Genomic health is dependent on long-term population demographic history

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Abstract

Current genetic methods of population assessment in conservation biology have been challenged by genome-scale analyses due to their quantitatively novel insights. These analyses include assessments of runs-of-homozygosity (ROH), genomic evolutionary rate profiling (GERP), and mutational load. Here, we aim to elucidate the relationships between these measures using three divergent ungulates: the white-tailed deer, caribou, and mountain goat. The white-tailed deer is currently expanding, while caribou are in the midst of a significant decline. Mountain goats remain stable, having suffered a large historical bottleneck. We assessed genome-wide signatures of inbreeding using the inbreeding coefficient F and %ROH (F_{ROH}) and identified evolutionarily constrained regions with GERP. Mutational load was estimated by identifying mutations in highly constrained elements (CEs) and sorting intolerant from tolerant (SIFT) mutations. Our results show that F and F_{ROH} are higher in mountain goats than in caribou and white-tailed deer. Given the extended bottleneck and low N_e of the mountain goat, this supports the idea that the genome-wide effects of demographic change take time to accrue. Similarly, we found that mountain goats possess more highly constrained CEs which are indicative of greater purifying selection. This is reflected by fewer mutations in CEs and deleterious mutations identified by SIFT. In contrast, white-tailed deer presented the highest mutational load with both metrics, while caribou were intermediate. Our results demonstrate that extended bottlenecks may lead to reduced diversity and increased F_{ROH} in ungulates, but not necessarily the accumulation of deleterious alleles, likely due to the purging of deleterious alleles in small populations.

Introduction

Organizations like the International Union for Conservation of Nature (IUCN) rely primarily on population sizes and trends when assigning wildlife conservation statuses (IUCN Standards and Petitions Committee, 2019). The predictive relationship between population size (N_C) and genetic diversity is challenging, as N_C, for example, tends to be substantially more variable than diversity metrics like nucleotide diversity (π), referred to as Lewontin's paradox (Ellegren & Galtier, 2016; Lewontin, 1974), and therefore rarely accounts for recent demographic history and the genetic health of populations. While genetic diversity estimates generally correlate with IUCN status (Peart et al., 2020; Petit-Marty et al., 2021), the relationship (i.e., direction) is varied and predictive association is low (Schmidt et al., 2022). The unprecedented loss of global biodiversity (Pascual et al., 2021) and ambiguous associations (Schmidt et al., 2022) necessitates the need for conservation-informative genetic metrics.

Both novel genome metrics and functional versus presumed neutral variation have garnered attention in the conservation genetics community. The importance of neutral genetic diversity for example, despite a long history (e.g. DeWoody et al., 2021; Kohn et al., 2006), was recently called into question (Teixeira & Huber, 2021). Citing Lewontin (1974), Teixera & Huber (2021) argued for a renewed focus on functional variation and gene-environment interactions, which prompted two high-profile responses (Garcia-Dorado & Caballero, 2021; Kardos et al., 2021). Kardos et al. (2021) in particular echoed earlier concerns of [functional] gene-targeted conservation (i.e. Kardos & Shafer, 2018), and highlighted some potential problems in Teixeira & Huber's simulations. Kardos et al. (2021) argued against abandoning genome-wide neutral markers; supporting this view would be empirical work from Peart et al. (2020) that showed genome wide estimates of Tajima's *D*, a composite statistic derived from

Watterson's θ and π (Tajima, 1989), appears predictive of conservation status, further reiterating the value of classic neutral summary statistics derived from genome-wide sequences.

The high correlation between nucleotide diversity and microsatellite heterozygosity suggests that genome-scale data reflect smaller targeted panels of loci (Väli et al., 2008). This observation is ultimately what formed the premise of Shafer et al. (2015) that argued for the use of quantitatively novel genomic metrics for conservation. Runs-of-homozygosity (ROH), genomic evolutionary rate profiling (GERP), and mutational load are metrics dependent on genome-scale data and have only recently been applied to species and populations of concern (Grossen et al., 2020; van der Valk et al., 2019; von Seth et al., 2021). ROHs are long stretches of homozygosity that arise as a result of inbreeding (Ceballos et al., 2018), and reflect the true inbreeding coefficient and can identify genes underlying inbreeding depression (Kardos et al., 2016). GERP profiling can be used to identify evolutionarily constrained genomic elements, defined as regions with fewer substitutions than expected under neutral evolution (Davydov et al., 2010), which are often the subject of selective pressures (Cooper et al., 2005; Davydov et al., 2010). Any mutations that appear at highly conserved sites are generally assumed to be deleterious (Cooper et al., 2005; Davydov et al., 2010). Likewise, tolerant and intolerant mutations – also referred to as SIFT scores (Vaser et al., 2015) – that are considered of human clinical interest (P. Kumar et al., 2009) – have now been applied to natural populations (Robinson et al., 2016). ROH, GERP and SIFT scores collectively allow for inferring the mutational load and genomic health of a population or species.

While quantitatively useful, the relationships between these metrics and conservation status and diversity metrics are generally not well defined and at-times counter to expectation. van der Valk et al. (2019) showed a weak relationship between conservation status and

mutational load across mammalian taxa; the highest mutational loads were in fact seen in the most abundant and diverse species. Similarly, species with the highest inbreeding (ROH) also had the lowest mutational load and GERP scores (van der Valk et al., 2019). This pattern could be due the purging of deleterious alleles in small populations, which is consistent with simulations by Kyriazis et al. (2021). van der Valk et al. (2019) argued that this process has implications for genetic rescue and suggested that individuals be selected for translocation based on low mutational load and not diversity. Work on the Sumatran rhinoceros (*Dicerorhinus sumatrensis*) added another layer of nuance, with populations showing low to moderate levels of inbreeding (ROH), but a relatively high mutational load (von Seth et al., 2021). In contrast, the Alpine ibex (*Capra ibex*) has experienced successive demographic bottlenecks that led to reduced diversity and increased ROH, but not the accumulation of highly deleterious alleles (Grossen et al., 2020). Thus, there seems to be a link between historical demography, ROH, and mutational load. Understanding such processes has important implications for both predicting long-term viability, but also selecting animals for genetic rescue.

Here, we analysed signatures of genomic health across populations of three divergent ungulates to elucidate these relationships. The caribou (*Rangifer tarandus*) is currently experiencing significant population declines (N_C) prompting international concern (COSEWIC, 2015). In Quebec, five of the major populations have seen drastic declines from their highest estimates ranging between 10% and 99%, depending on the herd (COSEWIC, 2015, 2017). The white-tailed deer (*Odocoileus virginianus*), in contrast, is expanding northward and has accordingly high genomic diversity (Kessler et al., 2022). While white-tailed deer currently number in the 10s of millions (Kauffman et al., 2018), they were historically extirpated across much their range due to colonial harvest and forestry practices (Kauffman et al., 2018); meaning

this demographic expansion is very recent. The mountain goat (*Oreamnos americanus*) underwent a massive bottleneck during the last glacial maximum (Martchenko et al., 2020) but has since remained relatively stable with a range-wide population of ~100,000 individuals (Festa-Bianchet & Côté, 2008). These three species, contrasted in their ecology but also conservation status, historical and demographic trajectories, present useful comparisons to explore patterns of novel genomic metrics as they pertain to conservation status. In line with recent empirical work, we predicted that populations with low diversity and high inbreeding do not necessarily have higher mutational load and that genome-wide effects of demographic change like ROH take time to accrue. Similarly, we expected to see greater purging of deleterious alleles in populations with long-term small N_e. Collectively, this means that poor genomic metrics of health would not correlate with current negative population trends.

Methods

Caribou samples were obtained from 67 individuals across Quebec and Labrador (Dedato et al. 2022); 53 white-tailed deer were sampled across their North American range (Kessler et al. 2022). Additionally, 35 mountain goats were sampled across their native range from Alaska to as far south as Idaho (Martchenko et al. 2022; Figure 1). All samples were sequenced with Illumina HiSeqX technology to approximately 5X coverage. Non-target sequences were trimmed from raw FASTQ files with Trimmomatic v.0.36 (Bolger et al., 2014), and reads were mapped to their reference genome using BWA-MEM v.0.7.17 and SAMtools v.1.15 (Li et al., 2009; Li & Durbin, 2009). Duplicate mapped reads were marked and removed using Picard v.2.23.0 (Broad Institute, 2019). Uniquely mapped reads were subset by Sambamba v.0.8.2 (Streit et al., 2012), and indels were realigned with GATK v.3.8 (Van der Auwera et al., 2013). We estimated genotype likelihoods using ANGSD v.0.934 using the GATK model: -doVcf 1 -doGeno -4 -

doPost 1 -gl 2 -SNP_pval 1e-6 -minMapQ 20 -minQ 20 -doCounts 1 -skipTriallelic 1 -doGlf 2 (Korneliussen et al., 2014). Hard calls required for analyses were performed by Beagle v.5.1 (Browning et al., 2018) with the following parameters: impute=false overlap=3000 window=100000 gprobs=TRUE parameters. Posterior genotype probabilities were estimated using allele frequencies, and only genotypes with a posterior probability above 0.95 were called. Tajima's *D* and nucleotide diversity (π) were calculated using VCFtools v.0.1.13 with windows of 10 Kb and using all positions, regardless of allele frequency (Danecek et al., 2011).

Inbreeding and Homozygosity

A subset of SNPs in approximate linkage equilibrium was obtained using PLINK v.1.9 with a window size of 50 SNPs, a window shift size of 10 SNPs, and a pairwise (R^2) linkage disequilibrium threshold of 0.1 (Chang et al., 2015; Purcell et al., 2007). We then identified runs of homozygosity (ROH) on scaffolds greater than 10 Mb in length and with a minor allele frequency cut-off of 0.05 (Foote et al., 2021). Sliding windows of 50 SNPs were used, and each ROH was defined as a sequence at least 100 Kb long with at least 50 homozygous SNPs (Foote et al., 2021). Each ROH required a minimum density of 1 SNP per 50 Kb and no more than 5 missing or heterozygous calls in a window. Any SNPs more than 1 Mb apart within a segment were used to split that segment in two. Once identified, each ROH was classified by size, and the percent ROH (F_{ROH}) for each scaffold and individual were calculated. PLINK parameters were taken from Foote et al. (2021) with output validated using the program ROHan (Renaud et al., 2019). ROHan was run on the same data with the same ROH size definition of 100 Kb and a transition to transversion ratio (Ts/Tv) of 2.25. This ratio was calculated using BCFtools v.1.14 and the genotypes provided by ANGSD (Li et al., 2009). The inbreeding coefficient (*F*) for each individual was also estimated using PLINK and the –het flag.

Evolutionary Constraint and Mutational Load

Twenty ungulate reference genomes from the NCBI GenBank genome database (Table S1) were selected to represent most extant families with genomes available in both the orders of Perissodactyla (odd-toed ungulates) and Artiodactyla (even-toed ungulates). Constrained regions for each individual of our three focal species were identified using the Genomic Evolutionary Rate Profiling (GERP++); this approach requires a phylogenetic tree which we constructed for all 23 species using TimeTree (Figure S1) (Kumar et al., 2017). The individual fastq files of all individuals of the three species were then aligned to each of the 20 reference genomes in 50 bp segments using BWA-MEM. All duplicate alignments were marked and removed with Picard. Then, the alignments of each individual were concatenated into one multiple alignment file (MAF), resulting in 155 total MAFs. The gerpcol function included in GERP++ was run on each MAF to generate rejected substitution (RS) scores for each alignment column. RS scores are used as a metric for constraint intensity and are calculated by subtracting the estimated evolutionary rate at each position from the neutral rate (Cooper et al., 2005; Davydov et al., 2010). Both rates were estimated by gerpcol using the aforementioned phylogenetic tree (Figure 2). RS scores quantify the number of differences between the expected and observed number of SNPs at a site, and a positive score is indicative of constraint (Cooper et al., 2005; Davydov et al., 2010). An RS score greater than 2 represents a highly constrained region whereas a score lower than 1 is putatively neutral and discarded (Cooper et al., 2005; Davydov et al., 2010). gerpcol was run with the previously calculated Ts/Tv ratio of 2.25. We also used GERP++ to identify constrained elements with the gerpelem function.

To estimate mutational load, we determined the number of SNPs within these constrained elements using BEDtools v.2.30.0 (Quinlan & Hall, 2010). These SNPs are putatively deleterious due to the common functional importance of constrained regions (Cooper et al., 2005; Davydov et al., 2010). For each species, the functional consequence of each SNP was determined using Ensembl's Variant Effect Predictor (VEP) (McLaren et al., 2016). The deleteriousness of each SNP across the genome was then estimated using functional tolerance scores provided by SIFT and SIFT4G (Vaser et al., 2015). Using multiple alignment information, SIFT and SIFT4G calculate normalised probabilities where substitutions with scores lower than 0.05 are predicted to be deleterious to protein function (Vaser et al., 2015).

Caribou did not have an available annotation at the time of analysis which is required for this analysis; thus, we used a mixture of *de novo* and homology-based predictions. We aligned protein sequences from *Bos taurus, Equus caballus, Homo sapiens,* and *Ovis aries* to the caribou genome using blastx v.2.7.1 (Altschul et al., 1990). These sequences were all obtained from Ensembl release 89 (Hunt et al., 2018). We also aligned transcripts from the NCBI mammalian RefSeq transcipt database v.1.0 using BLAT v.1.0 to help identify exons. Caribou RNAseq data (SRX12486288) from the NCBI Sequence Read Archive were aligned to the genome with HISAT v.2.10 for *de novo* prediction (Kim et al., 2015). Using these data, genes were predicted by Augustus v.3.1.1 as well as GeneMarkES v.4.69 (Lomsadze et al., 2005; Stanke et al., 2006). We generated a set of consensus genes using EVidenceModeler v.1.1.1 with the following weighting scheme (Haas et al., 2008): transcript alignment via BLAT (10x weight); protein alignment via blastx (5x weight); gene prediction via Augustus (1x weight); gene prediction via GeneMarkES (1x weight). PASA v.2.3.3 was used last to refine gene identifications (Haas et al., 2008).

For all individual metrics above we quantified the correlation to sample location (latitude, longitude). Because white-tailed deer were extirpated across much of their range, we used translocation data from McDonald & Miller, (2004) to determine if these metrics were correlated to restocking efforts in the United States. We examined correlations with stock start and end dates, estimated population lows, number of individuals stocked, and the number of states involved in stocking.

Results

Diversity and Inbreeding

Nucleotide diversity (π) was found to be highest in deer (mean = 0.00771) and lowest in mountain goats (mean = 0.00101) (Figure 1). Tajima's *D* was highest in mountain goats (mean = 0.625) and lowest in white-tailed deer (mean = -0.645) (Figure 1). For caribou, Tajima's D was positive (mean = 0.124), but lower than in mountain goats (t-test p-value < 2.2×10^{-16}). Mountain goats were found to have significantly higher F_{ROH} (mean = 0.038) than both caribou (none detected) and white-tailed deer (mean = 0.0020) (Figure 2a). This pattern is reflected in the relative values of *F* where mountain goats had the highest value (mean = -0.018) in comparison to caribou (mean = -0.44) and deer (mean = -0.27) (Figure 2b). For the mountain goat, most ROHs were found to be in the 500-1000 Kb range whereas deer possessed ROHs almost exclusively below 300 Kb (Figure 3). Despite a much higher number of ROHs overall, mountain goats possessed significantly fewer in the 100-300 Kb range than deer (Figure 3). Both measures of inbreeding were found to be correlated in mountain goats (Pearson's $r = 0.840$, $p < 0.05$) and deer (Pearson's $r = 0.395$, $p < 0.05$) (Figure S2), noting again there were no F_{ROH} in caribou preventing a similar assessment.

Evolutionary Constraint and Mutational Load

Mountain goats had the highest number of constrained elements (CEs) (mean = 2926) while caribou have the fewest (mean = 935; Figure S3). Highly constrained elements followed a nearly identical distribution between species with all three having approximately 50% as many CEs with RS > 2 (Figure S3). In addition to having the most CEs, mountain goats showed slightly longer CEs on average (Figure S4). The three species showed little variability in average CE RS-score (Figure S4, S5). Despite having significantly fewer CEs, white-tailed deer had a larger number of SNPs in CEs (mean = 411) followed by mountain goats (mean = 212) and caribou (mean = 74) (Table 1). The relative differences between species remained the same for highly constrained CEs (Table 1). We observed that a much higher fraction of constrained positions was deleterious in deer (mean = 0.625%) than in caribou (mean = 0.326%) and mountain goats (mean = 0.121%; Figure 4).

We found 42.1% of coding sequence consequences identified by VEP to be missense variants in white-tailed deer, 49.3% in caribou, and 57.4% in mountain goats (Figure 5a). Despite this distribution, white-tailed deer had the highest number of intolerant mutations (mean = 6852) identified by SIFT followed by caribou (mean = 4944) and mountain goats (mean = 3190) (Table 1). Similarly, we observed white-tailed deer to have the highest fraction of genes with deleterious mutations (mean = 0.330%) while caribou (mean = 0.149%) and mountain goats (mean = 0.143%) had the lowest (Figure 5b). None of these metrics were correlated to the number of genes in the annotation (Table 1). Lastly, we found F and latitude to be moderately correlated in caribou (Pearson's $r = -0.466$, $p < 0.05$; Figure S6) and deer (Pearson's $r = -0.513$, $p < 0.05$; Figure S6), but not in mountain goats (Pearson's $r = 0.003$, $p < 0.05$). There were no

significant correlations between the individual genome health metrics and restocking data in white-tailed deer (Pearson's $r < 0.4$, $p > 0.05$; Table S2).

Discussion

The interplay between nucleotide diversity (π), inbreeding, mutational load, and census size (N_C), is nuanced and full of contrasting examples. Here, we demonstrate that mutational load may be lower than expected in populations with relatively high inbreeding and low diversity. Our observed patterns suggest that the purging of deleterious alleles in these cases is stronger in small populations (Huber et al., 2020), and therefore would reduce risks from inbreeding depression. This supports our hypotheses that low diversity and high inbreeding do not on their own necessitate special conservation status, and that purging is likely occurring in populations with low N_e. We also predicted that genomic signatures of demographic events take time to accrue, which is supported by evidence of F_{ROH} and CEs requiring extended bottlenecks to surface and increase. Caribou have seen recent decreases in N_C yet have no detectable ROH; in contrast, mountain goats show elevated ROH likely due to the ancestral nature of their bottleneck (Martchenko et al., 2020). The varied relationships between different measures of genomic health and the lack of correlation to the population trends that are typically used for status assignment highlights the value of this comparative framework (see also Huber et al., 2020). This information is especially valuable for small populations of conservation concern, where mutational load combined with ROH and CE assessments is probably the most informative way forward, in addition to metrics like Tajima's *D* (Peart et al., 2020).

Inbreeding Effects

Genomic consequences stemming from demographic take time to accumulate (e.g. Grossen et al., 2020; von Seth et al., 2021). Several caribou populations are currently in the midst of a demographic bottleneck resulting in an extremely low contemporary effective population size (N_e) and nucleotide diversity (Dedato et al., 2022). In contrast, mountain goats underwent a large historical bottleneck during the last glacial maximum, and while they have not recovered, they have remained relatively stable, resulting in a moderately low N_e (Martchenko & Shafer, 2022). Low diversity stemming from bottleneck is supported by positive values of Tajima's *D* in these two species, which indicate a lack of rare alleles and population contraction (Tajima, 1989). The lower π in mountain goats is clearly due to their extended bottleneck that would have exacerbated any effects of inbreeding, but also their historically lower census sizes. These effects are reflected by F_{ROH}, where mountain goats were found to have 3.8% of the genome in ROH whereas caribou had virtually no detectable ROH segments over 100 Kb long. Here, despite a less drastic decline in N_C, mountain goats have accumulated a significantly greater F_{ROH} than caribou. These differences are also unlikely to be the result of genome assembly quality, as the mean number and length of analysed scaffolds were comparable (Table 1). Instead, it is likely that any stretches of homozygosity in caribou were too short to be classified as ROHs by our methods (<100 Kb). This is a common finding in mammalian genomes, where most species have no ROHs above this threshold and possess a low ROH burden (Brüniche-Olsen et al., 2018).

Compared to white-tailed deer, mountain goats have developed significantly longer and more numerous ROHs, which further outline the effects of a prolonged bottleneck. Caribou simply have not been subject to a low N_C for long enough to approach mountain goats or even differentiate themselves from deer in their levels of inbreeding. Of note, F_{ROH} was more correlated to *F* after an extended bottleneck (i.e., mountain goats vs. white-tailed deer; Figure

S2), which further implies that ROHs require both time and demographic change (Gillespie, 2004) to accumulate. Overall, these results support the idea that genome-wide effects of demographic change will not accrue unless given enough time; F and F_{ROH} are therefore unlikely to align with IUCN conservation decisions, similarly noted by Schmidt et al. (2022).

Mutational Load and Purging

The relationship between N_e, inbreeding, π, and mutational load are variable (Grossen et al., 2020; van der Valk et al., 2019; von Seth et al., 2021). Empirical studies on mammals have led us to predict that low diversity and high inbreeding do not necessarily result in the accumulation of deleterious alleles (Grossen et al., 2020; Robinson et al., 2018, 2018; van der Valk et al., 2019). Here, we found a much greater number of putatively deleterious mutations in white-tailed deer than in mountain goats and caribou by identifying CE-bound SNPs with GERP. These results were confirmed by the deleterious alleles identified by SIFT that showed a similar gap between white-tailed deer and the other species, regardless of the annotation quality (Table 1). This means that despite lower diversity and small N_e, caribou and mountain goats have relatively low mutational loads; further, despite a relatively low number of CEs, deer still possessed significantly more CE-bound SNPs indicative of deleterious mutations. Mountain goats, in contrast, were found to have the highest amounts of inbreeding but still not accumulated many deleterious alleles. This reiterates the finding that inbreeding does not necessarily result in high mutational load; in the case of caribou, if N_C can return (e.g., Peart et al. 2020) or if purging of deleterious alleles can occur (e.g., Huber et al., 2020), the genetic cost to the current decline in N_C could be minimized. This, however, remains an open question and wolves in Europe provide an informative example (Smeds & Ellegren, 2022).

The purging of deleterious alleles in small populations is rarely observed (Hedrick & Garcia-Dorado, 2016; Robinson et al., 2018; Xue et al., 2015), but there is evidence that it may occur in the presence of severe bottlenecks (Grossen et al., 2020). Mountain goats were shown to have low mutational load in conjunction with generally low diversity, while caribou present intermediate profiles. This would be consistent with N_C numbers pre-20th century for caribou, that were presumably much higher (Festa-Bianchet et al., 2011). Reductions in N_e from severe bottlenecks can increase the fitness effects of recessive deleterious alleles by increasing homozygosity (Garcia-Dorado, 2012, 2015); this strengthens the pressure of negative selection and can ultimately lower the mutational load of a population (Garcia-Dorado, 2012, 2015; Glémin, 2003). However, this effect can be insufficient for overcoming cases of extensive homozygosity. Sumatran rhinos for example have a very similar historical N_e to that of mountain goats (von Seth et al., 2021), but considerably more ROH. While purging might have played some role in some populations (von Seth et al., 2021), the overall negative effect of autozygosity and continued presence of high frequency deleterious alleles (e.g., Khan et al., 2021) was likely insurmountable from a population viability standpoint.

Purifying selection should remove SNPs in functionally important regions and constrain these areas against change, which is why we observed less SNPs than expected in CEs (Cooper et al., 2005; Davydov et al., 2010). The mountain goat's longer and more numerous CEs relative to the other species are possibly indicative of stronger purifying selection, the driving force behind constraint. In contrast, caribou have a smaller N_e than white-tailed deer; thus purging has either yet to take place or selection is less efficient, or a combination of both. The trajectory of caribou, would be predicted to be towards the patterns of mountain goat based on mammalian wide patterns and profiles (Cooper et al., 2005; Huber et al., 2020). It is also important to note

that purifying selection reduces nucleotide diversity by eliminating sites under direct selection and those linked through background selection (Charlesworth, 1996; Cvijović et al., 2018), which partially explains patterns of diversity seen here (Figure 1; Buffalo, 2021).

This work has implications for conservation decisions and the expected genomic consequences of numerical population declines. While deleterious alleles may be purged in small, inbred populations, high levels of autozygosity can render this benefit insignificant. Thus, we posit that high mutational load must be accompanied by high ROH to be of conservation concern, similar to what was observed in woolly mammoths (Rogers & Slatkin, 2017). Further, genetic rescue decisions should not rely solely on inbreeding and diversity metrics, especially when their relationships to fitness are not straightforward (Schmidt et al., 2022). This is also true for assessing restocking efforts, as we demonstrate that translocations are not necessarily related to population health metrics. Collectively, this provides further evidence of the time needed for genome-scale changes to accrue after significant changes in demography. Therefore, we recommend that genomic health metrics like F_{ROH}, mutational load, and evolutionary constraint be examined alongside classic metrics of population viability for a holistic view that can provide more accurate assessments of genome health.

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Data Availability and Benefit Sharing

All genome assemblies can be found on GenBank (details in Appendix Table S1). Re-sequencing data has been submitted to SRA and accession numbers are pending. Bash and R scripts for all analyses are available on GitLab: https://gitlab.com/WiDGeT_TrentU/undergrad-theses/-/tree/master/Wootton_2022

Benefits Generated: This research addresses important questions about the genomic health of populations of priority concern and illustrates relationships between genomic health and long-term demographic history. Collaborations were formed with researchers across North America, and all major collaborators are included as co-authors. All contributors are listed in the Acknowledgements. We are committed to institutional capacity-building and have shared the results of our research with our collaborators and the broader scientific community.

Author Contributions

ABAS and EW designed the research project. EW conducted all analyses. ABAS and EW wrote the manuscript using comments and contributions from all coauthors.

Tables and Figures

Table 1: The number of scaffolds > 10 Mb, mean scaffold length > 10 Mb, number of genes, and average number of deleterious mutations per species identified by GERP and SIFT. The number of genes was taken from the annotation of each species.

Species	Scaffolds > 10 Mb (n)	Mean Scaffold Length > 10 Mb	Genes (n)	SNPs in CEs (GERP)	SNPs in CEs with RS > 2 (GERP)	Deleterious mutations (SIFT)
Caribou	68	19,723,800	33,242	74	32	4,681
Mountain Goat	49	49,667,287	22,292	212	74	3,317
White-Tailed Deer	86	21,282,945	20,750	411	187	6,502

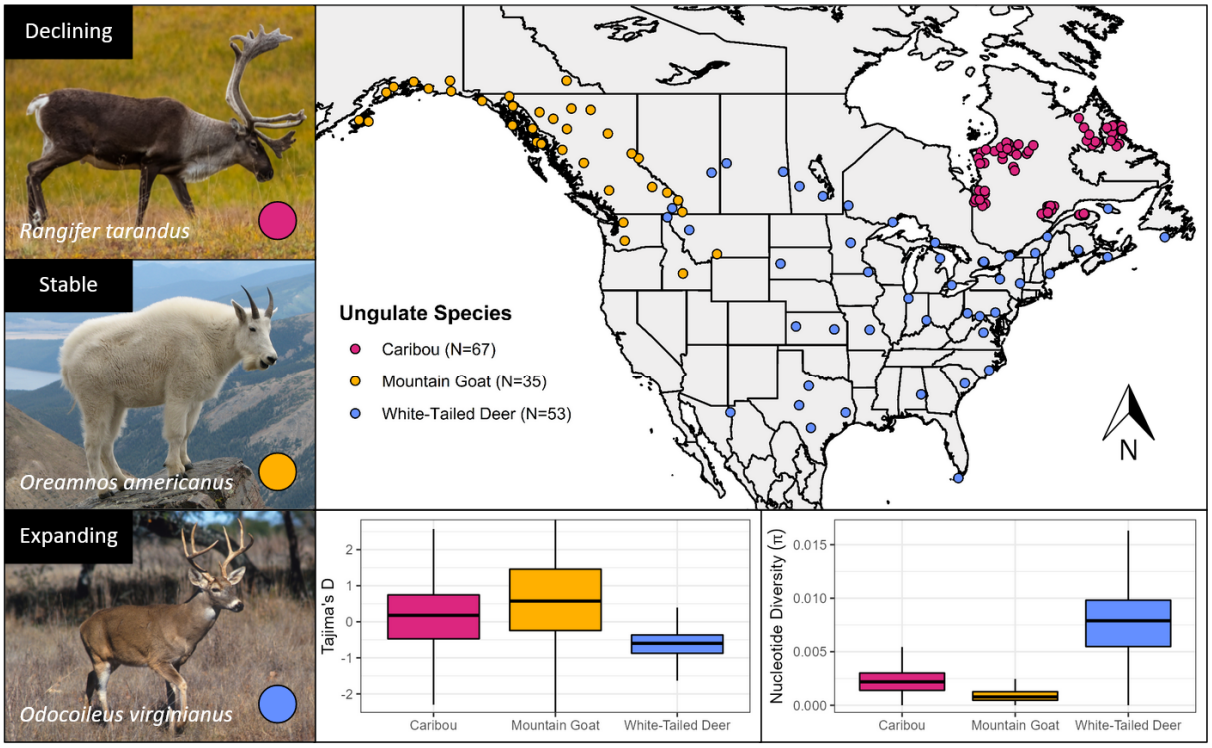

Figure 1: Sample locations and images of each species. (Red) *Rangifer tarandus* (N=67), (Yellow) *Oreamnos americanus* (N=35), (Blue) *Odocoileus virginianus* (N=53). Current demographic trends are outlined in the top left corner of each image. Boxplots showing Tajima's D and nucleotide diversity (π) per species are also pictured.

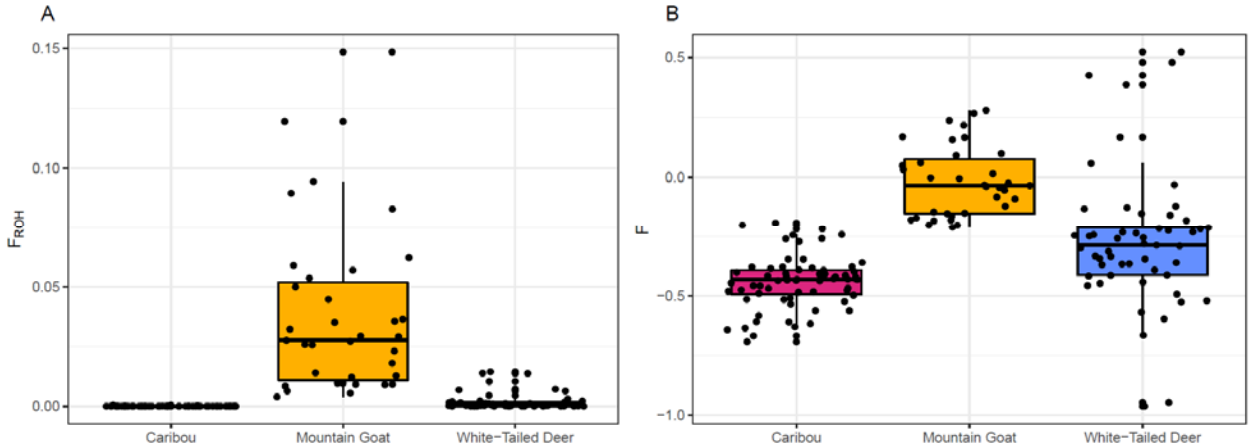

Figure 2: (A) Boxplot showing F_{ROH} as calculated by PLINK for all three species. Only ROHs greater than 100 Kb are included in each estimate. No ROHs were detected in caribou. (B) Boxplot showing the method-of-moments inbreeding coefficient (*F*) for each species as calculated by PLINK.

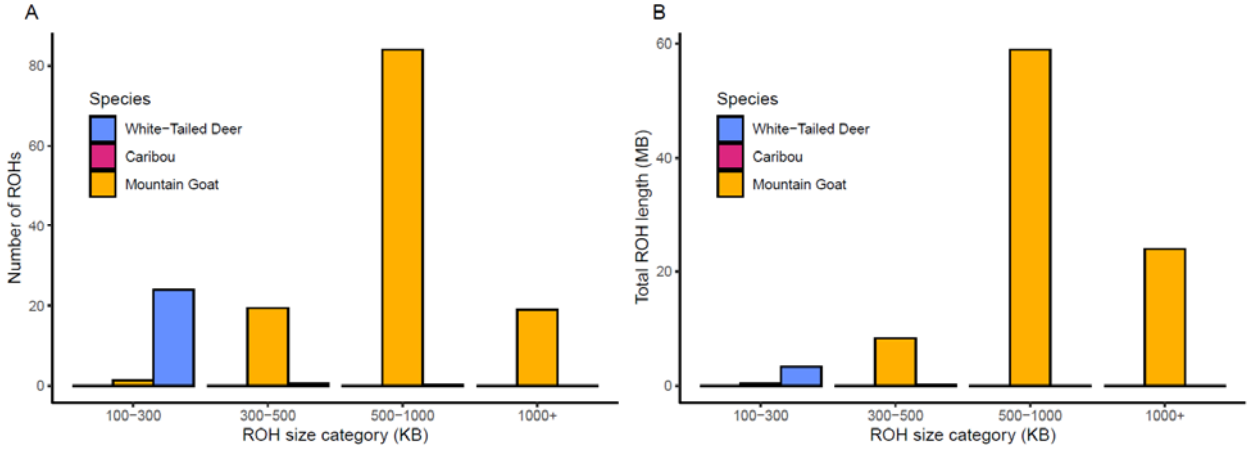

Figure 3: (A) Bar plot showing the total number of ROHs in each species stratified by ROH size category in Kb. No ROHs were detected in caribou. (B) Bar plot showing the total length (Mb) of all ROHs in each size category (Kb). No ROHs were detected in caribou.

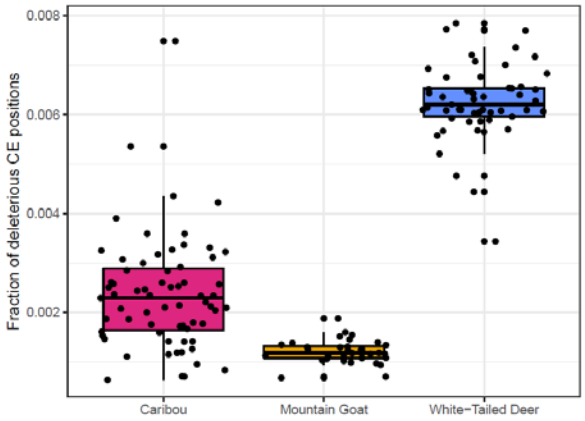

Figure 4: Boxplot showing the fraction of constrained positions with SNPs by species. Such SNPs are putatively deleterious, making this the fraction of positions that are deleterious in constrained elements (CEs).

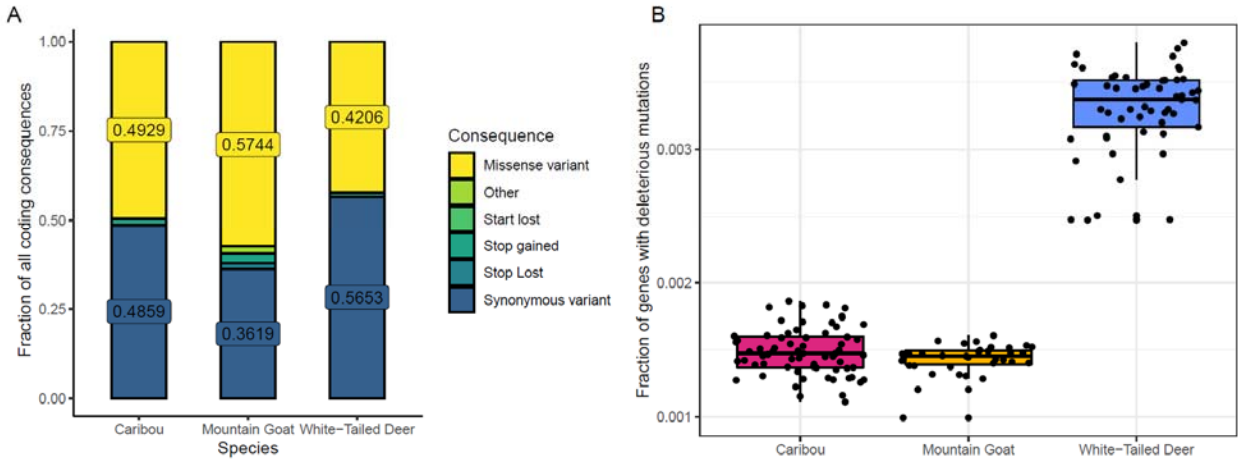

Figure 5: (A) Bar plot showing the fractions of each coding consequence identified by VEP in each species. (B) Boxplot showing the fraction of genes that possess deleterious mutations identified by SIFT.

Supplemental Information

Table S1: GenBank assembly accession numbers for each species used by GERP.

Species	**GenBank Assembly Accession**
Equus caballus	GCA_002863925.1
Equus asinus	GCA_016077325.2
Tapirus terrestris	GCA_004025025.1
Diceros bicornis	GCA_004027315.2
Ceratotherium simum	GCA_000283155.1
Camelus dromedarius	GCA_000803125.3
Camelus bactrianus	GCA_000767855.1
Hippopotamus amphibius	GCA_004027065.2
Tragulus javanicus	GCA_004024965.2
Moschus moschiferus	GCA_004024705.2
Moschus chryogaster	GCA_006461725.1
Bos taurus	GCA_002263795.2

Connochaetes taurinus	GCA_006408615.1
Ovis canadensis	GCA_004026945.1
Giraffa camelopardalis	GCA_017591445.1
Antilocapra americana	GCA_007570785.1
Alces alces	GCA_007570765.1
Cervus elaphus	GCA_910594005.1
Phacochoerus africanus	GCA_016906955.1
Sus scrofa	GCA_000003025.6
Rangifer tarandus	GCA_019903745.1
Oreamnos americanus	GCA_009758055.1
Odocoileus virginianus	GCA_014726795.1

Table S2: Correlation matrix between genomic health metrics and translocation data for white-tailed deer.

	F_{ROH}	F	**Fraction of deleterious CEs**	**Fraction of deleterious genes**
Stock start	0.032	0.127	0.076	-0.225
Stock end	0.372	0.392	0.205	0.347
Population low	-0.182	-0.059	-0.177	0.308
Number stocked	-0.102	-0.062	0.162	-0.103
Number of states	-0.161	-0.164	-0.093	0.096

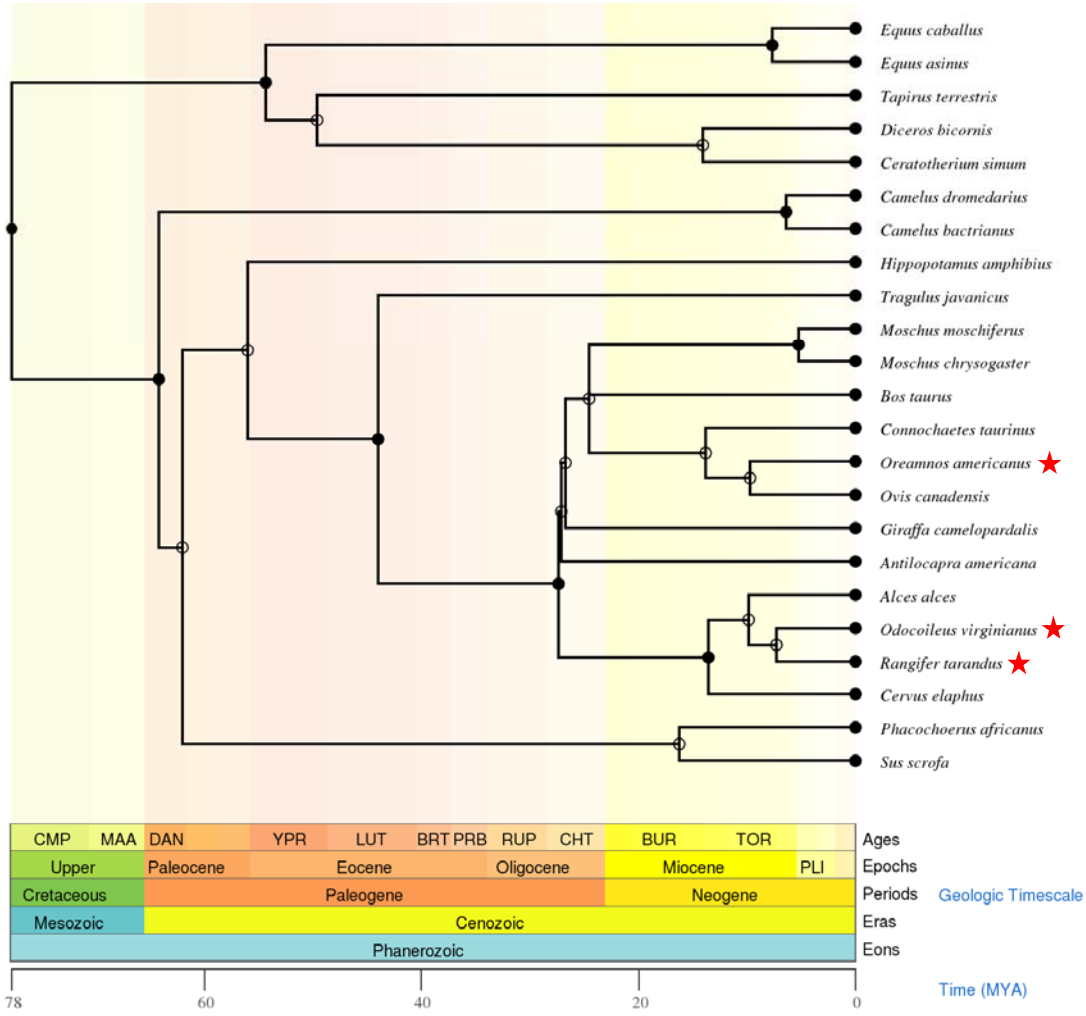

Figure S1: Phylogenetic tree generated by TimeTree showing all 23 ungulate species analysed by GERP++ and estimated times of divergence in MYA (Kumar et al., 2017). Study species with sample data are starred in red (*Odocoileus virginianus*, *Rangifer tarandus*, and *Oreamnos americanus*).

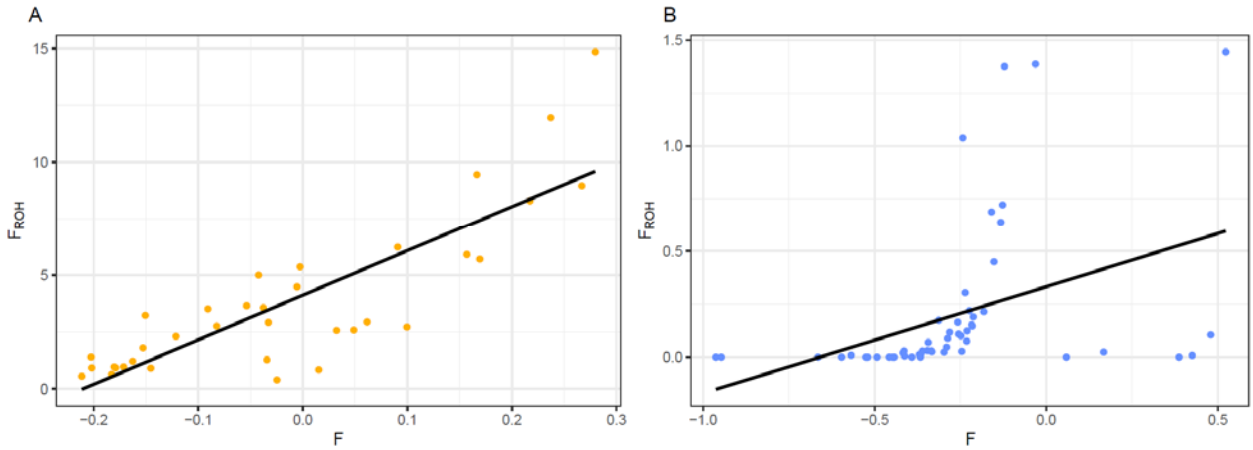

Figure S2: (A) Scatterplot and linear regression line showing the correlation between F and F_{ROH} in mountain goats (Pearson's r = 0.840, $p < 0.05$). (B) Scatterplot and linear regression line showing the correlation between F and F_{ROH} in white-tailed deer (Pearson's r = 0.395, $p < 0.05$).

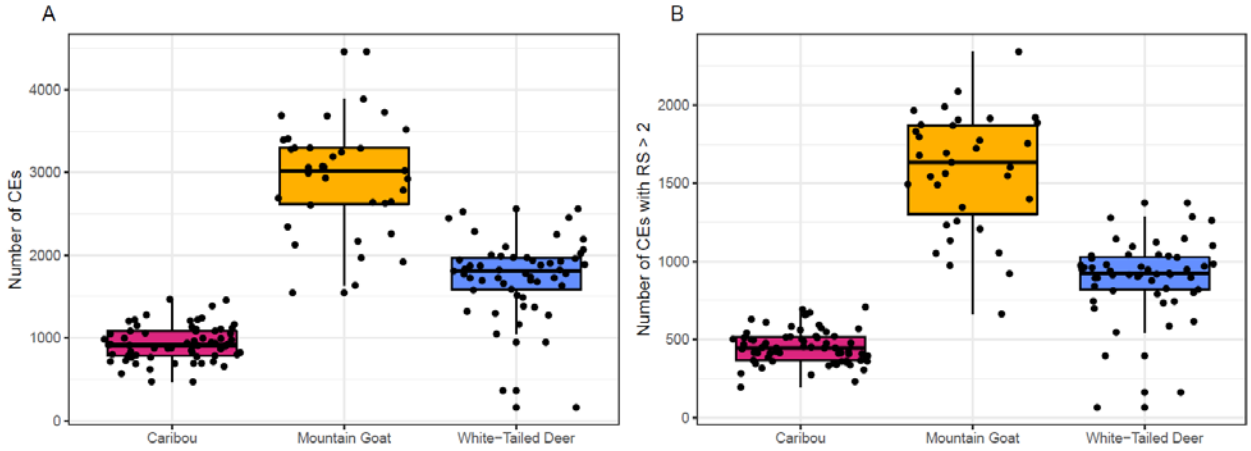

Figure S3: (A) Boxplot showing the number of constrained elements (CEs) identified per species by GERP++'s gerpelem function. (B) Boxplot showing the number of highly constrained CEs (RS > 2) identified in each species by gerpelem.

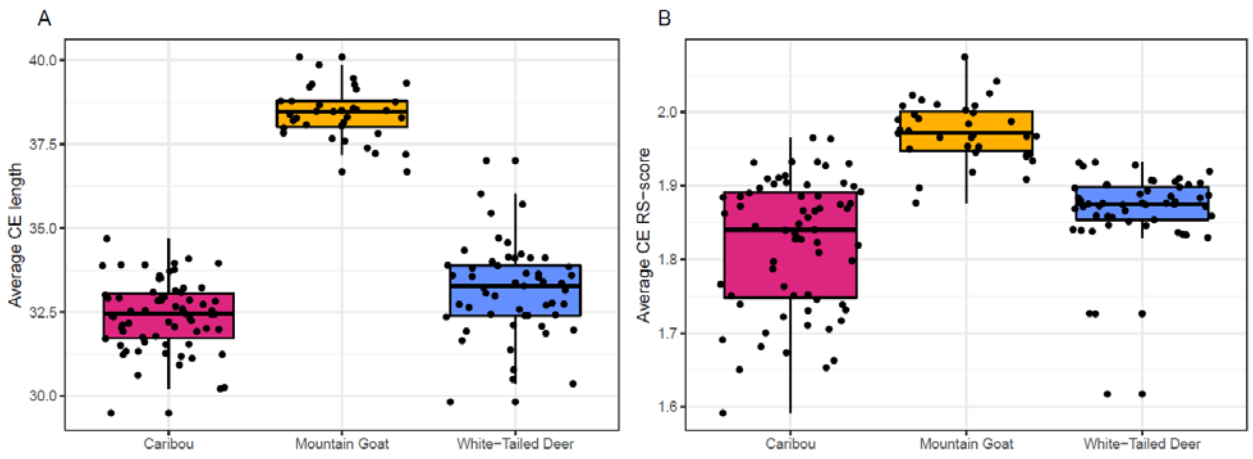

Figure S4: (A) Boxplot showing the average length of constrained elements (CEs) identified per species. (B) Boxplot showing the average intensity of constraint (RS-score) of CEs identified in each species.

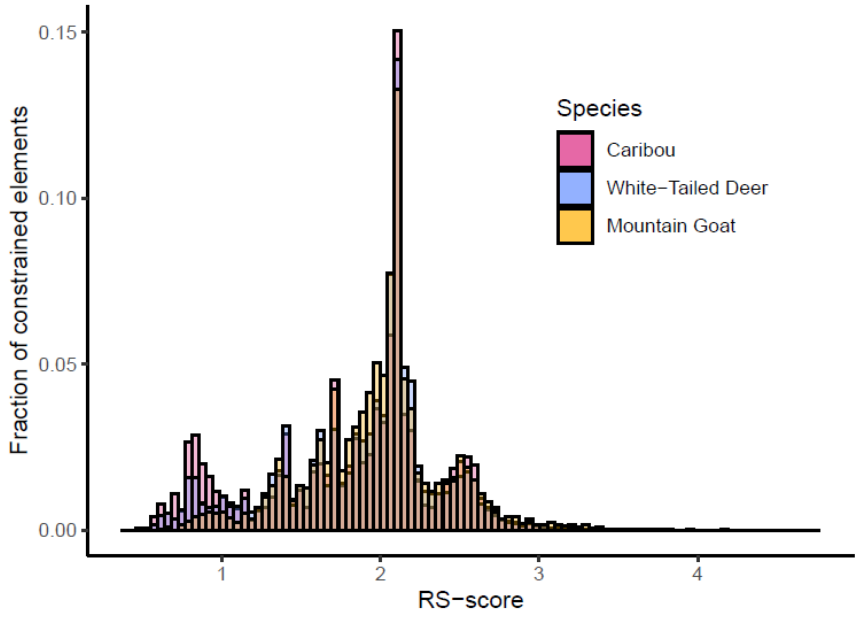

Figure S5: Histogram showing the fractional distribution of constrained element (CE) RS-scores.

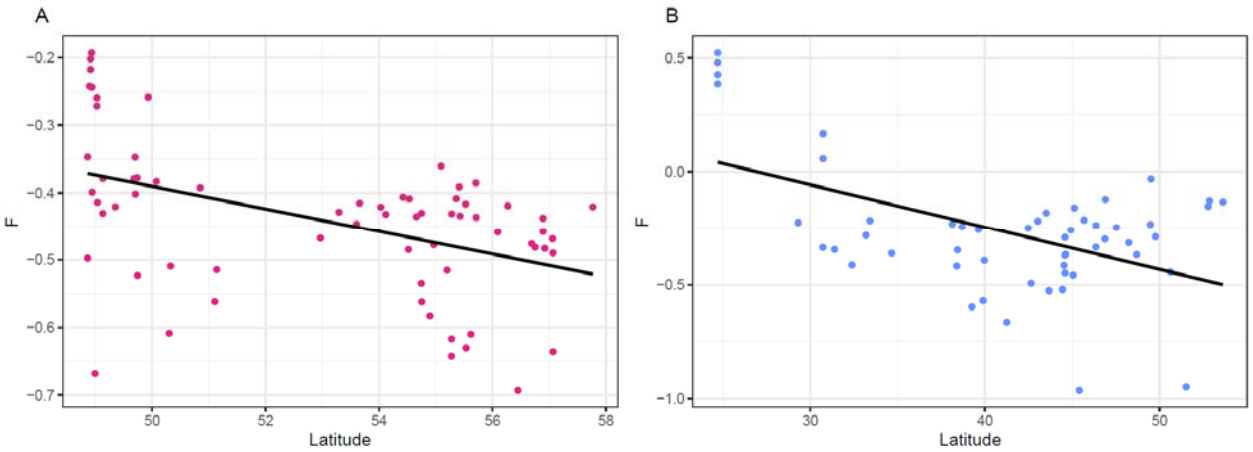

Figure S6: (A) Scatterplot and linear regression line showing the correlation between *F* and latitude in caribou (Pearson's r = -0.466, $p < 0.05$). (B) Scatterplot and linear regression line showing the correlation between *F* and latitude in white-tailed deer (Pearson's r = -0.513, $p < 0.05$).