

1

2

3 **Main Manuscript for**

4 Resurrection genomics provides molecular and phenotypic evidence of rapid
5 adaptation to salinization in a keystone aquatic species

6 Matthew J. Wersebe^{1,2} and Lawrence J. Weider¹

7 ¹Program in Ecology and Evolutionary Biology, Department of Biology, University of Oklahoma

8 ²730 Van Vleet Oval, Richards Hall 304, Norman, OK 73019

9 **Email:** matthew.wersebe@ou.edu

10 **Author Contributions:** This work represents a chapter of MJW's doctoral dissertation at the
11 University of Oklahoma. MJW conceived the study, conducted the research, and wrote the
12 manuscript with direction and advice from LJW.

13 **Competing Interest Statement:** No competing interest.

14 **Classification:** Evolutionary Biology; Environmental Science

15 **Keywords:** *Daphnia*, Salinization, Whole Genome Sequencing, LC50, Rapid Adaptation

16

17 **This PDF file includes:**

18 Main Text
19 Figures 1 to 4
20 Tables na
21

22 **Abstract**

23 Ecologists and evolutionary biologists are increasingly cognizant of rapid adaptation in wild
24 populations. Rapid adaptation to anthropogenic environmental change is critical for maintaining
25 biodiversity and ecosystem services into the future. Anthropogenic salinization of freshwater
26 ecosystems is quickly emerging as a primary threat, which is well documented in the northern
27 temperate ecoregion. Specifically, many northern temperate lakes have undergone extensive
28 salinization because of urbanization and the associated increase in impervious surfaces causing
29 runoff, and the extensive use of road deicing salts (e.g., NaCl). It remains unclear if increasing
30 salinization will lead to extirpation of species from these systems. Using a “resurrection
31 genomics” approach, we investigated whether the keystone aquatic herbivore, *Daphnia pulicaria*,
32 has evolved increased salinity tolerance in a severely salinized lake located in Minnesota, USA.
33 Whole genome resequencing of 54 *Daphnia* clones from the lake and hatched from resting eggs
34 that represent a 25-year temporal contrast demonstrates that many regions of the genome
35 containing genes related to osmoregulation are under selection in the study population. Tolerance
36 assays of clones revealed that the most recent clones are more tolerant to salinity than older
37 clones; this pattern is concomitant with the temporal pattern of stabilizing salinity in this lake.
38 Together, our results demonstrate that keystone species such as *Daphnia* can rapidly adapt to
39 increasing freshwater salinization. Further, our results indicate that rapid adaptation to salinity
40 may allow lake *Daphnia* populations to persist in the face of anthropogenic salinization
41 maintaining the food webs and ecosystem services they support despite global environmental
42 change.

43 **Significance Statement**

44 Rapid adaptation to human-induced environmental change is critical for preserving biodiversity
45 and ecosystem services into the future. A key question is whether populations of keystone
46 species can rapidly adapt to maintain the ecosystems they support. We investigated rapid
47 adaptation to anthropogenic salinization in *Daphnia pulicaria*, a keystone aquatic herbivore in lake

48 ecosystems. By resuscitating decades-old resting eggs, we investigate genomic changes across
49 an approximately 25-year temporal contrast from a severely salinized lake. We report that the
50 genes showing signatures of natural selection throughout the genome are related to
51 osmoregulation and ion regulation. Phenotyping clones for salinity tolerance revealed evidence
52 that genetic changes may underlie rapid evolution. We provide molecular genomic and
53 phenotypic evidence for rapid adaptation to salinity in *D. pulicaria*.

54
55

56 **Main Text**

57

58 **Introduction**

59 Ecologists and evolutionary biologists now recognize many examples of wild populations rapidly
60 evolving in the face of environmental change (1, 2). A population's ability to rapidly evolve is
61 critical for survival in the face of ever-increasing anthropogenic environmental change. This
62 capacity is especially important for organisms that provision key ecosystem services or are
63 keystone species because their extirpation would fundamentally alter ecosystem dynamics.
64 Despite this, studies demonstrating a mechanistic basis for rapid adaptation that integrates
65 information from the genome to the phenome of a population are rare (3). A key reason for the
66 paucity of such studies is that many loci of small effect are thought to contribute most to rapid
67 evolutionary change and do not align with classical "hard-sweep" models making their
68 identification difficult (4, 5). Additional analytical challenges are exacerbated by different
69 population-specific parameters (e.g., effective size or N_e) that influence the supply of new
70 potentially beneficial mutations ($\theta = 4N_e\mu$) and the resulting distribution of fitness effects ($N_e s$) (6,
71 7).

72

73 One way that rapid adaptation can be studied is using temporal genomic contrasts. Most
74 commonly, temporal contrasts take the form of so-called "evolve and re-sequence studies" which
75 follow a population across time during experimental evolution trials that typically employ
76 contrasting selection regimes (8–10). By finding the alleles that increase in frequency rapidly in
77 different treatments, the molecular basis of phenotypic shifts can be explored (8). Such studies
78 have been largely restricted to organisms such as bacteria (11) or *Drosophila* (10), which have
79 rapid generation times and are easily manipulated in the lab. Other types of temporal contrasts
80 encompass more natural experiments, such as the isolation and sequencing of ancient DNA,
81 which can give insight into past selection (12). However, with few exceptions, the genomes
82 sampled are divorced from the phenomes they produced; thus, inference is based solely on the

83 change in allele frequencies. A third way that temporal contrasts can be studied is through
84 “resurrection” studies that seek to hatch or germinate seeds, cysts or other resting stages of
85 organisms and compare genotypes and phenotypes from different points in time (13–15). For
86 instance, resurrection ecology (16–18), commonly used in animals from the freshwater
87 crustacean genus *Daphnia* has provided insight into the genetic basis of various traits (15, 19,
88 20). Thus far, however, this method has not allowed the identification of loci that can be plausibly
89 related to the phenotype under study because genetic markers are either too sparse (21) or the
90 traits under study are too highly integrated across the genome (19).

91

92 *Daphnia* are keystone species in freshwater food webs, connecting the flow of energy
93 from algal production to higher trophic levels such as fish (22, 23). Specific to many North
94 American lakes, *Daphnia pulicaria*, maintains water clarity, a key ecosystem service and supports
95 recreational fisheries with values in the millions of USD per lake per annum (24). Freshwater
96 ecosystems are among the most threatened ecosystems worldwide, impacted by various
97 anthropogenic stressors such as pollution, climate change and invasive species (25). One issue
98 threatening many freshwater ecosystems is salinization due to human activities (26), within
99 northern temperate lakes specifically salinization is particularly acute (27, 28). The causes and
100 scope of salinization have been well known (29, 30), while more recent studies have focused on
101 deciphering the ecological impacts of salinization (31, 32). In addition, we lack a more general
102 understanding of this widespread environmental issue from an evolutionary perspective, and of
103 the specific genetic architecture of adaptive responses. Such a perspective is critical because
104 recent research has shown that current water quality guidelines do not sufficiently protect aquatic
105 life from salinization (33).

106

107 To address this short coming, we sought to use resurrection ecology to study the
108 evolutionary response of *D. pulicaria* from a severely salinized lake located in Minnesota, USA.
109 Previous work on this lake has demonstrated the ecological dynamics of this population over the

110 last 150 years (31). Towards this goal, we resurrected genotypes from across approximately 25
111 years from the sediment egg bank isolated from a dated sediment core. Using whole genome
112 sequencing (WGS) of resurrected and extant individuals, we conducted numerous population
113 genomic analyses to depict population structure over time, reconstruct the demographic history,
114 and identify outlier genomic regions in the data. Additionally, we assayed a subset of genotypes
115 for tolerance to salinity. Specifically, we were interested in testing two main hypotheses; first, we
116 believed that the F_{st} outliers, or those genomic regions with extreme changes in frequency, would
117 contain genes related to osmoregulation as selection would favor higher salinity tolerance.
118 Second, this would be reflected in a higher mean tolerance of the more recent subpopulations. In
119 addition, we identify a list of candidate genes, which likely influence phenotypic variation
120 throughout the genome, and that can be targeted for further study.

121

122 **Results**

123 We studied the *D. pulicaria* population of Tanners Lake (TL; 44°57'02.2"N 92°58'54.2"W), a small
124 suburban hardwater lake located in Oakdale, Minnesota. The watershed includes approximately
125 32% impervious surfaces including parking lots, interstate highways and residential development
126 (31). TL has received significant inputs of chloride from the watershed primarily in the form of the
127 road deicer NaCl ($524 \text{ kg Cl}^- \text{ ha}^{-1} \text{ yr}^{-1}$) (34). The upper waters (i.e., surface/epilimnetic) chloride
128 concentration of TL has increased significantly in the last 75 years, from approximately 1-2 mg Cl⁻
129 L⁻¹ on average to over 150 mg Cl⁻ L⁻¹(35, 36). TL has also transitioned to a state of cultural
130 meromixis (37) with a persistent high salinity chemocline interrupting normal lake mixing dynamics.
131 We isolated and sequenced 54 *D. pulicaria* clones including 10 from the water column and 44

132 resurrected from lake sediments representing an approximately 25-year temporal contrast (~1994-
133 2019). In total we called 3,802,961 high confidence biallelic SNPs in the population.

134

135 *Population Structure and Genetic Divergence:*

136

137 Key to understanding the dynamics of the population across time is accurately describing
138 population structure to rule out possible extinction and recolonization. The 54 clones selected for
139 sequencing were separated into two clusters based on the first two principal component axes
140 explaining 7.6% and 4.6% of the variance in the LD-pruned SNP data, respectively (Figure 1A).
141 These two clusters largely separated the clones by depth with the older clones (layers 16-18 cm,
142 18-20 cm, and 22-24 cm) and more recent clones (Lake Clones, 2-4 cm, 6-8 cm, and 10-12 cm)
143 forming the two groupings. Subsequently, we decided to assign the clones into three groups for the
144 remainder of the analyses. We designated these as DEEP, encompassing all clones from 16-24
145 cm in depth (n = 18) and date from the mid to late 1990s. The second designation, called MID,
146 encompassed all clones from 6-12 cm in depth (n = 18) and date from the mid to late 2000s. The
147 third and final designation, referred to as TOP hereafter, included all clones collected from the water
148 column and 2-4 cm in depth in the core (n = 18) spanning from 2016 to 2019. We based our
149 assignment into the groupings on two observations: firstly, the TOP and DEEP subpopulations are
150 delineated by the PCA clusters, and thus warranted separate assignments. Secondly, the MID
151 subpopulation, while closely related to TOP, was intermediate in the time-scale –approximately 10
152 years prior to TOP and approximately 10 years after DEEP– and thus formed an appropriate
153 intermediate grouping. While the TOP and MID are indistinguishable using PCA (Fig 1A), their
154 temporal separation is a key feature of our study. Hence, we included each as a separate temporal
155 deme in our simulations and analysis. Conveniently, this scheme also allowed each grouping to
156 achieve equal sample size (i.e., 18 samples). Using Discriminate Analysis of Principal Components
157 (DAPC), which is a flexible group assignment method, we found good analytical support for
158 assignment of clones to the three *a priori* designated groups (Figure 1B). However, as with the

159 PCA results, generally MID clones had non-negligible assignment probabilities when compared to
160 the TOP subpopulation. Site-wise F_{st} was low across time, with an overall estimate of approximately
161 0.016 (Figure 1C). However, site-wise F_{st} estimates ranged from around 0 to as high as 0.398, with
162 most sites having an F_{st} of essentially 0. The pairwise genetic distance between the three temporal
163 subpopulations was related to the temporal distance with the TOP vs DEEP comparison being
164 highest (0.019) and the MID population being intermediate to both (Figure 1D). Patterns of
165 nucleotide diversity (π) were similar across all temporal subpopulations. Mean nucleotide diversity
166 ranged from 0.0066 for DEEP, to 0.0070 for MID and 0.0069 for TOP (Figures S3-5).

167

168 *Estimation of Effective Population Size and Simulations:*

169

170 To accurately parameterize tests for selection, we sought to estimate a demographic model for the
171 TL population. We were able to estimate the effective population size of the TL *D. pulicaria*
172 population using both linkage disequilibrium (LD) and coalescent simulations. The LD-based
173 results using just samples from the lake clone (LC) isolates showed that over the last few hundred
174 generations, there was a period of population expansion and contraction (Figure S1). The
175 population reached a peak N_e of 4000-6500 approximately 150 generations ago and the population
176 has contracted recently to an N_e of around 2000. For this analysis, we interpret “generations” here
177 to be sexual generations (i.e., LD is related to recombination and asexual generations are
178 ameiotic). Since sex may occur once or at most a few times per year in stable lake habitats (38,
179 39), we interpret a single generation to be equivalent to one year. The coalescent-based FSC2.7
180 (40) run with the highest likelihood estimated N_e to be 2931 individuals and had a signal of
181 population contraction with a population growth rate of 2.629×10^{-5} . The estimate of effective
182 population size was within the 95% confidence for 100 parametric bootstraps; however, the
183 estimate for population growth rate was not and the confidence intervals included zero (table S1).
184 We ran simulations in FSC2.7 using the maximum likelihood estimates to establish expectations
185 for F_{st} based on the modeled demographic parameters. The results from 100 independent runs of

186 FSC2.7 were pooled to develop a distribution of expected F_{st} values from simulated ~110000 SNPs.
187 Testing each LD-pruned SNP against this distribution and correcting for multiple testing using false
188 discovery rate (FDR) resulted in 178 outlier SNPs with corrected one-tailed p-values above a $p =$
189 0.05 significance threshold (Figure 2). There were outliers on every chromosome, ranging from a
190 low of 2 SNPs (CHR 08) to a high of 42 (CHR 04).

191

192 *Genes surrounding F_{st} outliers and GO term enrichment and Variant Annotation:*

193

194 One of our primary hypotheses was that the genes surrounding F_{st} outliers would be related to
195 osmoregulation and salinity tolerance. We searched for genes within 10Kb of the F_{st} outlier SNPs
196 (± 5 KB centered on the SNP) and in total we extracted 286 genes near these SNPs with known
197 function in the *D. pulicaria* genome. GO term enrichment analysis with PantherDB webtool (41)
198 yielded 59 enriched terms for this list of genes after correction for FDR (Figure 3). The enriched
199 terms and p-values are available in table S2. Notable among the enriched terms for molecular
200 function are chloride channel activity (GO:0005254; $p = 0.00925$); however, many different ion and
201 channel terms were enriched. After running variant effect prediction (42) on all the SNPs called in
202 the population, we identified 17181 variants with “high” predicted effects. Intersecting this list with
203 outlier genes, we found 78 of the 286 outlier genes had high effect variants, only one of these,
204 Chloride Channel 2 isoform X2 (*clcn2-x2*), located on chromosome 5 had any obvious relation to
205 osmoregulation or salinity tolerance. *Clcn2-x2* has five SNPs of high effect, including four
206 premature stop codon changes and a splice donor change that would likely severely interrupt
207 protein function. It is tightly linked to the F_{st} outlier site at CHR05:12238, which is an intronic SNP
208 within *clcn2-x2*. In addition to the high effect mutations, this gene has a total of 28 missense
209 mutations and 37 synonymous mutations classified as moderate and low impact respectively. Of
210 the clones surveyed for chloride tolerance (see next section), we found that the two most tolerant

211 clones were homozygous for the wildtype (i.e., functional) allele at *clcn2-x2*. Overall, 252 of the
212 outlier genes had effects that included moderate impacts to function such as a missense SNP.

213

214 However, *clcn2-x2*'s behavior as an outlier is not consistent with a single large effect locus.
215 The SNP at CHR05:12238 is the 88th most differentiated SNPs among the 178 outliers with an
216 overall F_{st} of just 0.1985 (FDR corrected $p = 0.0269$). The top 11 outlier SNPs (F_{st} 0.3983 – 0.3175)
217 all had F_{st} unobserved under neutral demography. One of the SNPs is an intergenic variant, while
218 the 10 remaining SNPs are intronic SNPs associated with 13 different genes, 10 of which have
219 annotated functions (table S3). The SNP located at CHR04:13394600 is the most differentiated
220 SNP observed and is associated with the genes *pank4* and *tda6*. The remaining genes span across
221 functional categories including the structural protein collagen alpha-1 chain and the calcium-binding
222 protein *fstl5*, to several genes related to development comprising *still life* (43), *daam1* (44), and *pnt*
223 (45), which contains two outlier SNPs, *rap1gtp* (46), and *rotund* (47).

224

225 *Chloride Tolerance LC₅₀:*

226

227 We expected that increasing chloride pollution in TL would result in higher tolerance of more recent
228 clones. Therefore, we conducted clone-specific assays to estimate 96-hour Lethal Concentration-
229 50% (LC₅₀). Clonal tolerance ranged from a low of 584.91 mg/L Cl⁻ (clones 10-12-12A & 18-20-04A)
230 to a high of 1047.16 mg/L Cl⁻ for (Clone LC-06). We observed a main effect of subpopulation in the
231 Kruskal-Wallis test ($p = 0.006$; Figure 4). Post-hoc testing using a pairwise Wilcoxon test found that
232 the Lake subpopulation (i.e., TOP) was more tolerant on average than either the DEEP (22-24 cm
233 & 18-20 cm) or MID (10-12 cm) subpopulations ($p = 0.0076$). The DEEP and MID subpopulations
234 did not differ from one another ($p = 0.9073$).

235

236 **Discussion**

237

238 Our resurrection ecology (RE) study across ~25-year temporal contrast has provided phenotypic
239 and molecular evidence of rapid evolution of salinity tolerance in the TL *Daphnia pulicaria*
240 population. Using whole genome resequencing to analyze population genetic structure and
241 demographic history, we have identified genomic regions putatively under selection, with support
242 from LC₅₀ chloride tolerance assays. We found that Chloride Channel-2 isoform X2 (*clcn2-x2*) has
243 a unique mutational history and may be affecting salinity tolerance in this population, as one of
244 many genes under selection in this population. We find support for both of our main hypotheses
245 — genes related to osmoregulation are enriched in outlier regions and that the population shows
246 increasing tolerance to salinity over time.

247

248 *Demographic History and Subpopulation Structure:*

249

250 A goal of our study was to establish a demographic model for the TL population. This would
251 enable the modeling of population genetic summary statistics (i.e., F_{st}) for establishment of a
252 distribution of expected or null estimates of F_{st} -based on the demographic history (discussed
253 below). We used two complementary methods – LD-based methods and coalescent simulation to
254 understand the recent demographic history of the TL population. Each method provides
255 incomplete information, and each has unique biases that need to be considered. Furthermore,
256 despite N_e being a critical parameter in population genetics, it is notoriously difficult to estimate
257 (46). For instance, it appears that SFS (coalescent) based methods are underpowered for recent
258 demographic history; however larger sample sizes may ameliorate some of these effects (47).
259 This may also explain why it was difficult to accurately estimate the population growth rate
260 parameter. The LD-based method we employed here appears to be strongly affected by the
261 number of homozygotes in the sample (48), and thus, is sensitive to analyzing genomes from
262 different generations. This prevented us from modeling each subpopulation separately.
263 Additionally, it also appears to be inaccurate for the first few generations, giving unrealistically
264 small numbers (see Figure S1). As such, we used these methods in tandem to increase the

265 confidence in our estimation of the demographic parameters. Both methods were concordant that
266 recent N_e was approximately 2000-3000 individuals, and both showed signatures of recent
267 population contraction. This effective population size is not unexpected for *D. pulicaria* because
268 populations are thought to delay sex and recombination for long periods of time (a year or more)
269 (36, 37). During this time, clonal selection is thought to winnow down the population to a
270 collection of ecologically equivalent clones (49). These facts, taken together with the current
271 understanding of zooplankton metapopulation dynamics (50, 51) suggests that *Daphnia* effective
272 populations should be small and insular.

273

274 Despite the small and insular nature of the TL population, it appears that drift does not
275 always predominate. Our PCA results demonstrate at least on a decadal scale (i.e., between MID
276 and TOP) that the population is temporally consistent. This likely reflects the dominance of the
277 few ecologically equivalent clones predicted by Lynch and Spitze (48). It is likely that only after a
278 rapid or pronounced change in environment can one expect to see a population structure across
279 time, as reflected in other studies of *Daphnia* utilizing RE (15, 52). Salinization in TL is ongoing
280 and likely started in the mid-20th century with the onset of widespread use of road deicing salts in
281 the 1950s (53). The sediment intervals dating to the early 1990s from which we recovered the
282 oldest samples in this study reflect a period of change in the sediment egg bank (e.g., low
283 ephippial fluxes) (30) and suggests a period of rapid environmental change in this lake. Wersebe
284 et al. (30), reported that by the early-1990s, TL had already reached a surface (waters) chloride
285 concentration of at least 100 mg/L. Thus, all the source periods examined in the present study
286 are typified by elevated chloride levels. Since we were unable to hatch eggs from before
287 salinization commenced or was comparatively low in TL, it is impossible to determine if the
288 structuring we observed was ongoing or more sudden. It is most likely that our temporal samples
289 represent a process of ongoing adaptation to very rapidly increasing salinization. Regardless,
290 since the clones are closely related across time at both the genomic and mitochondrial levels, it

291 remains unlikely that the population became extinct and was recolonized by migrant genotypes
292 (Fig. 1C; S2).

293

294 *Outliers and the Identification of Candidate Genes:*

295

296 The specification of a demographic model for the TL lake population allowed us to establish the
297 presence of statistical significance for each site in the genome scan. This analysis revealed 178
298 regions with F_{st} beyond what we could reasonably expect based on demography alone. Genes
299 surrounding these outliers had many functions, but as expected, genes involved in
300 osmoregulation were among the most enriched in the dataset according to GO terms for
301 molecular function. The presence of hundreds of outlier genes suggests that salinity tolerance
302 has a complex genetic architecture and that tolerance to increased salinity requires changes at
303 many loci of small effect. Transcriptomic studies of clonal isolates from within the *D. pulex*
304 complex (to which *D. pulicaria* belongs) support this assertion (54). Indeed, cross-referencing
305 these outlier genes with different mutational types (e.g., missense SNPs) showed that many of
306 these genes have sites that may be under selection. However, the only gene with a known
307 functional role in osmoregulation with high effect mutations was *clcn2-x2*. Chloride channels play
308 a key role in osmoregulatory physiology of all animals. *Daphnia* are known as hypo-
309 osmoregulators, meaning they attempt to maintain their hemolymph solute concentration above
310 the ambient media concentration (55). Osmoregulation occurs within the gill-epithelium of
311 *Daphnia*, and chloride channels like *clcn2-x2* play a major role in shuttling Cl⁻ ions into the
312 hemolymph across the basolateral membrane of the gill epithelium to maintain a hypo-osmotic
313 concentration in the hemolymph (56). The constellation of premature stop codon mutations in this
314 gene means that the protein is very unlikely to function properly because the channel is too short
315 to pass through the cell membrane. Individuals are at least heterozygous for each of the four
316 premature stop codon SNPs and the wild-type gene sequence annotated in the reference. Thus,
317 they can produce a functioning protein. The two most tolerant clones in this study were both

318 homozygous for the functional allele. This suggests that *clcn2-x2* is a critical gene requiring at
319 least one functional copy and it explains some portion of the variance in acute salinity tolerance.
320 However, a larger sample of genotyped and phenotyped individuals would be required to further
321 validate this statistically.

322

323 The most differentiated genes in the genome were involved in a few key functions. There
324 was a concentration of outlier SNPs in genes involved in regulating Rho-like GTPases which are
325 involved in the regulation of actin filaments and neuronal development. The reason why these
326 genes are the most differentiated is not initially clear. One potential explanation involves
327 phenotypic plasticity. Increased salinity tolerance may require the accommodation of new
328 developmental trajectories through phenotypic plasticity. Evolution via plasticity (59) requires that
329 central developmental pathways serve as fuel for phenotypic differentiation (60). Furthermore,
330 plasticity is thought to play an important role in rapid adaptation in *Daphnia*, a pattern we observe
331 in the TL population (61–63). Regardless, testing of this assertion would be difficult because
332 unlike other model arthropods, *Daphnia* development is not as well explored.

333

334 *Chloride Tolerance and Rapid Evolution:*

335

336 The clones observed in this study vary nearly two-fold in salinity tolerance (585 – 1047 mg/L),
337 with the most tolerant clones detected in the most recent (TOP) temporal subpopulation. It is
338 important to note that all clones in this study come from natal conditions that included elevated
339 salinity with all subpopulations likely experiencing surface chloride conditions of 100-150 mg/L
340 (31). Surface chloride conditions in TL are unlikely to exceed 150 mg/L in a year because of the
341 dynamic equilibrium between annual loading and flushing of chloride in the system (49).

342 However, TL more recently has transitioned to a state known as “cultural meromixis” where the
343 accumulation of Cl⁻ at depth has interrupted normal lake mixing. For instance, in July 2019, we
344 observed inferred Cl⁻ concentrations of approximately 275 mg/L directly above the chemocline

345 (47% of the lowest LC₅₀). Below the chemocline, inferred concentrations approached 483 mg/L
346 (82% of the lowest LC₅₀). Large-bodied *D. pulicaria*, undergo diel vertical migration to avoid
347 visually oriented predators like fish. This vertical movement means that a clone may have to
348 accommodate fluctuations in the ambient chloride concentration of 100s-of-miligrams over the
349 course of 24-hrs. We attempted to study the degree to which this might occur in the summer of
350 2021 (see the supplemental materials for details). We conducted a Diel Vertical Migration (DVM)
351 study in TL to track the spatial and temporal patterns of *D. pulicaria* distribution in the water
352 column. We observed that *D. pulicaria* do inhabit the deepest, saltiest parts of the water column
353 up to 12-m (Figure S6). However, in 2021, the chemical stratification of TL was much weaker
354 than what we observed in 2019 and DO was not depleted at depth (see 31, Figure 1A-C & Figure
355 S7 A-C). We believe the reason for the weaker chemical stratification is that the area surrounding
356 TL (Washington and Ramesy Counties, MN) were classified as being abnormally dry on
357 06/29/2021 and had been classified as such intermittently since 09/29/2020. Thus, our abilities to
358 determine the extent to which this might be reflected in a higher salinity year are diminished.
359 Regardless, this shows that a portion of the *Daphnia* population is moving to more anoxic and
360 saltier layers as part of the normal DVM behavior.

361 Such elevated Cl⁻ concentrations routinely observed in TL at depth are likely to have
362 several sublethal effects that reduce clonal fitness. Presumably, a clone that has high acute
363 tolerance has the physiological capacity to ameliorate the sublethal effects of increased chloride
364 and we attempted to approximate this physiological response with LC₅₀ assays. LC₅₀ may be a
365 quick measure of acute tolerance and it has an uncertain correlation with fitness components. As
366 such, our phenotyping results should be interpreted with caution when considering the potential
367 impact(s) on relative clonal fitness. Insofar as the pattern of rapid evolution of acute tolerance
368 observed may be not actively reflect the phenotype actually under selection across time.

369

370 *Caveats, Analytical Roadmap, and Conclusions:*

371

372 All studies integrating historical data require caution to avoid over-interpretation of the emergent
373 patterns. Our study is no different and relies heavily on a single core from a single lake. Our
374 previous studies with this lake have explored many of these caveats (31). Specific to resurrection
375 ecology (RE) studies, an acute limitation has always been hatching from the egg bank in the
376 deepest layers. There may be non-random patterns of egg mortality in the sediments or non-
377 random propensities in hatching success that skew phenotypic estimates and prevent accurate
378 estimation of allele frequencies. Some have termed this the "invisible fraction" (*sensu* (50)). In our
379 study this is most notable in our inability to hatch genotypes predating the most substantial
380 increases in salinity (e.g., from 1950 or before). In other resurrection studies that have been able
381 to hatch truly 'ancient' eggs (15), the utility of these samples in constructing a framework of
382 phenotypic evolution is limited by sample size because only one or a few isolates survive to be
383 cultured. In many circumstances, however, directly sequencing the eggs is an alternative but will
384 prevent any possible phenotypic characterization of sequenced individuals because eggs are
385 destructively sampled (51). These latter methods are still nonetheless difficult and involve costly
386 genome amplification steps with variable success rates and high levels of exogenous
387 contamination in the final libraries (51). This approach may help reduce the impacts of the
388 'invisible fraction' but will not eliminate it completely. Another caveat that we must highlight is that
389 we have only analyzed a single population, and as such, we cannot place our results within a
390 metapopulation context. We assume that gene flow should be negligible in producing allele
391 frequency changes. To accurately account for gene flow, one would need to sample many
392 additional spatially-distributed populations in addition to hatching temporal samples. While such
393 data sets are within technical reach, the practicality of such a sampling regime would be difficult
394 to amass for species such as *D. pulicaria*.

395

396 Regardless of the issues with RE data sets, we believe we can provide some insights into
397 an analytical framework for future resurrection genomic studies. One major goal of the field of
398 ecological and evolutionary genomics is to ultimately produce a comprehensive phenotype-to-

399 genotype map- especially for those traits that one deems “ecologically relevant.” Relevant
400 critiques of a QTL or QTN-centric research program aside (52), temporally-sampled genomic
401 datasets may provide some very convincing examples of phenotype-genotype maps (see (53)).
402 Resurrection-type studies are poised to do this in natural populations as well. This will require
403 robust demographic analysis of resurrected populations, integrated with simulations in a flexible
404 manner. One potential way forward is the use of flexible-forward genetic simulations (e.g., those
405 available in SLiM; (54)), which can allow more analytical power in genomic analysis of temporally
406 sampled populations.

407

408 In summary, resurrection ecology (RE), when paired with whole genome sequencing,
409 provides a unique and powerful way to study rapid evolution of populations *in situ*. Integration of
410 candidate loci identified with RE into study designs including breeding experiments (55), forward
411 mutation screens (56) or CRISPR technology (57) will provide unrivaled insight for the genetic
412 architecture of complex, ecologically relevant traits in the wild. Such an approach would be
413 beneficial here, for example in validating the effect of the null *clcn2-x2* allele on salinity tolerance
414 in the TL population. Overall, however, here RE provides both molecular and phenotypic
415 evidence of rapid adaptation in the TL population. Thus, the persistence of *D. pulicaria* in this
416 severely salinized lake is likely the result of this rapid adaptation. Our findings indicate that
417 keystone aquatic species such as lake *Daphnia* may continue to thrive in lakes that exceed
418 current water quality limits for salinization and thus maintain the stability of the food webs and
419 ecosystem services they support.

420

421 **Materials and Methods**

422

423 *Clone bank and Sequencing:*

424

425 On 2 July 2019, we collected duplicate sediment cores in TL from a 14 m deep station following
426 Wright (58). During the same period, we also collected *D. pulicaria* from the active plankton
427 community using several vertical tows of a Wisconsin net at the core sampling station. Animals
428 were isolated as single individuals in 125 mL plastic (screw-capped) cups in COMBO media (59).
429 A total of 10 of these clones were established in laboratory culture. Resting eggs (ephippia)
430 collected from throughout the cores were collected according to the methods outlined in Wersebe
431 et al. (31). *D. pulicaria* are cyclically parthenogenetic, meaning they produce clonal offspring
432 during the growing season and may occasionally engage in sex to produce resting eggs encased
433 in durable structures called ephippia (60). Ephippia identified as *D. pulicaria* were subjected to
434 hatching protocol described (15) (see supplemental). These hatchling individuals were expanded
435 in culture to establish upwards of 10 clones per sediment layer to establish a clone bank. Fifty-
436 four isoclonal lineages from the clone bank were selected for DNA extraction and whole genome
437 sequencing (average 10X) on an Illumina NovaSeq by the Oklahoma Medical Research
438 Foundation.

439

440 *Bioinformatics:*

441

442 Raw sequencing reads were quality trimmed and adaptor contamination removed using
443 Trimmomatic (61). Quality trimmed reads were aligned to the chromosome-level *D. pulicaria*
444 genome assembly (62) using the BWA mem algorithm (63). The resulting files were piped
445 through samtools (64) to mark duplicates, fix mates and sort the bam files. We called variants
446 using the bcftools mpileup and call pipeline using all individuals together (65). Using bcftools, the
447 resulting BCF files were concatenated together into a single genome-wide file and quality filtered
448 to a set of high confidence biallelic single nucleotide polymorphisms (SNPs).

449

450 *Population Structure and Genetic Divergence:*

451

452 The variants in the final quality-filtered VCF file were pruned for linkage disequilibrium (LD) using
453 Plink (66) independent pairwise function (settings: 50, 10, 0.1), providing an independent and
454 essentially random set of SNPs. The resulting variants were further filtered to SNPs with 0%
455 missingness to a set 27854 genome-wide SNPs. We conducted Principal Components Analysis
456 (PCA) in R (Version 4.2.0; R core team 2022) using the packages *adegenet* and *vcfr* (67, 68). We
457 also conducted Discriminant Analysis of Principal Components (DAPC); a flexible population
458 assignment method also implemented in *adegenet* (69). Using the cross-validation procedure
459 outlined in the vignette, we found that retention of 10 PCs performed best in population
460 assignment. We performed population assignment tests for each clone retaining 10 PCs and 2
461 discriminant functions and plotted the results as a bar plot to visual the probabilities of population
462 assignment. DAPC is flexible enough to handle mixed clonal and temporal sampling - two factors
463 that violate other assignment techniques (e.g., STRUCTURE (70). From the PCA and the DAPC,
464 results, we determined that the samples could be assigned to three “sub-populations” according
465 to the depth of their recovery (see results). Using these three subpopulations as designations, we
466 estimated overall site-wise F_{st} using the *basic.stats* function and mean pairwise genetic distance
467 *genet.dist* function in the R package *hierfstat* (71). In addition, we estimated nucleotide diversity
468 (π) in 10 kb windows throughout the genome for each of the temporal subpopulations using the
469 program PIXY (72).

470

471 *Estimation of Effective Population Size and Simulations:*

472

473 To parameterize our tests for selection, we sought to identify the recent demographic history of
474 the TL *D. pulicaria* population. To accomplish this, we used two different methods to estimate
475 effective population size (N_e) and growth trajectory of the population. The first method,
476 implemented in GONE (73) uses LD to estimate the recent population history. This method is
477 robust to non-equilibrium histories such as selection (74); however, it is suitable only for sample
478 pools collected from the same generation. Thus, for this method we used only the samples

479 collected from the water column in 2019. We ran six independent runs of this method using
480 random subsets of 600,000 SNPs from all the SNPs called in the population and a constant
481 recombination rate of 7.2 cm/mb estimated from the *D. pulicaria* genetic map (62). This analysis
482 does not assume a given model *a priori*, instead it produces a population size trajectory that when
483 inspected graphically can hint at different events (e.g., bottlenecks). In addition to LD-based
484 methods, we estimated demographic parameters of the TL population - estimating N_e and the
485 population growth rate - by fitting a demographic model to the folded site frequency spectrum
486 (SFS) implemented in FastSimcoal 2.7 (FSC2.7) (40). With the LD pruned SNPs used above in
487 the PCA analysis, we created folded 2-D SFS from the three temporal subpopulations. Using 100
488 independent runs of FSC2.7, we chose the run that maximized the likelihood of the observed
489 data. We fixed the sampling points in time for the temporal subpopulation (MID and DEEP) to be
490 50 and 100 generations in the past. This assumes approximately 4 asexual generations a year
491 and a single sexual generation for a total of five generations a year. Each run conducted 1-million
492 coalescent simulations and used 40 brent maximization cycles. Further, we estimated empirical
493 p-values for the site-wise F_{st} estimates using simulation (75). We chose the best fitting
494 demographic model estimated in FSC2.7 and conducted 100 separate runs of this model to
495 simulate approximately 1100 SNPs for each run. For each run, we estimated F_{st} using basic.stats
496 function in *hierfstat* (71). We pooled each simulation into an empirical distribution of probable F_{st}
497 values under the best fitting demographic model. We tested each observed site-wise F_{st} estimate
498 against this empirical distribution to estimate a p-value. We corrected these p-values for multiple
499 testing using a false discovery rate in R using the p.adjust function.

500

501 *Genes surrounding Fst outliers, GO term enrichment, and Variant Effects:*

502

503 We extracted the genes surrounding F_{st} outliers ($p \leq 0.05$ after correction) in 10 Kb windows
504 using the *D. pulicaria* RefSeq annotation (release 100, SC_F0-13Bv2) using bedtools (76). Next,
505 we created Panther Generic Mappings for the genes with known annotations using each gene's

506 protein sequences following the method outlined in (41). Using the generic mappings, we tested
507 for molecular function Gene Ontology (GO) term enrichment by testing against the *Daphnia pulex*
508 gene list using the PantherDB webtool using a false discovery rate correction (41). In addition to
509 testing for GO term enrichment, we also surveyed all genes for potential SNPs and small indels
510 mutations potentially driving selection using Ensembl's Variant Effect Predictor (VEP) (42). We
511 built a custom database using the RefSeq annotations in GFF format following the developer's
512 protocol. We extracted all "High" (e.g., premature stops) and "Moderate" (e.g., missense SNPs)
513 impact mutations predicted and cross-referenced these with the genes near F_{st} outliers.

514

515 *Chloride Tolerance:*

516

517 We selected a subset of 30 clones from the Lake (n = 10), 10-12 cm sediment layer (n = 10), 18-
518 20 cm (n = 3), and 22-24 cm (n = 7) subpopulations for estimating clone-specific tolerance to
519 chloride. See the Supplemental Materials for details on the experimental set-up. We estimated
520 LC_{50} for each clone separately by fitting a reduced-bias generalized linear model (GLM) to the
521 survival curve using the R package *brglm* (77). Data were not normally distributed, nor did they
522 meet the assumption of equal variances, so we performed a non-parametric ANOVA (Kruskal-
523 Wallis) test on the LC_{50} estimates to test for differences in the mean LC_{50} value for each
524 subpopulation.

525

526 **Acknowledgments**

527

528 We would like to thank M. Edlund who facilitated the collection of the Tanners Lake sediment
529 cores. T. Curb, Z. Arnold, A. Twumasi-Mensah, and A. Hemani, helped with the LC_{50}
530 experiments. G. Wiley aided in the preparation of sequencing libraries. The University of
531 Oklahoma Biological Station and the St. Croix Watershed Research Station facilitated field work.
532 Funding for this study was provided by the University of Oklahoma Department of Biology Adams

533 Summer Scholarship, Robberson Graduate College Grant, Hill Fund for Research in Biology,
534 Graduate Student Senate Research Grant, AMNH Theodore Roosevelt Grant and the
535 Biogeography of Behavior student seed grant (NSF DBI-2021880; PI L. Stein) awarded to MJW in
536 support of graduate research. Any opinions, findings and conclusions or recommendations
537 expressed in this material are those of the authors and do not necessarily reflect the views of the
538 National Science Foundation, the American Museum of Natural History or the University of
539 Oklahoma. This manuscript represents a portion of MJW's doctoral dissertation at The University
540 of Oklahoma. We thank three anonymous reviewers for constructive comments on earlier
541 versions of the manuscript.

542 **Data Availability:**

543 Sequencing reads will be made available on the NCBI SRA upon acceptance. Data and
544 appropriate metadata from this study will be archived in the University of Oklahoma's ShareOK
545 data repository (<https://shareok.org>). Code used in the analysis is available on GitHub at
546 https://github.com/mworsebe/Tanner_lake_genomics.

547

548

549

550 **References**

551

- 552 1. S. M. Rudman, *et al.*, What genomic data can reveal about eco-evolutionary dynamics.
553 *Nature Ecology & Evolution* **2**, 9–15 (2018).
- 554 2. N. G. Hairston, *et al.*, Lake ecosystems: Rapid evolution revealed by dormant eggs. *Nature*
555 **401**, 446–446 (1999).
- 556 3. F. Mallard, V. Nolte, R. Tobler, M. Kapun, C. Schlötterer, A simple genetic basis of
557 adaptation to a novel thermal environment results in complex metabolic rewiring in
558 *Drosophila*. *Genome Biology* **19**, 119 (2018).
- 559 4. P. W. Messer, D. A. Petrov, Population genomics of rapid adaptation by soft selective
560 sweeps. *Trends in Ecology and Evolution* **28**, 659–669 (2013).

- 561 5. P. W. Messer, S. P. Ellner, N. G. Hairston, Can Population Genetics Adapt to Rapid
562 Evolution? *Trends in genetics : TIG* **32**, 408–418 (2016).
- 563 6. A. Eyre-Walker, P. D. Keightley, The distribution of fitness effects of new mutations. *Nature*
564 *Reviews Genetics* 2007 8:8 **8**, 610–618 (2007).
- 565 7. P. D. Keightley, A. Eyre-Walker, What can we learn about the distribution of fitness effects
566 of new mutations from DNA sequence data? *Philosophical Transactions of the Royal*
567 *Society B: Biological Sciences* **365**, 1187–1193 (2010).
- 568 8. A. Long, G. Liti, A. Luptak, O. Tenaillon, Elucidating the molecular architecture of
569 adaptation via evolve and resequence experiments. *Nature Reviews Genetics* **16**, 567–582
570 (2015).
- 571 9. C. Schlötterer, R. Kofler, E. Versace, R. Tobler, S. U. Franssen, Combining experimental
572 evolution with next-generation sequencing: A powerful tool to study adaptation from
573 standing genetic variation. *Heredity* **114**, 431–440 (2015).
- 574 10. T. L. Turner, A. D. Stewart, A. T. Fields, W. R. Rice, A. M. Tarone, Population-Based
575 Resequencing of Experimentally Evolved Populations Reveals the Genetic Basis of Body
576 Size Variation in *Drosophila melanogaster*. *PLoS Genetics* **7**, e1001336 (2011).
- 577 11. R. E. Lenski, Convergence and divergence in a long-term experiment with bacteria.
578 *American Naturalist* **190**, S57–S68 (2017).
- 579 12. M. Dehasque, *et al.*, Inference of natural selection from ancient DNA. *Evolution Letters* **4**,
580 94–108 (2020).
- 581 13. L. J. Weider, P. D. Jeyasingh, D. Frisch, Evolutionary aspects of resurrection ecology:
582 Progress, scope, and applications-An overview. *Evolutionary Applications* **11**, 3–10 (2018).
- 583 14. M. Gros-Balthazard, *et al.*, The genomes of ancient date palms germinated from 2,000 y
584 old seeds. *Proceedings of the National Academy of Sciences of the United States of*
585 *America* **118** (2021).
- 586 15. D. Frisch, *et al.*, A millennial-scale chronicle of evolutionary responses to cultural
587 eutrophication in *Daphnia*. *Ecology Letters* **17**, 360–368 (2014).
- 588 16. W. C. Kerfoot, J. A. Robbins, L. J. Weider, A new approach to historical reconstruction:
589 Combining descriptive and experimental paleolimnology. *Limnology and Oceanography* **44**,
590 1232–1247 (1999).
- 591 17. W. C. Kerfoot, L. J. Weider, “Experimental paleoecology (resurrection ecology): Chasing
592 Van Valen’s Red Queen hypothesis” (2004).
- 593 18. L. Orsini, *et al.*, Temporal genetic stability in natural populations of the waterflea
594 *Daphnia magna* in response to strong selection pressure. *Molecular Ecology*
595 **25**, 6024–6038 (2016).

- 596 19. A. Chaturvedi, *et al.*, Extensive standing genetic variation from a small number of founders
597 enables rapid adaptation in *Daphnia*. *Nature Communications* **12**, 4306 (2021).
- 598 20. C. Cousyn, *et al.*, Rapid, local adaptation of zooplankton behavior to changes in predation
599 pressure in the absence of neutral genetic changes. *Proceedings of the National Academy*
600 *of Sciences of the United States of America* **98**, 6256–60 (2001).
- 601 21. D. Frisch, *et al.*, Paleogenetic records of *Daphnia pulicaria* in two North American lakes
602 reveal the impact of cultural eutrophication. *Global Change Biology* **23**, 708–718 (2017).
- 603 22. W. Lampert, U. Sommer, *Limnoecology: The Ecology of Lakes and Streams* (Oxford
604 University Press, 2007).
- 605 23. Ž. Ogorelec, C. Wunsch, A. J. Kunzmann, P. Octorina, J. I. Navarro, Large daphniids are
606 keystone species that link fish predation and phytoplankton in trophic cascades.
607 *Fundamental and Applied Limnology* **194**, 297–309 (2021).
- 608 24. J. R. Walsh, S. R. Carpenter, M. J. Vander Zanden, Invasive species triggers a massive loss of
609 ecosystem services through a trophic cascade. *Proceedings of the National Academy of*
610 *Sciences of the United States of America* **113**, 4081–5 (2016).
- 611 25. A. J. Reid, *et al.*, Emerging threats and persistent conservation challenges for freshwater
612 biodiversity. *Biological Reviews* **94**, 849–873 (2019).
- 613 26. S. S. Kaushal, *et al.*, Freshwater salinization syndrome on a continental scale. *Proceedings*
614 *of the National Academy of Sciences* (2018) <https://doi.org/10.1073/pnas.1711234115>.
- 615 27. H. A. Dugan, *et al.*, Lakes at Risk of Chloride Contamination. *Environmental Science and*
616 *Technology* **54**, 6639–6650 (2020).
- 617 28. H. A. Dugan, *et al.*, Salting our freshwater lakes. *Proceedings of the National Academy of*
618 *Sciences of the United States of America* **114**, 4453–4458 (2017).
- 619 29. W. D. Hintz, R. A. Relyea, A review of the species, community, and ecosystem impacts of
620 road salt salinisation in fresh waters. *Freshwater Biology* **64**, 1081–1097 (2019).
- 621 30. D. Cunillera-Montcusí, *et al.*, Freshwater salinisation: a research agenda for a saltier world.
622 *Trends in Ecology & Evolution* **0** (2022).
- 623 31. M. J. Wersebe, M. B. Edlund, L. J. Weider, Does salinization impact long-term *Daphnia*
624 assemblage dynamics? Evidence from the sediment egg bank in a small hard-water lake.
625 *Limnology and Oceanography Letters* **n/a** (2021).
- 626 32. S. Arnott, *et al.*, Road salt impacts freshwater zooplankton at concentrations below current
627 water quality guidelines. *Environmental Science & Technology* (2020)
628 <https://doi.org/10.1021/acs.est.0c02396> (August 11, 2020).

- 629 33. H. W. D., *et al.*, Current water quality guidelines across North America and Europe do not
630 protect lakes from salinization. *Proceedings of the National Academy of Sciences* **119**,
631 e2115033119 (2022).
- 632 34. E. V. Novotny, D. Murphy, H. G. Stefan, Increase of urban lake salinity by road deicing salt.
633 *Science of The Total Environment* **406**, 131–144 (2008).
- 634 35. J. M. Ramstack, S. C. Fritz, D. R. Engstrom, Twentieth century water quality trends in
635 Minnesota lakes compared with presettlement variability. *Canadian Journal of Fisheries*
636 *and Aquatic Sciences* **61**, 561–576 (2004).
- 637 36. J. M. Ramstack, S. C. Fritz, D. R. Engstrom, S. A. Heiskary, The Application of a Diatom-
638 based Transfer Function to Evaluate Regional Water-Quality Trends in Minnessota Since
639 1970. *Journal of Paleolimnology* **29**, 79–94 (2003).
- 640 37. R. J. Sibert, C. M. Koretsky, D. A. Wyman, Cultural meromixis: Effects of road salt on the
641 chemical stratification of an urban kettle lake. *Chemical Geology* **395**, 126–137 (2015).
- 642 38. D. E. Allen, M. Lynch, THE EFFECT OF VARIABLE FREQUENCY OF SEXUAL REPRODUCTION
643 ON THE GENETIC STRUCTURE OF NATURAL POPULATIONS OF A CYCLICAL PARTHENOGEN.
644 *Evolution* **66**, 919–926 (2012).
- 645 39. C. E. Cáceres, A. J. Tessier, Incidence of diapause varies among populations of *Daphnia*
646 *pulicaria*. *Oecologia* **141**, 425–431 (2004).
- 647 40. L. Excoffier, *et al.*, fastsimcoal2: demographic inference under complex evolutionary
648 scenarios. *Bioinformatics* **37**, 4882–4885 (2021).
- 649 41. H. Mi, *et al.*, PANTHER version 16: A revised family classification, tree-based classification
650 tool, enhancer regions and extensive API. *Nucleic Acids Research* **49**, D394–D403 (2021).
- 651 42. W. McLaren, *et al.*, The Ensembl Variant Effect Predictor. *Genome Biology* **17**, 122 (2016).
- 652 43. S. Masaki, *et al.*, Still life, a Protein in Synaptic Terminals of *Drosophila* Homologous to
653 GDP-GTP Exchangers. *Science* **275**, 543–547 (1997).
- 654 44. P. Aspenström, N. Richnau, A.-S. Johansson, The diaphanous-related formin DAAM1
655 collaborates with the Rho GTPases RhoA and Cdc42, CIP4 and Src in regulating cell
656 morphogenesis and actin dynamics. *Experimental Cell Research* **312**, 2180–2194 (2006).
- 657 45. C. Klambt, The *Drosophila* gene pointed encodes two ETS-like proteins which are involved
658 in the development of the midline glial cells. *Development* **117**, 163–176 (1993).
- 659 46. C. Fangli, B. Margaret, R. K. T., Q. Adrian, H. I. K., Biological characterization of *Drosophila*
660 Rapgap1, a GTPase activating protein for Rap1. *Proceedings of the National Academy of*
661 *Sciences* **94**, 12485–12490 (1997).

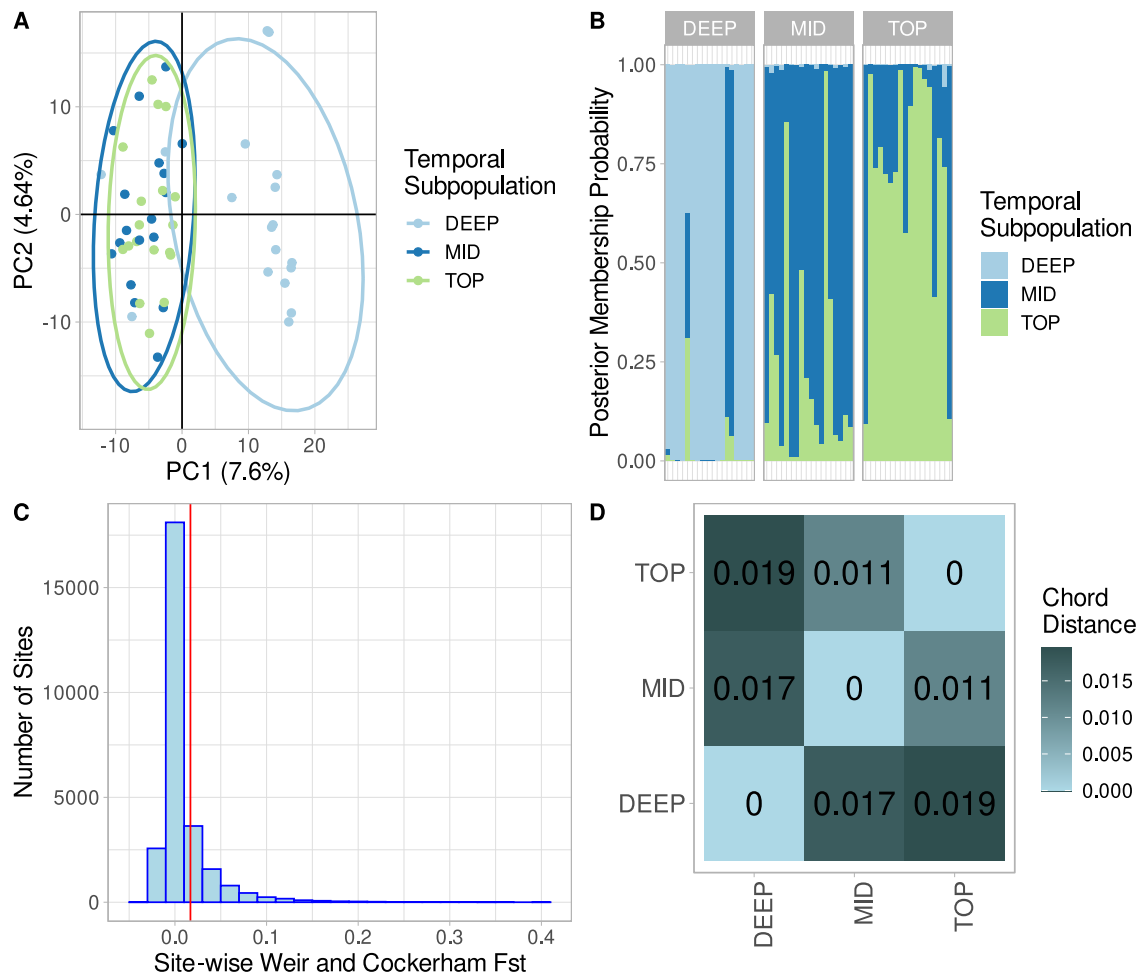
- 662 47. S. E. St Pierre, M. I. Galindo, J. P. Couso, S. Thor, Control of *Drosophila* imaginal disc
663 development by *rotund* and *roughened eye*: differentially expressed transcripts of the
664 same gene encoding functionally distinct zinc finger proteins. *Development* **129**, 1273–
665 1281 (2002).
- 666 48. M. Lynch, K. Spitze, “Evolutionary Genetic of *Daphnia*” in *Ecological Genetics*, L. Real, Ed.
667 (Princeton University Press, 1994).
- 668 49. E. V. Novotny, H. G. Stefan, Projections of Chloride Concentrations in Urban Lakes
669 Receiving Road De-icing Salt. *Water, Air, & Soil Pollution* **211**, 261–271 (2010).
- 670 50. A. E. Weis, Detecting the “invisible fraction” bias in resurrection experiments. *Evolutionary*
671 *Applications* **11**, 88–95 (2018).
- 672 51. J. B. Lack, L. J. Weider, P. D. Jeyasingh, Whole genome amplification and sequencing of a
673 *Daphnia* resting egg. *Molecular Ecology Resources* **18**, 118–127 (2018).
- 674 52. M. V. Rockman, THE QTN PROGRAM AND THE ALLELES THAT MATTER FOR EVOLUTION:
675 ALL THAT’S GOLD DOES NOT GLITTER. *Evolution* **66**, 1–17 (2012).
- 676 53. N. O. Therkildsen, *et al.*, Contrasting genomic shifts underlie parallel phenotypic evolution
677 in response to fishing. *Science (New York, N.Y.)* **365**, 487–490 (2019).
- 678 54. B. C. Haller, P. W. Messer, SLiM 3: Forward Genetic Simulations Beyond the Wright-Fisher
679 Model. *Molecular Biology and Evolution* **36**, 632–637 (2019).
- 680 55. R. E. Sherman, R. Hartnett, E. L. Kiehnau, L. J. Weider, P. D. Jeyasingh, Quantitative
681 genetics of phosphorus content in the freshwater herbivore, *Daphnia pulex*. *Journal of*
682 *Animal Ecology* **00**, 1365–2656.13419 (2021).
- 683 56. M. Snyman, T. V. Huynh, M. T. Smith, S. Xu, The genome-wide rate and spectrum of EMS-
684 induced heritable mutations in the microcrustacean *Daphnia*: on the prospect of forward
685 genetics. *Heredity* **127**, 535–545 (2021).
- 686 57. C. Hiruta, K. Kakui, K. E. Tollefsen, T. Iguchi, Targeted gene disruption by use of
687 CRISPR/Cas9 ribonucleoprotein complexes in the water flea *Daphnia pulex*. *Genes to Cells*
688 **23**, 494–502 (2018).
- 689 58. H. E. Wright, Coring tips. *Journal of Paleolimnology* **6**, 37–49 (1991).
- 690 59. S. S. Kilham, D. A. Kreeger, S. G. Lynn, C. E. Goulden, L. Herrera, COMBO: a defined
691 freshwater culture medium for algae and zooplankton. *Hydrobiologia* **377**, 147–159
692 (1998).
- 693 60. W. Lampert, *Daphnia: Development of a Model Organism in Ecology and Evolution*
694 (International Ecology Institute (ECI), 2011).

- 695 61. A. M. Bolger, M. Lohse, B. Usadel, Trimmomatic: a flexible trimmer for Illumina sequence
696 data. *Bioinformatics* **30**, 2114–2120 (2014).
- 697 62. M. J. Wersebe, R. E. Sherman, P. D. Jeyasingh, L. J. Weider, The roles of recombination and
698 selection in shaping genomic divergence in an incipient ecological species complex.
699 *Molecular Ecology* **n/a** (2022).
- 700 63. H. Li, R. Durbin, Fast and accurate short read alignment with Burrows-Wheeler transform.
701 *Bioinformatics* **25**, 1754–1760 (2009).
- 702 64. H. Li, *et al.*, The Sequence Alignment/Map format and SAMtools. *Bioinformatics* **25**, 2078–
703 2079 (2009).
- 704 65. P. Danecek, *et al.*, Twelve years of SAMtools and BCFtools. *GigaScience* **10**, 1–4 (2021).
- 705 66. C. C. Chang, *et al.*, Second-generation PLINK: Rising to the challenge of larger and richer
706 datasets. *GigaScience* **4**, 7 (2015).
- 707 67. T. Jombart, adegenet: a R package for the multivariate analysis of genetic markers.
708 *Bioinformatics* **24**, 1403–1405 (2008).
- 709 68. B. J. Knaus, N. J. Grünwald, vcfr: a package to manipulate and visualize variant call format
710 data in R. *Molecular Ecology Resources* **17**, 44–53 (2017).
- 711 69. T. Jombart, S. Devillard, F. Balloux, Discriminant analysis of principal components: A new
712 method for the analysis of genetically structured populations. *BMC Genetics* **11**, 94 (2010).
- 713 70. T. A. Joseph, I. Pe’er, Inference of Population Structure from Time-Series Genotype Data.
714 *American Journal of Human Genetics* **105**, 317–333 (2019).
- 715 71. J. Goudet, hierfstat, a package for r to compute and test hierarchical F-statistics.
716 *Molecular Ecology Notes* **5**, 184–186 (2005).
- 717 72. K. L. Korunes, K. Samuk, pixy: Unbiased estimation of nucleotide diversity and divergence
718 in the presence of missing data. *Molecular Ecology Resources* **21**, 1359–1368 (2021).
- 719 73. E. Santiago, *et al.*, Recent Demographic History Inferred by High-Resolution Analysis of
720 Linkage Disequilibrium. *Molecular Biology and Evolution* **37**, 3642–3653 (2020).
- 721 74. I. Novo, E. Santiago, A. Caballero, The estimates of effective population size based on
722 linkage disequilibrium are virtually unaffected by natural selection. *PLOS Genetics* **18**,
723 e1009764 (2022).
- 724 75. K. E. Lotterhos, M. C. Whitlock, The relative power of genome scans to detect local
725 adaptation depends on sampling design and statistical method. *Molecular Ecology* **24**,
726 1031–1046 (2015).

- 727 76. A. R. Quinlan, I. M. Hall, BEDTools: a flexible suite of utilities for comparing genomic
728 features. *Bioinformatics* **26**, 841–842 (2010).
- 729 77. I. Kosmidis, E. C. Kenne Pagui, N. Sartori, Mean and median bias reduction in generalized
730 linear models. *Statistics and Computing* **30**, 43–59 (2020).

731
732
733
734

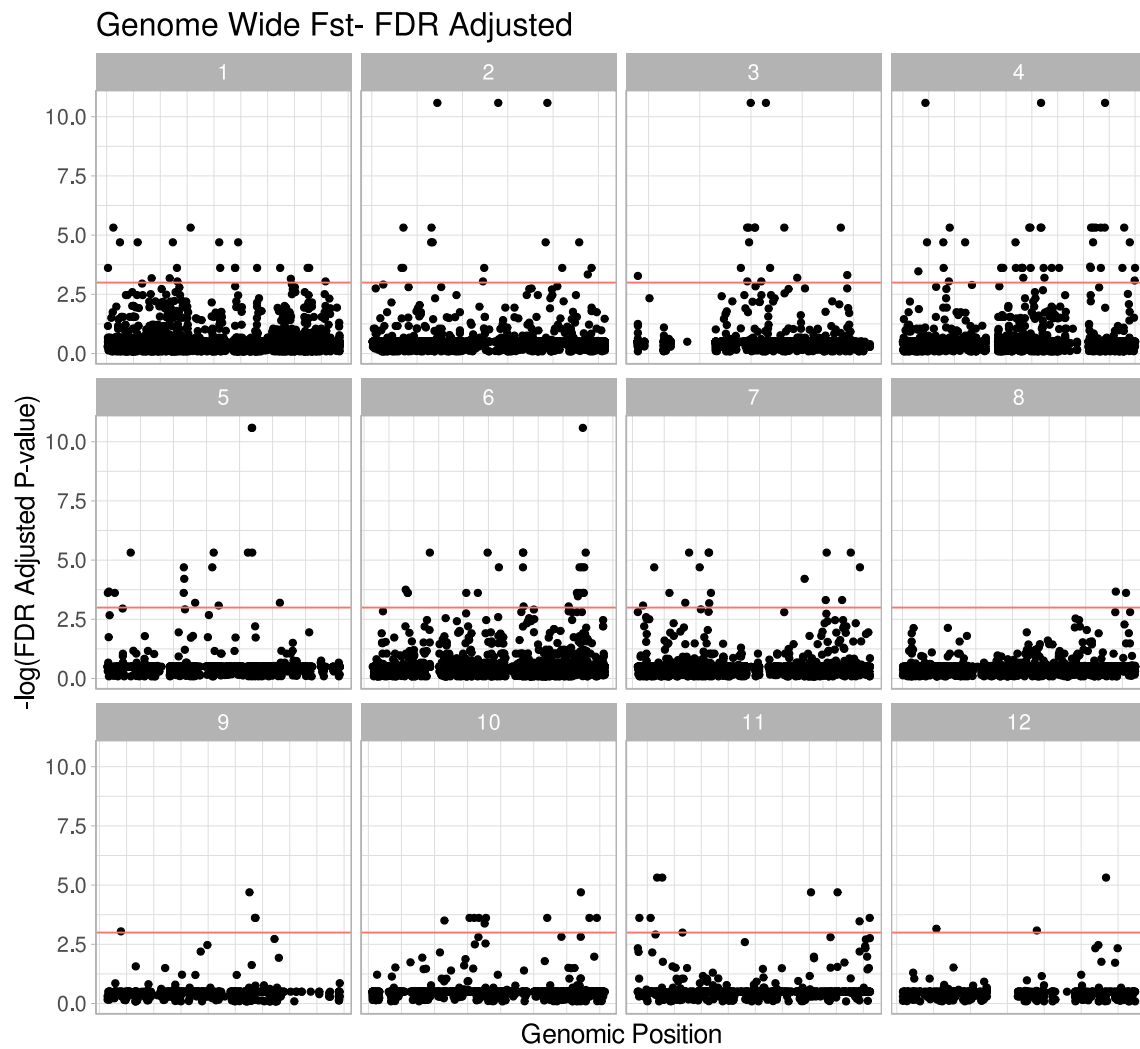
735 **Figures and Tables**
736



737

738 **Figure 1:** Genetic structure, divergence, and distance across time in Tanners Lake. A) Principal
739 Component Analysis (PCA) biplot of all clones color-coded according to the depth of recovery in
740 the core (cm). PC1 and PC2 explain 7.6 % and 4.6 % of variance observed in the SNP data,
741 respectively. Older clones (22-24 cm, 18-20 cm, & 16-18 cm) form a cluster distinct from more
742 recent clones (LC, 2-4 cm, 6-8 cm and 10-12 cm). B) Discriminant Analysis of Principal
743 Components (DAPC) bar plot showing the posterior probability of assignment to the three *a priori*
744 defined temporal subpopulations. C) Observed Weir and Cockerham site-wise F_{st} estimates.
745 Overall F_{st} was low (red vertical line) at only 0.0169. D) Pair-wise genetic chord distances
746 between the three temporal subpopulations (TOP, MID and DEEP)

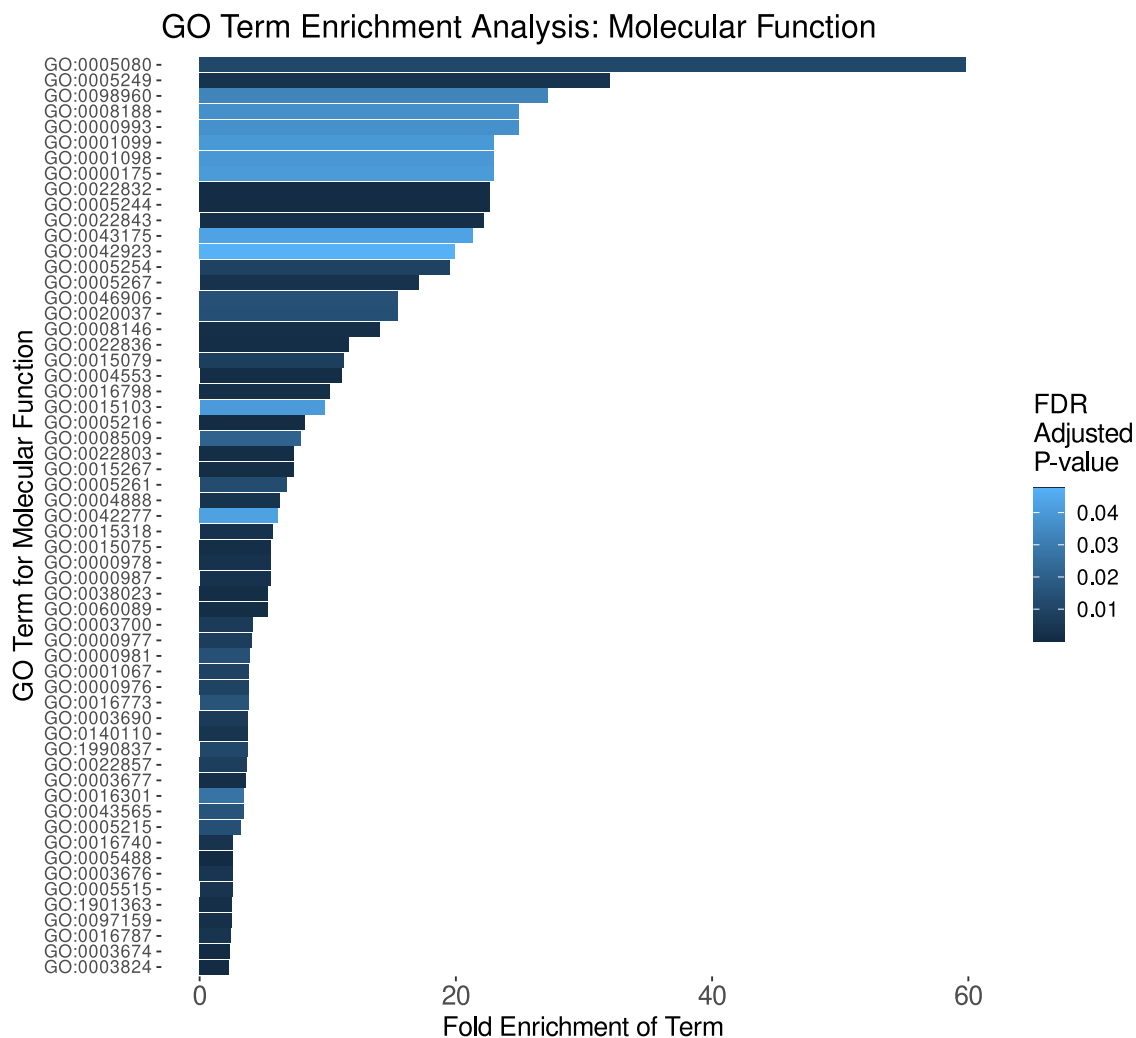
747



748

749 **Figure 2:** Genome-wide F_{st} Manhattan plots. Each panel represents a chromosome (1-12), each
750 point designates a Single Nucleotide Polymorphism (SNP), the horizontal line indicates genome-
751 wide significance at $p = 0.05$. Points above the red line are statistical outliers. Chromosome
752 lengths are normalized across panels.

753



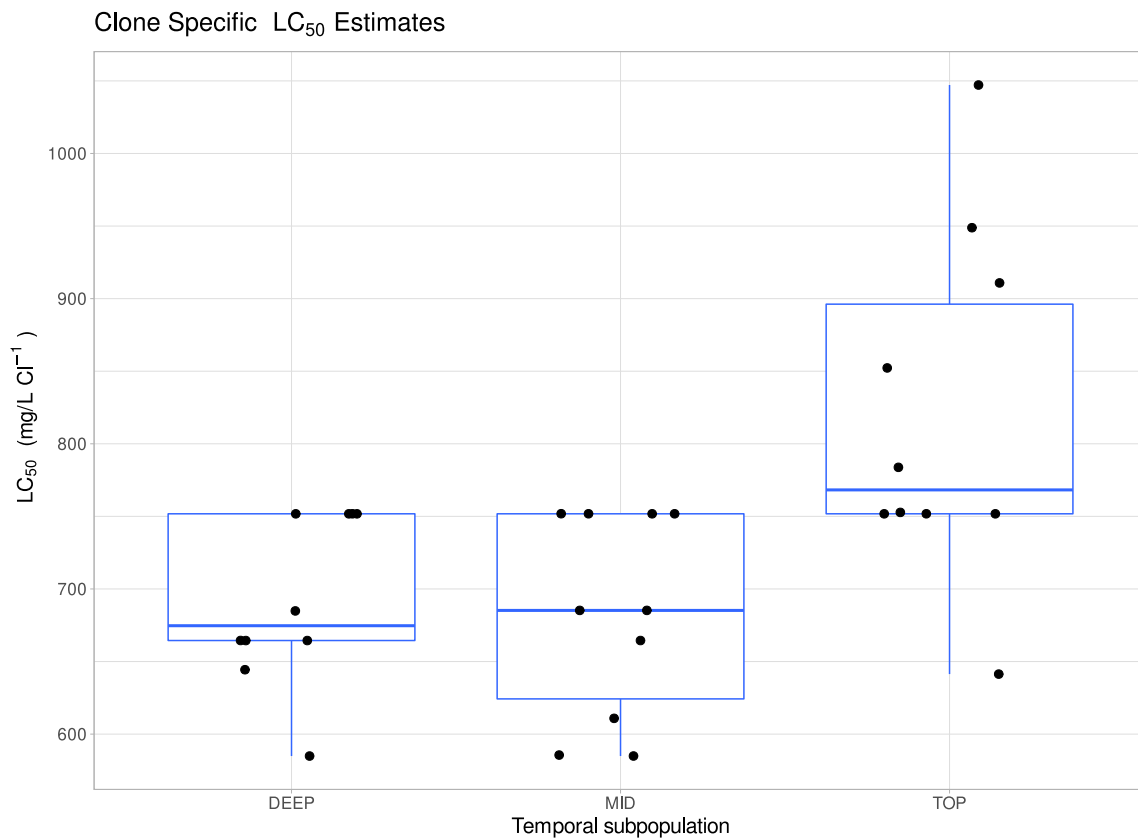
754

755

756 **Figure 3:** Enriched Gene Ontology (GO) terms for molecular function. Enriched terms are set
757 along the Y-axis. The length of each bar indicates the term's fold enrichment in the analyzed
758 gene list, and the bar color denotes its False Discovery Rate (FDR) corrected p-value. In total, 59
759 terms were enriched among the 286 genes found near outlier SNPs. GO term mappings are
760 provided in table S2.

761

762



763

764 **Figure 4:** 96-hour lethal concentration-50% assay (LC_{50}) estimates for clones evaluated in the
765 salinity tolerance experiments. Box plots indicate population medians and variances, each point is
766 a clone. Overall, non-parametric ANOVA (Kruskal-Wallis) indicated a significant main effect of
767 population. Post-hoc testing (pairwise Wilcoxon test) confirmed that the TOP population was
768 more tolerant than either the MID or DEEP populations, which were not different from one
769 another.

770