Nucleus of the lateral olfactory tract (NLOT): a hub linking water homeostasis-associated SON-AVP circuit and neocortical regions to promote social behavior under osmotic challenge

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Abstract

Homeostatic challenges increase the drive for social interaction. The neural activity that prompts this motivation remains poorly understood. Here, we identify direct projections from hypothalamic supraoptic nucleus (SON) to the cortico-amygdalar nucleus of the lateral olfactory tract (NLOT). Dual in situ hybridization (DISH) with probes for PACAP, and VGLUT1, VGLUT2, V1a and V1b revealed a population of vasopressin-receptive PACAPergic neurons in NLOT layer 2 (NLOT2). Water deprivation (48 hours, WD48) increased sociability compared to euhydrated subjects, assessed with the three-chamber social interaction test (3CST). Fos expression immunohistochemistry showed NLOT and its main efferent regions had further increases in rats subjected to WD48+3CST. These regions strongly expressed PAC1 mRNA. Microinjections of AVP into NLOT produced similar changes in sociability to water deprivation, and these were reduced by co-injection of V1a or V1b antagonists along with AVP. We conclude that during challenge to water homeostasis, there is a recruitment of a glutamatergic-multi-peptidergic cooperative circuit that promotes social behavior.

Keywords: V1a, V1b, PAC1, VGLUT1, agranular insular cortex, claustrum, fluorogold, dual ISH

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1. INTRODUCTION

Neuropeptides acting as neurotransmitters in the brain participate in circuits which monitor multiple environmental inputs, and access brain state-related information, in order to produce highly integrated responses that are appropriately tuned to the specifics of a given situation requiring motor decision-making (action) [1]. An area of research relevant to the general question of how multiple-circuit integration leading to specific behaviors occurs, is the study of the microanatomy, neurochemistry, and cellular and behavioral functions of vasopressinergic magnocellular neurosecretory neurons (AVPMNNs) of the mammalian brain. This is because AVPMNNs have very well-defined roles as neurosecretory neurons that release vasopressin into the bloodstream for hormonally-driven osmotic regulation (hydromineral *homeostasis*), and as well, via dual projections within the brain parenchima, release vasopressin as a neurotransmitter to affect limbic, hypothalamic, and other brain circuits that participate in behavioral prioritization responsive to competing homeostatic drives, i.e. *allostasis* [2].

In previous studies, we have demonstrated that activation of the AVPMNN system in the hypothalamic paraventricular nucleus (PVN) by alteration of osmotic balance through water deprivation or salt loading activates both limbs of this dually-projecting system. This results not only in changes in blood levels of vasopressin indicative of hormonal homeostatic regulation, but changes in electrical activity and even synaptic protein expression at vasopressinergic terminal field regions of hippocampus, amygdala, locus coeruleus and other locations. These are indicative of potentially profound effects on behaviors[3-9] and thus linking the original homeostatic osmoregulatory drive, to allostatic drives that must act in concert with it.

Social behavior is one of the core factors facilitating an individual's chances of survival. However, the neural substrates underlying this behavioral are only now being uncovered. Vasopressin has been shown to participate in several aspects of social behavior. For instance, mice deficient for V1a and V1b receptors show social deficits[10-12]; Brattleboro rats lacking AVP expression have reduced social interaction [13, 14]; and altered AVP transmission has been described in pathologies characterized by social deficits [15, 16].

In contrast to the PVN, the hypothalamic supraoptic nucleus (SON), which contains another major population of AVPMNNs responsible for controlling body water balance, has been less investigated regarding ascending projections, as well as concerning its roles in behavioral adaptation. This lack of information is mainly due to its deep location at the base of the brain. Using in vivo electrophysiological recording, Inyushkin et al [17] demonstrated a

73 clear dual projection system, neurohypophysial and central, from SON AVPMNNs, and
74 suggested the existence of projections from these neurons in the SON that are much more
75 widespread and longer than had previously been suspected. SON's innervation to dorsal and
76 ventral hippocampus and central amygdala was subsequently demonstrated using the
77 fluorogold method [6, 18].
78
The nucleus of the lateral olfactory tract (NLOT) is located in the anterior cortico-
amygdalar region adjacent to the ventral surface of the brain. This structure is connected
with the main olfactory bulb, and the piriform and insular cortices, and is implicated in
feeding behavior [19]. It has been described as a three-layered structure[20]. Its heterogeneous
neuronal composition suggests different neuroepithelial origins of the cells that populate its
three layers [21].
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The aim of the present study was to evaluate whether or not NLOT is a brain region
is a brain region containing vasopressin-responsive neurons important in mediating the
social effects of AVP. By analyzing Golgi-Cox staining, we observed direct projections from
SON to the nucleus of lateral olfactory tract (NLOT) of cortico-amygdalar complex. Fluoro-
Gold injection into the NLOT revealed retrogradely labelled AVP-positive somata in SON. We
previously reported that the main neuronal population of NLOT co-expresses the
neuropeptide PACAP and vesicular glutamate transporters 1 and 2 (VGLUT1 and VGLUT2).
Here, using the dual in situ hybridization method we demonstrate that the principal
population of PACAP/VGLUT1/VGLUT2 neurons co-expresses vasopressin receptors V1a and
V1b. Considering these observations, we devised a behavioral experiment using 48 hour
water deprivation (WD48) and a three-chamber social interaction test (3CST) [22, 23], designed
to quantify the level of sociability, defined as the tendency to approach and remain proximal
to an unfamiliar conspecific (we called "rat *stranger*" hereafter). WD48 significantly
increased the social behavior. Fos expression assessment showed AVP-MNNs and PACAP-
NLOT regions and their efferents had further increases in rats subjected to WD48+3CSI. The
efferent regions of PACAP-NLOT strongly expressed PAC1 mRNA. AVP and NLOT involvement
in this increased sociability and Fos expression was further demonstrated through
microinjections of AVP, AVP+V1a or V1b antagonists, targeting NLOT, that AVP
microinjection alone produced similar increase produced by WD24 but if applied together
with V1a or V1b antagonists, the AVP-stimulated increases were ablated. These results
suggest that under situations where homeostasis is compromised by osmotic challenge,
there is a recruitment of a glutamatergic-multi-peptidergic cooperative circuit that is able to
promote behavioral adaptation through social interaction.
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110   2. EXPERIMENTAL PROCEDURES
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<u>2.1 Animals</u>

One hundred and forty-four adult male Wistar rats of 280 ± 20 g were obtained from the

local animal facility. Rats were housed four per cage in a controlled environment

(temperature 24 ºC and illumination 12 h/12 h (lights on at 7:00 to 19:00 h) with water and

food *ad libitum*). All animal procedures were approved by the *Comision de Investigacion y*

Etica de la Facultad de Medicina, Universidad Nacional Autonoma de Mexico (approval

number: CIEFM-062-2016). Efforts were taken to minimize animal suffering throughout all

experimental procedures.

<u>2.2 Social behavior assessment under water deprivation</u>

A first experiment was devised to assess the effects of water deprivation on social behavior.

Twenty-four rats were randomly assigned to 48h of water deprivation (WD48, n=14) or water

ad libitum (control, n=12). The rationale for this manipulation was that WD48 up-regulates

the metabolic activity of the hypothalamic AVPMNNs with only a modest increase in plasma

osmolarity [5, 24]. Social behavior was tested in the three-chamber social interaction test

(3CST). The test was carried out essentially as described elsewhere [22, 23]. Briefly, testing was

conducted in a three-chambered box made of acrylic (100 cm x 30 cm x 30 cm), the central

chamber (20 cm x 30 cm) and two distal chambers (40 cm x 30 cm) each. Chambers were

connected via open doors (10 cm x 10 cm). One day before the experiment, the rats were

allowed to explore the apparatus freely for 10 min to be habituated to the experimental

device. On the day of the experiment, an empty cage was placed in a compartment (*non-*

social compartment), a similar cage containing a male rat *stranger* was placed in the opposite

chamber (*social* compartment). The experiment started by putting the experimental subjects

in the central chamber of the apparatus and allowed access to all chambers for 10 minutes.

The time the rat spent in the *social* chamber and the number of approaches/sniffing to the

rat *stranger* were scored.

<u>2.3 Immunohistochemistry and immunofluorescence</u>

Rats were deeply anesthetized with sodium pentobarbital (63 mg/kg, Sedalpharma, México)

and perfused transaortically with 0.9% NaCl followed by cold fixative containing 4%

paraformaldehyde in 0.1 M sodium phosphate buffer (PB, pH 7.4) plus 15% v/v saturated

picric acid solution for 15 min. Brains were immediately removed, blocked, thoroughly rinsed

with PB 0.1M, and sectioned at 70 µm from forebrain to hindbrain using a Leica VT 1000S

vibratome. Sections were blocked with 20% normal donkey serum in Tris-buffered (0.05 M,

pH 7.4), NaCl (0.9%), plus 0.3% of Triton X-100 (TBST) for 1h at room temperature,

immunoreacted overnight with rabbit anti-Fos primary antibody (SC-52, 1:1000, Santa Cruz

Biotechnology, Santa Cruz, CA, USA) or rabbit anti vasopressin primary antibody (kind gift
from Professor Ruud Buijs), washed and incubated with secondary biotinylated antibody
(goat anti-rabbit, vector BA1000) followed by incubation in Vectastain Elite ABC kit solution
(Vector Labs, Burlingame, CA, USA) and detection with a DAB-peroxidase reaction. For
Immunofluorescent processing we used a secondary donkey anti rabbit Alexa 594
fluorescent antibody. Some photomicrographs were presented digitally inverted (negative
mode) to enhance the visibility of DAB labelled AVP fibers.

2.4 Neuronal Activation assessment after 3CST

To assess the pattern of neuronal activation induced in the brain by the 3CST and/or water
deprivation, we formed four groups of n=5 rats: Control (same conditions as other groups
but undisturbed until perfusion time); Social interaction (rats that were under ad-libitum
water access until the 3CST); 48h WD (rats deprived of water during the 48h previous to the
perfusion with no other disturbance) and 48h WD + social Interaction (rats that underwent
48h of water deprivation before the 3CST experiment). Rats evaluated by the 3CST were
perfused 60 min after finishing the behavioral test; the other groups were perfused at the
same circadian time. After perfusion, immunohistochemistry against Fos was performed as
described above.

Regions of interest were identified using a low magnification objective with
referencing to a standard rat brain stereotaxic atlas [25]. The average number of Fos+ nuclei
in each identified region was quantified under a 40x objective ($0.196 \, mm^2$). Two comparable
fields per area in each rat were quantified by two independent researchers. A table
comparing the four groups was constructed with semi-quantitative criteria. i.e., "+": 1 to 25
per field; "++": 26 to 50 per field; "+++": 51 to 75 per field; "++++": 76 to 100 per field;
"+++++": >100 per field (field area: 0.196 mm2, for simplicity we used $0.2 \, mm^2$).

2.5 Fluorogold retrograde tracing

The Fluorogold retrograde method was performed as previously reported [6, 18]. Rats (male,
n=4, 280-300g) were deeply anaesthetized using a 1:1 mixture of xylazine (20 mg/ml, Procin,
Mexico) and ketamine (100 mg/ml, Inoketam, Virbac, Mexico) administered as a dose of 1
ml/kg body weight intraperitoneally. Rats were fixed in a stereotaxic frame, and the
retrograde tracer Fluoro-Gold (FG, Fluorochrome, LLC, Denver, Colorado 80218, USA),
dissolved to a concentration of 1% in 0.1 M cacodylate buffer (pH 7.5), was delivered into
the nucleus of the lateral olfactory tract (NLOT) using a glass micropipette with an inner tip
diameter of around 40 μm. Current used for iontophoresis was of 0.1 μA with a 5-s pulse-

duration and a 50% duty cycle during 20 min. The coordinates for positioning the pipette in
NLOT were: AP -1.4 mm, ML ± 3.2 mm from bregma, and DV -9.4 mm from the skull surface,
according to a stereotaxic atlas [25]. Additional time-lapse of 10 min was allowed to prevent
backflow of tracer up the injection track. Before recovering from anesthesia, rats received
0.4 mg/kg i.p. ketorolac (Apotex, Mexico) and 50 mg/ kg i.p. ceftriaxone (Kendric, Mexico) to
reduce pain and risk of infection. This therapeutic scheme was repeated once per 24h for
three consecutive days. Three weeks after the FG injections, the rats were perfused (*vide*
supra). Free floating coronal sections were obtained with a vibratome. AVP
immunofluorescence reaction was made using rabbit anti-AVP antibody as described above.
Observations were made using a Zeiss LSM 880 confocal microscope. FG was observed by
using a UV excitation filter, and images were artificially assigned a green color for better
visualization.
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202   2.6 Golgi-Cox impregnation and neuronal 2-D reconstruction
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For the argentic impregnation, four 300g male rats were deeply anaesthetized and
decapitated. Coronal blocks approximately 5 mm thick, containing the hypothalamus, were
cut with a sharp blade and briefly rinsed with PB 0.1M. Blocks were immersed in sequential
impregnation A/B and C solutions as indicated in the FD Rapid GolgiStain Kit (FD
Neurotechnologies, Ellicott City, MD), during the following two weeks, after which 150 µm
coronal sections containing the NLOT and SON were obtained using a vibratome with the
cutting chamber filled with solution C, and mounted on gelatin-coated microscope slices,
dried overnight at room temperature in the dark and stained with solution D/E. Slices were
then washed, dehydrated, cleared with xylenes and cover-slipped with Permount Mounting
medium (Fisher Medical, UN1294). Neurons that were identified with more complete
somatic/neurite impregnation at SON and NLOT were reconstructed in a 2-D plane using a
camera lucida attachment mounted on the Nikon Eclipse 50i at 40X magnification.
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217   2.7 Intracerebral cannula implantation
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For implantation of permanent guide cannula into the NLOT, rats were anaesthetized with a
mixture (1:1) of xylazine (20 mg/ml, Procin, Mexico) and ketamine (100 mg/ml, Inoketam,
Virbac, Mexico) administered at a dose of 1 ml/kg body weight intraperitoneally. Rat was
placed in a stereotaxic frame (Kopf Instruments, Tujunga, CA, USA). Body temperature was
maintained at 37 °C using a CMA/150 temperature controller (CMA/Microdialysis,
Stockholm, Sweden). Bilateral stainless-steel 26 gauge cannula (C315G, Plastics One,
Roanoke, VA, USA) were implanted (stereotaxic coordinates: antero-posterior -1.4 mm,
medio-lateral ± 3.2 mm from bregma, and dorso-ventral -8.6 mm from the skull surface.

Guide cannulae were affixed on the skull with stainless steel screws and dental acrylic
cement (Laboratorios Arias, México City, Mexico) and sealed with dummy cannulae (C315DC,
Plastics One). Ketorolac and ceftriaxone were administered as mentioned above for three
days following the surgery. Animals were housed in individual cages and allowed to recover
from the surgery for one week. Beginning after the 2nd week post-surgery, rats were handled
once daily for 5 min for three consecutive days. Rats with fallen cannulae, signs of infection,
or altered motricity were excluded from behavioral tests.

2.8 Social behavior assessment and neuronal activation after intracerebral drug
administration

Behavioral testing was performed on day 11 post-surgery. On the day of the experiment, four
experimental groups with rats meeting the selection criteria (*vide supra,* n=20, N=80) were
formed and rats were administered the following drugs: control (0.9% NaCl); AVP (1 ng AVP
(Sigma V9879, USA)); AVP + V1a antagonist (1 ng AVP +30 ng Manning compound (Bachem,
USA)) or AVP + V1b antagonist (1 ng AVP + 10 ng SSR149415 (Axon Medchem, Groeningen).
The doses used here were those previously reported [9].The substances were injected
bilaterally using two microdialysis pumps (CMA/Microdialysis, Stockholm, Sweden) in a
volume of 250 nl over five minutes, and the cannulae were kept in place for 1 min after the
injection to prevent backflow and to allow for diffusion as described previously [26].

Social behavior assessment using the 3CST was started 15 min after microinjections,
and the time spent in the social compartment of the 3CST and the number of approaches to
the rat *stranger* were quantified as above mentioned during a test period of 10 minutes. Five
rats from each experimental group were randomly chosen and perfused after 60 minutes of
the 3CST, and FOS immunohistochemistry was performed as described above. The number
of Fos positive nuclei in NLOT and structures known to be downstream targets of NLOT were
counted within an area of 0.2 mm^2. The rest of the rats were sacrificed with an overdose of
pentobarbital and guillotined. Brains were dissected and postfixed for canula location
assessment. Scores of rats whose canula tips were placed beyond 200 µm from the perimeter
of the NLOT were excluded.

2.9 RNAscope dual ISH assays

Rats were transcardially perfused with 4% paraformaldehyde (PFA) in 0.1M PBS (PBS tablets
Sigma P-4417). Brains were immediately frozen in 2-methylbutane (isopentane, Sigma-
Aldrich, cat 277258, MO, USA) cooled in powdered dry ice, sectioned at a thickness of 12 µm,
and mounted onto *Fisher Super Frost slides*. Sections were dried for one hour at 60ºC, then

treated with 1X target retrieval reagent (from 10x stock solution provided by the supplier) in
a boiling water bath for 5 min. Digestion with Protease-plus was carried out for 15 minutes.
The RNAscope duplex method for dual ISH hybridization was performed with RNA probes to
identify colocalization of PACAP (probe Rn Adcyap1) with V1b receptors (probe Rn-Avpr1b),
V1a receptors (probe Rn-Avpr1a), VGLUT1 (probe Rn Slc17a7), and VGLUT2 (probe Rn
Slc17a6) in the NLOT and hippocampus CA2, as well as the colocalization of PACAP receptor
1 (probe RN Pac1) and VGLUT1 (Slc17a7) in the insular and gustative cortex. Probes were
designed and provided by Advanced Cell Diagnostics (Hayward, CA, United States).
Amplification and staining steps were performed following the RNAscope®2.5 HD Assay
Duplex protocol for fixed frozen sections.
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276 2.10 Statistical analysis
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Results are expressed as means ± SEM. D'Agostino and Pearson test was used to evaluate
the normality of the data. Differences between means were evaluated by Student's t-test to
compare behavior and Fos expression in the water deprivation experiments. One-way
ANOVA followed by Dunnet´s multiple comparisons test was used to compare the effects of
pharmacological micro-infusion of AVP and combined infusion with its antagonists on social
behavior and Fos. Significance in all tests was set at * $p<0.05$, ** $p< 0.01$ and *** $p<0.001$.
All statistical analyses were computed using Prism (GraphPad Software, Version 9, San Diego,
CA).

3. RESULTS

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288 3.1 WD48 increased the time in the social chamber and number of approaches to the rat
289 _stranger_ during the social interaction test (3CST)
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We have previous observed that WD48 increases conditional anxiety (internal, homeostatic,
threat) evaluated by elevated plus maze [27] but reduces freezing behavior and increases
escaping attempts when rats are exposed to a predator [3]. In both cases, it seems there is a
crucial involvement of the hypothalamic vasopressinergic ascending system, but in a
differential circuit involvement, depending on the adversity the animal is facing, for instance,
projections to central amygdala in the conditional anxiety test [6] and projections to lateral
habenula in the escaping test [28]. From these observations, the question of social interaction
consequences of osmotic challenge, and its circuit involvement, arose. We devised a simple
social interaction test, the 3CST (Fig. 1A, for details see section 2.2), to evaluate if the up-
regulation of the hypothalamic AVPMNN system could influence social behavior. Water-
deprived rats showed a significant enhancement in time spent in the _social_ chamber where
a rat _stranger_ was located withing a wire cage (Fig. 1B, right) (control: 215.5 ±31.66 vs. WD48:

350.3 ±32.38; p<0.01) and in the number of approaches/sniffing to the rat *stranger* (control:
7.33 ±0.91 vs. 24h WD: 11 ±0.81; p<0.01) (Fig. 1B, left).

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Figure 1. Water deprivation during 48h (WD48) increased the time of social interaction. A:
schematic representation of the three-chamber social test (3CST) used during sociability
evaluations. B: (left) WD48 led a significant increase of the time spent in the social chamber
where the rat stranger was kept inside a wire cage as compared control rats. B: (right) WD48
increased the number of approaches/sniffing to the rat stranger compared to the control
*group. Results are expressed as means ± SEM. Ctrl n= 12, WD n= 14. ** p < 0.01. Unpaired*
Student´s t test.
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315 3.2 Fos expression triggered by WD48+3CST compared with basal, WD48 or 3CST alone
316 reveals a social behavioral neuronal network
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Fos expression analysis is a powerful approach for obtaining insight into patterns of neuronal
activation during internal/external stimulation and behavioral adaptation. To investigate Fos
activation, experimental subjects were submitted to one of four conditions: 1) basal
condition, with food and water *ad libitum* and un-disturbed before perfusion/fixation; 2)
WD48, with food *ad libitum* (usually food intake ceases after 12h of water deprivation), un-
disturbed before perfusion/fixation; 3) food and water *ad libitum*, subjected to 3CST - the
perfusion/fixation was performed 60min after the end of the test; 4) WD48 prior to 3CST -
the perfusion/fixation was performed 60 min after the end of the test. We systematically
examined increased Fos expression, as a measure of neuronal activation, throughout the
brain in these four groups and performed semi-quantitative assessment of Fos elevation
(Table, and see the analysis criteria in section 2.4).
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330   As expected, social interaction caused moderate to large increases in Fos expression
331 in regions involved in processing of exteroceptive and interoceptive information. These
332 regions include those involved in olfactory processing such as the anterior olfactory nucleus,

accessory olfactory bulb, nucleus of the lateral olfactory tract, piriform and entorhinal cortex, taenia tecta, cortical and posterior amygdala, olfactory tubercle, and endopiriform nucleus [29, 30]; regions involved in visual processing such as the primary and secondary visual cortex, pretectal nucleus, the lateral geniculate complex of the thalamus and the midbrain nucleus of the posterior commissure [31]; regions involved in auditory processing such as the inferior colliculus, medial geniculate nucleus and basolateral amygdala [32]; limbic regions that participate in the integration of emotional states such as central amygdala, bed nucleus of the stria terminalis, accumbens, ventromedial hypothalamus and septum; regions involved in memory and coding of space information such as the hippocampal formation, entorhinal cortex and retrosplenial cortex [33, 34]; and hypothalamic nuclei involved in stress, aggression, and arousal regulation such as the paraventricular and ventromedial hypothalamic nuclei.

In contrast, activation of neurons by water deprivation was dramatically greater in SON, and PVN, two nuclei rich in osmosensitive AVPMNNs, whereas the SCN, containing non-osmosensitive vasopressinergic neurons, and other non-vasopressin-expressing nuclei of brain, exhibited far less activation in response to water deprivation (Table 1). A high number of Fos positive nuclei was found in the BNST, a region reported to be targeted by projections from osmosensitive glutamatergic neurons of the subfornical organ [35, 36]

WD48+3CST-induced spatial neuronal activation pattern coincided with the main efferent regions of SON and the nucleus of the lateral olfactory tract (NLOT). Among brain regions showing neuronal activation (Fos expression) specifically associated with *combined* WE48 and 3CST exposure, the NLOT was of particular interest, since this region contains vasopressinergic fibers which were visibly increased by WD48 (Figure 2). The main efferent cortical regions of NLOT (see Allen mouse interaction map, Adcyap1-2A-Cre, Exp. 187269162, and Fig. 5A) also showed strong increase of Fos expression after WD48+3CST. These regions include anterior olfactory nucleus, piriform cortex, agranular insular cortex, gustatory cortex, dorsal insular cortex, basolateral amygdala, claustrum, and endopiriform nucleus (see Table, bold lettered regions).

3.3 Forty-eight hours of water deprivation (WD48) potentiated hypothalamic vasopressinergic system and activated Fos in neighboring NLOT

We examined further the source of origin of vasopressinergic fibers, as well as the nature of Fos-positive cells in NLOT after water deprivation. Concordant with progressive activation of secretion of AVP from posterior pituitary (Fig. 2A) following water deprivation, there was an increase in Fos expression both in PVN and SON (Fig. 2B'), as noted in Table, and in NLOT as well (Fig. 2B). Furthermore, AVP-positive terminals were found in close apposition to Fos-positive cells in NLOT (Fig. 2B´´), and the intensity of vasopressin terminals in NLOT was

corresponding­ly increased (Fig. 2C vs. 2D). Thus, it appeared that upregulation of the vasopressinergic system by osmotic stress facilitated an increase in the number and/or intensity of staining of AVP fibers in the NLOT.

Table 1. Semi-quantitative assessment of neural activation in 4 experimental conditions

Region	Abs.	Control	Social Interaction	48h WD	48h WD Social Inter.
Cortical plate (CTXpl)					
Anterior cingulate cortex	AC	-	++	+	++
Main olfactory bulb	MOB	-	-	+	+
Accessory olfactory bulb	AOB	-	++	-	++++
Agranular insular	**AI**	-	++	+	+++
Anterior olfactory nucleus	**AON**	-	+++	++	++++
CA1 (dorsal hippocampus)	CA1d	+	+	+	+
CA1 (ventral hippocampus)	CA1v	-	++	+	+++
CA3 (temporal hippocampus)	CA3t	-	++	-	++
CA2 (dorsal hippocampus)	CA2d	-	++	-	++
Dentate gyrus	DG	+	++	++	++*
Ectorhinal area	ECT	-	++	+	++++
Entorhinal area	ENT 4	-	++	+	+++
Gustatory cortex	**GU**	-	+	+	+++
Infralimbic cortex	ILA	-	++	-	+++
Nucleus of the lateral olfactory tract	**NLOT**	-	++	+	++++
Orbital cortex	ORB	-	++	+	+++
Pos-subiculum		-	+	-	++
Para-subiculum	PAR	-	-	-	+
Piriform amygdalar cortex	PAA	-	+	-	++
Piriform cortex	**PIR**	-	+++	+	++++
Prelimbic cortex	PL	-	++	-	++
Presubiculum	PRE	-	+	-	+++
Retrosplenial cortex	RSP2/3	-	+++	-	++++
Somatomotor cortex, L5	MO5	-	+	-	++
Somatosensory cortex, L2/3	SS2	-	++	-	+++
Somatosensory cortex, L5/6	SS5	-	++	-	+++
Subiculum	SUB	-	-	+	+
Taenia tecta	TT	-	+++	-	+++
Visual primary cortex L2/3	V-2/3	-	+++	-	++++
Visual primary cortex L6	V1 -6	-	+++	-	+++
Visual secondary cortex	V2	-	++	-	++++
Cortical subplate (CTXsp)					
Basolateral amygdalar nucleus	BLA	-	++	+	+++
Basomedial amygdalar nucleus	BMA	-	+	-	+
Cortical Amygdala	COA	-	+	+	+
Claustrum	**CLA**	-	++	+	+++
Endopiriform nucleus	**EPd**	-	+++	+	++++
Lateral amygdala	LA	-	+	-	+
Posterior amygdalar nucleus	PA	-	++	+	++
Hilus (dentate gyrus)	HYL	+	+	-	+
Cerebral nuclei (CNU)					
Bed nuclei of stria terminalis	BST	-	+++	+++	++++
Caudate putamen	**CP**	-	+	+	+++
Globus pallidus	GP	-	+	-	++
Central amygdalar nucleus	CEA	-	++	++	+++
Diagonal band nucleus	NDB	-	+++	++	++++
Lateral septum	LS	-	++	+	+++
Medial amygdalar nucleus	MEA	-	++	-	+++
Nucleus accumbens	**ACB**	-	++	+	+++
Olfactory tubercule	**OT**	-	++	++	++
Pallidum ventral	PAL	-	+++	+	+++
Sustancia inominata	SI	-	+	+	+
Interbrain (IB)					
Anterior hypothalamic nucleus	AHN	+	++	+	++
Arcuate hypothalamic nucleus	ARH	+	+	++	+++
medial geniculate	mGd	-	+	-	+
Dorsal lateral geniculate	LGd	-	++	-	+++
Dorsomedial hypothalamic nucleus	DMH	+	++	+	+++
Intergeniculate leaflet of the lateral geniculate complex	IGL	-	++	+++	++++
Lateral dorsal nucleus of the thalamus	LD	-	++	+	++
Lateral habenula	LH	-	+	++	+++
Lateral hypothalamic area	LHA	-	++	++	++++
Lateral preoptic area	LPO	-	+	+++	++++
Medial geniculate	MG	-	++	-	++
Medial habenula	MH		++	-	+++
Medial preoptic area	MPO	-	+	++	++
Nucleus of reuniens	RE	+	++	++	++
Paraventricular nucleus hypothalamus	PVH	++	++	+++++	+++++
Paraventricular thalamus	PVT	+	+	+	+++
Periaqueductal gray	PAG	-	+	+	+++
Posterior hypothalamic nucleus	PH	+	++	++	+++
Premammillary	PM	-	++	+	++++
Reticular nucleus of the thalamus	RT	+	++	+	++
Subthalamic nucleus	STN	+	++	+++	+++
Suprachiasmatic	SCH	+	+	+	+++
Supramammillary nucleus	SUM	-	+	++	+++
Supraoptic nucleus	SON	-	+	+++++	+++++
Ventromedial hypothalamic nucleus	VMH	-	+++	+++	++++
Zona incerta	ZI	+	++	++	+++
Midbrain (MB)					
Anterior pretectal nucleus	APN	-	+++	+	++++
Dorsal raphe nucleus	DR	-	++	+	+++
Inferior colliculus	IC	+	+++	++	+++
Midbrain reticular nucleus	MRN	+	+	+	++
Nucleus of the posterior commissure	NPC	+	+++	+	++++
Pretectal area	PRT	-	++	++	+++
Substantia nigra, compacta	SNc	-	++	++	+++
Substantia nigra, reticular	SNr	-	++	++	+++
Superior colliculus	SC	+	+	+	++
Ventral tegmental area	VTA	-	++	++	++
Hindbrain (HB)					
Parabrachial nuclei	PBN	-	++	+	++
Pontine reticular nucleus	PRN	-	++	+	++

Fos + nuclei were evaluated in a 0.2 mm^2 area and the following scores were assigned: (-) no Fos+ nuclei observed; (+) 1-25; (++) 26-50; (+++) 51 – 75, (++++) 76-100 and (+++++) >100.

Figure 2: Forty-eight hours of water deprivation (WD48) potentiated hypothalamic vasopressinergic system and activate Fos in neighboring NLOT. *A: Time course of dynamic changes in plasma arginine vasopressin (AVP) concentration during 48 h of water deprivation, measured in wild-type male rats (n=5) with ELISA (modified from Zhang et al, 2020, with permission). Bs: example of Fos expression in hypothalamus at supraoptic level and cortical amygdalar region (B': low magnification of B and B'', Fos and AVP IHC in NLOT after WD48). C and D: photomicrographs showing AVP immunoreactivity (ir) (digital photos in negative mode to enhance AVP immunopositive fibers' visibility) in the same region of B, in control and WD48 rats. Note in the high magnification insets that after WD48, the AVP + fibers clearly increased their visibility (white arrows). A vessel within the magnified region is indicated in dotted lines for anatomical reference. Abbreviatures: PVNmmd: paraventricular nucleus, medial magnocellular division; SON: supraoptic nucleus; NLOT: nucleus of lateral olfactory tract*

3.4 Vasopressinergic input to NLOT from SON of the hypothalamus revealed by Fluoro-Gold (FG) retrotracing and Golgi-Cox stained sample analysis

Retrograde tracing in conjunction with staining for AVP was employed to demonstrate that
SON AVPMNNs project to NLOT. Iontophoretic FG injection targeted to NLOT (see section
2.5) resulted in strong labeling in SON (Fig. 3A and B). Most of the labeled cells were
immunopositive for AVP (Fig. 3B). Microscopical observation of Golgi-Cox stained samples of
brain coronal sections (150 μm of thickness), containing NLOT and the SON, with only sparse
staining (Fig. 3C and 3D), revealed that SON magnocellular neurons could give rise to as many
as three main axons which coursed medial, lateral and dorsally. Figure 3Da and 3Db show
two magnocellular neurons, single-stained. Those two cells are designated as cells "a" and
"b" in Figure 4. Each of the cells emitted three main axons, numbered 1, 2, and 3, which
coursed medial, dorsal and laterally. Fig. 3Dc shows the axon coming from cell "b", axon #3,
with a huge varicosity, a characteristic of the magnocellular neurons, entering layer 1 of NLOT
(Fig. 4F)
407

*Figure 3. **Anatomical relationship and interconnections between rat hypothalamic supraoptic***
***nucleus (SON) and the nucleus of lateral olfactory tract (NLOT).** A and B: retrograde tracer*
fluorogold injected to NLOT labeled the SON vasopressin immunoreactive neurons. C. Golgi-

Cox staining of the rat brain coronal section (150 μm of thickness) containing NLOT and the
SON. D: higher magnification of C with squared region with NLOT and SON showed. Note the
clearly visible allocortical feature, three layers of the NLOT clearly visible. Da and Db show
two magnocellular neurons, single-stained within the SON, that they emitted 3 main axons
from soma or proximal dendrites and coursed medially (1), dorsally (2) and laterally (3) that
could be followed and reconstructed using adjacent sections (see Fig. 4A). Dc: showing the
axon coming from cell "b", axon #3, with huge varicosity, which is a characteristic of the
magnocellular neurons, entering the layer 1 of NLOT, could be followed in the two adjacent
sections. Hollow arrows indicate the main axons coming out from the magnocellular cells a
or b.
422
423 ### 3.5 Molecular signature of NLOT principal neurons (layer 2, NLOT2) and projection fields
424 reveals a cooperative role between hypothalamic vasopressinergic signaling and an NLOT
425 PACAPergic neuronal population
426
The NLOT is an isolated tri-laminar ovoid cell mass located in the anterior cortical amygdalar
region [19, 37]. Nissl staining reveals three cell layers within the NLOT nucleus. Layer 1 (NLOT1)
is a subpial molecular zone with scattered neurons, which receives mitral cell input from the
main olfactory bulb. Layer 2 (NLOT2) is a thick and dense corticoid aggregate of pyramidal
neurons of medium size with apical dendrites entering NLOT1 (Fig. 4F, for example). Layer 3
(NLOT3) is a multiform layer disposed more deeply than layer 2. It contains a mixture of small
and large neurons, some of them possibly representing inhibitory interneurons of subpallial
origin [21]. We next explored the neurochemical signature(s) of the neurons in NLOT that are
the presumptive targets of vasopressinergic terminals from SON, which terminate mainly in
the layer 1 where the pyramidal neurons apical dendrites branch. Figure 4A shows three
types of neuronal morphology reconstructed using camera lucida and Golgi-Cox stained
samples (section thickness: 150 μm). Pyramidal neurons of the NLOT2 are of particular
interest here. Using dual ISH (RNAscope duplex method), we identified a population of
PACAPergic, VGLUT1 and VGLUT2 expressing neurons in layer II of NLOT (Fig. 4C and 4D) [38]
representing pyramidal neurons (black cells) which, upon Golgi staining and reconstruction,
send dendritic processes into both layer I and layer III of NLOT where the SON magnocells
send their axons (Fig. 4A, red axons reconstructed with camera lucida in three adjacent
sections of thickness 150 μm each). As these neurons also express the vasopressin receptors
V1a and V1b (Fig. 4E and 4F), they are candidate target neurons of the vasopressinergic
projections from SON. In fact, these PACAPergic (and glutamatergic) neurons are also those
that exhibit Fos elevation after 48WD.
448

449
Figure 4. **Molecular signature of NLOT principal neurons (layer 2, NLOT2), glutamatergic, PACAPergic**
and V1a and V1b mRNA expressing. *A: coronal section showing the RNAscope ISH for Adcyap1, mRNA*
encoding PACAP, at the nucleus of lateral olfactory tract (NLOT) antero-posterior level. The NLOT
strongly expresses PACAP mRNA. A': amplification of squared region A. Note that most of PACAPergic
neurons are the layer 2 neurons (NLOT2). B-E, dual in situ hybridization (DISH) using RNAscope duplex
method showing the PACAPergic neurons of NLOT2 co-express Slc17a7 (mRNA for VGLUT1), Slc17a6
(mRNA for VGLUT2), Avpr1a (mRNA for vasopressin receptor V1a) and Avpr1b (mRNA for vasopressin
receptor V1b). Doble arrows indicate examples of co-expression. F: camera lucida reconstructions of
selective NLOT neurons, from Golgi-Cox-stained sample of Fig. 2C. Note the NLOT2 neurons (black
neurons) are mainly pyramidal neurons with the apical dendrites and their subsequent branching
toward the layer 1 and the brain ventral surface where most of AVP immunopositive fibers were
observed. Blue cells were camera lucida-reconstructed from the NLOT1 and green cells from the
NLOT3. Two SON (light beige shading magnocellular neurons with three axons emitted from cell
bodies or proximal dendrites, followed in the two adjacent sections. "1" indicate the axons coursing
toward the infundibulum, "2" indicate the axon coursing dorsally and "3" indicate axons coursing
laterally toward NLOT (light green shading) and cortical amygdala (CoA).
466

Cortical efferent regions of NLOT2 PACAPergic pyramidal neurons which also express
VGLUT2 [38] (Fig. 4D) and the transcription factor Sim1 [39] are revealed within the Allen
connectivity atlas (Fig. 5A and supplementary Fig. 1). Figure 5A depicts a map of NLOT
projections made with a mouse line Adcyap1-2A-Cre, Exp. 187269162, injected with a Cre-
dependent AAV tracer into the NLOT that expresses EGFP. The main efferent regions include
anterior olfactory nucleus, piriform cortex, agranular insular cortex, gustatory cortex, dorsal
insular cortex, basolateral amygdala, claustrum, and endopiriform nucleus (see Table, bold
lettered regions) which all strongly express the PACAP receptor PAC1 mRNA (Adcyap1r1, Fig.
5B) and also co-express VGLUT1 mRNA (Slc17a7, Fig. 5C and insets).

Fig. 5 Glutamate-PACAPergic NLOT main efferent regions analyzed in this study strongly co-
express mRNAs of PACAP receptor PAC1 (Adcyapr1r) and VGLUT1 (Slc17a7) and were
activated by 48WD. A: Axonal projections from PACAP transfected NLOT neurons. Obtained
from Allen mouse brain connectivity atlas (Experiment: 187269162 Transgenic line: Adcyap1-
2A-Cre injected into NLOT with an AAV Cre-dependent vector). B: coronal section showing the
expression of Adcyap1r1 (PAC1 receptor mRNA) with cortical regions that are target of NLOT
are delineated in dotted lines. Image obtained from Allen ISH brain atlas (Experiment:
74988667) C: Coronal section processed for dual ISH using Duplex RNAscope, to show the
colocalization of VGLUT1 and PAC1 in cortical regions targeted by projections from
PACAPergic NLOT neurons. D: Coronal section showing Fos expression after WD48+3CSI.
Panels: D1 – D5 show the key regions discussed in this study: SON: supraoptic nucleus; NLOT:
nucleus of lateral olfactory tract; CLA: claustrum; AIC: Agranular insular cortex; DI, dorsal
insular cortex; GC: gustatory cortex . The whole brain Fos expression semi-quantitative
analysis is reported in table 1.
490
491 3.6 Increased social behavior and Fos expression induced by vasopressin micro-infusion in
492 NLOT is blocked by both V1a and V1b receptor antagonists
493
In order to ascertain whether or not AVP can directly modulate neuronal activity in NLOT,
and influence social behavior, AVP (1 ng/side) was bilaterally infused into this region (Fig. 6A)
15 min before 3CST. To evaluate the participation of the V1b and V1a receptors, the
pharmacological antagonists SSR149415 (V1b antagonist) and Manning compound (V1a
antagonist) were co-injected with vasopressin at 30 ng/side in 20 rats per condition. All
cannula tips of the animals included in the behavioral or Fos analysis were shown to be within
a 200 μm perimeter of the NLOT boundaries
501
The microinjection of AVP significantly increased the time (350.34 ±21.3, p<0.01) that
the experimental rats spent in the *social* compartment as compared with the saline-treated
group (215.4 ±16.92). This effect was reduced to control level vasopressin injection was
accompanied by co-administration with SSR149415 (256.26 ±40.32) or Manning compound
(156.3 ±29.57)(Fig. 6C, right). A significant elevation in social approaches was seen after
intra-NLOT infusion of AVP (11.92 ±2.15) in comparison to the saline group (5.81 ±0.75;
p<0.001). The co-infusion of AVP+ SSR149415 prevented the increase in the number of
approaches to the social compartment (6.35 ±0.51) seen with AVP alone, showing a similar
value to the saline group (Saline: 5.81 ±0.75). Microinfusion of Manning compound along
with AVP similarly decreased the number of approaches (8.00 ±1.03) compared to AVP alone
(Fig. 6C, left).
513

To confirm that NLOT PACAPergic/glutamatergic neurons are plausible targets for
activation by vasopressin projections from SON, we examined Fos elevation in selected
regions known to be involved in social behavior (Fig 6D). One-way ANOVA followed by
Dunnett multiple comparison test showed that bilateral AVP (1 ng/side) infusion into the
nucleus of the lateral olfactory tract (NLOT) elicited an increase of Fos+ expression in NLOT
compared to saline (saline: 27.20 ±0.97, vs AVP: 44.00 ±2.21, $p<0.0001$), no significant
differences were observed between saline and AVP+SSR149415 (26.60 ±3.37), nor between
saline vs. AVP + Manning compound (26.80 ±1.35). In the anterior olfactory nucleus (AON)
we found that compared to saline (40.80 ±2.45), AVP increased the number of Fos positive
nuclei (68.40 ±1.53, p<0.001), and compared to control, there was a significant decrease in
the number of Fos positive nuclei after co-infusion of AVP+SSR149415 (24.2 ±1.93, $p<0.0001$)
or AVP + Manning compound (22.60 ±2.01, $p<0.0001$). In the basolateral amygdala (BLA),
AVP infusion into NLOT elicited significant increases in Fos activation compared to saline
infusion (saline: 26.00 ±1.87 *vs. AVP:* 41.00 ±2.91, $p<0.001$) and the effect of AVP infusion
was blocked with AVP+SSR149415 (24.00 ±1.87) and with Manning compound (22.00 ±2.55).
NLOT AVP infusion increased the number of Fos+ nuclei in agranular insular cortex (AI)
compared to saline (saline: 5.00 ±0.70 vs. AVP: 26.60 ±1.50; $p<0.0001$). While the effect of
AVP antagonists on AVP was not as complete here as in other areas (AVP+SSR149415: 18.4
±0.81, AVP + Manning compound: 15.8 ±1.35, $p<0.01$). In gustatory area (GU), compared to
control, AVP infusion into NLOT significantly increased the number of Fos+ nuclei (Saline:
12.60 ±1.32 *vs. AVP:* 76.60 ±7.43, $p<0.0001$) and V1a and V1b antagonists abolished this

effect as no differences were found between saline and SSR14915 (12.00 ±1.37), or Manning
compound (10.00 ±1.30)(Fig. 6D).
537
Figure 6. Vasopressin microinfusion into the nucleus of the lateral olfactory tract (NLOT)
induces both an increase in social behavior and Fos+ nuclei expression. (A) Representative
scheme showing a bilateral cannula placeman within the NLOT. (B) AVP infusion (1ng/side)
increased the time that the rats spent inside the social compartment these effects were fully
prevented by the simultaneous infusion of both AVP V1b receptor (10 ng/side, SSR149415
(SSR) antagonist, and AVP V1a receptor (10 ng/side, Manning compound antagonist. (C) A
significant difference was found in the number of entries between the infusion of AVP vs the
saline group these effects were blocked by the simultaneous infusion of AVP V1b and V1a
receptor antagonist. (D) AVP infusion into NLOT elicited an enhancement of Fos+ expression
in AON, BLA, AI, and GU, as compared with the respective saline-treated group. The
simultaneous infusion of AVP+V1b and AVP+V1a antagonist resulted in a decreased Fos
expression in all the areas studied in this work. Abbs: NLOT: nucleus of lateral olfactory tract;
AON: accessory olfactory nucleus; BLA: Basolateral amygdala; AI: agranular insular; GU:
*gustatory area. Results are expressed as means ± SEM. **P< 0.01; ***P<0.001. Two-way*
ANOVA followed by Dunnett´s multiple comparisons test was used to evaluate social behavior
(Saline n=20; AVP n=19; AVP+SSR n=19; AVP+Manning C. n=19) and One-way ANOVA
followed by Dunnett´s multiple comparisons test to Fos expression (n= 5 rats for each group).
555
556   DISCUSSION
557
558 In this study, we focused on a newly-characterized projection system from AVPMNNs of the
559 supraoptic nucleus (SON) of the hypothalamus to the nucleus of the lateral olfactory tract
560 (NLOT), the activation of these neurons by water deprivation, and ensuing effects on their
561 neurochemistry, their connections, and their potential effects on social behavior.
562
We report here that rats stressed osmotically via WD48 had a higher score in
sociability than euhydrated rats. It has been previously reported that osmotic stress potently
increases the metabolic activity of AVPMNNS in the hypothalamus, and that increased AVP
mRNA expression in PVN and SON correlate with levels of social interaction in mice [40, 41].
Intracerebroventricular infusion of AVP increases social contact in prairie voles [42] and novel
conspecific social interaction in the Syrian hamster [43]. Brattleboro rats lacking AVP due to a
genetic prohormone processing defect, show various behavioral impairments, including
altered social development [14] and less social playing and prosocial ultrasonic vocalizations
[13]. The duration of social recognition in rats [44] is dependent on the levels of AVP in the
lateral septum. Variation in expression of vasopressin receptor genes (AVPR1A, AVPR1B)
correlates with variation in social behavior in rhesus macaques [45]. Our novel finding here is
the identification of the direct innervation of the NLOT by AVP projections from the SON,

which has not been reported in the literature previously and represents a candidate circuit
activated by osmotic challenge induced by water deprivation.

We also observed the displayed increased Fos expression in the nucleus of the lateral
olfactory tract (NLOT) and its efferent cortical regions after WD48 + 3CST, indicating that the
homeostatic state is able to modulate neocortical social interaction processing centers via a
hub within the cortical amygdalar lateral olfactory tract nucleus, NLOT. We confirmed the
involvement of AVP within this hub by injecting vasopressin directly into the NLOT. A novel
finding in this study was that both V1a and V1b receptor antagonists reduced the AVP-
induced increase in sociability. The fact that this phenomenon was blocked by either V1a or
V1b receptor antagonists co-infused with AVP suggests that both types of receptor are
necessary, and either is insufficient, to mediate these behavioral effects.

NLOT projects to the accessory olfactory system which is activated during social
investigation behavior in which AVP participates through neuronal signaling to filter social
cues of odor [46]. Similarly, NLOT-lesioned rats spend significantly less time sniffing out odors
associated with social contacts than intact rats regardless of the number of times exposed to
the olfactory stimulus [47]. It is known that water deprivation enhances both AVP-
immunoreactivity and Fos expression in the hypothalamic paraventricular and supraoptic
nuclei [5] and therefore vasopressin effects could be exerted elsewhere than at the level of
the amygdala. However, the fact that exogenous administration of AVP into NLOT could
produce identical behavioral effects as those induced by WD support the conclusion that the
effects of WD are mediated by AVP release in the NLOT. The effects of AVP microinfusion in
NLOT on Fos activation in AON, BLA, AI, and GU, and blockade of Fos activation by AVP
antagonist co-administration in NLOT alone, is also consistent with the notion that AVP
affects activation of this behavioral network by its action as a neurotransmitter at the NLOT
'hub'. A previous study documented enhancement of social interaction after osmotic
dehydration (subcutaneous injection of 2 M NaCl) [48]. It seems likely that this osmotic
stressor, like water deprivation, may affect social behavior also through AVP release in NLOT

It is uncertain at the present time how altered social behavior (increased social
interaction) after water deprivation represents an integration of homeostatic and allostatic
drive that is beneficial to animal survival (and would, at least simplistically) explain the
evolution and stabilization of the pathway described here in mammalian brain. Broadly,
increased attention to the behavior and proximity of con-specifics during resource scarcity
may have both positive and negative effects. For example the borer and social mutations of
another neuropeptide-liganded GPCR, the NPY-like receptor of C. elegans, are posited to
drive foraging behavior when food is scarce, and social behavior (and increased mating)

when food is plentiful, respectively [49]. In this context, increased sociability during water
deprivation would seem to be disadvantageous, rather than the reverse. However, it must
be borne in mind that the behavioral model chosen here as a 'behavioral readout' for water
deprivation (social interaction) may be insufficiently rich/complex to capture the full range
of advantages and disadvantages of increased social interaction during periods of scarcity,
and indeed 'water deprivation' may not necessarily model resource scarcity per se. For
example, salt preference may be a behavioral characteristic that involves
vasopressinergic/dynorphinergic MNNs and helps to integrate water seeking and
consumption with salt balance ([2] and references therein). Thus, we postpone speculation
on the potential evolutionary advantages of a linkage between social behavior and water
deprivation, and note here only that this linkage exists, at least in part, because of a
vasopressinergic projection from SON (exclusive of PVN and SCN) to NLOT, in the rat.
625
626
Besides the vasopressinergic SON→NLOT PACAPergic projection system described
here, there are at least two other AVPMNN systems that link homeostasis and behavior that
have been recently reported [3, 6, 28, 50]. In these latter cases, the AVP target neurons (GERNs
in LHb, amygdalar neurons, noradrenergic neurons) have their own well-established
downstream connections that suggest how vasopressinergic inputs to them ultimately
affects behavior. For the present case, i.e. the SON→NLOT pathway, it is much less clear how
activation accomplishes enhanced social behavior, because the downstream connections of
NLOT2 PACAP/glutamate neurons are not yet well-established.
635
As a final note, it is worth remarking that receptor-specific pharmacological
manipulation of this pathway (and parallel ones) for potential therapeutic purposes could be
easily separate vasopressin-like effects on osmoregulation itself (mediated mainly via V2),
from behavioral effects. It will be of interest in future experiments to determine whether or
not a combination of V1a and V1b *agonists* can mimic the behavioral effects of AVP infusion
into the brain on behavioral independently of osmoregulatory effects intrinsic to arginine
vasopressin.
643

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Supplementary information

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Nucleus of lateral olfactory tract (NLOT) neurons with different neurochemical markers share similar projection
targets in cortical regions. Three dimensional projection patterns in dorso-ventral (left) and sagittal (right) views
of NLOT virally transfected neurons in which reporter gene expression was restricted by selective CRE
expression driven by the promoters of PACAP (Adcyap1 2A Cre) shown here in yellow , VGLUT2 (Slc17a6 IRES
Cre) shown in orange or *single minded homolog 1* (Sim1 Cre_KJ18) depicted in blue. Notice that for all the
experiments the cortical regions that were presented in bold letters in the table 1 (delineated by white dotted
lines) received strong innervation by glutamatergic / PACAPergic axons originated in NLOT (delineated by red
dotted lines). All of these target regions displayed increased Fos activity after 48WD+3CST. Reconstructions of
the projection pathways were generated using the *Brain Explorer 2* program of the Allen Institute for Brain
Science and the [data of the experiments 187269162, 294174996 and 159151138](#) that can be found in the
Mouse Brain Connectivity Atlas. Abbreviations: AON: anterior olfactory nucleus, AI: agranular insular cortex, BL
A: basolateral amygdala, CLA: claustrum, GU: gustatory cortex, NLOT: nucleus of the lateral olfactory tract, PIR:
piriform cortex.