Substrate binding and lipid-mediated allostery in the human organic anion transporter 1 at the atomic-scale

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Highlights

- Adefovir has at least two binding pockets on *h*OAT1 in the outward-facing conformation.
- The highly conserved B-motif within MFS is strongly involved in substrate binding.
- α-Ketoglutarate binds to the intracellular domain of *h*OAT1 and destabilizes its OF conformation.
- The lipid membrane bilayer plays an active role in the allosteric communication between intracellular and extracellular domains of *h*OAT1.

Abstract

The Organic Anion Transporter 1 is a membrane transporter known for its central role in drug elimination by the kidney. *h*OAT1 is an antiporter, *i.e.*, it translocates substrate in exchange for α-ketoglutarate. The pharmacological attention is drawn by *h*OAT1-mediated drug-drug interactions which may lead to adverse effects. Nonetheless, the understanding of *h*OAT1's structure and function remains limited due to the absence of resolved structure of *h*OAT1. However, owing to conserved structural and functional patterns shared within Major Facilitator Superfamily (MFS) that OAT1 belongs to, an opportunity appeared for structure modelling by protein structure prediction tools and molecular dynamics simulations.

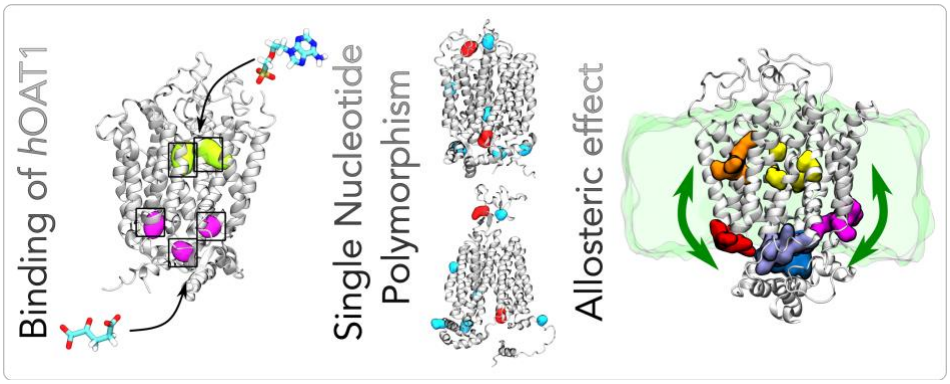
Taking advantage of formerly validated *h*OAT1 models, the present work investigated substrate and co-substrate binding to *h*OAT1, paying attention to allostery between key *h*OAT1 domains. Dynamic pictures were provided by microsecond-scaled molecular dynamic simulations of *h*OAT1 OF bounded to adefovir and/or α-ketoglutarate embedded into POPC-based membranes. Our computational approach has revealed key residues of two binding pockets for adefovir. α-Ketoglutarate might bind to the intracellular charge-relay system highly conserved motifs within MFS. Allostery between these binding pockets was also investigated highlighting the active role of the surrounding lipid bilayer. Lastly, using the structure of *h*OAT1 in OF as well as inward-facing conformation obtained from AlphaFold 2 (AF2) structure prediction tool, we proposed a structural rationalization of transport impairments experimentally observed for 2 significant single nucleotide polymorphisms, namely Arg50His and Arg454Gln.

Keywords: Membrane Transporters; Structural Pharmacology; Molecular Dynamics; Protein-lipid interactions; Major Facilitator Superfamily

Abbreviations

ABC	ATP-Binding Cassette
AF2	AlphaFold 2
AFV	Adefovir
***a*KG**	α-ketoglutarate
ANP	Acyclic Nucleoside Phosphonate
CDF	Cidofovir
Chol	Cholesterol
CKD	Chronic Kidney Disease
ECL	ExtraCellular Loop
H-bond	Hydrogen bond
IC	IntraCellular
IF	Inward-Facing
$\mathbf{K_M}$	Binding affinity
MD	Molecular Dynamics
MFS	Major Facilitator Superfamily
MTX	Methotrexate
NaDC3	Na^+/dicarboxylate transporter
OAT	Organic Anion Transporter
OF	Outward-Facing
OTA	Ochratoxin A
PAH	*p*-aminohippurate
PC	PhosphatidylCholine
PCA	Principal Component Analysis
PE	PhosphatidylEthanolamine
PK	PharmacoKinetics
PKTD	Proximal Kidney Tubular Dysfunction
PME	Particle Mesh Ewald
POPC	1-palmitoyl-2-oleoyl-sn-glycero-3-phosphocholine
POPE	1-palmitoyl-2-oleoyl-sn-glycero-3-phosphoethanolamine
PTC	Proximal Tubular Cells
RSST	Remote Sensing Signalling Theory
SLC	SoLute Carrier
SNP	Single Nucleotide Polymorphisms
TMH	TransMembrane Helix
TNF	Tenofovir

Graphical Abstract



Graphical abstract. The present work (from left): (*i*) reveals binding modes of adefovir (top) and α-ketoglutarate (bottom) to *h*OAT1; (*ii*) maps Single Nucleotide Polymorphisms on outward-facing (top) and inward-facing (bottom) conformation of *h*OAT1; (*iii*) asses the allosteric effect of lipidic environment and presence of substrates.

1. Introduction

Membrane transporters such as ATP-binding cassette (ABC) proteins and solute carriers (SLCs) are responsible for substrate translocation across cell membranes in a myriad of pharmacological and physiological events. At the cellular level, membrane transporters are involved in the selective influx and efflux of a broad range of compounds, including xenobiotics and endogenous substances. From the physiological perspective, the transepithelial transport of hormones [1], toxins [2], metabolites [3], and signalling molecules [4] at the interface between biofluids (e.g., blood/urine, blood/bile) plays an active role in body homeostasis as proposed by the "Remote Sensing and Signalling Theory" (RSST) which describe the transporter-mediated multi-organ regulatory system [4]. Membrane transporters are also involved in pharmacological events such as the drug absorption, distribution, metabolism, and elimination (ADME). Membrane transporters directly govern local pharmacokinetics (PK), *i.e.*, drug concentration at the target sites, whether linked with drug therapeutic or adverse effects [5,6]. Understanding the role of membrane transporters in systemic drug PK is thus essential since defects in their expression and/or function might alter drug exposure, disposition, and response [4,6]. The International Transporter Consortium has listed transporters of "emerging clinical importance" for which the functional evaluations are recommended for new drug development [6,7]. The human organic anion transporter 1 (*SLC22A6*/*h*OAT1) belongs to this list given its central role in drug elimination as well as in the RSST [6–8]. *h*OAT1 is localized at the basolateral membrane of kidney proximal tubular cells (PTCs), standing as a central mediator of renal elimination [9,10]. *h*OAT1 is involved in the translocation of a broad range of endogenous substrates such as metabolites and signalling molecules. *h*OAT1 is also responsible for the elimination process of xenobiotics including antiviral (*e.g.*, adefovir), anticancer and antituberculosis drugs [1].

*h*OAT1 is an antiporter, *i.e.*, the uptake of substrates from the blood into PTCs is coupled to the excretion of α-ketoglutarate (αKG) as a co-substrate into blood circulation [11]. This requires a high intracellular concentration of αKG in PTCs. That is maintained thanks to the so-called tertiary active transport model in which *h*OAT1 activity is coupled with Na^+/K^+-ATPase pump and NaDC3/*SLCA13A3* transporter [1,12]. Atomic-scaled mechanism of *h*OAT1-mediated transport remains unclear owing to the lack of experimentally resolved structure. Structural patterns of *h*OAT1 might provide insights into its roles regarding drug efficacy, toxicity, and elimination [13,14]. We recently proposed a structural and dynamic model of *h*OAT1 embedded in lipid bilayers [15]. *h*OAT1 is expected to adopt the Major Facilitator Superfamily (MFS) fold for which substrate translocation follows the alternating access model. It requires large-scale

conformational changes between at least two major conformational states, namely outward-facing (OF) or inward-facing (IF) conformations responsible for substrate binding and release events, respectively [16]. *h*OAT1 structure encompasses 12 transmembrane helices (TMH) organized into N- (TMH1-6) and C-bundles (TMH7-12). *h*OAT1 transmembrane domain exhibits pseudo symmetrical repeats [15] of functional helices, namely A-, B- and C-helices. Briefly, the inner cavity is made of A-helices (TMH1, 4, 7 and 10), which are expected to bind substrates and structurally adjust along the transport cycle. B-helices (TMH2, 5, 8, 11) stand at the interface of the bundles, contributing to the large conformational changes along transport cycle. Finally, the out-of-the-core C-helices (TMH3, 6, 9 and 12) were suggested to maintain the structural integrity of the transporter [17] through interactions with surrounding lipids. *h*OAT1 also exhibits a large extracellular loop (ECL) between TMH1 and TMH2 in which four glycosylation sites were described. ECL was suggested to be involved in trafficking; however, its glycosylation is non-essential for the transport function [18,19]. On the other side of the membrane, the intracellular region is involved in regulations of transport function and expression [19,20]. The intracellular region was suggested to consist in 6 intracellular helices which are tightly connected to TMH intracellular regions. It is featured by the presence of charged amino acids in conserved motifs within MFS proteins. The so-called A-, E[X_6]R and PETL motifs are repeated in both N- and C-bundles. The interactions between these motifs differ between IF and OF conformations [15]. Knowledge about binding modes of *h*OAT1 substrates remain fragmented while they are of particular importance for the understanding and prediction of transporter-mediated drug-drug interactions (DDIs) [21–23]. Likewise, investigating the structural impact of single nucleotide polymorphisms (SNPs) in *h*OAT1 may help to understand the mild transport impairments of *h*OAT1 observed experimentally, even though their clinical impacts may be rather limited owing to substrate overlap with other PTC influx transporters (e.g., *h*OAT3).

The present study aimed to model the binding modes of adefovir, an acyclic nucleoside phosphonate (ANP) antiviral, to *h*OAT1. Adefovir was chosen as a *h*OAT1 substrate prototype given the extensive experimental data available in the literature [1,24,25]. To better understand the interplay with co-substrate translocation, attention was paid to the binding of co-substrate αKG on the intracellular region as well. The allosteric communication between substrate and co-substrate through their binding sites was also studied, considering the contribution of surrounding lipids (i.e., phosphatidylcholine and phosphatidylethanolamine lipids as well as cholesterol) since it was previously described as being key for other MFS proteins [26–28].

2. Methods

2.1. Binding of (co-)substrates to *h*OAT1 model

From our previous μs-scaled MD simulations performed on the apo *h*OAT1 OF model, representative snapshots were extracted to investigate the binding modes of αKG and adefovir as co-substrate and substrate, respectively. Initial poses for (co-)substrates were obtained by means of molecular docking calculations using AutoDock Vina 1.1.2 [29]. αKG and adefovir were modelled as dianionic compounds given the physiological pH. αKG and adefovir initial structures were obtained from PubChem database [30] and optimized by quantum mechanical methods at the (CPCM)-M06-2X/6-31+G(d,p) level of theory using the Gaussian16 Rev. A package [31]. Molecular docking search volumes were defined with 66560 and 64768 $Å^3$ for the *h*OAT1 extracellular and intracellular sides respectively for adefovir and αKG given the antiporter transport (see Table S1 and Figure S1 for the definitions of box size and centre). For each substrate, 20 molecular docking calculations were carried out providing 20 molecular poses each, leading to 400 poses per substrate for each membrane. For each substrate, three initial binding poses were selected accounting for the calculated affinity scores as well as for the experimentally-identified key binding residues (see Figure S2) [32–34]. To confirm their relevance, molecular docking poses were then used as initial positions for μs-scaled MD simulations.

Three different systems were considered for further MD simulations, namely αKG-, adefovir- and αKG-adefovir-bound *h*OAT1. For each system, three binding modes were considered for further MD simulations (see Figure S2). In total, nine *h*OAT1 systems were used for MD simulations (see Figure S2). It is worth mentioning that two αKG molecules were systematically considered for αKG-based systems in order to optimize sampling. All systems were embedded into three types of 1-palmitoyl-2-oleoyl-sn-glycero-3-phosphocholine (POPC)-based membranes namely POPC, POPC and cholesterol (POPC:Chol (3:1)), POPC with cholesterol and 1-palmitoyl-2-oleoyl-sn-glycero-3-phosphoethanolamine (POPC:POPE:Chol (2:1:1)). In total, 27 different systems were considered in the present study. Systems were all solvated with water using a 0.154M NaCl concentration to mimic extracellular physiological conditions. System compositions and sizes are reported in supporting information (Table S2). It is worth mentioning that one position (namely, position 3 in Figure S2) was excluded since adefovir left the binding cavity in all simulations, except in POPC:POPE:Chol (2:1:1) membrane. However, in these simulations, adefovir mostly interacted with lipid components at the lipid-protein-water interface rather than with *h*OAT1 residues.

2.2. MD simulation setup

For the definitions of the protein, lipids and water, the following forcefields were applied Amber FF14SB [35], Lipids17 [36] and TIP3P [37], respectively. The parameters for counterions (Na^+, Cl^-) were obtained from Joung and Cheatham [38,39]. The definition of the co-substrate and substrate required a parametrization using GAFF2 and DNA.OL15 forcefield's parameters, considering the latter being a purine derivative. MD simulations were performed using a similar setup as previously established [15]. The simulations were carried out with using the CPU and GPU codes of the Amber18 package [40], [41]. The system was treated with periodic boundary conditions. Non-covalent interactions were considered within the cut-off of 10 Å for electrostatic and Lennard-Jones potentials. Long-range electrostatic interactions were treated using the particle mesh Ewald (PME) method [42]. The bonds involving hydrogen atoms were fixed by applying the SHAKE algorithm. Integration time step was set at 2 fs. To maintain the physiological conditions, the temperature was set at 310K and carried on using a Langevin thermostat [43]. Whereas the Monte Carlo barostat was applied to maintain the constant pressure boundary condition under semi-isotropic conditions [44].

The equilibration of the system was pursued by minimizing all atomic positions and followed by a smooth two-step thermalization. The system was heated up from 0 to 100 K during 200 ps under (N, V, T) conditions, while the second step of the thermalization up to 310K was carried out under semi-isotropic (N, P, T) conditions. Subsequently, equilibration simulations were performed for 5.5 ns. Finally, the MD simulations of substrate-bound *h*OAT1 resulted in a duration of 1.5 μs each, leading to a total of *ca.* 40.5 μs. Trajectory snapshots were saved every 10 ps.

2.3. Analysis

Given the previous validation of the model [15], structural analyses were based on our recent findings, as well as on the general knowledge of MFS patterns [16]**.** Structural analyses were performed using the PyTRAJ and CPPTRAJ AMBER modules [45] as well as the VMD software [46]. Based on the time-dependent backbone root-mean squared deviations (Figure S3), analyses were performed on the equilibrated section of the present trajectories, i.e., over the last 800 ns (Figure S3). H-bond analyses were performed using distance and angle cutoffs set at 3.0 Å and 135°. The minimum fraction threshold was set at 0.1 given the known uncertainties for side chain rotameric states in protein threading techniques. MD trajectories obtained from our previous study was also used for sake of comparison between apo and bound *h*OAT1 systems.

To unravel the allosteric communications between key regions of *h*OAT1, we carried out network analyses using Allopath tool [47]. The communication efficiency between two distant domains was calculated from the protein contact map and the mutual information matrix in which the contributions of interactors (*e.g.*, substrates and lipids) were included. Communication efficiencies were calculated unidirectionally from “source” to “sink” residues (see Ref. [47] for more details). In the present work, allosteric communications between binding pocket residues (B-like motif and the inner binding cavity, see Section 3.3) and the charge-relay system (A- and E[X_6]R motifs, see Table S3 and Section 3.3) were calculated. The contributions of interactors were assessed by calculating communication efficiencies on systems made of: (i) *h*OAT1 *apo*, (ii) *h*OAT1 substrate-bound, (iiI) *h*OAT1 *apo* with lipids, and (iv) *h*OAT1 substrate-bound with lipids.

3. Results and Discussion

3.1. Interactions of the *in silico h*OAT1 model with (co-)substrates

3.1.1. Substrate interactions with *h*OAT1 cavities

*h*OAT1 was shown to play a key role in the uptake of anionic endogenous and exogenous substrates, such as antiviral acyclic nucleoside phosphonates (ANP) (*e.g.*, tenofovir, adefovir) [34,48], urate, *p*-aminohippurate (PAH), β-lactam antibiotics and sulfate conjugates [49]. The presence of numerous cationic residues (i.e., lysine, arginine, and histidine, see Figure 1A) at the *h*OAT1 extracellular interface in OF and IF *h*OAT1 models are consistent with the expected high affinity for anionic substrates, by favouring electrostatic interactions. Cationic residues were found in the ECL between TMH1 and TMH2 as well as in the water-exposed cavity in the OF *h*OAT1 model. Molecular docking calculations suggested three possible binding positions for adefovir as well as for αKG in the intracellular regions (Figure S2). MD simulations confirmed the importance of A-helices in the water-exposed cavity, mainly TMH1 and TMH4. The latter was previously reported as key for substrate binding and translocation owing to the presence of the conserved motif RXX[Q/S]G [50]. The so-called B-like motif was observed in several MFS antiporters, even though it cannot be considered as an antiporter fingerprint [50]. MD simulations reveal that adefovir may strongly bind to the B-like motif Arg192 residue thanks to a salt-bridge between phosphonate and guanidinium moieties as pictured by calculated large H-bond fraction at 1.950 over the MD simulation (see Figure 1B and Table S4).

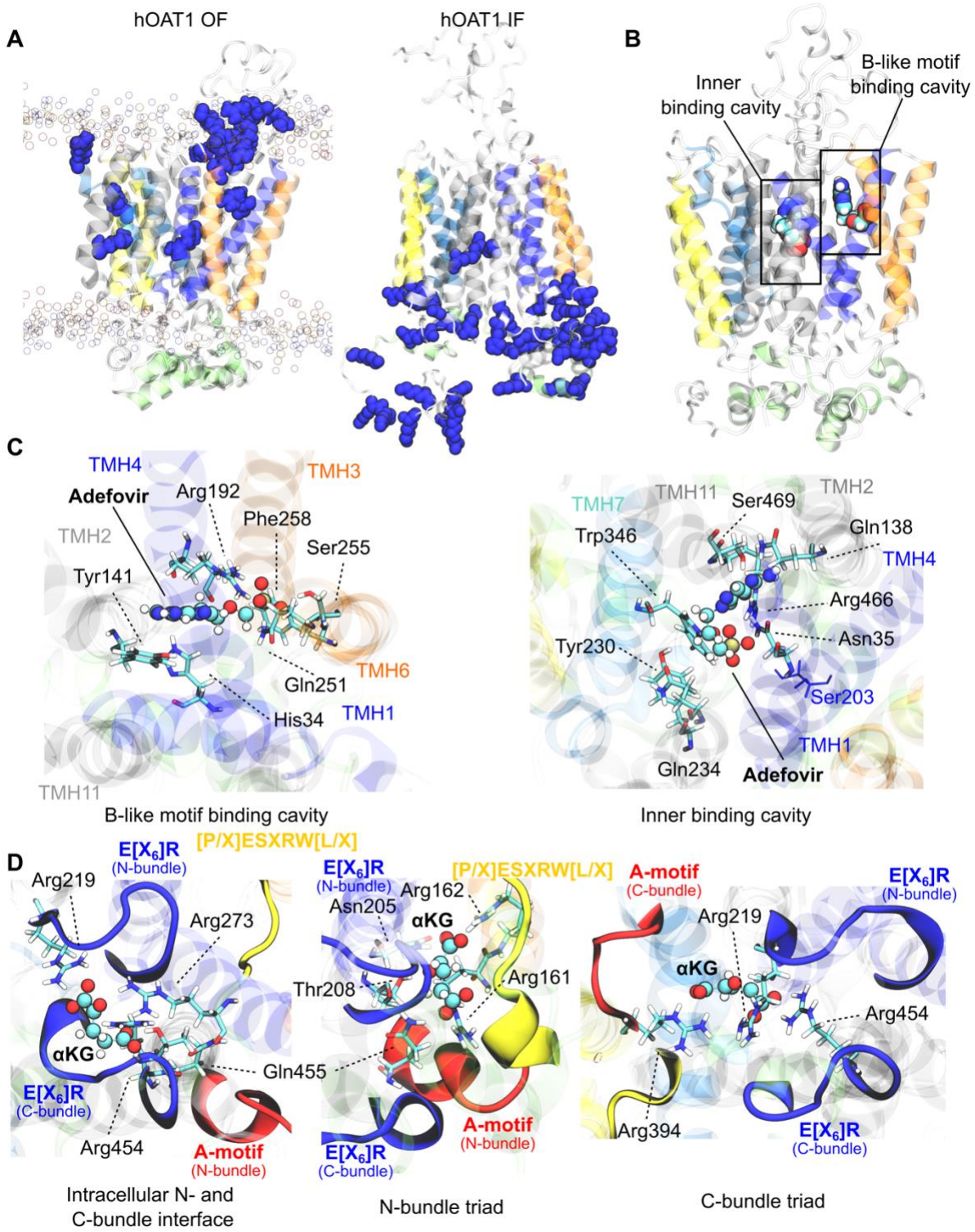


Figure 1. Interactions with (co-)substrate. A) Representation of cationic residues present in extracellular and intracellular sites of the OF (left) and IF (right) models, respectively. B) The water-exposed cavity exhibits 2 binding pockets for adefovir: the B-like motif and the inner binding cavity involving TMH1 and TMH4 (dark blue). C) Residues involved in adefovir binding in the B-like motif (left) and inner (right) binding cavity. D) Three mains αKG binding spots and the most frequently interacting residues. A-, B- and C-helices are coloured blue-, grey- and yellow-ish, respectively.

H-bond analyses also highlighted the key role of several polar and aromatic residues, namely Ser255, Gln251, Ser195 and Tyr141. π-Stacking interactions with Tyr141 were also observed between the adefovir adenosyl moiety and tyrosine phenol moiety contributing to the stabilization of the substrate. This is in line with the *h*OAT1 substrate spectrum which includes various anionic aromatic compounds (*e.g.*, ANPs, PAH, urate). It is worth mentioning that the *h*OAT1 OF model suggested two possible accesses to the B-like motif cavity. On one hand, substrates may enter directly from the water phase through the ECL cationic residue network. On the other hand, the OF *h*OAT1 model suggests that there may exist an access channel between TMH3 and TMH6 connecting the high-density polar head region of the outer leaflet membrane to the B-like motif cavity. This is in line with biophysical studies showing that amphiphilic substrates might partition below polar head region of the lipid bilayer membrane [51].

Molecular docking calculations also suggested the existence of an inner binding cavity at the interface between N- and C-bundles, involving mostly TMH1 and TMH7. The substrate is again stabilized thanks to a strong H-bond network (Figure 1D). Cationic Arg466 is likely to play a central role for anionic substrate translocation (Table S4 and Figure 1C). This is in good agreement with experimental observations in which site-directed mutagenesis of Arg466 was associated to substrate-dependent loss of transport capacity (Table S5) [52]. Likewise, several residues of the site-directed mutagenesis which were associated with the loss of transport function are present in either the first- or the second shell around adefovir (*e.g.*, Asn39 [18], Tyr230 [32], Tyr353 [53], Trp346 [53], see Table S5). This confirms the relevance of the region obtained from MD-refined molecular docking calculations. Particular attention was also paid to Ser203 (Figure 1C) which was described as central for the binding of several ANPs. In our simulations, Ser203 belongs to the second-shell of contact residues with adefovir. Therefore, no direct interactions between adefovir and Ser203 were observed in our calculations. However, it is worth mentioning that binding to Ser203 was suggested from static structural investigations performed using an IF *h*OAT1 homology model [34]. Therefore, in line with experimental investigations, we can hypothesize that Ser203 is likely involved in the substrate translocation by carrying substrate from the outer to the inner leaflet as part of the OF-to-IF large-scale conformational transition. Furthermore, this is strengthened by the recent AF2 IF model [54] in which Ser203 is deeply located in the MFS core, precluding direct substrate binding in the OF conformation.

MD simulations suggest that the inner binding cavity is slightly less accessible than the B-like motif pocket, likely due to the absence of an access channel. Interestingly, the two cavities are contiguous, around TMH1. Therefore, present results as well as experimental observations

regarding mutations of residues located in the inner binding cavity [32,33,52,53] suggest that substrate binding might occurs first in the B-like motif cavity. Substrate binding to B-like motif pocket is thus expected to rapidly lead to local conformational changes which may open an access channel to the inner binding cavity. Binding to inner cavity is in turn expected to be then pivotal for substrate translocation events along the transport cycle. Finally, rocker-switch large-scale TMH conformational changes occurring during the translocation event might then decrease substrate binding affinity, favouring substrate release in the intracellular medium.

3.1.2. Investigations on α-ketoglutarate binding at the *h*OAT1 intracellular interface

*h*OAT1 transport cycle requires large conformational changes for the OF-to- IF transition. *h*OAT1 being an antiporter, substrate uptake is coupled with the co-transport of αKG in the opposite direction [11]. However, the sequence of events (*i.e.*, αKG efflux and substrate influx) remains unclear as well as triggers. However, our present model can be used to assess binding modes of αKG in the intracellular region of *h*OAT1. As stated for adefovir binding at the extracellular interface, intracellular domains are also rich in cationic residues favouring electrostatic interactions with anionic αKG molecules. These cationic residues are exposed to the intracellular medium independently on the conformational state (Figure 1A and 1B). Even though αKG translocation is expected to reset the OF conformation, our simulations suggest that αKG can also bind intracellular domains, regardless of the *h*OAT1 conformational state.

MD-refined molecular docking calculations also exhibited three preferential binding modes for αKG: (i) on the N- and (ii) C-bundle motif triads; or (iii) at the interface between N- and C-bundle E[X_6]R motifs (Figure 1D). Owing to its two anionic carboxylate moieties, αKG favours electrostatic interactions with the charge-relay system cationic residues, as pictured by described H-bond network from MD simulations (Figure 1D, Tables S4 and S6). For instance, strong salt-bridges were observed along the simulations between αKG and A-motif arginine residues: Arg162 and Arg161, or Arg394 for binding poses in the N- or C-bundle triad, respectively (H-bond fractions=2.019, 1.698 and 1.376 for Arg162, Arg161 and Arg394, respectively). Likewise, the intracellular charge-relay system may play a central role in αKG binding at the interface between N- and C-bundle E[X_6]R motifs (*e.g.*, Arg219, Arg454). In turn, MD simulations with αKG bound system to the intracellular domain also reveal that the charge-relay system might be disrupted by the presence of αKG (Table S6). Assuming that αKG can bind *h*OAT1 in the OF conformation, present results suggest that αKG may play favour substrate translocation by destabilizing intracellular N- and C-bundle interactions, which is necessary for OF-to-IF transition along with

substrate translocation. These findings pave the way for further investigations to establish whether the IF-to-OF transition resetting *h*OAT1 conformation is driven by αKG efflux or if αKG and the other substrate are simultaneously transported; even though the latter is expected to be less likely [16].

3.2. Allosteric communication between the substrate binding pockets and intracellular domains

The comparison of the intracellular structural arrangement between OF and IF apo *h*OAT1 models revealed that the charge-relay system must be disrupted along transport cycle. This might ultimately unlock the IC gating required for substrate release [15]. Conformational changes are expected to be triggered upon substrate and/or co-substrate binding, suggesting the existence of a distant communication between substrate binding pockets and MFS conserved intracellular motifs. Allosteric communications were thus monitored providing structural insights about the efficiency regarding the different domains located across the membrane (see Section 2.3 and Ref. [47] for technical details). Particular attention was paid to the plausible communication between binding domains (B-like motif and inner substrate binding site) with the charge-relay system (A- and E[X_6]R motifs). It is worth mentioning that (i) N- and C-bundle IC motifs were considered separately and (ii) allostery was monitored in both directions, *i.e.*, from EC to IC regions and *vice versa* leading to 16 different pathways (Figures 2A). The role of substrate, co-substrate and surrounding lipid bilayer were also taken into account. However, results provided here are only qualitatively discussed since the resolution of the present model, especially regarding side chains, precludes quantitative conclusions (Figure 2 and S4-S11).

As an example, Figure 2B shows efficiencies of selected allosteric pathways. MD simulations revealed the existence of a reciprocal distant communication between intracellular motifs and substrate binding pocket. While considering only the role of *h*OAT1 protein, allosteric pathway analyses revealed an efficient communication from A-motifs to substrate binding pockets, while other pathways remain inherently low. Furthermore, these allosteric pathway efficiencies exhibit very similar profiles for both B-like motif and inner binding pockets. Interestingly, efficiencies calculated for allosteric pathways from A-motifs to substrate binding pockets were not affected by the presence of substrates. In contrast, the presence of substrate and/or co-substrate strongly potentiated the efficiency of the allosteric communications from binding pockets to intracellular motifs. In other words, substrate binding events are thus expected to modulate the dynamics of intracellular motifs. Such observations are in line with the proposed mechanism of *h*OAT1 transport for which binding events might trigger the conformational changes in the IC region

required for substrate translocation. The contributions of *h*OAT1 residues to allosteric networks were also calculated underlining the pivotal roles of TMH2, TMH3 and TMH4 which are involved either in binding pockets or in A-motifs.

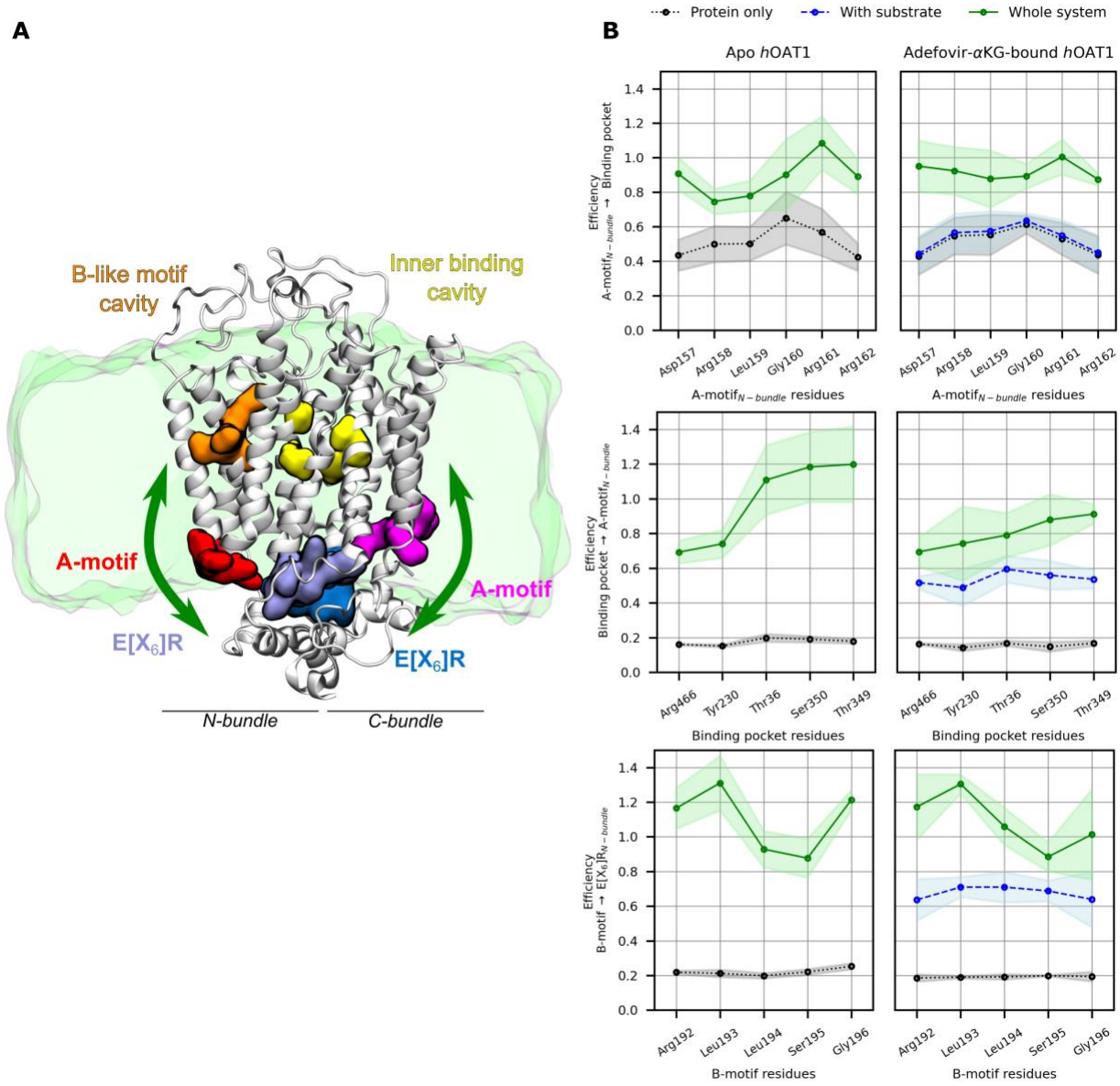


Figure 2. The allosteric effect of the membrane lipid bilayer in the presence and absence of substrates on the *h*OAT1 communication efficiency from biding pockets toward charge-relay system and reverse. (A) Visualisation of binding pockets (B-like motif cavity on left and inner binding cavity on right) and motifs of charge-relay system divided into N and C domain. The wide green arrows represent strong communication in presence of lipids. (B) The schemes plot the communication efficiency calculated for pure protein (black dashed line), protein in presence of substrates (blue dashed line) and protein accounting lipids in apo (left column) or substrate-bound state (right column) in green line.

Besides, it is important to note that MFS protein functions have been experimentally shown to strongly depend on the membrane composition [26,27,55]. Likewise, PE lipids were shown to non-covalently bind with the residues standing at the interface between N- and C bundles in apo *h*OAT1 structure [15]. Therefore, out of the structural interplay between surrounding lipid bilayer and *h*OAT1 protein, the active role of lipid bilayer membrane in the communication between extracellular and intracellular regions was also investigated. Efficiencies drastically increased while considering the membrane. This confirms that the membrane plays an active part in allosteric signalling as recently shown for other membrane proteins [47], including transporters [56]**.** This also underlines the central role of protein-lipid interactions and the need for representative and realistic membranes, including PE and cholesterol, for computational and experimental investigations on *h*OAT1. At the body level, *in situ* alteration or modulation of cell membranes may affect the activity of *h*OAT1 either by modifying the global structure and dynamics of *h*OAT1, or its intrinsic function [57,58]. Such observations may be extended to other MFS transporters given the high protein-lipid dependency.

3.3. Structural mapping of *h*OAT1 single nucleotide polymorphisms

Beyond pharmacologically relevant mechanistic insights into *h*OAT1-mediated drug transport, the present structural model substrate can also be used to understand the modulation of substrate transport across cell membranes by naturally-occurring polymorphisms in genes coding transporters [59–62]. Several SNPs in *h*OAT1 were identified and classified according to ethnical origins and species [34]. Since most SNPs are located in untranslated intronic regions, the focus will be given here to non-silent mutations (Table 1 and Figure 3), which correspond to a limited number of SNPs reported in the literature. Moreover, most of them exhibit no or limited alteration of transport function. The present IF and OF *h*OAT1 models can help understand the structural changes induced by genetic polymorphisms. Particular attention will be paid to those which are not associated with either protein expression or altered membrane trafficking. Indeed, protein structure prediction tools should be carefully considered when variants exhibit significant folding differences [63].

Table 1. List of *h*OAT1 SNPs and rare variants. The reported impairment of *h*OAT1 function is shown in bold.

Nucleotide change	Variant	Position on OAT1 gene sequence[a]	Amino acid change	*h*OAT1 location	Effects	References
T>C	rs1415632329	20 (Exon 1)	p.Leu7Pro	N-terminal region	No data	[64–66]
G>A	**rs11568626**	**149 (Exon 1)**	**p.Arg50His**	**ECL1**	**No change in transport of PAH, OTA, MTX; Decreased transport affinity for AFV, CDF and TNF**	[59,62,64,65, 67]
C>T	rs11568627	311 (Exon 1)	p.Pro104Leu	ECL1	No change in transport of PAH, OTA, MTX	[62,65]
T>C	rs11568623	677 (Exon 4)	p.Ile226Thr	TMH5	No change in transport of PAH, OTA, MTX	[62,65]
C>T	rs11568624	767 (Exon 4)	p.Ala256Val	TMH6	No change in transport of PAH, OTA, MTX	[62,65]
C>T	rs45607933	877 (Exon 5)	p.Arg293Trp	Intracellular region	No change in transport of PAH, OTA, MTX	[62,65]
G>A	**rs11568634**	**1361 (Exon 8)**	**p.Arg454Gln**	**C-bundle E[X_6]R**	**Loss of transport activity of PAH, OTA and MTX; not associated with PKTD induced by AFV treatment for chronic hepatitis B**	**[24,62,65]**
A>T	nd	1575 (Exon 10)	p.Lys525Ile	C-terminal region	No change in transport of PAH, AFV, CDF, TNF	[59,65]

[a] Gene sequence position are related to the ATG start site.

Abbreviations: AFV, adefovir disproxyl fumarate; CDF, cidofovir; MTX, methotrexate; OTA ochratoxin A; PAH, p+-aminohippurate; PKTD, proximal kidney tubular dysfunction; TNF, tenofovir, not determined.

Eight SNPs have been considered and mapped on the present IF and OF *h*OAT1 models (Figure 3). They can be classified according to their location on the model, as follows: (i) in the N-terminal domain (p.Leu7Pro); (ii) in the long ECL between TMH1 and TMH2 (p.Arg50His and p.Pro104Leu); (iii) in TMH5 and TMH6 (p.Ile226Thr and p.Ala256Val); and (iv) in the intracellular domain (p.Arg293Trp, p.Arg454Gln and p.Leu525Ile). SNPs leading to p.Pro104Leu, p.Ile226Thr, p.Ala256Val, p.Arg293Trp and p.Lys525Ile substitutions were not associated with alteration of substrate intake [24,59,62,64,67]. Only two SNPs were reported to impair *h*OAT1 function, namely rs11568626 and rs11568634 leading to p.Arg50His and p.Arg454Gln protein mutations, respectively. The rs11568626 SNP was found to be specific to the African population. The resulting p.Arg50His mutation was shown to be associated with decreased transport affinities

(K_m) of phosphate analogues such as adefovir, cidofovir and tenofovir in Xenopus oocytes-expressed p.Arg50His variant as compare to wild-type *h*OAT1 (rs15914676). However, Xenopus oocytes-expressing p.Arg50His variant exhibited normal uptake of PAH, ochratoxin A and methotrexate [62].

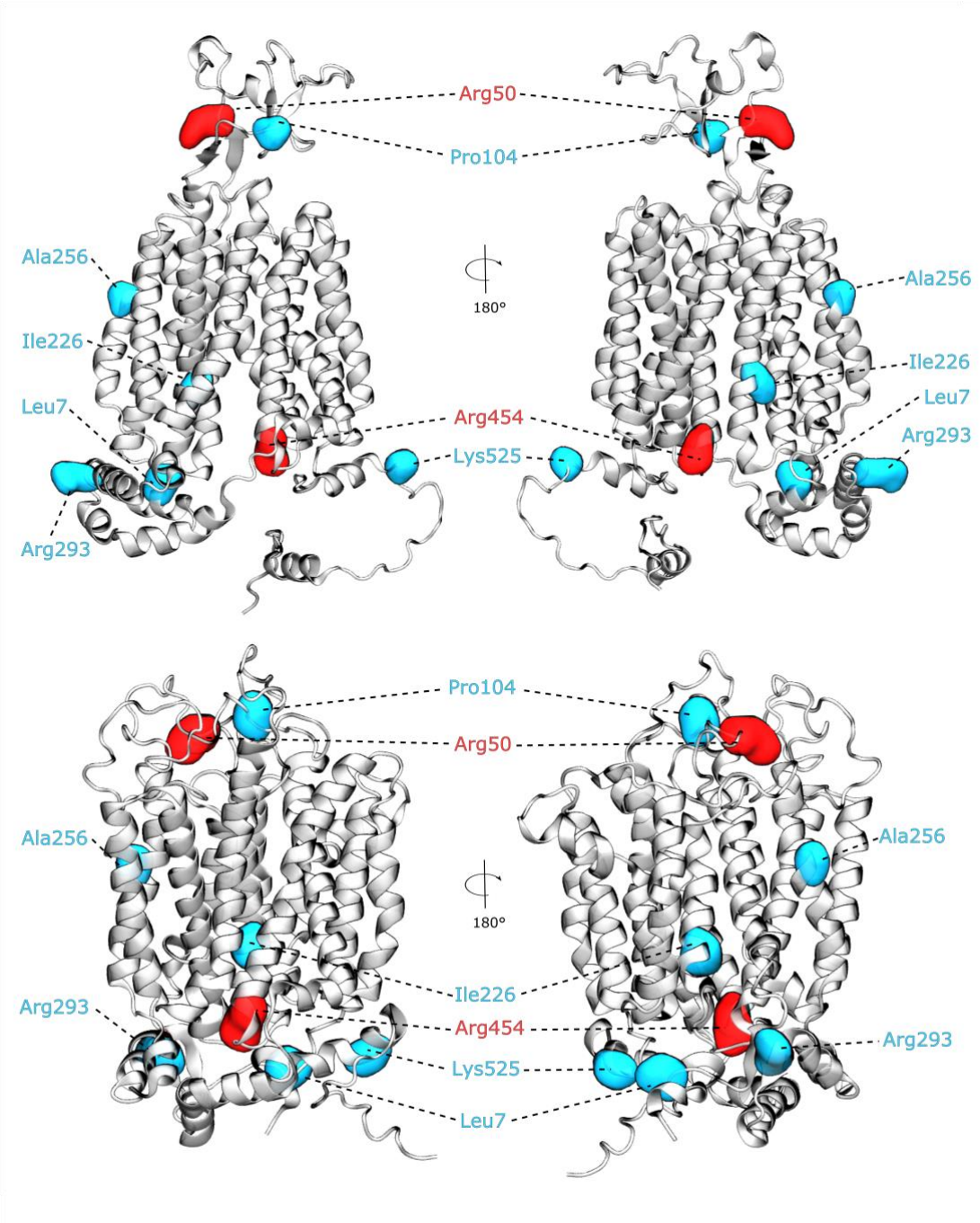


Figure 3. Mapped Single Nucleotide Polymorphisms on IF (top) and OF (bottom) conformations of *h*OAT1. Substitutions of Arg50 and Arg454 were shown to impact drug transport while no impact was reported for substitutions of Pro104, Ala256, Arg293, Ile226 and Leu7.

Interestingly, Arg50 is located in a conserved motif on the aforementioned ECL1 pattern, rich in cationic residues and likely involved in anionic substrate access to the B-like motif binding pocket. Therefore, we can hypothesize that the p.Arg50His substitution may lead to a lower electrostatic potential between the anionic substrate and the binding access channel given the poor ECL resolution of the present OF model, this must be confirmed by further experiments or through the experimental structure resolution of *h*OAT1. The rs11568634 SNP leads to the substitution of Arg454 by a glutamine residue. Interestingly, this SNP was associated to decreased uptake of PAH, ochratoxin A and methotrexate, suggesting a loss of function [62]. This is in perfect agreement with present structural observations which underlined the central role of Arg454 in the intracellular charge-relay system: Arg454 is likely involved in salt-bridges between the C-bundle A-motif and the PETL motif. The p.Arg454Gln substitution might weaken the local supramolecular arrangement, which in turn is likely to disrupt the charge-relay system essential for the MFS transport cycle.

It is worth mentioning that no *h*OAT1 SNP was associated with pathological conditions at the clinical level. Owing to the central role of OATs in RSST [4], the substrate overlap between *e.g.,* *h*OAT1 and *h*OAT3 [11] might lead to compensation activity between transporters. However, systemic compensation between transporters might not be sufficient to overcome *h*OAT1 impairment in chronic conditions such as chronic kidney diseases. Recently, particular attention has been paid to the loss of tubular function in CKD [68], where *h*OAT1 impairment is expected to play a key role [69]. Further clinical investigations are required to examine *h*OAT1 SNPs as risk factors for the still unclear patient variability in *e.g.*, long-term ANP nephrotoxicity [25,59,62,70].

4. Conclusion

The present study investigated substrate binding events of both substrate and co-substrate to the human *SLC22A6*/OAT1 at the atomic scale. Molecular docking calculations and μs-scaled MD simulations revealed two plausible binding spots for adefovir, consistent with experimental observations, in the B-like motif and inner binding pockets. A-helices (essentially TMH1, TMH4) residues were shown to play an essential role for substrate binding event. In the intracellular region, MD simulations also suggested the binding of αKG involved residues of the charge-relay system, located either within the motif triads (within A-motif, E[X_6]R and PETL) or at the interface between them. The presence of αKG may interfere with the salt-bridge network of intracellular conserved motifs. This is expected to favour the opening of IC gating which might be key in driving

large-scale conformational changes required for *h*OAT1 alternated access. Besides, the allosteric pathways between substrate cavities and intracellular motifs were revealed on *h*OAT1 dynamic pictures obtained using MD simulations and highlight the central role of *e.g.*, TMH2, 3 and 4. Likewise, our simulations showed that the surrounding lipids play an active role in *h*OAT1 transport function. Non-synonymous SNPs were then mapped onto the *h*OAT1 structural model. Transport impairments experimentally observed for p.Arg50His and p.Arg454Gln were respectively attributed to (i) the decrease of substrate binding affinity or (ii) disrupted intracellular domain interactions owing to the weakening of electrostatic interactions. Each new SNP or site-direct mutagenesis mutant can now be implemented in the present model to elucidate its role at an atomistic resolution.

Altogether, the present study paves the way for the structural understanding of *h*OAT1 function in a pharmacological context. The present model can be used to understand and predict transporter-xenobiotic interactions as well as DDIs involving this membrane transporter. The allosteric pathways described may be involved in non-competitive DDIs, by which a *h*OAT1 inhibitor may interact with an allosteric site involved in the communication between key regions.

5. Declaration of interest

None.

6. CRediT (Contributor Role Taxonomy) author contributions

Angelika Janaszkiewicz: Conceptualization, methodology, formal analysis, investigation, writing (original draft, reviewing & editing). **Ágota Tóth:** Methodology, validation, formal analysis, investigation, writing (reviewing & editing). **Quentin Faucher:** Conceptualization, formal analysis, investigation, writing (reviewing & editing). **Hélène Arnion:** formal analysis. **Nicolas Védrenne:** formal analysis, writing (reviewing & editing). **Chantal Barin-Le Guellec:** Conceptualization, supervision, writing (reviewing & editing). **Pierre Marquet:** Conceptualization, supervision, funding acquisition, writing (reviewing & editing). **Florent Di Meo:** Conceptualization, methodology, validation, formal analysis, investigation, writing (original draft, reviewing & editing), supervision, funding acquisition.

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9. Supplementary Information

In Supplementary information are reported: (i) molecular docking and MD technical details; (ii) calculated H-bond and contact fractions; (iii) list of site-directed mutagenesis reviewed from the literature; (iv) Allosteric communication efficiencies for all systems. Data are available upon reasonable request.

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