

1 **Differential patterns of taxonomic and functional diversity for two groups of canopy**
2 **arthropods across spatial scales**

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20
21 **Authorship:** TOC originally conceived of the study and collected samples; all authors conceived
22 of current manuscript; HJP and KUC identified spiders and ants from samples; MBM conducted
23 statistical analyses; MBM, HJP, and KUC wrote the first draft of the manuscript; all authors
24 revised the manuscript

25
26 **Biosketch:** This research team aims to better understand and quantify arthropod community
27 structure and diversity at landscape scales through local experiments and broad scale
28 observational approaches. They are interested in refining our understanding of the structuring of
29 biodiversity to improve conservation efforts.

1 **Differential patterns of taxonomic and functional diversity for two groups of canopy**
2 **arthropods across spatial scales**

3 **Running Title:** Scale impacts canopy arthropod diversity

4 **Key Words:** beta diversity, functional diversity, diversity partitioning, forest canopy,
5 Formicidae, Araneae

6 **Abstract**

7 *Aim*

8 Arthropod diversity is often linked to variation in resource use, dispersal ability, habitat
9 connectivity, and climate factors that differ across spatial scales. The aim of this research was to
10 examine how species richness, functional diversity, and community composition of two taxa
11 differing in functional roles and dispersal ability are structured across spatial scales and to
12 identify the importance of vegetation, climate, and landscape in explaining these patterns at
13 different scales.

14 *Location*

15 96 trees in 24 stands of 6 deciduous forest sites in 2 ecoregions of the eastern USA (North-
16 Central Till Plain and Western Allegheny Plateau)

17 *Time period*

18 2000

19 *Major taxa studied*

20 Canopy dwelling ants (Hymenoptera: Formicidae) and spiders (Araneae)

21 *Methods*

22 Organisms were collected from tree canopies using insecticidal fogging. Ant and spider
23 taxonomic and functional beta diversity were partitioned across four hierarchical spatial scales

24 (individual tree, forest stand, site, and ecoregion). The contribution of climactic, landscape, and
25 vegetation variables was determined using model selection.

26 *Results*

27 Ant and spider species richness, functional diversity, and community composition differed
28 between taxa and across spatial scales. Alpha diversity (within trees) was lower than expected for
29 both taxa and types of diversity, with host tree species supporting different species of ants and
30 spiders. While beta components of species diversity among trees and forest stands was greater
31 than expected for both taxa, spiders also showed significant levels of beta diversity among sites.
32 Functional beta diversity was less scale-dependent than taxonomic beta diversity. Stand-level
33 patterns of beta diversity were significantly predicted by variation in climate and landscape
34 connectivity.

35 *Main conclusions*

36 Effects of climate and landscape fragmentation on the diversity and community structure of both
37 taxa indicate that anthropogenic climate change and land use change will alter canopy arthropod
38 communities. Results also suggest that patterns of diversity among fragmentation metrics is
39 influenced by differences in dispersal ability.

40 1 INTRODUCTION

41 Understanding drivers of species diversity and distributions are primary goals of ecology
42 and can be used to inform conservation efforts. Measures of biodiversity span local (alpha
43 diversity, α) and regional (gamma diversity, γ) scales. Studies of the variation, or turnover, in
44 biodiversity among sites or communities (beta diversity, β), first proposed by Whittaker (1960),
45 have exploded over the last few decades (Anderson *et al.*, 2011; Socolar *et al.*, 2016; Mammola
46 *et al.*, 2021). Beta diversity has been applied to the spatial scaling of biodiversity, temporal
47 change in communities for conservation monitoring, changes across latitudinal and elevational
48 gradients, and environmental filtering of functional traits and subsequent assembly processes
49 (Crist *et al.*, 2003; Kraft *et al.*, 2011; Barton *et al.*, 2013; Siefert *et al.*, 2013; Jarzyna & Jetz,
50 2018).

51 Beta diversity has been primarily focused on measures of taxonomic diversity but not
52 functional diversity (i.e. based on the ecological roles and traits of species). To a certain extent,
53 species diversity and functional diversity are correlated, but not reliably so, as not all species are
54 equivalent in a given system and their relative roles may change depending on the presence of
55 and interactions with other species. Species and functional diversity are, therefore, not
56 interchangeable. Indeed, one could imagine two communities comprised of entirely different
57 species (high species beta diversity) that carry out the same functions (low functional beta
58 diversity; Swenson, 2011; Swenson *et al.*, 2012; Siefert *et al.*, 2013). Because functional diversity
59 quantifies ecological impacts and species interactions, studies including the functional diversity
60 of organisms provide more robust data for conservation and restoration than studies on species
61 presence and abundance alone (Cadotte *et al.*, 2009).

62 Conservation and restoration efforts are landscape-level problems requiring an
63 understanding of how both species and functional diversity scales from local to regional
64 perspectives. Using taxa, such as arthropods, that are widespread, abundant, speciose, and
65 functionally diverse can help researchers test diversity hypotheses across multiple scales (Kemp
66 *et al.*, 2017; McCreddie & Adler, 2018). Ants and spiders play several important roles in
67 ecosystem functioning through direct and indirect species interactions (Pearce & Venier, 2006;
68 Maleque *et al.*, 2009). Specifically, ants are numerically dominant in most ecosystems, affecting
69 ecological processes through their nest building, predation, and mutualisms (Folgarait, 1998;
70 Crist, 2009); therefore, shifts in ant communities can have multifaceted impacts across trophic
71 levels. Similarly, spiders are obligate predators of other arthropods and may be more susceptible
72 to changes in landscape fragmentation, and loss of key predators is linked to trophic cascades
73 (Bonte *et al.*, 2004; Voigt *et al.*, 2007).

74 The mechanisms that might structure arthropod taxonomic and functional beta diversity
75 across spatial scales (e.g., region, site, and forest stand) include characteristics such as land use
76 and topography, management history, and habitat characteristics, respectively. Forest
77 management practices (e.g., logging, fire regimes, and patch size) affect arthropods across
78 multiple scales. At landscape and regional scales, forest fragmentation due to logging or land-use
79 changes lead to decreased habitat area and isolation as well as greater edge to area ratio and
80 altered biotic and abiotic conditions along edge boundaries (Gluck & Rempel, 1996; Fahrig,
81 2003; Haddad *et al.*, 2015). At the stand level, decreased forest area and increased edge cause
82 shifts in tree stand pattern, vegetation structure, and soil characteristics (e.g. moisture, organic
83 matter, texture; Johnston & Elliott, 1996). The effects of these changes on arthropods are many
84 and varied. Fragment size impacts both taxonomic and functional diversity of ants and spiders as

85 well as shifting community composition of these taxa (Pearce *et al.*, 2005). Forest habitats with
86 greater tree diversity and structural variability and complexity give rise to a greater variety of
87 niches, and, thus, species and roles to inhabit those niches (Tews *et al.*, 2004). Thus, vegetation
88 or habitat complexity can drive patterns of arthropod species richness through direct effects on
89 foraging success and resource availability and indirect effects on ant nest site selection via soil
90 moisture and shading (Mcnett & Rypstra, 2000; Wang *et al.*, 2001; Lassau & Hochuli, 2004).

91 Taken together, beta diversity and composition of key arthropod communities may be
92 affected at a range of spatial scales, potentially driven by several biotic and abiotic
93 characteristics. However, many studies of arthropod diversity focus on a single spatial scale and
94 on taxonomic diversity alone. In this study, we evaluated the role of spatial scale on the
95 taxonomic and functional diversity and the community composition of two arthropod groups
96 (ants and spiders). We quantified these measures across four spatial scales: trees, forest stands,
97 sites, and ecoregions. Specifically, we asked: 1) What spatial scales explain the greatest variation
98 in taxonomic and functional beta diversity and in the community composition of these two taxa?
99 and 2) What are potential mechanisms of scale-associated community structuring, including
100 variation in tree species, forest area and connectivity, and seasonality in climate? We
101 hypothesized that both taxonomic and functional diversity have greater variation at finer spatial
102 scales (tree and stand levels) than at broader spatial scales (site and ecoregion). We also expected
103 that spiders, with greater dispersal abilities, would exhibit greater levels of beta diversity at
104 broader spatial scales than ants, which are more dispersal limited. Similarly, we hypothesized that
105 the biotic and abiotic factors structuring these patterns of diversity differ for spiders and ants
106 given inherent differences in resource use and dispersal ability.

107

108 2 METHODS

109 2.1 Sampling Design, Study Sites, and Arthropod Collection

110 Samples were collected from tree canopies of southern Ohio and southeastern Indiana
111 using a hierarchically nested design with four levels. The broadest level – ecoregion – was
112 represented by the glaciated North-Central Till Plain and unglaciated Western Allegheny Plateau
113 (Figure S1). Each ecoregion varies in soil type, forest composition, and topography. The second
114 level – site – comprised 3 sites in each ecoregion for a total of 6 sites (Figure S1). The third level
115 – stand – comprised 4 forest stands within each site (24 total). Within each site, two stands were
116 located in uplands, and two stands were located in lowland topographic positions. The fourth
117 level – tree – included 8 individual trees (96 total) within a 1-ha area of each stand. Each tree was
118 sampled using canopy fogging and arthropods were collected in an array of 12 1-m² funnel arrays
119 placed under the crown of the fogged tree. Arthropods were knocked down by insecticidal
120 fogging (0.5 L of 0.5% pyrethrin-based insecticide) for 3 minutes using a Curtis Dyna-Fogger
121 hoisted into the tree crown and collected by funnels with attached vials of ethanol. Insecticidal
122 fogging is not dependent on arthropod activity and lethality is non-specific (Basset *et al.*, 1997;
123 Stork & Hammond, 1997). Sampling was completed during early (22 May-20 June 2000) and
124 late (2-25 August 2000) summer to capture seasonal variation, with early and late season samples
125 pooled for analyses. For more details on sampling methods, see Gering & Crist (2002). Ants were
126 identified to species using the *Ants of Ohio* (Coovert, 2005) and ant functional groups were trait-
127 based (see below) rather than explicitly based on taxonomy or behavior (Andersen, 1997; Crist,
128 2009). Spider adults were identified to species based on the *Spiders of North America* (American
129 Arachnological Society, 2005), *Spiders of Connecticut* (Kaston, 1981), and a provisional list of
130 Ohio spiders (Bradley, 2017), while juveniles were identified to family level when possible.

131 Spider families were used as an indicator of functional guild, based on foraging strategy, prey
132 range, habitat stratification, and circadian activity, as these traits are highly correlated with family
133 (Cardoso *et al.*, 2011).

134 We refer to these samples as canopy or arboreal arthropods because they were collected
135 by fogging tree crowns. Although some of the common species we recorded are primarily
136 arboreal (e.g., *Aphaenogaster mariae*, Table S3), most of the species of ants and spiders are
137 known to move between strata and nest or overwinter in the ground. Nonetheless, Crist and
138 Campbell (2017) recorded significant differences in the community composition of ants from
139 canopy fogging and pitfall traps samples at the same study sites in the North-Central Till Plain.

140

141 2.2 Data Analyses

142 2.2.1 Ant functional groups

143 To classify ants according to functional traits, we selected 10 binary, categorical, and
144 continuous traits that may influence the ecological role of ants (Table S1; (Del Toro *et al.*, 2015;
145 Record *et al.*, 2018; Mahon, 2019). We used trait definitions and data from Del Toro *et al.*
146 (2015), Record *et al.* (2018), Coovert (2005), and AntWiki (2022). Missing morphological data
147 were supplemented with measurements taken from 2-10 mounted specimens per species collected
148 during this study. Functional groups were formed from a dendrogram based on functional
149 dispersion and the Ward clustering method using the dbFD function (FD package, R; Laliberté &
150 Legendre, 2010); functional groups were delineated by setting six functional groups, where we
151 noted a clear break of functional groups (Table S1, Figure S2).

152

153 2.2.2. General patterns and species accumulation curves

154 For all statistical analyses, early and late sampling periods were pooled and all univariate
155 analyses were conducted in R v4.0.0 (R Core Team, 2022). We constructed sample-based
156 rarefaction curves of ants and spiders by tree species to assess differences in species richness
157 among the most common host trees. To determine the effectiveness of our overall sampling
158 effort, we also conducted species rarefaction and extrapolation curves for ants and spiders using
159 the iNEXT package in R (Chao *et al.*, 2014). We also estimated species richness by host trees and
160 overall richness using the Chao1 estimator (Chao, 1984; Colwell & Coddington, 1994).

161

162 2.2.3 Diversity partitioning

163 To analyze taxonomic diversity, we partitioned species richness and functional group
164 richness ($q = 0$) across hierarchical levels, with multiplicative partitioning methods based on the
165 PARTITION software developed by Crist *et al.* (2003) using the R package, PARTITIONR
166 (Mahon *et al.*, 2019). Multiplicative partitions express beta diversity as the effective number of
167 distinct communities, whereas alpha diversity is in units of species richness (Anderson *et al.*,
168 2011). Using hierarchical diversity partitioning, beta components can be separated into nested
169 hierarchical levels (i.e. spatial scale). Hierarchical multiplicative partitioning calculates beta
170 diversity at a given level (i) by dividing the average alpha at the $i+1$ level, $\beta_i = \alpha_{i+1}/\alpha_i$. We
171 tested the significance of α_1 (within trees), β_1 (among trees), β_2 (among stands), β_3 (among
172 sites), and β_4 (between ecoregions) against null, random distributions using 1000 sample-based
173 randomizations. This type of randomization preserves intraspecific aggregation at each
174 hierarchical level, and thus tests the null hypothesis that observed patterns of species diversity at

175 each level are similar to those expected by randomized aggregation of samples at each level of
176 the hierarchical design; alternatively, the null hypothesis is rejected if observed patterns are
177 significantly different from those expected from null distributions, supporting non-random
178 hierarchical species assemblages that are structured by ecological processes (Crist *et al.*, 2003).
179 Partitioning of alpha, beta diversity in this manner therefore accounts for issues of spatial
180 pseudoreplication that can arise in local-to-regional comparisons of species richness (Gering &
181 Crist, 2002). We used a two-tail probability of $p = 0.05$ ($p = 0.025$ for each tail of the null
182 distribution) to determine whether observed diversity patterns were higher or lower than expected
183 via randomizations. To compare relative deviations of the null distributions from the expected
184 values across hierarchical levels and endpoints, we calculated standard effect sizes (SES; also
185 termed beta and alpha deviations) from the mean and standard deviation of the null distributions
186 for each hierarchical level ($SES = (I_{obs} - I_{exp})/\sigma$) (Gotelli & McCabe, 2002; Kraft *et al.*, 2011).

187 To analyze variation in community composition across hierarchical levels, we conducted
188 analyses for multivariate location using PRIMER-E and PERMANOVA+ v6 (Anderson, 2001).
189 We used permutational multivariate analysis of variance (PERMANOVA) to partition the
190 variation in community composition across hierarchical levels. PERMANOVAs were conducted
191 using 9999 permutations of the data, with a nested design. Since multivariate analyses were
192 conducted on square-root transformed abundance data with a Bray-Curtis dissimilarity, variance
193 components of PERMANOVA can be interpreted as percent dissimilarity (Anderson, 2001).

194

195 2.2.4 Landscape and environmental variable analyses

196 To address potential mechanisms of diversity structuring, we compiled and evaluated
197 landscape, climactic, and vegetation variables for forest stands and surrounding areas. Landscape
198 variables were habitat fragmentation measures (estimated using ESRI arcGIS Desktop v10.6.1
199 (ESRI, 2019), Fragstats v4.2.1 (McGarigal *et al.*, 2012)) and land use data (USDA NASS Crop
200 data layer for 2008, the closest year with most reliable data for both Indiana and Ohio (USDA
201 National Agricultural Statistics Service, 2022)). Land cover/land use composition and
202 configuration was collected for of 1.0, 3.0, 6.5, and 10.0 km buffers surrounding each stand to
203 accommodate the potential dispersal distances of both spiders and ants, and to minimize overlap
204 between stand buffers (Thomas *et al.*, 2003). Initial model selection (see methods below) was
205 conducted to determine which buffer size explained the most variation for both ants and spiders;
206 data at the 6.5 km buffer were used for the remainder of the statistical analyses. Each landscape
207 was analyzed using a “no-sampling” strategy and an 8-cell neighborhood rule for the following
208 class level metrics as specified in Fragstats: CLUMPY, PLAND, GYRATE_MN,
209 GYRATE_AM, and PARA (McGarigal *et al.*, 2012). CLUMPY (fragmentation index) is an
210 index of fragmentation of deciduous forest within the measured landscape where -1 is highly
211 fragmented forest and 1 is a complete, unfragmented forest; the CLUMPY index is not
212 confounded by changes in forest area. PLAND is the percentage of deciduous forest within the
213 landscape. GYRATE_MN is the mean distance (m) to forest edge from patch centroid.
214 GYRATE_AM (patch connectedness), is the area-weighted distance (m) to forest edge from
215 patch centroid, or patch connectedness. PARA (patch edge:area ratio) is the mean edge:area ratio
216 for all deciduous forest patches. We obtained 19 bioclimatic variables from WorldClim v2.0
217 (Fick & Hijmans, 2017); due to the relatively course resolution (~1 km²) of the bioclimatic
218 variables, 2 of our stands (within Brookville) had identical climatic data, but all other stands and

219 sites varied. We included a stand-level vegetation measure of tree species richness ($\text{dbh} \geq 10$ cm)
220 by recording all tree species present within the same 1-ha stands where trees were fogged (Crist
221 *unpublished data*).

222 We accounted for collinearity among landscape and environmental variables by removing
223 those that were highly correlated (Pearson $r \geq 0.80$). In total, 10 variables were included: 4
224 landscape variables, 5 climatic variables, and 1 vegetation variable (Table S2). Prior to all
225 analyses, variables were standardized to z-scores to aid in model fitting and inference. To
226 determine climatic, landscape, and vegetative influence on patterns of taxonomic and functional
227 richness, we used linear regression models (lm function, stats package, R; (R Core Team, 2022)
228 with response variables of mean alpha (α_1 , within trees) and beta (β_1 , among trees) diversity for
229 taxonomic and functional diversity at each stand. Model selection allowed for the additive term
230 of all environmental predictors. To identify best models, we used the lowest Akaike's
231 Information Criterion with bias-correction (AICc). For best models, we calculated AICc weights
232 (w) and appropriate R^2 . We tested for spatial autocorrelation using Moran's I (Moran.I function,
233 ape package, (Paradis & Schliep, 2019), which indicated no spatial autocorrelation present in the
234 residuals of our univariate models.

235 Similarly, we used DISTLM and distance-based redundancy analyses (dbRDA) in
236 PRIMER and PERMANOVA+ (McArdle & Anderson, 2001) to assess influences of our
237 environmental variables on community composition at the stand level. We performed stepwise
238 model selection based on AIC values to examine the relationship between the explanatory
239 variables and community composition. We only included variables with significance of $p < 0.10$
240 in preliminary marginal tests to reduce likelihood of overfitting multivariate analyses.

241 Significance values and variance explained by selected predictors were found using bootstrap
242 tests based on 9999 iterations. Multivariate analyses were conducted on square-root transformed
243 abundance data with Bray-Curtis dissimilarity.

244

245 **3 RESULTS**

246 We collected 3,053 individual ants representing 23 species with 2 singleton species (Table
247 S3). Estimated Chao1 richness was 24 ± 2 species, with rarefaction curves indicating ant richness
248 had plateaued (Figure S3). We collected 5,221 individual spiders representing 23 families, 67
249 genera, and 97 species. Of these, 83.7% were juveniles, we identified the remaining 925 adults to
250 morphospecies (Table S3). Of these species, 15 were represented by a single specimen. Dominant
251 spider families were Araneidae (23%), Linyphiidae (17%), Salticidae (16%), Anyphaenidae
252 (12%), and Theridiidae (10%). Estimated Chao1 richness was 108 ± 6 species, with rarefaction
253 indicating spider richness began to plateau and likely would have been reached by 150 tree
254 samples (Figure S3).

255 *3.1 Hierarchical diversity partitioning*

256 Spider species alpha and beta diversity were higher than ant species diversity, ant
257 functional diversity, and spider functional diversity (Table 1). Multiplicative beta diversity based
258 on species richness ($q = 0$) exhibited similar patterns between taxa, with beta decreasing as scale
259 increased (Figure 1). For species richness, the observed β_{tree} components were 2x the α_{tree} of 4.8
260 species of ants per tree and 2.8x of 6.4 species of spiders per tree, respectively. The β_{stand} values
261 were 1.6x the combined of 9.5 species of ants per stand, and 2.4x the 17.9 species of spiders per
262 stand. The β_{site} components were 1.3x the 15.7 species of ants per site and 1.7x the 42.9 spider

263 species per site, and $\beta_{\text{ecoregion}}$ were $<1.3x$ the mean richness of ants and spiders per ecoregion
264 (Table 1). The functional group diversity components mirrored those of species richness, except
265 that the α_{tree} and β_{tree} levels comprised most of variation in total functional diversity of ants with
266 additional variation explained by β_{stand} for spiders. Randomization tests of species richness
267 indicated α_{tree} was significantly lower than expected across taxa ($p < 0.001$), β_{tree} was higher than
268 expected across taxa ($p < 0.001$), β_{stand} was higher than expected for ant species and spiders ($p <$
269 0.001), β_{site} was higher for spider species ($p = 0.005$), and β_{region} was not different across taxa ($p >$
270 0.025 , Table 1). These patterns emerged, despite having similar diversity across scales for
271 diversity for ant species, ant functional groups, and spider functional groups (Figure 1, Table 1).
272 At local scales (within and among trees), for both taxonomic and functional endpoints, the
273 deviations from observed diversity were greater for ants than spiders, but the opposite pattern
274 emerged at broader scales (stand, site, and region; Figure 1), indicating differential hierarchical
275 patterns of species diversity between ants and spiders.

276

277 *3.2 Community composition*

278 We tested for multivariate differences in ant and spider species composition across
279 hierarchical levels based on variation in Bray-Curtis dissimilarities (Table 2). For both taxa, a
280 greater proportion of the variation in dissimilarities was explained at site and ecoregion levels
281 than with univariate diversity partitions, but components still decreased with increasing spatial
282 scale (Table 2). Spiders also showed greater residual variability among trees (56%) compared to
283 ants (40%). Similarly, spider species composition showed greater variance among stands within
284 sites than ant species composition (Table 2). Ant assemblages showed significant variability

285 across all spatial scales; however, this was not the case for spiders, as the variance component for
286 ecoregion was not different from zero (Table 2).

287 For both ants and spiders, functional composition was less variable than taxonomic
288 composition across scales (Table 2). Ant and spider functional assemblages were significantly
289 variable at the stand and site levels, but the variance component for ecoregion was not different
290 from zero (Table 2). While spider functional composition showed reduced variance components
291 with increasing spatial scale, ant functional composition showed little differentiation among
292 stand, site, and ecoregion scales, but variance was highest among sites (Table 2), indicating
293 stronger variability across sites within ecoregion than among stands within sites. Nonetheless,
294 both ant and spider functional composition was most variable among trees within stands (Table
295 2), suggesting functional differentiation of these communities among tree species.

296

297 *3.3 Patterns of diversity and community composition*

298 Tree identity influenced spider and ant species richness. For ants, white oak (*Quercus*
299 *alba*), red oak (*Q. rubra*), American beech (*Fagus grandifolia*), sugar maple (*Acer saccharum*),
300 and hickory (*Carya* spp.) hosted similar levels of richness, while tulip poplar (*Liriodendron*
301 *tulipifera*) maintained the lowest richness (Figure 2B). Ant richness began to plateau on most tree
302 species (but see hickory, Figure 2A). For spiders, white oak, sugar maple, and American beech
303 had greater richness than tulip poplar, red oak, and hickory, as indicated by rarefaction (Figure
304 2B). Unlike for ants, most tree species did not begin to plateau for spiders, except for red oak.

305 Average alpha (α_1 , within trees) and beta (β_1 , among trees within stand) diversity for
306 taxonomic and functional endpoints were driven by a combination of landscape and climatic

307 variables measured at the stand level. Temperature evenness (isothermality) and precipitation
308 seasonality were important predictors of alpha diversity of spider functional groups, ant species,
309 and ant functional groups as well as beta diversity of spider species, spider functional groups, and
310 ant species (Table 3). Specifically, precipitation seasonality was negatively associated with alpha
311 diversity, but positively associated with beta diversity (Table 3). This suggests that increased
312 variability in precipitation may simultaneously reduce average alpha within trees while increasing
313 differentiation of arthropod communities among tree species (beta). Stand-level richness of tree
314 species was positively associated with beta diversity of ant species, but negatively associated
315 with beta diversity of spider functional groups (Table 3). Finally, patch connectedness, size, and
316 shape were important predictors of alpha diversity of ant and spider functional groups and of beta
317 diversity of ant functional groups (Table 3).

318 Similar to univariate patterns of alpha and beta diversity, variation in multivariate
319 community composition using dbRDA was driven by a similar combination of landscape and
320 climatic drivers. Variation in ant community composition was best explained by precipitation
321 variability ($R^2 = 16.3\%$) (Figure 3A). Variation in ant functional group composition was best
322 explained by isothermality ($R^2 = 40\%$), patch connectedness (14.2%), and precipitation
323 seasonality (11.1%) (Figure 3B). Variation in spider community composition was best explained
324 by isothermality ($R^2 = 17.6\%$), maximum temperature (15.6%), and habitat fragmentation
325 (10.4%) (Figure 3C). Variation in spider functional guild composition was best explained by
326 patch connectedness ($R^2 = 25.2\%$), isothermality (16%), stand tree richness (9.6%), and habitat
327 fragmentation (8.9%) (Figure 3D).

328

329 **4 DISCUSSION**

330 4.1 Spatial structure of diversity

331 Our results indicate differential patterns of beta diversity and community structure
332 between canopy-dwelling ants and spiders and that taxonomic diversity is more variable at larger
333 spatial scales than functional diversity. In support of our hypothesis that spider diversity would be
334 more affected by broader spatial scales than ants, we saw greater diversity and greater deviation
335 from expected diversity (SES) at broader spatial scales for spiders than for ants, but there was
336 more variation between ecoregions for ant communities than for spider communities. While
337 functional and taxonomic diversity exhibited similar patterns across scales, taxonomic diversity
338 exceeded functional diversity at coarser spatial scales (Site and Ecoregion), suggesting trait
339 clustering and functional redundancy at broader spatial scales, consistent with the findings of
340 Jarzyna & Jetz (2018). Essentially, the loss or addition of species at the site and ecoregion scales
341 has little influence on the loss or addition of ecological functions – as determined by functional
342 groups – at these same scales.

343 The finding that broad-scale (Ecoregion) beta components of diversity were not different
344 than expected suggests the effects of ecoregions do not structure the taxonomic and functional
345 diversity of canopy-dwelling ants and spiders. Our results contrast with the significant
346 differentiation of beetle diversity between ecoregions from these same samples (Gering *et al.*,
347 2003) but are consistent with those of Summerville *et al.* (2003) who found no significant
348 deviation from expected diversity in forest moth species richness among ecoregions. Nonetheless,
349 both ant and spider communities showed similar patterns of decreasing community variation with
350 increasing spatial scale, indicating strong differences in beta diversity at the tree and stand levels,
351 consistent with the findings of Gering *et al.* (2003) and Summerville *et al.* (2003). Here, we also

352 conducted multivariate partitions of Bray-Curtis dissimilarity across scales, which showed
353 slightly larger components of variation at the site and ecoregion levels than univariate partitions.
354 This suggests that shifts in the relative abundance of species or functional group across sites and
355 ecoregions were more important than shifts in species or functional group composition.

356 Lower variation than expected at the individual tree-level implies that tree species support
357 distinct levels of species richness and, likely, functional diversity of arthropod taxa, as supported
358 by rarefactions of individual tree species for both ants and spiders. Differences in tree species-
359 arthropod species richness relationships (estimated and observed richness) indicate potential tree-
360 species specific constraints such as prey species abundance and competition/territoriality in ants
361 (Majer & Delabie, 1999; Yasuda & Koike, 2009) and nesting/web site limitations for spiders
362 (Nicolai, 1986; Larrivé & Buddle, 2010). Thus, the maintenance of temperate, canopy-dwelling
363 arthropod communities is, at least somewhat, dependent on maintaining the diversity of host
364 trees. As such, the subsequent emerald ash borer (*Agilus planipennis*) driven loss of ash trees
365 (*Fraxinus* spp.; Herms & McCullough, 2014) and long-term declines of several oak species in
366 eastern forests (McEwan *et al.*, 2011) since the time of sampling has likely resulted in the loss of
367 distinct arboreal arthropod communities and, possibly, species since the early 2000s.

368

369 4.2 Landscape and environmental influences

370 Previous studies have found differential environmental drivers between taxonomic and
371 functional diversity (Longhi & Beisner, 2010; Pool *et al.*, 2010; Meynard *et al.*, 2011),
372 suggesting that different environmental filters act on taxonomic and functional diversity of the
373 same taxa. While we saw similar scaling patterns between ants and spiders, taxonomic and

374 functional diversity, and community structure, the climatic, landscape, and vegetation drivers of
375 these patterns did, indeed, differ (Table 3, Figure 3). Climatic characteristics showed strong
376 influence on alpha and beta diversity as well as both taxonomic and functional composition,
377 while landscape characteristics were important in explaining patterns of functional alpha diversity
378 and functional composition. Nonetheless, taxonomic and functional community composition was
379 explained by shared climatic variables (precipitation seasonality and isothermality), indicating
380 that both taxonomic and functional assemblages are partially driven by the same climatic
381 conditions.

382 Climate variables were more common than landscape variables in univariate (diversity)
383 and multivariate (community composition) ant and spider models. Precipitation seasonality was
384 the most common predictor in explaining variation in taxonomic and functional diversity (Table
385 3), while isothermality (temperature evenness) was the most common predictor for explaining
386 multivariate variation in taxonomic and functional community composition (Figure 3). These
387 differences in climate variables as predictors of univariate and multivariate variation in diversity
388 may also reflect variation in species distributions versus shifts in relative abundances. The
389 broader implication of climate variability as an important driver of diversity and composition of
390 canopy arthropod communities is that climate change mediated increases in climate variability
391 (increased precipitation seasonality and decreased isothermality) will likely greatly alter these
392 canopy-dwelling communities (Westerling, 2016; Jump *et al.*, 2017; Neumann *et al.*, 2017).

393 Habitat configuration was more important in explaining patterns of ant and spider alpha
394 and beta diversity and community structure than total habitat availability, but spider functional
395 richness was related to patch size and shape. In line with our results, previous research suggests

396 that ants show weak support for species-area relationships, while spiders tend to exhibit strong
397 positive species-area relationships (Crist, 2009; Cardoso *et al.*, 2010). Similarly, previous studies
398 have found higher ant richness in habitat fragments with higher connectivity, while habitat
399 connectivity does not influence spider species richness (Abensperg-Traun *et al.*, 1996; Suarez *et*
400 *al.*, 1998; Cardoso *et al.*, 2010). The disparities in the influence of habitat fragmentation and
401 patch size and shape between ants and spiders is likely due to differences in dispersal ability
402 (Thompson & Townsend, 2006). Our results suggest that patterns of arthropod diversity among
403 habitat patches is influenced by dispersal ability, with connectivity being an important predictor
404 of poor dispersing arthropods and patch size and shape being an important predictor of diversity
405 of stronger dispersing arthropods. Taken together, this suggests that the importance of habitat
406 configuration and area of habitat in determining species richness (Haddad *et al.*, 2017) may be
407 somewhat dependent upon the dispersal ability of organisms. Yet, connectivity and fragmentation
408 of habitat patches were important for composition of both taxa, suggesting that while spider
409 richness may not be influenced by habitat configuration, the identities of spider species is
410 influenced by habitat configuration. Therefore, conservation efforts should focus on increasing
411 both patch connectivity and size to provide the largest benefits to diversity of all arthropods.
412 Nevertheless, future research should expand upon the relative roles of habitat configuration and
413 patch size and shape in driving patterns of diversity for taxa across a dispersal gradient.

414 *4.3 Conclusions*

415 Our findings demonstrate stronger scaling patterns of taxonomic diversity than functional
416 diversity from local to regional scales, suggesting functional redundancy at broader spatial scales.
417 Further, taxonomic and functional diversity and community assemblages change along different

418 environmental gradients. The controls of climate and landscape fragmentation on the diversity
419 and structure of canopy-dwelling ants and spider communities indicate that climate change via
420 increased variability will likely further alter the diversity, composition, and function of arboreal
421 arthropods that are already threatened by forest fragmentation and land use changes. Our findings
422 provide further support for the consideration of functional components of diversity and multiple
423 measures of beta diversity in monitoring and conservation (Cadotte *et al.*, 2011; Socolar *et al.*,
424 2016; Isbell *et al.*, 2017; Jarzyna & Jetz, 2018).

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Code and Data accessibility: All data and R code are available on github:

<https://github.com/mahonmb/CanopyArthropodDiversity>. PRIMER and PERMANOVA+ code is available by request.

Tables.

Table 1. Multiplicative partition of alpha and beta diversity components across a hierarchically nested study of canopy ant and spider diversity. *P* values <0.05 are bolded

| Source | Observed Diversity | Expected Diversity | <i>p</i> | SES |
|---|--------------------|--------------------|--------------|--------|
| Ant Species | | | | |
| Ecoregion (β_{region}) | 1.10 | 1.10 | 0.687 | -0.01 |
| Site (β_{site}) | 1.34 | 1.33 | 0.386 | 0.52 |
| Stands (β_{stand}) | 1.66 | 1.55 | 0.001 | 3.94 |
| Among Trees (β_{tree}) | 1.98 | 1.57 | 0.001 | 24.67 |
| Within Trees (α_{tree}) | 4.78 | 6.02 | 0.001 | -19.6 |
| Ant Functional Group | | | | |
| Ecoregion (β_{region}) | 1.00 | 1.00 | 0.500 | 0.00 |
| Site (β_{site}) | 1.06 | 1.04 | 0.348 | 1.27 |
| Stands (β_{stand}) | 1.16 | 1.16 | 0.520 | 0.15 |
| Among Trees (β_{tree}) | 1.56 | 1.33 | 0.001 | 15.23 |
| Within Trees (α_{tree}) | 3.13 | 3.66 | 0.001 | -13.01 |
| Spider Species | | | | |
| Ecoregion (β_{region}) | 1.30 | 1.28 | 0.291 | 0.92 |
| Site (β_{site}) | 1.74 | 1.67 | 0.003 | 2.98 |
| Stands (β_{stand}) | 2.40 | 2.23 | 0.001 | 5.97 |
| Among Trees (β_{tree}) | 2.79 | 2.60 | 0.001 | 7.01 |
| Within Trees (α_{tree}) | 6.42 | 6.88 | 0.001 | -6.56 |
| Spider Functional Group | | | | |
| Ecoregion (β_{region}) | 1.15 | 1.15 | 0.803 | 0.19 |
| Site (β_{site}) | 1.24 | 1.23 | 0.475 | 0.36 |
| Stands (β_{stand}) | 1.32 | 1.26 | 0.001 | 4.27 |
| Among Trees (β_{tree}) | 1.38 | 1.32 | 0.001 | 7.12 |
| Within Trees (α_{tree}) | 8.89 | 9.32 | 0.001 | -6.80 |

Table 2. Permutational multivariate analyses of variance (PERMANOVA) based on Bray-Curtis dissimilarity of square-root transformed abundance data for species and functional groups for both taxa. *P*-values are based on Monte-Carlo randomization, *p* values <0.05 are bolded.

Variance is the square root of the estimated component of variance, to put values on the scale of Bray-Curtis dissimilarities (percent difference among assemblages).

| Endpoint and Source of Variation | <i>df</i> | MS | Pseudo-F | <i>p</i> | Variance | Percent Variance Explained |
|----------------------------------|-----------|--------|----------|------------------|----------|----------------------------|
| Ant Species | | | | | | |
| Ecoregion | 1 | 14407 | 2.41 | 0.042 | 13.3 | 15.9% |
| Site(Ecoregion) | 4 | 5970.3 | 2.31 | <0.001 | 14.5 | 17.4% |
| Stand(Site(Ecoregion)) | 18 | 2589.7 | 1.65 | <0.001 | 16.0 | 19.2% |
| Residual | 72 | 1566.1 | | | 39.6 | 47.5% |
| Ant Functional Groups | | | | | | |
| Ecoregion | 1 | 10354 | 2.71 | 0.066 | 11.7 | 17.8% |
| Site(Ecoregion) | 4 | 3821.6 | 2.61 | 0.003 | 12.1 | 18.5% |
| Stand(Site(Ecoregion)) | 18 | 1463.4 | 1.63 | 0.003 | 11.9 | 18.1% |
| Residual | 72 | 899.2 | | | 30.0 | 45.7% |
| Spider Species | | | | | | |
| Ecoregion | 1 | 15822 | 1.82 | 0.058 | 12.2 | 11.7% |
| Site(Ecoregion) | 4 | 8706 | 1.82 | <0.001 | 15.7 | 15.1% |
| Stand(Site(Ecoregion)) | 18 | 4778.7 | 1.53 | <0.001 | 20.4 | 19.6% |
| Residual | 72 | 3120 | | | 55.9 | 53.7% |
| Spider Functional Groups | | | | | | |
| Ecoregion | 1 | 4849.3 | 2.01 | 0.089 | 7.1 | 13.5% |
| Site(Ecoregion) | 4 | 2415.2 | 1.80 | 0.023 | 8.2 | 15.5% |
| Stand(Site(Ecoregion)) | 18 | 1343.8 | 2.42 | <0.001 | 14.0 | 26.5% |
| Residual | 72 | 554.9 | | | 23.6 | 44.5% |

Table 3. Multiple regressions of tree-level alpha and beta diversity as predicted by stand-level measurements of climate, landscape structure and tree species richness. Regression coefficients of best models as indicated by AICc model selection. All predictor variables were converted to Z scores (SE) to allow for comparison of strength of effects.

| | <i>df</i> | <i>Predictor Variables</i> | | | | | <i>R</i> ² |
|--------------------|-----------|----------------------------|--------------------|------------------|---------------|--------------------|-----------------------|
| | | Patch Connectedness | Patch Edge:Area | Tree Richness | Isothermality | Temp Warm Month | |
| Alpha | | | | | | | |
| Ant Species | 2 | | | | | | -0.73 (0.22) 0.53 |
| Ant Func. Group | 3 | 0.44 (0.18) | | | | | -0.60 (0.18) 0.72 |
| Spider Species | 1 | | | | | | --- |
| Spider Func. Guild | 3 | | -0.99 (0.25) | | | | -1.27 (0.24) 0.70 |
| Beta | | | | | | | |
| Ant Species | 4 | 0.82 (0.21) | | 0.49 (0.20) | | | 0.86 (0.21) 0.63 |
| Ant Func. Group | 2 | | | | | -0.58 (0.26) | 0.33 |
| Spider Species | 2 | | | | -0.54 (0.27) | | 0.29 |
| Spider Func. Guild | 3 | | | -0.52 (0.23) | | | 0.49 (0.23) 0.44 |

Figure 1. Standard effect sizes (SES) of the observed diversity partitions and the diversity deviations from a null model across the hierarchical scales (within trees, among trees, stand, site, and region) for taxonomic and functional diversity. Blue triangles are spiders, pink circles are ants.

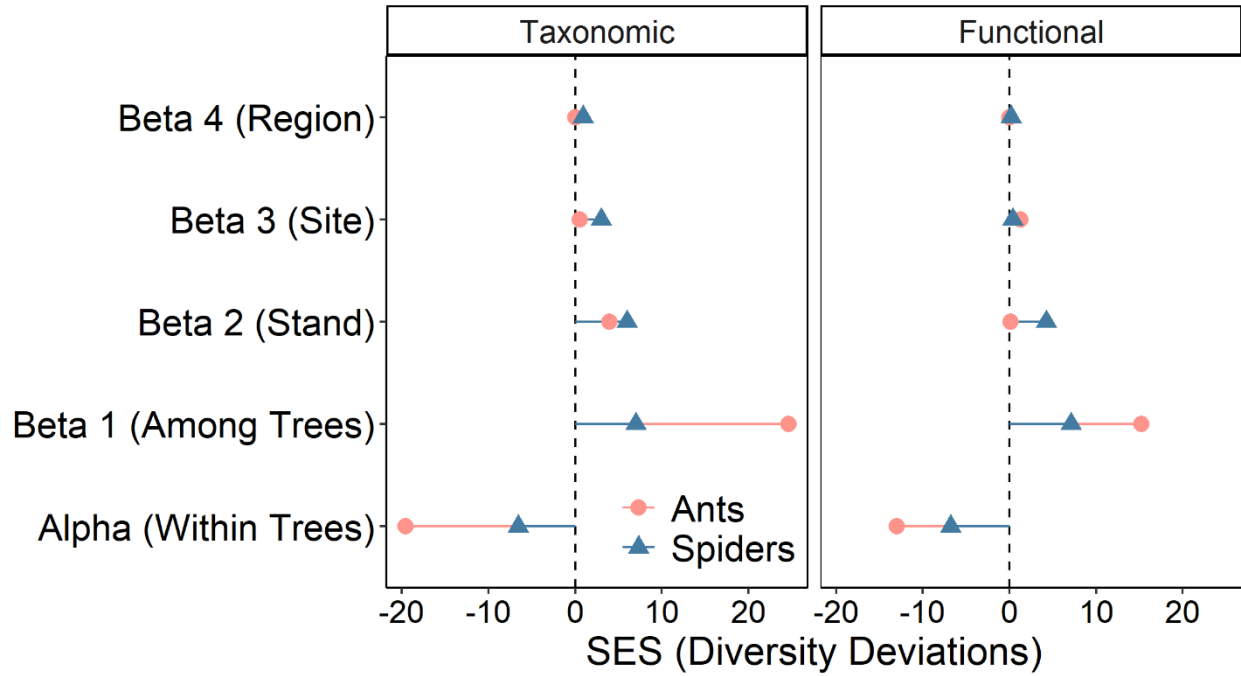


Figure 2. Sample-based rarefaction curves of ant (A) and spider (B) species by number of host trees sampled. Ash, Hackberry, Sycamore, and Walnut trees are not shown, because of few individual trees sampled (< 5 for each species). Number of points on a curve and length of the curve represent the number of individual trees fogged. Ants and spiders show differential patterns among tree species of observed (O) and Chao1 estimated (E) species richness.

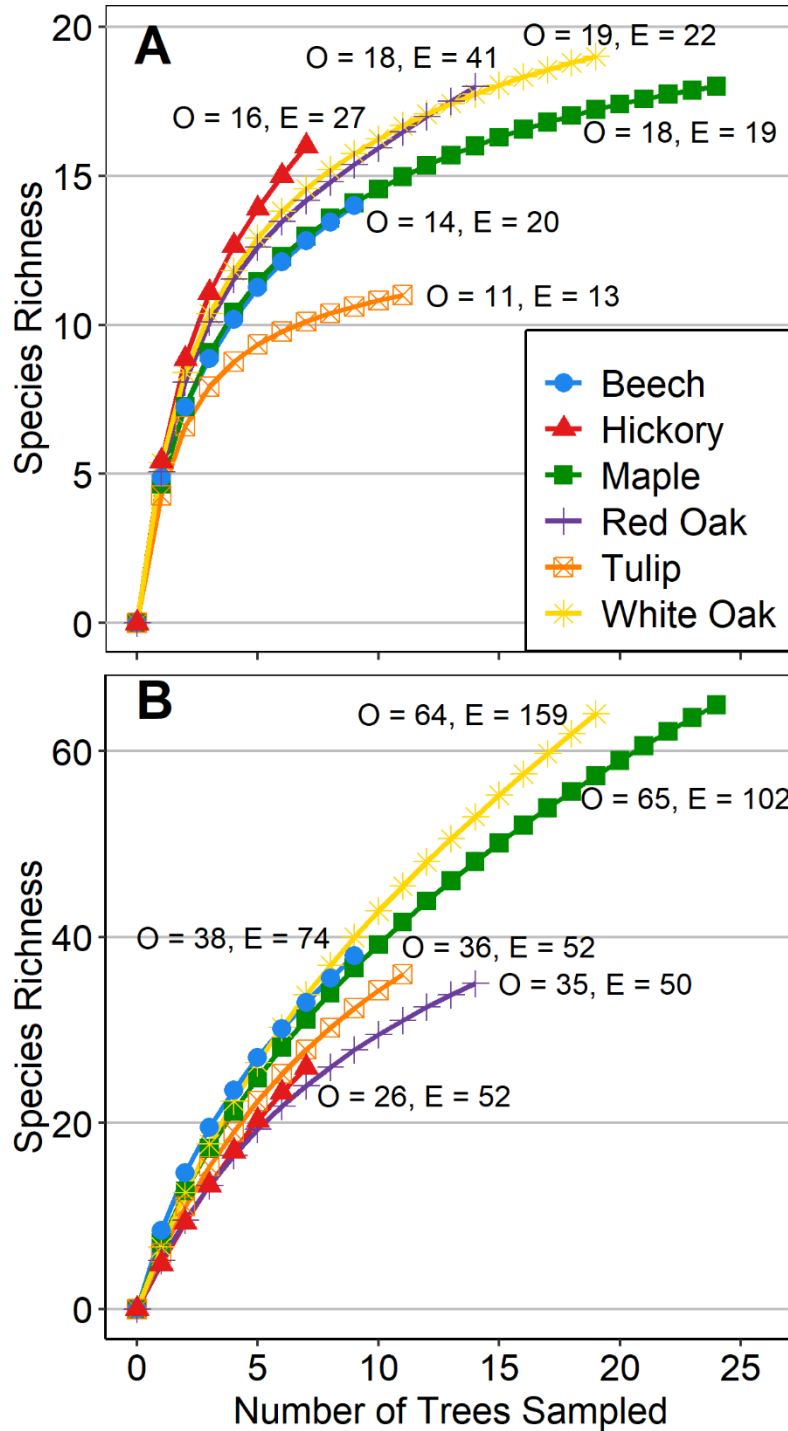


Figure 3. Distance-based redundancy analysis (dbRDA) of stand-level variation in composition based on Bray-Curtis dissimilarities of square-root transformed abundance of ant species (A), ant functional groups (B), spider species (C), and spider functional groups (D). Vectors correspond to predictor variables in best model as determined by model selection. The circles correspond to vector lengths that would have a correlation coefficient of 1 with a given axis, strength of correlation for vectors are scaled to this circle. Yellow symbols are stands within sites in the North-Central Till Plains and purple symbols are stands within sites in the Western Allegheny Plateau. Sites are designated by different symbol shapes.

