bioRxiv preprint doi: https://doi.org/10.1101/2022.08.12.503800; this version posted August 25, 2022. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

# Parasitoid pressures and silence evolution

Megha R. Suswaram<sup>1\*</sup>, Justin D. Yeakel<sup>2</sup> Chaitanya S. Gokhale<sup>3</sup>

<sup>1</sup>Quantitative Systems Biology, University of California Merced,

<sup>2</sup>Life and Environmental Sciences, University of California Merced,

<sup>3</sup>Research Group for Theoretical Models of Eco-evolutionary Dynamics,

Department of Evolutionary Theory, Max Planck Institute for Evolutionary Biology

\*Corresponding author: msuswaram@ucmerced.edu

#### Abstract

Acoustic signals used by organisms to attract mates are known to attract parasitoid 5 flies. The parasitoid flies lay their eggs inside the host signaler, eventually killing the 6 host. We build a host-parasitoid acoustic model to investigate the effect of parasitoid flies 7 on the signalling host's eco-evolutionary dynamics. We used field crickets as a system 8 to build the framework of the model. We explore how the sex ratio and the female para-9 sitoid fecundity impact the evolution of the acoustic signal and population density of the 10 signalling hosts. We also explore the stability of the host populations with an increase in 11 parasitoid load. We find that up to a threshold value, an increase in parasitoid load leads 12 to a thriving yet silent host population. Consistent with field observations, we show how 13 this emergence of silence as an evolutionary strategy is immediate. Our results show that 14 a drastic increase in the parasitoid load can rapidly push the signalling host population 15 towards instability and extinction. 16

Keywords: acoustic signal, singing crickets, parasitism, reproduction, eco-evolutionary dy namics

## **Introduction**

1

2

3

4

Acoustic signalling is the primary mode of communication shared by roughly 8.7 million species ranging from arthropods to mammals [1], inhabiting terrestrial and marine environments. Acoustically signalling species are incredibly diverse, from crickets, anurans, and birds to several marine organisms. The reproductive fitness can be attributed to the mating success of the individual signalers in many acoustically signalling species [2–4]. A consistent life-history trade-off is the one between reproduction and survival citekolluru1999effects. This trade-off determines the evolution of the signalling trait whether visual [5], chemical [6] or indeed acoustic acoustic [7, 8]. Signalling populations evolve to amplify or diminish their conspicuousness based on their natural chorus and other environmental pressures [9–12]. Many organisms solely use acoustic signals to secure mates. In most of these systems, the acoustic signals transmit mating opportunities with the females approaching the calling males to copulate [13]. Females cue into the signal, with some species performing phonotaxis, and locate the male. The females assess the song to assess the quality of the male [14].

Conspicuous sexual signallers, however, also garner unwanted attention. The signals risk 33 attracting potential predators, and exploiters [15]. Male field crickets (Gryllus campestris) use 34 stridulation to produce the chirp by rubbing their front wings together against the underneath 35 of their wing, called the scraper [16]. The parasitoid flies (Euphasiopteryx ochracea) locate 36 the signalling males by eavesdropping on their song by cuing in on the sound produced by the 37 stridulation [17, 18]. The female fly then places larvae on the cricket that then burrow into the 38 cricket's body cavity [19]. They develop there for seven to ten days before emerging. Some 39 parasitoid females show specificity to particular host species [15]. Studies have shown that 40 males switch their mate securing strategies and resort to alternative mating strategies when 41 there is an increased risk of parasitism or can even lose the signalling ability altogether [20]. 42 Both polymorphism and plasticity has been observed in cricket populations [21, 22]. With 43 increased parasitoid densities, male singing crickets have evolved to become silent [23]. The 44 males develop flat wings which are incapable of producing a song. The silent males still have 45 a chance at reproduction, as they become satellite males of the few signalling males in the 46 population. They steal the mates from these signalling males. Additionally, added ecological 47 pressures, compound with parasitoid population density, sex ratio, and fecundity to change 48 the course of host signal evolution [24]. Furthermore, population dynamics create a feedback 49 process that controls the reproduction and mortality rates, thus changing the evolutionary tra-50 jectory of sexual signals [25]. As a culmination of all the confounding factors, the evolutionary 51 loss of the signalling traits can be remarkably rapid, in less than twenty generations, where the 52 population becomes largely silent with a few signalers and many satellite males [23, 26]. 53

Numerous studies focus on the evolutionary significance of parasitoid exploiters and their acoustically signalling insect hosts, such as the above-described crickets [27]. However, a quantitative estimate of the exact parasitoid densities, the proportion of parasitoid females, and other environmental pressures influencing the speed of signal loss are lacking. The existing host-parasitoid models predict the population dynamics as a version of the classical predatorprey cycles. We develop a theoretical model specific to such an extraordinary host-parasitoid system by incorporating reproductive costs and benefits of the acoustic signal.

61 Our mechanistic eco-evolutionary model goes beyond classical evolutionary game-theoretic

reasoning that invokes negative frequency dependence. Specifically, we focus on the parasitoid 62 sex-ratio and the parasitoid fecundity's influence on the acoustic signal evolution and the host 63 population density dynamics. We find a threshold parasitoid load at which there is a rapid evo-64 lutionary transition from conspicuousness to silence in the hosts. We also find that an increase 65 in the parasitoid load decreases the stability of the host population, and the population dynam-66 ics become cyclic, chaotic and eventually go extinct. Our model will thus provide insights 67 into the fundamental mechanisms that affect the evolution of acoustic signals in the presence 68 of parasitoids. 69

### 70 Model

We begin by recapitulating an existing host-parasitoid insect population model [28]. The pop-71 ulation size of the host at a given time t is  $H_t$ , and the population size of the parasitoid is  $P_t$ . 72 The proportion of parasitoid females in the population is q. Simultaneously, a is the search 73 efficiency of the parasitoid and  $F_{max}$  is the maximum fecundity of the parasitoid. The repro-74 ductive rate of the host is represented by r. This model assumes that there are i) q proportion 75 of parasitoid females, ii) a parasitoid female can examine area  $\alpha$  ("area of discovery") during 76 its lifetime, and iii) there is a maximum parasitoid fecundity,  $F_{max}$ . The population dynamics 77 of the host and parasitoid is given by: 78

$$H_{t+1} = H_t \left( e^{-\frac{\alpha P_t F_{max}}{F_{max} + \alpha H_t}} \right) r$$
$$P_{t+1} = H_t \left( 1 - e^{-\frac{\alpha P_t F_{max}}{F_{max} + \alpha H_t}} \right) q \tag{1}$$

Ecologists have extensively used this particular approach to study parasitised insect pop-79 ulations. Host-parasitoid theoretical models typically generate oscillating populations of in-80 creasing amplitude and are by themselves unstable [29-31]. However, this does not accu-81 rately represent what happens in nature. In nature, additional ecological processes like in-82 traspecific competition and spatial heterogeneity can partially or completely stabilise the sys-83 tem. The model developed by Rogers provides a realistic depiction compared to Thompson's 84 1922 model [28] and Nicholson and Bailey's 1935 model [32]. It is a further development of 85 Holling's disc equation [33] including realised fecundity instead of a potential fecundity [28]. 86

### <sup>87</sup> Reproductive fitness of the host based on acoustic trait

We assume that the signalling host's acoustic trait is represented by a single acoustic character, the syllable rate. While we have considered syllable rate for building the model, it can be

any feature of the acoustic signal like amplitude, frequency or intensity. We assume that the 90 syllable rate, z, varies from 0 to 100 units in time. A chorus is formed when many individuals 91 signal together with varying syllable rates. The chorus will thus have a mean syllable rate, 92  $\bar{z}$ . A low syllable rate is when an individual has  $z < \bar{z}$ . A silent individual who does not 93 signal is represented by z = 0, and a high syllable rate when  $z > \overline{z}$  indicates a conspicuous 94 signaler. Given an environment devoid of acoustic interference, we can assume that the mean 95 syllable rate,  $\bar{z}$ , sets the reproductive fitness standard for all calling individuals. If the syllable 96 rate of a calling individual is higher than that of the chorus mean, then the individual stands 97 out from the overall population, is conspicuous, and can be easily distinguished by listening 98 mates. Therefore, it has a higher chance of securing more mates. We can model this as an 99 individual caller, whose syllable rate is higher than that of the chorus' mean,  $z > \overline{z}$ , having a 100 higher reproductive fitness component. While we assume that securing more mates increases 101 the reproductive component of fitness, there is a maximum reproductive reward within a given 102 time frame. The reproductive fitness with maximum reproductive reward is denoted by  $r_{max}$ . 103 Hence, even when the individual's syllable reaches maximum conspicuousness, the reproduc-104 tive fitness component saturates at  $r_{max}$  (Fig. 1). 105

Similarly, suppose the syllable rate of a calling individual is lower than that of the chorus' 106 mean. In that case, the chorus deafens the individual's signal, and therefore it has a lower 107 chance of securing mates. So, we can model an individual whose syllable rate is lower than 108 the chorus' mean,  $z < \overline{z}$ , with a lower reproductive fitness component. Nevertheless, we 109 also assume that even if a caller is silent (a non-calling individual), it can encounter a mate 110 by random chance, with its movement within the habitat. This gives the silent individual a 111 minimum reproductive reward value,  $r_{min}$ . By  $\beta$  we denote the sensitivity of the reproductive 112 curve, considered to be the phonotactic selectivity of the receiver citegerhardt2008phonotactic 113 (Fig. 1). The reproductive component of fitness of an individual signaler is therefore given by: 114

$$r(z|\bar{z}) = r_{min} + \frac{r_{max} - r_{min}}{1 + e^{-\beta(z-\bar{z})}}$$
(2)

#### 115 Search efficiency of the parasitoid based on acoustic trait

We then modelled the search efficiency with a type II functional response. Hence, as the syllable rate increases, the more conspicuous the individual is, the parasitoid can better find the signaller. After a certain threshold of the syllable rate, the search efficiency of the parasitoid saturates (Fig. 1). The search efficiency  $\alpha$  is,

$$\alpha = \frac{z}{1+z} \tag{3}$$

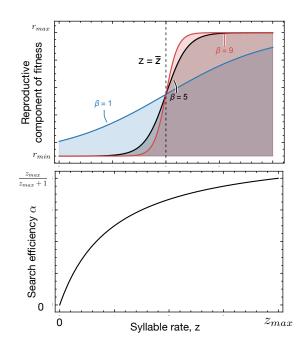


Figure 1: Reproductive component of fitness function of an individual host Eq. (2), Search efficiency (Eq. (3)) of a parasitoid as a function of mean chorus syllable rate. The reproductive component of fitness is dependent on how far the syllable rate of the individual, z is from the population chorus mean  $\bar{z}$ , and the phonotactic selectivity,  $\beta$ .  $r_{min}$  and  $r_m ax$  are the minimum and maximum reproductive rewards respectively. With high phonotactic selectivity there is a larger benefit in reproductive component of fitness. The search efficiency of the parasitoid,  $\alpha$ , increases with increase in host syllable rate and saturates at a high syllable rates.

#### **120 Density dependence**

We add density dependence to the insect population host-parasitoid framework. This entails replacing the host's reproduction and search efficiency of the parasitoid with the newly formulated acoustic character-based modifications. By substituting Eq. (2) and q. (3) in Eq. (1), and adding density dependence with K being the carrying capacity of the habitat, we get:

$$H_{t+1} = H_t \underbrace{e^{\left(r_{min} + \frac{r_{max} - r_{min}}{1 + e^{-\beta(z-\bar{z})}}\right)(1 - H_t/K) - \left(\frac{\bar{z} + z}{F_m ax + \frac{z}{1 + z} H_t}\right)}_{\text{prob. of esc. parasitism (density dependent)}}$$

$$P_{t+1} = H_t \underbrace{\left(1 - e^{-\frac{\bar{z} + z}{F_m ax + \frac{z}{1 + z} H_t}\right)}_{\text{prob. of parasitism}} q \qquad (4)$$

#### 125 Simulation of the eco-evolutionary dynamics

Because an analytical solution for the average fitness of the population  $\bar{w}(\bar{z})$  is intractable, 126 we track changes in population abundance and the trait distributions of a cricket population 127 with an individual-based model. As such, we numerically track the evolution of the full trait 128 distribution of z, denoted by f(z), over time, in addition to population size N(t). Each time 129 step represents a generation where all adults are assumed to die at the end of each time step 130 such that generations are non-overlapping. Such dynamics are generally the case for cricket 131 populations [34]; however, this assumption would not hold for many other acoustically sig-132 nalling organisms, including most bird species. Offspring inherit trait values from parents 133 with variability  $\sigma$  such that, 134

$$z_{offspring}(t+1) = z_{parent}(t) + g,$$
(5)

where  $g \sim N(0, \phi)$ , and we set  $\phi = 1$ . The number offspring for each individual *i* with trait trais  $z_i$  is determined by its fitness with respect to the chorus mean,  $w_i(z_i|\bar{z})$ , where the sum across reproducing individuals determines the future population size N(t + 1), such that

$$N(t+1) = \sum_{i=1}^{N(t)} w_i(z_i | \bar{z}).$$
(6)

We initiate the host population to be conspicuous  $\bar{z}(0) = 90$  with a standard deviation of  $\sigma = 10$ , and simulate dynamics over the course of 100 generations. We confirmed 100 generations to be adequate for calculating steady-state conditions. We assumed steady state when the population density did not change more than  $\epsilon = 0.00001$  for more than 50 timesteps. We set  $r_{min} = 0.01$ ,  $r_{max} = 2 \beta = 0.5$ , and K = 1000. The initial host population is represented in Fig. 2 (t = 1). bioRxiv preprint doi: https://doi.org/10.1101/2022.08.12.503800; this version posted August 25, 2022. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

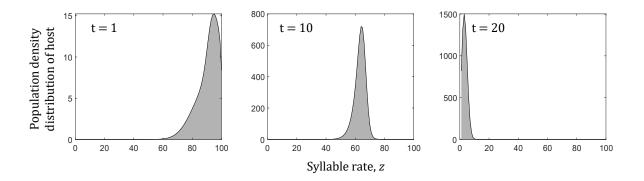


Figure 2: Initial distribution of the host population. With increasing generations, the host population evolves from a conspicuous chorus mean to a silent chorus mean within 20 generations. We see the evolution of silence within 20 generations in a population that has been attacked by parasitoids. The silent population has a higher population density. H(0) = 500,  $\bar{z}(0) = 90$ ,  $\sigma = 10$ ,  $r_{min} = 0.01$ ,  $r_{max} = 2 \beta = 0.5$ , and K = 1000.

### **Results**

#### **145** Quick transition from conspicuousness to silence

Female biased sex-ratio. As the proportion of parasitoid females increases, the conspicu-146 ously signalling population quickly evolves into silence. The reproductive fitness of the host 147 individual is influenced by the chorus mean,  $\bar{z}$ . The conspicuous individuals' fitness is ini-148 tially high, resulting in reproductive gains. As the parasitoid females attack the conspicuous 149 host individuals, mortality increases and the fitness of conspicuous individuals is reduced. In 150 contrast, the silent host individuals escape parasitoid attack, reproduce, and increase the pro-151 portion of silent individuals in the host population in the next generation, reducing the chorus 152 mean. With an increasing proportion of parasitoid females, there are more attacks on the con-153 spicuous hosts, and the evolution of silence is favoured (Fig. 3). This evolutionary transition is 154 rapid, and it usually happens within ten generations in our study. The silent individuals survive 155 and reproduce, increasing the population density to the carrying capacity (Fig. 3). Beyond a 156 specific female parasitoid load, all the host individuals die, and the population goes extinct 157 (Fig. 3). Thus a female-biased sex ratio of parasitoids influences the signalling trait evolution 158 and drives population extinction of the host. 159

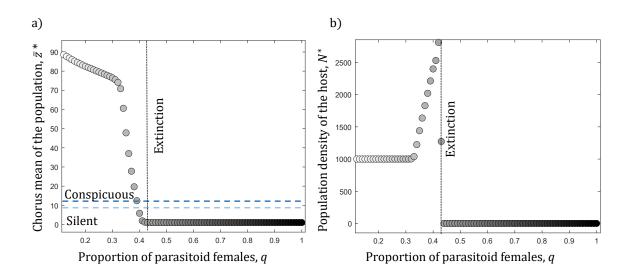


Figure 3: Quick transition from conspicuous to silence. a) Change in chorus mean of the host population represented by circle shapes. b) Change in population size of the host population. A more female-biased sex ratio in parasitoid populations results in a host population that is silent and large. Beyond a threshold of female-biased population in the parasitoids, the host population goes extinct. H(0) = 500,  $\bar{z}(0) = 90$ ,  $\sigma = 10$ ,  $r_{min} = 0.01$ ,  $r_{max} = 2$ ,  $\beta = 0.5$ ,  $F_{max} = 3$  and K = 1000.

Parasitoid fecundity influences the signalling trait evolution and population dynamics of 160 **the host** As the maximum number of viable offspring of the parasitoid females increases, 161 the conspicuously signalling population abruptly transitions to silence (Fig. 4). The increase 162 in parasitoid fecundity increases their population density in the next generation. This results 163 in more number of attacks on conspicuous hosts. The conspicuous individuals die, and the 164 proportion of silent individuals increases in the population, driving the chorus mean to silence. 165 This evolutionary transition of the host is rapid and occurs within ten generations. As we 166 further increase the fecundity of parasitoids, the population goes extinct (Fig. 4). 167

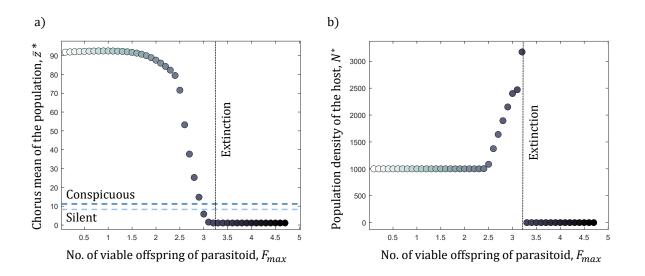
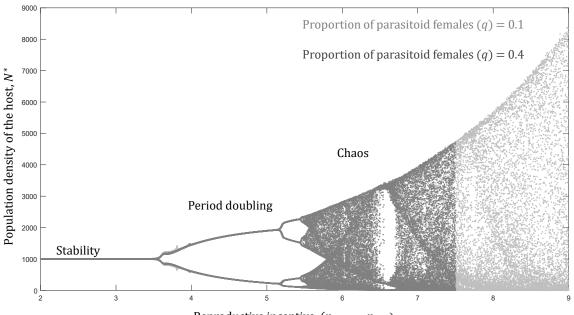


Figure 4: Quick transition from conspicuous to silence. a) Change in chorus mean of the host population. b) Change in population size of the host population. Increase in the fecundity of parasitoid females results in a host population that is silent and large. Beyond a threshold of fecundity of the female parasitoids, the host population goes extinct. H(0) = 500,  $\bar{z}(0) = 90$ ,  $\sigma = 10$ ,  $r_{min} = 0.01$ ,  $r_{max} = 2$ ,  $\beta = 0.5$ , q = 0.4 and K = 1000.

#### 168 Stability of the system and bifurcation analysis

The reproductive incentive to signal is given by the difference in the maximum reproductive 169 fitness  $r_{max}$  and the minimum reproductive reward  $r_{min}$ . Keeping the reproductive reward 170 constant at a low value,  $r_{min} = 0.01$ , as we increase the maximum reproductive fitness, we 171 increase the reproductive incentive. With the increase in reproductive incentive to signal, 172 multiple population steady-state values emerge, such that the host population enters cyclic 173 oscillations. We first see period-doubling in the host population density as the reproductive 174 incentive increases. With a high reproductive incentive, the population size is similarly high. 175 The costs of overpopulation are then determined by the carrying capacity, driving the pop-176 ulation extinct. The stability of the system and the start of period-doubling changes with 177 parasitoid load. As we increase the parasitoid load (increasing fecundity or the proportion of 178 females), the populations go extinct Fig. 5. 179



Reproductive incentive,  $(r_{max} - r_{min})$ 

Figure 5: **Bifurcation diagram of the host system.** Outcome of host population when parasitoid population is male-biased shown in light grey and female-biased shown in dark grey. A female-biased parasitoid population causes the host population to be less stable and drive it towards extinction sooner.

We also plotted the chorus mean and the size of the host population for a varying sex ra-180 tio and fecundity of the parasitoid (Fig. 6). When the parasitoid population is more male or 181 female biased, and the fecundity of the parasitoid is low, the host population remains conspic-182 uous. If the sex ratio relatively is male-biased, but there is high fecundity, then the population 183 is polymorphic with a mix of singing and silence. The host population goes extinct with a 184 female-biased sex ratio and high fecundity (Table. 1). The system is less stable and pushed to-185 wards chaos for a female-biased parasitoid population with high fecundity and the population 186 is subsequently driven to extinction. 187

Parasitoid Condition		Host Outcome	
Sex-ratio	Fecundity	Chorus	Population density
Male-biased	Low	Conspicuous	High
Male-biased	High	Intermediate	Intermediate
Female-biased	Low	Conspicuous	High
Female-biased	High	Silent	Low

Table 1: Effect of parasitoid condition on the host population

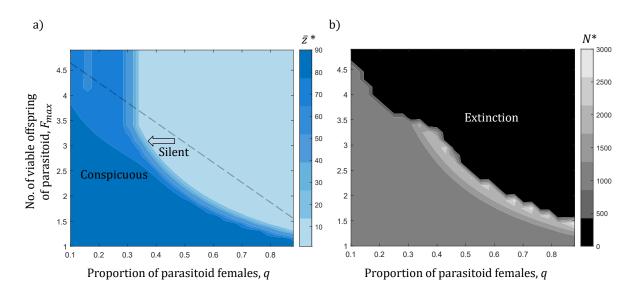


Figure 6: Stability analysis of the host system a) Chorus mean of the host population. Dark blue represents a conspicuous population and light blue represents a silent population. The blue transparent dashed line shows approximate extinction in population. Anything above the line is extinction and not silence. b) Population size of the host population. A high parasitoid load, pushes the host population to silence with a high population density. Beyond, a certain threshold of parasitoid load, the population enters cycles, followed by chaos, and then goes extinct. H(0) = 500,  $\bar{z}(0) = 90$ ,  $\sigma = 10$ ,  $r_{min} = 0.01$ ,  $r_{max} = 2$ ,  $\beta = 0.5$ , and K = 1000.

### **Discussion**

Acoustic signal evolution can result from diversification of the signal [35], amplification of 189 the existing signal to a conspicuous population citemhatre2016stay, diminishing of a signal 190 into a silent population [36] and, loss of signal by a morphological change in the signal pro-191 ducing organ [23], loss of preference for conspicuous signalers from the receiver [37]. This 192 evolution of the acoustic signal can be a result of population dynamics [38, 39], and environ-193 mental pressures [40]. Parasitism is one of the major drivers of diminishing acoustic signals in 194 various signalling species [41]. Several empirical studies have shown that with an increase in 195 parasitoid load, the acoustic signal is diminished and lost within a few generations [25, 42, 43] 196 Various aspects of the effect of parasitoids on host signalling traits and population sizes have 197 been documented [41, 44]. As discussed previously, the existing host-parasitoid models do not 198 predict the host's trait evolution and only focus on the population densities [28, 31, 32]. We 199 modelled the parasitoid load, its impacts on the host population densities, and their signalling 200 trait evolution. We have elucidated the critical components of the parasitoid load that affect 201

the host population and identify the threshold load at which the host population responds by the loss of the signal and evolution of silence.

Our results show that the sex ratio and the fecundity of the parasitoid population play a key 204 role in the switch from conspicuousness to silence. Specifically, the sex ratio and fecundity 205 in the parasitoid population drive the signalling trait evolution, which in turn influences host 206 population dynamics and stability (Fig. 3, Fig. 4, and Fig. 5). The evolution from conspicu-207 ousness to silence occurs within ten generations. Empirical studies corroborate these results. 208 Among field crickets, it is known that signalling males are more affected by the parasitoid fly 209 of the genus Ormia [18]. Female parasitoid flies find a specific cricket host depositing their 210 parasitoid offspring. The offspring then grow and feed within the host. Within ten days, the 211 fully grown larvae kill the host as they emerge [45]. Therefore one would estimate that the 212 sex ratio in parasitoid populations and the offspring size play vital roles in shaping the host's 213 survival and signalling behaviour. Our results show that the increasing female-biased sex ratio 214 and fecundity quickly drive the populations to evolve silence. Zuk et al. (2006) showed that on 215 the Hawaiian island of Kauai, where there was high parasitoid density, most of the population 216 evolved silence through selection for a flatwing morphology that rendered the males incapable 217 of generating an acoustic signal within twenty generations. To reproduce, they used alterna-218 tive mating strategies such as increased locomotive behaviour to encounter females randomly 219 [44]. The resulting rapid and short transition period towards silence conforms with the results 220 of our model. 221

A high reproductive incentive causes the host population destabilization, period doubling, 222 chaos, and extinction (Fig. 5). Population densities of the host can be highly susceptible, and 223 the risk of infection increases with parasitoid density [46, 47]. When subject to varying preda-224 tor densities, Allee effects or parasitism, fast-growing populations are highly sensitive to initial 225 population densities [48]. Such populations can exhibit chaotic dynamics that may promote 226 extinction risk [49–53]. We show that the reproductive rate drives the population expansion, 227 stability, and ultimately extinction (Fig. 5). Our results corroborate natural observations of 228 rapidly growing populations which first thrive, reach a critical threshold, exhibit chaotic dy-229 namics and then go extinct [49, 54, 55]. Also, rapid oscillations in population densities can 230 lead to species extinction [56]. Such oscillations are dictated by the reproductive incentive, 231 moulded by the parasitoid load. An increased parasitoid load pushes the system faster toward 232 instability (Fig. 5). This observation implies a threshold beyond which parasitoid load cannot 233 increase in nature as it drives the host population extinct. 234

Empirical studies have independently shown that parasitoids cause shifts in host traits [15]. Theoretical models and other empirical findings also point to the effects of parasitoids on host populations [28, 31, 32]. We outline the missing link: the connection between the parasitoid load, acoustic trait evolution and population density. One would expect the parasitoids first to change the host population density and the evolution of silence to be a response to the population change. Counter-intuitively, we show that the parasitoid load first alters the reproductive rate of the host population, causing an evolutionary signal adaptation in the host. The population density then responds to this evolutionary signal adaptation (Fig. 6).

Our model provides a general framework for organisms that use acoustic signals to se-243 cure mates and exploited by predators or parasitoids. This framework to predict the signal 244 evolution change applies to crickets, cicadas, anurans, sparrows, etc. Further, the model can 245 be expanded and developed to understand the effects of parasitism on host sex ratios [57]. 246 The model could also be further developed to understand the evolution of multi-component 247 auditory signals, anthropogenic effects, and specifically to climate change acoustic behaviour 248 distributions [58–60]. Changes in the environment can cause significant changes in the re-249 productive incentive of signalers [40, 61, 62]. Changes in the environment can also shift the 250 parasitoid load [63]. At different parasitoid loads, we have shown that drastic changes in evo-251 lutionary adaptations of signalling hosts and their population stability are possible for a given 252 reproductive incentive. Together with environmental conditions that alter the reproductive in-253 centive and parasitoid load, our model's findings can provide insight into their reproduction, 254 trait evolution and population densities. In future studies, we aim to leverage our findings 255 to develop strategies for conserving the acoustic communicating populations under changing 256 environmental conditions. Artificial manipulation of the parasitoid population size, sex ratios, 257 and fecundity may provide a path forward. 258

Acknowledgements. We thank Irina Birskis-Barros, Taran Rallings, and Ritwika VPS for their helpful comments and suggestions. This manuscript benefited from University of California, Merced's School of Natural Sciences Dean Fellowship. Chaitanya S. Gokhale acknowledges funding from the Max Planck Society.

Data Availability and Analysis. All code and preliminary plots are available on GitHub at https://github.com/meghasr92/parasitismsilence.

**Code availability**. https://github.com/meghasr92/parasitismsilence.

### 266 **References**

- [1] Chen, Z. & Wiens, J. J. The origins of acoustic communication in vertebrates. *Nature communications* 11, 1–8 (2020).
- [2] Gerhardt, H. C. & Huber, F. Acoustic communication in insects and anurans: common
   problems and diverse solutions (2003).

- [3] Kumar, A. Acoustic communication in birds. *Resonance* **8**, 44–55 (2003).
- [4] Hedwig, B. Insect hearing and acoustic communication (2014).
- [5] Nokelainen, O., Hegna, R. H., Reudler, J. H., Lindstedt, C. & Mappes, J. Trade-off
   between warning signal efficacy and mating success in the wood tiger moth. *Proceedings* of the Royal Society B: Biological Sciences 279, 257–265 (2012).
- [6] Penn, D. & Potts, W. K. Chemical signals and parasite-mediated sexual selection. *Trends in Ecology & Evolution* 13, 391–396 (1998).
- [7] Stevens, M. Sensory ecology, behaviour, and evolution (Oxford University Press, 2013).
- [8] Barbosa, F., Rebar, D. & Greenfield, M. D. Reproduction and immunity trade-offs con strain mating signals and nuptial gift size in a bushcricket. *Behavioral Ecology* 27, 109–
   117 (2016).
- [9] Wollerman, L. & Wiley, R. H. Background noise from a natural chorus alters female
   discrimination of male calls in a neotropical frog. *Animal Behaviour* 63, 15–22 (2002).
- [10] Vélez, A., Schwartz, J. J. & Bee, M. A. Anuran acoustic signal perception in noisy
   environments. In *Animal communication and noise*, 133–185 (Springer, 2013).
- [11] Parris, K. M., Velik-Lord, M. & North, J. M. Frogs call at a higher pitch in traffic noise.
   *Ecology and Society* 14 (2009).
- [12] Slabbekoorn, H. & Peet, M. Birds sing at a higher pitch in urban noise. *Nature* 424, 267–267 (2003).
- [13] Searcy, W. A. & Andersson, M. Sexual selection and the evolution of song. *Annual Review of Ecology and Systematics* 17, 507–533 (1986).
- [14] Otter, K., Chruszcz, B. & Ratcliffe, L. Honest advertisement and song output during the
   dawn chorus of black-capped chickadees. *Behavioral Ecology* 8, 167–173 (1997).
- [15] Zuk, M. & Kolluru, G. R. Exploitation of sexual signals by predators and parasitoids.
   *The Quarterly Review of Biology* 73, 415–438 (1998).
- [16] Chapman, R. F. & Chapman, R. F. *The insects: structure and function* (Cambridge university press, 1998).
- [17] Cade, W. Acoustically orienting parasitoids: fly phonotaxis to cricket song. *Science* 190, 1312–1313 (1975).

- <sup>300</sup> [18] Robert, D., Amoroso, J. & Hoy, R. R. The evolutionary convergence of hearing in a <sup>301</sup> parasitoid fly and its cricket host. *Science* **258**, 1135–1137 (1992).
- [19] Adamo, S., Robert, D. & Hoy, R. Effects of a tachinid parasitoid, ormia ochracea, on
   the behaviour and reproduction of its male and female field cricket hosts (gryllus spp).
   *Journal of Insect Physiology* 41, 269–277 (1995).
- [20] Heinen-Kay, J. L. & Zuk, M. When does sexual signal exploitation lead to signal loss?
   *Frontiers in Ecology and Evolution* 7, 255 (2019).
- <sup>307</sup> [21] Rüppell, O. & Heinze, J. Alternative reproductive tactics in females: the case of size <sup>308</sup> polymorphism in winged ant queens. *Insectes Sociaux* **46**, 6–17 (1999).
- [22] Ellers, J. & Liefting, M. Extending the integrated phenotype: Covariance and correlation
   in plasticity of behavioural traits. *Current Opinion in Insect Science* 9, 31–35 (2015).

[23] Zuk, M., Rotenberry, J. T. & Tinghitella, R. M. Silent night: adaptive disappearance of
 a sexual signal in a parasitized population of field crickets. *Biology letters* 2, 521–524
 (2006).

- Rotenberry, J. T. & Zuk, M. Alternative reproductive tactics in context: how demogra phy, ecology, and behavior affect male mating success. *The American Naturalist* 188, 582–588 (2016).
- [25] Wilkins, M. R., Seddon, N. & Safran, R. J. Evolutionary divergence in acoustic signals:
  causes and consequences. *Trends in ecology & evolution* 28, 156–166 (2013).
- [26] Tinghitella, R. Rapid evolutionary change in a sexual signal: genetic control of the mutation 'flatwing'that renders male field crickets (teleogryllus oceanicus) mute. *Heredity* **100**, 261–267 (2008).
- [27] Lehmann, G. U. & Lakes-Harlan, R. Adaptive strategies in life-history of bushcrickets
   (orthoptera) and cicadas (homoptera) to parasitoids pressure on their acoustic communi cation systems—a case for sociality? *Frontiers in Ecology and Evolution* 7, 295 (2019).
- [28] Rogers, D. Random search and insect population models. *The Journal of Animal Ecology* 369–383 (1972).
- [29] Jansen, V. A. The dynamics of two diffusively coupled predator-prey populations. *Theoretical Population Biology* 59, 119–131 (2001).

- [30] Briggs, C. J. & Hoopes, M. F. Stabilizing effects in spatial parasitoid–host and predator– prey models: a review. *Theoretical population biology* **65**, 299–315 (2004).
- [31] May, R. M. Stability and complexity in model ecosystems. In *Stability and Complexity in Model Ecosystems* (Princeton university press, 2019).
- [32] Nicholson, A. J. & Bailey, V. A. The balance of animal populations.—part i. In *Proceedings of the zoological society of London*, vol. 105, 551–598 (Wiley Online Library, 1935).
- <sup>336</sup> [33] Holling, C. S. Some characteristics of simple types of predation and parasitism1. *The* <sup>337</sup> *canadian entomologist* **91**, 385–398 (1959).
- [34] Masson, M. V. *et al.* Bioecological aspects of the common black field cricket, gryllus
   assimilis (orthoptera: Gryllidae) in the laboratory and in eucalyptus (myrtaceae) planta tions. *Journal of Orthoptera Research* 29, 83–89 (2020).
- [35] Symes, L., Ayres, M., Cowdery, C. & Costello, R. Signal diversification in oecanthus
   tree crickets is shaped by energetic, morphometric, and acoustic trade-offs. *Evolution* 69, 1518–1527 (2015).
- [36] Gray, B., Bailey, N. W., Poon, M. & Zuk, M. Multimodal signal compensation: do field
   crickets shift sexual signal modality after the loss of acoustic communication? *Animal behaviour* 93, 243–248 (2014).
- [37] Bailey, N. W. & Zuk, M. Acoustic experience shapes female mate choice in field crickets.
   Proceedings of the Royal Society B: Biological Sciences 275, 2645–2650 (2008).
- [38] von Helversen, D., von Helversen, O. & Heller, K.-G. When to give up responding
   acoustically in poecilimon bushcrickets: a clue to population density. *Articulata* 27,
   57–66 (2012).
- [39] Reichard, D. G. & Anderson, R. C. Why signal softly? the structure, function and
   evolutionary significance of low-amplitude signals. *Animal Behaviour* 105, 253–265
   (2015).
- [40] Bowen, A. E., Gurule-Small, G. A. & Tinghitella, R. M. Anthropogenic noise reduces
   male reproductive investment in an acoustically signaling insect. *Behavioral Ecology and Sociobiology* 74, 1–8 (2020).

[41] Kolluru, G. R. *The effects of an acoustically-orienting parasitoid fly Ormia ochracea on reproduction in the field cricket, Teleogryllus oceanicus: a trade-off between natural and sexual selection* (University of California, Riverside, 1999).

- [42] Garamszegi, L. Z. Bird song and parasites. *Behavioral Ecology and Sociobiology* 59, 167–180 (2005).
- [43] Rayner, J., Aldridge, S., Montealegre-Z, F. & Bailey, N. W. A silent orchestra: convergent song loss in hawaiian crickets is repeated, morphologically varied, and widespread.
   *Ecology* (2019).
- <sup>366</sup> [44] Balenger, S. L. & Zuk, M. Roaming romeos: male crickets evolving in silence show
   <sup>367</sup> increased locomotor behaviours. *Animal Behaviour* 101, 213–219 (2015).
- [45] Zukl, M., Simmons, L. W. & Cupp, L. Calling characteristics of parasitized and unpar asitized populations of the field cricket teleogryllus oceanicus. *Behavioral Ecology and Sociobiology* 33, 339–343 (1993).
- <sup>371</sup> [46] Anderson, R. M. & May, R. M. Regulation and stability of host-parasite population <sup>372</sup> interactions: I. regulatory processes. *The journal of animal ecology* 219–247 (1978).
- [47] Steinhaus, E. A. Crowding as a possible stress factor in insect disease. *Ecology* 503–514
   (1958).
- [48] Hart, E. M. & Avilés, L. Reconstructing local population dynamics in noisy metapopulations—the role of random catastrophes and allee effects. *PloS one* 9, e110049 (2014).
- <sup>377</sup> [49] Schreiber, S. J. Allee effects, extinctions, and chaotic transients in simple population <sup>378</sup> models. *Theoretical population biology* **64**, 201–209 (2003).
- <sup>379</sup> [50] Hastings, A. Transients: the key to long-term ecological understanding? *Trends in ecology & evolution* 19, 39–45 (2004).
- [51] Gokhale, C. S., Papkou, A., Traulsen, A. & Schulenburg, H. Lotka-Volterra dynamics
   kills the Red Queen: population size fluctuations and associated stochasticity dramati cally change host-parasite coevolution. *BMC Evolutionary Biology* 13, 254 (2013).
- <sup>384</sup> [52] Schenk, H., Traulsen, A. & Gokhale, C. S. Chaotic provinces in the kingdom of the Red
   <sup>385</sup> Queen. *Journal of Theoretical Biology* 431, 1–10 (2017).
- <sup>386</sup> [53] Hastings, A. *et al.* Transient phenomena in ecology. *Science* **361** (2018).

- <sup>387</sup> [54] Sinha, S. & Parthasarathy, S. Unusual dynamics of extinction in a simple ecological <sup>388</sup> model. *Proceedings of the National Academy of Sciences* **93**, 1504–1508 (1996).
- [55] Schreiber, S. J. Chaos and population disappearances in simple ecological models. *Journal of Mathematical Biology* 42, 239–260 (2001).
- [56] Misra, J. & Mitra, A. Instabilities in single-species and host-parasite systems: period doubling bifurcations and chaos. *Computers & Mathematics with Applications* 52, 525–
   538 (2006).
- <sup>394</sup> [57] Venkateswaran, V. R., Roth, O. & Gokhale, C. S. Consequences of combining sex-<sup>395</sup> specific traits. *Evolution* **75**, 1274–1287 (2021).
- Isological conservation 143, 1635–1645 (2010).
- <sup>398</sup> [59] Roca, I. T. *et al.* Shifting song frequencies in response to anthropogenic noise: a meta <sup>399</sup> analysis on birds and anurans. *Behavioral Ecology* 27, 1269–1274 (2016).
- [60] Desjonquères, C. *et al.* Acoustic species distribution models (asdms): A framework
   to forecast shifts in calling behaviour under climate change. *Methods in Ecology and Evolution* (2022).
- [61] Nabi, G. *et al.* The possible effects of anthropogenic acoustic pollution on marine mammals' reproduction: an emerging threat to animal extinction. *Environmental science and pollution research* 25, 19338–19345 (2018).
- [62] Lengagne, T. Traffic noise affects communication behaviour in a breeding anuran, hyla
  arborea. *Biological conservation* 141, 2023–2031 (2008).
- <sup>408</sup> [63] Phillips, J. N., Ruef, S. K., Garvin, C. M., Le, M.-L. T. & Francis, C. D. Background
   <sup>409</sup> noise disrupts host–parasitoid interactions. *Royal Society open science* 6, 190867 (2019).