200317. doi: 10.1002/jlb.2mr0220-549r. PubMed PMID: 32182390; PubMed Central PMCID:
  PMC7977775.
- 529 20. Hyun YM, Lefort CT, Kim M. Leukocyte integrins and their ligand interactions. Immunol
  530 Res. 2009;45(2-3):195-208. Epub 20090129. doi: 10.1007/s12026-009-8101-1. PubMed PMID:
  531 19184539; PubMed Central PMCID: PMC2990789.
- 532 21. Ramos TN, Bullard DC, Barnum SR. ICAM-1: isoforms and phenotypes. J Immunol.
- 533 2014;192(10):4469-74. doi: 10.4049/jimmunol.1400135. PubMed PMID: 24795464; PubMed 534 Central PMCID: PMC4015451.
- 535 Bohnacker S, Hartung F, Henkel F, Quaranta A, Kolmert J, Priller A, et al. Correction to: 22. 536 Mild COVID-19 imprints a long-term inflammatory eicosanoid- and chemokine memory in 537 monocyte-derived macrophages. Immunol. Mucosal 2022:1. Epub 20220513. doi: 538 10.1038/s41385-022-00526-7. PubMed PMID: 35562559; PubMed Central PMCID: 539 PMC9098895.
- 540 23. Cihan M, Doğan Ö, Ceran Serdar C, Altunçekiç Yıldırım A, Kurt C, Serdar MA. 541 Kynurenine pathway in Coronavirus disease (COVID-19): Potential role in prognosis. J Clin Lab
- 542 Anal. 2022;36(3):e24257. Epub 20220129. doi: 10.1002/jcla.24257. PubMed PMID: 35092710;
- 543 PubMed Central PMCID: PMC8906035.
- 544 24. Holtze M, Asp L, Schwieler L, Engberg G, Karlsson H. Induction of the kynurenine
- 545 pathway by neurotropic influenza A virus infection. J Neurosci Res. 2008;86(16):3674-83. doi:
- 546 10.1002/jnr.21799. PubMed PMID: 18655201.

547 25. Wickström R, Fowler Å, Goiny M, Millischer V, Ygberg S, Schwieler L. The Kynurenine
548 Pathway is Differentially Activated in Children with Lyme Disease and Tick-Borne Encephalitis.
549 Microorganisms. 2021;9(2). Epub 20210204. doi: 10.3390/microorganisms9020322. PubMed

550 PMID: 33557172; PubMed Central PMCID: PMC7913947.

Drewes JL, Meulendyke KA, Liao Z, Witwer KW, Gama L, Ubaida-Mohien C, et al.
Quinolinic acid/tryptophan ratios predict neurological disease in SIV-infected macaques and
remain elevated in the brain under cART. J Neurovirol. 2015;21(4):449-63. Epub 20150317. doi:
10.1007/s13365-015-0334-2. PubMed PMID: 25776527; PubMed Central PMCID:
PMC4512924.

Tong M, Jiang Y, Xia D, Xiong Y, Zheng Q, Chen F, et al. Elevated Expression of Serum
Endothelial Cell Adhesion Molecules in COVID-19 Patients. J Infect Dis. 2020;222(6):894-8. doi:
10.1093/infdis/jiaa349. PubMed PMID: 32582936; PubMed Central PMCID: PMC7337874.

559 28. Kiluk M, Lewkowicz J, Pawlak D, Tankiewicz-Kwedlo A. Crosstalk between Tryptophan 560 Metabolism via Kynurenine Pathway and Carbohydrate Metabolism in the Context of Cardio-

- 561 Metabolic Risk-Review. J Clin Med. 2021;10(11). Epub 20210604. doi: 10.3390/jcm10112484.
  562 PubMed PMID: 34199713; PubMed Central PMCID: PMC8199979.
- Lionetto L, Ulivieri M, Capi M, De Bernardini D, Fazio F, Petrucca A, et al. Increased
  kynurenine-to-tryptophan ratio in the serum of patients infected with SARS-CoV2: An
  observational cohort study. Biochim Biophys Acta Mol Basis Dis. 2021;1867(3):166042. Epub
  20201216. doi: 10.1016/j.bbadis.2020.166042. PubMed PMID: 33338598; PubMed Central
  PMCID: PMC7834629.
- 568 30. Badawy AA-B, Guillemin G. The Plasma [Kynurenine]/[Tryptophan] Ratio and 569 Indoleamine 2,3-Dioxygenase: Time for Appraisal. International Journal of Tryptophan Research. 570 2019;12:1178646919868978. doi: 10.1177/1178646919868978. PubMed PMID: 31488951.
- 31. Badawy AA. Kynurenine Pathway of Tryptophan Metabolism: Regulatory and Functional
  Aspects. Int J Tryptophan Res. 2017;10:1178646917691938. Epub 20170315. doi:
  10.1177/1178646917691938. PubMed PMID: 28469468; PubMed Central PMCID:
  PMC5398323.
- 575 32. Sarkar SA, Wong R, Hackl SI, Moua O, Gill RG, Wiseman A, et al. Induction of 576 indoleamine 2,3-dioxygenase by interferon-gamma in human islets. Diabetes. 2007;56(1):72-9. 577 doi: 10.2337/db06-0617. PubMed PMID: 17192467.
- 578 33. Liang Y, Yu Z, Song Y, Wang T, Xiao B. Indoleamine 2,3-Dioxygenase Activation by
- 579 Interferon Gamma in Vascular Endothelial Rat Cells Requires Noncanonical NF-κB Signaling.
- 580 Transplant Proc. 2019;51(6):2141-5. Epub 20190712. doi: 10.1016/j.transproceed.2019.03.043.
  581 PubMed PMID: 31307771.
- 582 34. Ellul MA, Benjamin L, Singh B, Lant S, Michael BD, Easton A, et al. Neurological
  583 associations of COVID-19. Lancet Neurol. 2020;19(9):767-83. Epub 20200702. doi:
  584 10.1016/s1474-4422(20)30221-0. PubMed PMID: 32622375; PubMed Central PMCID:
  585 PMC7332267.
- 35. Okamoto N, Natsuyama T, Igata R, Konishi Y, Tesen H, Ikenouchi A, et al. Associations
  Between the Kynurenine Pathway, Proinflammatory Cytokines, and Brain-Derived Neurotrophic
  Factor in Hospitalized Patients With Chronic Schizophrenia: A Preliminary Study. Front
  Psychiatry. 2021;12:696059. Epub 20210728. doi: 10.3389/fpsyt.2021.696059. PubMed PMID:
- 590 34393855; PubMed Central PMCID: PMC8357143.
- 591 36. Sims MD, Maine GN, Childers KL, Podolsky RH, Voss DR, Berkiw-Scenna N, et al. 592 Coronavirus Disease 2019 (COVID-19) Seropositivity and Asymptomatic Rates in Healthcare

593	Workers	s Are A	Associat	ed with J	ob Fu	nction an	d Maskin	g. Clin Infect	Dis. 2021	73(Suppl	2):S154-
594	s62. dc	oi: 10	).1093/c	id/ciaa16	684.	PubMed	PMID:	33150375;	PubMed	Central	PMCID:
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#### Table 1. Demographics of the patients included in the current study

Mild SARS-CoV-2 Demographics					
	<b>SARS-CoV-2</b> <b>Negative</b> (n = 106)	SARS-CoV-2 Positive (n = 44)			
Age					
(Mean <u>+</u> SD)	45.65 <u>+</u> 13.39	44.18 <u>+</u> 13.12			
<b>Sex</b> F (%)	47 (44.3%)	25 (56.8%)			
Race n (%)					
Asian	8 (7.5%)	7 (15.9%)			
Black/African	5 (4.7%)	0 (0%)			
White/Caucasian	84 (79.2%)	35 (79.5%)			
Other	4 (3.8%)	2 (4.5%)			
No Answer	5 (4.7%)	0 (0%)			

- 620 SD: Standard Deviation
- 621 F: female



Figure 1. Significantly higher levels IL-6 in patients with mild COVID-19 when compared to controls. a) Significantly higher levels of IL-6 were found in patients with COVID-19 when compared to the negative controls (data adjusted for age and sex, ANOVA test F: 5.260, p=0.002 \*\*). b) No differences were found in the level of TNF- $\alpha$ between the two groups (data adjusted for age and sex, ANOVA test F: 2.347, p= 0.075 ns). Graphs are represented by median with 95% of confidence interval (CI).



Figure 2. Significantly higher levels of serum ICAM-1, VCAM-1, and neopterin in

**patients with COVID-19.** When the levels of all three proteins were analyzed, patients with COVID-19 presented significantly higher levels of **a**) ICAM-1 (data adjusted for age and sex, ANOVA test F: 5.823, p<0.001 \*\*\*), **b**) VCAM-1 (data adjusted for age and sex, ANOVA test F: 3.307, p= 0.022 \*), and **c**) neopterin (data adjusted for age and sex, ANOVA test F:3.309, p= 0.022\*). Graphs are represented by median with 95% of

682 confidence interval (CI).



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684 Figure 3. The kynurenine pathway is altered in patients with mild COVID-19, who present 685 increased levels of neurotoxic metabolites. Significantly increased levels of kynurenine (data adjusted for age and sex, ANOVA test F: 11.195, p<0.001 \*\*\*), 3-hydroxykynurenine (data 686 adjusted for age and sex, ANOVA test F: 3.390, p=0.009 \*\*), anthranilic acid (data adjusted for 687 age and sex, ANOVA test F: 4.024, p=0.009 \*\*), and quinolinic acid (data adjusted for age and 688 sex, ANOVA test F: 8.492, p<0.001 \*\*\*) were found in patients with mild COVID-19 when 689 690 compared to controls. When the ratio of the metabolites was analyzed, significantly increased 691 levels of KYN/TRP (data adjusted for age and sex, ANOVA test F: 6.377, p<0.001 \*\*\*) and 692 QUIN/TRP (data adjusted for age and sex, ANOVA test F: 5.837, p<0.001 \*\*\*), as well as QUIN/KYNA (data adjusted for age and sex, ANOVA test F: 2.847, p= 0.040 \*) were found in 693 694 patients with COVID-19. Graphs show the median with 95% of CI. Abbreviations: IDO, 695 Indoleamine 2,3-dioxygenase; TDO, Tryptophan 2,3-dioxygenase; KATs, Kynurenine 696 aminotransferase; KYNA, kynurenic acid; KMO, Kynurenine 3-monooxygenase; AA, anthranilic 697 acid; 3-HK, 3-hydroxykynurenine; KYNU, Kynureninase; 3-HAA, 3-hydroxyanthranilic acid; 698 3HAO, 3-hydroxyanthranilate oxidase: ACMSD, Aminocarboxymuconate-semialdehyde 699 decarboxylase, and NAD, Nicotinamide adenine dinucleotide.



Figure 4. There is correlation between inflammatory cytokines and metabolites of the kynurenine pathway in patients with mild COVID-19. Patients with mild COVID-19 present a positive correlation between TNF- $\alpha$  and kynurenine (data adjusted for age and sex, Pearson R: 0.4512; p=0.0021 \*\*), TNF- $\alpha$  and 3-hydroxykynurenine (data adjusted for age and sex, Pearson R: 0.4567; p=0.0018 \*\*), TNF- $\alpha$  and quinolinic acid (data adjusted for age and sex, Pearson R: 0.4812; p=0.0009 \*\*\*), IL-6 and 3-hydroxykynurenine (data adjusted for age and sex, Pearson R: 0.3252, p=0.0312 \*) and between IL-6 and quinolinic acid (data adjusted for age and sex, Pearson R: 0.4161, p=0.0050 \*\*).



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Figure 5. Increased metabolites and ratio in COVID-19 positive hypertensive patients when compared to non-hypertensive COVID-19 patients. Higher levels of IL-2 (fold change 2.66, p=0.003\*\*), IL-6 (fold change 1.45, p-value=0.014\*), and TNF-a (fold change 1.38, p=0.012\*) in COVID-19 hypertensive patients when compared to COVID-19 non-hypertensive patients. Weak evidence of increased levels in 3-HK (fold change 1.26, p=0.08°), and the QUIN/TRP ratio (fold change 1.14, p = 0.089°). Graphs are represented by median with 95% of confidence interval (CI).