Title: Long term maintenance of heterologous symbiont association in *Acropora palmata* on

natural reefs

Running Head: Symbiont community dynamics in out-planted *Acropora palmata*

Holland Elder[1]*

Wyatt C. Million[1]

Erich Bartels[2]

Cory Krediet[3]

Erinn Muller[4]

Carly D. Kenkel[1]

1. University of Southern California, 3616 Trousdale Parkway, AHF 231, Los Angeles, CA 90089

2. Mote Marine Laboratory, 24244 Overseas Hwy, Summerland Key, FL 33042

3. Eckerd College, 4200 54th Ave., St. Petersburg, FL 33711

4. Mote Marine Laboratory, 1600 Ken Thompson Parkway, Sarasota, FL 34236

*corresponding author

760-622-9116

hollandelder@gmail.com

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Abstract

The sensitivity of reef-building coral to elevated temperature is a function of their symbiosis with dinoflagellate algae in the family Symbiodiniaceae. Changes in the composition of the endosymbiont community in response to thermal stress can increase coral thermal tolerance. Consequently, this mechanism is being investigated as a human-assisted intervention for rapid acclimation of coral in the face of climate change. Successful establishment of novel symbioses that increase coral thermal tolerance have been demonstrated in laboratory conditions; however, it is unclear how long these heterologous relationships persist in nature. Here, we show that *Acropora palmata* can form a novel symbiosis with *Durusdinium spp*. when reared in land-based aquaria. We tested the stability of this heterologous relationship by outplanting clonal replicates (ramets) of five *A. palmata* host genotypes to natural reefs in the lower Florida Keys. Amplicon sequencing analysis of ITS2-type profiles revealed that the majority of surviving ramets remained dominated by *Durusdinium spp*. two years after transplantation. However, 15% of ramets, including representatives of all genotypes, did exhibit some degree of symbiont shuffling or switching at six of eight sites, including complete takeover by site-specific strains of the native symbiont, *Symbiodinium fitti*. The predominant long-term stability of the novel symbiosis supports the potential effectiveness of symbiont modification as a management tool. However, the finding that 6–7 year-old coral can alter symbiont community composition in the absence of bleaching indicates that Symbiodiniaceae communities are indeed capable of greater flexibility under ambient conditions.

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Increasingly frequent and severe thermal stress events are the predominant threat to reef

ecosystems in the Anthropocene. Thermal stress perturbs the photosynthetic mechanism and can

result in loss or expulsion of the dinoflagellate algae in the process known as bleaching [1]. While

persistent bleaching can result in coral death due to the loss of autotrophic carbon contributed by

the symbionts [2], bleaching can also precipitate an acclimatory change in the composition of the

endosymbiont community that can subsequently increase coral thermal tolerance [3,4]. For coral

hosting multiple symbiont types, changes in their relative abundance, or 'shuffling', can facilitate

survival and recovery [3]. The ability for adult coral to acquire completely new symbiont types

following bleaching was also recently confirmed [5]. Although this capacity for flexibility is not

universal [6], and is influenced by environmental cues and physiological trade-offs [7], symbiont

shuffling and/or switching has been proposed as a key target for human-assisted evolution [8].

Successful establishment of novel symbioses that increase coral bleaching tolerance have been

demonstrated in laboratory conditions [9], however it is unclear how long these heterologous

relationships persist, especially in natural reef environments. In a key test of the ultimate utility

of this approach, we investigate the long-term stability of a heterologous coral symbiosis in the

context of reef restoration.

Acropora palmata were once one of the dominant habitat builders of shallow Caribbean

reefs [10]. Unprecedented demographic declines since the mid-1970's have resulted in extensive

propagation efforts in both land and ocean-based nursery systems to both maintain existing

genetic diversity and produce biomass for outplanting efforts aimed at restoring ecosystem

structure and function [11]. Naturally occurring populations of *A. palmata* are highly specific in

their symbiosis with *Symbiodinium fitti*, formerly ITS2 type A3, [12,13]. Single nucleotide

polymorphism (SNP) genotyping of *ex situ* nursery-reared lines of *A. palmata*, however,

revealed an almost exclusive association with *Durusdinium spp.* (Fig S1A, Supplementary

Methods). This provided a unique opportunity to test the maintenance of this heterologous

symbiotic relationship after outplanting back to the reef environment.

In April 2018, we transplanted three clonal replicates (ramets) of five exclusively *ex situ*

nursery-reared *A. palmata* genotypes to nine reefs in the lower Florida Keys (n=27 ramets per

genet) to evaluate the long-term stability of the *Durusdinium* association on natural reefs using

periodic phenotypic monitoring and ITS2 amplicon sequencing for symbiont typing using the

SymPortal pipeline [14], (Supplemental methods, Figure 1). A random sub-sampling of 10-12

ramets per genet confirmed that no *Symbiodinium fitti* were detectable prior to outplanting (T0,

Supplemental methods, Figure 2A). Four genets were exclusively dominated by a *Durusdinium*

ITS2 type, whereas genet 13-XK showed a consistent co-infection with a *Cladocopium* ITS2

type composed of three co-occurring C1 amplicon sequence variants (Figure 2A). A PCoA using

Bray-Curtis dissimilarity of T0 samples confirmed a significant effect of genotype ($p < 0.001$)

with 13-XK driving divergence (Figure 2B).

No bleaching was observed during quarterly site visits, but site-specific mortality was

evident, with 100% ramet mortality at Bahia Honda following 18 months (Supplemental

Methods, Table S2). *Durusdinium* remained the dominant ITS2 profile after two years of

outplanting (T24, Fig. 2B, C). Surviving co-infected 13-XK ramets also shuffled to *Durusdinium*

dominance across sites (Fig. 2C) and a PERMANOVA confirmed genotype was no longer a

significant factor ($p = 0.764$). Uptake of novel symbionts was detected, however, with site

specific patterns (PERMANOVA, $p < 0.031$, Fig. 2C). Acquisition of the homologous symbiont,

Symbiodinium fitti (A3) was observed at four sites, with ITS2 type profiles indicating potential

sub-species or strain-level differences between Dave's Ledge, Eastern Dry Rocks, Looe Key,

and Maryland Shoals (Fig. 2C). Two ramets of 13-X7 acquired *Cladocopium* types, including

one apparently novel strain (C66) although their relative abundance remained low (Figure 2C). A

Gerakladium (G) type profile was also detected in ramets of two different genets at two different

sites, also at low relative abundance (Fig. 2C).

The general long-term stability of the heterologous symbiosis between *A. palmata* and

Durusdinium spp. on natural reefs supports the potential effectiveness of symbiont modification

as a management intervention. Infection of coral larvae from mass spawning events with

thermally tolerant symbionts and outplanting them to degraded reefs could also be a restoration

tool [15]. However, the occasional acquisition of homologous, site-specific strains of *Symbiodinium*

fitti indicates that the temporal dynamics of symbiosis establishment in coral may be more

complicated than previously thought. While corals can take up symbionts in their larval phase,

the current consensus is that the majority of symbionts are taken up after settlement. New

recruits and juvenile corals can form a variety of novel symbiont associations [16], but this

diversity is generally pruned to one numerically dominant taxon per genus over the first few

years of life [17]. Subsequent changes to the dominant taxon occur in response to bleaching-level

stress [5,18,19].

The *A. palmata* lines used in this study were produced using individual larvae from batch

crosses in 2013/14. Although physically small, at the time of outplanting, experimental ramets

were 4-5 years old, which under normal field conditions is the age of sexual maturity for this

species [20]. The finding that adult corals can switch their symbionts *in situ* in the absence of

bleaching suggests that acquisition and winnowing may be size-dependent rather than age-

dependent process. It is also possible that Symbiodiniaceae communities are indeed capable of

greater flexibility under ambient conditions [21]. However, the stability of the heterologous

symbiosis in most individuals suggests that infection with a novel symbiont type as a recruit and

maintenance till maturity could produce a stable symbiosis.

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Code and Data Availability

Sequence Access: NCBI, Accession: PRJNA868513

https://github.com/hollandelder/OutplantedA.palmITS2

Authors' contributions

C.D.K., E.M., C.K, W.M., and H.E. conceived the experiments and analysis. W.M., E.B., C.K.,

and C.D.K. conducted experiments. E.M. and E.B. provided advice and logistical support

through the experiment. H.E. conducted genetic and statistical analysis and wrote the manuscript.

C.D.K. and E.M. provided advice and support for analysis. C.D.K., E.M. and C.K. helped write

the manuscript.

Conflict of Interest Statement

All authors approve the submission of this manuscript and declare no conflicts of interest.

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Figure 1. Experimental setup showing genotypes subsampled at time point 0 (T0), the locations of outplanting sites, the locations of water quality monitoring stations, and the genotype array strategy at each site. All surviving corals were sampled at time point 24 (T24).

Figure 2. Relative abundance and PCoA plots showing that symbiont community did not

significantly change after two years of outplanting. (**A.**) The relative abundance plot shows the

ITS2 symbiont profiles of each individual coral grouped by genotype that was subsampled at T0.

Each bar represents an individual and each is labeled with either their ex situ nursery frag label

or their outplant site ID. Color represents symbiont type. (**B.**) The relative abundance plot shows

the ITS2 symbiont profiles of each individual coral grouped by genotype that was sampled at

T24. Each bar represents an individual. They are grouped by site, then genotype, and then frag

id. Asterisks (*) identify individuals that were also sampled at T0. Color represents symbiont

type. (**C.**) A PcoA plot that shows that symbiont profiles did not significantly change after two

229   years of being outplanted. Color represents coral genotype and shape represents site. The ex situ

230   nursery and T0 is included as a site.