Diffusive lensing as a mechanism of intracellular transport and compartmentalization

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Abstract

While inhomogeneous viscosity has been identified as a ubiquitous feature of the cellular interior, its implications for particle mobility and concentration at different length scales have remained unexplored. In this work, we use agent-based simulations of diffusion to investigate how diverse manifestations of heterogenous viscosity affect movement and concentration of diffusing particles. We propose that a mode of membraneless compartmentalization arising from the convergence of diffusive trajectories into viscous sinks, which we call "diffusive lensing," can occur in a wide parameter space and is thus likely to be ubiquitous in living systems. Our work highlights the phenomenon of diffusive lensing as a potentially key driver of mesoscale dynamics in the cytoplasm, with possible far-reaching implications for biochemical processes.

Introduction

Diffusion is a fundamental phenomenon of transport at scales ranging from atoms to galaxies. In cells, diffusion of individual components occurs in a complex, crowded milieu (Ellis, 2001; Luby-Phelps, 1999; van den Berg et al., 2017) that demonstrates position-dependent diffusivity (Berret, 2016; Garner et al., 2022; Śmigiel et al., 2022; Xiang et al., 2020). Diffusion can occur within or between cellular compartments, where concentrated components carry out chemical reactions. The rich interaction of diffusion and compartmentalization provides the context for cellular biochemistry. Diffusivity varies inversely with viscosity, a key biophysical parameter of the cytoplasm (Bausch et al., 1999; Hu et al., 2017) that dictates translational and rotational mobility of proteins and, by extension, possibly influences their activity (Huang et al., 2021; Lippincott-Schwartz et al., 2001; Pan et al., 2009). While viscosity has been implicated in modulating or driving a range of cellular processes (Molines et al., 2022; Persson et al., 2020; Xie et al., 2022), the role of *inhomogeneous* viscosity in shaping biochemistry by regulating biomolecular concentration and dynamics remains poorly understood.

Any conceptualization of diffusion in the presence of position-dependent viscosity must confront the so-called 'Itô-Stratonovich dilemma' according to which the equilibrium

concentration distribution of a diffusing tracer depends not only on the localized viscosity distribution but also on conventions based on microscopic parameters not captured in a coarse-grained model of diffusion (Bo et al., 2021; Lau and Lubensky, 2007; Sokolov, 2010; Tupper and Yang, 2012; Van Kampen, 1988). It is possible, under certain such conventions, for modeled particles to accumulate in areas of low diffusivity, leading to a concentration gradient (Volpe and Wehr, 2016), as has been experimentally observed (Bringuier, 2011; Pesce et al., 2013; Volpe and Wehr, 2016); however, the implication of such diffusive lensing for dynamics occurring at timescales and length scales relevant to biology remain largely unexplored. In particular, such accumulation may represent a novel mechanism of compartmentalization, a key activity for cells in regulating biochemical processes.

In this work, we employ agent-based modeling to determine how position-dependent viscosity affects the distribution of tracer particles. We show that viscophoresis (transport due to a viscosity gradient) leads to diffusion trajectories being biased toward areas of higher viscosity, leading to compartmentalization and the growth of concentration gradients; we call this effect "diffusive lensing," in non-quantitative analogy to the effects on light rays of media with inhomogeneous refractive index, including refraction and the formation of caustics. Analyzing particle trajectories, we show that viscophoretic transport manifests as anomalous diffusion, albeit unlike the canonical forms seen in the case of homogeneous diffusion in confined setups. We conclude that inhomogeneous diffusivity has diverse implications for intracellular transport, from sequestering particles to modulating where and when higher-order processes such as clustering happen, in a way that is not predictable from equivalent homogeneous-diffusivity models and may affect biochemical reactions.

Results

Viscophoresis drives particle accumulation

We probed the effect of inhomogeneous diffusion on particle concentration using agent-based modeling of Brownian dynamics (Fig. S1A; see *Methods*). The expected macroscale behavior is dictated by the 1D diffusion equation: $\frac{\partial c}{\partial t} = \frac{\partial^2 cD}{\partial x^2}$, where $c(x,t)$ denotes the concentration distribution and $D(x)$ denotes the position-dependent diffusivity (Oksendal, 2013). This equation conforms to the Itô interpretation of inhomogeneous diffusivity (Volpe and Wehr, 2016); it is non-anticipatory in that for a modeled particle traversing the sharp viscosity interface, the step size distribution is defined by the viscosity at its *present* position. Other equally consistent interpretations (such as the entirely anticipatory "isothermal" interpretation) produce different macroscale behaviors (Fig. S1B). The range of physically-incompatible possibilities resulting from different interpretations is known as the Ito-Stratonovich dilemma (Lau and Lubensky, 2007; Sokolov, 2010; Tupper and Yang, 2012; Van Kampen, 1988; Volpe and Wehr, 2016).

Over the course of the simulation, particles accumulated in the higher-viscosity zone (Fig. 1A), consistent with steady state closed form Itô-convention solutions (Oksendal, 2013). This

accumulation entailed the transient depletion of particles on the less-viscous side of the interface. A similar accumulation was observed in a smooth viscosity gradient (Fig. 1B). Thus, agent-based simulations demonstrate that under the Itô convention, areas of increased viscosity lead to increases in the concentration of diffusing particles. This is the phenomenon which we term "diffusive lensing."

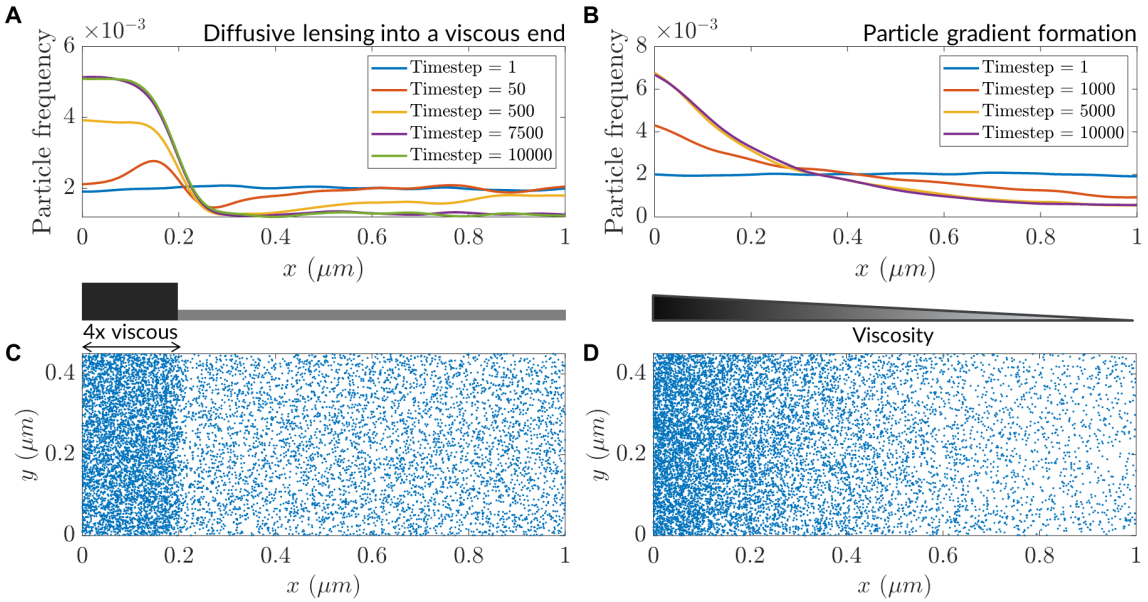

Figure 1: Viscophoresis leads to accumulation of particles. (**A**) Particle distribution at various timesteps of a simulation with a step-like higher-viscosity region. (**B**) Particle distribution at various timesteps for a simulation with a viscosity gradient. (**C**) Equilibrium particle distribution for the simulation in (A). (**D**) Equilibrium particle distribution for the simulation in (B).

Interaction-mediated clustering is affected by viscophoresis

Diffusive lensing is an interaction-free mode of concentrating particles that stands in contrast to a more typical paradigm of particle accumulation: interaction-driven formation of higher-order structures like protein complexes, gels, crystals and phase-separated condensates (Banani et al., 2017; Vekilov, 2010). How might interaction-induced clustering be modulated by inhomogeneous diffusion in a cellular context? To address this question, we heuristically modeled inter-particle interactions via a neighbor-sensing scheme in high and low interaction-strength regimes. The scheme involved using a step size for the modeled particle, which decreases as the number of particles in the vicinity increases (see *Methods*). At low interaction strength, clustering occurred only at the high-viscosity end of a gradient (Fig. 2A), while the same interaction strength was insufficient to produce clusters in a uniform viscosity distribution (Fig. S2A, S2C). In contrast, a high interaction strength resulted in robust clustering manifesting before particle gradient formation reached the steady state, leading to clustering towards the low-viscosity side of the simulation region as well (Fig. 2B). At this high interaction strength, the clustering rate remained the same throughout the region in the absence of a gradient (Fig. S2B, S2D). Taken together, the results reveal that viscophoresis can modulate clustering and under certain circumstances cause viscosity-dependent localized

cluster formation, and furthermore that the relative strengths and timescales of each phenomenon quantitatively dictate whether increased clustering will preferentially occur in viscous zones. Effects of diffusive lensing on particle concentration may additionally regulate reaction rates and drive stochastic clustering of enzymes (Jilkine et al., 2011).

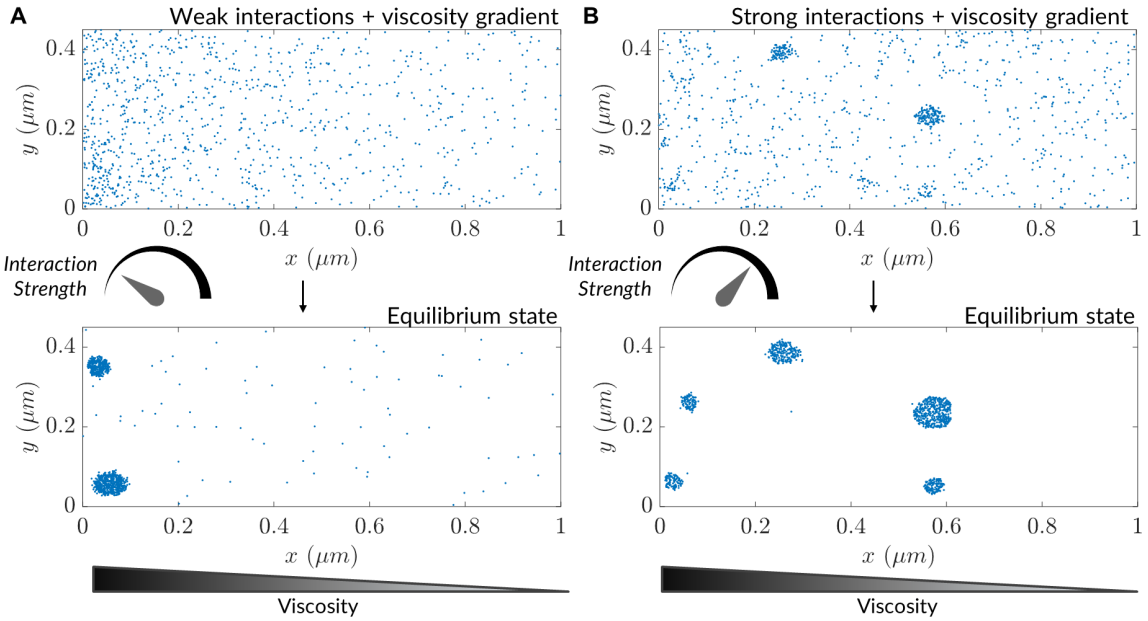

Figure 2: Interaction-driven clustering is modulated by viscophoresis. (**A**) Progress of a simulation comprising particles possessing weak interactions ($k = 0.04$ is the interaction strength; see *Methods*), initialized with a uniform concentration of particles. (**B**) Progress of a simulation comprising particles possessing strong interactions ($k = 0.1$), initialized with a uniform concentration of particles.

In silico microrheology shows that viscophoresis manifests as anomalous diffusion

The diffusion coefficient is a fundamental biophysical parameter that affects numerous other phenomena, including biochemical reaction rates. To elucidate particle diffusion at the microscale in the context of viscophoresis, we used an *in silico* implementation of microrheology to analyze particle trajectories (see *Methods*; Fig. S3A). We computed the mean squared displacements (MSDs) for uniform viscosity simulations (in the case of unencumbered and confined diffusion) and used these to understand how MSD is affected by heterogenous viscosity in two cases: a continuous viscosity gradient and a discrete step in viscosity.

Particle diffusion was unencumbered in the case of large bounds (relative to step size) (Fig. 3A) and confined in the case of small bounds (Fig. 3B), with the latter demonstrating a transition from anomalous to normal diffusion with time, all in agreement with earlier results (Dix and Verkman, 2008; Saxton, 2007). The MSD at saturation in homogeneously viscous systems was found to be agnostic to the underlying uniform viscosity of the system, indicating that it is exclusively determined by the simulation region size. In contrast, particles in a viscosity gradient exhibited dynamics intermediate to those of homogeneous high and

low viscosity cases, both in the diffusion coefficient and saturation MSD (Fig. 3C, inset). The lowering of the saturation MSD reflects particle diffusion occurring within apparent simulation region bounds that confine more than the actual simulation region size. We note that such modifications of apparent geometry are also a feature of optical lensing. Apparent bounds were also found to occur in the two-zone viscosity case (as in Fig. 1A) where, at steady-state, particles populated the simulation region non-uniformly (Fig. S3B). For most of the viscosity ratio parameter space, irrespective of whether the smaller zones were more fluid or viscous relative to the bulk, a reduction in MSD was seen indicating effectively lower diffusion bounds (Fig. 3D). The magnitude of reduction depended on whether most particles resided in the larger or smaller of the two zones. In one observed case ($\frac{\mu_i}{\mu_o} = 4$), however, the saturation MSD was higher than what was seen in the homogeneous diffusion scenario possibly due to particles robustly populating the bulk milieu followed by directed motion into the viscous zone (similar to that of a Brownian ratchet, (Peskin et al., 1993)). The saturation MSD was also found to depend on the location of the viscous zone: a more-centered zone resulted in a lowered saturation value, possibly due to weaker ratchet effects (Fig. S3C, D). Note that lensing may cause particle displacements to deviate from a Gaussian distribution, which could explain anomalous behaviors observed in our simulations and experiments in cells (Parry et al., 2014). These results point to the insufficiency of using the diffusion coefficient alone to describe diffusion in heterogenous milieu. They also indicate a rich interplay between heterogenous viscosity and anomalous diffusion that requires further investigation.

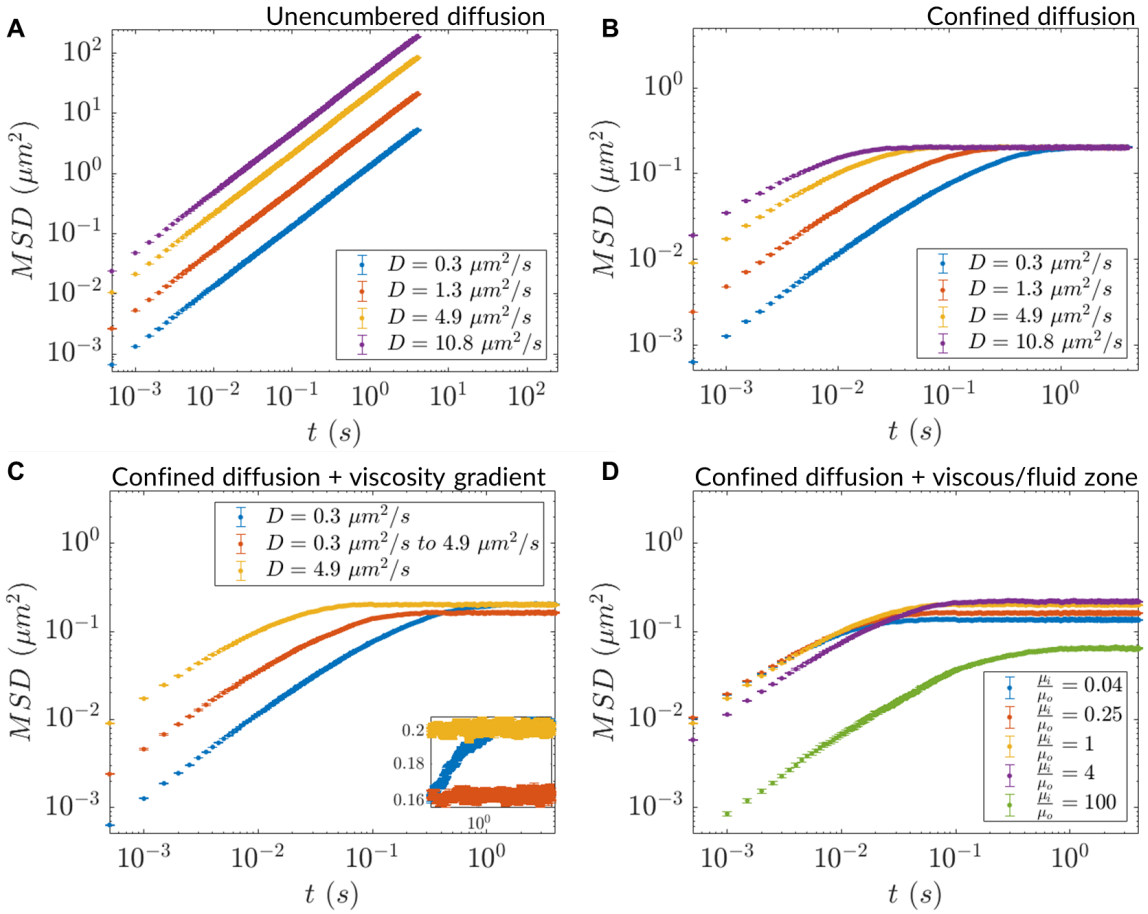

Figure 3: Inhomogeneous diffusivity can manifest as anomalous diffusion. (**A**) MSD versus time for homogeneous diffusion of 10,000 particles in a 5 mm x 5 mm simulation region. (**B**) Same as (A) for homogeneous diffusion in a more tightly bounded simulation region (1 μm x 0.45 μm). (**C**) MSD versus time for inhomogeneous diffusion in a viscosity gradient versus homogeneous diffusion in the extreme viscosity cases (simulation region size: 1 μm x 0.45 μm). Inset: zoomed region showing differential saturation of the MSD. (**D**) MSD versus time for inhomogeneous diffusion due to a stepwise viscosity distribution with viscosity ratio $\frac{\mu_i}{\mu_o}$ relative to the bulk (simulation region size: 1 μm x 0.45 μm). In all cases, n = 10,000 particles for MSD calculation (error bars denote SEM).

***In silico* FRAP in heterogeneously viscous environments reveals drivers of mesoscale dynamics**

The *in silico* microrheology analysis we performed provided insights into dynamics at the single-particle level (i.e., the microscale). To explore collective, emergent behaviors at the mesoscale while continuing to make contact with feasible experiments, we employed an *in silico* version of fluorescence recovery after photobleaching (*in silico* FRAP) (Fig. S4A, B), in more cell-like inhomogeneous environments. In particular, we modeled viscous patches/granules in a cell using a three-parameter disc-packing setup comprising granule radius (r), packing density (ϕ) and the ratio of granule viscosity to bulk viscosity ($\frac{\mu_i}{\mu_o}$) (see *Methods*). We investigated the effect on dynamics of varying these parameters individually, with the goal of gaining understanding of the effects of varying the amount, nature and distribution of viscogens in cells. In all cases, the *in silico* "photobleaching" event was conducted after equilibration (Fig. S4C,D,E). To explain observed changes in recovery, we probed how the mean dwell time spent by particles in viscous granules varies as a function of these parameters. An increase in the viscosity ratio ($\frac{\mu_i}{\mu_o}$) at fixed ϕ and r resulted in a decline in measured particle mobility, as characterized by an increase in the simulated FRAP $t_{1/2}$ values (Fig. 4A). Increasing $\frac{\mu_i}{\mu_o}$ from 1 to 10 caused a doubling of $t_{1/2}$ (or halving of diffusivity). Similar reduction in mobility was observed upon variation of ϕ or r separately, keeping the other (and the viscosity ratio, $\frac{\mu_i}{\mu_o}$) constant (Fig. 4B, C). The decrease in average mobility in all three cases arose from changes in flux between the viscous and bulk zones, as reflected by an increase in mean dwell times of particles within viscous granules (Fig. S4F,G,H). Furthermore, such reductions in mobility were emergent in that they arose from the interplay between granular viscosity and bulk-granule fluxes, as the regions of interest in the simulated photobleaching events comprised granules and the surrounding bulk environment. To investigate whether particle dynamics is affected by the underlying topography realizing the system's viscosity, we averaged the granular and bulk viscosity values to produce weighted-average viscosity values, and compared *in silico* recovery in these simulations to that of the equivalent granule-comprising simulations. Such an averaging of the viscosity to cause an effective uniform mobility for all resident particles, resulted in slower dynamics than that of the equivalent granule-comprising simulations (Fig. 4D). We conclude that inhomogeneity in viscosity drives rapid effective dynamics via fluxes between

the granular ("interior") and bulk ("exterior") environments, creating expressways for particles to move rapidly between viscous regions. The diffusive lensing of particles into viscous zones, and their consequent dwelling in these regions, can be tuned by modulating the underlying viscosity distribution in myriad ways.

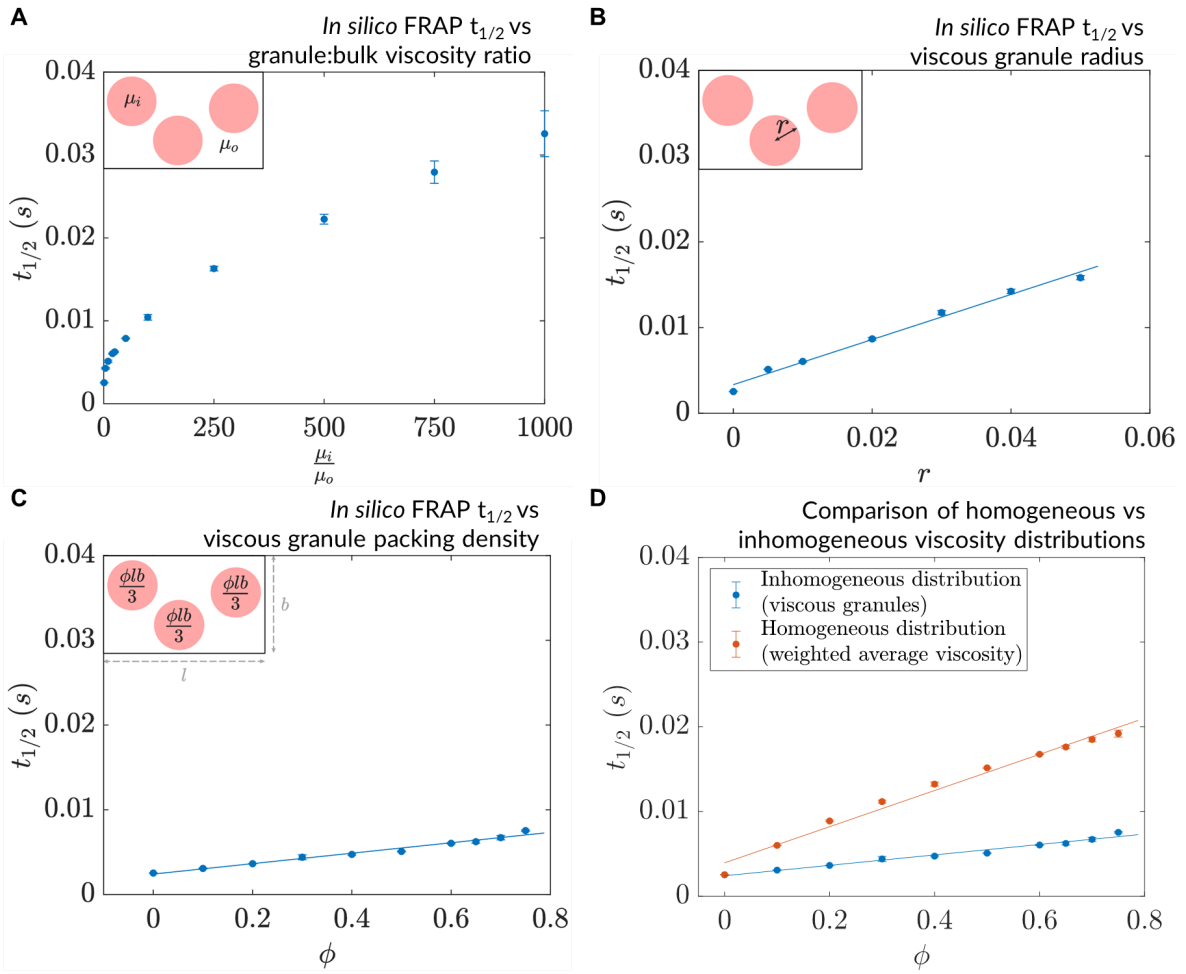

Figure 4: An increase in granule viscosity, radius, or packing density slows down mesoscale dynamics. (**A**) Simulated FRAP $t_{1/2}$ as a function of granule:bulk viscosity ratio (r = 0.01 μm, ϕ = 0.6). (**B**) Simulated FRAP $t_{1/2}$ as a function of granule radius ($\frac{\mu_i}{\mu_o}$ = 20, ϕ = 0.6). (**C**) Simulated FRAP $t_{1/2}$ as a function of granule packing density ($\frac{\mu_i}{\mu_o}$ = 20, r = 0.01 μm). (**D**) Simulated FRAP $t_{1/2}$ for homogeneous and inhomogeneous viscosity setups realizing the same effective viscosities ($\frac{\mu_i}{\mu_o}$ = 20, r = 0.01 μm). In all cases, n = 3 ROIs were chosen for the simulated photobleaching (error bars denote SEM).

Discussion

The complex, heterogeneous milieu of the cellular interior has been recently shown to cause heterogeneous diffusion (Garner et al., 2022; Huang et al., 2021; Parry et al., 2014; Śmigiel et al., 2022; Xiang et al., 2020), yet the consequences of such inhomogeneity on compartmentalization and mesoscale molecular dynamics have remained unclear. Through agent-based modeling of diffusion, we have shown that heterogenous viscosity can lead to

simulated particle trajectories converging into viscous hotspots, causing the accumulation of diffusing particles into membraneless compartments defined by the higher-viscosity zones. We term this mode of transport "diffusive lensing". We show that it has wide-ranging effects on particle distribution and dynamics and, furthermore, that it can occur across a wide parameter space. This leads us to speculate that diffusive lensing is a ubiquitous phenomenon in living systems, which are inherently removed from equilibrium by energy-driven processes (Phillips et al., 2012) and, wherein, non-isothermal interpretations of heterogeneous diffusion may be meaningful (Wang et al., 2020).

We found that inhomogeneous viscosity allows for particle mobility at the microscale and mesoscale to be different from that expected in the presence of homogeneous diffusion. Such an expectation is in line with predicted and observed deviations from normal diffusion in cells (Bancaud et al., 2012; Baum et al., 2014). The relative strengths of viscophoresis and inter-particle interactions (if any) determined the extent to which clustering was modulated by diffusive lensing: this interplay may be important for determining the effects of inhomogeneous viscosity on biochemical reaction rates. Spatially-averaged effective diffusion timescales were found to depend on the microscopic viscosity distribution: the same average viscosity can give rise to slower or faster dynamics depending on whether it is realized via homogeneous or heterogenous viscosity distributions. In the latter case, the bulk region interspersed between the viscous hotspots provides "expressways" that contribute to large fluxes at the viscosity interface, thereby accounting for the faster dynamics. Such expressways and their associated fluxes may impact reaction kinetics by altering substrate turnover rates, congruent with the model of unusual transport processes potentially modifying reaction kinetics (Bénichou et al., 2010). In the context of subcellular viscous regions (Garner et al., 2022), cells may compensate for geometry-imposed constraints on packing density and size of these regions by altering the viscosity ratio (against the bulk milieu) instead. To map the detailed effects of inhomogeneous viscosity on reaction rates, however, our work suggests that a key prerequisite is to chart a suitable set of metaparameters that provide an adequate description of inhomogeneous diffusion (Jin and Verkman, 2007), as a one-parameter description relying exclusively on the average diffusion coefficient is insufficient.

Changes in viscosity have been shown to occur in the context of cellular processes including cell death (Kuimova et al., 2008), stress adaptation (Persson et al., 2020) and protein aggregation (Thompson et al., 2015). At any given time point, intracellular transport dynamics arises emergently from contributions across length scales ranging from crowding in the bulk milieu due to proteins (Wang et al., 2010) and large biomolecules (Delarue et al., 2018) to cytoskeleton (Carlini et al., 2020; Chaubet et al., 2020) and active flows in the cytoplasm (Arcizet et al., 2008), all leading to unusual anomalous diffusive behaviors at the mesoscale (Banks and Fradin, 2005; Bressloff, 2014; Dix and Verkman, 2008; Höfling and Franosch, 2013; Kuznetsova et al., 2015; Swaminathan et al., 1997; Zhou et al., 2008). These diffusive behaviors cannot be decoupled from the intrinsic heterogeneity in biomolecular properties themselves (Heald and Cohen-Fix, 2014; Milo and Phillips, 2015). The effects of all of these subcellular determinants and processes on how position-dependent diffusivity is maintained in a cell remains unclear. Our work underscores the need to not only examine

viscosity distributions *in vivo* as a function of local composition and the environment, but also to study their time-evolution in response to external stimuli. More speculatively, we suggest that diffusive lensing serves as a potential candidate for a rudimentary mode of pre-biotic compartmentalization. Viscophoresis-driven accumulation of diverse biomolecules may have served to produce chemically enriched spaces, acting as an antecedent of more sophisticated, membrane-bound and membraneless organizational modalities; such a protocell organization is orthogonal to currently studied models (Monnard and Walde, 2015). This work demonstrates that diffusive lensing can have strong effects on transport and may be common in cellular contexts, modulating both passive and active flows. Future experimental and theoretical work will elucidate the extent of lensing inside and outside of cells and its effects on the biochemical reactions that sustain life.

Methods

Agent-based modeling (random walk simulations)

Agent-based modeling of diffusion was conducted via 2D random walk simulations. Non-interacting point particles were initialized uniformly in a 2D simulation region with an aspect ratio matching that of an E. coli bacterium (Phillips et al., n.d.). During each time step (also termed epoch or frame), every particle was moved along each coordinate by step sizes sampled from a uniform distribution, $\mathcal{U}(-S, S)$, where S denotes the step size limit. Across a large number of steps, the distribution of displacements converges to the normal distribution by virtue of the central limit theorem. While sampling was not performed via the normal distribution directly by using the diffusion coefficient (D) as a parameter, the diffusion coefficient was instead arrived at as an emergent property of trajectories comprising a simulation, in a ground-up fashion. Reflection off the wall was modeled using a mirror-image rule. To model a zone of differential viscosity relative to bulk viscosity (either a fluid or a viscous zone), particle step sizes were sampled from zones characterized by different viscosities, noting that the diffusion coefficient and viscosity are inversely related (Phillips et al., n.d.) and $S \propto \sqrt{D}$. At all times, step sizes were sampled from distributions defined by the viscosity around the present position in accordance with the Ito interpretation of multiplicative noise (Volpe and Wehr, 2016). In all simulations, a set seed of 1 was used for the random number generator. Simulations were run on MATLAB R2020a on Sherlock (a high-performance computing cluster at Stanford).

In the simulations which included inter-particle interactions, these interactions were modeled via a neighbor-sensing approach. The step size limit was modified as per the relation, $S_{eff} = Se^{-kn}$, where k denotes the sensing strength and n denotes the number of neighbors (defined as those particles lying within a cutoff span around the particle in question). Such a rule-based approach modeled an effective attractive potential for the inter-particle interactions. Local density calculation used the same cutoff and the data were normalized to the mean local density of particles during initialization. Considering the computational work due to neighbor-sensing, a smaller number of particles (10^3) were deployed, for a longer period of 2×10^4 epochs.

In the viscous granule simulations, the granules were modeled as disks with randomly initialized centers and fixed radii (r), covering the simulation region up to a desired packing density, ϕ. The algorithm saturated for $\phi \geq 0.6$, in which case, the disks were generated as per cubic close packing and their positions were incrementally deviated over 10^9 steps to reduce local ordering as much as possible. The ratio of viscosity inside the granules to viscosity outside the granules ($\frac{\mu_i}{\mu_o}$) was the third parameter under consideration. No two disks were allowed to overlap and all disks were kept confined within the boundaries of the simulation region. The default setup is as follows: r = 0.01 μm (uniform), ϕ = 0.6 (that is, 60% of the simulation region is covered by the granules) and $\frac{\mu_i}{\mu_o}$ = 20. Titration of one of these three parameters involved keeping the other two at the specified levels.

Numerical methods for the diffusion equations

The Fokker-Planck equations corresponding to the Ito, Stratonovich and isothermal interpretations of inhomogeneous diffusion are as follows (Oksendal, 2013; Tupper and Yang, 2012) (here $c(x,t)$ denotes the concentration distribution and $D(x)$ denotes the position-dependent diffusivity):

Ito interpretation: $\frac{\partial c}{\partial t} = \frac{\partial^2 cD}{\partial x^2}$,

Stratonovich interpretation: $\frac{\partial c}{\partial t} = \frac{\partial}{\partial x}\left(\sqrt{D}\frac{\partial c\sqrt{D}}{\partial x}\right)$,

Isothermal interpretation: $\frac{\partial c}{\partial t} = \frac{\partial}{\partial x}\left(D\frac{\partial c}{\partial x}\right)$.

These equations were numerically evaluated via forward time centered space (FTCS) schemes, with length and time increments set as 10^{-2} and 10^{-5} arbitrary units, respectively, and the number of time steps was set to 5000. A gaussian well profile was used for the diffusion coefficient and the initial condition for the concentration distribution was a uniform distribution (Fig. S1B). Numerical analysis and data visualization were performed on MATLAB R2019a.

***In silico* microrheology**

Analysis of particle trajectories was carried out via quantifying the mean squared displacements (MSD). These were calculated from 10^4 trajectories (each 10^5 timesteps in duration) per simulation. The timestep was set as $50\ \mu s$ so that the diffusion coefficient was $\approx 5\ \mu m^2/s$ (order of magnitude for a small protein's mobility in the *E. coli* cytoplasm (Milo and Phillips, 2015)).

***In silico* FRAP**

In silico fluorescence recovery after photobleaching (FRAP) studies were performed on the diffusion simulations to quantify emergent dynamics at the mesoscale. 10^5 particles were

deployed for a total duration of 0.5 s (10^4 epochs). Circular regions (radius of $0.2\ \mu m$) were chosen as the regions of interest (ROIs). *In silico* photobleaching was instantaneously performed and involved assigning the particles in the ROI the photobleach status. The background was chosen from a uniform viscosity setup to ensure that the normalization is standardized. The outward turnover of these particles and the simultaneous inward flux of unbleached particles were captured via $t_{1/2}$, the time taken for recovery up to the 50% of the steady state level of unbleached particles in the ROI (Sprague and McNally, 2005). In these simulations, $t_{1/2}$ connotes the time taken for the number of "unbleached" particles in the ROI to reach 50% of the steady-state value. To dissect particles' behavior during the simulation (in terms of bias towards inhabiting the viscous granules), we calculated the mean dwell time across all particles, per simulation. This involved averaging the periods (of any duration) spent by particles inside viscous granules. For normalization, the total simulation duration was used (0.5 s).

Code and data availability

Code and data utilized in this study will be made available upon reasonable request.

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Supplementary Figures

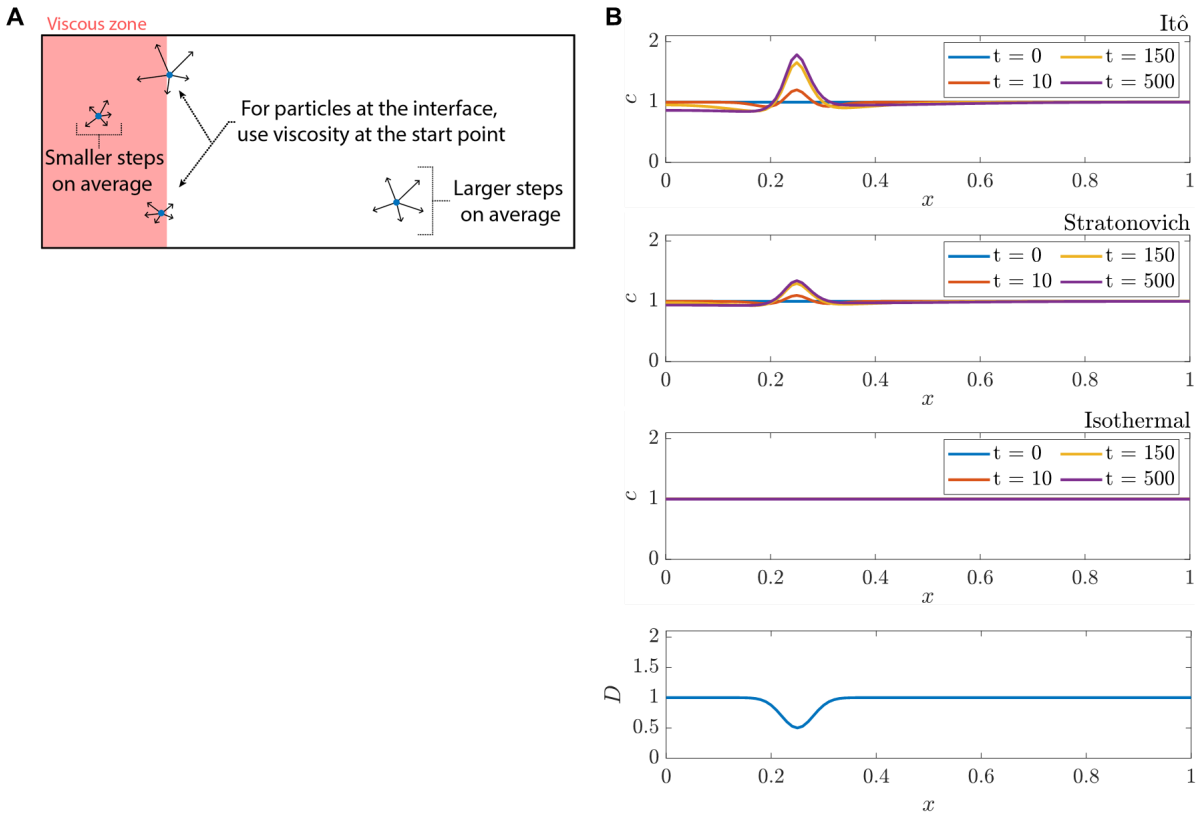

Figure S1: Itô convention leads to Fokker-Planck diffusion, contrasting canonical ("Fickian") homogenization. (**A**) Agent-based modeling of Brownian dynamics used in this study. Choosing the viscosity at the start point of a particle hop is in line with the Itô interpretation. (**B**) Numerical solutions for drift-less Fokker-Planck equations with an inhomogeneous diffusion coefficient, for the Itô, Stratonovich and isothermal conventions.

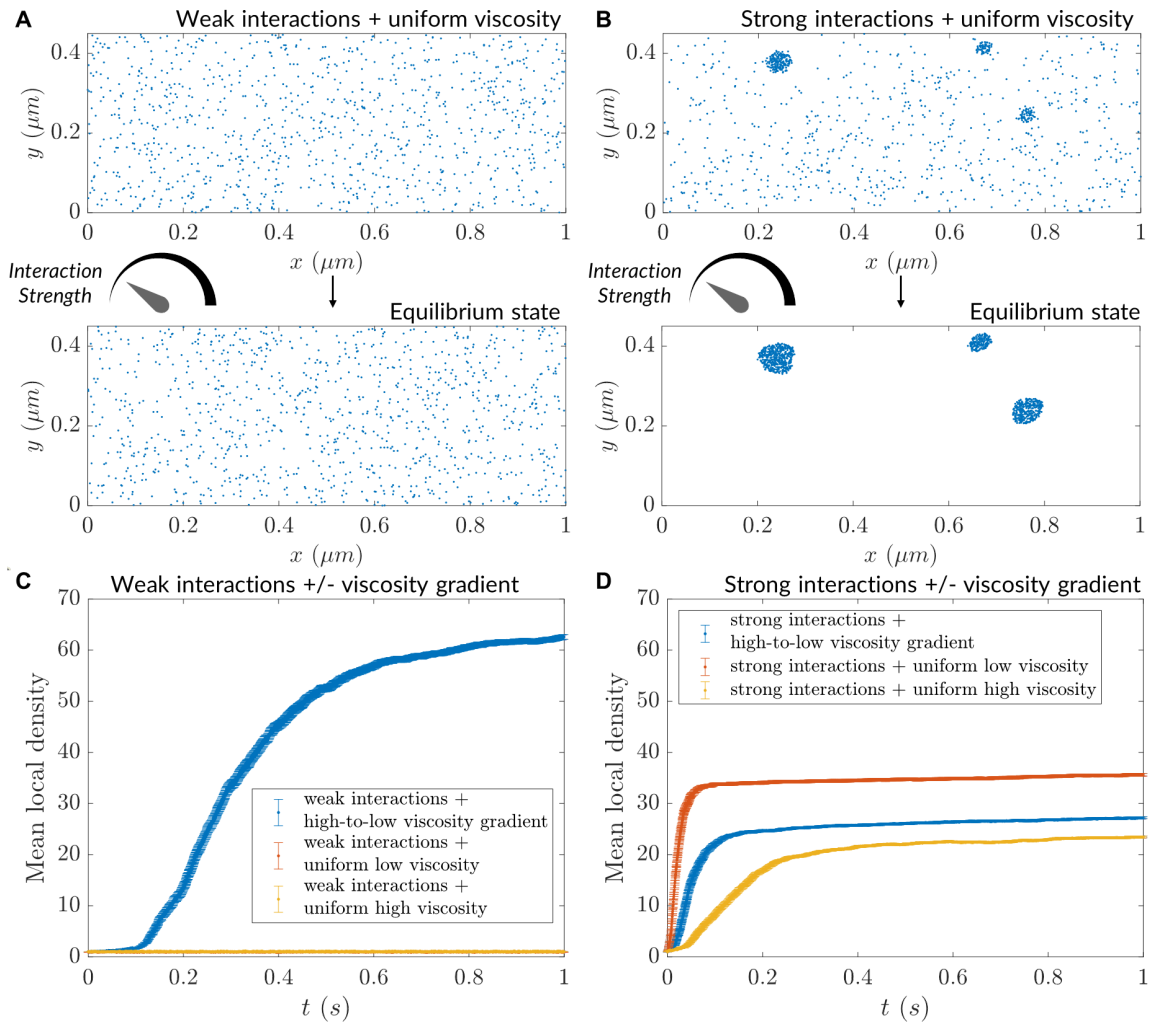

Figure S2: Particle clustering at different strengths in homogeneous versus heterogeneous viscosity environments. (**A**) Progress of a simulation comprising particles possessing weak interactions ($k = 0.04$), initialized with a uniform concentration of particles; no viscosity gradient used here. (**B**) Progress of a simulation comprising particles possessing weak interactions ($k = 0.1$), initialized with a uniform concentration of particles; no viscosity gradient used here. (**C**) Mean local density versus time for particles possessing weak interaction strength. (**D**) Mean local density versus time for particles possessing strong interaction strength. For (C) and (D), n = 1000 particles for mean local density calculation (error bars denote SEM).

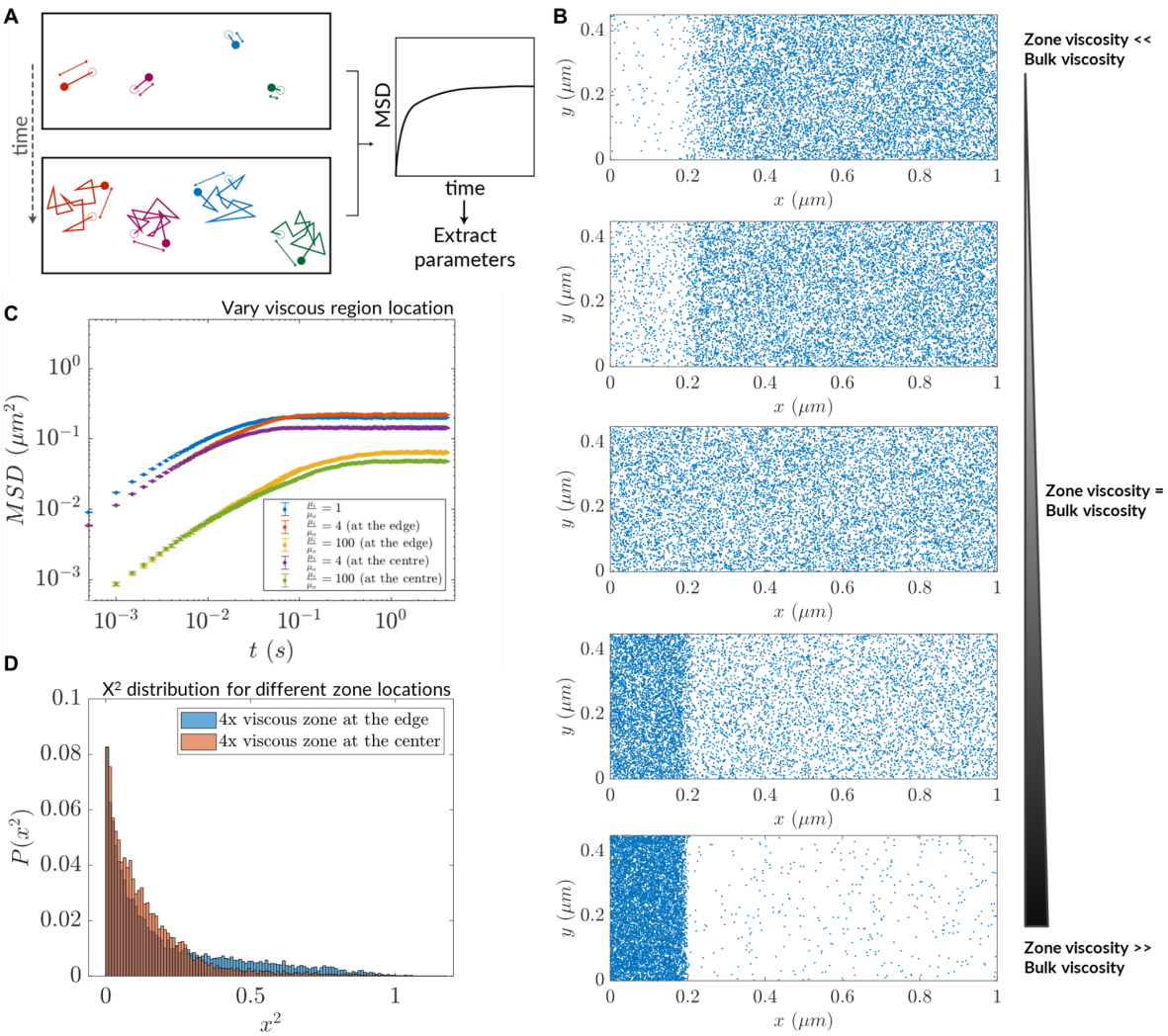

Figure S3: Magnitude and distribution of inhomogeneity in viscosity affects diffusive lensing. (**A**) Analysis of simulation trajectories via *in silico* microrheology. (**B**) Increasing viscophoretic extent due to variation of the zone viscosity in a chamber comprising a viscous end. (**C**) Mean squared displacement after transition to normal diffusion (saturation MSD) depends both on the magnitude of viscosity difference and the location of the zone itself. n = 10,000 particles for MSD calculation (error bars denote SEM). (**D**) Histogram of $\|X\|^2$ at the end of the run for the cases of the 4x viscous zone located at the simulation region edge versus the center.

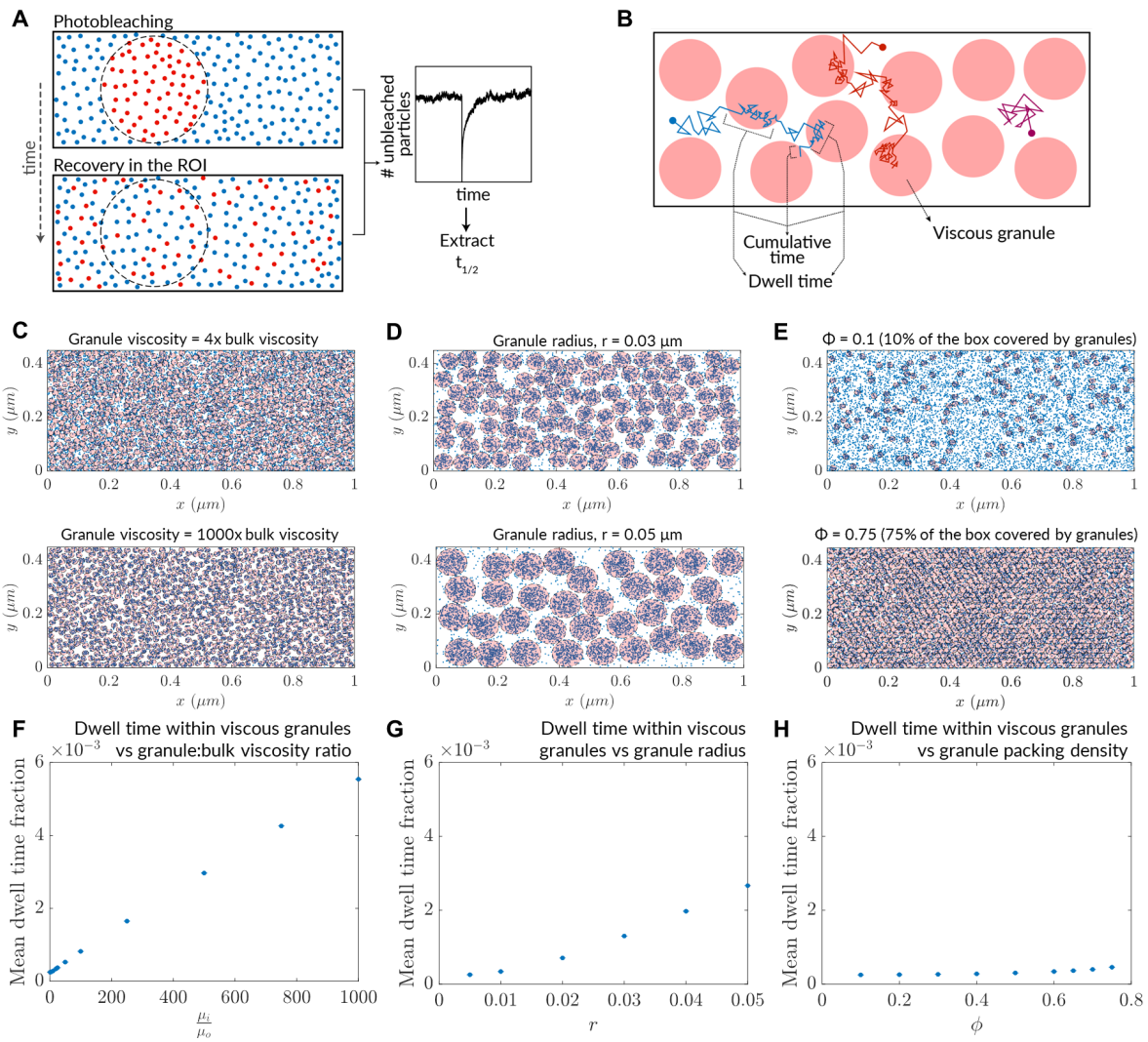

Figure S4: Dwell times for particles in viscous granules dictate FRAP kinetics. (**A**) The *in silico* implementation of FRAP used in this study. (**B**) Methodology for determining mean dwell time of particles in viscous granules, from a set of simulation trajectories. (**C**) Equilibrium state of systems in the variation of granule viscosity, before commencing *in silico* FRAP. (**D**) Equilibrium state of systems in the variation of granule radius, before commencing *in silico* FRAP. (**E**) Equilibrium state of systems in the variation of granule packing density, before commencing *in silico* FRAP. (**F**) Mean dwell time fraction variation as a function of granule:bulk viscosity. (**G**) Mean dwell time fraction variation as a function of granule radius. (**H**) Mean dwell time fraction variation as a function of granule packing density. For (F)-(H), n = 10,000 particles used for calculation (error bars denote SEM).