

1     **Evolution of median fin patterning and modularity in living and fossil osteichthyans**

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3     **Short title:** Median fin patterning in osteichthyans

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20

## 21 **Abstract**

22           Morphological and developmental similarities, and interactions among developing  
23 structures are interpreted as evidences of modularity. Such similarities exist between the  
24 dorsal and anal fins of living actinopterygians: (1) both fins differentiate in the same  
25 direction [dorsal and anal fin patterning module (DAFPM)], and (2) radials and  
26 lepidotrichia differentiate in the same direction [endoskeleton and exoskeleton module  
27 (EEM)]. To infer the evolution of these common developmental patternings among  
28 osteichthyans, we address (1) the complete description and quantification of the DAFPM  
29 and EEM in a living actinopterygian (the rainbow trout *Oncorhynchus mykiss*) and (2) the  
30 presence of these modules in fossil osteichthyans (coelacanth, lungfishes, porolepiforms  
31 and ‘osteolepiforms’). In *Oncorhynchus*, sequences of skeletal elements are determined  
32 based on (1) apparition (radials and lepidotrichia), (2) chondrification (radials), (3)  
33 ossification (radials and lepidotrichia), and (4) segmentation plus bifurcation  
34 (lepidotrichia). Correlations are then explored between sequences. In fossil osteichthyans,  
35 sequences are determined based on (1) ossification (radials and lepidotrichia), (2)  
36 segmentation, and (3) bifurcation of lepidotrichia. Segmentation and bifurcation patterns  
37 were found crucial for comparisons between living and extinct taxa. Our data suggest that  
38 the EEM is plesiomorphic at least for actinopterygians, and the DAFPM is plesiomorphic  
39 for osteichthyans, with homoplastic dissociation. Finally, recurrent patterns suggest the  
40 presence of a Lepidotrichia Patterning Module (LPM).

41

## 42 **Introduction**

43           In the past two decades, the median fins [i.e., dorsal, anal, and caudal fins] of fishes  
44 have been the focus of an overwhelming body of research in evolutionary developmental  
45 biology. Primary interest for these so-called unpaired fins lies in their locomotor functions  
46 [1–3], ecological implications [4,5], comparative morpho-anatomy [6], molecular [7] and  
47 developmental [8] patterning, as well as morphological disparity [9]. Indeed, among  
48 median fins, the dorsal and anal fins of piscine osteichthyans show a great morphological  
49 disparity, reflecting the evolvability of this system [9–11].

50           Osteichthyans primitively display two dorsal fins and a single anal fin [12].  
51 Independently and repeatedly in actinopterygians and sarcopterygians, the number of dorsal  
52 fins is reduced by loss or fusion with the caudal fin [9,11,13]. The loss of the anal fin is less  
53 frequent in piscine osteichthyans, although loss or fusion with the caudal fin occurs in some  
54 teleosts and dipnoans [11,14]. Among ‘elpistostegalians’, the extinct transitional taxa  
55 between fishes and tetrapods, the condition is poorly documented (e.g., *Panderichthys*,  
56 *Tiktaalik*). However, *Elpistostege*, considered a basal tetrapod, has lost the dorsal fin while  
57 an anal fin is present [15]. And finally, the absence of both the dorsal and anal fins is  
58 considered a synapomorphy shared by aquatic and terrestrial tetrapods [14] with the  
59 exception of *Elpistostege* [15]. Even with such morphological disparity, the structure and  
60 development of the median fins is expected to be broadly similar among osteichthyans  
61 because these fins have similar constituents.

62           In a forerunner comparative study, Mabee et al. [16] revealed the recurrence of  
63 similar developmental patterning (i.e., sequences and direction of development among

64 endoskeletal and exoskeletal elements) in the dorsal and anal fins among living  
65 actinopterygians, which they interpreted as evidence of modularity. They found out that the  
66 similar patterning of these fins might be indicative of two modules: (1) the Dorsal and Anal  
67 Fin Patterning Module (DAFPM), where the skeletal elements (radial bones and  
68 lepidotrichia) of both fins differentiate in the same direction and (2) the Endoskeleton and  
69 Exoskeleton Module (EEM), where the directions of development of the endoskeleton  
70 (radials) and exoskeleton (lepidotrichia) are similar. The DAFPM and EEM are considered  
71 to be maintained during actinopterygian phylogeny [16] but a phylogenetic inference across  
72 osteichthyans was not possible owing to the absence of comparative data on early  
73 actinopterygians and sarcopterygians. A broad phylogenetic sampling of actinopterygians  
74 and sarcopterygians is thus necessary to document the patterns of developmental similarity  
75 between dorsal and anal fins throughout osteichthyan evolution. However, in order to  
76 validate the prospective existence and distribution of median fin modules among  
77 osteichthyans (i.e., specifically the DAFPM and EEM *sensu* [16]), it is mandatory to  
78 include extant as well as extinct taxa within a comparative framework.

79         The rarity of fossilized ontogenies [17] and a bias toward the preservation of hard  
80 tissues (bones *versus* cartilages) limit our assessment of early developmental patterns in  
81 extinct taxa. However, the presence of fossilized individual ontogenies (i.e., anatomical  
82 structures found in adults that have recorded individual developmental patterns), can  
83 potentially broaden the phylogenetic sampling. The complex structure of osteichthyan fin  
84 rays (e.g., [18,19]) provides such developmental data because fin rays are accretional

85 structures for which structural elements are added without substantial remodelling, thus  
86 allowing the preservation of early developmental patterning.

87 The main objectives of this study are (1) to provide a complete description of the  
88 patterns of developmental similarity of the dorsal and anal fins in a living actinopterygian,  
89 the rainbow trout (*Oncorhynchus mykiss*), (2) to describe the dorsal and anal fins patterning  
90 in fossil osteichthyans, and (3) to compare the similarity of the developmental patterns in  
91 order to discuss the prospective existence of the DAFPM and EEM modules within  
92 osteichthyans. Patterns of developmental similarity were investigated by using relative  
93 developmental sequences and direction of development among endoskeletal and  
94 exoskeletal elements of the dorsal and anal fins. We expect developmental sequences to be  
95 (1) significantly congruent between dorsal and anal fins (i.e., indicative of DAFPM), and  
96 (2) significantly congruent between the endoskeleton and exoskeleton within each fin (i.e.,  
97 indicative of EEM).

98

## 99 **Nomenclature**

100 In order to facilitate comparisons during the description of fin morphologies and its  
101 constituents, a brief review of the main elements composing the median fins of  
102 osteichthyans is presented here. We will detail the different types of fins rays encountered  
103 in osteichthyans (i.e., lepidotrichia and actinotrichia) and the events used to define the  
104 median fin developmental patterning.

## 105 **Lepidotrichia**

106           Lepidotrichia are osseous fin rays of dermal origin. Each lepidotrichium is  
107 composed of two parallel and symmetrical elements called hemirays. Lepidotrichia are  
108 usually segmented (i.e., “jointed” lepidotrichia). The “joints” correspond to very narrow,  
109 non-mineralized spaces between adjacent segments connected by collagenous ligaments  
110 called Sharpey’s fibres. Adjoining segments are sequentially added distally during growth  
111 [20] before the ossification of the ray, which begins proximally. The most proximal  
112 segment is always longer than the others from the very first stages of development. This  
113 pattern has been observed in many living and fossil species (e.g., the actinopterygians  
114 *Gobius*, *Pygosleus*, *Cottus*, and *Blennius*, among others, and the sarcopterygian *Miguashaia*  
115 and *Eusthenopteron* [21–23]. The lepidotrichia articulate with the most distal endoskeletal  
116 elements (e.g., radials or pterygiophores; or the phalanges as in *Elpistostege* [15]) in the  
117 paired and median fins. The hemirays are contralaterally arranged on both sides of the  
118 endoskeleton. Usually, each radial bone carries more than one lepidotrichium, however  
119 certain derived taxa (e.g., teleosts and coelacanths) show a 1:1 ratio between the radials and  
120 the fin rays in the median fins. In their most distal portion, the lepidotrichia usually  
121 bifurcate (i.e., branched lepidotrichia) as if the ray was split in two. Numerous episodes (or  
122 orders) of bifurcation can occur in a single ray. Lepidotrichia represent a synapomorphy of  
123 crown osteichthyans [24]. Extant dipnoans (i.e., *Neoceratodus*, *Lepidosiren* and  
124 *Protopterus*) display a unique kind of partially ossified lepidotrichia termed camptotrichia  
125 [25].

## 126 **Actinotrichia**

127 Actinotrichia are flexible fin rays formed by long fibres of collagen known as  
128 elastoidine [19]. The actinotrichia form the main support of the osteichthyan fins in larval  
129 and juvenile stages of the ontogeny and are found in the most distal part of the adult fins  
130 arranged in contralateral palisades. During the formation of the lepidotrichia, actinotrichia  
131 are progressively resorbed, both within hemirays and between lepidotrichia, leaving only a  
132 narrow distal fringe. The formation of actinotrichia is followed by the apparition and  
133 development of the endoskeletal elements (e.g., radials), and the formation of the  
134 lepidotrichia [16,26–28]. Mesenchymal cells (osteoblasts) may then use the actinotrichia as  
135 a scaffold during the initial stages of formation of the lepidotrichia [19,26,29].

### 136 **Median fin patterning**

137 Median fin patterning can be defined in terms of a series of events rather than solely  
138 on formation (or differentiation, *sensu* [16]). An event defines a unit of transformation with  
139 concomitant phenotypic changes (e.g., lepidotrichia ossification) and an event may have  
140 different properties (e.g., onset, offset, duration) [30]. The chronological order of events  
141 corresponds to a sequence. Fins being composed of different elements, we referred to a  
142 developmental sequence when comparing the same developmental state (e.g., apparition,  
143 chondrification, ossification) among elements. We referred to an ontogenetic sequence  
144 when comparing different developmental states for a single element [31].

145 Of the numerous developmental events associated with fin formation in living  
146 actinopterygians [8,28,32–38], our study focused on 11 skeletogenic events: (1) apparition  
147 (i.e., collagenous matrix precursor) of actinotrichia, (2) apparition (i.e., mesenchymal  
148 condensation) of proximal radials, (3) chondrification of proximal radials, (4) apparition of

149 lepidotrichia, (5) apparition (i.e., mesenchymal condensation) of distal radials, (6)  
150 chondrification of distal radials, (7) segmentation of lepidotrichia, (8) ossification of  
151 lepidotrichia, (9) bifurcation of lepidotrichia, (10) ossification of proximal radials, and (11)  
152 ossification of distal radials. The establishment of these developmental events is a powerful  
153 tool that could be used to uncover patterns of developmental similarity and shed light on  
154 patterns of modularity across osteichthyan evolution.

155

## 156 **Material and methods**

### 157 **Living material**

158         Developmental sequences of the dorsal and anal fins were obtained from embryo-  
159 juvenile specimens of the rainbow trout (*Oncorhynchus mykiss*) ranging from 5 days pre-  
160 hatching to 100 days post-hatching (dph). Alevins-juveniles were reared in swimming  
161 channels under constant water velocity (0.4 cm/s) in 2005 [see [39] for rearing conditions].  
162 Specimens were sampled every day up to 34 dph, every other day from 34 to 80 dph, and  
163 every four days up to 100 dph. Samples were fixed in neutral buffered formalin for 48h,  
164 and then preserved in 70% ethanol. One specimen for each sampling day, plus one or two  
165 replicates for specimens between 0 to 24 dph, were cleared-and-double stained with  
166 Alizarin red S for bones and Alcian blue for cartilages [40]. Replicates were used to palliate  
167 with staining problems [41]. Pre-hatching specimens were removed from their egg capsule  
168 prior to clearing and Alcian blue staining. Digital pictures were taken before staining and 5-  
169 10 days after staining to avoid interpretive errors owing to destaining. In total, eighty



170 specimens were used to reconstruct developmental sequences of serial skeletal elements  
171 from the dorsal and anal fins (Figs 1 and 2) for all the events associated with fin  
172 development. All specimens were reared and used for a previous experiment [39], for  
173 which protocols were approved by the Université du Québec à Rimouski's animal care and  
174 use committee.

175

176 **Fig 1. Details of the dorsal and anal fins of the rainbow trout (*Oncorhynchus mykiss*)**  
177 **based on a 31.33 mm long juvenile specimen.** The main morphological features of these  
178 fins are identified. The serial elements, the proximal radials (PR), the distal radials (DR)  
179 and the lepidotrichia (L), are numbered from the anterior to the posterior of the fins. Act,  
180 actinotrichia; an.f, anal fin; bif, bifurcation; caud.f, caudal fin; DR, distal radial; dors.f,  
181 dorsal fin; L, lepidotrichia; PR, proximal radial; pect.f, pectoral fin; pelv.f, pelvic fin; seg,  
182 segmentation.

183

184 **Fig 2. Cleared and stained specimens of the rainbow trout (*Oncorhynchus mykiss*).** The  
185 cartilage is stained in red and the cartilages in blue. A: Dorsal fin showing the beginning of  
186 lepidotrichia ossification (specimen SL = 28.36 mm); B: Anal fin showing the beginning of  
187 lepidotrichia ossification (specimen SL = 19.80 mm); C: Dorsal fin showing the beginning  
188 of radial ossification (specimen SL = 24.90 mm); D: Anal fin showing the beginning of radial  
189 ossification (specimen SL = 24.90 mm). Arrows point anteriorly. L, lepidotrichia; PR,  
190 proximal radial; seg, segmentation.

191

192 Observations were made under a Leica MZ16A binocular mounted with a digital  
193 camera. Standard length (SL) was measured prior to staining with Northern Eclipse  
194 Software (Version 6.0). Since SL and dph are highly correlated ( $r^2 = 0.952$ ;  $P < 0.001$ ) and  
195 SL is recognized as a better proxy for morphological development in fishes [42], SL was  
196 used for all statistical analyses.

197 Event coding was based on colour uptake by skeletal elements. In this study, three  
198 states are recognized for radials: (1) present (mesenchymal cell condensations without stain  
199 uptake), (2) cartilaginous (blue), and (3) ossified (red). Four developmental states are  
200 recognized for lepidotrichia: (1) present (collagenous matrix), (2) ossified (red), (3)  
201 segmented (number of segments per lepidotrichium), and (4) bifurcated (position of the  
202 bifurcation along the lepidotrichia). Surveyed specimens with their size (SL) and event  
203 coding of the skeletal elements are listed in S1 File.

204 To manage with the inter-individual variation in the number of radials and  
205 lepidotrichia, positional homologies and numbering of elements were inferred *a posteriori*  
206 by lining up all specimens with the third radial (variation being more important in  
207 peripheral areas) and by comparing similarities among sequences of similar-sized  
208 specimens. Myomere counts (from cranial to caudal) were used in the earliest stages as a  
209 topographical criterion to identify the first proximal radials to differentiate. The dorsal and  
210 anal fins are positioned at the level of myomeres 21-32 and 40-50, respectively.

211 Logistic regressions were used to estimate the SL at which 50% (SL<sub>50</sub>) of the  
212 specimens have reached a given developmental state (i.e., present, cartilaginous, ossified,  
213 segmented and bifurcated) for each skeletal element (see [30] for further details).

214 Significance of the logistic regressions were tested using the Likelihood ratio statistic [43].  
215 To interpret a regression for a given element, the significance level was calculated using the  
216 Bonferroni correction; the collective significance level of 0.05 was divided by the number  
217 of elements to get the nominal significance level for each regression. Statistical analyses  
218 were performed with R Studio for Windows v. 1.3.1093 (library: MASS R; [44]).

219 In order to investigate the median fins patterning and developmental similarity, the  
220  $SL_{50}$  values of each skeletal element (i.e., derived from the logistic regressions) were used  
221 to order the serial elements in relative developmental sequences within a fin. The relative  
222 order of a skeletal element within a developmental sequence was then converted by  
223 attributing a rank value. Spearman rank correlation coefficients were then used to describe  
224 the relations between the developmental sequences in the dorsal and anal fins, and in the  
225 endoskeleton and exoskeleton (see [30,45] for the detailed procedure). Only the elements  
226 for which the logistic model was significant (under the nominal significance level) were  
227 considered for Spearman correlations. Logistic regressions do not produce  $SL_{50}$  when the  
228 elements are present in all specimens. The actual sizes of the smallest specimens were  
229 included in the Spearman correlations involving the apparition of the proximal radials in  
230 the dorsal and anal fins, since few skeletal elements were already present in these  
231 specimens. These already present proximal radials are ranked as the first appeared in the  
232 developmental sequence for the Spearman correlations.

### 233 **Fossil material**

234 The phylogenetic sampling includes six Palaeozoic (Devonian-Carboniferous)  
235 species of osteichthyans comprising a ‘palaeonisciform’ actinopterygian (*Elonichthys*

236 *peltigerus*) and five sarcopterygians including coelacanths (*Miguashaia bureaui* and  
237 *Rhabdoderma exiguum*), lungfishes (*Dipterus valenciennesi*), porolepiforms (*Quebecius*  
238 *quebecensis*) and ‘osteolepiforms’ (*Eusthenopteron foordi*). Specimens were chosen  
239 according to their exceptional state of preservation (articulated postcranial material and  
240 undistorted fins) and, whenever possible, availability of ontogenetic series. Specimens of *E.*  
241 *peltigerus* and *R. exiguum* come from the Upper Carboniferous (middle Pennsylvanian)  
242 Francis Creek Shale (Mazon Creek area, Illinois) and studied specimens are housed in the  
243 Field Museum of Natural History (FMNH; Chicago, IL, USA). Specimens of *M. bureaui*,  
244 *Q. quebecensis* and *E. foordi* come from the Upper Devonian (middle Frasnian) Escuminac  
245 Formation (Miguasha, Quebec, Canada); studied specimens are housed in the Musée  
246 d’Histoire Naturelle de Miguasha (MHNM, parc national de Miguasha, Quebec, Canada),  
247 the American Museum of Natural History (AMNH, New York, NY, USA), the Musée de  
248 géologie René-Bureau from Université Laval (ULQ, Quebec, Canada), and The University  
249 of Kansas Biodiversity Institute and Museum of Natural History, Division of Vertebrate  
250 Paleontology (KUVVP, Lawrence, KS, USA). Finally, specimens of *D. valenciennesi* come  
251 from the Middle Devonian (Givetian) Achanarras beds (Scotland, UK); studied specimens  
252 are housed in the Natural History Museum (BMNH, London, UK). A complete list of  
253 surveyed specimens with their size [SL or total length (TL)] are listed in S1 Table.

254 Fossil specimens were examined under a Leica MZ9.5 binocular equipped with a  
255 drawing tube and were photographed with an Olympus Camedia C5060. Developmental  
256 states include (1) ossification (presence; radials and lepidotrichia), (2) segmentation  
257 (lepidotrichia), and (3) bifurcation (lepidotrichia).

258

259

## 260 **Results**

### 261 **Developmental patterning of the dorsal and anal fins of**

#### 262 *Oncorhynchus*

263           The dorsal and anal fins of juvenile *Oncorhynchus* are similar in their anatomy and  
264 shape but differ slightly in meristic counts of skeletal elements and fin size (Fig 1; Table 1).  
265 Generally, the dorsal and anal fins are composed of 29 (i.e., 15 proximal and 14 distal  
266 radials) and 25 (i.e., 13 proximal and 12 distal radials) endoskeletal elements, and 18 and  
267 15 exoskeletal elements (i.e., lepidotrichia), respectively. In both fins, proximal radials  
268 (PR), distal radials (DR), and lepidotrichia (L) are organized in a one-to-one relationship,  
269 with the exception of the first proximal and distal radials that support four and three  
270 lepidotrichia in the dorsal and anal fins, respectively, while the last proximal and distal  
271 radials support two lepidotrichia in both fins. Nine of the eleven developmental events  
272 previously described (see Nomenclature) were analysed in *Oncorhynchus*; the relative  
273 developmental sequence in each fin is ordered based on SL<sub>50</sub> (Table 2). The presence of  
274 actinotrichia (event 1) was not analysed except for its initial position in the sequence of  
275 events. The ossification of distal radials (event 11) occurs after 100 dph, beyond the  
276 timeframe of our study thus no data were available.

277

278 **Table 1. Meristic counts for the skeletal elements, proximal radials (PR), distal radials**  
279 **(DR) and lepidotrichia (L) and proportions of the dorsal (D) and anal (A) fins in living**  
280 **and extinct osteichthyans.**

281

Taxa	Dorsal fin			Anal fin			Proportion
	PR	DR	L	PR	DR	L	
<i>Oncorhynchus</i>	12-15	12-14	15-18	11-13	11-13	12-16	D > A
<i>Elonichthys</i>	5-12	1-17	34-40	3-9	1-17	40-47	D < A
<i>Miguashaia</i>	1	NA	21-28	1	NA	19-25	D = A
<i>Rhabdoderma</i>	?	?	12-21	?	?	9-21	D = A
<i>Quebecius</i>	?	?	30-35	?	?	31-36	D = A
<i>Dipterus</i>	1 <sup>1</sup>	5 <sup>1</sup>	42-48	1 <sup>1</sup>	4 <sup>1</sup>	22-28	D > A
<i>Eusthenopteron</i>	1	3	19-26	1	3	20-25	D = A

282 NA, non applicable

283 ?, endoskeletal elements are unknown

284 <sup>1</sup> Following Ahlberg and Trewin [46]; there is more than one row of distal radials; data are

285 for one row.

286  
 287 **Table 2. Values of the SL<sub>50</sub> for the serial skeletal elements, i.e. the proximal radials (PR), the distal radials (DR) and**  
 288 **the lepidotrichia (L) during nine out of eleven developmental events of the developmental sequence of the dorsal and**  
 289 **anal fins of *Oncorhynchus*.** See Nomenclature for the complete list of events. The serial skeletal elements are ordered from  
 290 the anterior to the posterior of the fins and their column is representative of their position in the fin (see Fig 1). A PR or DR is  
 291 found in more than one column when more than one L is articulated with it. The sites of initiation of the different events are  
 292 identified with a grey shading.

Events	Serial elements (SL <sub>50</sub> mm)																	
	Dorsal fin																	
<b>2. Apparition of PR<sup>1</sup></b>	PR1	PR1	PR1 (15.249)	PR2 (12.692)	PR3 (10.478)	PR4 (8.608 <sup>ns</sup> )	PR5 (NA)	PR6 (NA)	PR7 (NA)	PR8 (NA)	PR9 (NA)	PR10 (9.362)	PR11 (9.362)	PR12 (9.362)	PR13 (11.553)	PR14 (12.573)	PR15 (12.080 <sup>n</sup> <sub>s</sub> )	PR15
<b>3. Chondrification of PR</b>	PR1	PR1	PR1 (19.422)	PR2 (15.613)	PR3 (13.490)	PR4 (12.608)	PR5 (12.608)	PR6 (12.608)	PR7 (12.608)	PR8 (12.608)	PR9 (12.608)	PR10 (12.890)	PR11 (13.390)	PR12 (14.580)	PR13 (15.128)	PR14 (17.288)	PR15 (16.993 <sup>n</sup> <sub>s</sub> )	PR15

<b>4. Apparition of L</b>	L01 (18.315)	L02 (16.908)	L03 (15.488)	L04 (14.579)	L05 (13.949)	L06 (13.722)	L07 (13.285)	L08 (13.285)	L09 (13.285)	L10 (13.285)	L11 (13.285)	L12 (13.722)	L13 (14.561)	L14 (14.932)	L15 (15.191)	L16 (16.030)	L17 (18.085)	L18 (16.962 <sup>n</sup> <sub>s</sub> )
<b>5. Apparition of DR</b>				DR1 (23.756)	DR2 (16.045)	DR3 (15.535)	DR4 (15.059)	DR5 (15.059)	DR6 (14.815)	DR7 (14.815)	DR8 (14.815)	DR9 (14.815)	DR10 (15.308)	DR11 (16.058)	DR12 (16.564)	DR13 (16.901)	DR14 (17.332)	DR15 (17.332)
<b>6. Chondrification of DR<sub>2</sub></b>				DR1 (34.430 <sup>n</sup> <sub>s</sub> )	DR2 (18.541)	DR3 (17.142)	DR4 (17.002)	DR5 (16.744)	DR6 (16.947)	DR7 (17.489)	DR8 (17.489)	DR9 (17.489)	DR10 (17.786)	DR11 (18.459)	DR12 (18.459)	DR13 (18.601)	DR14 (18.601)	DR15 (18.848)
<b>7. Segmentation of L<sup>3</sup></b>	L01 (NA)	L02 (42.618)	L03 (39.910)	L04 (25.307)	L05 (19.511)	L06 (18.160)	L07 (17.782)	L08 (17.555)	L09 (17.671)	L10 (17.671)	L11 (17.671)	L12 (17.173)	L13 (17.896)	L14 (17.947)	L15 (17.947)	L16 (17.927)	L17 (18.792)	L18 (16.962 <sup>n</sup> <sub>s</sub> )
<b>8. Ossification of L</b>	L01 (26.217)	L02 (22.830)	L03 (21.657)	L04 (19.913)	L05 (19.509)	L06 (19.374)	L07 (19.374)	L08 (19.374)	L09 (19.777)	L10 (19.913)	L11 (19.913)	L12 (20.696)	L13 (21.856)	L14 (22.920)	L15 (24.634)	L16 (27.151)	L17 (26.971 <sup>n</sup> <sub>s</sub> )	L18 (NA)
<b>9.</b>	L01	L02	L03	L04	L05	L06	L07	L08	L09	L10	L11	L12	L13	L14	L15	L16	L17	L18



<b>Bifurcation of L</b>								(31.085)	(28.338)	(28.119)	(24.798)	(27.177)	(24.067)	(23.598)	(27.883)	(30.672)	(32.704 <sup>ns</sup> )	(NA)
<b>10. Ossification of PR</b>	PR1	PR1	PR1 (30.096)	PR2 (27.485)	PR3 (25.880)	PR4 (25.520)	PR5 (25.894)	PR6 (26.254)	PR7 (29.754)	PR8 (30.937)	PR9 (31.206)	PR10 (31.206)	PR11 (32.016)	PR12 (34.553 <sup>ns</sup> )	PR13 (34.607 <sup>ns</sup> )	PR14 (34.945 <sup>ns</sup> )	PR15 (32.704 <sup>ns</sup> )	PR16 (NA)

**Ana  
lfin**

<b>2. Apparition of PR<sup>1</sup></b>	PR1	PR1	PR1 (15.066)	PR2 (13.290)	PR3 (12.309)	PR4 (10.507 <sup>ns</sup> )	PR5 (10.507 <sup>ns</sup> )	PR6 (NA)	PR7 (NA)	PR8 (NA)	PR9 (7.465 <sup>ns</sup> )	PR10 (7.462 <sup>ns</sup> )	PR11 (12.484)	PR12 (13.997)	PR13 (15.353 <sup>ns</sup> )	PR14 (NA)	PR15 (NA)	PR16 (NA)	PR17 (NA)
<b>3. Chondri- fication of PR</b>	PR1	PR1	PR1 (19.674)	PR2 (16.550)	PR3 (14.463)	PR4 (13.569)	PR5 (13.569)	PR6 (12.852)	PR7 (12.852)	PR8 (12.846)	PR9 (14.159)	PR10 (15.016)	PR11 (16.395)	PR12 (17.488)	PR13 (18.591)	PR14 (NA)	PR15 (NA)	PR16 (NA)	PR17 (NA)
<b>4. Apparition of L</b>	L01 (19.135)	L02 (17.783)	L03 (15.523)	L04 (14.405)	L05 (13.138)	L06 (12.884)	L07 (12.884)	L08 (12.884)	L09 (12.884)	L10 (13.451)	L11 (14.059)	L12 (15.084)	L13 (16.331)	L14 (16.870)	L15 (17.184)	L16 (17.257 <sup>ns</sup> )	L17 (NA)	L18 (NA)	L19 (NA)
<b>5.</b>				DR1	DR2	DR3	DR4	DR5	DR6	DR7	DR8	DR9	DR10	DR11	DR12	DR13	DR14	DR15	DR16

<b>Apparition of DR</b>				(40.547 <sup>n</sup> <sub>s</sub> )	(16.698)	(15.485)	(15.762)	(15.762)	6(15.762)	(15.762)	(16.057)	(16.057)	0(16.057)	1(16.325)	2(17.347)	3(18.404)	3	
<b>6. Chondri-fication of DR<sup>2</sup></b>				DR1(40.917 <sup>n</sup> <sub>s</sub> )	DR2(18.705)	DR3(18.195)	DR4(18.195)	DR5(18.195)	DR(6(18.440))	DR7(18.440)	DR8(18.597)	DR9(18.597)	DR1(0(18.751))	DR1(1(18.865))	DR1(2(19.209))	DR1(3(19.830))	DR1(3)	
<b>7. Segmentation of L<sup>3</sup></b>	L01(NA)	L02(NA)	L03(35.789 <sup>n</sup> <sub>s</sub> )	L04(20.827)	L05(17.996)	L06(17.996)	L07(17.671)	L08(17.671)	L09(17.671)	L10(17.671)	L11(17.844)	L12(17.996)	L13(18.143)	L14(18.235)	L15(18.645)	L16(19.170 <sup>n</sup> <sub>s</sub> )	L17(NA)	
<b>8. Ossification of L</b>	L01(28.357)	L02(24.287)	L03(20.417)	L04(19.938)	L05(19.515)	L06(19.515)	L07(19.516)	L08(19.646)	L09(19.646)	L10(20.168)	L11(20.168)	L12(20.988)	L13(21.697)	L14(22.650)	L15(23.927)	L16(25.310 <sup>n</sup> <sub>s</sub> )	L17(30.071 <sup>n</sup> <sub>s</sub> )	
<b>9. Bifurcation of L</b>	L01	L02	L03	L04	L05	L06(35.399 <sup>n</sup> <sub>s</sub> )	L07(29.525)	L08(26.506)	L09(25.230)	L10(25.174)	L11(26.800)	L12(26.147)	L13(26.286)	L14(27.036)	L15(29.923)	L16(43.001 <sup>n</sup> <sub>s</sub> )	L17(30.071 <sup>n</sup> <sub>s</sub> )	
<b>10. Ossification</b>	PR1	PR1	PR1	PR2(30.483)	PR3(29.349)	PR4(28.255)	PR5(28.635)	PR6(29.680)	PR7(29.680)	PR8(32.897)	PR9(33.131)	PR1(0)	PR1(1)	PR1(2)	PR1(3)	PR1(4)	PR1(4)	

<b>ion of PR</b>			(37. 937 <sup>n</sup> s)								(32. 834	(33. 117)	(32. 640)	(46. 057 <sup>n</sup> s)	(NA )		
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293 NA, non applicable

294 <sup>ns</sup>, these SL<sub>50</sub> results are not significant

295 <sup>1</sup> Few proximal radials of the dorsal and anal fins were present in the smallest specimens (8.650 mm and 10.894 mm in SL,  
296 respectively).

297 <sup>2</sup> Events 6 and 7 are inverted in the anal fin, i.e. lepidotrichia segmentation (event 7) occurs prior to distal radial  
298 chondrification (event 6).

299 <sup>3</sup> Event 7 corresponds to the overall initiation of segmentation; the following events of segmentation (up to 6 events of  
300 segmentation; data not shown) are not included in the series of 11 events.

301           The apparition of proximal radials (event 2) begins before hatching. Few proximal  
302 radials, located centrally, are present in the dorsal and anal fins of all the specimens  
303 examined, even in the smallest pre-hatching specimens (8-10-mm SL), thus, no SL<sub>50</sub> were  
304 obtained for these elements (Table 2; Fig 3A). These proximal radials are interpreted as the  
305 initiation sites of the development of the dorsal (PR 5-9) and anal (PR 6-8) fin  
306 endoskeletons. These sites are congruent in the dorsal and anal fins (Table 2; Fig 3A). In  
307 both fins, proximal radials appear by proceeding bilaterally from the initiation site (Table  
308 3). This indicates a bidirectional pattern for the development of the proximal radials; the  
309 most peripherally located proximal radials are the last to appear. The relative  
310 developmental sequences are not simultaneous but are significantly correlated between fins  
311 (Table 4). All following events occur after hatching.

312

313 **Fig 3. Comparisons between nine out of eleven developmental sequences for the dorsal**  
314 **(black) and anal (gray) fins of the rainbow trout (*Oncorhynchus mykiss*).** See

315 Nomenclature for the complete list of events. Skeletal elements are ordered from anterior to  
316 posterior. Filled and empty symbols represent significant and non-significant results for

317 SL<sub>50</sub>, respectively.

318

319 **Table 3. Directions of developmental sequences of ten out of eleven events for the**  
320 **dorsal (D) and anal (A) fins in living and extinct osteichthyans.** See Nomenclature for

321 the complete list of events.

322

Events	<i>Oncorhynchus</i>		<i>Elonichthys</i>		<i>Miguashaia</i>		<i>Rhabdoderma</i>		<i>Quebecius</i>		<i>Dipterus</i>		<i>Eusthenopteron</i>	
	D	A	D	A	D	A	D	A	D	A	D	A	D	A
2. Apparition of proximal radials	B	B	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	NA 1	NA 1	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	NA 1	NA 1
3. Chondrification of proximal radials	B	B	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	NA 1	NA 1	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	NA 1	NA 1
4. Apparition of lepidotrichia	B	B	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	NA 1	NA 1	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	NA 1	NA 1
5. Apparition of distal radials	B	B	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	NA 1	NA 1	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	NA 1	NA 1
6. Chondrification of distal radials	B	B	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	NA 1	NA 1	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	N A <sup>1</sup>	NA 1	NA 1
7. Segmentation of lepidotrichia	B	B	B	B	B	B	B	B	?	?	B	B	B	B
8. Ossification of lepidotrichia	B	B	?	?	?	?	B	B	?	?	?	?	?	?
9. Bifurcation of lepidotrichia	B	B	N A <sup>2</sup>	N A <sup>2</sup>	B	B	NA 2	NA 2	B	B	B	B	B	B

10. Ossification of proximal radials	B	B	?	?	N A <sup>3</sup>	N A <sup>3</sup>	?	?	?	?	?	?	NA <sub>3</sub>	NA <sub>3</sub>
11. Ossification of distal radials	NA <sup>4</sup>	NA <sup>4</sup>	B-A P <sup>5</sup>	B-A P <sup>5</sup>	N A <sup>3</sup>	N A <sup>3</sup>	?	?	?	?	?	?	PA	PA

323 B, Bidirectional; AP, Antero-posterior; PA, Postero-anterior

324 ?, the skeletal elements and/or sequence are unknown

325 NA: non-applicable

326 <sup>1</sup> This event is not observable in fossil taxa

327 <sup>2</sup> Bifurcation is unknown

328 <sup>3</sup> Only one endoskeletal element (for *Miguashaia*, the condition is unknown for the anal  
329 fin, however, the dorsal fin has at least three distal radial and one large basal plate)

330 <sup>4</sup> The ossification occurs after 100 dph.

331 <sup>5</sup> Data suggest an antero-posterior direction but a bidirectional direction is possible (see the  
332 text)

333

334

335 **Table 4. Spearman correlations between developmental sequences of (a) the dorsal**  
336 **and anal fins and (b) fin exoskeleton and endoskeleton of the dorsal (D) and anal (A)**  
337 **fins of *O. mykiss* to validate the presence of the DAFPM and the EEM respectively.**

338 See Nomenclature for the complete list of events.

<b>Events</b>	<b>n</b>	<b><math>r_s</math></b>	<b><math>P</math></b>
<b>Dorsal and anal fin patterning module</b>			
2. Proximal radial differentiation	8	0.8553628	<0.01
3. Proximal radial chondrification	13	0.9185589	<0.001
4. Lepidotrichia differentiation	15	0.8604663	<0.001
5. Distal radial differentiation	12	0.6288943	<0.05
6. Distal radial chondrification	12	0.9019622	<0.001
7. Lepidotrichia segmentation	12	0.6691664	<0.05
8. Lepidotrichia ossification	15	0.9576292	<0.001
9. Lepidotrichia bifurcation	8	-0.40476	ns
10. Proximal radial ossification	10	0.9268293	<0.001
11. Distal radial ossification	NA	NA	NA
<b>Endoskeleton and exoskeleton module</b>			
Events 2 and 4: Proximal radial and lepidotrichia differentiation (D)	14	0.903	< 0.001
Events 2 and 4: Proximal radial and lepidotrichia differentiation (A)	12	0.730	< 0.01
Events 4 and 5 Lepidotrichia and distal radial differentiation (D)	14	0.972	< 0.001
Events 4 and 5 Lepidotrichia and distal radial differentiation (A)	12	0.891	< 0.001

Events 8 and 10 Lepidotrichia and proximal radial ossification (D)	11	0.854	< 0.001
Events 8 and 10 Lepidotrichia and proximal radial ossification (A)	11	0.839	< 0.01

339 NA, non applicable: The ossification of distal radials occurs after 100 dph

340 ns, non significant

341

342 The chondrification of the proximal radials (event 3) quickly follows their  
343 appearance. In both fins, the chondrification sequences show similar patterns as to their  
344 sequences of apparition (S1 Table); chondrification starts from the same initiation site (i.e.,  
345 the most centrally located proximal radials) and further proceeds bidirectionally (Table 3).  
346 The bidirectional sequences of chondrification are significantly correlated and are almost  
347 simultaneous between dorsal (PR 4-9) and anal (PR 8) fins (Fig 3B, Table 4).

348 Lepidotrichia appear (event 4) slightly after the differentiation of the first proximal  
349 radial and subsequently articulate with the distal radials. The initiation site of the  
350 lepidotrichia is similar in both fins (L7-11 in the dorsal fin and L6-9 in the anal fin) and  
351 corresponds in position to the early apparition of the proximal radials (S1 Table). Similar to  
352 proximal radials, new lepidotrichia appear by proceeding bidirectionally from the initiation  
353 site (Table 2). The sequences of apparition of lepidotrichia are highly correlated between  
354 fins and almost simultaneous (Fig 3C, Table 4).

355 The first distal radials (event 5) to appear differ in position between the dorsal  
356 (DR6-9) and anal (DR3) fins (Table 2). In both fins, the sequences of apparition proceed



357 bidirectionally from the initiation site. The sequences are significantly correlated (Fig 3D,  
358 Table 4).

359 The chondrification of distal radials (event 6) follows rapidly, starting from similar  
360 initiation sites, DR5 in the dorsal fin and DR3-5 in the anal fin (Table 2). Starting from the  
361 initiation site, the chondrification sequences proceed bidirectionally and are almost  
362 simultaneous as well as highly correlated between fins (Fig 3E, Table 4).

363 Lepidotrichia grow by distal addition of new segments, a process called  
364 segmentation (event 7). Through growth, five and six segmentation events occur in the anal  
365 and dorsal fins, respectively. The lepidotrichia with the higher numbers of segments in  
366 most specimens are the L10-14 in the dorsal fin and L8-10 in the anal fin and the number of  
367 segments on the adjacent lepidotrichia decreases in a bidirectional pattern. The first  
368 segmentation event occurs slightly posteriorly in the dorsal fin (L12) comparatively to the  
369 anal fin (L7-10) (Fig 3F). The site for the initiation of segmentation corresponds to the  
370 lepidotrichia with the highest number of segments, suggesting that the longest lepidotrichia,  
371 in terms of number of segments, are the first lepidotrichia to segment in the developmental  
372 sequence (Fig 1). All sequences of segmentation are bidirectional (Table 3). The sequences  
373 of the first segmentation are almost simultaneous and significantly correlated between fins  
374 (Fig 3F, Table 4).

375 The initiation site for the ossification of lepidotrichia (event 8) is similar in both fins  
376 (L6-8 in the dorsal fin and L5-6 in the anal fin) (Fig 3G). Sequences of ossification are  
377 bidirectional, simultaneous, and highly correlated (Tables 3 and 4). There are small,  
378 positional differences for the initiation site of apparition, segmentation and ossification of

379 the lepidotrichia (Fig 3G). Nevertheless, sequences of apparition are significantly correlated  
380 with sequences of segmentation of the lepidotrichia in the dorsal (0.670;  $p < 0.01$ ) and anal  
381 (0.768;  $p < 0.01$ ) fins, while the results differ between sequences of segmentation and  
382 ossification of the lepidotrichia (dorsal, 0.098;  $p > 0.05$ ; anal, 0.485;  $p > 0.05$ ) (Table 4).

383 The bifurcation of the lepidotrichia (event 9) is initiated at different positions in the  
384 dorsal (L14) and anal (L10) fins (Fig 3H). The bidirectional sequences of bifurcation  
385 between fins are not significantly correlated and not simultaneous (Fig 3H, Table 4).  
386 Generally, in the dorsal fin, the lepidotrichia located at the initiation site show more  
387 proximal bifurcations comparatively to the lepidotrichia located bilaterally of the initiation  
388 site; this pattern is not as clear in the anal fin. A single order of bifurcation is present.

389 The ossification of proximal radials (event 10) is initiated from PR4, located in the  
390 same anterior portion of both fins (Fig 3I). In both fins, the sequences of ossification  
391 proceed in a bidirectional direction, and are highly correlated, but not simultaneous (Fig 3I,  
392 Table 4).

393 All nine developmental events analysed show a certain degree of congruence  
394 between the dorsal and anal fins (e.g., similar initiation site and/or similar direction) and  
395 eight events are significantly correlated between fins (events 2-8 and 10; Table 4).  
396 Moreover, in congruence with the one-to-one relationship observed between radials and  
397 lepidotrichia in both fins, the first lepidotrichia to appear are related to the first apparition  
398 of proximal and distal radials (Fig 4); this is corroborated by the strong correlations  
399 between sequences of apparition of the endoskeletal and exoskeletal elements within each

400 fin (Table 4). Highly significant correlations are also found for sequences of ossification of  
401 the endoskeleton and exoskeleton within both fins (Fig 4, Table 4).

402

403 **Fig 4. Comparisons between developmental sequences (apparition and ossification) of**  
404 **the endoskeleton (radials; black) and exoskeleton (lepidotrichia; grey) of the dorsal**  
405 **(A, C, E) and anal fins (B, D, F) of the rainbow trout (*Oncorhynchus mykiss*).** Skeletal  
406 elements are ordered from anterior to posterior. Filled and empty symbols represent  
407 significant and non-significant results for SL50 respectively.

408

## 409 **Developmental patterning in the dorsal and anal fins of fossil** 410 **osteichthyans**

411 For the six fossil osteichthyans surveyed (Table 1, S1 Table), only the events  
412 dealing with the lepidotrichia [i.e., segmentation (event 7), ossification (event 8), and  
413 bifurcation (event 9)] and the ossification of endochondral elements [i.e., proximal radials  
414 (event 10) and distal radials (event 11)] are available to study due to the nature of fossils  
415 and the rarity of fossilized ontogenies. Meristic counts (Table 1) are based on mean values  
416 from all specimens examined for each taxon, including both juveniles and adults whenever  
417 possible. Congruence of events between the dorsal and anal fins, and between the  
418 endoskeleton and exoskeleton are inferred based on initiation sites, and when available, the  
419 direction of sequences.

420

421 **Actinopterygians**

422 ***Elonichthys peltigerus***

423 The dorsal and anal fins of *Elonichthys* are of similar shape but differ slightly in size (Table  
424 1; Fig 5) [47]. The dorsal fin is composed of 16-19 distal radials whereas the anal fin has  
425 19-23 proximal radials. The general relationship between radials and lepidotrichia is 1:2,  
426 with some supernumerary lepidotrichia occurring in the anterior and posterior margins.

427

428 **Fig 5. The Carboniferous actinopterygian *Elonichthys peltigerus*.** A, Fossil specimen  
429 (FMNH PF 7502) and drawings of its dorsal (B) and anal (C) fins. Arrows point anteriorly.  
430 An. f, anal fin; caud. f, caudal fin; Dors. f, dorsal fin; DR, distal radial; L, lepidotrichia; Pect.  
431 f, pectoral fin; Pelv. f, pelvic fin; PR, proximal radial; seg, segmentation.

432

433 Lepidotrichia are the first structures to ossify in both fins. Segmented lepidotrichia  
434 (event 7) are present in all specimens. The number of segments varies little with respect to  
435 SL (from 4-5 to 6-7 segments per lepidotrichium). In both fins, lepidotrichia from the  
436 anterior part of the fin are usually the longest (L7-15 in the dorsal and L10-13 in the anal  
437 fin) and articulate with distal radials 5-6, which seem among the first ones to ossify (see  
438 below). The number of segments decreases bilaterally from these lepidotrichia.

439 Lepidotrichia are already numerous in the smallest specimen (38 dorsal and 46 anal  
440 lepidotrichia); therefore, it is not possible to infer a sequence of ossification (event 8). None  
441 of the specimens show bifurcated lepidotrichia (event 9).

442 Ossification sequences are difficult to reconstruct for the proximal (event 10) and  
443 distal radials (event 11). Proximal radials are visible at the anterior portion of the dorsal and  
444 anal fins; remaining radials are probably hidden under the scale cover. Distal radials have  
445 only been clearly identified in the dorsal fin (FMNH PF 7502) (Fig 5). When present, both  
446 ossified proximal and distal radials do not reach the posterior margin of both fins.

447

## 448 **Actinistians**

### 449 *Miguashaia bureaui*

450 The second dorsal and anal fins of *Miguashaia* are overly similar in size and shape  
451 (Table 1; Figs 6 and 7) with narrow, rectangular bases and pointed anterior corners ([22],  
452 fig. 1B). Only the lepidotrichia can be described (27-28 in the dorsal fin and ca. 25 in the  
453 anal fin), a single specimen (MHNM 06-1809) partially shows the radials articulating with  
454 the second dorsal basal plate.

455

456 **Fig 6. The Devonian coelacanth *Miguashaia bureaui*.** A, Juvenile specimen (ULQ 120)  
457 and drawings of its second dorsal (B) and anal (C) fins. Arrows point anteriorly. An. f, anal  
458 fin; bif, bifurcation; Caud. f, caudal fin; Dors. f, dorsal fin; L, lepidotrichia; seg,  
459 segmentation.

460

461 **Fig 7. The Devonian coelacanth *Miguashaia bureaui*.** A, Adult specimen (MHNM 06-41)  
462 and drawings of its second dorsal (B) and anal (C) fins. Note the occurrence of merging  
463 segments (m.seg) at the base of the fins. Arrows point anteriorly. An. f, anal fin; bif,

464 bifurcation; Caud. f, caudal fin; Dors. f, dorsal fin; L, lepidotrichia; Pect. f, pectoral fin; Pelv.  
465 f, pelvic fin; PR, proximal radial; seg, segmentation.

466

467 Numerous segmentations (up to five) are visible in the lepidotrichia (event 7) of the  
468 dorsal and anal fins of the smallest specimen (MHNM 06-1633, 64.5 mm TL). The longest  
469 lepidotrichia are L4-7 in the dorsal fin and L5-8 in the anal fin in specimen MHNM 06-41  
470 (Fig 7) and the number of segments decreases gradually bilaterally from these sites. The  
471 basal proximal segment is always longer than the others in all specimens examined. In  
472 specimen MHNM 06-41, the segment immediately distal to the first proximal segment  
473 appears to be half-fused with this basal segment (Fig 7; L11, m.seg), thus evidencing that  
474 the increase in length of the basal segment during growth is the result of its merging with  
475 other proximal segments.

476 Ossified lepidotrichia are already numerous (28 dorsal and ca. 22 anal lepidotrichia)  
477 in the smallest specimen (MHNM 06-1633); it is thus not possible to infer an ossification  
478 sequence (event 8).

479 Few bifurcations (event 9) are seen on L15 and L14-17, respectively in the dorsal  
480 and anal fins of the small specimen ULQ 120 (85 mm TL; Fig 6). The most proximal  
481 bifurcations are positioned approximately between L12-15 in all specimens (Figs 6 and 7).  
482 Bifurcations are present anteriorly and posteriorly to L12-15 in large specimens.  
483 Bifurcations are gradually displaced distally anterior to the most proximally bifurcated  
484 lepidotrichium (L15) resulting in a higher proportion of proximally bifurcated lepidotrichia

485 in the posterior portion of the fins. Up to three orders of bifurcation have been observed in  
486 specimen MHNM 06-494 (400 mm TL).

487 Ossified endoskeletal supports (events 10 and 11) are poorly documented. A single  
488 basal plate in the first dorsal fin is present in one partial specimen (MHNM 06-1232), while  
489 the basal plate of the second dorsal fin shows articulating surfaces for three radials, the  
490 posterior one being well-preserved (MHNM 06-1809).

491

492 ***Rhabdoderma exiguum***

493 The second dorsal and anal fins of *Rhabdoderma* are almost identical in size and  
494 shape (Table 1, Fig 8). They display a narrow-based fan-like outline and, as for  
495 *Miguashaia*, only the lepidotrichia are preserved.

496

497 **Fig 8. The Carboniferous coelacanth *Rhabdoderma exiguum*.** A, Small specimen (FMNH  
498 PF 9954) and drawings of its second dorsal (B) and anal (C) fins. Arrows point anteriorly.  
499 An. f, anal fin; Caud. f, caudal fin; Dors. f, dorsal fin; L, lepidotrichia; Pelv. f, pelvic fin; seg,  
500 segmentation.

501

502 All specimens show segmented lepidotrichia (event 7). The number of segments  
503 increases with SL (from 5-6 to 13-14 segments). In both fins, the longest lepidotrichia  
504 occur between L9-12 and the number of segments gradually decreases bilaterally from this  
505 site.

506           The number of lepidotrichia increases up to 21 in both fins during growth (event 8).  
507   Lepidotrichia at the anterior and posterior extremities are absent in the dorsal and anal fins  
508   of small specimens, suggesting that central lepidotrichia might have ossified before the  
509   anterior and posterior ones (Fig 8). This fin comparison between small (less than 21  
510   lepidotrichia; e.g., FMNH PF7528; 33 mm SL) and larger specimens (21 lepidotrichia; e.g.,  
511   FMNH PF7338; 48 mm SL) was done by lining up their respective longest lepidotrichia.

512           None of the specimens show bifurcation of the lepidotrichia (event 9), and no  
513   ossified endoskeletal supports are preserved in embryos and larvae examined (events 10  
514   and 11).

515

## 516   **Porolepiforms**

### 517   *Quebecius quebecensis*

518           The second dorsal and anal fins of *Quebecius* have a similar size and shape (Table  
519   1) [48 (figs 1, 7), 49 (fig. 2)]. Endoskeletal supports remain unknown in this taxon and only  
520   the lepidotrichia can be described (between 30 and 40 lepidotrichia in both fins).

521           Segmented lepidotrichia (event 7) are already visible in the smallest specimen  
522   (MHNM 06-1474a); however, the number of segments is unclear in all specimens owing to  
523   preservation issues. Lepidotrichia are well developed, and their number is similar in the  
524   small as well as in the larger specimens (35 dorsal lepidotrichia and 34 anal lepidotrichia),  
525   it is thus not possible to infer an ossification sequence (event 8).

526           The first bifurcation (event 9) is seen in L22 of the anal fin (MHNM 06-1474a). The  
527   most proximal bifurcations occur between L18-21 in longer specimens, with other



528 branched lepidotrichia located anteriorly and posteriorly to this area. In both fins, the  
529 posterior lepidotrichia display more proximal bifurcations than the anterior lepidotrichia.

530

## 531 **Dipnoans**

### 532 *Dipterus valenciennesi*

533 The second dorsal and anal fins of *Dipterus* differ in size and shape (Table 1; Fig 9), with  
534 the second dorsal fin being longer and higher than the somewhat pointed, leaf-shaped anal  
535 fin. Lepidotrichia are more numerous than their supporting radials, which are known to be  
536 grossly similar between fins with minor differences in terms of the number and shape [46].

537

538 **Fig 9. The Devonian dipnoan *Dipterus valenciennesi*.** A, Fossil specimen of (BMNH  
539 P.22187) and drawings of its second dorsal (B) and anal (C) fins. Note the development of  
540 scales (Sc) covering the proximal portion of the lepidotrichia. Arrows point anteriorly. An.  
541 f, anal fin; bif, bifurcation; Caud. f, caudal fin; Dors. f, dorsal fin; L, lepidotrichia; Pect. f,  
542 pectoral fin; seg, segmentation.

543

544

545 Lepidotrichia show multiple segmentations (event 7). The longest lepidotrichia are  
546 not found at the same position in the dorsal (L16-26) and anal (L8-12) fins; these areas are  
547 interpreted as different initiation sites of segmentation. In both fins, the number of  
548 segments gradually decreases bilaterally from these sites. The basal segment is  
549 considerably long, comprising one third of each lepidotrichium total length.

550 Even the smallest specimens show numerous well ossified lepidotrichia (44 dorsal  
551 and 25 anal), it is thus not possible to infer a sequence of ossification (event 8).

552 Up to three orders of bifurcation (event 9) are present. Bifurcations are present  
553 between L14-18 and L39-42 in the dorsal fin and between L11-12 and L17-22 in the anal  
554 fin. Specimen BMNH P22187 (Fig 9A) is informative concerning the sequence of  
555 bifurcation: (1) two orders of bifurcation are present in the dorsal fin: 1<sup>st</sup> order: L15-42 (bif  
556 1), 2<sup>nd</sup> order: L28-39 (bif 2) (Fig 9B), and three orders in the anal fin: 1<sup>st</sup> order: L10-20 (bif  
557 1); 2<sup>nd</sup> order: L11-18 (bif 2); 3<sup>rd</sup> order: L13-16 (bif 3) (Fig 9C); (2) the most proximal  
558 bifurcations are in L36-37 in the dorsal fin and in L13-16 in the anal fin (Figs 9B and 9C),  
559 and (3) bifurcated lepidotrichia are found anteriorly and posteriorly to these sites. Thus, in  
560 both fins, the posterior-most lepidotrichia show more proximal bifurcations whereas the  
561 anterior-most lepidotrichia are more distally branched.

562 Ossified endoskeletal supports (events 10 and 11) were not observed due to the  
563 scale cover (Sc, Figs 9B and 9C). Their sequence of ossification is unknown.

564

## 565 **'Osteolepiforms'**

### 566 *Eusthenopteron foordi*

567 The second dorsal and anal fins of *Eusthenopteron* are similar in size and shape  
568 (Table 1; Figs 10 and 11), displaying a narrow-based and posteriorly pointed profile [50].

569 The median fins display a broad basal plate on which generally three distal radials  
570 articulate, carrying numerous lepidotrichia (up to 25 in each fin).

571

572 **Fig 10. The Devonian ‘osteolepiform’ *Eusthenopteron foordi*.** A, Very small specimen  
573 (MHNM 06-1754) and a drawing of its anal fin (B). Arrows point anteriorly. An. f, anal fin;  
574 bif, bifurcation; Caud. f, caudal fin; Dors. f, dorsal fin; DR, distal radial; L, lepidotrichia;  
575 Pect. f, pectoral fin; Pelv. f, pelvic fin; seg, segmentation.

576

577

578 **Fig 11. The Devonian ‘osteolepiform’ *Eusthenopteron foordi*.** A, Small specimen (MHNM  
579 06-1769) and drawings of its second dorsal (B) and anal (C) fins. Arrows point anteriorly.  
580 Note the preservation of a basal plate (BPL) and a reduced scale covering (Sc) at the base of  
581 the fins. An. f, anal fin; bif, bifurcation; Caud. f, caudal fin; Dors. f, dorsal fin; DR, distal  
582 radial; L, lepidotrichia; Pect. f, pectoral fin; Pelv. f, pelvic fin; PR, proximal radial; seg,  
583 segmentation.

584

585 The first segmentations (event 7) are seen in L11-14 in the dorsal fin and in L8-13  
586 in the anal fin (MHNM 04-1293p10-Ef1; 40.8 mm SL). The position of these first  
587 segmented lepidotrichia is congruent with the location of the longest lepidotrichia in larger  
588 specimens. In these specimens, most lepidotrichia are segmented and the number of  
589 segments gradually decreases bilaterally from L8-14.

590 Lepidotrichia are the first structures to ossify in both fins (event 8); they are  
591 numerous in the smallest specimen (MHNM 04-1293p9-Ef1, 27.4 mm SL; 18 and 21  
592 lepidotrichia in dorsal and anal fins, respectively) and the highest number of lepidotrichia is

593 reached early (MHNM 06-1754; 49.3 mm SL; Fig 10), thus it is not possible to infer an  
594 ossification sequence.

595 Bifurcations (event 9) are restricted between L6-7 to L25 in large specimens and up  
596 to five orders of bifurcation are present in both fins. First bifurcations are present in L13  
597 and 17 in the dorsal fin and in L18-19 in the anal fin (MHNM 06-1754; Fig 10B) Most  
598 proximal bifurcations and subsequent orders of bifurcation are initiated in this area (e.g.,  
599 MHNM 06-1769: 2<sup>nd</sup> order: L9-17 in the dorsal fin and L13-18 in the anal fin; 3<sup>rd</sup> order:  
600 L15-17 in the anal fin; Figs 11B and 11C).

601 Radial ossification (DR; event 11) occurs prior to the ossification of the basal plate  
602 (event 10) (BPL, Figs 11B and 11C). A postero-anterior pattern of ossification for dorsal  
603 and anal distal radials is suggested because few small specimens (43-56-mm SL) show only  
604 the posterior, or the posterior and central radials in the dorsal or anal fins. The basal plate is  
605 first seen at 88.9-mm SL and 69.6-mm SL in the dorsal and anal fins, respectively.

606

## 607 **Discussion**

608 Our study described a series of 11 skeletogenic events in the formation of the endo-  
609 and exoskeletal components of the dorsal and anal fins of the living actinopterygian  
610 *Oncorhynchus mykiss* between 5 days pre-hatching up to 100 days post-hatching. We also  
611 documented some of these events in one Carboniferous actinopterygian species, and five  
612 Devonian-Carboniferous sarcopterygian species. For the first time, we quantified the  
613 segmentation and bifurcation of lepidotrichia supporting a lepidotrichial patterning module.

614 The similarity and synchronicity of these developmental events between the dorsal and anal  
615 fins support the hypothesis that these two fins form a developmental module.

616

## 617 **Median fins in vertebrates**

618 The fins of vertebrates can be described as membranous lateral outgrowths of the  
619 body walls reinforced internally by elongated elements, which can be of endoskeletal (e.g.,  
620 radial bones) or dermal (e.g., fin rays) origin.

621 Median fins are present in stem vertebrates, such as myllokunmingiids, in the form  
622 of dorsal and ventral skin folds reminiscent of the median larval fin fold observed during  
623 the early ontogeny of more advanced fishes (e.g., [6,9,11,51]). The earliest ‘agnathans’  
624 display well-developed median fins, which include, in most cases, a caudal fin and elongate  
625 dorsal and ventral fins. A separate anal fin has been confidently identified in  
626 petromyzontiforms, anaspids [52,53], the anaspid-like *Euphanerops*, and in certain  
627 thelodonts (e.g., *Loganellia*) [54]. In *Euphanerops*, the anal fins are paired fins [55,56].

628 The presence of an anal fin in hagfishes has been suggested in *Myxinikela* [57], but  
629 it is currently difficult to confirm given the poor fossil record of Myxiniiformes. An anal fin  
630 could be a plesiomorphic characteristic of vertebrates if the ventral fin of *Myxinikela* is  
631 resolved as homologous to the anal fin of anaspids and gnathostomes. However, the  
632 distribution of the anal fin is variable among vertebrates [11]. The differentiation of an anal  
633 fin from a ventral fin fold might have occurred early in vertebrate history since anal fins  
634 supported by endoskeletal elements have been identified in fossil lampreys (e.g.,  
635 *Hardistiella* and possibly *Mayomyzon*), despite its absence in extant forms (e.g.,

636 *Petromyzon, Lampreta*). In these, an anal ‘crest’ made of a skin fold devoid of fin rays  
637 occurs, but an anal fin may develop in certain atavistic specimens [54,58]. There is no  
638 evidence of an anal fin in heterostracans [12] or arandaspids [59] so its absence can be  
639 considered a derived loss from the primitive condition of fossil lampreys. Janvier (2007;  
640 [54]) suggested that the ‘horizontal caudal lobe’ of osteostracans might be a modified anal  
641 fin. Galeaspids and pituriaspids appear to lack an anal fin [60,61] but uncertainty is due to  
642 the poor preservation of their postcranial skeleton. In gnathostomes, the anal fin is not  
643 preserved usually in ‘placoderms’, probably due to its reduced size and the cartilaginous  
644 nature of the radials, but has been illustrated in some cases (e.g., arthrodiros like  
645 *Africanaspis* and *Dunkleosteus*) [62,63]). The anal fin is primitively present in all crown  
646 gnathostomes but may be absent in certain chondrichthyans [9,11]. Living and fossil  
647 actinopterygians (with only a few exceptions in Osteoglossiformes, Anguilliformes,  
648 Lampridiformes, Siluriformes, and Syngnathiformes) and most piscine sarcopterygians  
649 (with exception in derived dipnoans) possess an anal fin. Among ‘elpistostegalians’,  
650 *Elpistostege* is known to retain an anal fin [15], which is definitely lost in tetrapods, while  
651 *Tiktaalik* might have lost the anal fin.

652         In the case of dorsal fins, paleontological and developmental evidence reveal that it  
653 is not constrained in its anterior extent and position, as opposed to the anal fin (which  
654 cannot extend anteriorly to the position of the anus), resulting in a variable occurrence of  
655 long-based and short-based dorsal fins in the earliest vertebrates [9]. *Pikaia* and  
656 Myllokunmingiida are the first and oldest early vertebrates in which a separate dorsal fin  
657 occurs. Lampreys display elongate dorsal fins that are not supported by radials. Extant

658 lampreys display two dorsal fins, separated by a gap but the ‘posterior dorsal fin’ is now  
659 assumed to corresponds to the anterior extension of the caudal epichordal lobe seen in  
660 fossil lampreys (e.g., *Mesomyzon*; [64]). Fossil lampreys (e.g., *Hardistiella* and  
661 *Mayomyzon*) show no gap separating the anterior and posterior dorsal fins as in extant  
662 lampreys, suggesting that the double dorsal fins of lampreys and gnathostomes is a  
663 convergent feature [54]. Duplication of short-based dorsal fins is also a common case in the  
664 evolution of vertebrates [9]. Several osteostracans have two dorsal fins (e.g., *Ateleaspis*,  
665 *Aceraspis*, and *Hirella*), but the anterior one lacks a fin web and resembles more a scale-  
666 covered hump than a proper fin [54]. On the other hand, the second dorsal fin of  
667 osteostracans clearly displays a fin web made of small scales arranged in a lepidotrichial  
668 pattern overlying numerous delicate radials. However, it is still debated whether this fin  
669 should be considered homologous to the posterior dorsal fin of gnathostomes or to the  
670 anterior part of the epichordal lobe of the caudal fin of lampreys, anaspids, and thelodonts.  
671 Among gnathostomes, ‘placoderms’ can either display single (e.g., antiarchs, stensionellids,  
672 rhenanids, and arthrodires) or double (e.g., ptyctodontids) dorsal fins [9]. Nevertheless, the  
673 plesiomorphic condition for crown gnathostomes, including chondrichthyans,  
674 ‘acanthodians’, and osteichthyans, is the occurrence of two dorsal fins supported by radials,  
675 fin rays, and sometimes associated spines [65–67]. Actinopterygians loss the anterior dorsal  
676 fin (from *Cheirolepis* onwards) but may regain a second dorsal fin either spinous (e.g.,  
677 acanthomorphs) or adipous (e.g., euteleosts) [9]. Many sarcopterygians primitively retain  
678 two dorsal fins, with the exception of the derived loss of the dorsal fins in post-Devonian  
679 dipnoans and ‘elpistostegalians’ + tetrapods.

680

681 **Comparison of the median fin developmental patterning in**  
682 **osteichthyans**

683 Fin and fin ray development have been well studied by developmental biologists  
684 since the middle of the 19<sup>th</sup> century (e.g., [68–70]) and numerous studies have dealt with  
685 the morphological and molecular features of fin ray development and regeneration (see a  
686 review in [7] and references therein). Indeed, fin rays are a good tool to better understand  
687 vertebrate ontogenetic development [17] and the connections between gene expression  
688 (during normal development, regeneration, and mutagenesis) and morphological and  
689 structural variation of anatomical traits.

690 Fin and fin ray development has been thoroughly surveyed in osteichthyans through  
691 the zebrafish *Danio rerio* [7]. Other studies on fin anatomy and development have been  
692 performed mostly in extant actinopterygians such as *Salmo* [26], *Medaka* [71], *Tilapia* [72],  
693 *Amia* [73], *Polyodon* [6,74], and *Acipenser* [74], but also in sarcopterygians such as the  
694 dipnoans *Neoceratodus* [75,76], *Protopterus* [25], and *Lepidosiren* [77], and the coelacanth  
695 *Latimeria* [78,79]. Our new data on extant (*Oncorhynchus*) and extinct (*Elonichthys*,  
696 *Miguashaia*, *Rhabdoderma*, *Dipterus*, *Quebecius*, and *Eusthenopteron*) osteichthyans allow  
697 us to accurately depict similarities in the developmental patterning of the median fins of  
698 bony fishes dealing with the morphological and temporary characteristics of appearance,  
699 chondrification, and ossification of endoskeletal elements (proximal and distal radials) and  
700 dermal fin rays (actinotrichia and lepidotrichia).



## 701 **Apparition, chondrification, and ossification of radials and lepidotrichia**

702           In extant actinopterygians (e.g., *Oncorhynchus*, *Salvelinus*, *Danio*) ([8,37,80]; this  
703 study) all developmental sequences (apparition, chondrification, and ossification) of the  
704 endoskeleton and exoskeleton for the dorsal and anal fins are bidirectional. This  
705 bidirectional pattern is corroborated by (1) common initiation sites for most corresponding  
706 events, (2) significant correlations between developmental sequences in both fins (dorsal  
707 and anal), and between the endoskeleton and exoskeleton (proximal/distal radials and  
708 lepidotrichia), and (3) a certain degree of simultaneity between sequences of  
709 apparition/chondrification (radials and lepidotrichia) and for the ossification of  
710 lepidotrichia. Despite little discrepancies among the initiation sites for  
711 apparition/chondrification/ossification of radials seen in *Oncorhynchus*, and between  
712 apparition/ossification of lepidotrichia, all these events can be confidently considered to be  
713 initiated from a unique initiation site. Unfortunately, the ossification patterns of many fossil  
714 osteichthyans are difficult to infer due to the preservation biases associated with the nature  
715 of fossilisation. However, in certain exceptional cases, ossification patterns can be  
716 tentatively reconstructed, such as in the coelacanth *Rhabdoderma* in which a bidirectional  
717 sequence occurred for the lepidotrichia Table 3, Fig. 8), whereas in the ‘osteolepiform’  
718 *Eusthenopteron* ossification of the distal radials proceeded postero-anteriorly [17,81]  
719 (Table 3); however, the narrowness of the fin and the reduced number of radials in  
720 *Eusthenopteron* might explain why the pattern is unidirectional.

721           In the case of lepidotrichia, fossil evidence confirms that dermal fin rays always  
722 ossify relatively early during ontogeny and before the endoskeletal radials ([82]; this

723 study), a condition identical to that of *Oncorhynchus* and other extant osteichthyans.

724 However, as for the radials, it is not easy to infer an ossification sequence for the

725 lepidotrichia in immature fossil specimens.

## 726 **Segmentation of lepidotrichia**

727 Lepidotrichial growth is achieved by successive addition of distal segments at the

728 extremity of the forming lepidotrichia [28]. The process of segmentation is congruent

729 between the dorsal and anal fins in *Oncorhynchus* in terms of: (1) similar initiation sites,

730 (2) common bidirectional sequences of segmentation, (3) simultaneity, and (4) significantly

731 correlated sequences. The correlations among the lepidotrichia apparition, segmentation,

732 and ossification sequences within both fins suggest that all three sequences are initiated

733 from the same site and proceed in a bidirectional sequence. This congruence also matches

734 the apparition/chondrification/ossification patterns described in radials.

735 In fossil taxa, an important aspect concerns the identification of the initiation site of

736 segmentation. Considering the observation made in *Oncorhynchus* (i.e. longer lepidotrichia

737 are the ones for which segmentation started earlier), the location of the longest lepidotrichia

738 in the dorsal and anal fins can be confidently identified as the initiation site of segmentation

739 in fossil specimens. This scenario has been proposed in all extinct osteichthyans surveyed

740 (i.e., *Elonichthys*, *Miguashaia*, *Rhabdoderma*, *Quebecius*, *Dipterus*, and *Eusthenopteron*)

741 (Table 3).

742 These similarities in patterning imply that sequences of segmentation may as well

743 be generally used as proxies for sequences of ossification in osteichthyans. Considering the

744 impracticality to observe an ossification sequence for lepidotrichia in fossil specimens,  
745 segmentation patterns are essential for comparisons between living and extinct taxa.

## 746 **Bifurcation of lepidotrichia**

747 Bifurcations are the results of the distal branching of an individual lepidotrichial  
748 segment after the intersegmental joint [38]. As for segmentation, bifurcation patterns can be  
749 compared in extant and extinct taxa. However, the bifurcation pattern is not clear in  
750 *Oncorhynchus*: (1) the sequences of bifurcation of both fins are bidirectional but not  
751 correlated, (2) the initiation site for bifurcation is posterior to the other initiation sites  
752 (apparition, ossification, segmentation) in the dorsal fin but similar in the anal fin, and (3)  
753 the initiation site corresponds to the lepidotrichia with the most proximal bifurcations in the  
754 dorsal fin but not in the anal fin. These results might be artefactual owing to small sample  
755 size and inter-individual variation. However, despite these potential biases, bifurcation  
756 sequences are likely similar between fins.

757 The bifurcation patterns found in both fins of extinct sarcopterygians (e.g.,  
758 *Miguashaia*, *Quebecius*, *Dipterus*, *Eusthenopteron*) corroborate the observations made on  
759 the dorsal fin of *Oncorhynchus*: (1) an initiation site for bifurcation located posteriorly to  
760 the initiation site for segmentation, (2) concordance between the initiation site and the  
761 lepidotrichia with the most proximal bifurcations, and (3) a bidirectional sequence of  
762 bifurcation. These similarities between extant and extinct osteichthyans suggest a shared  
763 pattern of bifurcation in which bifurcation is initiated in a different position than the other  
764 events for the lepidotrichia.

765

## 766 **Median fin modularity and evolution in osteichthyans**

767           Modularity is a fundamental property of organisms playing an important role in  
768 their evolution [83–85]. Anatomical modules refer to an internal organization of anatomical  
769 structures into distinct units, or modules, which develop and vary in quasi-autonomy, but  
770 within which the constituents interact and vary together [83,85,86]. This quasi-autonomy  
771 among modules allows three main evolutionary processes: dissociation,  
772 duplication/divergence, and co-option [83]. Different categories of modularity, and  
773 modules, have been defined over the past few decades. Among these categories, Zelditch &  
774 Goswami [85] emphasized the intricate relationship between developmental and functional  
775 modularity. Developmental modules are often represented as networks depicting their  
776 physical location, spatial extent and genetic specification, while functional modules are  
777 represented by anatomical elements integrated as structural components of a functional (or  
778 physiological) system [85]. However, developmental and functional modules have not been  
779 investigated methodologically as thoroughly as variational and evolutionary modules (see  
780 [85] for an exhaustive critical review of the methods). Thus, the identification of  
781 developmental and functional modules is frequently only suggested without being tested.  
782 Herein, we have proposed a method to compare similarities among sequences of  
783 developmental events helping to assess developmental modules.

784           Building on the original idea of Mabee et al. [16] of comparing sequence of  
785 formation (e.g., chondrification and ossification; e.g., in [87]), few studies have proposed to  
786 quantify the phenotypic patterning using correlation. Previous examples include correlation  
787 of relative sequence of events [88] and correlation of neural branching patterns in the skull

788 [89]. Patterning is closely associated to the concept of developmental modularity [83,89].  
789 Thus, methodologically one has to compare similar patterning to infer developmental  
790 modules.

791 In our study, we are comparing our results that are methodologically constrained  
792 with previous developmental modules proposed by Mabee et al. [16] for actinopterygians:  
793 (1) the Endoskeleton and Exoskeleton Module (EEM) and (2) the Dorsal and Anal Fin  
794 Patterning Module (DAFPM). We will discuss the evidences supporting the occurrence of  
795 these modules in the median fins of the surveyed taxa, in osteichthyans, and in vertebrates  
796 as a whole.

797

## 798 **Endoskeleton and Exoskeleton Module**

799 The median and paired fins of osteichthyans are constituted of two skeletons, or  
800 modules, formed by distinct developmental processes: (1) the endoskeleton and (2) the  
801 exoskeleton [90–92]. In extant vertebrates, and more particularly in gnathostomes, the  
802 Endoskeleton and Exoskeleton Module (EEM) explains the similarities in the direction of  
803 development of the endoskeleton from the paired and median fins (fin radials and girdles)  
804 and the exoskeleton (fin rays). The EEM is thus composed of two interacting submodules  
805 (SM): (1) the endoskeleton submodule (EnSM) and (2) the exoskeleton submodule  
806 (ExSM). The EnSM as a whole probably originated at the base of the Gnathostomata with  
807 the evolution of pelvic fins in ‘placoderms’, homologous to those of osteichthyans [54,93],  
808 while the ExSM is related to the origin of lepidotrichia in osteichthyans [24,94] (Fig 12).

809

810 **Fig 12. Vertebrate phylogeny illustrating the evolution of fin related characters and**  
811 **median fin modules.** A, Interrelationships of the main groups of vertebrates and  
812 distribution of fin characters. Taxa in bold have living representatives. Fins have been  
813 plotted in the tree according to their definition as membranous outgrowths of the body  
814 walls internally supported by endoskeletal (e.g., radial bones) or exoskeletal (e.g., fin rays)  
815 elements and based on consensual hypotheses of homology (see main text for a discussion  
816 on the distribution of fins in the selected taxa). Preferred tree topology combined from [95–  
817 97] for non-gnathostomes, [98,99] for actinopterygians, and [15,100] for sarcopterygians;  
818 B, Distribution of median fin modules (DAFPM and EEM) in the studied species. DAFPM,  
819 dorsal and anal fin patterning module; EEM, endoskeleton exoskeleton module; LPM,  
820 lepidotrichia patterning module.

821

822 The endoskeletal components of a vertebrate fin includes series of radial bones,  
823 which support both paired (pectoral and pelvic) and median (dorsal, anal and caudal) fins.  
824 Endoskeletal radials have been proposed to be present in the median fins of the earliest  
825 vertebrates (e.g., myllokunmingiids like *Haikouichthys*) [16,101]. However, these  
826 supposedly cartilaginous structures have been reinterpreted as either epidermal folds or  
827 collagenous structures [11,54,102] and the presence of true radials is thus now restricted to  
828 crown vertebrates as present in the caudal fin of hagfishes, lampreys, and even conodonts  
829 [11,54,103]. Radial elements (either osseous or cartilaginous) have been identified in the  
830 caudal fin of many other subsequent groups of vertebrates (e.g., heterostracans, anaspids,  
831 osteostracans and gnathostomes) [52], but it is not always clear whether the other median

832 fins (i.e., dorsal and anal) were also supported by radials. The ribbon-shaped anteroventral  
833 paired fins of certain ‘agnathans’ (e.g., *Euphanerops*, anaspids) present numerous thin,  
834 parallel radials, lacking any fin support or girdle [55], whereas in osteostracans, possibly  
835 pituriaspids, and gnathostomes a few large radials articulate with a massive girdle forming  
836 stout, paddle-shaped paired fins [11,54].

837 Dermal fin rays are absent in cyclostomes (hagfishes and lampreys), but they are  
838 known in non-osteichthyans (e.g., in ‘agnathans’ like *Euphanerops* and in gnathostomes  
839 like ‘acanthodians’ and chondrichthyans) [55,63,104,105]. However, the structure and  
840 histological nature of the fin rays in early vertebrates is difficult to decipher. In  
841 *Euphanerops*, the paired anteroventral fins are composed of ‘fin rays’ made of stacked  
842 chondrocytes, which articulate with the cartilaginous radials [55]. In anaspids (e.g.,  
843 *Birkenia*), the anal and caudal fin are covered with small scales and in the epichordal lobe  
844 of the caudal fin the scales are arranged in rows, recalling the jointed structure of the  
845 lepidotrichia in osteichthyans [52]. The small median fins of thelodonts (e.g., *Phlebolepis*)  
846 are also covered by minute scales, closely stacked together and sometimes forming a fin  
847 web [106], similar to the configuration of the caudal fin of the arandaspid *Sacabambaspis*  
848 [58]. In osteostracans (e.g., *Escuminaspis*), the second dorsal fin is supported by numerous  
849 narrow radials and covered by small scales arranged in rows [52,107]. Among  
850 gnathostomes, chondrichthyans possess ceratotrichia [104,108–111], large fibrous fin rays  
851 but homologous in all senses to the actinotrichia of osteichthyans [19,78,112,113].  
852 Ceratotrichia have also been identified in ‘placoderms’ [e.g., *Bothriolepis* [114–117],  
853 dunkleosteids [118], stensionellids [115]. ‘Acanthodians’ possess dermal fin rays with an

854 ossified proximal portion and a distal, non-ossified portion, which, according to Géraudie  
855 and Meunier [78] correspond to large fibrous rays, similar to ceratotrichia. Wide  
856 ceratotrichia may thus represent the primitive condition from which the slender  
857 actinotrichia evolved in osteichthyans. Completely ossified fin rays (lepidotrichia) are  
858 solely present in osteichthyans.

859         The exoskeleton submodule (ExSM) might thus have been present in the last  
860 common ancestor of chondrichthyans and osteichthyans, since partially ossified fin rays  
861 (i.e., potential lepidotrichia) may be present in the caudal fin of certain ‘acanthodians’  
862 [119,120], currently considered as stem chondrichthyans [65, 121,122]. On the other hand,  
863 this module is lost in the limbs of tetrapods [91,123], in which dermal fin rays are absent  
864 from the paired fins but still retained in the caudal fin of Devonian forms (e.g.,  
865 *Acanthostega*, *Ichthyostega*) [124,125]. This pattern of lepidotrichial reduction at the  
866 transition between fishes and tetrapods has been documented in *Tiktaalik* by looking at the  
867 dorso-ventral asymmetry of hemirays [126]. The origin of the EEM most likely coincides  
868 with the common presence of both submodules (EnSM and ExSM) in crown gnathostomes  
869 (Fig. 12). Interactions between these two modules resulted in the morphological disparity in  
870 terms of relative size, shape, and position of the paired and median fins.

871         Based on the sequence of nine events surveyed during *Oncorhynchus* fin  
872 development, the patterning of the EEM in living actinopterygians is composed of five  
873 events, all starting from the same initiation site and following the same direction: (1)  
874 apparition/chondrification of radials, (2) apparition of lepidotrichia, (3) segmentation of  
875 lepidotrichia, (4) ossification of lepidotrichia, and (5) ossification of proximal radials. The



876 ossification of distal radials is probably the sixth step, but it was not observed in our  
877 juvenile specimens. Based on our observations, the EEM can thus be confidently  
878 considered plesiomorphic at least in actinopterygians, since it was not possible to confirm  
879 its presence in fossil sarcopterygians.

## 880 **Dorsal and Anal Fin Patterning Module**

881         The anatomical composition, disparity, phylogenetic distribution, and modularity of  
882 fins in all orders of extinct and living fishes have been surveyed by Larouche et al. [9,11].  
883 Larouche et al. [9] recognized that the dorsal and anal fins form an evolutionary module  
884 nested within a median fin module. A Dorsal and Anal Fin Module (DAFM) most likely  
885 evolved early in stem-gnathostomes (Fig. 12). Larouche et al. [11] suggested that fin  
886 modules, including DAFM, re-expressed within the topographic boundaries of fin-forming  
887 morphogenetic fields.

888         Mabee et al. [16] described the DAFPM based on a similar direction of  
889 differentiation (i.e., apparition of the skeletal elements) for the dorsal and anal fins of  
890 actinopterygians. However, our data allow to expand Mabee et al.'s original description of  
891 the DAFPM to also include the patternings of chondrification, ossification, segmentation,  
892 and bifurcation (Tables 2 and 3; Fig 3). Further evidences of the DAFPM are given on  
893 extant actinopterygians by similar coordination of phenotypic plastic response between  
894 both fins [37] and similar pattern of correlated shape between both fins [127]. Mabee et al.  
895 [16] considered the bidirectional direction of development as plesiomorphic at least for  
896 teleosts. Our data show that the sequences of all events are effectively bidirectional in  
897 *Oncorhynchus* and most fossil osteichthyans (Table 3) thus suggesting that developmental

898 bidirectionality is most likely plesiomorphic for osteichthyans. This general bidirectional  
899 development may differ in some sarcopterygians (e.g., radials in *Eusthenopteron*), most  
900 likely owing to a reduction of size in both fins, but does not necessarily compromise the  
901 presence of the primitively shared DAFPM among osteichthyans.

902 The Dorsal and Anal Fin Patterning Module (DAFPM) is the developmental module  
903 corresponding to the anatomical DAFM of Larouche et al. [9]. In addition, Mabee et al.  
904 [16] suggested that the DAFPM originated from a Dorsal and Anal Fin Positioning Module  
905 (DAFPoM), one in which the antero-posterior position of the dorsal and anal fins are  
906 correlated. Most likely the basal condition of the DAFPoM in actinopterygians is a  
907 condition in which these median fins occupy a symmetrical position [16]; this condition is  
908 observable in Devonian actinopterygians (e.g., *Dialipina*, *Pickeringius*, *Howqualepis*,  
909 *Mimipiscis* [128–130]), while the anal fin is located slightly anterior to the dorsal fin in  
910 *Cheirolepis* [131] or the opposite (e.g., *Gogosardina*, *Moythomasia*, *Limnomis* [132–134]).  
911 Mabee et al. [16] mentioned that there was a high level of dissociability of the positioning  
912 module from the patterning module among actinopterygians. Outside actinopterygians, the  
913 generalized condition in actinistians (*Miguashaia*; [22]), dipnomorphs (e.g., *Holoptychius*,  
914 *Quebecius*, *Glyptolepis*, *Uranolophus*; [48,49]) and tetrapodomorphs (e.g., *Cabonnichthys*,  
915 *Eusthenopteron*, *Gyroptychius*, *Heddleichthys*; [50,135,136]) also corresponds to the  
916 symmetrical positioning. Not only the position of the dorsal and anal fins is symmetrical,  
917 but the shape, size and number of endoskeletal elements correspond to a mirror image. The  
918 DAFPM of osteichthyans implies that the skeletal elements (either of endochondral origin  
919 such as the radials or dermal origin such as the lepidotrichia) of both median fins (anal and

920 dorsal) differentiate in the same direction and thus share common developmental properties  
921 [16].

922 The DAFPM is confirmed by our data in actinopterygians (*Oncorhynchus*,  
923 *Elonichthys*), as well as in sarcopterygians like coelacanth (*Miguashaia*, *Rhabdoderma*),  
924 porolepiforms (*Quebecius*), and ‘osteolepiforms’ (*Eusthenopteron*) (Tables 2 and 3). The  
925 DAFPM can also be recognised in early lungfishes (e.g., *Dipterus*, *Barwickia*) based on  
926 morphological similarities between these fins, particularly with respect to the oar-shaped  
927 fin support supporting the distal radials [137]. However, due to the heterogeneity of fin  
928 morphologies in dipnoans it is difficult to confidently reconstruct the evolution of the  
929 DAFPM across lungfishes [17]. Consequently, a dissociation of the DAFPM is inferred  
930 during dipnoan evolution. Multiple dissociations have also occurred in actinopterygians  
931 every time either one of the dorsal or anal fin is absent or fused to the caudal fin (e.g.,  
932 Osteoglossiformes, Anguilliformes, Siluriformes, Lampridiformes, Sygnathiformes) [9].  
933 Among other sarcopterygians non-surveyed in our study, the DAFPM can be certainly  
934 inferred in onychodonts (e.g., [138]) and rhizodonts (e.g., [139]) based on the size and  
935 shape similarities between both median fins. The DAFPM is lost definitely at the basis of  
936 the clade including the ‘elpistostegalians’ and tetrapods with the loss of the dorsal fins (Fig  
937 12).

### 938 **Lepidotrichial Patterning Module**

939 Molecular mechanisms and grafting experiments in zebrafish suggest that a  
940 lepidotrichium can be grafted to a new location and grows quite normally by the addition of  
941 new segments [140,141]. However, interactions among lepidotrichia are necessary to

942 achieve the original morphology because the position of segmentation and bifurcation are  
943 dependant of the position of the lepidotrichia within fins [140,141]. This suggests a smaller  
944 modular unit within the exoskeleton module (ExM), the lepidotrichium itself. The hemi-  
945 lepidotrichium (or hemiray) could be the smallest unit of regeneration (and development) in  
946 the hierarchical modular organization of fins [142], because developmental interactions are  
947 recognized to control coordination of segmentation and bifurcation between hemirays  
948 [141]. Segmentation and bifurcation patterning similarities found in this study may be  
949 indicative of molecular mechanism conservatism in osteichthyans.

950       Lepidotrichia patterning in *Oncorhynchus* includes: (1) apparition, (2)  
951 segmentation, (3) ossification, and (4) bifurcation. Data from fossil specimens agree with  
952 this sequence. Therefore, the patterning of the lepidotrichia appears conserved in  
953 osteichthyans and a “Lepidotrichia Patterning Module” (LPM) may be generalized in all  
954 the fins of all osteichthyans.

955       The molecular basis behind the establishment of segmentation has been explored by  
956 using regenerative experiments in zebrafish. Part of the molecular machinery involved in  
957 segmentation is recruited for lepidotrichia bifurcation [18,143]. Molecular transcripts  
958 involved in segmentation are: (1) *evx1* acts as an on/off switch defining the putative  
959 boundaries between two successive segments [28], mutant zebrafish for this gene grow  
960 normal lepidotrichia, but joint formation between successive segments is impaired [38]; (2)  
961 three genes of the Sonic Hedge Hog signalling pathway (i.e., *shh*, *bmp2*, *ptc1*) are thought  
962 to be involved in the patterning of the lateral limits of segments [18]; (3) *hoxa13b* may

963 participate in the elaboration of the next segment [144]; and (4) *cx43*, a particular gene  
964 allowing intercellular communication involved in joint position [145].

965         Segmentation and bifurcation processes might be independently regulated. In *evx1*  
966 zebrafish mutants, bifurcation occurs normally while joint formation is down-regulated  
967 [38]. At least two genes are expressed during the bifurcation process: (1) prior to  
968 bifurcation, *shh* is expressed centrally where a new segment is forming, then laterally in the  
969 presumptive twin-segments [18,143], and (2) *bmp2* (required for bone synthesis in the  
970 central region of the segment) is restricted in the two lateral domains copying *shh* prior to  
971 branching [18]. Fossil evidence shows that bifurcation likely originated at the base of  
972 osteichthyans. One notable exception is the puzzling *Dialipina salgueiroensis*, which lacks  
973 bifurcation (RC, pers. obs.); *Dialipina* is either considered a stem actinopterygian [146] or  
974 a stem osteichthyan [24]. The absence of bifurcation is highly homoplastic in living  
975 [32,147] and fossil actinopterygians as well as in some sarcopterygians (e.g., it constitutes a  
976 common derived feature of post-Devonian coelacanths) [148]. Because bifurcation is the  
977 last event of the lepidotrichia developmental sequence, it might be more susceptible to be  
978 affected by epigenetic phenomena [37].

979

## 980 **Conclusions**

981         Our analysis of median fin development in *Oncorhynchus* has allowed the  
982 quantification and validation of two median fin modules in a living actinopterygian: the  
983 Dorsal and Anal Fin Patterning Module (DAFPM) and the Endoskeleton and Exoskeleton  
984 Module (EEM). Comparison with other extinct osteichthyans, comprising both

985 actinopterygians and sarcopterygians (coelacanth, dipnoans, porolepiforms and  
986 ‘osteolepiforms’) has corroborated the data on extant taxa, but highlighted the difficulties  
987 of confidently identifying developmental sequences based on fossil specimens. The  
988 DAFPM and EEM modules incorporate all the events associated with fin patterning  
989 including the sequences of segmentation and bifurcation of the lepidotrichia that are crucial  
990 for comparisons and inferences of developmental sequences in fossil osteichthyans.

991 We suggest that: (1) the EEM includes the apparition, segmentation, and  
992 ossification sequences, and, based on our results, is plesiomorphic at least for  
993 actinopterygians; (2) the DAFPM includes the apparition, chondrification, and ossification  
994 sequences plus the segmentation and bifurcation sequences, and is plesiomorphic for  
995 osteichthyans with multiple dissociations along osteichthyan phylogeny. Additionally, the  
996 recurrence of the developmental pattern of the lepidotrichia in living and fossil  
997 osteichthyans suggests an additional developmental module within fins, the Lepidotrichia  
998 Patterning Module (LPM), where the constitutive units, the hemirays, have a synchronous  
999 and similar development. The median fins of osteichthyans have thus been shown to be  
1000 important representatives for the study of modularity across the evolution of vertebrates.

1001

## 1002 **Acknowledgments**

1003 For the loan of specimens, we thank J. Maisey, I. Rutzky and J. Galkin (AMNH), L.  
1004 Grande, B. Simpson and E. Zeiger (FMNH), J. Kerr, N. Parent and J. Willett (MHNM), A.  
1005 Lévesque (ULQ), and J. Gauthier and M.A. Turner (YPM). S. Cumbaa (NMC), Z.  
1006 Johanson (NHM), and K. Mickle (KU) provided specimen photographs. I. Béchar

1007 digitized the drawings. I. Béchar, L. Fischer-Rousseau, Z. Johanson, and C. Riley  
1008 provided constructive comments on earlier versions of this manuscript. B. Vincent (UQAR)  
1009 provided statistical and programming advices. P. Janvier (MNHN) and M. Marí-Beffa  
1010 (UMA) are warmly thanked for their insightful comments on fin morphogenesis, fin ray  
1011 structure and fin diversity.

1012

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## 1402 **Supporting information**

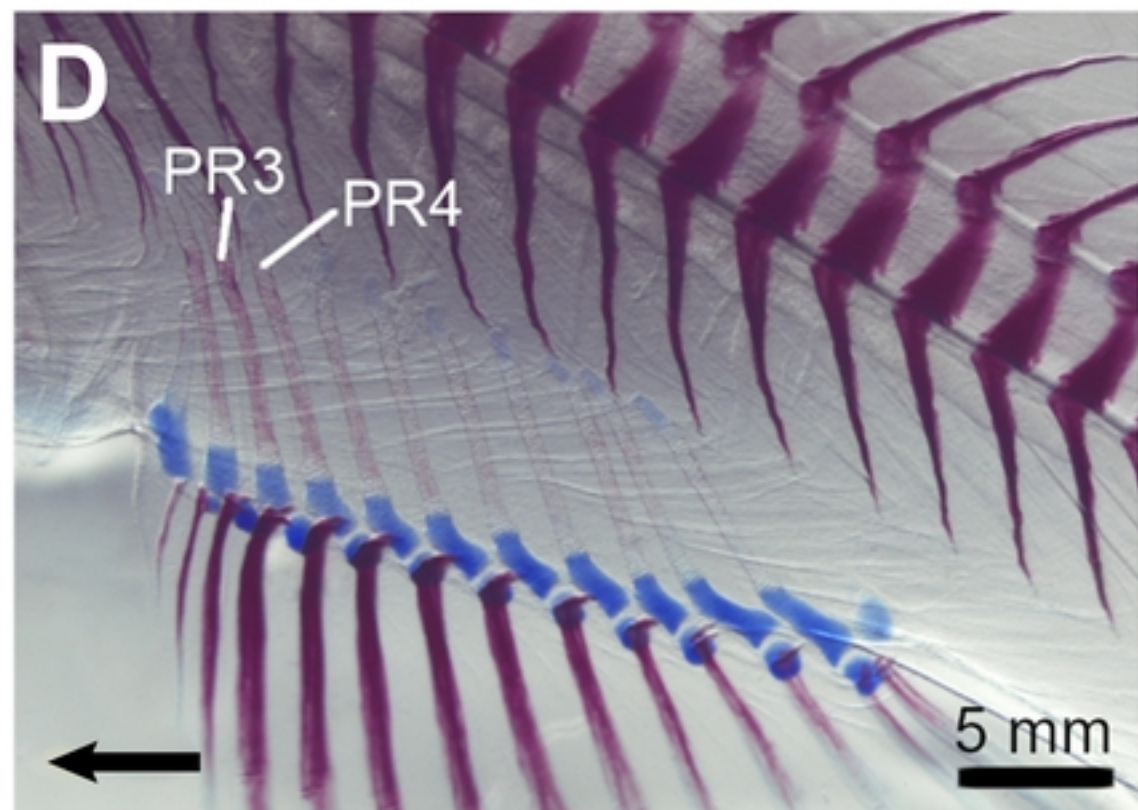
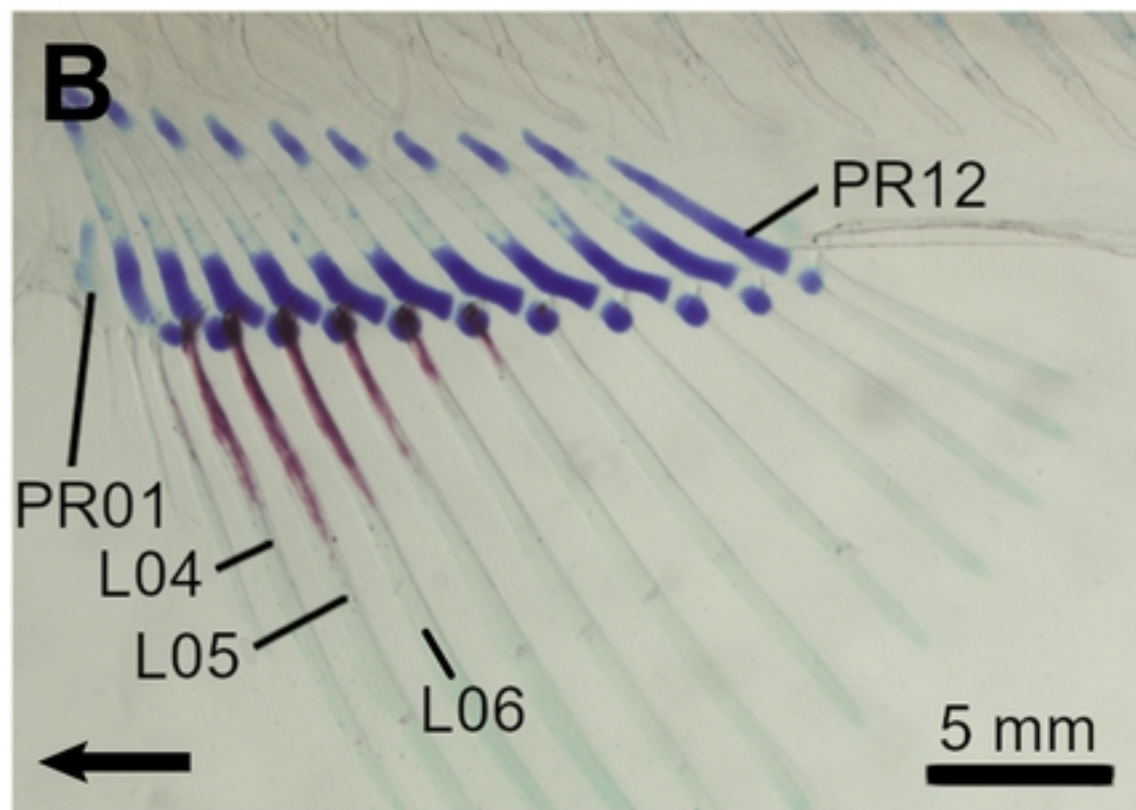
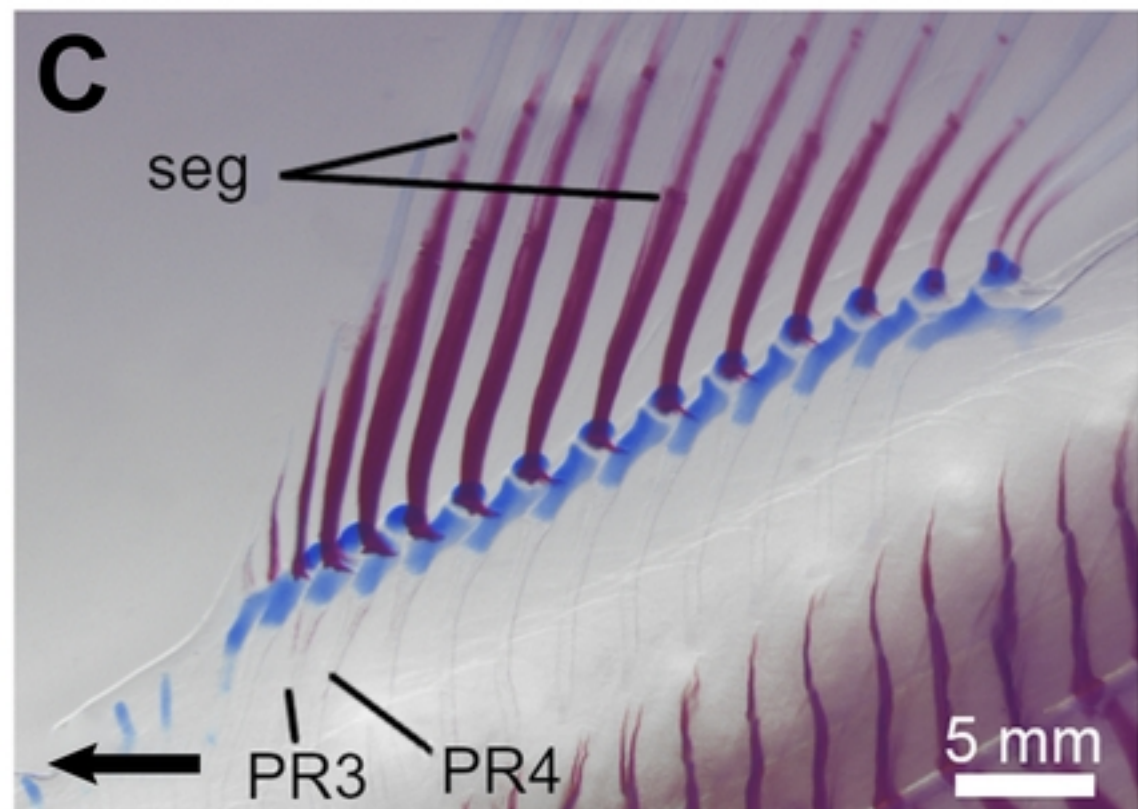
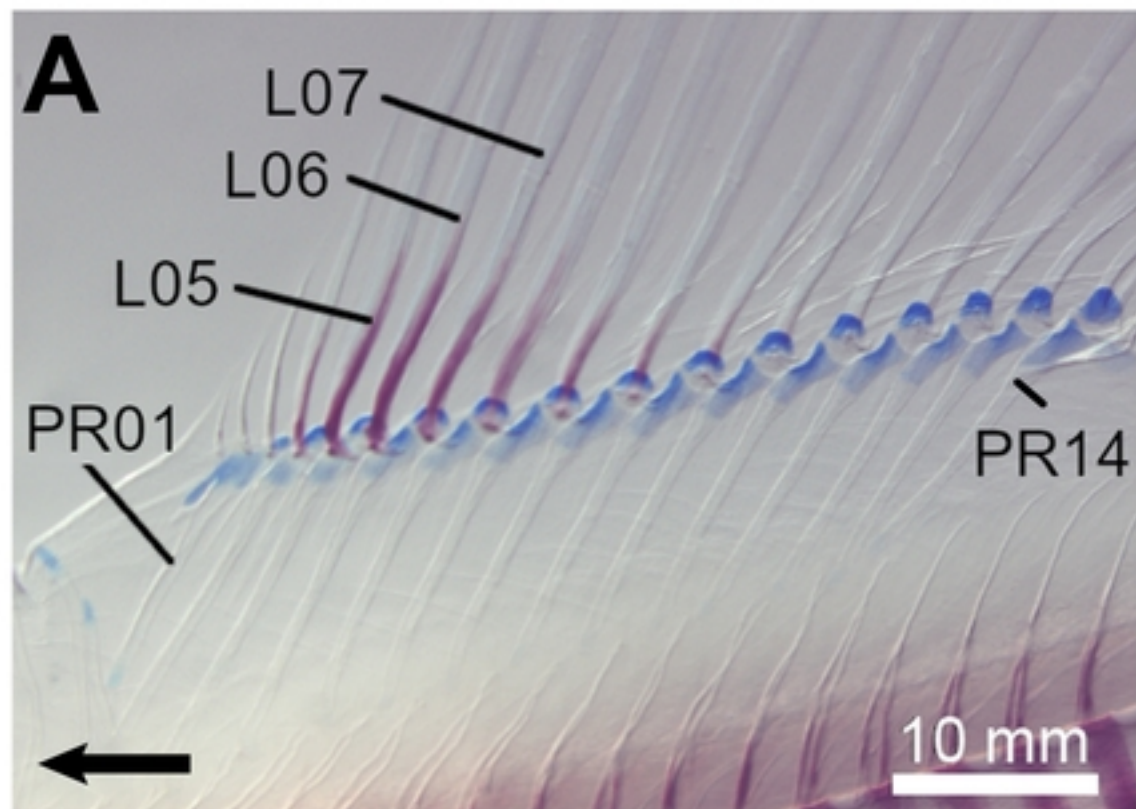
1403

1404 **S1 Table. Fossil specimens examined given in size order.** For each species, the size  
1405 series is given with SL or TL (mm) and specimen number. Only *D. valenciennesi* is not  
1406 represented by immature specimens.

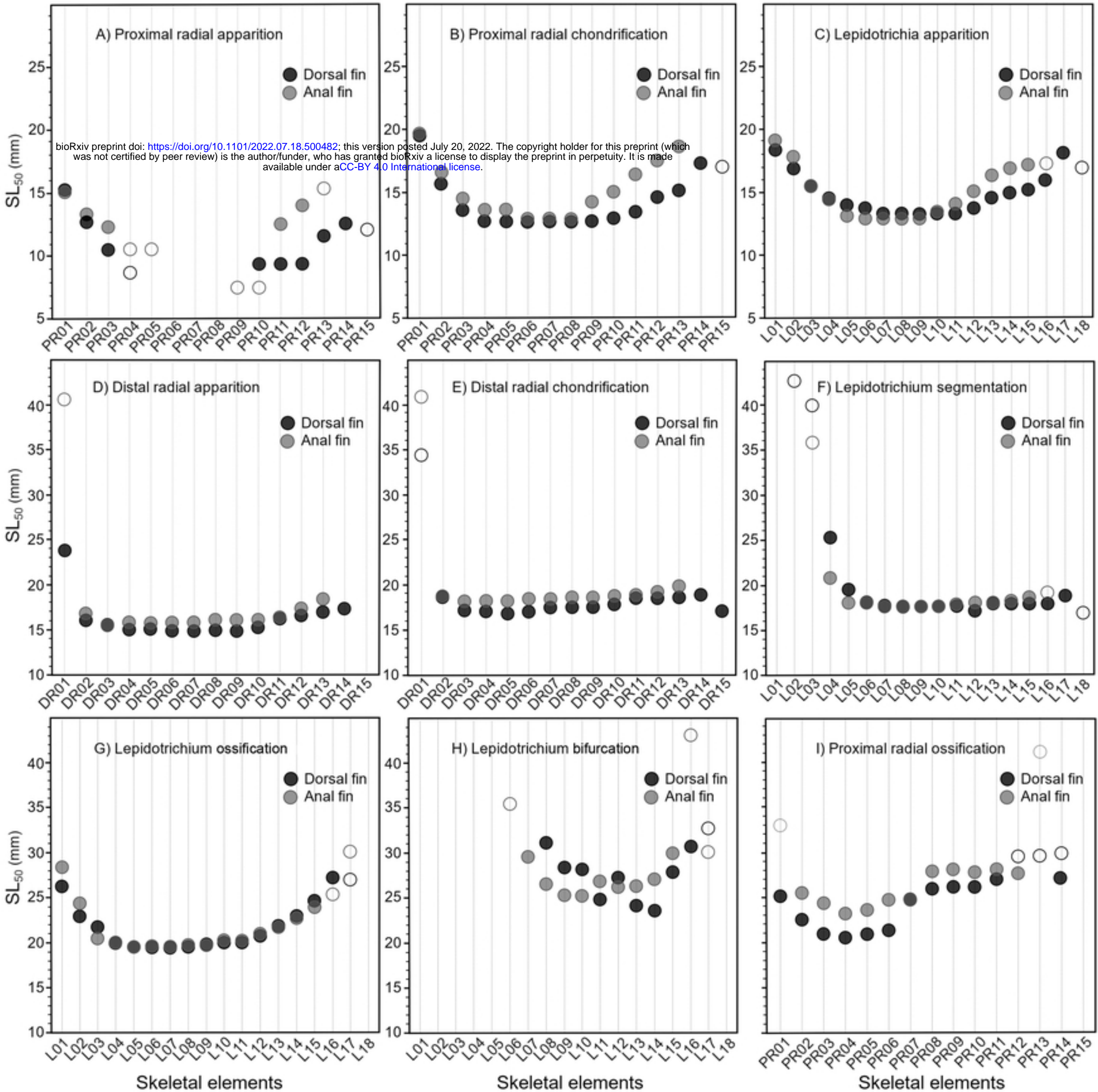
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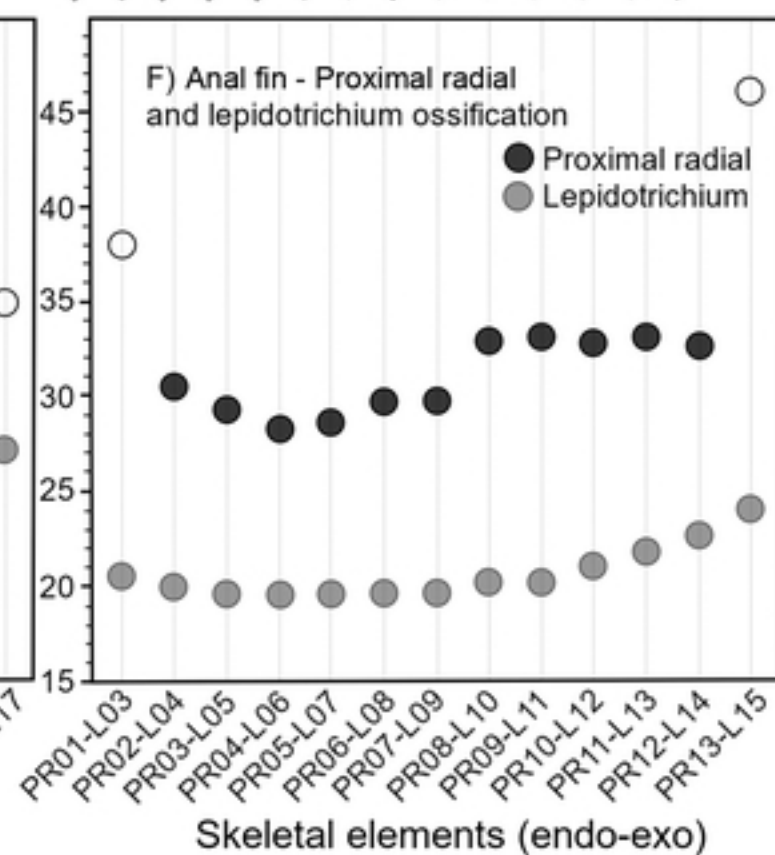
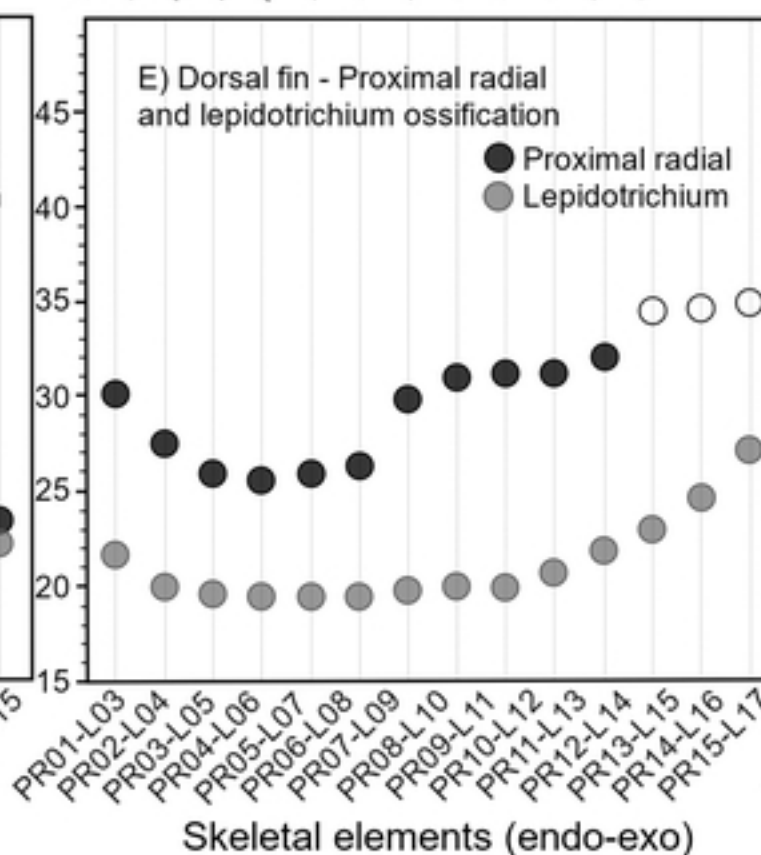
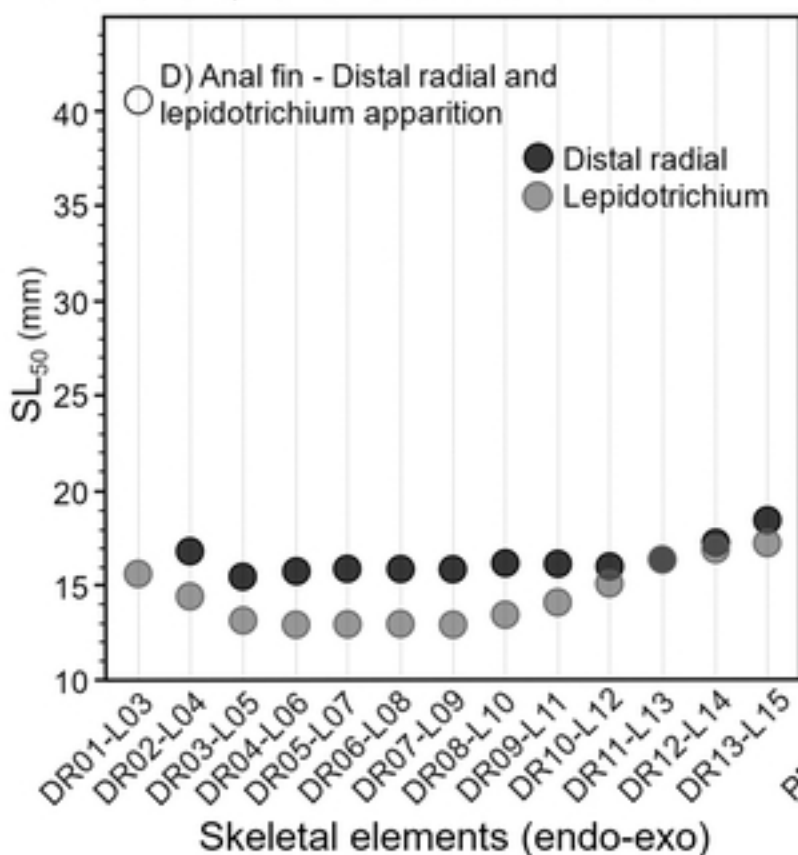
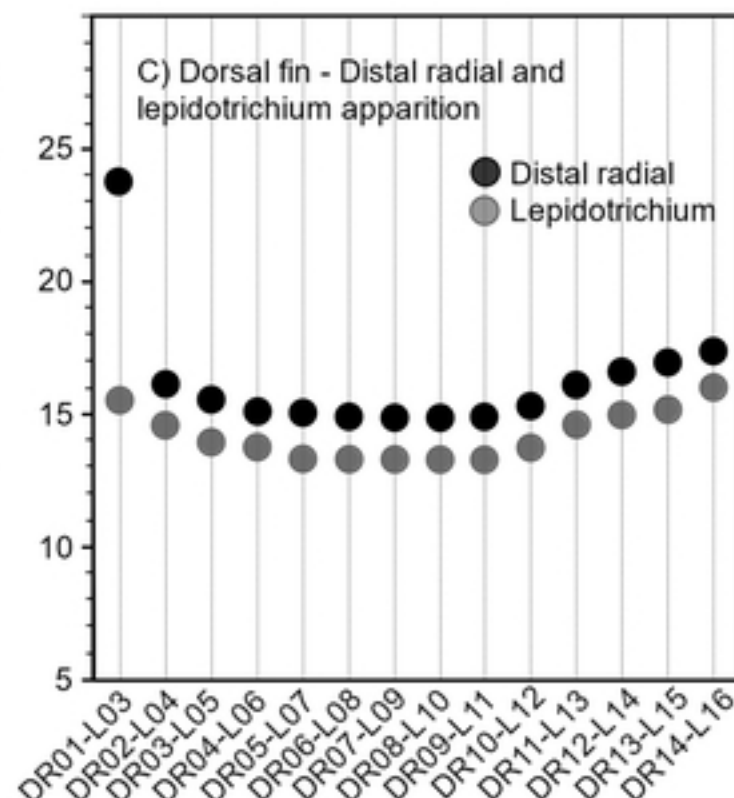
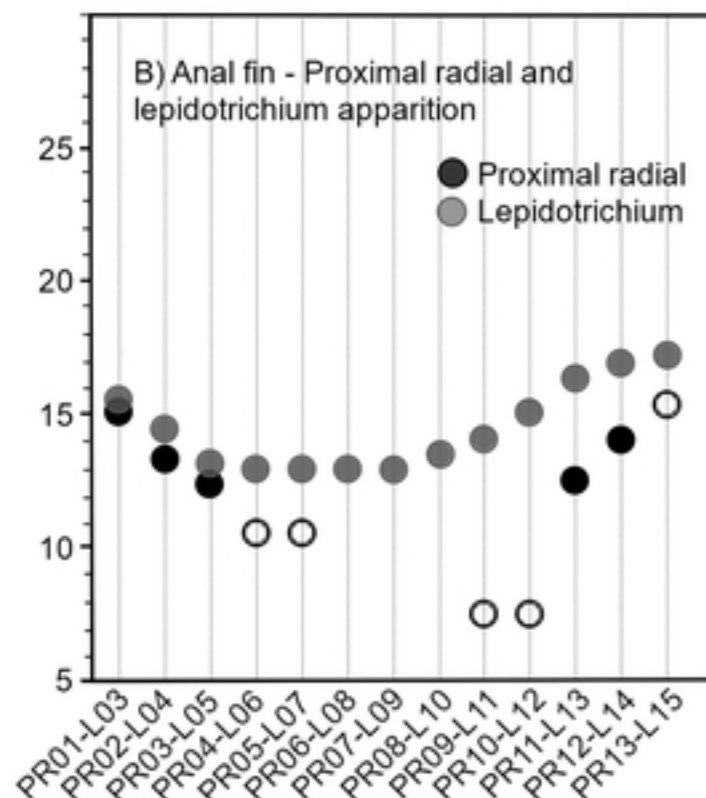
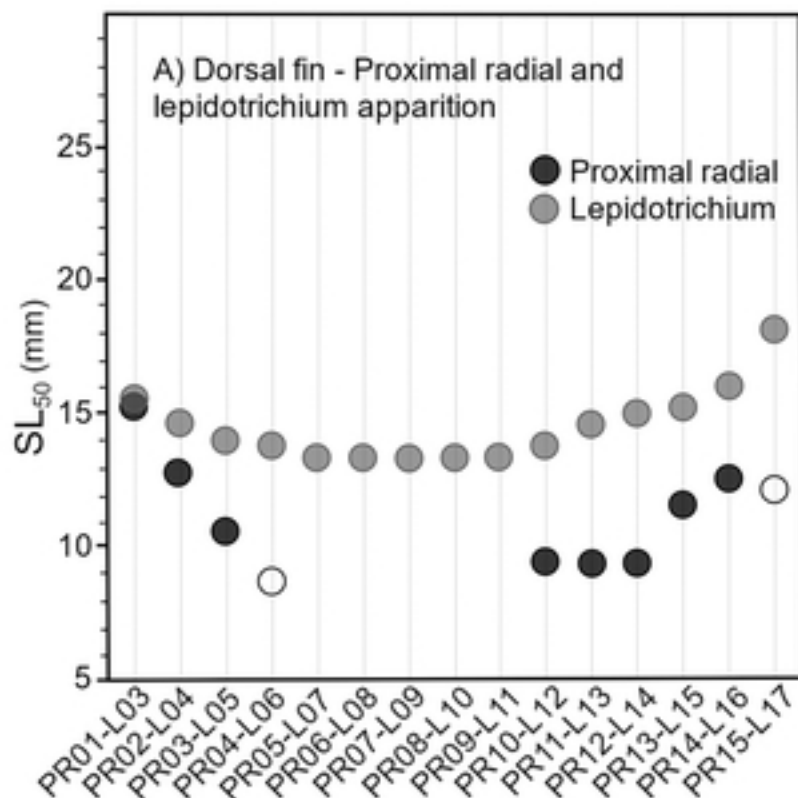
1408 **S1 File. Coding of the serial skeletal elements, i.e. the proximal radials (PR), the distal**  
1409 **radials (DR) and the lepidotrichia (L), during nine out of eleven developmental events**  
1410 **of the developmental sequence of the dorsal and anal fins of *Oncorhynchus mykiss*.** For  
1411 each specimen, the size is given in SL (mm) and the skeletal elements are coded with 0, 1 or  
1412 NA. The definitions of these codes are different for each developmental event, they are  
1413 provided.

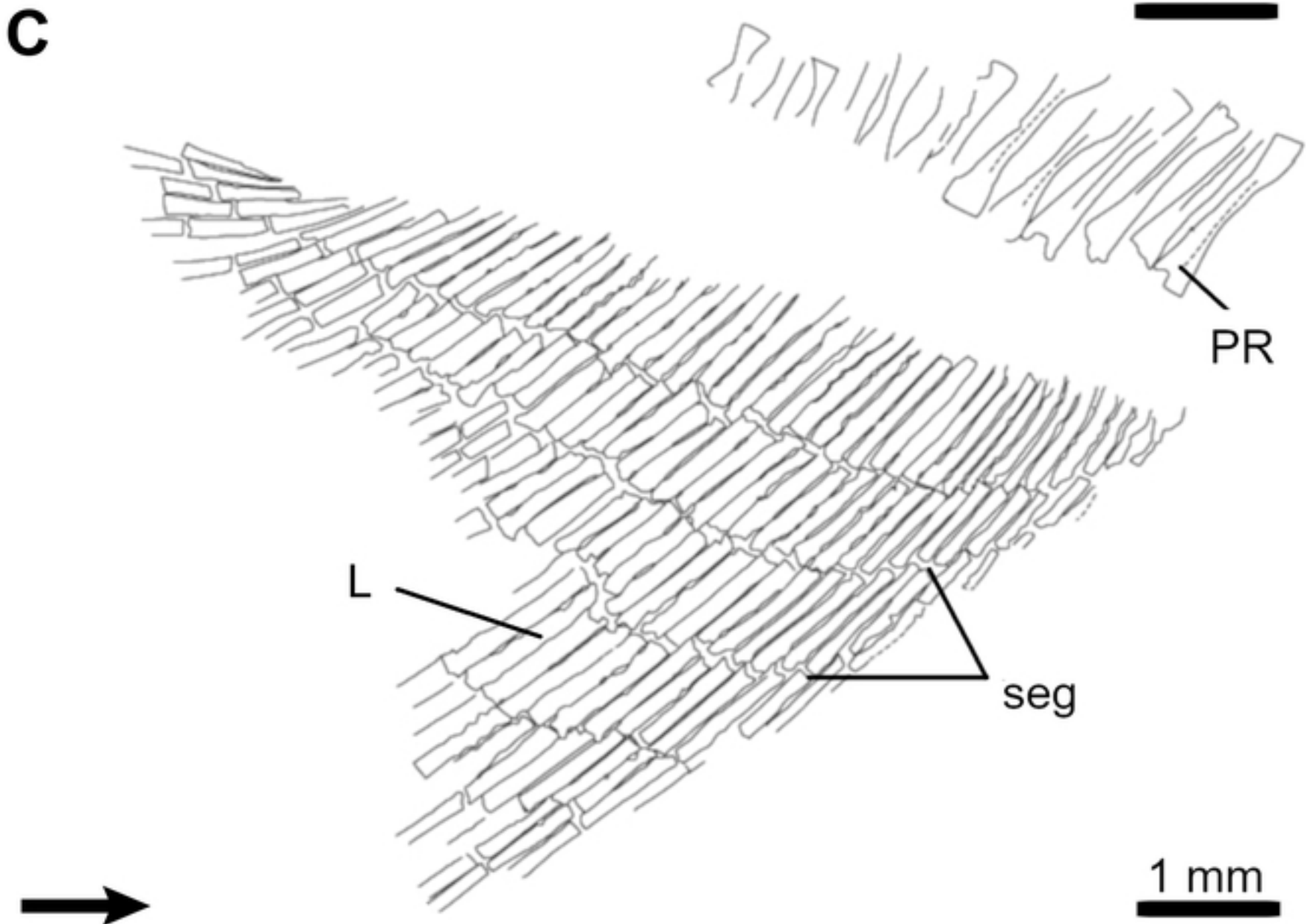
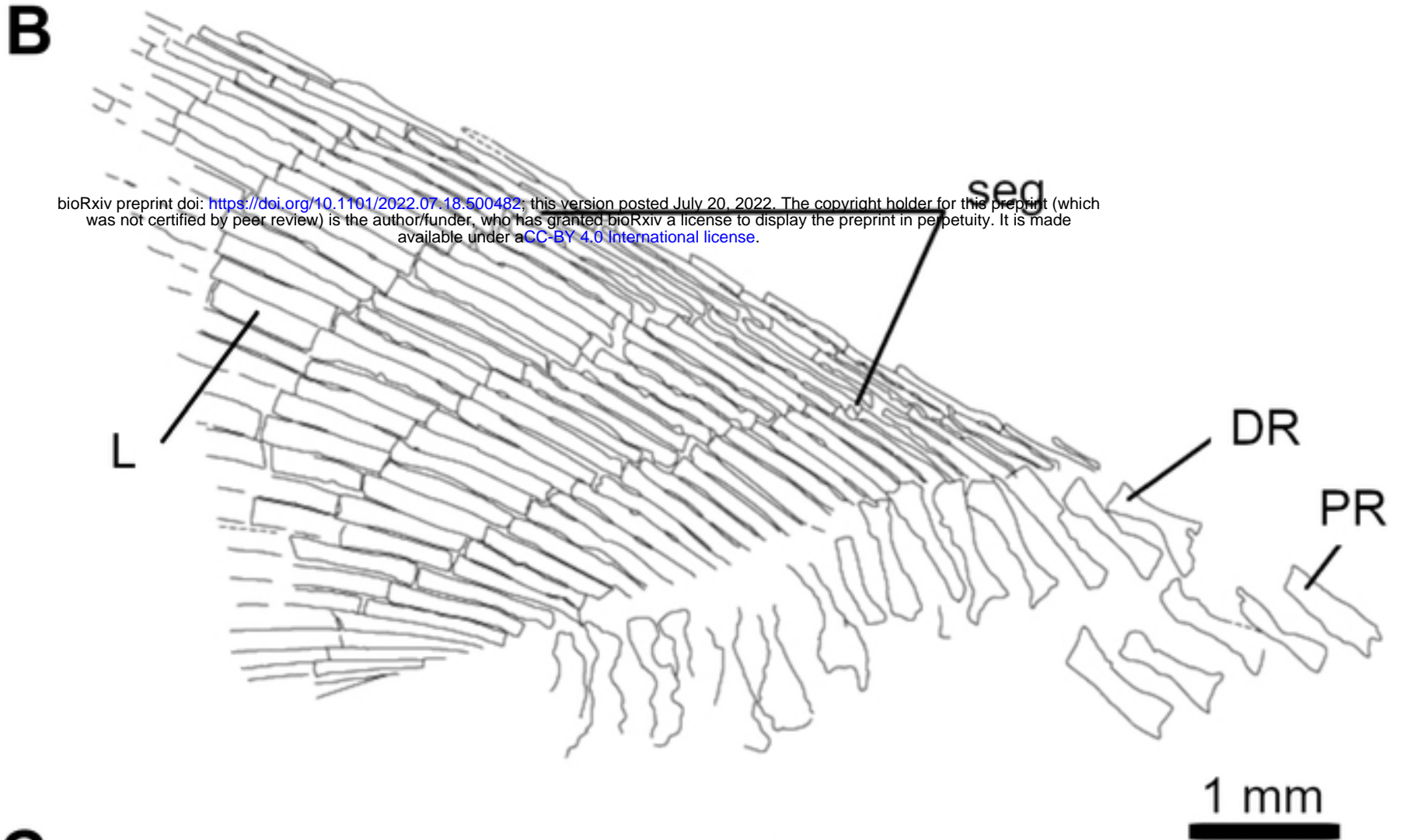
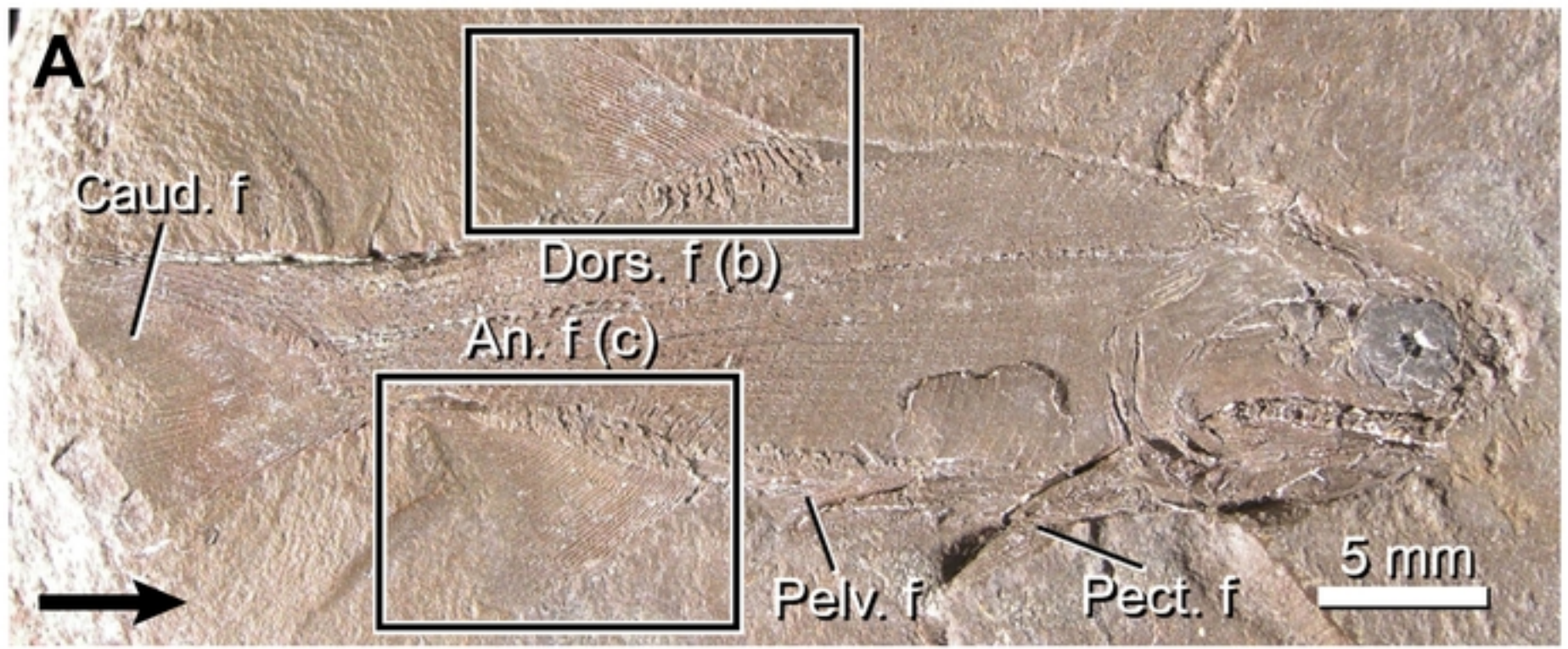
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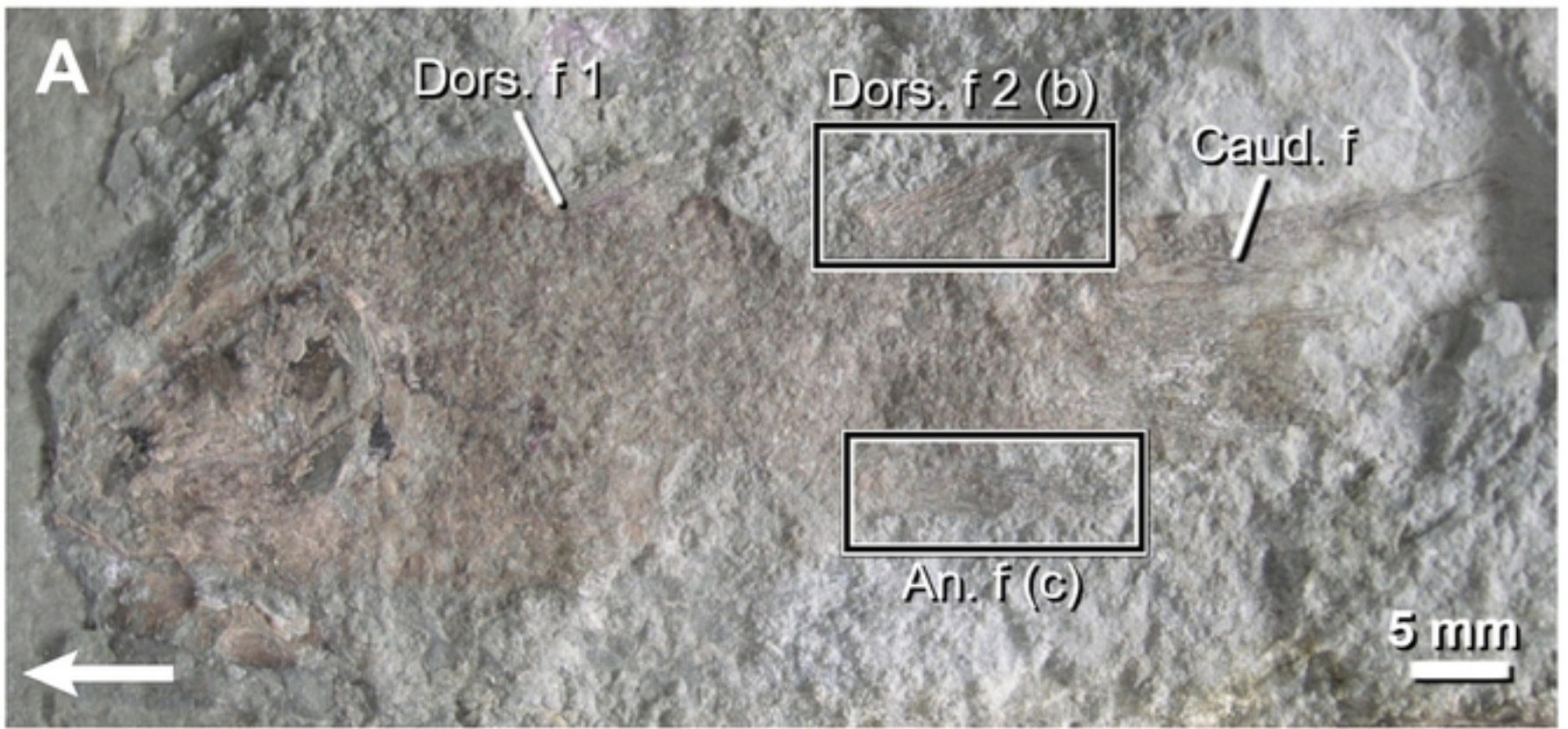


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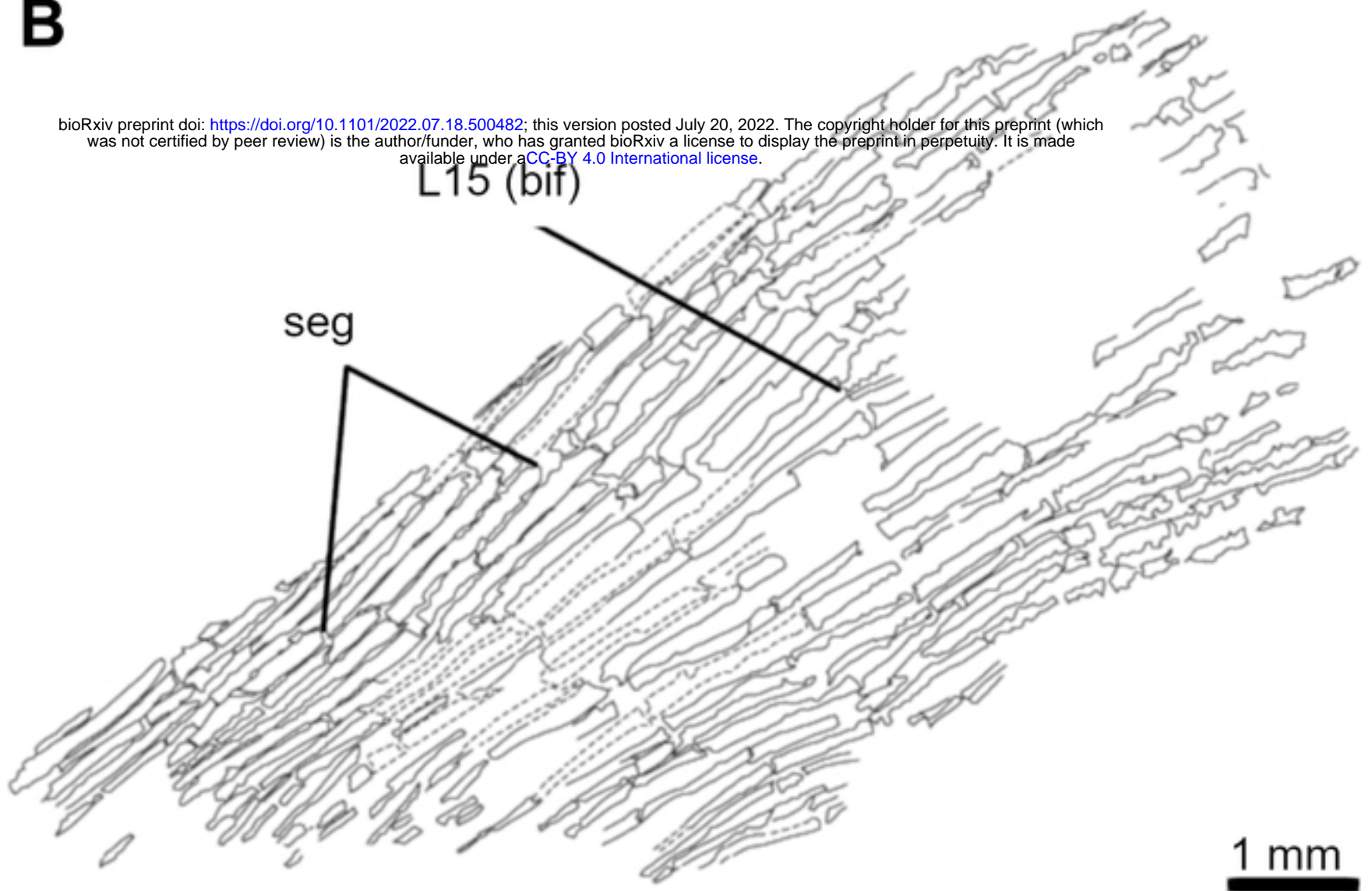




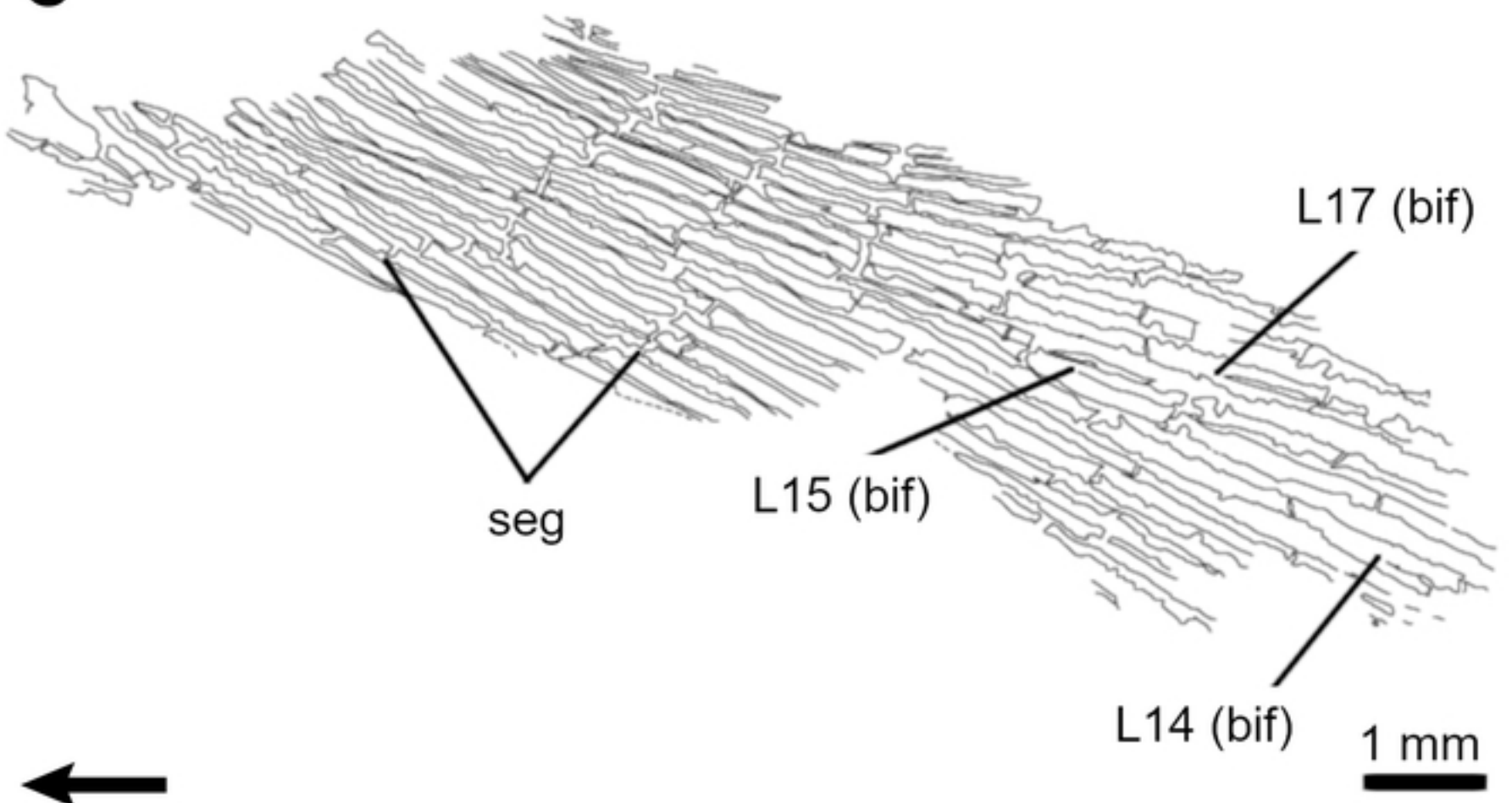


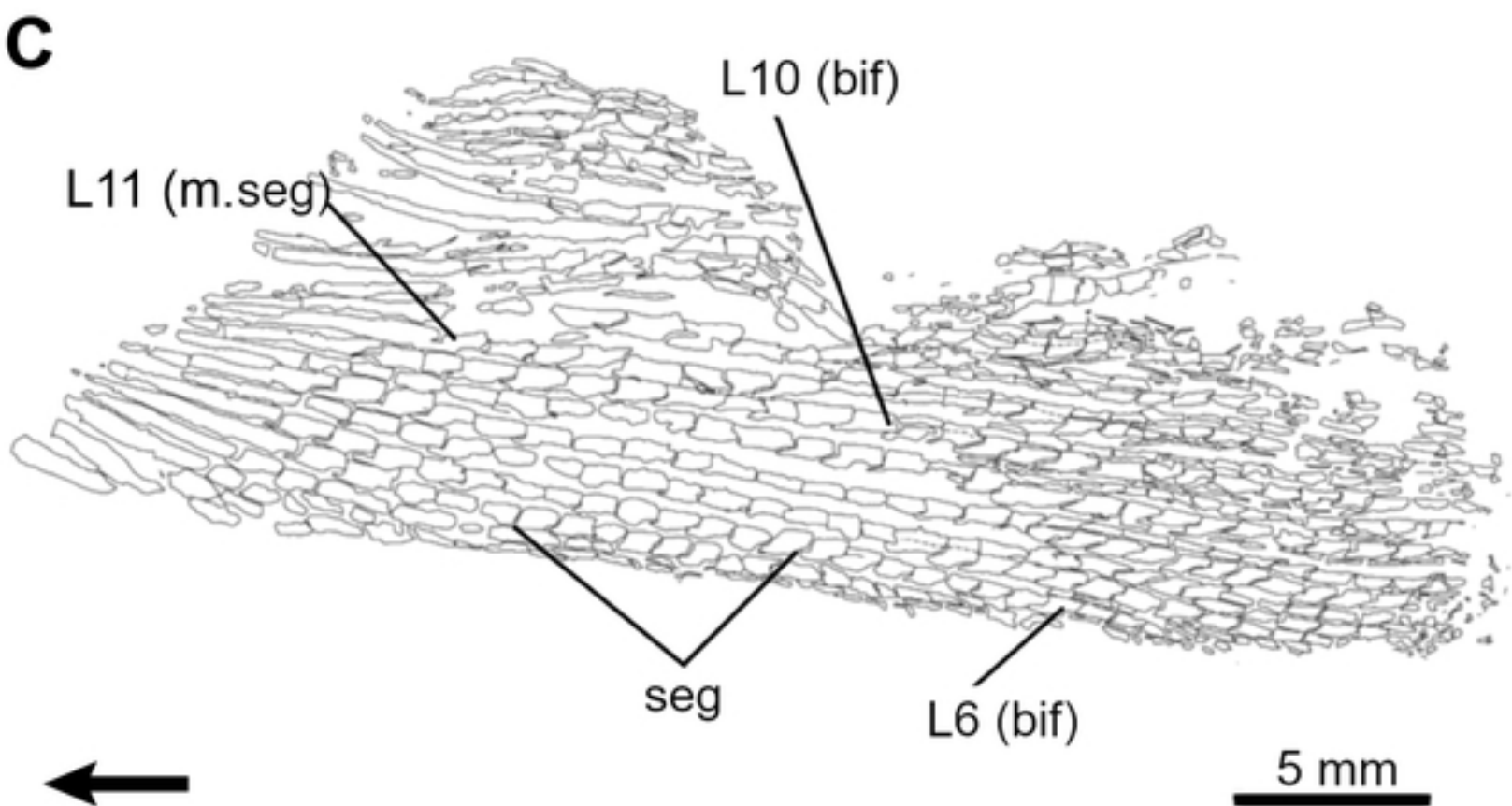
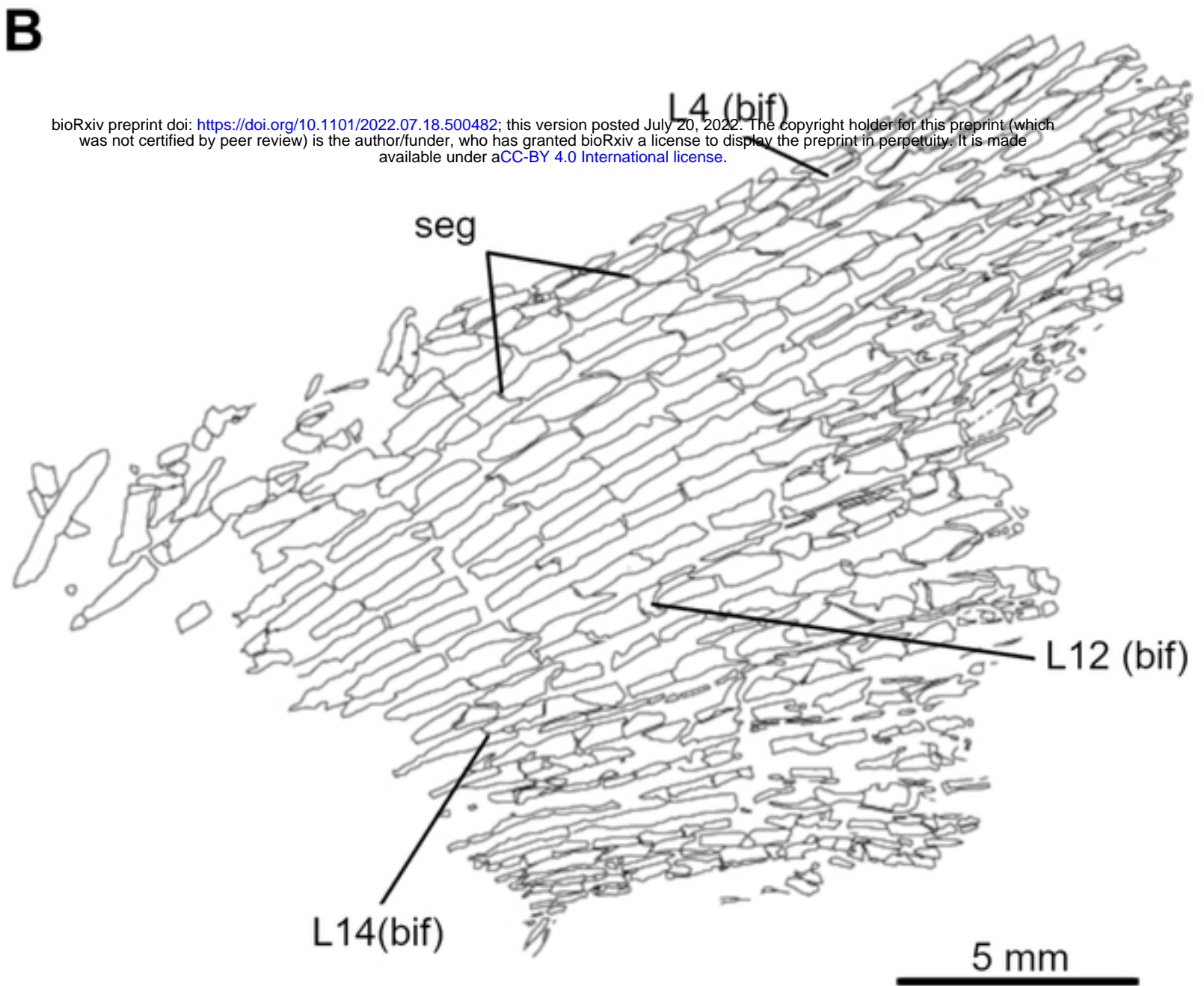
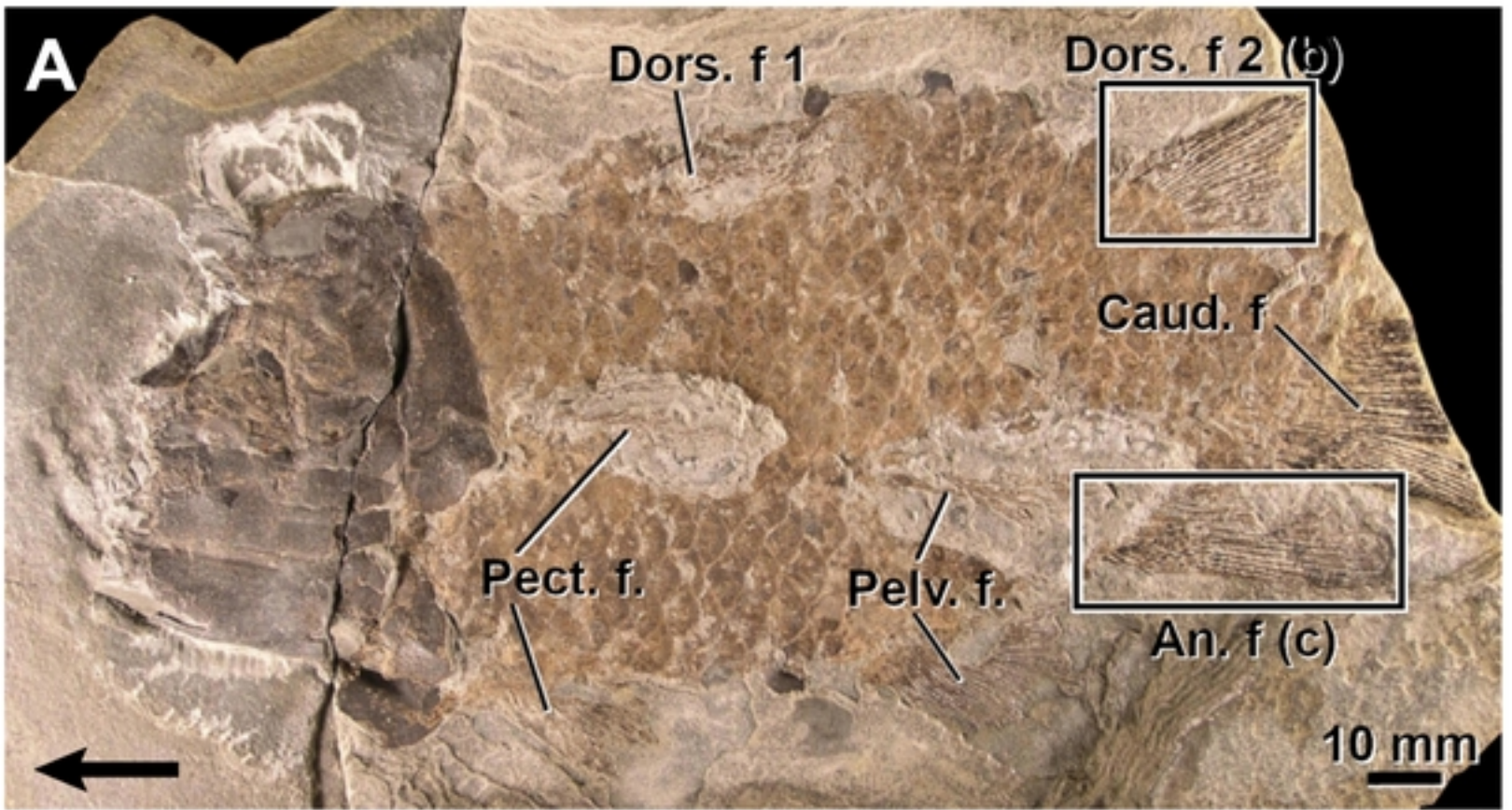
**B**

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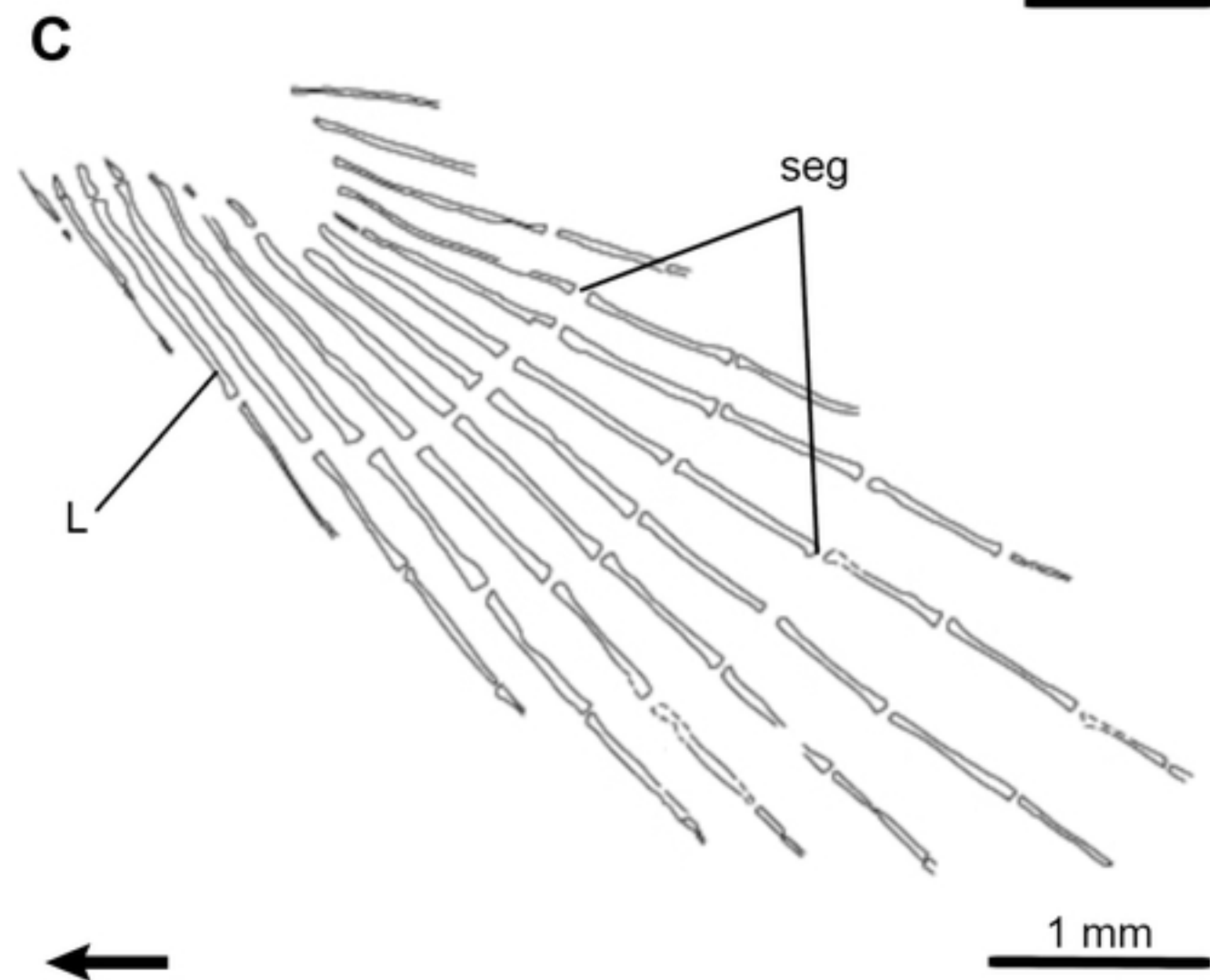
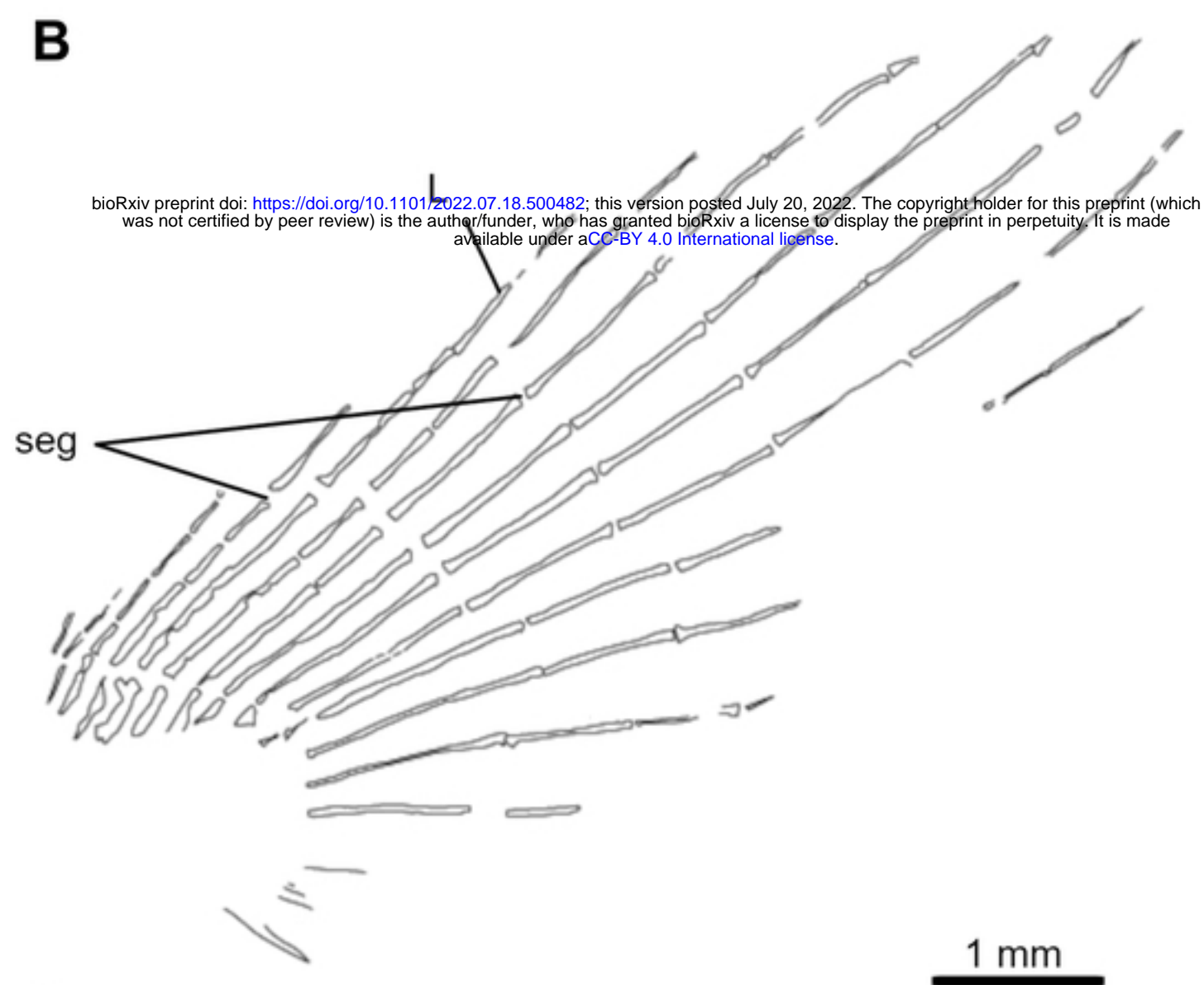
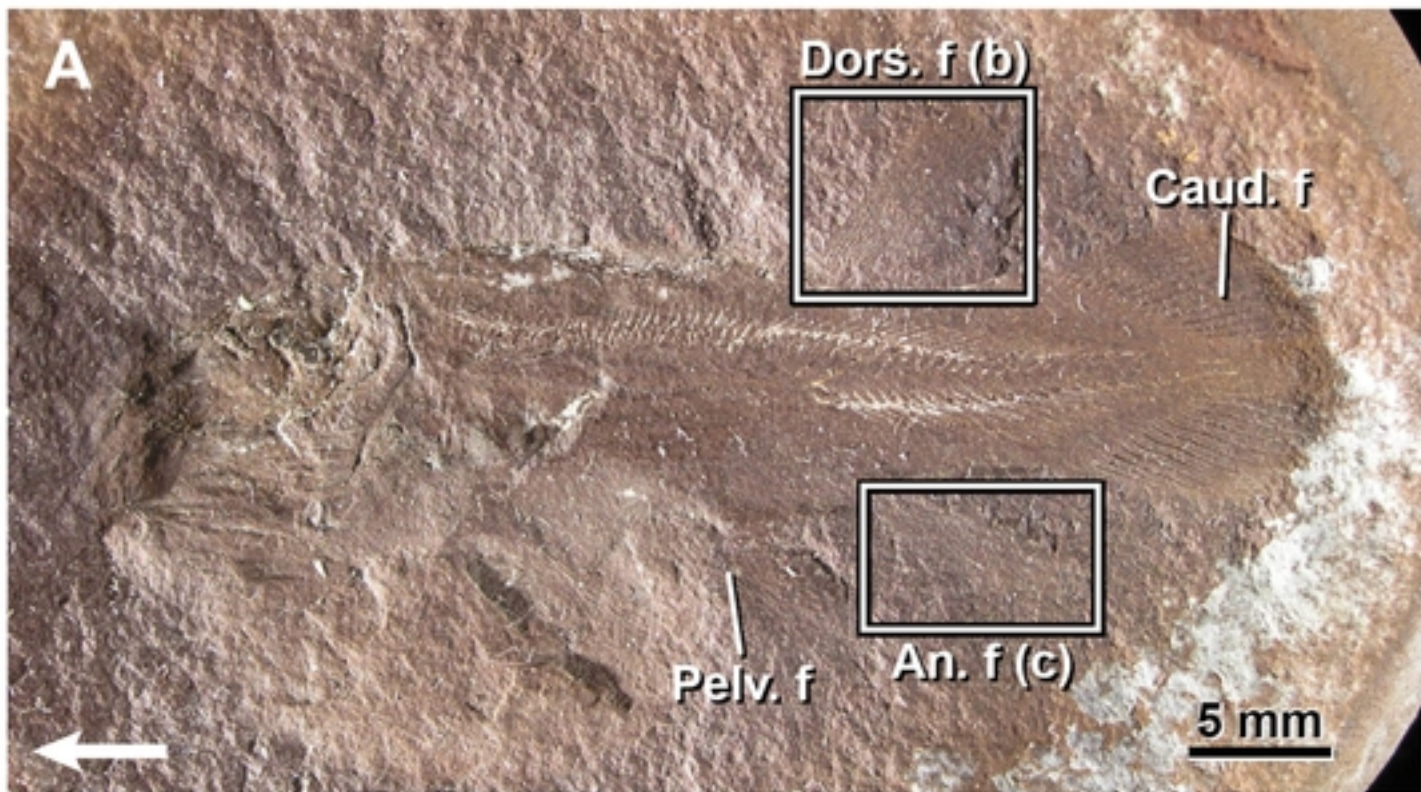


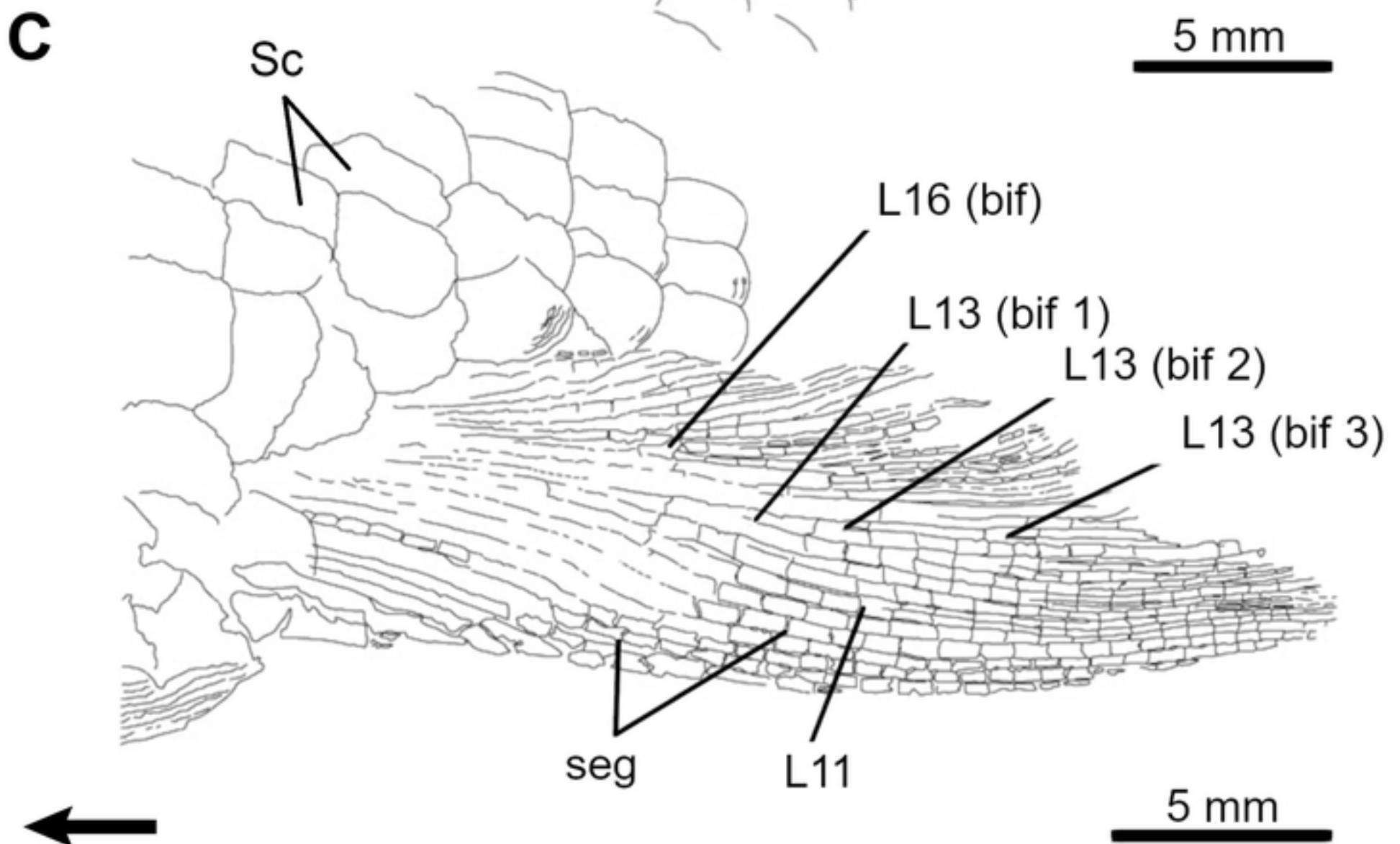
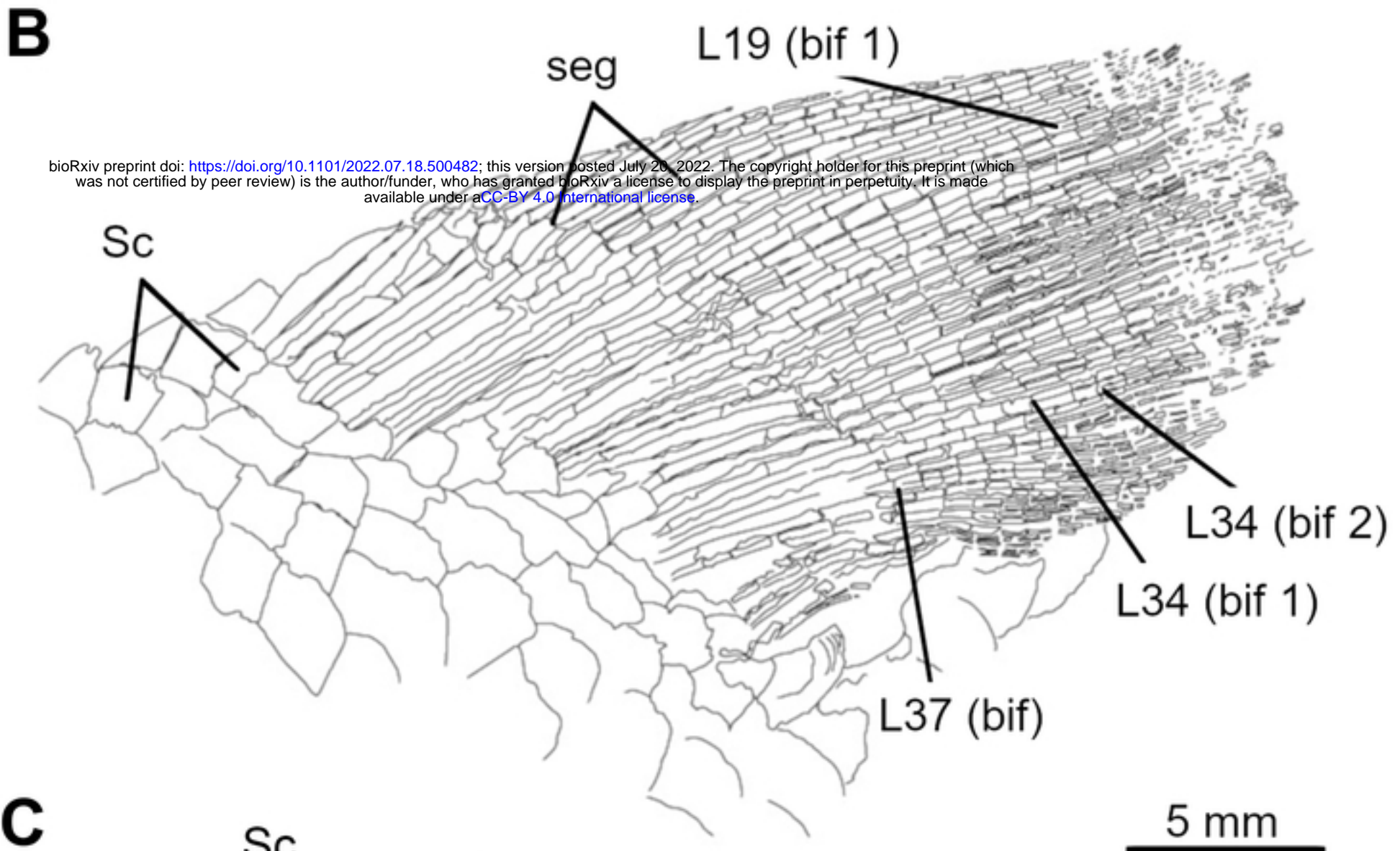
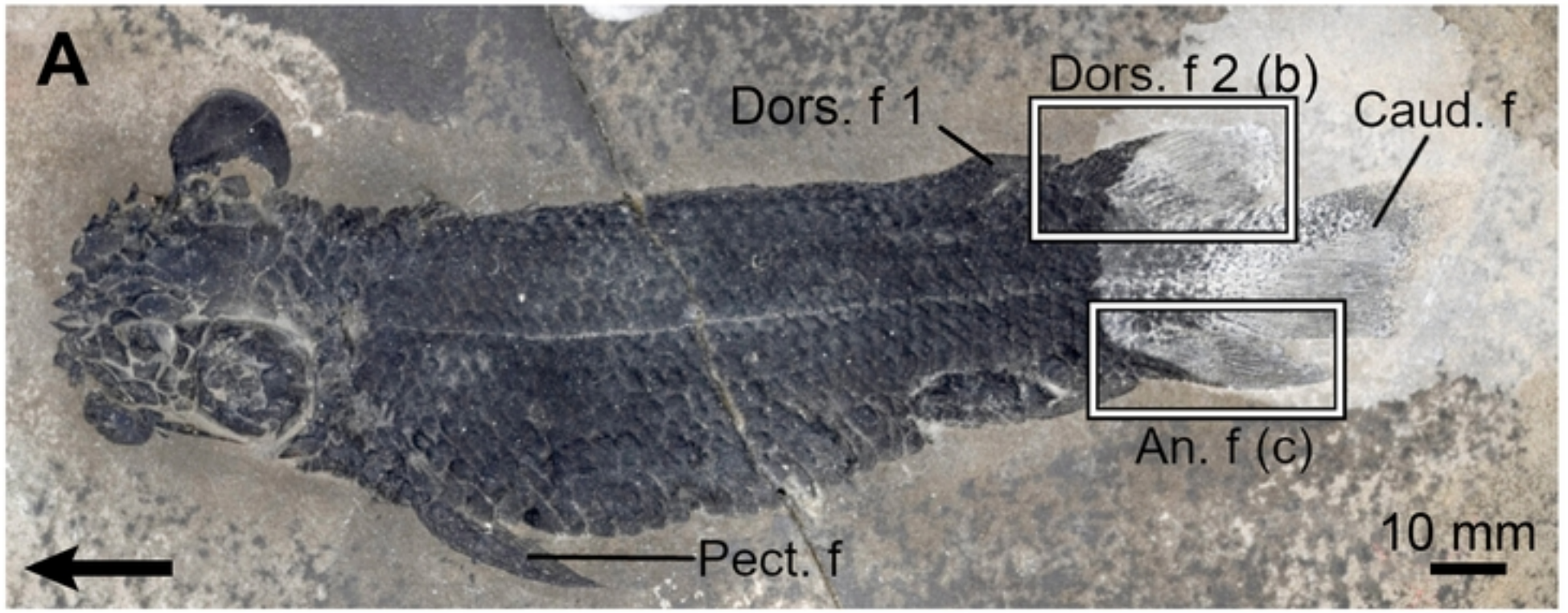
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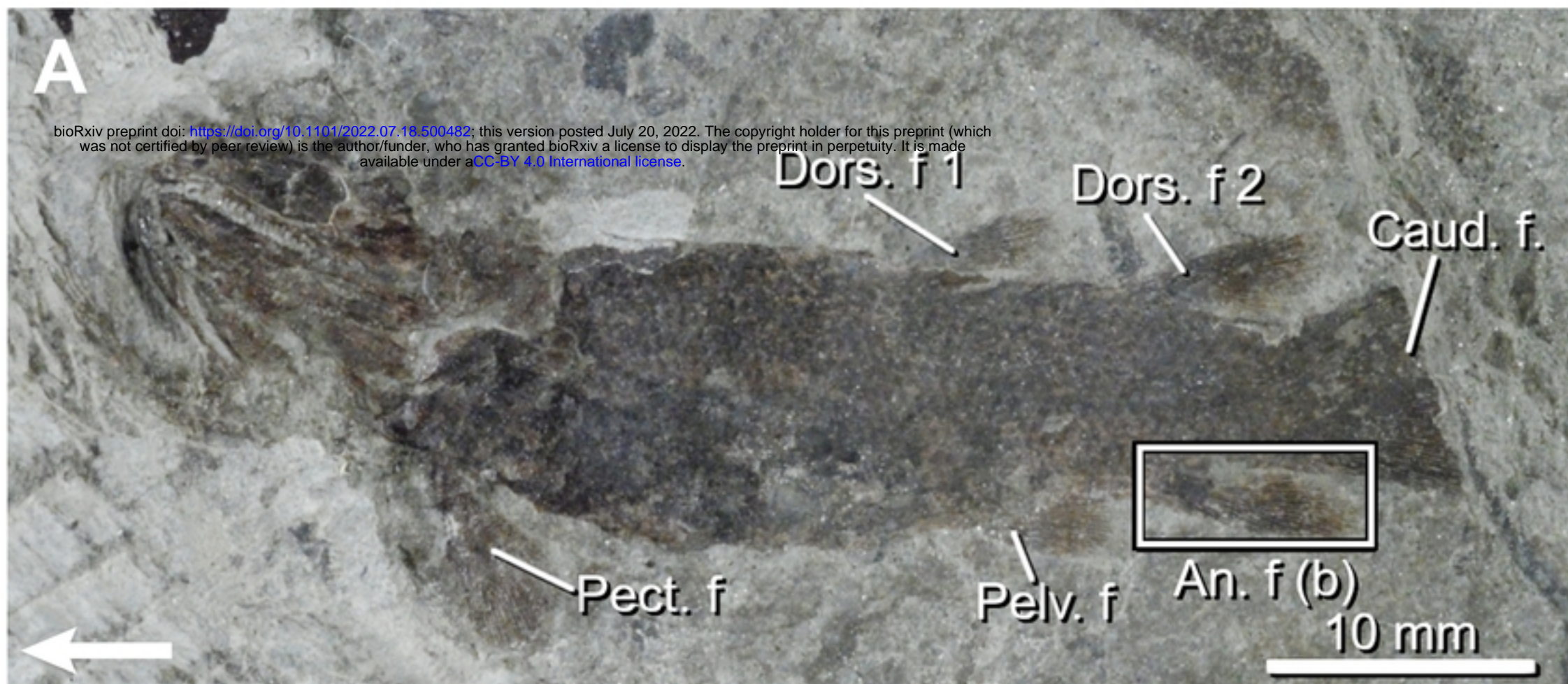
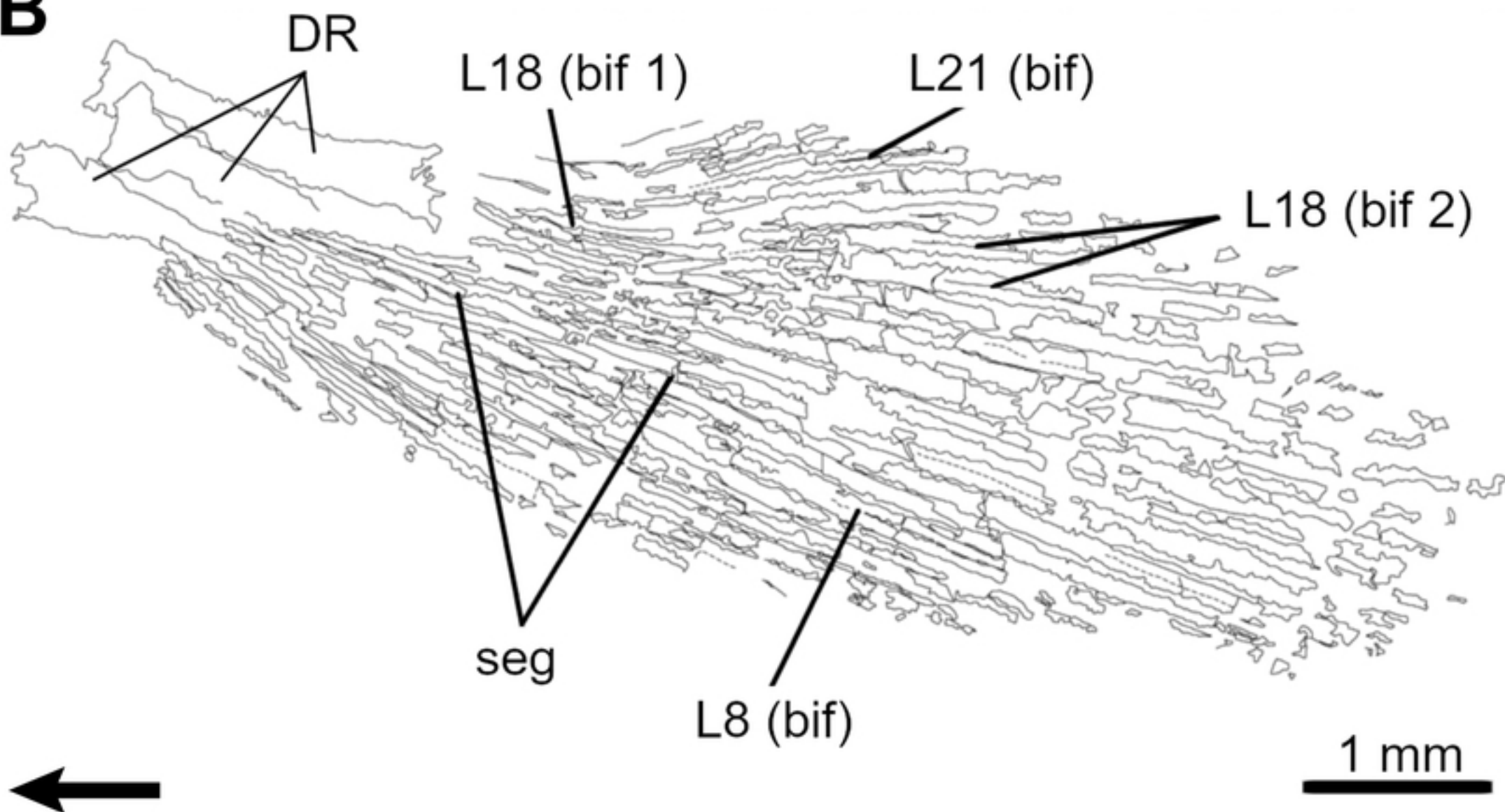


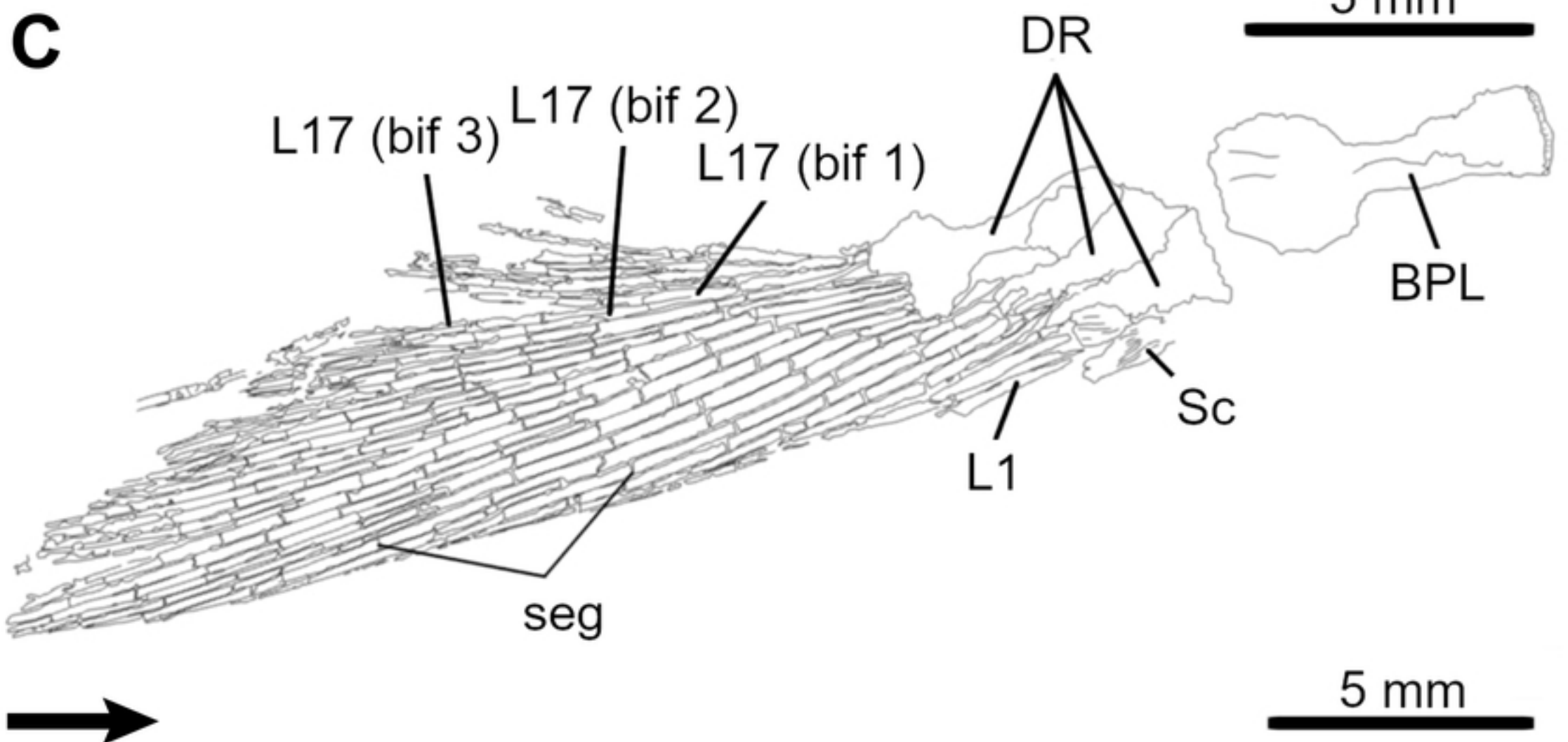
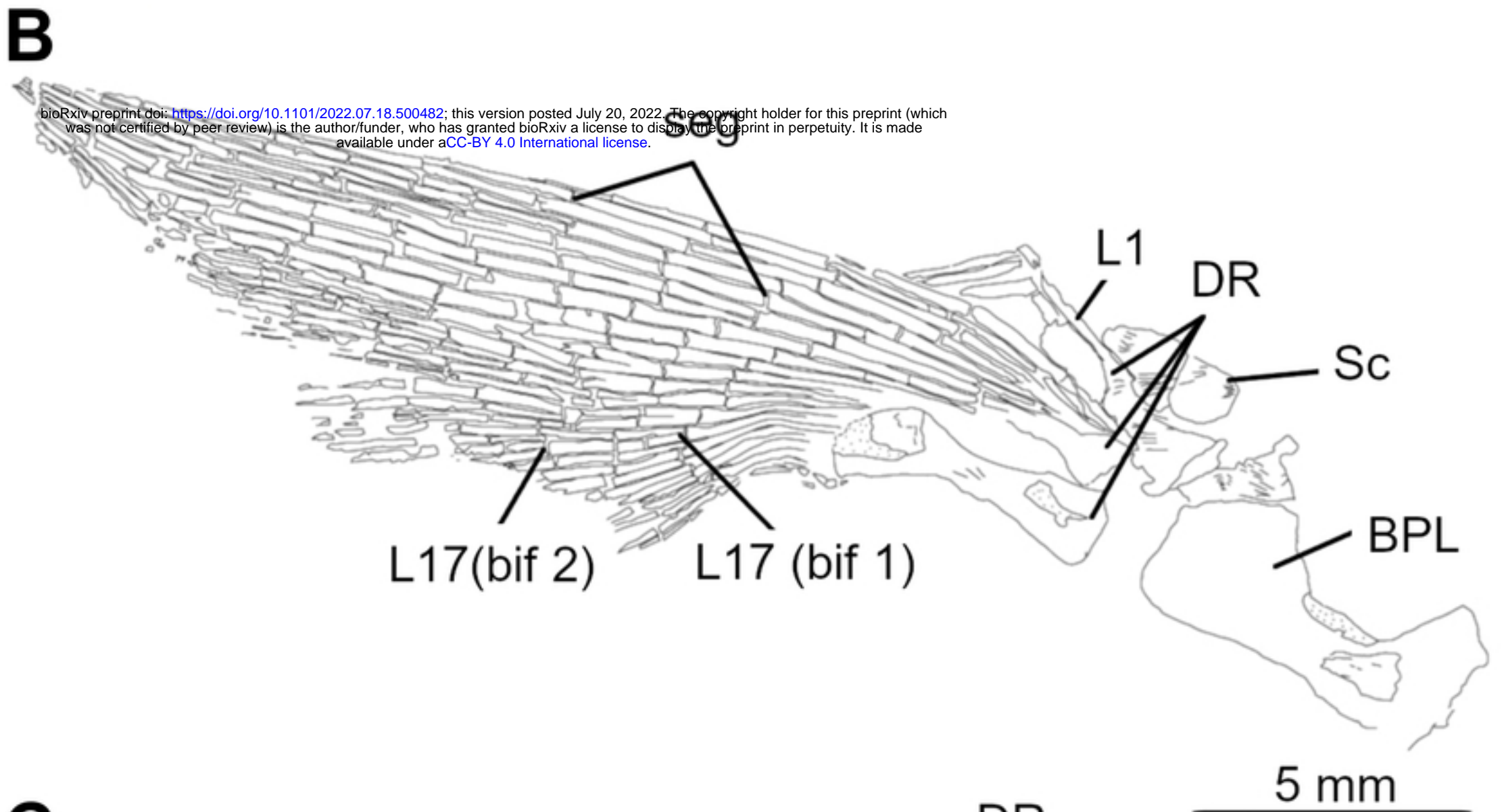
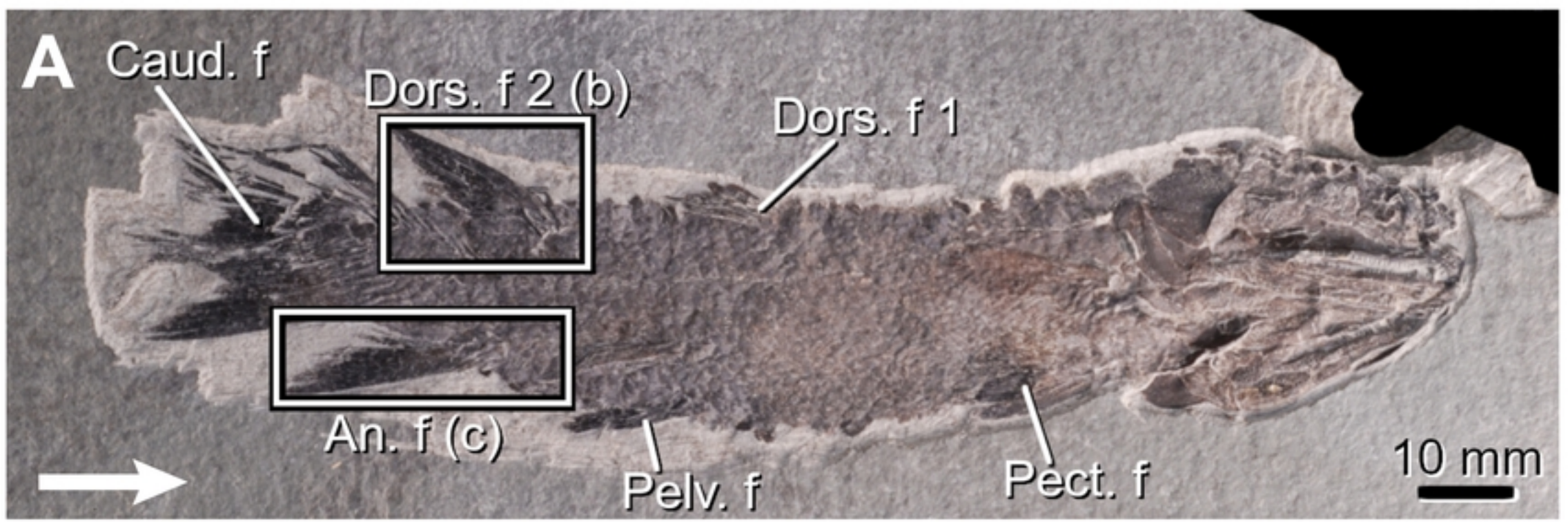


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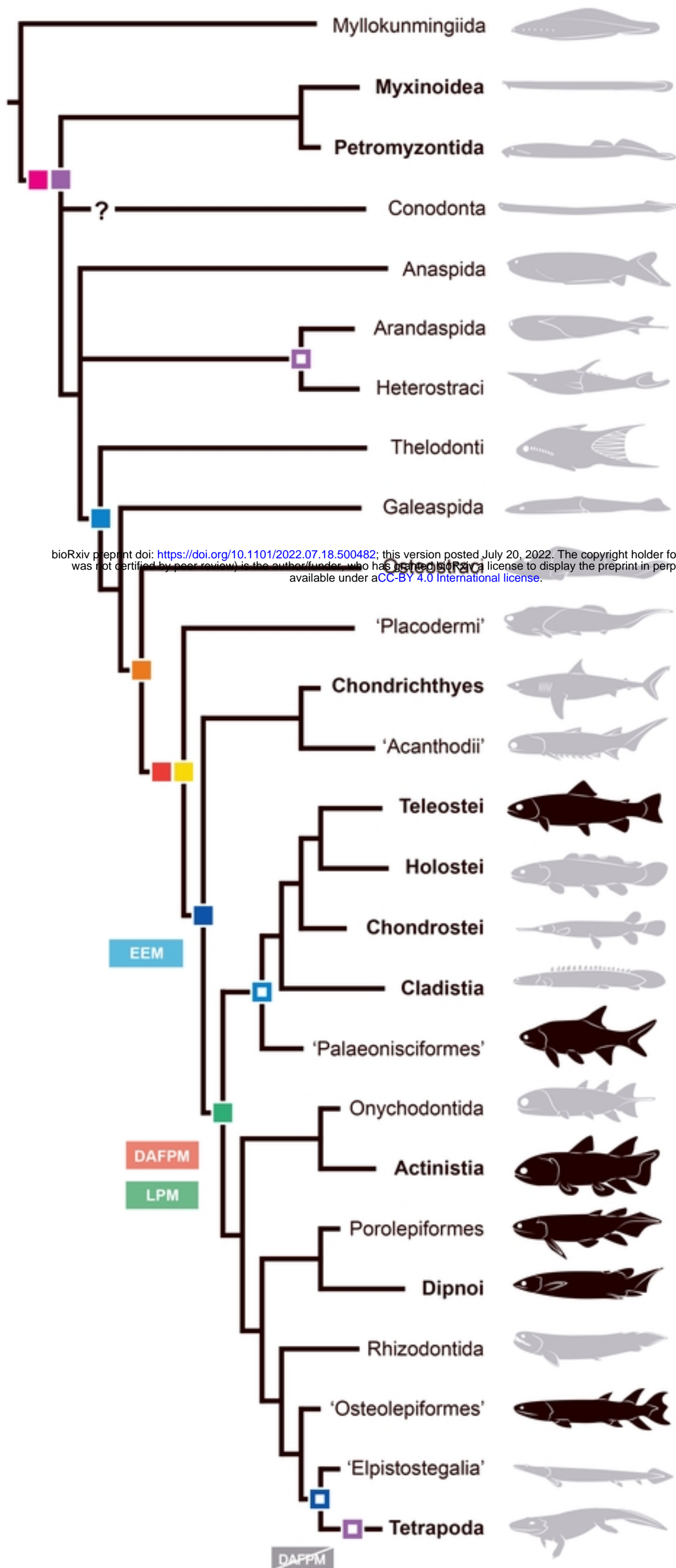
**A**

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**B**

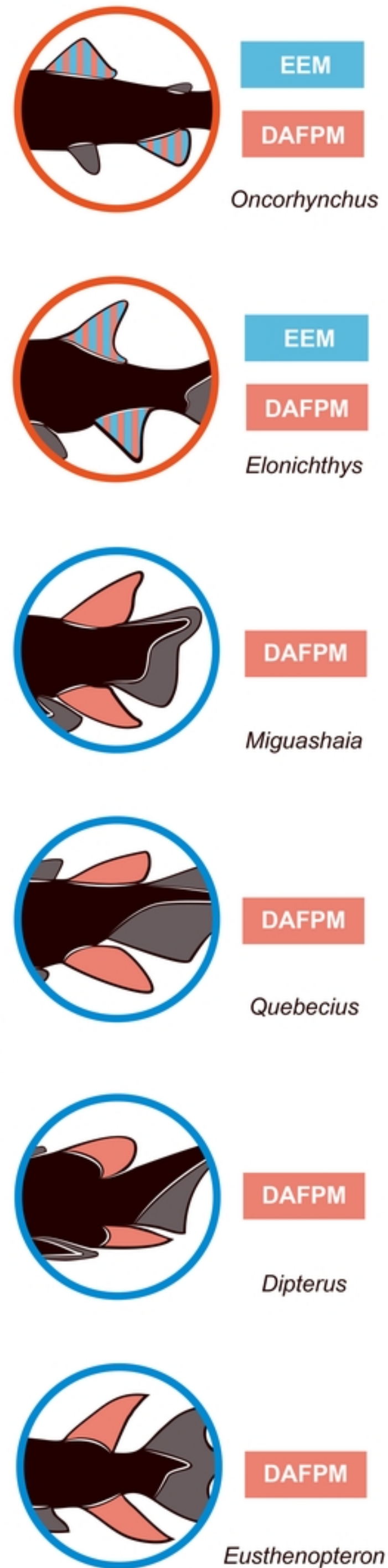


A



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B



□ Loss of second dorsal fin

□ Loss of dorsal fins

□ Loss of anal fin

■ Actino-ceratotrachia

■ Dorsal fin

■ Two dorsal fins

■ Pectoral fins

■ Lepidotrichia

■ Anal fin

■ Median fin radials

■ Pelvic fins

Gnathostomata

Actinopterygii

Osteichthyes

Sarcopterygii

EEM

DAFPM

*Oncorhynchus*

EEM

DAFPM

*Elonichthys*

DAFPM

*Miguashaia*

DAFPM

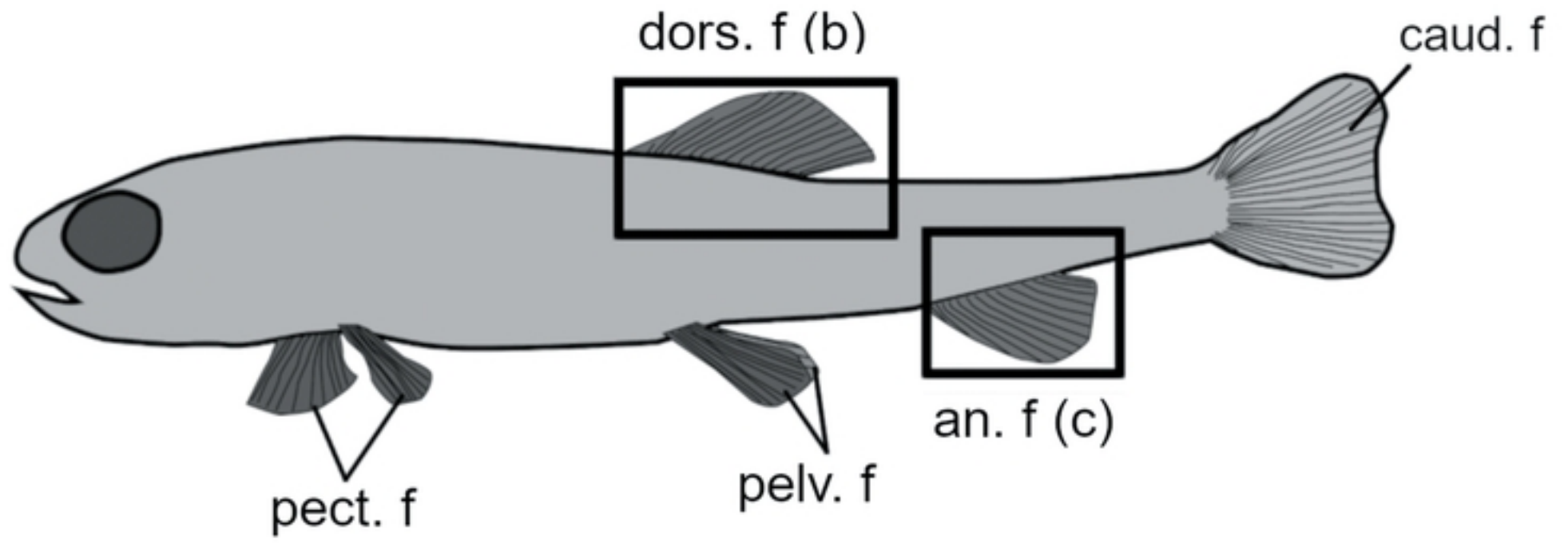
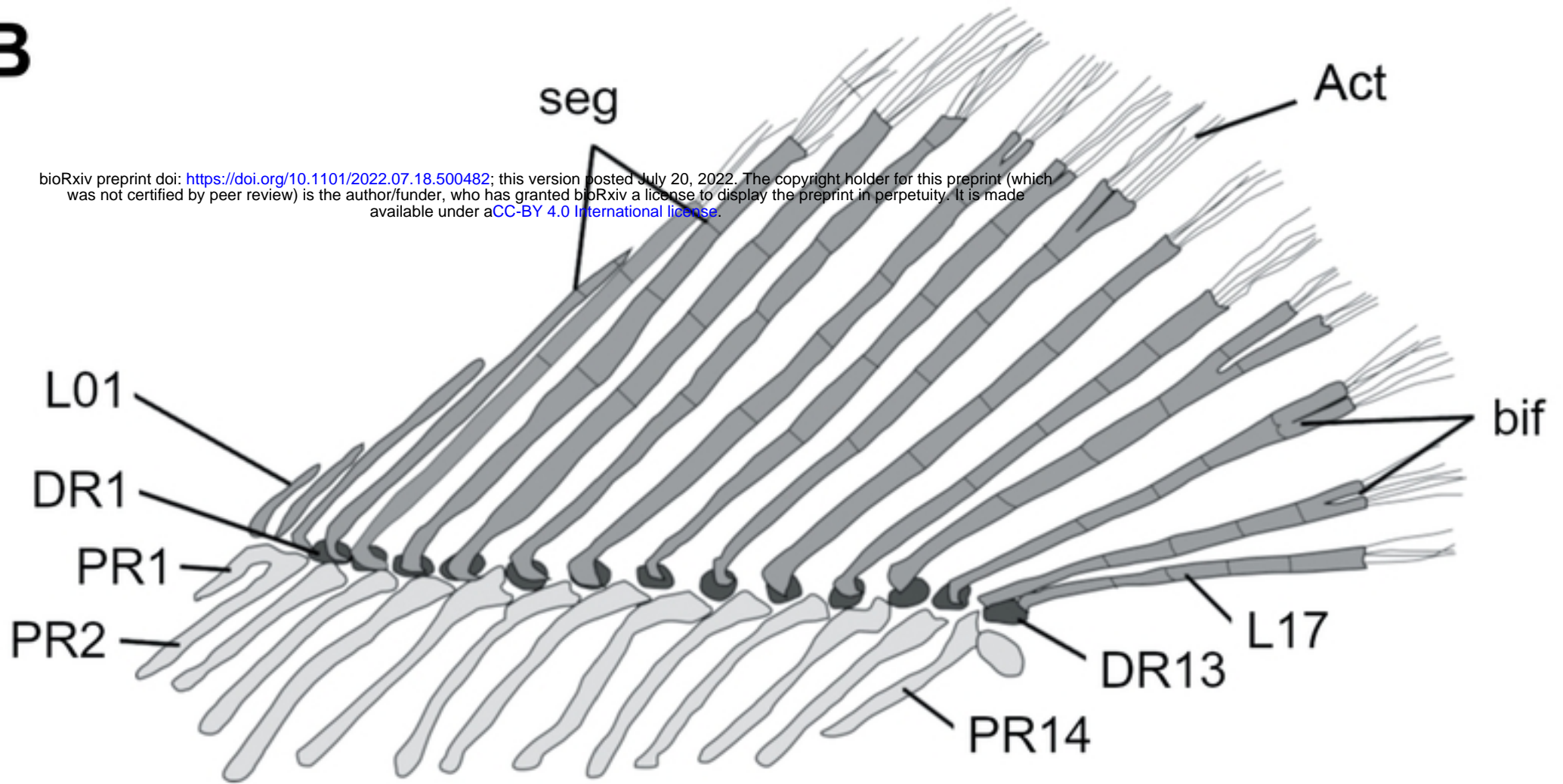
*Quebecius*

DAFPM

*Dipterus*

DAFPM

*Eusthenopteron*

**A****B****C**