Trait-specific patterns of community ecospace occupancy in an insular mammal radiation

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Abstract

Communities that vary in the richness of closely related species but occupy similar environments provide a powerful system to investigate how trait variation and species richness interact to fill ecological space. In tropical mountain habitats on the island of Sulawesi, Indonesia, local murine rodent community richness ranges from 7 to 23 species. We measured 17 morphological, ecological, and isotopic traits to test for the expansion or packing of community ecospace among nine murine communities. Traits related to cranial shape (but not jaw shape), trophic level, and body shape showed the strongest evidence of expansion. Locomotion emerged as a fundamental axis of ecological variation, expanding ecospace and enabling niche packing of other traits such as diet and body size. Though trait divergence often explains functional diversity in island communities, we found that phylogenetic diversity facilitates community ecospace expansion in some conserved traits such as cranial shape, while more labile traits are overdispersed both within and between island clades. Our results evoke competition, trait evolvability, and the independent evolutionary trajectories of each of Sulawesi's murine clades as central to generating the exceptional functional diversity and species richness in this insular radiation.

Komunitas-komunitas dengan variasi kekayaan spesies berkerabatan dekat tetapi menempati lingkungan yang serupa menyediakan sistem yang kuat untuk menyelidiki bagaimana variasi sifat dan kekayaan spesies berinteraksi untuk mengisi ruang ekologi. Kami mengukur 17 karakter morfologi, ekologi, dan sifat isotop untuk menguji perluasan atau pengemasan volume ruang ekologi komunitas di antara sembilan komunitas hewan pengerat yang terdiri dari 7 hingga 23 spesies di pulau Sulawesi. Sifat-sifat yang berhubungan dengan bentuk tengkorak (tetapi tidak bentuk rahang), tingkat trofik, dan bentuk tubuh menunjukkan bukti ekspansi yang kuat. Sistem penggerak muncul sebagai sumbu fundamental dari variasi ekologi, mengembangkan ruang ekologi, dan memungkinkan pengemasan relung pada sifat lain seperti makanan dan ukuran tubuh. Meskipun divergensi sifat dianggap melampaui konservatisme ceruk di komunitas pulau, kami menemukan bahwa keragaman filogenetik memfasilitasi ekspansi ruang ekologi komunitas dalam beberapa sifat yang dilestarikan seperti bentuk tengkorak, sementara sifat yang lebih labil tersebar secara berlebihan baik di dalam maupun di antara klad pulau. Hasil kami menunjukkan persaingan, sifat evolvabilitas, dan lintasan evolusi independen dari setiap klad murinae Sulawesi sebagai pusat untuk menghasilkan keragaman fungsional yang luar biasa dan kekayaan spesies dalam radiasi insular ini.

Introduction

Local species richness is often positively correlated with the complexity of habitat structure and

the diversity of available resources. Different localities with similar habitat structure, resource

availability, and historical access often contain ecologically similar communities. These

observations precipitated the study of species assembly (Hutchinson 1957, MacArthur and May

1972, Brown and Lieberman 1973, Brown 1975) and gave rise to the prediction that species only

co-occur if they partition niche space along some axis, otherwise one will be excluded through

competition (May and MacArthur 1972, Pianka 1974, M'Closkey 1978). In areas with high

species richness, individual species may use a narrower breadth of resources than in lower

species richness communities, thereby limiting niche overlap with co-occurring species, and

producing a more densely packed community niche space ("Niche Packing", Figure 1,

MacArthur 1970). Among animal communities, this prediction has found support with a variety

of phenotypic and behavioral proxies for resource use, such as body size and proportions, wing

shape, skull shape, time of activity, and microhabitat use (Brown 1975, Karr and James 1975,

M'Closkey 1978, MacArthur and MacArthur 1961, Pianka 1974, Pigot et al. 2016, Van de Perre

et al. 2020). However, individual species in the richest communities may exploit habitats or

resources that are unused or non-existent in low-resource, species-poor communities. In this

case, estimates of community niche space should be positively correlated with species richness

("Niche Expansion", Figure 1; MacArthur 1965, 1970, Pigot et al. 2016, Oliveira et al. 2020).

This mode of resource partitioning has also found support, including in isolated communities of

many closely related species (Lister 1976). Importantly, individual traits can reflect distinct

processes (Pigot et al. 2016), with packing on one axis (e.g., body size) and expansion on another

(e.g., diet, see Spasojevic and Suding, 2012).

The foundational work describing competition's role in community assembly relied on

empirical data from continental communities with equal biogeographic accessibility but variation

in primary productivity or habitat complexity (Brown and Lieberman 1973, Brown 1975,

M'Closkey 1978, Karr and James 1975, Pianka 1974, Weiher and Keddy 1995), or from

archipelagos where island size or geographic complexity determines resource availability and

habitat area, both of which influence overall species richness (Wilson 1961, Diamond 1975,

Lister 1976, Gillespie 2004, Losos 2009, Losos and Ricklefs 2009). Less explored are insular

areas of similar habitat structure and resource availability, but with discrete communities that

vary widely in species richness ("species richness anomalies", Swenson et al. 2016 pg. E83).

Such areas often house many closely related species and provide an opportunity to investigate

the role of competition in the evolution of functional diversity and community structure.

Insular settings such as oceanic islands often contain high levels of endemism. Lineages

that arrive and diversify in isolated locations regularly undergo ecological shifts that result in

behaviors or phenotypes that are uncommon or non-existent in source communities (Pinto et al.

2008, Esselstyn et al. 2012, 2015, Stroud and Losos 2016). However, the degree to which closely

related species in isolated locations share similar life history traits may vary (Losos et al. 2003,

Hiller et al. 2019, Dorey et al. 2020, Stroud 2021). The strength of phylogenetic niche

conservatism, or the tendency for similar niche requirements or trait values to be clustered within

clades, mediates the impact of phylogenetic diversity on community niche space. If traits are

conserved within clades, then increasing phylogenetic diversity inevitably expands community

niche space. However, high trait lability means that community phylogenetic diversity can be

decoupled from functional diversity. Multiple lineages evolving in sympatry in isolated settings

may reduce the degree of niche conservatism and promote niche divergence (Losos et al. 2003),

thereby limiting the impact of phylogenetic diversity on community niche breadth. Testing the

effects of trait conservatism and lability on community ecological space in insular settings

requires replicated communities assembled from the same clades, but that vary in species

richness.

Figure 1: Conceptual diagram of expected patterns from community niche expansion (upper) and packing (lower).
Expansion results in a larger ecospace area with the same density of points as the low-richness community, while
packing results in a higher ecospace density in the same ecospace area.
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We investigate the patterns of niche occupancy among communities of murine rodents

(Murinae: Muridae: Rodentia) from the oceanic island of Sulawesi, Indonesia, a mountainous,

tropical island (Figure 2) at the center of the Wallacean biodiversity hotspot. The murine

diversity of Sulawesi is exceptional, with 41 endemic species stemming from seven colonist

ancestors that arrived from circa 6 Ma to < 1 Ma (Figure S1, Rowe et al. 2019, Handika et al.

2021, Nations et al. 2021). While some colonists spawned small radiations of species and others

are evidenced by only a single living species (Table 1, Figure S1), each clade is widespread on

Sulawesi and most contribute to species richness of all local murine communities on the island.

These local communities, defined as the murine species living on a mountain from lowlands to

peak, range from 7 to 23 species, with the upper limit being perhaps the most diverse local

community of closely related terrestrial small mammals on Earth. Importantly, variation in local

community richness does not appear to be determined by environmental or habitat differences,

suggesting that environmental filtering, a central process of community assembly (Webb et al.

2002, Cavender-Bares et al. 2004), does not affect community functional diversity. Maximum

elevation, which may affect the area occupied by different habitats, appears to play only a minor

role (Figure 2A), and net primary productivity is nearly constant across the island's montane

regions (Imhoff et al. 2004). Instead, it appears that biogeography is the strongest predictor of

local diversity, as richness for many taxa is highest in the central part of the island and generally

lower on the peninsulas (Iskandar & Nio 1996, Musser 2014, Esselstyn et al. 2021). Such a

disparity in species richness between geographically and ecologically similar mountains provides

a natural experiment for testing alternative hypotheses of how communities fill ecological space.

Functional traits, or properties that quantify an organism's performance, microhabitat

use, or dietary preferences, are often evaluated as niche axes in vertebrate communities

(M'Closky 1978, Ricklefs and Travis 1980, McGill et al. 2006, Pianka et al. 2017). Gathering

direct functional data can be difficult in communities of motile, nocturnal, or secretive species.

In these cases, functional morphology provides a window into the ecological niche (Ricklefs and

Travis 1980, Pigot et al. 2016, 2020). Differences in head shape (Lister 1976, Maestri et al. 2016,

Kohli and Rowe 2019, Van de Perre 2020), body size (Woodward and Hildrew 2002), and body

proportions (Karr and James 1975, Ricklefs and Travis 1980, Nations et al. 2019, Nations et al.

2021) indicate differences in dietary and locomotor niches. Other indirect measures of resource

use, such as trophic dimensions revealed through stable isotope ratios and estimates of locomotor

mode, offer a quantitative assessment of diet and microhabitat use (Baltensperger et al. 2015,

Galetti et al. 2016). The murine fauna of Sulawesi is an intriguing system for sampling

functional disparity, as it includes some of the most unusual rodent forms (Figure 2B, Esselstyn

et al. 2012, 2015, Rowe et al. 2014) living in sympatry with more typical "rat-like" morphologies

and ecologies. Sulawesi murines have an array of cranial shapes that likely reflect their dietary

preferences (Esselstyn et al. 2012, 2015, Martinez et al. 2018), have body sizes ranging from 10

to 500g, occupy a variety of locomotor modes, and consume a broad range of foods such as fruit,

seeds, fungi, leaves, roots, and earthworms (Rowe et al. 2016a).

Table 1: Ages of six clades descended from Sulawesi colonists.
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Clade	Crown Age	Species Sampled
Echiothrix	5.11 (4.49-5.69)	10 (10)
Maxomys	3.72 (3.21-4.29)	5 (5)
Bunomys	3.45 (3.09-3.81)	15 (15)
Margaretamys	2.92 (2.29-3.51)	2 (4)
Rattus*	1.57 (1.22-1.95)	6 (6)
Haeromys[†]	Indeterminate	1 (1)

NOTE.- Median age in millions of years is reported with 95% credible intervals in parentheses. Species sampled
reports the number of species in this study with overall clade richness in parentheses and demonstrates near
complete sampling. The two unsampled *Margaretamys* species were not detected in the localities in this study. All
ages taken from Rowe et al. 2019. *There were likely two colonizations by the ancestors of native *Rattus* spp., the
second of which occurred 1.16-0.6 Ma (Rowe et al. 2019), however, all *Rattus* in this study form a clade relative to
other Sulawesi murines. Two human-commensal *Rattus* spp. found on Sulawesi were excluded. [†]The age of arrival
of the Haeromys clade is unknown as other species of Haeromys from Borneo have yet to be included in
phylogenetic analyses.
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Here, we test niche packing and niche expansion as alternative explanations of how trait

variation relates to species richness among nine montane communities of murine rodents on

Sulawesi. We quantify both the volume and density of community ecological space

("ecospace"), which we define as the sum of the n-dimensional niches of the species therein,

from 12 individual functional and ecological traits and five multivariate trait complexes.

Together, these traits reflect the full complexity of biotic interactions. Additionally, we examine

the mode of niche filling in these nine montane communities by quantifying the influence of

phylogenetic diversity and inter-clade variation on ecospace occupancy.

Figure 2: Small mammals surveys of nine mountains A) on the oceanic island of Sulawesi revealed varying murine
rodent species richness across the island. Each mountain on the map contains the maximum elevation and the
number of murine rodent species present. All but Latimojong were surveyed within 600m of the summit. B) The
diversity of Sulawesi murines is exceptional and includes unique forms that live alongside species with more
"typical" ecologies and morphologies, as demonstrated from surfaces of cranial μCT scans of *i – Rattus hoffmanni,* a
"typical" murine morphology and ecology, *ii –* the shrew rat *Echiothrix leucura* of the Echiothrix Division with its
extremely elongate rostrum and soft invertebrate diet, *iii –* the arboreal *Haeromys minahassae*, with a short rostrum
and very small size. C) Photographs: *i – Rattus hoffmanni*, *ii –Echiothrix leucura*, and *iii – Haeromys minahassae.*

Materials and Methods

The distribution of species- We compiled small mammal inventories based on surveys of nine

mountain regions on Sulawesi, Indonesia. These included one well-documented mountain

surveyed from 1973 to 1976 by Guy Musser and colleagues (Mt. Nokilalaki; Musser, 2014) and

eight mountains surveyed from 2011 to 2016 (Ambang, Bawakaraeng, Buliohuto, Dako,

Gandang Dewata, Katopasa, Latimojong, and Torompupu; Figure 2A). All surveys began in

lower primary forest, typically near the line of anthropogenic forest clearing, and extended to

montane forests. All surveys extended to within 600m elevation of the summit except for

Latimojong (highest survey site at 2535m, summit at 3400m). Historical surveys by Musser

lasted several months and were conducted over four years, employing a mix of snap traps and

live traps. Modern surveys (2011-16) lasted an average of 17 days (11-25) and employed similar

collection methods, including a mix of snap traps, live traps, and 20-30L pitfall buckets. All the

murine rodent species known from the sampled localities (Musser 2014, Wilson et al. 2019) were

collected during these modern surveys, indicating a thorough sampling effort. Five new taxa that

were discovered during these expeditions have been described (Musser 2014, Esselstyn et al.

2012, Rowe et al. 2014, Esselstyn et al. 2015, Rowe et al. 2016b) and several new locality

records resulted (Achmadi, et al., 2014; Handika, et al., 2021). Specimens from all surveys were

deposited in the Museum Zoologicum Bogoriense (MZB), Bogor, Indonesia; the American

Museum of Natural History (AMNH), New York, USA; Museums Victoria (MV), Melbourne,

Australia; the Museum of Vertebrate Zoology (MVZ), Berkeley, USA; the Field Museum of

Natural History (FMNH), Chicago, USA; and the Louisiana State University Museum of Natural

Science (LSUMZ), Baton Rouge, USA.

Functional trait data collection and processing- We compiled or generated functional trait

values for 11 continuous traits and one discrete trait for all available Sulawesi murines from each

of the nine communities. Detailed information on traits, sample sizes, data processing, and

multivariate trait composition are available in Table S1. All data, models, and output files are

available in Dryad Repository (to be added prior to publication).

Morphological data collection- We assembled external measurement data from 630 specimens,

including head-body length (mm), tail length (mm), hind-foot length (mm), ear length (mm), and

mass (g), from previously published sources (Wilson et al. 2019, Nations et al. 2021) and online

museum databases. Measured specimens were from the nine surveyed mountains and other

localities on Sulawesi. To obtain ecologically relevant features of external measurements and

mitigate the influence of size in some of our analyses, we calculated three commonly used ratios:

Relative tail length (tail length / (head-body length + tail length)), relative hind-foot length (hind-

foot length / head-body length), and relative ear length (ear length / head-body length) (Nations

et al. 2021, Table S1).

We used a 3-dimensional geometric morphometric approach to calculate the variation in

skull shape for each species. We generated µCT scans of the cranium of 64 specimens from 38

species (Table S1) and the mandible of 61 specimens from 36 species (Table S1). Scans were

generated from specimens collected in the nine surveys, as well as from previously collected

museum materials. Stacks of 2D Tiff files were imported to MorphoDig, where 3D landmarks

were placed on cropped volume renderings (Lebrun, 2018). We placed 67 cranial landmarks

(Figure S2) on the left side of the skull, unless damage caused us to use the right side, in which

case we reversed the rendering on the Z-axis. In separate renderings, we placed 20 landmarks on

the left dentary of the mandible (Figure S2). Landmarks were exported from MorphoDig as .stv

files and imported into the R package geomorph v.4.0.3 for processing (Adams et al. 2021,

Baken et al. 2021). Missing landmarks (11 of 4288 cranial and three of 1220 mandible

landmarks) were imputed with estimate.missing() , a generalized Procrustes analysis (GPA)

superimposition was performed using gpagen(), and shape coordinates were subjected to a

principal components analysis with gm.prcomp(). We retained the centroid size (an estimate of

total size) and the scores from the first 36 principal components of the cranium and 20 axes of

the mandible, each representing >95% of the shape variation of the element.

Stable isotope data collection- Approximately 1-2 grams of hairs were plucked from the rump of

286 dry museum specimens collected on six focal surveys. Isotopic values can vary regionally

(Fry 2006), therefore we collected hair samples from multiple individuals of each species from

each locality (mean = 4.6 specimens/species/locality, range = 1-14). Nitrogen stable isotope

values (δ^{15}N) act as a proxy for consumer trophic position as they generally increase by 3–5%

per trophic level (DeNiro and Epstein 1981). Carbon isotope values (δ^{13}C) generally exhibit little

to no change with trophic position, and are commonly used as proxies of consumer's basal

carbon resource use (e.g. use of differing primary production energy pathways; DeNiro and

Epstein 1978). Combined, these two metrics are commonly used to quantify the "isotopic niche"

of consumers, which can act as a useful proxy of species dietary niche (Newsome et al. 2007,

Ben-David and Flaherty 2012). Stable isotope values are reported in delta notation in per mil

units. Samples were processed at the Stable Isotope Ecology Laboratory, Department of

Oceanography & Coastal Sciences, Louisiana State University. Details on sample cleaning,

processing, and stable-isotope analysis are in the Supporting Methods.

Locomotor mode data collection- Nations et al. (2019, 2021) classified Sulawesi murines into

one of four discrete locomotor modes: Arboreal, which means climbing is integral to survival;

General, or known to navigate a variety of substrates and habitat strata; Terrestrial, or only

known to be on the ground surface; and Amphibious, or a dependence on aquatic habitats for

foraging. Nations et al. (2021) showed that locomotor transitions in murines occur in a set order

(Arboreal ↔ General ↔ Terrestrial ↔ Amphibious) and therefore we treat these modes as

ordinal categories.

Combined traits – We combined our 11 continuous traits into five multivariate traits that

represent biologically meaningful groupings: (1) head shape (cranium shape PC 1-36 and

mandible shape PC 1-20); (2) isotopic niche space (δ^{15}N and δ^{13}C stable isotope values); (3)

body proportions (head-body length, relative tail length, relative hind-foot length, and relative

ear length); (4) body size (log(mass) and head-body length); and (5) total morphological shape

(all nine morphological traits). We did this by combining the predicted trait values for each

community (see below), each represented as a column, into a single dataframe of multiple

columns (Table S1).

Estimating species' trait values- The sample size of morphological and isotopic data varied

between species and mountain communities, and in some cases was limited to one individual,

such as with the amphibious *Waiomys mammasae,* a species known from a single specimen

(Rowe et al. 2014). To mitigate uneven sampling, we estimated a probability distribution of

species trait values using partial-pooling in a multilevel Bayesian model. Unlike complete

pooling (one global mean estimated for all combined samples) or no-pooling (one mean

estimated per species, independent of all others), partial-pooling estimates a mean for each

species as well as the variation among species, which serves as an adaptive prior that is common

to all the species' means (McElreath 2020). The Bayesian partial-pooling modeling allowed us to

incorporate intraspecific variation in trait value predictions while avoiding point estimates such

as averages, which discard valuable information, and prevent unbalanced estimates by using the

trait-variance probability estimates from well-sampled species to inform variance estimates of

species with fewer samples (Gelman and Hill 2006, McElreath 2020). All analyses were

conducted in the probabilistic programming language Stan (Carpenter et al. 2017) within the R

library brms v. 2.17.0 (Bürkner 2018). Subsequent data processing and figuring relied on the R

libraries tidyverse v. 1.3.1 (Wickham et al. 2020), furrr v. 0.3.0 (Vaughan 2021), and tidybayes

v. 3.0.2 (Kay 2020). All data, scripts, and output files are available in Dryad Repository (to be

added prior to publication).

Model, prior, and chain-estimation details are described in the Supporting Methods.

Briefly, for each continuous trait (Table S1), we used the trait value as the response variable, and

used species as a group-level predictor. All traits were scaled to unity prior to analyses. We used

the student-*t* distribution to describe the response variable to minimize the influence of rare,

extreme observations (a.k.a. 'robust regression', Kruschke 2013, McElreath 2020). Each model

included four chains with 4000 iterations of warm-up and 1000 sampling iterations. Posterior

predictions from the four chains were combined, resulting in 4000 samples per trait per species.

To mitigate the potential geographic signal in stable isotope values (Fry 2006), and facilitate

randomization (see below), the posterior estimates of the $\delta^{15}N$ and $\delta^{13}C$ stable isotope values

were estimated with models that included an additional 'community' group-level effect, but

otherwise were identical to the models described above (Supporting Methods). These stable

isotope models generated posterior values of species' isotopic measurements conditioned on the

locality, enabling comparisons among communities and randomized sampling for null models

(Table S1, Figure S3).

Estimating ecospace volume – We defined ecospace volume as the total volume of n-

dimensional trait space occupied by species in a community, and for consistency and clarity we

used the term 'ecospace volume' for all individual and multivariate traits, regardless of

dimension. We used the sum of the variance of each trait value, which outperforms other

commonly used metrics such as ellipse or convex hull volumes (Guillerme et al. 2020), to

estimate ecospace volume. We estimated a posterior distribution of variance of each trait for

each community by first grouping the species' trait value estimates by community. Then, for

each of the 4000 posterior draws (one draw ranging between 7-23 values, depending on the

richness of the community), we estimated the variance of the trait, resulting in a distribution of

4000 variance values for each trait for each community. We estimated the combined multivariate

trait space variance by summing the variances of each trait, then dividing by the number of traits.

To estimate a total community variance for head shape traits (cranium and mandible), we

estimated the variance of each axis for each draw, as above, then calculated a weighted value of

each PC axis by multiplying the value by the contribution of each axis to the multivariate

variation of the PCA, and then summed the normalized values (Cooney et al. 2017). This

avoided both overestimating the impact of PC axes with low importance and arbitrarily dropping

PC axes that may contain valuable information (Cooney et al. 2017, Uyeda et al. 2015). We

estimated the 89% probability values for the variance of each individual and combined trait

space. We estimated the size of locomotor-mode space by calculating the variance of each

community.

Estimating ecospace density- We define ecospace density as the density of species within trait

space. We used the mean nearest neighbor (NN) to estimate the density of trait values among

species (Guillerme et al. 2020) and estimated the density of species in trait space in a similar

manner to estimating the variance. First, we grouped species' trait values by community, then

estimated the individual trait density for each posterior draw using the NN metric in the

disparity() function of the R library dispRity (Guillerme 2018), resulting in a distribution of 4000

NN values for each trait. As NN can be calculated for any number of dimensions, we performed

the same steps for the combined trait spaces. All axes in an NN estimate are considered equally,

and we were unable to weigh cranial and dentary PC axes by their variances as we did above.

Therefore, we conducted a broken stick analysis on the principal components using the R

package PCDimension v.1.1.11 (Coombs & Wang 2019) and found the first two principal

components of the cranial dataset and the first three components of the dentary dataset to be

significant. We used the significant axes in NN estimates of cranial shape, dentary shape, and

head shape density. To estimate locomotor mode density, we dummy coded locomotor mode into

four binary columns (one per mode) and performed a redundancy analysis (Legendre and

Legendre 2012) with the rda() function in the vegan package (Oksanen et al. 2019). We then

extracted the PC scores for each species, grouped the species by community, and estimated the

NN value for each community as with the continuous traits above, in this case generating only a

single NN value per community rather than a distribution.

Null models of ecospace volume and density- We used null models to determine the difference

between the ecospace of our nine communities and randomly assembled communities. For each

community we created 1000 lists of $n_{community}$ species of Sulawesi murines with the

independent swap algorithm, which draws samples using species-occurrence frequency and

community species richness (Gotelli 2001, Swenson 2014), resulting in 1000 randomized null

communities for each of the nine localities. Each null community vector was equal in length to

the richness of the community it represented. We then calculated both volume and density as

above for each individual (n=12) and combined (n=5) trait for each of the 1000 randomized

community samples. We determined the standardized effect size (SES) of each individual and

combined trait value for each community using the formula:

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$$SES = \frac{Trait_{observed} - Mean(Trait_{Random})}{SD(Trait_{Random})},$$

where $Trait_{observed}$ is the vector of 4000 samples of the density or volume of the given trait,

$Mean(Trait_{Random})$ and $SD(Trait_{Random})$ are the mean and standard deviation of the density

or volume values of the given trait from the 1000 random species assemblies. This method

produced distributions of SES values for both the density and volume of each individual and

combined trait for each community. Positive SES values indicate greater than random ecospace

volume or lower than random ecospace density (overdispersion), meaning that species in that

community are occupying ecospace outside the range of other communities and/or their niches

are farther apart in ecospace (niche expansion). Negative SES values indicate lower than random

ecospace volume and greater than random ecospace density (underdispersion or clustering),

meaning that species in that community are occupying less ecological space than other

communities and/or their niches are closer together in ecospace (niche packing; Oliveria et al.

2020).

Effect of species richness on community ecospace volume and density- If niche expansion is the

primary mode of resource partitioning in Sulawesi murine communities, then we expected that

ecospace volume will increase with increasing species richness, and that ecospace density will

remain stable as richness increases. Whereas if niche packing is occurring, we expected that

ecospace density will increase with species richness, and that there will be no effect of richness

on ecospace volume (Figure 1). To quantify the effect size of richness ($\beta_{richness}$) on ecospace

volume and density, we used Bayesian linear regression models that include the measurement

error of the predictor variable to estimate the effect size of species richness on volume or density.

Measurement error models contain vastly more information per observation than a single point

observation and provide robust estimates despite the small number of sampled communities

(Bürkner 2018, McElreath 2020). The models, priors, and chain estimations are detailed in the

Supporting Methods. Briefly, we used species richness as the response variable, and the mean

and standard error of the trait estimates for each community as the predictor. We ran one model

for the SES volume or density of each individual and combined trait space. We also calculated

the Bayesian R^2 value for each regression (Gelman et al. 2019).

Phylogenetic diversity as a path to trait disparity- The ecological space occupied by a

community may depend on which lineages or clades are present (Webb 2000). Trait disparity

appears to vary among clades of Sulawesi murines, and if so, then the volume and densities of

ecospaces may be more influenced by phylogenetic diversity than ecological factors. To

determine how phylogenetic diversity mediates ecospace occupancy in the nine murine

communities, we estimated the phylogenetic diversity (PD) of each community using Faith's

metric of branch length (Faith 1996). We removed all but the Sulawesi species (n = 35) from a

time-calibrated phylogenetic hypothesis of Murinae (Nations et al. 2021). For the analyses

described above, we had trait data for four species that are not included in this phylogeny:

Maxomys wattsi, Rattus bontanus, *Rattus mollicomulus*, and *Rattus marmosurus*. We manually

added these four species into the tree using the bind.tip() function in phytools (Revell 2012).

Details on the phylogenetic positioning of these species is found in the Supplemental Methods.

We estimated Faith's metric of PD and, because PD is not independent of species richness, the

SES value of PD using the ses.pd() function in picante (Kembell et al. 2010). As previously

described, we used the independent swap algorithm to sample 1000 random communities and

used linear modeling in brms to estimate the effect size of species richness on SES PD.

Haeromys minahassae is the only representative of its genus on Sulawesi, and, due to its large

phylogenetic distance from other Sulawesi murines, may have an oversized impact on estimates

on community PD. Therefore, we repeated the estimates of SES PD and effect size above

without including *Haeromys minahassae* to ensure unbiased results. To quantify the trait

disparity within and among Sulawesi murine clades, we grouped the posterior distributions of the

species' predicted trait values by clade, then estimated the variance of each of the 12 univariate

traits. We plotted the trait value variance for each community along with the species' predicted

trait values (Figure S4)

Figure 3: Most estimates of community ecospace volume increase with species richness. X-axes show the $\beta_{richness}$
estimates (effect sizes) for each trait space (y-axis) on the left and the Bayesian R^2 for each $\beta_{richness}$ estimate on the
right. Colored point intervals show 89% probability of $\beta_{richness}$ estimates (effect size), with color varying by
$\beta_{richness}$ value. Black point intervals show 50% posterior estimations of Bayesian R^2 for each trait space.
Four of the five combined traits (A) and seven of the 11 individual traits (B), show an increase in volume with
increased richness (i.e., positive $\beta_{richness}$). Phylogenetic diversity also increases with species richness (B). All
combined traits (C) and individual traits (D) show a stable or, surprisingly, decreasing ecospace density (increased
NN distance) with increased species richness.

                                                    Results

Models of trait values- All parameters in the Bayesian multilevel model estimates of trait-value

distributions and measurement-error estimates of the species richness effect on trait values had

ESS > 1000 and a Gelman-Rubin diagnostic $\hat{R} \leq 1.01$, indicating convergence. Raw size and

density values differed from the size and density of SES. To avoid the confounding influence of

species richness in our size and density estimates we only report and discuss the SES values

(Swenson 2014).

Effects of Species Richness on Ecospace Volume and Density- Ecospace volume is positively

correlated with species richness in seven of the 12 traits (locomotor mode, tail length, head-body

length, cranium shape, mass, $\delta^{15}N$, and hind foot length), as shown by the positive effect sizes

from the Bayesian linear models (Figures 3, Table S3) and the greater variance with higher

community richness (Figure 4). The remaining five traits (ear length, cranium size, dentary size,

dentary shape, and $\delta^{13}C$) do not substantially increase in volume with increased species richness.

The SES values of three of the five combined traits increased substantially with species richness

(total morphological shape, external proportions, and head shape) whereas body size and isotopic

niche space showed a moderate probability of increasing with species richness (Figures 3, 4,

Table S2, S3).

Linear regression showed that species richness had little to no effect on ecospace density

(NN SES values) for 10 of the 12 individual traits and three of five multivariate traits (Figures 3,

S4, Table S2). The exceptions were a positive effect size (i.e., decreased density with richness)

on locomotor mode, tail length, total morphological shape, and external proportions, suggesting

an overdispersion of morphology and locomotor diversity in the richest communities.

Phylogenetic Diversity and Ecospace- Phylogenetic diversity (PD) increased with species

richness (Figures 3, 4, Table S2). Including *Haeromys minahassae* in the estimates of PD

changed the PD SES values of the individual communities but had a very small impact on the

linear regressions (Table S4). Results including *Haeromys minahassae* had a slope ($\beta_{richness}$) of

0.158 (89% C.I. of 0.043, 0.272) while excluding *H. minahassae* generated a slope of 0.168

(0.039, 0.297), and we therefore present the results that include *H. minahassae*. Katopasa, a

community of 12 murines on Sulawesi's Eastern Peninsula, has the lowest PD SES value

followed by Bawakaraeng on the Southwestern Peninsula (seven species). Gandang Dewata, the

richest community, has the highest PD SES value (Figure 4A, Table S2). As we suspected,

ecospace volumes vary between clades, though not consistently (Figure S5). For example, the

Echiothrix clade has the highest variance in cranial shape, dentary shape, and dentary size but

low trait variance for hind-foot length and $\delta^{15}N$. In contrast, the Maxomys clade has low variance

for all trait values except tail length and $\delta^{15}N$ (Figure S5). This results in different densities of

ecospace occupation among traits, where some trait volumes are strongly influenced by

phylogeny, such as cranial space, while other traits such as isotopic niche space have high and

low values distributed among clades (Figure 5).

Figure 4: Estimated trait volume SES plotted against species richnesss: A) Combined trait space volumes, B) Skull traits, C) External measurements, and D) Isotopic measures, locomotor mode, and phylogenetic diversity. Points represent the mean SES values and error bars indicate the 89% credible intervals. X-axis tick marks show the species richness values for each of the communities by mountain. Values equal to zero are consistent with null expectations, positive values indicate overdispersion, and negative values show underdispersion (trait clustering). Dotted lines depict the 89% interval of the null distribution. The plot of trait densities is shown in the supporting information (Figure S4)

Figure 5: The mode of ecospace filling varies between traits. The bivariate plots in the left-hand columns depict the 2D ecospaces. Each black shape within the ecospace is the mean value of a particular species in each clade (shown in legend). The convex hull colors match the color of the community variance SES distributions in the right-hand columns. The thick convex hull lines are mean values, and 100 random samples from the posterior of each trait are shown in thin hull lines. The number of species in each community and each clade used in this study is shown next to the SES distributions in parentheses. A) Large values of hind foot length belong to members of two clades, while large and small tail length values are dispersed among all six clades, reducing the influence of phylogenetic diversity on tail length disparity. B) Cranial morphospace values are mostly clustered tightly near the mean of each axis, apart from Echiothrix species and the single Haeromys species. Intraspecific variance is also relatively low on these axes. C) Intraspecific variance is high in isotopic niche space. High and low δ^{15}N values are distributed among clades, though δ^{13}C values less so.

Discussion

The recognition that species richness often covaries with habitat complexity and resource availability gave rise to the theory of limiting similarity, which suggests that the degree of niche overlap governs the intensity of interspecific competition (MacArthur and Levins 1967, Brown 1975). In the richest communities, the narrowing of species' niches and the packing of resource space appears to be a common form of resource partitioning and is particularly evident at high taxonomic levels and broad spatial scales (Karr and James 1975, Pigot et al. 2016, Cooney et al. 2017, McClain et al. 2018, Hughes et al. 2022). However, interpreting the distribution of functional richness across large resource gradients may be difficult due to covarying factors such as environmental changes or taxonomic turnover (Swenson et al. 2016). Communities of closely related species that inhabit similar environments provide a unique window to explore how interspecific competition may affect community functional richness absent confounding factors like taxonomic or environmental variation (Roughgarden 1976, Swenson et al. 2016), but such communities are rare in nature (Swenson et al. 2016, Van de Perre et al. 2020). Here we use a large dataset of ecological and morphological variables along with a novel probabilistic approach to estimate the changes in community ecospace occupancy across a richness gradient. Our results

from nine murine communities living on similar tropical mountains revealed that the relationship

between ecospace volume and density with species richness varies among traits. The most

common relationship was an increase of ecospace volume with increasing richness, consistent

with niche expansion. The cranial shape, body shape, body size, and locomotor mode ecospaces,

all thought to be ecologically important traits in murines and vertebrates in general (Peters 1986,

Losos 2009, Martinez et al. 2018, Nations et al. 2021), increase with richness (Figure 3).

Measurements of skull size, dentary shape and size, ear length, and $\delta^{13}C$ values all demonstrate

either a weak signal of niche packing, or no signal. While the size of many trait ecospaces

increase with richness, ecospace density shows little correlation with species richness and in

some cases, surprisingly, decreases with increased richness. These results, along with a general

underdispersion of trait values in low-richness communities and overdispersion in high-richness

communities (Table S2, Figures 4 & S4), suggest that species in rich communities mitigate

competition by occupying underused niche space. Our results contrast theoretical predictions on

resource partitioning in insular communities. On remote islands, niche packing is expected to

occur with increased species richness if resources are constant across communities, as in our

Sulawesi localities, but if resources covary with species richness, expansion is expected to occur

(Roughgarden 1976). We also found that phylogenetic diversity increases with species richness

and ecospace volume beyond random expectation. Combined, these results point to species

interactions as a mechanism for both phenotypic and phylogenetic overdispersion (Webb et al.

2002, Cavender-Bares et al. 2004), and suggest that, given adequate resources, structural

complexity, and evolutionary time, species can adapt to unique regions of ecospace, often far

from the average trait value, which minimizes niche overlap and cultivates exceptional richness.

Whether in a single desert valley (Brown 1975), or across continental (Maestri and

Patterson 2016) and global latitudinal gradients (Karr and James 1975, Pellissier et al. 2018),

structurally complex habitats are thought to foster higher species diversity. High plant diversity,

in particular, creates a more complex, vertically structured habitat matrix for other plant and

animal species to occupy and has long been tied to higher animal richness (Hutchinson 1959,

MacArthur and MacArthur 1961, Scheffers et al. 2013, Oliveira and Scheffers 2018). Our

estimates of locomotor-mode occupancy clearly demonstrate that vertical habitat partitioning is

critical to maintaining high species richness in Sulawesi murines (Figure 3, Table S2). Strikingly,

the density of locomotor trait space decreases along the richness gradient, indicating a very high

level of trait value overdispersion (Figures 3C & S4). Arboreal, Terrestrial, and Amphibious

locomotor modes each provide access to different microhabitats that contain similar resources.

On Mt. Gandang Dewata, the richest community with 23 species, the amphibious *Waiomys*

mamasae, the terrestrial *Paucidentomys vermidax*, and the arboreal *Sommeromys macrorhinos*,

all consume invertebrates and have similar $\delta^{15}N$ values, yet they are unlikely to compete for

resources due to their distinct microhabitat use, a pattern observed in other insular communities

of closely related vertebrates (e.g., Jamaican *Anolis*; Shoener 1974). Morphological

measurements are often used to infer locomotor mode in a variety of eco-evolutionary contexts

(Ricklefs and Travis 1980, Samuels and Van Valkenburgh 2009, Pianka et al. 2017, Verde-

Arregoitia et al. 2019), and our estimates of tail-length variance, a trait correlated with

locomotion in murines (Nations et al. 2021), increases with richness at nearly the same rate as

locomotor mode (Figure 3). However, we demonstrate that categorical locomotor modes

themselves, especially if shown to be structured in an ordinal manner, can be used directly to

estimate community locomotor variance and provide a simple way to test hypotheses of niche

structure and variance across wide taxonomic levels and geographic scales where continuous

trait data are lacking.

Combining many traits into one multivariate measure of functional diversity is a common

approach in evolutionary ecology. Ordination techniques were especially promoted to overcome

pitfalls from early community ecology studies that used few, largely subjective measures of

resource use (Ricklefs and Travis 1980). However, merging traits into multivariate axes masks

trait-specific processes related to functional space (Spasojevic and Suding 2012, Astor et al.

2014). Indeed, our trait volume and density estimates reveal distinct patterns between individual

and combined traits. For example, isotopic niche space, a combined signal of $\delta^{15}N$ and $\delta^{13}C$

values commonly used in terrestrial ecological studies, exhibits an equivocal signal of niche

packing and expansion (Figure 3B), but individual isotopic values reveal that the packing signal

originates from the static $\delta^{13}C$ value variance along the species richness gradient. The $\delta^{15}N$

value, a signal of trophic level, expands along the richness gradient, a result that would be

overlooked in multivariate analyses. In contrast to $\delta^{15}N$, dentary shape exhibits an equivocal

signal of packing and expansion, despite its presumed relationship to dietary niche (Maestri et al.

2016, Kohli et al. 2019). These opposing patterns are only illuminated by analyzing differences

in the functional diversity of individual traits (Spasojevic and Suding 2012). If we performed this

study using only cranial size and dentary shape, which are thought to capture the important axes

of size and diet in murine rodents (Rowsey et al. 2019, 2020) and mammals in general (Prevosti

et al. 2012, Grossnickle 2020), our results would support niche packing as the dominant pattern.

The theory of limiting similarity is based on both the mean and variance of species' niche

space (MacArthur and Levins 1967), and intraspecific trait value variation may equal or exceed

interspecific variation (Des Roches et al. 2017). Ignoring the differences between individuals of

a species may taint inferences of functional richness within communities (Violle et al. 2012).

Here we demonstrate a multilevel approach (Bürkner 2018) to model species' trait distributions

and estimate richness and disparity metrics that fully incorporates intraspecific variation, thereby

circumventing the typical concerns associated with point estimates or undersampling (Des

Roches et al. 2017). We then used those values to estimate distributions of ecospace volume and

density. This approach has several favorable properties. First, it allows for any level of

intraspecific variation, and seamlessly incorporates this variation in predictions of community

ecospace volume and density. Additionally, including uncertainty in ecospace values provides

information about the trait itself. Using isotopic niche space as an example, the measure of

uncertainty in species-level stable isotope values is far greater than the intraspecific variation

estimated for other traits such as tail length or cranial shape (Figures 4 and 5). This result

suggests substantial flexibility within species' diets, demonstrating how diversifying dietary

items may aid niche partitioning within communities (Grant 1986).

The disparity of traits within a clade can mediate the impact of phylogenetic diversity on

community ecospace volume. For example, if trait values are phylogenetically clustered, then

increasing phylogenetic diversity, such as through invasion by new clades, is necessary for

community niche expansion to occur. But if trait values are phylogenetically overdispersed,

niche packing or expansion could result from increased phylogenetic diversity. This phenomenon

is especially important for communities that are assembled through a mix of island colonization

and in situ speciation such as Sulawesi murines, Caribbean anoles, or Hawaiian spiders

(Gillespie 2004, Losos 2009, Rowe et al. 2019; Handika et al 2022). Niche divergence has been

hypothesized to overcome niche conservatism in communities with an extended history of

coevolution, such as those on oceanic islands, likely diminishing the signal of phylogenetic trait

clustering (Losos et al. 2003). Contrarily, we found that for most traits ecospace volume

increases with species richness and phylogenetic diversity. For example, the distinct skull

features of species in the Echiothrix division, the descendants of the first murine colonists on

Sulawesi, set them apart from other clades in skull shape and, despite their relatively low

abundance, these species overcontribute to community cranial and dentary shape volumes

(Figures 4B, 5B, S5). However, we also found that the effect of phylogenetic diversity on

ecospace size and density varies between traits. Unlike cranial shape, trophic level estimates

from $\delta^{15}N$ values are notably dispersed (Figures 5C, S5), and high phylogenetic diversity is not

necessary for high isotopic niche space estimates. Are there innate differences in these traits that

could lead to opposing patterns of niche conservatism?

The evolutionary lability of ecologically important traits determines the rate of

convergence and divergence possible in a given time frame and can directly influence dispersion

of trait values among species (Cavender-Bares et al. 2004). The traits with the highest within-

clade variance — $\delta^{15}N$, tail length, body size — are all thought to be evolutionarily labile.

Changes in tail length, body size, and intestinal tract morphology can occur on brief evolutionary

time scales (Powell and King 1997, Naya et al. 2008, Kingsley et al. 2017, 2021), whereas

morphological changes in cranial shape, such as substantial rostral elongation and the reduction

of molar grinding area, may take much longer. Indeed, insular species are known to have rapidly

expanded breadths of diet and labile morphological traits following colonization (Stuart et al.

2014) and subsequent speciation (Wilson 1959, 1961, Lister 1976, Rowe et al. 2016a). Yet the

extreme morphologies of some Sulawesi murines, particularly those of the oldest radiation on the

island (Echiothrix division), are likely the result of a long process of in-situ evolution (Rowe et

al. 2019). Our results point toward evolvability of traits as a determinant of patterns of trait value

dispersion (Webb et al. 2002, Cavender-Bares et al. 2004), which directly relates to our

inferences of niche expansion and packing in Sulawesi murines. Less understood is how varying

evolutionary lability affects dispersion of trait values. The theory of niche complementarity

suggests that a pair of species that are similar in one trait should diverge in another trait

(Schoener 1974). Therefore, similarities in some slowly evolving traits may lead to higher

divergences in more labile traits, or vice versa. Linking the degree of trait lability with niche

complementarity in an island system has important implications for the generation of functional

and taxonomic diversity and may further illuminate the process of niche filling in endemic

radiations.

Literature Cited

Achmadi, A. S., K. C. Rowe, and J. A. Esselstyn. 2014. New records of two rarely encountered,

endemic rats (Rodentia: Muridae: Murinae) from Gunung Gandangdewata, West

Sulawesi province. Treubia 41:51-60.

Adams, D. C., M. Collyer, and A. Kaliontzopoulou. 2021. Geomorph: Software for geometric

morphometric analyses. R package version 4.0.3.

Astor, T., J. Strengbom, M. P. Berg, L. Lenoir, B. Marteinsdóttir, and J. Bengtsson. 2014.

Underdispersion and overdispersion of traits in terrestrial snail communities on islands.

Ecology and Evolution 4:2090-2102.

Baken, E. K., M. L. Collyer, A. Kaliontzopoulou, and D. C. Adams. 2021. Geomorph v4.0 and

gmShiny: enhanced analytics and a new graphical interface for a comprehensive

morphometric experience. Methods in Ecology and Evolution. 12:2355-2363.

Baltensperger, A. P., F. Huettmann, J. C. Hagelin, and J. M. Welker. 2015. Quantifying trophic

niche spaces of small mammals using stable isotopes (δ^{15}N and δ^{13}C) at two scales across

Alaska. Canadian Journal of Zoology 93:579-588.

Ben-David, M., and E. A. Flaherty. 2012. Stable isotopes in mammalian research: a beginner's

guide. Journal of Mammalogy 93:312-328.

Brown, J. H., and G. A. Lieberman. 1973. Resource utilization and coexistence of seed-eating

desert rodents in sand dune habitats. Ecology 54:788-797.

Brown, J. H. 1975. Geographical Ecology of Desert Rodents. Pages 315-341 *in* M. L. Cody and

J. M. Diamond, editors. Ecology and Evolution of Communities. The Belknap Press,

Cambridge.

618 Bürkner, P.-C. 2018. Advanced Bayesian Multilevel Modeling with the R Package brms. The R

619   Journal 10:395–411.

620 Carpenter, B., A. Gelman, M. D. Hoffman, D. Lee, B. Goodrich, M. Betancourt, M. Brubaker, J.

621   Guo, P. Li, and A. Riddell. 2017. Stan: A Probabilistic Programming Language. Journal

622   of Statistical Software 76.

623 Cavender-Bares, J., D. D. Ackerly, D. A. Baum, and F. A. Bazzaz. 2004. Phylogenetic

624   Overdispersion in Floridian Oak Communities. The American Naturalist 163:823-843.

625 Coombes, K. R. and M. Wang (2019). PCDimension: Finding the Number of Significant

626   Principal Components. R package version 1.1.11.

627 Cooney, C. R., J. A. Bright, E. J. Capp, A. M. Chira, E. C. Hughes, C. J. Moody, L. O. Nouri, Z.

628   K. Varley, and G. H. Thomas. 2017. Mega-evolutionary dynamics of the adaptive

629   radiation of birds. Nature 542:344-347.

630 DeNiro, M. J., and S. Epstein. 1978. Influence of diet on the distribution of carbon isotopes in

631   animals. Geochimica et Cosmochimica Acta 42:495-506.

632 DeNiro, M. J., and S. Epstein. 1981. Influence of diet on the distribution of nitrogen isotopes in

633   animals. Journal of Archeological Science 45:341-351.

634 Des Roches, S., D. M. Post, N. E. Turley, J. K. Bailey, A. P. Hendry, M. T. Kinnison, J. A.

635   Schweitzer, and E. P. Palkovacs. 2018. The ecological importance of intraspecific

636   variation. Nature Ecology & Evolution 2:57-64.

637 Diamond, J. M. 1975. Assembly of Species Communities. Pages 342-444 *in* M. L. Cody and J.

638   M. Diamond, editors. Ecology and Evolution of Communities. The Belknap Press,

639   Cambridge.

Dorey, J. B., S. V. Groom, E. H. Freedman, C. S. Matthews, O. K. Davies, E. J. Deans, C.

Rebola, M. I. Stevens, M. S. Lee, and M. P. Schwarz. 2020. Radiation of tropical island

bees and the role of phylogenetic niche conservatism as an important driver of

biodiversity. Proceedings of the Royal Society B 287:20200045.

Esselstyn, J. A., A. S. Achmadi, and K. C. Rowe. 2012. Evolutionary novelty in a rat with no

molars. Biology Letters 8:990-993.

Esselstyn, J. A., A. S. Achmadi, H. Handika, and K. C. Rowe. 2015. A hog-nosed shrew rat

(Rodentia: Muridae) from Sulawesi Island, Indonesia. Journal of Mammalogy 96:895-

907.

Esselstyn, J. A., A. S. Achmadi, H. Handika, M. T. Swanson, T. C. Giarla, and K. C. Rowe.

2021. Fourteen New, Endemic Species of Shrew (Genus *Crocidura*) from Sulawesi

Reveal a Spectacular Island Radiation. Bulletin of the American Museum of Natural

History 454:1-108.

Faith, D. P. 1996. Conservation priorities and phylogenetic pattern. Conservation Biology

10:1286-1289.

Fry, B. 2006. Stable isotope ecology. Springer, New York.

Galetti, M., R. R. Rodarte, C. L. Neves, M. Moreira, and R. Costa-Pereira. 2016. Trophic Niche

Differentiation in Rodents and Marsupials Revealed by Stable Isotopes. PLOS One

11:e0152494.

Gelman A, Hill J. 2006. Data Analysis Using Regression and Multilevel/Hierarchical Models.

Cambridge: Cambridge University Press.

Gelman, A., B. Goodrich, J. Gabry, and A. Vehtari. 2019. R-squared for Bayesian regression

models. The American Statistician 73:307-309.

Gillespie, R. 2004. Community assembly through adaptive radiation in Hawaiian spiders.

Science 303:356-359.

Gotelli, N. J. 2001. Research frontiers in null model analysis. Global Ecology and Biogeography

10:337-343.

Grant, P. R. 1986. Ecology and Evolution of Darwin's Finches. Princeton University Press,

Princeton.

Grossnickle, D. M. 2020. Feeding ecology has a stronger evolutionary influence on functional

morphology than on body mass in mammals. Evolution 74:610-628.

Guillerme T. 2018. dispRity: A modular R package for measuring disparity. Methods in Ecology

and Evolution 9:1755-1763.

Guillerme, T., M. N. Puttick, A. E. Marcy, and V. Weisbecker. 2020. Shifting spaces: Which

disparity or dissimilarity measurement best summarize occupancy in multidimensional

spaces? Ecology and Evolution 10:7261-7275.

Handika, H., A. S. Achmadi, J. A. Esselstyn, and K. C. Rowe. 2021. Molecular and

morphological systematics of the Bunomys division (Rodentia: Muridae), an endemic

radiation on Sulawesi. Zoologica Scripta 50:141-154.

Hiller, A. E., M. S. Koo, K. R. Goodman, K. L. Shaw, P. M. O'Grady, and R. G. Gillespie. 2019.

Niche conservatism predominates in adaptive radiation: comparing the diversification of

Hawaiian arthropods using ecological niche modelling. Biological Journal of the Linnean

Society 127:479-492.

Hughes, E. C., D. P. Edwards, J. A. Bright, E. J. Capp, C. R. Cooney, Z. K. Varley, and G. H.

Thomas. 2022. Global biogeographic patterns of avian morphological diversity. Ecology

Letters 25:598-610.

Hutchinson, G. E. 1957. Concluding remarks. Cold Spring Harbor Symposia on Quantitative

Biology 22:415-427.

Hutchinson, G. E. 1959. Homage to Santa Rosalia or Why Are There So Many Kinds of

Animals? The American Naturalist 93:145-159.

Imhoff, M. L., L. Bounoua, T. Ricketts, C. Loucks, R. Harriss, and W. T. Lawrence. 2004.

Global patterns in human consumption of net primary production. Nature 429:870-873.

Iskandar, D., and T. Nio. 1996. The amphibians and reptiles of Sulawesi, with notes on the

distribution and chromosomal number of frogs. Pages 39-46 *in* Proceedings of the First

International Conference on Eastern Indonesian-Australian Vertebrate Fauna. Manado,

Indonesia.

Karr, J. R., and F. C. James. 1975. Ecomorphological Configurations and Convergent Evolution.

Pages 258-291 *in* M. L. Cody and J. M. Diamond, editors. Ecology and Evolution of

Communities. The Belknap Press, Cambridge.

Kay, M. 2020. Tidybayes: Tidy data and geoms for Bayesian models. R package version 2.1.1.

Kembel, S. W., P. D. Cowan, W. K. Helmus, W. K. Cornwell, H. Morlon, D. D. Ackerly, S. P.

Blomberg, and C. O. Webb. 2010. Picante: R tools for integrating phylogenies and

ecology. Bioinformatics 26:1463-1464.

Kingsley, E. P., K. M. Kozak, S. P. Pfeifer, D. S. Yang, and H. E. Hoekstra. 2017. The ultimate

and proximate mechanisms driving the evolution of long tails in forest deer mice.

Evolution 71:261-273.

Kingsley, E. P., E. R. Hager, J. M. Lassance, K. M. Turner, O. S. Harringmeyer, C. Kirby, B. I.

Neugeboren, and H. E. Hoekstra. 2021. Adaptive tail-length evolution in deer mice is

associated with differential Hoxd13 expression in early development. bioRxiv

2021.12.18.473263.

Kohli, B. A., and R. J. Rowe. 2019. Beyond guilds: the promise of continuous traits for

mammalian functional diversity. Journal of Mammalogy 100:285-298.

Kohli, B. A., R. D. Stevens, E. A. Rickart, and R. J. Rowe. 2021. Mammals on mountainsides

revisited: Trait-based tests of assembly reveal the importance of abiotic filters. Journal of

Biogeography 48:1606-1621.

Kruschke, J. K. 2013. Bayesian estimation supersedes the t test. Journal of Experimental

Psychology: General 142:573.

Lebrun, R. 2018. MorphoDig, an open-source 3D freeware dedicated to biology. IPC5, Paris,

France.

Legendre, P., and L. Legendre. 2012. Numerical ecology. Elsevier, Amsterdam.

Lister, B. C. 1976. The nature of niche expansion in West Indian Anolis lizards I: ecological

consequences of reduced competition. Evolution 30:659-676.

Losos JB. 2009. Lizards in an evolutionary tree: ecology and adaptive radiation of anoles.

University of California Press, Berkeley.

Losos, J. B., and R. E. Ricklefs. 2009. Adaptation and diversification on islands. Nature

457:830-836.

M'Closkey, R. T. 1978. Niche separation and assembly in four species of Sonoran Desert

rodents. The American Naturalist 112:683-694.

MacArthur, R. H. 1965. Patterns of species diversity. Biological Reviews 40:510-533.

MacArthur, R. H. 1970. Species packing and competitive equilibrium for many species.

Theoretical Population Biology 1:1-11.

MacArthur, R., and R. Levins. 1967. The limiting similarity, convergence, and divergence of

coexisting species. The American Naturalist 101:377-385.

MacArthur, R. H., and J. W. MacArthur. 1961. On bird species diversity. Ecology 42:594-598.

Maestri, R., B. Patterson, R. Fornel, L. Monteiro, and T. De Freitas. 2016. Diet, bite force and

skull morphology in the generalist rodent morphotype. Journal of Evolutionary Biology

29:2191-2204.

Maestri, R., and B. D. Patterson. 2016. Patterns of species richness and turnover for the South

American rodent fauna. PLoS One 11:e0151895.

Martinez, Q., R. Lebrun, A. S. Achmadi, J. A. Esselstyn, A. R. Evans, L. R. Heaney, R. P.

Miguez, K. C. Rowe, and P.-H. Fabre. 2018. Convergent evolution of an extreme dietary

specialization, the olfactory system of worm-eating rodents. Scientific Reports 8:17806.

May, R. M., and R. H. Mac Arthur. 1972. Niche overlap as a function of environmental

variability. Proceedings of the National Academy of Sciences 69:1109-1113.

McElreath R. 2020 Statistical Rethinking v2. Taylor & Francis Group, Boca Raton.

McClain, C. R., C. Nunnally, A. S. A. Chapman, and J. P. Barry. 2018. Energetic increases lead

to niche packing in deep-sea wood falls. Biology Letters 14:20180294.

McGill, B. J., B. J. Enquist, E. Weiher, and M. Westoby. 2006. Rebuilding community ecology

from functional traits. Trends in Ecology & Evolution 21:178-185.

Musser, G. G. 2014. A systematic review of Sulawesi Bunomys (Muridae, Murinae) with the

description of two new species. Bulletin of the American Museum of Natural History

392.

Nations, J. A., L. R. Heaney, T. C. Demos, A. S. Achmadi, K. C. Rowe, and J. A. Esselstyn.

2019. A simple skeletal measurement effectively predicts climbing behavior in a diverse

clade of small mammals. Biological Journal of the Linnean Society 128:323–336.

Nations, J. A., G. G. Mount, S. M. Morere, A. S. Achmadi, K. C. Rowe, and J. A. Esselstyn.

2021. Locomotory mode transitions alter phenotypic evolution and lineage diversification

in an ecologically rich clade of mammals. Evolution 75:376-393.

Naya, D. E., F. Bozinovic, and W. H. Karasov. 2008. Latitudinal trends in digestive flexibility:

testing the climatic variability hypothesis with data on the intestinal length of rodents.

The American Naturalist 172:122-134.

Newsome, S. D., C. Martinez del Rio, S. Bearhop, and D. L. Phillips. 2007. A niche for isotopic

ecology. Frontiers in Ecology and the Environment 5:429-436.

Oksanen, J, F. Guillaume Blanchet, M. Friendly, R. Kindt, P. Legendre, D. McGlinn, P. R.

Minchin, R. B. O'Hara, G. L. Simpson, P. Solymos, M. Henry, H. Stevens, E. Szoecs and

H. Wagner (2019). Vegan: Community Ecology Package. R package version 2.5-6.

Oliveira, B. F., and B. R. Scheffers. 2019. Vertical stratification influences global patterns of

biodiversity. Ecography 42:249-249.

Oliveira, B. F., J. M. Flenniken, R. P. Guralnick, S. E. Williams, and B. R. Scheffers. 2020.

Historical environmental stability drives discordant niche filling dynamics across

phylogenetic scales. Journal of Biogeography 47:807-816.

Pellissier, V., J. Y. Barnagaud, W. D. Kissling, Ç. Şekercioğlu, and J. C. Svenning. 2018. Niche

packing and expansion account for species richness–productivity relationships in global

bird assemblages. Global Ecology and Biogeography 27:604-615.

Peters, R. H. 1986. The Ecological Implications of Body Size. Cambridge University Press,

Cambridge.

Pianka, E. R. 1974. Niche overlap and diffuse competition. Proceedings of the National

Academy of Sciences 71:2141-2145.

Pianka, E. R., L. J. Vitt, N. Pelegrin, D. B. Fitzgerald, and K. O. Winemiller. 2017. Toward a

Periodic Table of Niches, or Exploring the Lizard Niche Hypervolume. The American

Naturalist 190:601-616.

Pigot, A. L., C. H. Trisos, and J. A. Tobias. 2016. Functional traits reveal the expansion and

packing of ecological niche space underlying an elevational diversity gradient in

passerine birds. Proceedings of the Royal Society B: Biological Sciences 283:20152013.

Pigot, A. L., C. Sheard, E. T. Miller, T. P. Bregman, B. G. Freeman, U. Roll, N. Seddon, C. H.

Trisos, B. C. Weeks, and J. A. Tobias. 2020. Macroevolutionary convergence connects

morphological form to ecological function in birds. Nature Ecology & Evolution 4:230-

239.

Powell, R. A., and C. M. King. 1997. Variation in body size, sexual dimorphism and age-specific

survival in stoats, Mustela erminea (Mammalia: Carnivora), with fluctuating food

supplies. Biological Journal of the Linnean Society 62:165-194.

Prevosti, F. J., G. F. Turazzini, M. D. Ercoli, and E. Hingst-Zaher. 2012. Mandible shape in

marsupial and placental carnivorous mammals: a morphological comparative study using

geometric morphometrics. Zoological Journal of the Linnean Society 164:836-855.

Revell, L. J. 2012. Phytools: an R package for phylogenetic comparative biology (and other

things). Methods in Ecology and Evolution 3:217-223.

Ricklefs, R. E., and J. Travis. 1980. A Morphological Approach to the Study of Avian

Community Organization. The Auk 97:321-338.

Rowe, K. C., A. S. Achmadi, and J. A. Esselstyn. 2014. Convergent evolution of aquatic

foraging in a new genus and species (Rodentia: Muridae) from Sulawesi Island,

Indonesia. Zootaxa 3815:541-564.

Rowe K.C., A.S. Achmadi, and J.A. Esselstyn. 2016a. Repeated evolution of carnivory among

Indo-Australian rodents. Evolution 70:653-665.

Rowe, K. C., A. S. Achmadi, and J. A. Esselstyn. 2016b. A new genus and species of

omnivorous rodent (Muridae: Murinae) from Sulawesi, nested within a clade of endemic

carnivores. Journal of Mammalogy 97:978-991.

Rowe, K. C., A. S. Achmadi, P. H. Fabre, J. J. Schenk, S. J. Steppan, and J. A. Esselstyn. 2019.

Oceanic islands of Wallacea as a source for dispersal and diversification of murine

rodents. Journal of Biogeography 46:2752-2768.

Rowsey, D. M., L. R. Heaney, and S. A. Jansa. 2019. Tempo and mode of mandibular shape and

size evolution reveal mixed support for incumbency effects in two clades of island-

endemic rodents (Muridae: Murinae). Evolution 73:1411-1427.

Rowsey, D. M., R. M. Keenan, and S. A. Jansa. 2020. Dietary morphology of two island-

endemic murid rodent clades is consistent with persistent, incumbent-imposed

competitive interactions. Proceedings of the Royal Society B 287:20192746.

Roughgarden, J. 1976. Resource partitioning among competing species–a coevolutionary

approach. Theoretical Population Biology 9:388-424.

Samuels, J. X., and B. Van Valkenburgh. 2008. Skeletal indicators of locomotor adaptations in

living and extinct rodents. Journal of Morphology 269:1387-1411.

Scheffers, B. R., B. L. Phillips, W. F. Laurance, N. S. Sodhi, A. Diesmos, and S. E. Williams.

2013. Increasing arboreality with altitude: a novel biogeographic dimension. Proceedings

of the Royal Society B 280:20131581.

Schoener, T. W. 1974. Resource Partitioning in Ecological Communities: Research on how

similar species divide resources helps reveal the natural regulation of species diversity.

Science 185:27-39.

Spasojevic, M. J., and K. N. Suding. 2012. Inferring community assembly mechanisms from

functional diversity patterns: the importance of multiple assembly processes. Journal of

Ecology 100:652-661.

Stroud, J. T., and J. B. Losos. 2016. Ecological Opportunity and Adaptive Radiation. Annual

Review of Ecology, Evolution, and Systematics 47:507-532.

Stroud, J. T. 2021. Island species experience higher niche expansion and lower niche

conservatism during invasion. Proceedings of the National Academy of Sciences 118:

e2018949118.

Stuart, Y. E., T. S. Campbell, P. a. Hohenlohe, R. G. Reynolds, L. J. Revell, and J. B. Losos.

2014. Rapid evolution of a native species following invasion by a congener. Science

346:463-466.

Swenson, N. G. 2014. Functional and Phylogenetic Ecology in R. Springer, New York.

Swenson, N. G., M. D. Weiser, L. Mao, S. Normand, M. Á. Rodríguez, L. Lin, M. Cao, and J.-C.

Svenning. 2016. Constancy in functional space across a species richness anomaly. The

American Naturalist 187:E83-E92.

Uyeda, J. C., D. S. Caetano, and M. W. Pennell. 2015. Comparative Analysis of Principal

Components Can be Misleading. Syst Biol 64:677-689.

Van de Perre, F., M. R. Willig, S. J. Presley, I. J.-C. Mukinzi, M. S. Gambalemoke, H. Leirs, and

E. Verheyen. 2020. Functional volumes, niche packing and species richness:

biogeographic legacies in the Congo Basin. Royal Society Open Science 7:191582.

Vaughan, D, and M. Dancho 2021. furrr: Apply Mapping Functions in Parallel using Futures. R

package version 0.2.2.

Verde Arregoitia, L. D., D. O. Fisher, and M. Schweizer. 2017. Morphology captures diet and

locomotor types in rodents. Royal Society Open Science 4:160957.

Violle, C., B. J. Enquist, B. J. McGill, L. Jiang, C. H. Albert, C. Hulshof, V. Jung, and J.

Messier. 2012. The return of the variance: intraspecific variability in community ecology.

Trends in Ecology & Evolution 27:244-252.

Webb, C. O. 2000. Exploring the phylogenetic structure of ecological communities: an example

for rain forest trees. The American Naturalist 156:145-155.

Webb, C. O., D. D. Ackerly, M. A. McPeek, and M. J. Donoghue. 2002. Phylogenies and

community ecology. Annual Review of Ecology and Systematics 33:475-505.

Weiher, E., and P. A. Keddy. 1995. Assembly rules, null models, and trait dispersion: new

questions from old patterns. Oikos:159-164.

Wickham, H., M. Averick, J. Bryan, W. Chang, L. D. A. McGowan, R. François, G. Grolemund,

A. Hayes, L. Henry, and J. Hester. 2019. Welcome to the Tidyverse. Journal of Open

Source Software 4:1686.

Wilson, D. E., R. A. Mittermeier, and T. E. Lacher. 2019. Handbook of the Mammals of the

World. Lynx Editions, Barcelona.

Wilson, E. O. 1959. Adaptive shift and dispersal in a tropical ant fauna. Evolution 13:122-144.

Wilson, E. O. 1961. The Nature of the Taxon Cycle in the Melanesian Ant Fauna. The American

Naturalist 95:169-193.

Woodward, G., and A. G. Hildrew. 2002. Body-size determinants of niche overlap and intraguild

predation within a complex food web. Journal of Animal Ecology 71:1063-1074.

References Only Cited in the Online Supplement

Achmadi, A. S. 2010. Taxonomic Status of Spiny Rats (Maxomys Jentink, Rodentia) from

Indonesia and Malaysia Based on Morphological Study. Treubia 37:49-82.

Fry, B. 2006. Stable isotope ecology. Springer, New York.

Hückstädt, L., J. Burns, P. Koch, B. McDonald, D. E. Crocker, and D. Costa. 2012. Diet of a

specialist in a changing environment: the crabeater seal along the western Antarctic

Peninsula. Marine Ecology Progress Series 455:287-301.

Kruschke, J. K. 2013. Bayesian estimation supersedes the t test. Journal of Experimental

Psychology: General 142:573.

McElreath R. 2020 Statistical Rethinking v2. Taylor & Francis Group. Boca Raton.

Musser, G. G., and M. D. Carleton. 2005. Superfamily Muroidea. Pages 894–1531 in D. E.

Wilson, Reeder D.M., editor. Mammal species of the world: a taxonomic and geographic

reference. Johns Hopkins Univ. Press, Baltimore.

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