PROPOSED TITLE: "Comparing the evolutionary dynamics of predominant SARS-CoV-2 virus lineages co-circulating in Mexico"

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ABSTRACT (150/150 WORDS)

Up to November 2021, over 200 different SARS-CoV-2 lineages circulated in Mexico. To investigate lineage replacement dynamics, we applied a phylodynamic approach to explore the evolutionary trajectories of five dominant lineages that circulated during the first year of the local epidemic. For most lineages, peaks in sampling frequencies coincided with different epidemiological waves of infection in the country. Lineages B.1.1.222 and B.1.1.519 showed comparable dynamics, represented by clades likely originating in Mexico and persisting for over a year. Lineages B.1.1.7, P.1 and B.1.617.2 also displayed similar dynamics, characterized by multiple introduction events leading to a few successful extended local transmission chains that persisted for several months. We further explored viral movements across the country, applied within the largest clades identified (belonging to lineage B.1.617.2). Many clades were located within the south region of the country, suggesting that this area played a key role in the spread of SARS-CoV-2 in Mexico.

MAIN TEXT (7179/5000 WORDS)

INTRODUCTION

Genome sequencing efforts for the surveillance of the Severe Acute Respiratory Syndrome Coronavirus-2 (SARS-CoV-2) has granted public access to a massive number of virus genomes generated worldwide (https://www.gisaid.org/). Exploring SARS-CoV-2 genome data using molecular evolution tools has enabled to characterize an increasing virus diversity [1], to track emerging viral subpopulations, and to explore virus evolution in real-time both at local and global scales (for examples see [2–6]). Throughout the development of the COVID-19 pandemic, viral variants have emerged and circulated across different regions of the world [2,7], displaying specific mutations that define their phylogenetic clustering patterns [8,9]. SARS-CoV-2 virus lineages have been classified using a dynamic nomenclature system ('Pango system', Phylogenetic Assignment of Named Global Outbreak Lineages), developed to consistently assign newly generated virus genomes to existing lineages, and to designate novel virus lineages according to their phylogenetic identity and epidemiological relevance [10,11].

The emergence and spread of SARS-CoV-2 virus lineages has been routinely monitored since early 2021 (https://www.gisaid.org/), prompted by public health authorities to inform on the ongoing responses to the pandemic [12]. Virus lineages that may pose an increased risk to global health are classified as Variants of Interest (VOIs), Variants under Monitoring (VUMs), and Variants of Concern (VOC), and may display some of the following biological properties [9,13] : increased transmissibility [14], decreasing the effectiveness of available diagnostics or therapeutic agents (such as monoclonal antibodies)[15], and evasion of immune responses (including vaccine-derived immunity) [12,16,17]. Up to date, five distinct SARS-CoV-2 lineages (including their descending sub-lineages) have been designated as VOC: B.1.1.7 (Alpha), B.1.351 (Beta), P.1 (Gamma), B.1.617.2 (Delta), and more recently, the B.1.1.529 lineage (Omicron) [12,18,19].

The spread of variants in different regions of the world has been linked to human movement, with VOC often replacing previously circulating virus lineages to become dominant (for example, see [20]). Virus lineages that dominate across distinct geographic regions are likely to denote an evolutionary advantage, driven in part by a genetic background prompting some increase in fitness (such as enhanced transmissibility and/or immune escape) [20–24]. However, whilst some lineages may become dominant in some geographic regions, others fail to do so [20]. Exploring lineage replacement

dynamics remains challenging, as it represents a complex scenario impacted by numerous factors,

such as: i) differential and stochastic growth rates linked to lineage-specific spread patterns varying

across geographic regions, ii) a genetic background and a shifting immune structure of the host

population (linked to viral pre-exposure levels, as well as vaccination rates),[20,24] and iii) changing

behavioural patterns, such as fluctuating human mobility and the implementation of local non-

pharmaceutical interventions across time [20,25,26].

Mexico has been severely impacted by the COVID-19 pandemic, evidenced by the high

number of cumulative deaths reported relative to other countries in Latin America [27]. Since the first

introductions of the virus into the country during early 2020 [28], the development of the local epidemic

has been characterized by an epidemiological curve fluctuating between three waves of infection

observed in July 2020, January 2021, and August 2021 (up to November 2021) [27–29]. Prior to the peak

of the first wave, non-pharmaceutical interventions including social distancing and suspension of non-

essential activities were implemented at a national scale (from March 23rd 2020 to May 30th 2020),

whilst a reopening plan for the country was announced on May 13th 2020. Further, the national

vaccination campaign did not began until December 24th 2020 [27].

During February 2021, a sequencing consortium for the surveillance of SARS-CoV-2 in

Mexico was launched (Mexican Consortium for Genomic Surveillance, abbreviated CoViGen-Mex [30]).

In an attempt to further homogenize the spatial and temporal representation of the viral genomes

sequenced, the sampling scheme was upgraded in May 2021, for which approximately 1,200

representative samples selected across all states following official case reports were sequenced

monthly [30]. This resulted in approximately one third of the virus genome sequences from Mexico

publicly available. Independent to CoViGen-Mex, other national institutions significantly contributed to

the genomic surveillance of the virus (https://www.gisaid.org/). Sequencing efforts at a local scale

have enabled to characterize viral diversity, and to identify the viral lineages circulating within national

territory [30–32]. Over 200 different virus lineages have been detected in Mexico, including all VOC [19,33].

Many lineages also co-circulated across time and space, an observation of particular importance in

the context of the recently described recombinant SARS-CoV-2 linages that emerged in North

America during 2021 [34].

Using a phylodynamic approach, we used publicly available virus genome data collected in

Mexico to investigate the introduction, spread and replacement patterns of predominant virus lineages

(B.1.1.222, B.1.1.519, B.1.1.7, P.1 and B.1.617.2) that circulate(d) in the country up to November

2021. To generate datasets, we implemented a migration and a phylogenetically-informed

subsampling approach, aiming to increase robustness for tailored phylogeographic analyses. We

further explored the correlation between human mobility and the inferred viral movements for the

largest clades (C5d and C6d) belonging to the B.1.617.2 lineage, which also represented the most

extended transmission chains. Analysis revealed similar dynamics for the B.1.1.222 and B.1.1.519,

with both of these likely originating in Mexico and showing a sustained transmission over a year. For

the P.1, B.1.1.7 and the B.1.617.2, multiple introduction events were identified, with few large

transmission chains representing successful viral spread across the country. For C5d, spread

occurred from the south, with subsequent movement into the central and northern region of the

country. Our study further provides the basis for in-depth exploration of SARS-CoV-2 genomic data

from Mexico, and highlights the need to investigate the spread of transmission chains at a state level

resolution.

RESULTS

Prior to the introduction of 'Omicron' (B.1.1.529) into Mexico, only the B.1.1.222, B.1.1.519, B.1.1.7

(Alpha), P.1 (Gamma) and B.1.617.2 (Delta) virus lineages showed a progressive replacement pattern

to become dominant over other co-circulating virus lineages, with peaks in sampling frequency often

coinciding with the epidemiological waves of infection (except for B.1.1.7 and the P.1; **Figure 1a and**

b)[19,31,32]. During this time, Mexico reported a daily mean test rate ranging between 0.13-0.18

test/1,000 inhabitants, approximately 19 times lower than the global daily median test rate recorded at

the time[29]. Despite testing limitations, the number of viral genomes generated at a national scale

correlates with the cumulative number of cases recorded throughout the development of the epidemic

(up to March 2022), with the majority of genomes coming from Mexico City (**Supplementary Figure**

1). In average, 100 genomes per 10,000 cases have been generated, corresponding to approximately

1% of the official COVID-19 cases reported. The distribution of genomes sequenced across states

apparently coincides with the reported numbers of active and cumulative confirmed cases at the time

[35,36,37]. Nonetheless, this is likely impacted by an uneven genome sequencing across states

(particularly evident during the first year of the epidemic), and by differences in the sequencing

intensity across time.

- B.1.1.222

The B.1.1.222 lineage circulated between April 2020 and September 2021, mostly sampled from the

USA (77% of all sampled sequences) and Mexico (22% of all sequences), with a limited detection in

other regions of the world (as reported in https://cov-lineages.org/) [38]. However, the apparent

proportion of genome sequences sampled from different countries reflects sequencing biases across

locations. The first B.1.1.222-assigned genome was collected from Mexico on the 04/04/2020

(Mexico/CMX-INER-0026/2020-04-04) [38], whilst the last B.1.1.222-assigned genome was collected

from the USA on the 14/09/2021 (USA/CA-CDPH-1002006730/2021-09-14). The last B.1.1.222-

assigned genome from Mexico dates to the 26/07/2021 (Mexico/CHH_INER_IMSS_1674/2021-07-

26), two months before the latest collection date for the B.1.1.222 lineage at an international scale.

The B.1.1.222 lineage was continuously sampled from the country between April 2020 and

May 2021, followed by a steady decline in detection after July 2021 (**Figure 1b**). Most of the genome

sampling corresponds to the central region of the country, represented by Mexico City (CMX; **Figure**

2a). Data corresponding to the first year of the epidemic showed that by February 2021, the B.1.1.222

had reached a cumulative frequency of approximately 10% relative to other virus lineages co-

circulating within national territory [32]. Our results show that the sampling frequency of the B.1.1.222 in

Mexico progressively increased after June 2020, initially coinciding with the peak of the first

epidemiological wave in the country, further reached a highest of up to 35% in October 2020 **(Figure**

1b). Thus, the B.1.1.222 lineage is likely associated with the observed increase in the number of

cases at that time. An initial frequency underestimation for the B.1.1.222 lineage is attributable to a

limited and centralized virus genome sequencing during the first year of the epidemic in the country.

Phylogenetic analysis revealed a main clade deriving from a single MRCA (most recent

common ancestor; here denoting lineage emergence), with subsequent introductions into other

countries (namely the USA; **Figure 2a**). The inferred date for this earliest MRCA corresponds to

March 2020, a month before the earliest collection date of the first B.1.1.222-assigned genome

(Mexico/CMX-INER-0026/2020-04-04; **Figure 2a**). During this time, the B.1.1.222 was likely to be

seeded into the USA from Mexico multiple times; thus, all subsequent inferred introduction events are

interpreted as 're-introductions'. In this context, our analysis revealed a minimum of 237 distinct

introduction events from Mexico into the USA (95% HPD interval = [225-250]), and a minimum of 106

distinct events from the USA into Mexico (95% HPD interval = [95-122]; **Figure 2a**). The dates

inferred for re-introduction events ranged from October 2020 to July 2021. The B.1.1.222 lineage

displayed a total persistence of up to 16 months.

●   B.1.1.519

Descending from the B.1.1.222, the B.1.1.519 lineage was first detected in North America, being

mostly sampled in the USA (58% of all sequences) and Mexico (32% of all sequences, as reported in

https://cov-lineages.org/) [32,39–41]. Again, the proportion of genomes from different countries reflect

sequencing biases across geographic regions. The first B.1.1.519-assigned genome was collected in

the USA on the 31/07/2020 (USA/TX-HMH-MCoV-45579/2020-07-31) [42], with the lineage being last

detected on December 2021 [43]. During our initial phylogenetic assessment, most of the B.1.1.519-

assigned genomes collected after November 2021 were identified as outliers, and were excluded from

further analyses. Of note, these genomes were sampled from countries where the lineage was not

reported to be locally circulating (namely Turkey and Africa).

135   In Mexico, the B.1.1.519 lineage was first detected on the 28/08/2020 (Mexico/YUC-NYGC-

39037-20/2020-08-28)   and   was   last   detected   on   the   01/11/2021

(Mexico/CAM_IBT_IMSS_4057/2021-11-01) [32]. By late February 2021, the B.1.1.519 represented the

majority of the viral genomes from the country [32,41]. The spread of the B.1.1.519 lineage raised

awareness in public health authorities, [32,40,41], leading to its designation as a VUM in June 2021 [12].

Cumulative virus genome data shows that by January 2021, the B.1.1.519 progressively displaced

other co-circulating virus lineages, representing up to 75% of all virus genomes generated from the

country, and dominating during the peak of the second wave of infection (**Figure 1a and b**). Most of

the genome sampling again corresponds to the central region of the country, represented by the state

of Puebla (PUE; **Figure 2b**).

145   Phylogenetic analysis revealed a pattern similar to the one observed for the B.1.1.222, with

an earliest MRCA leading to a single clade from which subsequent dispersal events into other

countries occurred (namely USA; **Figure 2b**). This earliest MRCA dates to July 2020, again

representing lineage emergence within the country. Our analysis suggests that the B.1.1.519

circulated cryptically in Mexico for four months prior to its initial detection, being seeded back and

forth between the USA and Mexico during this time (**Figure 2b**). Such observations are congruent

with the collection date of the earliest B.1.1.519 genome sampled globally (July 2020, USA/TX-HMH-

MCoV-45579/2020-07-31), positioned basally to the B.1.1.519 clade. As only good-quality sequences

were included within our dataset, two earlier genome sequences from Mexico excluded from our

datasets collected between August and October 2020 (Mexico/YUC-NYGC-39037-20/2020-08-28 and

Mexico/CMX-INER-IBT-0485/2020-10-13) suggest that the cryptic circulation period for the lineage

could have been shorter. We estimated a minimum number of 121 introduction events from the USA

into Mexico (95% HPD interval = [109-132]), compared to a minimum number of 391 introduction

events from Mexico into the USA (95% HPD interval = [380-402]), all occurring between July 2020

and November 2021 (the latter represented by sequence Mexico/CAM_IBT_IMSS_4057/2021-11-01;

Figure 2b). The B.1.1.519 lineage displayed a total persistence of approximately 16 months (earliest

collection date: 28/08/2020, latest collection date: 01/11/2021).

● B.1.1.7

The B.1.1.7 lineage was first detected in the UK during September 2020, spreading to over 175

countries in over a year [44]. The earliest B.1.1.7-assigned genome from Mexico was first collected on

the 31/12/2020 (Mexico/TAM-InDRE-94/2020-12-31), and the latest on the 13/10/2021

(Mexico/QUE_InDRE_FB47996_S8900/2021-10-13). Cumulative data showed a continuous lineage

detection for over ten months, with a sampling peak observed around May 2021, followed by a steady

decline with almost no detection by late August 2021 (**Figure 1b**). Relative to the epidemic in the

country, the circulation of the B1.1.7 in Mexico coincided with a lower number of recorded cases

observed during May-June 2021 [27,29,45]. Overall, the B.1.1.7 lineage reached a lower genome

sampling frequency (of up to 25%), compared to the other virus lineages studied here (**Figure 1b**).

Most of the genome sampling corresponds to the northern region of the country, represented by the

state of Chihuahua (CHH; **Figure 3a**).

Phylogenetic analysis revealed a minimum of 224 introduction events into Mexico (95% HPD

interval = [219-231]; **Figure 3a**), with an earliest MRCA dating to October 2020, denoting a cryptic

circulation period of approximately two months. Subsequent introduction events occurred between

March and June 2021, with a total lineage persistence of ten months. Seven clades (C1a to C7a)

were identified to represent large local transmission chains (composed of >20 sequences from Mexico

with ancestral nodes supported by a PP value >.80; **Figure 3a, Supplementary Data 2**). The earliest

genome sequences were collected from the north-east region of the country (mostly represented by

the state of Chihuahua, CHH) [32,46] and fall within the C2a identified here, with an MRCA dating to late

January 2021 (**Supplementary Data 2**). Of note, the collection date of the earliest B.1.1.7 genome

from Mexico coincides with at least four independent introduction events that occurred synchronously

during December 2020 (**Figure 3a**).

The C3a and C7a were identified as the largest clades, with both MRCAs dating to February

2021, denoting again independent and synchronous introduction events (**Figure 3a**). C3a was

sampled from 22/32 states in the country, mostly from Mexico City (CMX), followed by southern states

of Yucatán (YUC) and Quintana Roo (ROO; **Supplementary Figure 4, Supplementary Data 2**). The

MRCA for C3a dates to late February 2021, showing a persistence of three months (earliest collection

date: 02/03/2021, latest collection date: 28/06/2021). Within C7a, ~70% of the genomes were

sampled from 20/32 states of the country (mostly represented by the southern state of Tabasco;

TAB), followed by the norther-eastern state of Tamaulipas (TAM; **Supplementary Data 2**). As for the

C3a, the MRCA node for C7a dates to late February 2021, showing a persistence of four months

(earliest collection date: 10/03/2021, latest collection date: 05/07/2021).

• P.1

The P.1 lineage was first detected in Brazil during October 2020 [47], after which it diverged into >20

sub-lineages circulating in different countries[19]. Relevant to the epidemic in Mexico, the P.1.17 virus

sub-lineage was sampled mostly from the USA (66% of all sequences) and Mexico (29% of all

sequences; https://cov-lineages.org/) [19], again keeping in mind the sequencing bias across

geographic regions. We detected at least 13 P.1 sub-lineages in the country, with the majority of

genomes assigned to the P.1.17 sub-lineage (66%), and to a lesser extent to the parental P.1 lineage

(25%) [33]. As our dataset comprises genomes assigned to the P.1 and descending sub-lineages, it is

henceforth referred here as a P.1+. The earliest P.1+ genome from Mexico was collected in January

2021   (Mexico/JAL-InDRE_245/2021-01-28)   and   the   latest   in   November   2021

(Mexico/ROO_IBT_IMSS_4502/2021-11-19). Cumulative data shows that genome sampling

frequency for the P.1+ peaked around June 2021, with almost no detection after September 2021.

Most of the genome sampling corresponds to the southern region of the country, represented by the

states Yucatan and Quintana Roo (YUC and ROO; **Figure 3b**). As for the B.1.1.7, the P.1+ showed

an overall lower genome sampling frequency (of 25%) relative to the other lineages studied here, with

spread coinciding with a decreasing number of cases following the second wave of infection in the

country (**Figure 1b**) [31–33].

Phylogenetic analysis revealed a minimum number of 126 introduction events into Mexico

(95% HPD interval = [120-132]; **Figure 3b**). We identified two well-supported clades named here

C1_P1 (corresponding to P.1) and C1_P1_17 (corresponding to P.1.17; **Figure 3b, Supplementary**

Data 2). Genomes within C1_P1 were mostly collected from the central region of the country (>40%

of the sequences from Mexico City and the State of Mexico; CMX and MEX), and directly descend

from genomes from South America (**Supplementary Figure 5**). The MRCA of clade C1_P1 dates to

March 2021, showing a persistence of seven months (earliest collection date: 07/03/2021, latest

collection date: 01/10/2021). Subsequent smaller introduction events were identified for the P.1, with

most MRCAs positioned closely together (referred here as clade C1_P1 MRCAs 1 and 2; **Figure 3b**).

Genomes within clade C1_P1_17 we mostly sampled from Mexico (75%), and to a lesser

extent from the USA (20%). 'Mexico' genomes are positions basally, and were collected

predominantly from the southern region of the country (90%, represented by the states of Quintana

Roo and Yucatán, ROO and YUC; **Supplementary Figure 5**). The MRCA of C1_P1_17 dates to

October 2020, corresponding to the earliest introduction event inferred in the tree. However, the long

branch between the MRCA and the earliest sequence within the clade suggests a significant under-

sampling, indicating a lag in detection after lineage emergence/introduction (**Figure 3b**). Considering

tip dates only, the C1_P1_17 showed a persistence of five months (earliest collection date:

01/04/2021, latest collection date: 17/09/2021). As for P.1, subsequent P.1.17 introduction events

were identified as smaller clusters, with their MRCAs positioned closely together (referred here as

C1_P1_17 MRCAs; **Figure 3b**). Our results indicate that the P.1 parental lineage was introduced

independently to the P.1.17 and from distinct geographic locations. Contrasting to the P.1 parental

lineage, the P.1.17 showed a limited number of introductions but spread successfully, showing a

persistence of up to five months and establishment within the southern region of the country.

● B.1.617.2

Initially detected in India during October 2020, the B.1.617.2 lineage was associated with an increase

in the number of COVID cases recorded globally after March 2021, spreading globally to become

dominant [48,49]. The parental B.1.617.2 lineage further diverged into >230 descending sub-lineages

(designated as the AY.X sub-lineages) that circulate(d) in different regions of the world [19,48,50]. Again,

as our dataset comprises both B.1.617.2 and AY. X-assigned genomes, it is henceforth referred here

as a B.1.617.2+. The first 'B.1.617.2-like' genome from Mexico was collected on September 2020

(Mexico/AGU-InDRE_FB18599_S4467/2020-09-22), followed by a sporadic genome sampling

throughout January 2021 (with <10 sequences) [51]. However, lineage expansion only occurred after

April 2021 (representative earliest sequence: Mexico/JAL-InDRE_FB13752_S1722/2021-04-21). By

the end of August 2021, the B.1.617.2+ reached a relative frequency of >95%, coinciding with the

peak of the third wave of infection recorded in the country (**Figure 1b**) [52]. Over 80 AY.X sub-lineages

were detected in Mexico, with most genomes being assigned to the AY.20 (22%), AY.26 (13%), and

AY.100 (5%), followed by the AY.113 and other sub-lineages (AY.62 and AY.3) [53]. All of these were

initially characterized as being most prevalent within North America [54–58]. Compared to the other virus

lineages studied here, genome sampling of the B.1.617.2+ lineage in Mexico displayed a more

homogeneous distribution across states, associated with an enhanced genome sampling and

sequencing after May 2021 (**Figure 4**).

Phylogenetic analysis revealed a minimum number of 142 B.1.617.2+ introduction events into

Mexico (95% HPD interval = [125-148]), and a total lineage persistence of seven months (up to

November 30th 2021). We identified six major clades (C1d-C6d), with C1d, C3d, C5d and C6d being

the largest (**Figure 4, Supplementary Data 2**). At least four synchronous introduction events were

identified as the earliest MRCAs dating to April (including the introduction events leading to C3d, C4d

and C6d). Within the smaller clades, C2d comprises genomes from Mexico assigned to the AY.62,

sampled mainly from the state of Yucatán. Clade C4d comprises genomes from Mexico assigned to

the AY.3, sampled from the north, central and south of the country (**Supplementary Data 2**). Within

the larger clades, C1d is composed of genomes from Mexico assigned to AY.26, sampled mostly from

the north (>60%; BCS, SIN, JAL), followed by central (CMX) and south-eastern states (VER, ROO

and YUC). The MRCA of C1d dates to May 2021, showing a persistence of six months (earliest

collection date: 17/05/2021, latest collection date: 12/11/2021). Comparably, C3d is composed of

genomes from Mexico assigned to AY.26, sampled mostly from the north (37%; SIN, BCS and SON),

and to a lesser extent from central, east and southern states. The MRCA of C3d dates to April 2021,

displaying a persistence of seven months (earliest collection date: 15/05/2021, latest collection date:

30/11/2021; **Figure 4, Supplementary Data 2**). Thus, C1d and C3d represent two independent

introductions events of the AY.26 sub-lineage into the country.

Clade C5d is one of the largest clades identified (n=~1500), represented by genomes

assigned to the AY.100 (44%), to the parental B.1.617.2 (40%), and to the AY.113 sub-lineage (12%).

Within C5d, the AY.100 and B.1.617.2 genomes are separated by a central sub-cluster of AY.113-

assigned genomes (**Figure 4, Supplementary Data 2**). Approximately, 70% of the genomes within

C5d were sampled from Mexico (mostly assigned to the AY.100 and AY.113), and 30% from the USA

(mostly assigned to the B.1.617.2). Genomes from Mexico are positioned basally and distally, and

were sampled from all 32 states, but predominantly from the north, centre and south of the country

(>50%; represented by CHH, DUR, NLE, CMX, MEX, CAM, YUC, TAB, CHP and ROO; **Figure 4,**

Supplementary Data 2). Thus, C5d represents the most genetically and geographically diverse clade

identified. The MRCA of C5d dates to May, showing a persistence of up to six months (earliest

collection date: 28/05/2021, latest collection date: 29/11/2021). Finally, C6d is the second largest

clade (n=~1150), composed of genomes mostly assigned AY.20 sub-lineage, mainly sampled from

central region of the country (>60%; represented by CMX, MEX, MOR, MIC and HID), suggesting a

restricted geographic distribution for this transmission chain [59]. The MRCA of C6d dates to April,

displaying a persistence of seven months (earliest collection date: 06/05/2021, latest collection date:

30/11/2021; **Figure 4**).

•   Spread of the B.1.617.2 lineage

Being the largest clades, we further explored viral spread for the C5d and C6d under a

phylogeographic approach (see Methods section 4). For C5d, viral spread within the country is likely

to have occurred from the south (represented by the states of Chiapas and Campeche; CHP and

CAM) into the rest of the country (**Figure 4**). Well-supported transitions (scored under a with a

BFR>100 and a PP>.90, **Supplementary Table 1**) were mostly inferred from the southern state of

Campeche (CAM) into central and northern states, and subsequently from the northern state of

Chihuahua (CHH) into the central and northern region of the country (with some bidirectionality

observed). Additional well-supported transitions were observed from Baja California into Chihuahua

(BCN/BCS into CHH), and from Chihuahua into the USA (arbitrarily represented by the geographic

coordinates of the state of California; **Figure 4**). Contrastingly, for C6d, viral spread within the country

is likely to have occurred from the central region (represented by Mexico City, CMX) into central,

northern and southern states (again with some bidirectionality observed). Additional well-supported

transitions were inferred from the southern state of Chiapas into central and northern states (**Figure 4,**

Supplementary Table 1). Animated visualizations of the spread of C5d and C6d across Mexico is

available as **Supplementary Data 3 and 4**.

●   Linking virus spread and human mobility data

Analysis of human mobility data collected between 01/01/2020 and 31/12/2021 at a national scale

(see Methods section 5) revealed two peaks in human mobility observed across time (**Figure 1c**). The

first occurred between February and April 2021, coinciding with the introduction and spread of the

B.1.1.7 and P.1+ lineages, and with the withdrawal of the B.1.1.519. The second peak was observed

between August and November 2021, coinciding with the spread of the B.1.617.2+. Increased

movements (represented by the cumulative number of trips into a given state) were observed for

Mexico City, followed by Jalisco, the State of Mexico, Nuevo León and Puebla, and to a lesser extent

for Coahuila, Guanajuato and Veracruz (**Figure 1c**). Mean intra-state connectivity within national

territory revealed intensified movements from Mexico City into the State of Mexico, Morelos, Hidalgo,

Puebla, Veracruz and Jalisco, and from Jalisco into Michoacán and Guadalajara (**Supplementary**

Figure 6). However, for both the C5d and C6d clades, no correlation between transitions and the

mean intrastate connectivity was observed (C5d: Adjusted R-squared: 0.006577, F-statistic: 7.15 on 1

and 928 DF, p-value: 0.007628. C6d: Adjusted R-squared: 0.3216, F-statistic: 470.8 on 1 and 990 DF,

p-value: < 2.2e-16), nor with the pairwise distance between states (C5d: Adjusted R-squared:

0.01086, F-statistic: 4.051 on 1 and 277 DF, p-value: 0.04511. C6d: Adjusted R-squared: 0.02296, F-

statistic: 2.715 on 1 and 72 DF, p-value: 0.1038). Many of the clades identified were localized within

the southern region of the country (clades C3a and C7a belonging to the B.1.1.7, C1_P1_17

belonging to the P.1.17, and C2d and C5d belonging to the B.1.617.2). Ranking connectivity between

the southern region of the country (represented by the states of Yucatán, Quintana Roo, Chiapas and

Campeche) and the remaining 28 states did reveal a consistently high number of bidirectional

movements between CAM, CMX and VER (**Supplementary Figure 6**).

DISCUSSION

Our analyses revealed contrasting evolutionary dynamics between lineages, with some

identifiable patterns. The B.1.1.222 and B.1.1.519 are likely to have originated in Mexico, with viral

spread characterized by single and extended transmission chains sustained for over a year. During

this time, these lineages were seeded back and forth into Mexico and the USA, remaining endemic,

but never dominating across the USA. The B.1.1.222 lineage reached a maximum national frequency

of 3.5% in the USA, whilst in Mexico it reached a maximum of 35% (up to October 2020). Similarly,

the B.1.1.519 lineage reached a maximum national frequency of 5% in the USA (up to April 2021),

whilst in Mexico it reached a maximum of 76% (up to March 2021) [38,42]. Thus, a greater number of

genomes sampled from the USA compared to the numbers from Mexico (https://cov-lineages.org/),

reflect sequencing disparities across countries, an important consideration for genomic epidemiology

analyses [1].

The introduction and spread B.1.1.7, P.1 and B.1.617.2 lineages in Mexico was characterized by

multiple introduction events resulting in a few successful extended local transmission chains, with

many clades constrained to specific regions, and displaying a persistence shorter periods of time.

Further similarities were observed between the P.1 and B.1.1.7, which overlapped both spatially and

temporally, coinciding with a decrease in the number of cases following the second wave of infection

in the country. Compared to the B.1.1.222 and the B.1.1.519 (and later to the B.1.617.2), and

contrasting to what was observed for other countries (like UK, USA and Brazil) [20,47,60], the P.1 and

B.1.1.7 lineages did not dominate in Mexico [32,46]. Nonetheless, the epidemiological dynamics of these

VOC in the country are comparable to those observed for other regions of the world [20,47,60]. For

instance, in the USA, the earliest introductions of the B.1.1.7 were synchronous to those observed in

Mexico (occurring between October and November 2020), with a similar spread pattern characterized

by a few extended transmissions chains, often constrained to specific states [60]. Thus, viral spread

does not seem to be linked to a higher number of introduction events, but rather to the extent and

distribution of successful transmission chains, with the size of clades likely reflecting virus

transmissibility [61].

Increasing growth rates (*Rt;* the instantaneous reproduction number measuring how an infection

multiplies [62]) for viral variants within different geographic locations are partially explained by a genetic

background increasing viral fitness [20,47,60]. For instance, in the UK, the B.1.617.2 lineage sweep

occurred at a growth rate of around 60% greater than for the B.1.1.7, resulting from a combination

specific mutations increasing transmissibility and immune evasion from the host population [20]. Within

Mexico, the B.1.1.519 likely displayed an enhanced fitness over the previously dominating B.1.1.222.

In agreement with this observation, an *Rt* of 2.9 was estimated for the B.1.1.519, compared to an *Rt*

of 1.93 for the B.1.1.222 (calculated during epidemic week 2020-46, coinciding with the early

expansion of the B.1.1.519 in the country [40]). As these two lineages were directly phylogenetically

related, they were almost genetically identical [41]. Relative to the parental B.1.1.222, the B.1.1.519

displayed only two amino acid changes within the Spike protein: T478K and P681H [41]. Mutation

T478K is located within the RBD domain, with a possible impact in antibody-mediated neutralization

[63,64]. On the other hand, mutation P681H is located upstream the furin cleavage site within an epitope

signal hotspot [65] and may potentially enhance viral cell entry [66], reduce antibody-mediated recognition

[65], and confer Type I interferon resistance [67].

We speculate that at least one of these mutations may have contributed to the dominance of the

B.1.1.519 lineage over the B.1.1.222 by increasing the *Rt*, as has been documented for other SARS-

CoV-2 subpopulations [21,22,68]. Noteworthy, the mutations observed in the B.1.1.519 were not exclusive

to the lineage, as P681H emerged later and independently in the B.1.1.7 (and to a lesser extent in

P.1, corresponding to 5% of all sampled genomes) [69,70]. Moreover, mutation T478K subsequently

appeared in the B.1.617.2 [63,66,67]. In this scenario, we propose that a somewhat 'shared' genetic

background between the B.1.1.519 and the B.1.1.7 (represented by mutation P681H) may have

limited the spread of this VOC in Mexico. Although the impact of specific mutations on viral fitness

would need to be assessed experimentally, current data derived from the natural virus population

evidences that amino acid changes observed at site 681 of the Spike protein have been

predominantly observed in VOCs, and are likely to yield an evolutionary advantage [63,66,67,71–73].

As for other regions of the world, lineage replacement dynamics during the first year of the

epidemic in Mexico were probably driven by increased virus transmissibility. However, lineage

establishment was likely to be shaped by the changing immune landscape of the local host population

[74]. Relatively constant exposure levels to widespread and genetically similar virus subpopulations for

extended periods of time (represented by the B.1.1.222 and the B.1.1.519 lineages), may have

yielded consistently increasing immunity levels in a somewhat still naïve population (assuming a lower

infection attack rate at that time). As more genetically divergent virus lineages were introduced and

began to spread (the P.1 and B.1.1.7 lineages, and later the B.1.617.2), this resulted in a shift in the

local immune landscape, impacted by a virus genetic background prompting (at least a partial)

evasion of the existing immune responses. In Mexico, a vaccination rate of >50% (reflecting the

proportion of the population immunized with at least two doses) was only reached after December

2021 [29,75], suggesting that immunity levels in the population depended mostly on virus pre-exposure

levels. By December 2020, a nationwide representative seroprevalence was estimated to be of

~33.5% [76,77].

Differences in the proportion of genomes sequenced across states did not always mirror the

geographic distribution and extension of the transmission chains identified, but are likely reflecting an

increasing sampling and sequencing intensity through time and space. Many clades identified were

distributed within the south region of the country, suggesting this area has played a key role in the

spread of SARS-CoV-2, comparable to observations made for other viral epidemics in Mexico

(Gutierrez *et al.*, *in preparation*). Although geographic distances and human mobility trends at state

level resolution did not correlate with the spread pattern of clades C5d and C6d, connectivity

measures between the southern region and Mexico City do mirror some of the transitions observed.

Mobility network analyses were also consistent with decrease in the number cases following

epidemiological peaks (as observed for the second and third wave of infection in the country). Thus,

fluctuating human mobility patterns likely reflect changes in the colour-coded system used to regulate

restrictions leveraged by the risk of infection (including travel), applied at a state level [78].

As geographic distances and human mobility within the country are not potential predictors of viral

spread, the patterns observed are most likely to be explained (at least partially) by both terrestrial and

aerial movements across borders. Nonetheless, determining the geographic origin of introduction

events is limited by genome sampling biases both at a global and local scale [79,80]. Such biases also

affects the viral spread reconstructions we present, likely to be impacted by *i*) an uneven genome

sampling across all Mexico states, by *ii*) an uneven sampling of foreign locations at a regional level,

and by *iii*) a poor viral genome representation for many countries (*i.e.*, Central and South America)

[51,81]. Taking this into consideration, it has been hypothesized that the introduction and spread of

SARS-CoV-2 in Mexico would be strongly linked to mobility between Mexico and the USA (for an

example, see [46]). Although human movements across these countries have indeed contributed to

spread of some lineages (clearly for the B.1.1.222 and the B.1.1.519), some of the viral diffusion

patterns observed across national territory are more likely to be congruent with human mobility linked

to migration routes from South and Central America into Mexico. This raises the question of the

potential role of irregular migration on virus dispersal across countries, and further highlights the need

of using multiple sources for human mobility data across borders (such as mobile-usage, in addition to

air traffic) to inform phylogeographic analyses [82–85].

METHODS (1592/3000 WORDS)

1.   Data collation and initial sequence alignments

Global genome datasets assigned to each PANGO lineage [10] under investigation (B.1.1.222,

B.1.1.519, B.1.1.7, P.1 and B.1.617.2) were downloaded with their associated metadata from the

GISAID platform as of November 30th 2021 [51,86]. The total number of sequences retrieved sampled

globally for each virus lineage were the following: B.1.1.222 = 3,461, B.1.1.519 = 19,246, B.1.1.7 =

913,868, P.1 = 87,452, and B.1.617.2 = 2,166,874. This also included the virus genome sequences

available from Mexico up the sampling date of this study. Sampling date represents the peak for

B.1.617.2 frequency in Mexico before the introduction of 'Omicron' into the country [30,31]. Sequences

were further excluded if they displayed incomplete collection dates, and/or if >1000 nt shorter than full

genome length, and/or if showing >10% of sites coded as ambiguities (including N or X). Individual

datasets were then processed using the Nextclade pipeline to filter sequences according to sequence

quality [87]. In addition, a set of the earliest SARS-CoV-2 sequences sampled from 2019-2020

(including reference sequence Wuhan-Hu-1, GenBank accession ID: MN908947.3), and a set of viral

genomes representing early 'Pango' diversity sampled up to 31/05/2020 were added for rooting

purposes   (https://github.com/BernardoGG/SARS-CoV-2_Genomic_lineages_Ecuador).   Output

datasets were mapped to the reference sequence Wuhan-Hu-1 (GenBank: MN908947.3) using

Minimap2 [88] to generate whole genome alignments. The main viral ORFs (Orf1ab and S) were

extracted to generate reduced-length alignments of approximately 25,000 bases long, comprising only

the largest and most phylogenetically informative coding genome regions, to exclude smaller ORFs,

UTRs, and short intergenic sequences.

2.   Migration data and phylogenetically-informed subsampling

To provide an overview for global introductory events into Mexico as a proxy for dataset reduction, we

used the Google COVID-19 Aggregated Mobility Research Database, describing anonymized relative

human mobility flow into different geographical regions based on mobile data usage [89,90]. For any

given dataset, all 'non-Mexico' sequences are sorted according to their location, selecting only the top

5 countries representing the most intense human mobility flow into Mexico. Then, 'Mexico' sequences

are subsampled to ~4,000 in proportion to the total number of cases reported in the country across

time (corresponding to epidemiological weeks) according to publicly accessible epidemiological data

[91]. This enables a homogeneous epidemiological and temporal representation of genomes. In case of

a given lineage reporting sub-lineages, datasets are further reduced by selecting the top 5 sub-

lineages that circulate(d) within the country. For 'globally sampled sequences', datasets are further

reduced randomly to an approximate 1:1 ratio relative to the number of 'Mexico' sequences (keeping

sequences with the earliest and latest collection dates sampled both from Mexico and globally). This

migration-informed approach, coupled with balancing the number of sequences from different

countries, aims to mitigate the effects of geographical over-representation impacting phylogeographic

analyses, in which clades tend to not segregate fully, and certain regions may appear as more

frequent seeding sources than they are in reality [1].

For each dataset, a preliminary Maximum Likelihood (ML) tree is inferred using IQ-TREE

(command line: iqtree -s -m GTR+I+G -alrt 1000) [92]. The pipeline then uses a modified version of

Treemmer v0.3 (https://github.com/fmenardo/Treemmer/releases) to reduce the size and redundancy

of the resulting ML trees with a minimal loss of diversity [93]. Subsampling is based on phylogenetic

clustering patterns, reducing the noise derived from clonal and phylogenetically overrepresented

sequences. For this, the -lm command is initially used to protect 'Mexico' sequences and those added

for rooting purposes. During the pruning iterations, the -pp command is then used to protect 'Mexico'

clusters and pairs of 'non-Mexico' sequences that are immediately ancestral or directly descending

from these. This results in reduced datasets that render local computational analyses feasible, in

which clades may appear to be smaller relative to the raw counts of genomes publicly available, but

reflect the sampled viral genetic diversity. Datasets are used for re-estimating ML trees as described,

to be then inputted for a time-scaled phylogenetic analysis (see Methods section 4). Plots

representing genome data distribution before and after implementing our subsampling approach are

available in the **Supplementary Figure 4**. The pipeline is publicly accessible at

(https://github.com/rhysinward/Mexico_subsampling).

3. Dataset construction

Given the reduced size of the original B.1.1.222 dataset, all sequences retained after initial quality filtering were used for analyses. This resulted in a 3,849-sequence alignment (including 760 genomes from Mexico) used for ML tree inference using IQ-TREE (iqtree -s -m GTR+I+G -alrt 1000; **Supplementary Data 1**). The resulting tree was further inputted for a time-scaled analysis (see Methods section 4). All other datasets (B.1.1.519, B.1.1.7, P.1 and B.1.617.2) were processed under the pipeline described above to render informative datasets for phylogeographic analysis (see Methods section 2 and 4). The B.1.1.519 final dataset resulted in a 5,001-sequence alignment, including 2,501 genomes from Mexico. The B.1.1.7 final dataset resulted in a 7,049-sequence alignment, including 1,449 genomes from Mexico. The P.1 final dataset resulted in a 5,493-sequence alignment, including 2,570 genomes from Mexico. The B.1.617.2 final dataset resulted in a 5,994-sequence alignment, including 3,338 genomes from Mexico (**Supplementary Data 1**). Individual datasets were then used for ML tree inference as described, with the resulting trees used as input for time-scaled analyses (described in Methods section 4).

4. Time-scaled analysis

Output ML trees were assessed for temporal signal using TempEst v1.5.3 [94], removing outliers and re-estimating trees when necessary. Trees were then time-calibrated informed by tip sampling dates using TreeTime [95] (command line: treetime -aln --tree --clock-rate 8e-4 --dates --keep-polytomies --clock-filter 0). Due to a low temporal signal, a fixed clock rate corresponding to a reported virus evolutionary rate estimate of 8×10^{-4} substitutions per site per year was used [96,97]. Root-to-tip regression plots for the ML trees (prior to time calibration and excluding rooting sequences) are available in the **Supplementary Figure 3**. In order to investigate viral introductions into Mexico, the time-calibrated trees were used as input for a discrete trait analysis (DTA, or 'discrete phylogeographic inference') using BEAST v1.10.4 to generate maximum clade credibility (MCC) trees [98–100]. Using fixed 'time-calibrated' tree as an input for DTA in an effective way of circumventing the restrictions of computationally-expensive analyses on large datasets [98]. However, although this approach allows inferring dates and number for introduction events into the study area, it does not consider the uncertainty associated with phylogenetic inference (*i.e.*, the dates of inferred MRCAs do not come with a credibility intervals). Introduction events into the country are associated with the most

recent common ancestor (MRCA) node of each 'Mexico' clade. Following a similar strategy as

described in du Plessis et al.[3] 'Mexico' clades were identified as those composed by a minimum of

two sister 'Mexico' sequences directly descending from another 'Mexico' sequence. For all

introduction events identified in the MCC trees, the mean and associated HPD interval were

assessed.

Analyses for the B.1.1.7 and the P.1 and B.1.617.2 datasets were performed to estimate the

number of introduction events into Mexico from other (unknown) geographic regions, and only two

locations were considered: 'Mexico' and 'other'. For the B.1.1.222 and B.1.1.519 dataset, we

estimated the number of transitions between Mexico and the USA, thus considering three distinct

geographic locations: 'Mexico', 'USA' and 'other'. Each Monte Carlo Markov Chain (MCMC) was run

for 10^6 iterations, sampling every 10^3 iterations. To assess if sufficient effective sample size values

(*i.e.,* ESS>200) were associated with the estimated parameters, we then inspected MCMC

convergence and mixing using the program Tracer 1.7 [101]. To explore viral spread across the country,

two additional DTAs were performed within each of the largest monophyletic clades identified within

the B.1.617.2+ tree (C5d and C6d, see Results section). These additional DTAs were based on the

definition of 33 distinct sampling locations (all 32 states from Mexico + 'other' for genomic sequences

sampled from outside the country). Visualization of viral spread patterns was performed using

SpreadViz (https://spreadviz.org/home), an updated web implementation of the Spatial Phylogenetic

Reconstruction of Evolutionary Dynamics software SpreaD3 [102], also used to estimate Bayes Factor

Rate (BFR) values to identify well-supported transitions between locations [100] (**Supplementary Table**

1).

5. Human mobility data analysis and exploring correlations with genomic data

Human mobility data used for this study derived from anonymized mobile device locations provided by

the company Veraset collected between 01/01/2020 and 31/12/2021 within national territory. The

source dataset includes anonymized identifiers for mobile devices, geographical coordinates (latitude

and longitude) and a timestamp. The dataset was used to construct aggregated intra-state mobility

networks, where nodes are defined as each of the 32 states from the country, whilst (weighed and

directed) edges represent the normalized volume of observed trips between nodes [103]. The resulting

networks were then used to quantify the number of cumulative trips from any state into a given

specific state across time, the geographic distances among states, the mean intrastate connectivity

observed between April 2021 and November 2021 (corresponding to the expansion period for the

B.1.617.2 lineage, see **Figure 4b**), and for ranking connectivity between the southern region of the

country (represented by Yucatán, Quintana Roo, Chiapas and Campeche) and the remaining 28

states (see **Supplementary Figure 8,** matrixes available as **Supplementary Data 5**). The

connectivity measure was defined as the sum of the weights for edges that go from any given node

into other node(s), reflecting the number of trips in any direction. We then used the 'PhyCovA'

software tool (https://evolcompvir-kuleuven.shinyapps.io/PhyCovA/) to perform preliminary analysis

for exploring the human mobility data from the country as a potential predictor of viral transition

among locations [104]. 'PhyCovA' was chosen as an explanatory approach over a fully-integrated GLM

implemented in the Bayesian BEAST framework, as the last one would imply a high computational

burden related to our datasets [99].

DATA AVAILABILITY

Virus genome IDs and GISAID accession numbers for the sequences used in each dataset are

provided in the **Supplementary Data 1** file. All genomic and epidemiological data supporting the

findings of this study is publicly available from GISAID/GenBank, from the Ministry Of Health

Mexico[105], and/or from the 'Our world in data coronavirus pandemic web portal [29]. For the GISAID

data used, the corresponding acknowledgements table is available on the 'GISAID Data

Acknowledgement Locator' under the EPI_SET_20220405qd and EPI_SET_20220215at keys [51]. Our

bioinformatic pipeline implementing a migration data and phylogenetically-informed sequence

subsampling approach is publicly available at https://github.com/rhysinward/Mexico_subsampling.

AUTHOR CONTRIBUTIONS

HGCS, LD, BG, RPDI, SD, and MEZ performed research and analysed data. RPDI developed the

bioinformatic pipeline, whilst RPDI, NMV and CB processed data. MK, BG and RPDI, together with

the Mobility Team in Mexico [MHR, AM, MF, OF, PG, GOA, GAJ] performed the analysis of human

mobility data integrated with genomic data. Other members of CoViGen-Mex and ACG, BT, SZ, SL,

CFA, AL and MEZ contributed to the initial experimental design by leading evolutionary analyses part

of the duties of the consortium. OGP and SD contributed to revising the methodological approach.

MEZ supervised and coordinated data analysis. MEZ wrote the manuscript, with contributions and

comments from all authors.

COMPETING INTERESTS

The authors declare no competing interests.

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obtaining the specimens, and their submitting laboratories for generating the genetic sequence and

metadata and sharing via the GISAID initiative, on which this research is based.

FIGURES

Figure 1. Overview of the SARS-CoV-2 epidemic in Mexico

(a) Time-scaled phylogeny for representative viral genomes from Mexico within a global context, highlighting the positioning of the B.1.1.222, B.1.1.519, B.1.1.7, P.1 and B.1.617.2 virus lineages. (b) The epidemic curve for COVID-19 in Mexico from January 2020 up to November 2021, showing the average number of daily cases (red line) and the associated excess mortality (grey line). The peak for the first, second and third waves of infection are highlighted in yellow shadowing, whilst the lower panel represents the percentage of SARS-CoV-2 genomes from Mexico assigned to distinct 'Pango' lineages within the given timeframe. Lineage B.1.1.222 is shown in light green, B.1.1.519 in yellow, P.1 in red (Gamma), B.1.1.7 (Alpha) in dark green, and B.1.617.2 (Delta) in teal. Other lineages displaying a lower sampling frequency are jointly shown in purple. The time period for the implementation of non-pharmaceutical interventions at national scale is highlighted in grey shadowing. (c) Heatmap displaying the volume of trips into a given state from any other state recorded from January 2020 up to November 2021 derived from anonymized mobile device geolocated and time-stamped data. The dashed red line corresponds to the start date national vaccination campaign (December 2020), whilst the dashed black line represents the implementation date of a broader viral genome sampling and sequencing scheme for the surveillance of SARS-CoV-2 in Mexico (after May 2021).

Figure 2. Time-scaled phylogenetic analyses for the B.1.1.222 and B.1.1.519 lineage

Maximum clade credibility (MCC) trees for the (a) B.1.1.222 and (b) B.1.1.519 lineages, in which the main clades corresponding to distinct introduction events into Mexico are highlighted. Nodes shown as outline circles correspond to the most recent common ancestor (MRCA) of clades resulting from a distinct introduction event into Mexico (in teal) or the USA (in ochre). The estimated circulation period in Mexico for a given lineage is highlighted in yellow shadowing. The dashed purple line represents the collection date for the earliest lineage-assigned genome sampled from Mexico, with its position indicated with an arrow. The dashed yellow line represents the implementation date (May 2021) of a broader viral genome sequencing scheme for the surveillance of SARS-CoV-2 in Mexico. Below, map graphs show the cumulative proportion of viral genomes from Mexico assigned to each virus lineage relative to the total number of genome sequences reported for any given state (taken from GISAID, corresponding to raw data before subsampling).

Figure 3. Time-scaled phylogenetic analyses for the B.1.1.7 and P.1+ lineages

Maximum clade credibility (MCC) trees for the (a) B.1.1.7 and the (b) P.1+ B.1.617.2 lineages, in which major clades identified as distinct introduction events into Mexico are highlighted. Nodes shown as red outline circles correspond to the most recent common ancestor (MRCA) of clades resulting from a distinct introduction event into Mexico. The estimated circulation period for a given lineage is highlighted in yellow shadowing. For each panel, the dashed purple line represents the collection date for the earliest lineage-assigned genome sampled from Mexico, with its position within the tree indicated with an arrow. The dashed yellow line represents the implementation date (May 2021) of a broader viral genome sampling and sequencing scheme for the surveillance of SARS-CoV-2 in Mexico. Below, map graphs show the cumulative proportion of viral genomes from Mexico assigned to each virus lineage of interest relative to the total number of genome sequences reported for any given state (taken from GISAID, corresponding to raw data before subsampling).

Figure 4. Time-scaled and phylogeographic analysis for the B.1.617.2+ lineage

Maximum clade credibility (MCC) tree for the B.1.617.2+ lineage, in which major clades identified as distinct introduction events into Mexico are highlighted. Nodes shown as red outline circles correspond to the most recent common ancestor (MRCA) of clades resulting from a distinct introduction event into Mexico. The estimated circulation period is highlighted in yellow shadowing. The dashed purple line represents the collection date for the earliest lineage-assigned genome sampled from Mexico, with its position within the tree indicated with an arrow. The dashed yellow line represents the implementation date (May 2021) of a broader viral genome sampling and sequencing scheme for the surveillance of SARS-CoV-2 in Mexico. On the left, a map graph sows the cumulative proportion of sequenced viral genomes from Mexico assigned as B.1.617.2+, relative to the total number of genome sequences reported for any given state (taken from GISAID, corresponding to raw data before subsampling). On the right, a zoom-in to the C5d and C6d clades shows sub-lineage composition and the inferred location for nodes. The geographic spread of these clades across Mexico is shown from the inferred discrete phylogeographic analysis (see Methods section 4). Viral lineage transition events between Mexican states are represented by curved lines coloured according to sampling location, showing only well-supported transitions (Bayes factor rate >100 and a posterior probability >0.9). Bayes factor rate values are all reported in the Supplementary Table 1.

REFERENCES

1. Hill, V., Ruis, C., Bajaj, S., Pybus, O. G. & Kraemer, M. U. G. Progress and challenges in virus genomic epidemiology. *Trends Parasitol.* **37**, 1038–1049 (2021).

2. Kraemer, M. U. G. *et al.* Spatiotemporal invasion dynamics of SARS-CoV-2 lineage B.1.1.7 emergence. *Science* **373**, 889–895 (2021).

3. du Plessis, L. *et al.* Establishment and lineage dynamics of the SARS-CoV-2 epidemic in the UK. *Science* **371**, 708–712 (2021).

4. Worobey, M. *et al.* The emergence of SARS-CoV-2 in Europe and North America. *Science* **370**, 564–570 (2020).

5. Candido, D. S. *et al.* Evolution and epidemic spread of SARS-CoV-2 in Brazil. *Science* **369**, 1255–1260 (2020).

6. COVID-19 Genomics UK (COG-UK) consortiumcontact@cogconsortium.uk. An integrated national scale SARS-CoV-2 genomic surveillance network. *Lancet Microbe* **1**, e99–e100 (2020).

7. Chen, Z. *et al.* A global analysis of replacement of genetic variants of SARS-CoV-2 in association with containment capacity and changes in disease severity. *Clin. Microbiol. Infect.* **27**, 750–757 (2021).

8. Li, J., Lai, S., Gao, G. F. & Shi, W. The emergence, genomic diversity and global spread of SARS-CoV-2. *Nature* **600**, 408–418 (2021).

9. Tao, K. *et al.* The biological and clinical significance of emerging SARS-CoV-2 variants. *Nat. Rev. Genet.* **22**, 757–773 (2021).

10. Rambaut, A. *et al.* A dynamic nomenclature proposal for SARS-CoV-2 lineages to assist genomic epidemiology. *Nat Microbiol* **5**, 1403–1407 (2020).

11. Pango Network – Helping track the transmission and spread of SARS-CoV-2. https://www.pango.network/.

12. Tracking SARS-CoV-2 variants. https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/.

13. CDC. Science brief: Emerging SARS-CoV-2 variants. *Centers for Disease Control and Prevention* https://www.cdc.gov/coronavirus/2019-ncov/science/science-briefs/scientific-brief-emerging-variants.html?CDC_AA_refVal=https://www.cdc.gov/coronavirus/2019-ncov/more/science-and-research/scientific-brief-emerging-variants.html (2022).

14. Horby, P. *et al.* https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/961037/NERVTAG_note_on_B.1.1.7_severity_for_SAGE_77__1_.pdf.

15. Weisblum, Y. *et al.* Escape from neutralizing antibodies by SARS-CoV-2 spike protein variants. *Elife* **9**, e61312 (2020).

16. Greaney, A. J. *et al.* Mapping mutations to the SARS-CoV-2 RBD that escape binding by different classes of antibodies. *Nat. Commun.* **12**, 4196 (2021).

17. Greaney, A. J. *et al.* Complete mapping of mutations to the SARS-CoV-2 spike receptor-binding domain that escape antibody recognition. *Cell Host Microbe* **29**, 44-57.e9 (2021).

18. Classification of Omicron (B.1.1.529): SARS-CoV-2 variant of Concern. https://www.who.int/news/item/26-11-2021-classification-of-omicron-(b.1.1.529)-sars-cov-2-variant-of-concern.

19. Cov-lineages. https://cov-lineages.org/.

20. Vöhringer, H. S. *et al.* Genomic reconstruction of the SARS-CoV-2 epidemic in England. *Nature* **600**, 506–511 (2021).

21. Martin, D. P. *et al.* The emergence and ongoing convergent evolution of the N501Y lineages coincides with a major global shift in the SARS-CoV-2 selective landscape. *medRxiv* 2021.02.23.21252268 (2021).

22. Escalera-Zamudio, M. *et al.* Identification of site-specific evolutionary trajectories shared across human betacoronaviruses. *bioRxivorg* (2021) doi:10.1101/2021.05.24.445313.

23. Kumar, S. *et al.* An evolutionary portrait of the progenitor SARS-CoV-2 and its dominant offshoots in COVID-19 pandemic. *Mol. Biol. Evol.* **38**, 3046–3059 (2021).

24. Caniels, T. G. *et al.* Emerging SARS-CoV-2 variants of concern evade humoral immune responses from infection and vaccination. *Sci. Adv.* **7**, eabj5365 (2021).

25. Zhang, X. *et al.* Viral and host factors related to the clinical outcome of COVID-19. *Nature* **583**, 437–440 (2020).

26. Boutin, S. *et al.* Host factors facilitating SARS-CoV-2 virus infection and replication in the lungs. *Cell. Mol. Life Sci.* **78**, 5953–5976 (2021).

27. https://globalhealthsciences.ucsf.edu/sites/globalhealthsciences.ucsf.edu/files/mexico-covid-19-case-study-english.pdf.

28. Taboada, B. *et al.* Genomic analysis of early SARS-CoV-2 variants introduced in Mexico. *J. Virol.* **94**, (2020).

29. Ritchie, H. *et al.* Coronavirus Pandemic (COVID-19). *Our World in Data* (2020).

30. MexCoV2. http://mexcov2.ibt.unam.mx:8080/COVID-TRACKER/.

31. Vigilancia de variantes del virus SARS-CoV-2. *Vigilancia de variantes del virus SARS-CoV-2* https://salud.conacyt.mx/coronavirus/variantes/.

32. Taboada, B. *et al.* Genetic analysis of SARS-CoV-2 variants in Mexico during the first year of the COVID-19 pandemic. *Viruses* **13**, 2161 (2021).

33. MexCoV2. http://mexcov2.ibt.unam.mx:8080/COVID-TRACKER/tablero.

34. Gutierrez, B. *et al.* Emergence and widespread circulation of a recombinant SARS-CoV-2 lineage in North America. *Cell Host Microbe* (2022) doi:10.1016/j.chom.2022.06.010.

35. https://www.gob.mx/cms/uploads/attachment/file/571250/Comunicado_Tecnico_Diario_COVID-19_2020.08.19__1_.pdf.

36. https://www.gob.mx/cms/uploads/attachment/file/626111/Comunicado_Tecnico_Diario_COVID-19_2021.03.30.pdf.

37. https://www.gob.mx/cms/uploads/attachment/file/648612/Comunicado_Tecnico_Diario_COVID-19_2021.06.25.pdf.

38. Cov-lineages. https://cov-lineages.org/lineages/lineage_B.1.1.222.html.

39. https://cov-lineages.org/lineages/lineage_B.1.1.519.html,.

40. Cedro-Tanda, A. *et al.* The evolutionary landscape of SARS-CoV-2 variant B.1.1.519 and its clinical impact in Mexico city. *Viruses* **13**, 2182 (2021).

41. Rodríguez-Maldonado, A. P. *et al.* Emergence and spread of the potential variant of interest (VOI) B.1.1.519 of SARS-CoV-2 predominantly present in Mexico. *Arch. Virol.* **166**, 3173–3177 (2021).

42. Cov-lineages. https://cov-lineages.org/lineages/lineage_B.1.1.519.html.

43. Outbreak.Info. *outbreak.info* https://outbreak.info/situation-reports?pango=B.1.1.519.

44. Cov-lineages. https://cov-lineages.org/global_report_B.1.1.7.html.

45. https://www.gob.mx/cms/uploads/attachment/file/655969/Comunicado_Tecnico_Diario_COVID-19_2021.07.21.pdf.

46. Zárate, S. *et al.* The Alpha Variant (B.1.1.7) of SARS-CoV-2 Failed to Become Dominant in Mexico. *Microbiol Spectr* **10**, e0224021 (2022).

47. Faria, N. R. *et al.* Genomics and epidemiology of the P.1 SARS-CoV-2 lineage in Manaus, Brazil. *Science* **372**, 815–821 (2021).
48. Cov-lineages. https://cov-lineages.org/global_report_B.1.617.2.html.
49. Tian, D., Sun, Y., Zhou, J. & Ye, Q. The global epidemic of the SARS-CoV-2 Delta variant, key spike mutations and immune escape. *Front. Immunol.* **12**, 751778 (2021).
50. https://cov-lineages.org/lineage_list.html,.
51. GISAID - initiative. https://www.gisaid.org/.
52. SSA/SPPS/DGE/DIE/InDRE/UIES/Informe técnico. COVID. https://www.gob.mx/cms/uploads/attachment/file/684495/Comunicado_Tecnico_Diario_COVID-19_2021.11.30.pdf.
53. Taboada, B. *et al.* Dominance of three sublineages of the SARS-CoV-2 Delta variant in Mexico. *Viruses* **14**, 1165 (2022).
54. Cov-lineages. https://cov-lineages.org/lineage.html?lineage=AY.20.
55. Cov-lineages. https://cov-lineages.org/lineage.html?lineage=AY.26.
56. Cov-lineages. https://cov-lineages.org/lineage.html?lineage=AY.100.
57. Cov-lineages. https://cov-lineages.org/lineage.html?lineage=AY.113.
58. Cov-lineages. https://cov-lineages.org/lineage.html?lineage=AY.3.
59. Taboada, B. *et al.* Dominance of three sublineages of the SARS-CoV-2 Delta variant in Mexico. (2022) doi:10.20944/preprints202205.0131.v1.
60. Washington, N. L. *et al.* Emergence and rapid transmission of SARS-CoV-2 B.1.1.7 in the United States. *Cell* **184**, 2587-2594.e7 (2021).
61. Grenfell, B. T. *et al.* Unifying the epidemiological and evolutionary dynamics of pathogens. *Science* **303**, 327–332 (2004).
62. https://rss.org.uk/RSS/media/File-library/Publications/Special.
63. Liu, C. *et al.* Reduced neutralization of SARS-CoV-2 B.1.617 by vaccine and convalescent serum. *Cell* **184**, 4220-4236.e13 (2021).
64. Wilhelm, A. *et al.* Antibody-mediated neutralization of authentic SARS-CoV-2 B.1.617 variants harboring L452R and T478K/E484Q. *Viruses* **13**, 1693 (2021).
65. Haynes, W. A., Kamath, K., Lucas, C., Shon, J. & Iwasaki, A. Impact of B.1.1.7 variant mutations on antibody recognition of linear SARS-CoV-2 epitopes. *bioRxiv* 2021.01.06.20248960 (2021) doi:10.1101/2021.01.06.20248960.
66. Lubinski, B. *et al.* Functional evaluation of the P681H mutation on the proteolytic activation the SARS-CoV-2 variant B.1.1.7 (Alpha) spike. *bioRxivorg* (2021) doi:10.1101/2021.04.06.438731.
67. Lista, M. J. *et al.* The P681H mutation in the Spike glycoprotein confers Type I interferon resistance in the SARS-CoV-2 alpha (B.1.1.7) variant. *bioRxiv* 2021.11.09.467693 (2021) doi:10.1101/2021.11.09.467693.
68. Dolan, P. T., Whitfield, Z. J. & Andino, R. Mapping the evolutionary potential of RNA viruses. *Cell Host Microbe* **23**, 435–446 (2018).
69. Emergence and spread of SARS-CoV-2 P.1 (Gamma) lineage variants carrying Spike mutations Δ141-144, N679K or P681H during persistent viral circulation in Amazonas, Brazil. *Virological* https://virological.org/t/emergence-and-spread-of-sars-cov-2-p-1-gamma-lineage-variants-carrying-spike-mutations-141-144-n679k-or-p681h-during-persistent-viral-circulation-in-amazonas-brazil/722 (2021).
70. CoVariants: 20J (gamma, V3). https://covariants.org/variants/20J.Gamma.V3.
71. auspice. https://nextstrain.org/groups/neherlab/ncov/S.P681?c=gt-S_681.
72. https://observablehq.com/@spond/spike-trends,.
73. CoVariants: S:P681. https://covariants.org/variants/S.P681.
74. Gupta, S. & Anderson, R. M. Population structure of pathogens: the role of immune selection. *Parasitol. Today* **15**, 497–501 (1999).
75. Bhatia, G., Dutta, P. K., McClure, J. & Reuters Graphics. Mexico: the latest coronavirus counts, charts and maps. *Reuters* (2020).
76. Basto-Abreu, A. *et al.* Nationally representative SARS-CoV-2 antibody prevalence estimates after the first epidemic wave in Mexico. *Nat. Commun.* **13**, 589 (2022).
77. Muñoz-Medina, J. E. *et al.* SARS-CoV-2 IgG antibodies seroprevalence and Sera neutralizing activity in MEXICO: A national cross-sectional study during 2020. *Microorganisms* **9**, 850 (2021).
78. https://coronavirus.gob.mx/semaforo/.
79. Kalkauskas, A. *et al.* Sampling bias and model choice in continuous phylogeography: Getting lost on a random walk. *PLoS Comput. Biol.* **17**, e1008561 (2021).
80. De Maio, N., Wu, C.-H., O'Reilly, K. M. & Wilson, D. New routes to phylogeography: A Bayesian STructured coalescent Approximation. *PLoS Genet.* **11**, e1005421 (2015).
81. No title. https://academic.oup.com/ve/article/7/2/veab051/6292076?login=true.
82. Kraemer, M. U. G. *et al.* The effect of human mobility and control measures on the COVID-19 epidemic in China. *Science* **368**, 493–497 (2020).
83. Migration data in central America. *Migration data portal* https://www.migrationdataportal.org/regional-data-overview/migration-data-central-america.
84. Migrants trapped in the Mexican vertical border. *Oxford Law Faculty* https://www.law.ox.ac.uk/research-subject-groups/centre-criminology/centreborder-criminologies/blog/2018/06/migrants-trapped (2018).
85. París-Pombo, M. D. & El Colegio de la Frontera Norte, México. Trayectos peligrosos: inseguridad y movilidad humana en México. *Papeles Poblac.* **22**, 145–172 (2016).
86. Shu, Y. & McCauley, J. GISAID: Global initiative on sharing all influenza data - from vision to reality. *Euro Surveill.* **22**, (2017).
87. Aksamentov, I., Roemer, C., Hodcroft, E. & Neher, R. Nextclade: clade assignment, mutation calling and quality control for viral genomes. *J. Open Source Softw.* **6**, 3773 (2021).
88. Li, H. Minimap2: pairwise alignment for nucleotide sequences. *Bioinformatics* **34**, 3094–3100 (2018).
89. Kraemer, M. U. G. *et al.* Mapping global variation in human mobility. *Nat. Hum. Behav.* **4**, 800–810 (2020).
90. Inward, R. P. D., Parag, K. V. & Faria, N. R. Using multiple sampling strategies to estimate SARS-CoV-2 epidemiological parameters from genomic sequencing data. *bioRxiv* 2022.02.04.22270165 (2022) doi:10.1101/2022.02.04.22270165.
91. COVID-19 tablero México. *COVID - 19 Tablero México* https://datos.covid-19.conacyt.mx/.
92. Minh, B. Q. *et al.* IQ-TREE 2: New Models and Efficient Methods for Phylogenetic Inference in the Genomic Era. *Mol. Biol. Evol.* **37**, 1530–1534 (2020).

93. Menardo, F. *et al.* Treemmer: a tool to reduce large phylogenetic datasets with minimal loss of diversity. *BMC Bioinformatics* **19**, (2018).

94. Rambaut, A., Lam, T. T., Max Carvalho, L. & Pybus, O. G. Exploring the temporal structure of heterochronous sequences using TempEst (formerly Path-O-Gen). *Virus Evol.* **2**, vew007 (2016).

95. Sagulenko, P., Puller, V. & Neher, R. A. TreeTime: Maximum-likelihood phylodynamic analysis. *Virus Evol.* **4**, vex042 (2018).

96. Su, Y. C. F. *et al.* Discovery and genomic characterization of a 382-nucleotide deletion in ORF7b and ORF8 during the early evolution of SARS-CoV-2. *MBio* **11**, (2020).

97. MacLean, O. A., Orton, R. J., Singer, J. B. & Robertson, D. L. No evidence for distinct types in the evolution of SARS-CoV-2. *Virus Evol.* **6**, veaa034 (2020).

98. Dellicour, S. *et al.* A phylodynamic workflow to rapidly gain insights into the dispersal history and dynamics of SARS-CoV-2 lineages. *Mol. Biol. Evol.* **38**, 1608–1613 (2021).

99. Suchard, M. A. *et al.* Bayesian phylogenetic and phylodynamic data integration using BEAST 1.10. *Virus Evol.* **4**, vey016 (2018).

100. Lemey, P., Rambaut, A., Drummond, A. J. & Suchard, M. A. Bayesian phylogeography finds its roots. *PLoS Comput. Biol.* **5**, e1000520 (2009).

101. Rambaut, A., Drummond, A. J., Xie, D., Baele, G. & Suchard, M. A. Posterior summarization in Bayesian phylogenetics using tracer 1.7. *Syst. Biol.* **67**, 901–904 (2018).

102. Bielejec, F. *et al.* SpreaD3: Interactive visualization of spatiotemporal history and trait evolutionary processes. *Mol. Biol. Evol.* **33**, 2167–2169 (2016).

103. Fontanelli, O. *et al.* Intermunicipal travel networks of Mexico (2020-2021). *arXiv [cs.SI]* (2022).

104. Blokker, T., Baele, G., Lemey, P. & Dellicour, S. Phycova - a tool for exploring covariates of pathogen spread. *Virus Evol.* **8**, veac015 (2022).

105. https://covid19.sinave.gob.m.