Altered visual cortex excitability in premenstrual dysphoric disorder: evidence from magnetoencephalographic gamma oscillations and perceptual suppression

Viktoriya O. Manyukhina (mvo96@inbox.ru)[1,2], Elena V. Orekhova (orekhova.elena.v@gmail.com)[1]*, Andrey O. Prokofyev (prokofyevao@mgppu.ru)[1], Tatiana S. Obukhova (obukhovats@mgppu.ru)[1], Tatiana A. Stroganova (stroganova56@mail.ru)[1]

[1] Center for Neurocognitive Research (MEG Center), Moscow State University of Psychology and Education,
Moscow, Russian Federation.
[2] National Research University Higher School of Economics, Moscow, Russian Federation.

* Corresponding author

A short running title: Altered visual cortex excitability in PMDD

Three key points:

- Atypical modulation of visual gamma power by excitatory drive in women with PMDD suggests the presence of neuronal hyperexcitability, which is exacerbated during the symptomatic luteal phase, possibly due to altered sensitivity to neurosteroids.

- The peak frequency of gamma oscillations does not differ between women with PMDD and control women and demonstrates the luteal phase-specific effect of the 1st scan in both groups, which suggest normal functioning of inhibitory neurons in the visual cortex in PMDD.

- The presence of normal perceptual spatial suppression also speaks against inhibitory deficit in the visual cortex in PMDD.

Abstract

Premenstrual dysphoric disorder (PMDD) is a psychiatric condition characterized by extreme mood shifts, anxiety, and irritability during the premenstrual period. Abnormal sensitivity to allopregnanolone, a neurosteroid that normally potentiates inhibition, and an increased ratio of neural excitation-to-inhibition (E/I) have been linked to the pathophysiology of PMDD. We hypothesized that in subjects with PMDD these factors will lead to an altered frequency of magnetoencephalographic (MEG) visual gamma oscillations, altered modulation of their power by the strength of excitatory drive and to an altered perceptual spatial suppression.

We examined women with PMDD and age-matched control women twice: during the asymptomatic follicular and symptomatic luteal phases of the menstrual cycle (MC). MEG gamma oscillations were recorded while modulating excitatory drive to the visual cortex by increasing drift rate of high-contrast visual gratings. Perceptual suppression was assessed as a degree of impairment of the discrimination of visual motion direction with increasing stimulus size.

In women with PMDD, the gamma response (GR) peak frequency and its modulation by the drift rate were normal, whereas modulation of the GR power was significantly altered. A moderate increase in drift rate had an unusually strong facilitating effect on the GR power in PMDD, regardless of the MC phase. In contrast, the effect of increasing drift rate, which normally suppresses GR power, was attenuated in PMDD in the luteal phase and predicted symptom severity estimated on the same day. Perceptual spatial suppression did not differ between the groups, but decreased from the follicular to the luteal phase only in PMDD subjects.

The atypical GR power modulation suggests that neuronal excitability in the visual cortex is constitutively elevated in women with PMDD, and that their E/I ratio is additionally shifted toward excitation during the luteal phase, possibly due to an abnormal sensitivity to neurosteroids. However, the unchanged frequency of GR and normal spatial suppression in women with PMDD speak against the dysfunction of their inhibitory neurons, at least those involved in generation of visual gamma oscillations.

Keywords: visual gamma oscillations, premenstrual dysphoric disorder, excitation/inhibition balance, magnetoencephalography, perceptual spatial suppression

Introduction

Female steroid hormones not only support reproduction but also regulate excitability of the neural cells. These hormones enter the brain by crossing the blood-brain barrier or are synthesized in the brain (Azcoitia et al., 2011; Bixo et al., 1986; Micevych & Sinchak, 2008) and affect emotions, perception, and memory (Farage et al., 2008). In the majority of females of reproductive age hormone fluctuations during the menstrual cycle (MC) cause only mild changes in physical state and mood a few days before menstruation, known as premenstrual symptoms (PMS) (Biggs & Demuth, 2011; Direkvand-Moghadam et al., 2014). However, in 2-5% of women they result in disabling psychiatric problems, also known as premenstrual dysphoric disorder (PMDD) (Halbreich et al., 2003; Tschudin et al., 2010). Over the past decades, PMDD has attracted attention of both clinicians and the scientific community (Gao et al., 2021), as it is associated with substantial disruption of womens' everyday activities, inability to work and high risks of suicide (Chan et al., 2021; Wikman et al., 2022; Yang et al., 2021).

Normal as well as pathological changes in the brain during the MC to a large extent depend on varying concentration of two steroid hormones – progesterone and estradiol. Although mechanisms of their action are not fully understood, there is evidence that progesterone and estradiol either directly or indirectly modulate neuronal excitability and affect excitation-to-inhibition (E/I) balance in the brain (Finocchi & Ferrari, 2011).

There is evidence that PMDD symptoms are related to an altered neural inhibition which results from abnormal sensitivity to neurosteroids rather than their altered levels in the brain (Bäckström et al., 2011; Dubey et al., 2017; Schmidt et al., 1998; Timby et al., 2016; Turkmen et al., 2011). The neuroactive effects of progesterone associated with PMS are thought to be mediated by its neurosteroid metabolite allopregnanolone (ALLO) (Bäckström et al., 2011; Hantsoo & Epperson, 2020; Timby et al., 2016). ALLO is a positive allosteric modulator of $GABA_A$ receptors ($GABA_A$Rs) that primarily facilitates tonic inhibition mediated by δ subunit-containing $GABA_A$Rs (δ$GABA_A$Rs) (Reddy, 2010). However, under certain conditions it can also produce a paradoxical excitatory effect (Bäckström et al., 2014; A. M. I. Barth et al., 2014; Lovick, 2014; Wohlfarth et al., 2002). Estradiol also affects neuronal excitability via several pathways (Balthazart & Ball, 2006; Clemens et al., 2019; Huang et al., 2015; Kow et al., 2005; Schroeder et al., 2017; Schwarz et al., 2008; X. Wu et al., 2013; Y. W. C. Wu et al., 2014). In general, both progesterone and estradiol produce complex effects on neuronal excitability that depend on the brain area, neurosteroid levels, and other factors.

Symptoms of PMDD, such as depressed mood, anger, irritability, sleep problems, etc. appear to be primarily associated with functional abnormalities in the brain regions involved in emotional regulation (Dan et al., 2020; Dubol et al., 2020; Petersen et al., 2019). However, changes associated with PMDD are not limited to these regions (Baller et al., 2013; Bannbers et al., 2012; Comasco et al., 2014; Niederhofer, 2009; Smith et al., 2003). In particular, activity of the visual cortex is affected by neurosteroids (Jeong et al., 2011; Neill Epperson et al., 2002; Schloemer et al., 2020; Sergeeva et al., 2018) and may reflect MC-related abnormalities in the E/I balance regulation in women with PMDD. Visual cortex displays the highest concentration of $GABA_A$ and $GABA_B$ receptors among other brain areas in humans (Zilles et al., 2002), and its GABAergic tone is sensitive to fluctuations in ALLO (Sergeeva et al., 2018). Estradiol is another potent modulator of visual cortex excitability (Jeong et al., 2011).

Evidence for sensitivity of neural activity in the visual cortex to steroid hormones during the MC also comes from several electrophysiological studies. Amplitude and latency of cortical visual evoked potentials (Avitabile et al., 2007; De Lima Resende et al., 2000; Lusk et al., 2015; Yilmaz et al., 1998), as well as magnitude of the transcranial magnetic stimulation (TMS) paired-pulse suppression of visual evoked potentials (Schloemer et al., 2020) change with phase of the MC. Parameters of the visual alpha rhythm in women without severe PMS are also affected by phase of the MC (Bazanova et al., 2014; Brötzner et al., 2015; Haraguchi et al., 2021; Lin et al., 2013). Besides, in a recent study Sumner and colleagues have shown that frequency of gamma oscillations induced by static and moving visual gratings also changes during the MC in women without severe PMS (Sumner et al., 2018).

Functional changes in the visual cortex during the MC may reflect modulatory effects of neurosteroids on excitatory and inhibitory neurotransmission. Gamma oscillations may be particularly sensitive to these modulatory effects, as they closely reflect local cortical interaction of excitatory and inhibitory neurons and are sensitive to the balance of their activity (E/I balance) (Bartos et al., 2007; Buzsáki & Wang, 2008; Ray & Maunsell, 2015). These oscillations are effectively induced by high-contrast regular patterns, such as gratings, and can be most reliably captured in humans with magnetoencephalography (MEG) (Hoogenboom et al., 2006; Muthukumaraswamy & Singh, 2013).

Our previous MEG studies suggest that individual variations in regulation of the E/I balance are better captured by stimulation-related *changes* of oscillatory gamma response (GR) rather than by GR amplitude or frequency measured in a single experimental condition (Manyukhina et al., 2021; Orekhova et al., 2019; Orekhova, Rostovtseva, et al., 2020). Strong and predictable modulations of GR power and

frequency can be achieved when modulating the excitatory drive to the visual cortex by increasing drift rate of a large high contrast visual grating: while the frequency of gamma oscillations increases nearly linearly with increasing drift rate, the GR power usually increases with transition from static to slowly drifting grating and then decreases with further increase of the drift rate. As we discussed elsewhere (Orekhova et al., 2018, 2019; Orekhova, Prokofyev, et al., 2020; Orekhova, Rostovtseva, et al., 2020), this pattern of changes in GR power may reflect efficiency of neuronal inhibition: some level of activity of inhibitory interneurons is necessary for synchronization of gamma oscillations (Mann & Mody, 2010; Rodriguez et al., 2004), but their 'too strong' activation – which in our experimental paradigm was associated with 'medium' and 'fast' drift rate (Orekhova, Prokofyev, et al., 2020) – may lead to a disruption of synchronization between principal cells and interneurons resulting in reduced amplitude of gamma oscillations (Börgers & Kopell, 2005; Cannon et al., 2014). Following this logic, a higher neural E/I ratio would lead to less suppression of GR for the same level of excitatory drive (i.e., at the same drift rate of a grating), because in this case more inhibition is required to desynchronize strongly excited principle cells (Börgers & Kopell, 2005). We therefore predicted that the presumably elevated E/I ratio in the visual cortex in women with PMDD during the symptomatic premenstrual period would result in relatively weaker GR suppression with increasing drift rate.

Next, we predicted that MC phase-related changes in E/I ratio in women with PMDD would affect the frequency of their gamma oscillations. Animal studies suggest that gamma frequency strongly depends on the tonic excitability of inhibitory neurons, which is regulated through δGABA$_A$Rs, so that the lower the tonic inhibition of the inhibitory interneurons, the higher the gamma frequency (Ferando & Mody, 2015; Mann & Mody, 2010). The δGABA$_A$Rs are highly sensitive to neurosteroids, and their numbers increase following an increase in progesterone levels (Barbaccia et al., 1998; Mody & Maguire, 2012; Purdy et al., 1991). However, Sumner et al. (2018) found that in women without severe PMS, the frequency of visual gamma oscillations was *higher* during high-progesterone-and-ALLO luteal phase than during low-progesterone-and-ALLO follicular phase. The authors hypothesized that these changes might be caused by developing tolerance to ALLO during late luteal phase. Since abnormal sensitivity to ALLO has previously been suggested to be one of the mechanisms of PMDD (Turkmen et al., 2011), we hypothesized that it may manifest as a change in GR frequency during the luteal phase.

Thirdly, we expected that abnormal neural inhibition in women with PMDD during their symptomatic luteal phase would affect visual perception functions that are highly sensitive to the strength of neural inhibition in the visual cortex. To test this

prediction, we assessed in our participants perceptual spatial suppression – the capacity of the visual system to suppress perception of large background-like motion (Tadin et al., 2019). Spatial suppression is reflected in an increase in the time it takes to discriminate the direction of motion of regularly patterned visual objects as their size increases (Tadin, 2015; Tadin et al., 2003). This perceptual phenomenon can be explained by an increase of surround inhibition associated with stimulation of the 'far surround' of the neurons' receptive fields (Angelucci et al., 2017; Nurminen & Angelucci, 2014). Indeed, elderly people (Betts et al., 2005, 2012) or individuals with disorders associated with weakened neural inhibition (Golomb et al., 2009; Linares et al., 2020; Tadin et al., 2006; Zhuang et al., 2017) demonstrated decreased spatial suppression.

Herein, to test these predictions, we investigated 1) visual MEG gamma oscillations and 2) perceptual spatial suppression in women with PMDD and women without severe PMS during the early-to-mid follicular and the mid-to-late luteal phases of the MC, when the PMS are least and most pronounced, respectively. For this purpose, we invited the same women twice, counterbalancing the phases of the first visit (follicular and luteal) between the participants.

Materials & methods

Participants

The participants from the PMDD group were recruited via media advertisements (social networks and publications in the media). The control women were recruited among members of the internet 'healthy life style' group and students. The criteria for exclusion, common for both groups, included (a) pregnancy or lactation, (b) irregular MC, (c) intake of hormones (contraceptive pills, thyroid hormones, etc.) or psychoactive drugs (nootropics, antidepressants, tranquilisers, etc.). Control women also had to have no personal or family history of neurological or psychiatric disorders. Women with PMDD who had been diagnosed with depression at some point in their lives (two of twenty women) were still included in the study because PMDD is often comorbid with this psychiatric condition (Cirillo et al., 2012; de Carvalho et al., 2018; Kim et al., 2004).

The potential subjects of the clinical group reported severe changes in mood associated with the MC, which had a significant impact on their lives, relationships, daily activities, and job. To test whether these symptoms met criteria for PMDD, we used Carolina Premenstrual Assessment Scoring System (C-PASS, Eisenlohr-Moul et

al., 2017). The C-PASS measures severity of PMS according to DSM-5 PMDD criteria (American Psychiatric Association, 2013). It includes 20 statements related to 11 symptoms, which should be rated from 1 ('not at all') to 6 ('extreme symptoms') according to their severity. All volunteers of the PMDD group were asked to fill in the C-PASS scales daily for two or three MCs. They were included in the study only if they met the C-PASS criteria for PMDD in at least two MCs. All but two PMDD subjects continued to fill in the C-PASS during the period when the MEG and psychophysical investigation were performed, which allowed us to assess the severity of their PMDD symptoms on the day of examination.

To estimate the symptom severity on the day of examination (hereafter 'same-day PMS scores'), we averaged scores obtained on this day for the symptoms that were reliably associated with PMDD according to the C-PASS (i.e., the symptoms which were strong enough and varied during the MC; see Eisenlohr-Moul et al., 2017). The same-day PMS scores were available in 18 out of 20 participants with PMDD.

To ensure that participants in the control group did not have severe PMS, they were asked to retrospectively assess their symptoms on a visual analogue premenstrual tension syndrome scale (PMTS-VAS, Steiner et al., 2011) for one week after the start of their last menstruation (early follicular phase) and for a week before their last menstruation (late luteal phase), when the PMS are expected to be least and most pronounced, respectively. PMTS-VAS consists of 12 statements which satisfy the DSM-4 criteria for PMDD and should be rated from 0 ('not at all') to 100 ('extreme symptoms') across a continuum of values. In each control participant, we estimated PMS score as a difference between PMTS-VAS scores for the luteal and follicular phase (Steiner et al., 2011; Steiner & Streiner, 2005). The resulting PMS score could thus vary from 0 (no PMS) to 100 (maximal symptom severity). In control subjects, the resulting PMS score averaged over 12 symptoms varied from 0 to 33.3 (mean ± standard deviation (S.D.): 16.59 ± 10.41), and PMS score averaged over core mood symptoms (depression, anxiety, irritability, and liability of mood) varied from 0 to 37.5 (mean ± S.D.: 18.53 ± 13.87), which is below the cut-off for severe PMS according to Steiner et al. (2005).

Neurophysiological (MEG) and behavioral (psychophysical) data were collected from 20 women who fulfilled criteria for PMDD (PMDD group) and 27 age-matched control women. PMDD and control subjects did not differ in age (Student's t-test, $t(45)=0.81$, $p=0.42$), average length of MC (Student's t-test, $t(45)=0.03$, $p=0.98$) or day of MC during the visit (follicular: Mann-Whitney $U=241.0$, $p=0.27$; luteal: Mann-Whitney $U=267.0$, $p=0.48$). The day of the MC normalized by the average MC length also did not differ between the groups (p's > 0.26). For PMDD participants, we also got

information about the day when their next menstruation started after their 'luteal phase visit'. Detailed information about the participants is summarized in Table 1.

Table 1. Characteristics of the experimental groups

Characteristic	PMDD group (N=20)	Control group (N=27)	Group difference	
			Student's t-test	*P*-value
	Mean (S.D.) [range]	Mean (S.D.) [range]		
Age (years)	29.05 (5.38) [18-39]	27.70 (5.82) [18-40]	0.81	0.42
Average reported length of MC	28.50 (2.74) [24-34]	28.48 (2.01) [25-33]	0.03	0.98
Day of MC, follicular phase visit	5.35 (2.56) [2-10]	4.81 (2.08) [2-10]	0.79	0.43
Day of MC, luteal phase visit	23.30 (2.79) [18-30]	23.33 (2.22) [20-29]	-0.05	0.96
Luteal phase visit: days before the next menstruation	5.60 (2.72) [1-10]			

N – number of subjects; S.D. - standard deviation; MC – menstrual cycle.

To test whether subjects from the two groups differed in their level of trait-like anxiety and level of anxiety during their visit to the laboratory, participants were asked to complete the State-Trait Anxiety Inventory (STAI) questionnaire (Spielberger, 1983) right before the MEG experiment. Besides, participants with PMDD filled in the screening tests for bipolar disorder (Bipolar Spectrum Diagnostic Scale, BSDS (Ghaemi et al., 2005); Mood Disorder Questionnaire, MDQ, (Hirschfeld et al., 2000); Hypomania Check List, HCL-32 (Vieta et al., 2007)), and for depressive disorder (Beck's Depression Inventory, BDI (Beck et al., 1988)). The bipolar disorder questionnaires were completed during the asymptomatic follicular phase, and the BDI was completed twice: during the follicular and symptomatic late luteal phases.

All the participants got blood tests for hormones and underwent MEG recording and psychophysical testing on the same day, once during the early-to-mid follicular (hereafter 'follicular') and once during mid-to-late luteal (hereafter 'luteal') phases of the MC. The phase of the 1st visit to the laboratory (follicular or luteal) was counterbalanced between participants. The local ethics committee approved the study and each subject gave written informed consent.

Blood samples

The blood samples used to confirm the phase of the MC and to analyze the effect of steroid hormone levels on visual gamma oscillations were collected in the morning. Plasma levels of estradiol and progesterone were measured using a solid phase chemiluminescence immunoassay. The analyzes were performed by INVITRO, a certified commercial laboratory that performs a wide range of medical tests for clinical and diagnostic purposes (https://www.invitro.ru).

MEG paradigm

Participants watched a sequence of large high-contrast circular gratings (18°, 100% contrast, spatial frequency 1.66 cycles per degree) that either drifted with one of three velocities ('slow': 1.2°/s, 'medium': 3.6°/s, 'fast': 6.0°/s) or remained static (see Fig. 1, upper panel). The presentation time for each stimulus ranged randomly from 1.2 to 3 seconds. After this period, the moving grating stopped or the static grating began to move. Participants were instructed to press a button as soon as this change occurred. Each new trial started with a fixed 1.2 s prestimulus interval, continued with the presentation of the stimulus, and ended immediately after the button press. To reduce fatigue and boredom, participants were shown short (3-6 s) cartoon animations after every 2-5 gratings. For each participant, 90 gratings of each type were randomized and presented in three sessions. For additional details, see Orekhova et al. (2018; 2019).

MRI data acquisition and processing

All participants underwent magnetic resonance imaging (MRI; 1.5 T, voxel size 1 mm x 1 mm x 1 mm). The T1-weighted images were processed with the FreeSurfer's (v.6.0.0) default 'recon-all' pipeline. Its main included steps were motion correction, spatial normalization, skull stripping, gray/white matter segmentation.

MEG data acquisition, preprocessing, and analysis

MEG data were acquired at the Center for Neurocognitive Research (MEG-center) of the Moscow State University of Psychology and Education using an Electa VectorView Neuromag 306-channel MEG detector array (Helsinki, Finland) consisting of 102 magnetometers and 204 planar gradiometers.

The MEG signal was registered with 0.03 Hz high-pass and 300 Hz low-pass in-built filters and sampled at 1000 Hz. Temporal signal-space separation method (tSSS) (Taulu & Simola, 2006), a temporal extension of SSS, with 0.9 correlation limit and with movement compensation implemented in MaxFilter software (v.2.2), was applied

to raw MEG signal in order to reduce interference of external artifact sources and compensate for movement-related distortions.

All subsequent processing steps were performed using MNE python software (v.0.22.0). Raw data were down-sampled to 500 Hz and independent component analysis (ICA) was applied in order to detect and remove ICs that corresponded to biological artifacts (eye blinks and heart beats). The number of removed ICs did not differ between the groups and/or phases of the MC (all p's>0.08). The data were then filtered using 30 Hz high-pass and 115 Hz low-pass finite impulse response filter with default parameters (symmetric, with 'humming' window; the window length and width of the transition band at the low/high cut-off was set to 'auto') and epoched from -1 s to 1.2 s relative to the stimulus onset. Epochs contaminated by instrumental and myogenic artifacts were excluded based on visual inspection of unfiltered epochs. The number of resulting artifact-free epochs did not differ significantly between the groups for any of the four velocity conditions and/or phases of the MC (all p's>0.19). Average number of artifact-free epochs for 'static', 'slow', 'medium', and 'fast' conditions in the control group was 83.41, 83.56, 83.11, 83.56 in the follicular phase vs. 81.78, 82.78, 81.62, 82.52 in the luteal phase; and in the PMDD group was 84.05, 84.25, 84.40, 83.40 in the follicular phase vs. 83.35, 83.55, 82.70, 82.85 in the luteal phase.

MEG source analysis

MEG data were co-registered with the structural MRI of the head, and a single-layer boundary element model was constructed. We then created surface-based source space with 4096 vertex sources in each hemisphere and estimated the forward solution. Noise covariance matrices and data covariance matrices were estimated based on the prestimulus (-1 – 0 s, relative to the stimulus onset) and stimulation (0 – 1.2 s, relative to the stimulus onset) time intervals, respectively, of all conditions combined. Linearly constrained minimum variance (LCMV; Van Veen et al., 1997) beamformer spatial filters with a source orientation that maximizes power and a regularization coefficient 0.05 were created separately for each stimulus type and were applied individually to each epoch. As we expected to observe gamma oscillations in the visual cortex (Hoogenboom et al., 2006; Orekhova et al., 2018, 2019; Orekhova, Prokofyev, et al., 2020; Orekhova, Rostovtseva, et al., 2020), the inverse solution was limited to visual and adjacent cortical areas, same as in Manyukhina et al. (2021).

Time-frequency analysis of the MEG data at the source level

The approach to the analysis of visual gamma parameters used in the present study is described in detail by Manyukhina et al. (2021). The time-frequency analysis was

performed for the signal at each vertex source using multi-taper method, separately for the four velocity conditions and for the prestimulus and stimulation time intervals. The following parameters were used: bandwidth = 10 Hz, frequency resolution ~ 1.11 Hz, time step = 2 ms. Then, at each vertex source we estimated the normalized response power as (Stim – PreStim)/PreStim, where PreStim and Stim are the spectral powers estimated at -0.9 – 0 s and 0.3 – 1.2 s time intervals relative to the stimulus onset, respectively. Among all the four conditions, we then selected 26 vertices with maximum response power in gamma range (40-80 Hz), and averaged the spectra over these vertices, separately for each condition. See Figure 1 (upper panel) for a brief review of MEG data analysis steps.

For each velocity condition, the GR power was calculated as the average of those spectrum values that exceeded 2/3 of the peak value in the frequency range 35-90 Hz. The GR frequency was estimated as a center of gravity of the spectrum values used to calculate the GR power. The GR was considered reliable if, according to Wilcoxon signed-rank test, the probability of no difference in the frequency corresponding to the peak gamma power in prestimulus and stimulation time intervals was below p=0.0001 (see, e.g., Orekhova et al., 2018; Orekhova, Rostovtseva, et al., 2020). In all our participants and velocity conditions, the GRs met this reliability criterion.

Psychophysical perceptual spatial suppression test

To assess perceptual suppression associated with increasing stimulus size, we used the spatial suppression paradigm (Tadin et al., 2003, 2006), a modification detailed in our previous study (Orekhova, Rostovtseva, et al., 2020).

Visual stimuli were presented using PsychoToolbox for Matlab (MathWorks). Subjects sat at 60 cm distance from the monitor (Benq XL2420T, 24″W LED, 1920 × 1080 resolution, 120 Hz). Before the testing, all participants completed a training session.

The stimuli were 12°, 2.5°, and 1° vertical high-contrast sinusoidal gratings moving at a constant rate of 4°/s in each trial, randomly to the right or left (Fig. 1, lower panel). For the purpose of the present study, only responses to the large (12°) and small (1°) stimuli were analyzed. The participants were asked to indicate the direction of the visual motion by pressing the corresponding (right or left) button. No response time limit was given. The stimulus presentation time started from 150 ms and was adjusted in the following trials using 'one-up two-down' staircases procedure with the initial step of 16.7 ms that decreased to 8.3 ms after the first two reversals. Separate staircases were used for the three types of the stimuli. The block continued until all staircases completed at least 7 reversals. The subject completed two blocks

during each visit. The duration threshold was computed by averaging the presentation times over all the reversals excluding the first two, and then over the two blocks. As a result, for each visit and stimulus size we estimated the minimal exposure time required for the subject to discriminate the direction of motion of the small and large grating (in each visit: $\text{Threshold}_{\text{SMALL}}$ and $\text{Threshold}_{\text{LARGE}}$).

To estimate strength of perceptual suppression, we calculated spatial suppression index (SSI) as:

$$SSI = \log10(\text{Threshold}_{\text{LARGE}}) - \log10(\text{Threshold}_{\text{SMALL}}).$$

For a brief review of all the steps of psychophysical data analysis, see Figure 1 (lower panel).

Figure 1. Experimental procedure and analyses of MEG (upper panel) and psychophysical (lower panel) data. The procedures were identical between the two visits.

Statistical analyses

To analyze group and phase-related differences in gamma parameters, we used General Linear Models (GLM) implemented in the Rstatix and Stats packages in R 4.0.3 (R Core Team, 2013). GR power values were log10-transformed to normalize the distributions. According to the Shapiro-Wilk test, distributions of the GR frequency and GR log10-transformed power did not differ significantly from normal (all p's>0.05). Since the preliminary analysis showed the GR power and frequency depend on age (see below), standardized Age was included as a factor in the linear models for GR

frequency and log10-transformed GR power. The between-group factors were Group (control, PMDD), Visit-Order (1st visit during the follicular phase, 1st visit during the luteal phase). The repeated measures factors were Phase (follicular, luteal) and Velocity (4 levels). We tested for the effects of Group, Visit-Order, Group x Visit-Order, Age, and their interactions with repeated measures factors Phase and Velocity. When appropriate, the Greenhouse-Geisser correction was used to adjust for the lack of sphericity.

Other steps of statistical analysis were performed using the standard Python 3.8.5 libraries Numpy (Harris et al., 2020) and Scipy (Virtanen et al., 2020). The data were tested for normality using Shapiro-Wilk test, homogeneity of variance was evaluated using Levene's test. When assumptions for parametric testing were violated, non-parametric tests were implemented, i.e., Wilcoxon signed-rank test for within- and Mann-Whitney U test for between-samples comparisons. Alternatively, parametric Student's t-test for related or independent groups was used.

Pearson correlation coefficient was estimated to assess the relationship between the normally distributed variables. When assumptions of normality were violated, Spearman's rank correlation coefficient was estimated. Partial Pearson's and Spearman's rank correlation coefficients were used wherever necessary to control for confounding variables.

3. Results

3.1. Premenstrual symptom severity in women with PMDD

Figure 2 shows the daily symptom severity scores in women with PMDD, averaged over two or three MCs. For the subjects to meet criteria for PMDD on the C-PASS, at least five out of 20 symptoms need to meet the PMDD criteria, i.e., the symptoms need to be strong enough and be reliably associated with the MC. The curves in Figure 2 represent averages over all 20 symptoms. Despite this rough average, one can see that symptom severity substantially increased during the week that precedes onset of menstruation (luteal phase; the score >=3 of maximum 6) and then gradually declined in the early follicular phase. According to a recent study of the PMDD subtypes (Eisenlohr-Moul et al., 2020), these average symptom scores indicate severe rather than mild PMDD in most participants in the clinical group.

Figure 2. Daily symptom severity scores (from 1 = 'Not at all' to 6 = 'Extreme') averaged over 20 symptoms (C-PASS questionnaire (Eisenlohr-Moul et al., 2017)) and two or three menstrual cycles in participants with PMDD. The thin lines show individual participants; the thick line is the group average. The days are numbered relative to the start of menstruation (day 1).

3.2. Psychometric characteristics of women with PMDD

3.2.1. Bipolar and depressive disorders questionnaires

The questionnaire scores for the PMDD group are summarized in Table 2. According to BSDS, 55% (11 out of 20) of PMDD subjects screened positive for bipolar disorder (Ghaemi et al., 2005). However, none of PMDD subjects met criteria for bipolar disorder according to MDQ, as none indicated 'moderate' or 'serious' problems related to the described symptoms (Hirschfeld et al., 2000). In addition, according to HCL-32, 13 PMDD subjects (65%) demonstrated hypomanic features. In general, this result is consistent with the high prevalence of bipolar disorder in women with PMDD (de Carvalho et al., 2018).

It has been previously demonstrated that about 23% of women with PMDD suffer from concurrent major depressive disorder (de Carvalho et al., 2018). BDI scores in our PMDD sample are consistent with this finding: when assessed during the asymatomatic period (follicular phase), 30% (6 out of 20) of PMDD subjects had mild-to-moderate depression (Beck et al., 1988). In contrast, during the premenstrual

period (luteal phase), 90% (18 out of 20) of women with PMDD were rated as having mild-to-moderate or more severe depression according to BDI.

Table 2. Questionnaire scores in the PMDD and control groups

Scales	PMDD Mean (S.D.) [range]	Control Mean (S.D.) [range]	Group difference Mann-Whitney U test	P-value
STAI, trait $N_{PMDD}/N_{Control}$ = 20/27	47.23 (10.04) [30-72]	39.83 (9.37) [27-62]	163.5	0.011
STAI, state (follicular) $N_{PMDD}/N_{Control}$ = 20/26	37.85 (8.09) [25-53]	34.92 (10.83) [22-62]	234.0	0.22
STAI, state (luteal) $N_{PMDD}/N_{Control}$ = 20/27	47.6 (11.56) [26-71]	34.33 (9.46) [20-56]	105.5	0.0002
BDI (follicular) N_{PMDD} = 20	8.35 (7.42) [2-29]			
BDI (luteal) N_{PMDD} = 20	18.25 (9.69) [4-44]			
BSDS N_{PMDD} = 20	12.35 (5.43) [2-24]			
MDQ N_{PMDD} = 20	6.50 (3.58) [0-11]			
HCL-32 N_{PMDD} = 20	14.8 (6.88) [1-24]			

N – number of subjects; S.D. – standard deviation; STAI – State-Trait Anxiety Inventory; BSDS – Bipolar Spectrum Diagnostic Scale; MDQ – Mood Disorder Questionnaire; HCL-32 – Hypomania Check List; BDI – Beck's Depression Inventory

3.2.2. State and trait anxiety during the MC

According to the STAI questionnaire, the 'trait anxiety' was higher in women with PMDD than in control women (Mann-Whitney U test, p=0.011, Table 2). The 'state anxiety' was higher in the PMDD than control group during the luteal phase (Mann-Whitney U test, p=0.0002, Table 2), while no group differences were found during the follicular phase (Mann-Whitney U test, p=0.22, Table 2). Also, women with PMDD had higher situational anxiety when they visited the laboratory during the luteal compared

to the follicular phase (Wilcoxon signed-rank test, Z=40.0, p=0.014), which was not the case in control women.

We expected our participants to be more anxious during their first visit to the laboratory, irrespective of the MC phase. Indeed, in the combined group of subjects, state anxiety was higher during the first visit compared to the second (Mann-Whitney U=319.5, p=0.025). In the individual groups, however, this comparison did not reach the level of significance (both p's>0.12).

3.3. Plasma levels of steroids and PMDD symptom severity

In all participants, plasma level of progesterone increased from the follicular to the luteal phase (Fig. 3a,b). Estradiol plasma level also increased from the follicular to the luteal phase in the majority of participants (Fig. 3c,d). Thus, the hormone levels were in the range consistent with the follicular and luteal phases at each visit.

Figure 3. (a-d) Plasma levels of estradiol and progesterone in the control **(a, c)** and PMDD **(b, d)** groups in two phases of the menstrual cycle (follicular, luteal). **(e-g)** Violin plots show estradiol and progesterone concentrations and estradiol to progesterone ratios in the luteal phase of the menstrual cycle in control subjects and subjects with PMDD.

*p<0.05; **p<0.01

The luteal progesterone plasma level was reduced in women with PMDD (Student's t-test, t(45)=-2.53, p=0.015; Fig. 3f), while their luteal estradiol did not differ from that in the control group (Student's t-test, t(45)=0.65, p=0.52; Fig. 3e). The ratio of estradiol to progesterone concentrations in the luteal phase was significantly elevated in the PMDD group (Mann-Whitney U=150.0, p=0.005; Fig. 3g).

To test for the link between steroid hormone levels and symptom severity in PMDD, we estimated Pearson correlation between the hormone concentrations and the same-day PMS scores (number of days prior menstruation onset was partial out of the correlation). The correlation was not significant (N_{PMDD}=18; estradiol: $r_{partial}$=0.32, p=0.22; progesterone: $r_{partial}$=0.25, p=0.33).

3.4. MEG results

3.4.1. Behavioral performance in MEG task

The percentage of epochs with omission (reaction time<150 ms) and commission (no response or reaction time>=1000 ms) errors did not differ significantly between the groups and MC phases (all p's>0.19). The reaction time was significantly longer in women with PMDD than in control participants during the luteal phase of the MC (Mann-Whitney U=1995, p=0.035), but not during the follicular phase (Mann-Whitney U=2252.0, p=0.23).

3.4.2. Dependence of the GR parameters on age

In line with the previous studies (Gaetz et al., 2012; Muthukumaraswamy et al., 2010; Orekhova et al., 2015), GR frequency, averaged over phases, was negatively correlated with age in control women (Table 3). In women with PMDD, none of the correlations reached the level of significance. However, there was no significant difference in the correlation coefficients between the groups (all p's>0.15, two-sided).

Phase-averaged GR power increased with age in both groups of participants and in all velocity conditions (Table 3).

Table 3. Pearson's correlations between Age and gamma response (GR) power (log10-transformed) and frequency in the two groups of participants (control and PMDD). Gamma parameters were averaged over the two phases of the menstrual cycle.

Grating's motion velocity	Control group (N=27)	PMDD group (N=20)
	GR power	
Static, 0 °/s	**r=0.62, p=0.001**	r=0.40, p=0.08
Slow, 1.2 °/s	**r=0.56, p=0.002**	**r=0.46, p=0.04**
Medium, 3.6 °/s	**r=0.65, p=0.001**	**r=0.49, p=0.03**
Fast, 6.0 °/s	**r=0.66, p=0.001**	**r=0.54, p=0.01**
	GR frequency	
Static, 0 °/s	r=-0.33, p=0.097	r=-0.06, p=0.80
Slow, 1.2 °/s	**r=-0.38, p=0.049**	r=-0.29, p=0.21
Medium, 3.6 °/s	**r=-0.47, p=0.012**	r=-0.23, p=0.33
Fast, 6.0 °/s	r=-0.34, p=0.080	r=0.07, p=0.77

N - number of subjects; significant correlations are highlighted in bold.

3.4.3 Effects of diagnosis, MC phase, and visit order on GR frequency

Results of GLM with factors Group (control, PMDD), Visit-Order (1st visit during the follicular phase, 1st visit during the luteal phase), repeated measures factors Phase (follicular, luteal) and Velocity (4 levels), and Age as a covariate are presented in Tables 4a and 4b for GR frequency and GR power, respectively.

Table 4a. ANCOVA results for GR frequency

Effect	*F*	*df*	*G-G epsilon*	*Generaliz ed eta squared*	*P / adj. p*
Group	0.159	1, 42		0.003	0.69
Visit-Order	0.142	1, 42		0.003	0.71
Group * Visit-Order	0.520	1, 42		0.009	0.47

	F	df	ε	p	
Phase	**8.259**	**1, 42**		**0.004**	**0.006**
Phase * Group	0.000	1, 42		$3.7*10^{-8}$	0.99
Phase * Visit-Order	**17.968**	**1, 42**		**0.009**	**0.0001**
Phase * Group * Visit-Order	0.105	1, 42		0.00005	0.75
Velocity	**410.088**	**3, 126**	0.61	**0.63**	**$2.5*10^{-40}$**
Velocity * Group	0.185	3, 126	0.61	0.0008	0.81
Velocity * Visit-Order	0.859	3, 126	0.61	0.004	0.42
Velocity * Group * Visit-Order	0.747	3, 126	0.61	0.003	0.47
Phase * Velocity	0.347	3, 126	0.58	0.0003	0.68
Phase * Velocity * Group	0.078	3, 126	0.58	0.00006	0.90
Phase * Velocity * Visit-Order	2.845	3, 126	0.58	0.002	0.072
Phase * Velocity * Group * Visit-Order	0.248	3, 126	0.58	0.0002	0.75

Significant effects are highlighted in bold.

There was a highly significant effect of *Velocity*: GR frequency almost linearly increased with increasing grating's motion velocity (Table 4a, Fig. 4a,b). There was also a significant effect of *Phase* (luteal>follicular), which however can be better understood by inspecting the highly significant interaction between *Phase* and *Visit-Order* ($F_{(1,42)}=18.0$, p=0.0001). This result shows that the difference in GR frequency between the luteal and follicular phases was present only if the luteal phase occurred during the first visit to the MEG laboratory (Fig. 4c,d, right panels), but not during the second visit (Fig. 4c,d, left panels). Notably, the effect of *Phase* and *Visit-Order* interaction – higher GR frequency during the luteal phase if it coincided with the first visit – was the same in both PMDD and control groups (Fig. 4c,d). Indeed, the *Phase* and *Visit-Order* interaction remained highly significant when tested separately in the control and PMDD groups (Control: $F_{(1,24)}=16.3$, p=0.0005; PMDD: $F_{(1,17)}=11.5$, p=0.003).

The effect of *Group* and interactions of *Group* with *Phase*, *Visit-Order*, or *Velocity* were all not significant.

Figure 4. Effect of the phase of the menstrual cycle on the frequency of gamma response (GR) in four motion velocity conditions ('static': 0°/s, 'slow': 1.2°/s, 'medium': 3.6°/s, 'fast': 6.0°/s). **(a)** and **(b)** show comparison of the GR frequency in the luteal and follicular phases separately for all control **(a)** and all PMDD **(b)** subjects. **(c, d)** show comparison of the GR frequency in the luteal and follicular phases separately for those subjects, who came for the first investigation during their follicular **(c, d left panel)** or luteal **(c, d right panel)** phase. Error bars represent 95% confidence intervals.

*p<0.05; **p<0.01

3.4.4. Effects of diagnosis and MC phase on GR power

The group average GR power spectra for participants from the control and PMDD groups are shown in Figure 5.

Figure 5. Grand average spectra of gamma response (GR) ([stimulation-prestimulus]/prestimulus) in four motion velocity conditions ('static': 0°/s, 'slow': 1.2°/s, 'medium': 3.6°/s, 'fast': 6.0°/s), two phases of the menstrual cycle (follicular, luteal), and two groups of subjects (control **(a)**, PMDD **(b)**). Shaded areas represent 95% confidence intervals.

Table 4b. ANCOVA results for log10-transformed GR power

Effect	F	df	G-G epsilon	Generalized eta squared	P / adj. p
Group	0.833	1, 42		0.014	0.37
Visit-Order	0.608	1, 42		0.010	0.44
Group * Visit-Order	3.502	1, 42		0.055	0.068
Phase	0.070	1, 42		0.0002	0.79
Phase * Group	0.155	1, 42		0.0004	0.70
Phase * Visit-Order	0.909	1, 42		0.002	0.35
Phase * Group * Visit-Order	2.714	1, 42		0.007	0.11
Velocity	**152.058**	**3, 126**	**0.71**	**0.375**	**3.5*10⁻³⁰**
Velocity * Group	**3.464**	**3, 126**	**0.71**	**0.013**	**0.033**
Velocity * Visit-Order	1.661	3, 126	0.71	0.007	0.19
Velocity * Group * Visit-Order	0.618	3, 126	0.71	0.007	0.55
Phase * Velocity	1.615	3, 126	0.76	0.0008	0.20
Phase * Velocity * Group	2.441	3, 126	0.76	0.001	0.085
Phase * Velocity * Visit-Order	**3.020**	**3, 126**	**0.76**	**0.001**	**0.047**
Phase * Velocity * Group * Visit-Order	0.317	3, 126	0.76	0.0002	0.76

Significant effects are highlighted in bold.

For GR power, there was a highly significant effect of *Velocity* (Table 4b): the power increased from the 'static' to the 'slow' condition and then decreased with further increase of grating's drift rate (Fig. 5). There were no main effects of *Phase* or *Group*. However, there was a significant *Velocity* and *Group* interaction ($F(3, 126)=3.5$, epsilon = 0.71, adj. p=0.033). There was also a tendency for *Phase * Velocity * Group* interaction ($F(3, 126)=2.4$, epsilon = 0.76, adj. p=0.085). Since we had a clear prediction of a reduction of GR suppression in women with PMDD during the luteal phase and of the association between this reduction and PMS severity, we further analyzed this prediction even though the *Phase * Velocity * Group* interaction was only a trend.

3.4.5. Modulations of GR power by visual motion velocity

To investigate group differences in velocity-related modulations of GR power, we used post-hoc planned comparisons. There were no group differences in GR powers in any MC phase or velocity condition (all p's>0.1). However, suppression of the GR power from the 'slow' to the 'medium' condition was lower in women with PMDD than in control women during the luteal phase ($F(1, 42)=5.1$, p=0.03; Fig. 6a, right panel). This group difference was absent during the follicular phase ($F(1, 42)=0.01$, p=0.9; Fig. 6a, left panel). Figure 6c shows this *GR suppression* in percent of GR power in the 'slow' condition, which induced maximal power of GR at the group level; *GR suppression* = (1 - GR Power$_{Medium}$/GR Power$_{Slow}$)*100%. Larger values indicate stronger GR power decrease under higher sensory load.

   The GR suppression increased from the follicular to the luteal phase in control women (Student's t-test, $t(26)=2.2$, p=0.041; Fig. 6c), but not in women with PMDD (Student's t-test, $t(19)=-1.5$, p=0.16; Fig. 6c), in which the direction of change was even inverted. As a result, in the luteal phase women with PMDD had lower GR suppression than control participants (Student's t-test, $t(45)=-2.5$, p=0.017; Fig. 6c). The lower luteal GR suppression in women with PMDD correlated with higher severity of their same-day PMS scores ($N_{PMDD}=18$, Pearson's r=-0.49, p=0.04; Fig. 6d).

   We then tested for the presence of group differences in *GR facilitation*, i.e., an increase of GR power from the 'static' to the 'slow' condition, which likely has a different mechanism from that of GR suppression (see Orekhova, Prokofyev et al., 2020 and Discussion section). The increase of GR power from the 'static' to the 'slow' condition was steeper in PMDD than in control participants in both MC phases (follicular: $F(1, 43)=4.2$, p=0.047; luteal: $F(1, 43)=7.0$, p=0.01; combined phases: F(1,

43)=6.1, p=0.02; Fig. 6a). This means that women with PMDD show a stronger facilitation of GR power from the 'static' to the 'slow' condition regardless of the MC phase. Figure 6b presents GR facilitation in percent of GR power in the 'slow' condition, which induced maximal power of GR at the group level: *GR facilitation* = (1 - GR Power$_{Static}$/GR Power$_{Slow}$)*100%. Larger values indicate stronger GR facilitation with a moderate increase of sensory load. Similarly to the planned comparison analysis, this measure indicates greater GR facilitation in PMDD than in control subjects during both follicular (Student's t-test, t(45)=2.2, p=0.034) and luteal (Student's t-test, t(45)=2.9, p=0.006) phases of the MC (Fig. 6b).

Figure 6. Group differences in gamma response (GR) power. **(a)** Log10-transformed GR power modulation in four velocity conditions ('static': 0°/s, 'slow': 1.2°/s, 'medium': 3.6°/s, 'fast': 6.0°/s) in the follicular (left panel) and luteal (right panel) phases of the menstrual cycle separately in control and PMDD subjects. **(b)** Comparison of velocity-related GR power facilitation from the 'static' to the 'slow' velocity condition, estimated in percent of the GR power in the 'slow' condition, which induced maximal GR at the group level: (1 - GR Power$_{Static}$/GR Power$_{Slow}$)*100%. **(c)** Comparison of velocity-related GR power suppression from the 'slow' to the 'medium' velocity condition, estimated in percent of the GR power in the 'slow' condition: (1 - GR Power$_{Medium}$/GR Power$_{Slow}$)*100%. **(d)** Correlation between luteal GR power suppression

from the 'slow' to the 'medium' condition and premenstrual symptom (PMS) severity on the day of investigation in PMDD subjects. Error bars and shaded areas represent 95% confidence intervals.

To sum up, we found that the modulation of GR power by velocity differed in women with PMDD and control participants. GR facilitation from the 'static' to the 'slow' condition (i.e., on the ascending branch of the GR power modulation curve) was greater in women with PMDD than in control women during both phases of the MC. On the other hand, GR suppression from the 'slow' to the 'medium' velocity condition (i.e., at the descending branch of the GR power modulation curve) was reduced in PMDD participants specifically during the luteal phase, and correlated with the severity of PMS on the day of the visit.

3.4.6. Relationship between GR power and GR frequency modulations

We have previously reported a significant correlation between an increase in GR frequency and suppression of GR power from the 'slow' to the 'medium' visual motion velocity condition (Orekhova et al., 2018).

Consistently with the previous findings, we found in control participants a correlation between the 'slow'/'medium' GR power ratio and an increase in GR frequency from the 'slow' to the 'medium' condition ($N_{Control}$=27, follicular: Pearson's r=0.39, p=0.04; luteal: Pearson's r=0.56, p=0.002). This correlation was absent in the PMDD sample (N_{PMDD}=20, follicular: Pearson's r=-0.15, p=0.5; luteal: Pearson's r=-0.08, p=0.7). The between-group difference in correlation coefficients was significant in the luteal phase (p=0.03) and approached significance level in the follicular phase (p=0.09).

3.4.7. Relationship between GR parameters and plasma levels of steroid hormones

To test for the link between plasma levels of steroid hormones and GR parameters, we estimated partial Spearman's correlation coefficients between GR frequency or GR power in the corresponding phase of the MC and (A) follicular estradiol, (B) luteal estradiol, or (C) luteal progesterone separately in the control and PMDD groups (see Supplementary table S1). Age was partialled out of the correlations because it affected

the GR parameters (see Table 3). Correlations with follicular progesterone were not assessed because the poor sensitivity of commercial immunoassay systems to low progesterone concentrations (<5 nmol/L) precludes its reliable measurement (Shankara-Narayana et al., 2016). In control participants, there were positive correlations between follicular estradiol and GR frequency in the 'medium' and 'fast' velocity conditions. None of these correlations, however, survived correction for multiple comparisons.

3.5. Results of the psychophysical experiment: directional sensitivity to motion

Spatial suppression was estimated in 19 of 20 subjects with PMDD and in 26 of 27 control subjects. Two women (1 control, in both phases; 1 PMDD, in the follicular phase) demonstrated a persistent illusion of reverse motion when presented with large moving grating, which did not allow to reliably estimate motion direction discrimination thresholds (hereafter, 'duration thresholds'), in these participants.

In both groups and during both phases of the MC, subjects demonstrated reliable perceptual spatial suppression: it took them longer to discriminate direction of motion of a large than of a small visual grating (all F's>9.0, p's<0.0001; Fig. 7a). Based on duration thresholds, we calculated the spatial suppression index (SSI; SSI = $log10(Threshold_{SMALL}/Threshold_{LARGE})$) and tested for the *Group* and *Phase* differences in spatial suppression using repeated measures ANOVA. This analysis revealed the main effect of *Phase* (F(1,43)=4.37, p=0.043) and *Group * Phase* interaction (F(1,43)=4.10, p=0.049), which is illustrated in Figure 7b. In the PMDD group, SSI decreased from the follicular to the luteal phase (Student's t-test, t(18)=2.49, p=0.022). Importantly, this decrease was mainly due to an *improvement* in discriminating the direction of motion for the *large* stimulus during the luteal phase (mean follicular Threshold_{LARGE}=127 ms, mean luteal Threshold_{LARGE}=110 ms; Student's t-test, t(18)=2.02, p=0.06), rather than a change in the duration threshold for the small one (mean follicular Threshold_{SMALL}=39 ms, mean luteal Threshold_{SMALL}=44 ms; Student's t-test, t(18)=1.3, p=0.19). There was also a tendency for stronger spatial suppression in women with PMDD than in control women during the follicular phase (Student's t-test, t(18)=1.8, p=0.08; Fig. 7b).

Figure 7. Menstrual cycle effect (follicular, luteal phase) on the results of the motion direction discrimination task in the two groups of participants (control, PMDD): **(a)** duration thresholds for the small and large stimuli **(b)** and spatial suppression index (SSI). Error bars represent 95% confidence intervals.

$*p < 0.05$; $**p < 10^{-8}$

3.6. Relationship between perceptual spatial suppression and GR power suppression

In control women, the percent of GR suppression from the 'slow' to the 'medium' condition (Fig. 6c) correlated with the SSI ($N_{Control}=26$; follicular Pearson's r=0.53, p=0.005; luteal Pearson's r=0.31, p=0.12). This result reproduces that obtained in our previous study in a smaller sample of neurotypical women (13 of them were included in the control sample in this study) where we estimated GR suppression as a 'gamma suppression slope' based on tree velocity conditions (Orekhova, Rostovtseva, et al., 2020; for the same analysis of the present data see the Supplementary materials). In the PMDD group, this correlation was absent ($N_{PMDD}=19$; follicular Pearson's r=-0.12, p=0.6; luteal Pearson's r=-0.25, p=0.3).

4. Discussion

We hypothesized that in women with PMDD, the paradoxical response to neurosteroids, which results in changes of neural E/I balance during the luteal phase of the MC, would affect parameters of visual gamma oscillations and visual perception

that are highly dependent on E/I balance, and that these abnormalities would be specific to the symptomatic luteal phase. To test this hypothesis, in women with and without PMDD we measured visual gamma responses (GRs) to drifting gratings with MEG and, in a separate psychophysical experiment, estimated perceptual spatial suppression. By using drift rate of a visual grating as a proxy of excitatory drive and changing it from 'static' to 'slow' and further to 'medium' and 'fast' (Orekhova, Prokofyev, et al., 2020), we probed the dynamics of gamma generating circuitry response, which strongly depends on the neural E/I balance.

Our predictions were only partially confirmed. The only parameter that distinguished women with PMDD from women with the healthy MC specifically during the luteal phase was the attenuated suppression of GR from the 'slow' condition optimal for GR generation to the 'medium' one. This attenuation indicates an abnormally decreased inhibition during the luteal phase in women with PMDD compared to control women. Moreover, the degree of GR suppression during the luteal phase in women with PMDD correlated with the severity of clinical symptoms assessed on the same day. Another characteristic feature of the PMDD group was an atypically strong facilitation of GR power from the 'suboptimal' for gamma generation 'static' to the optimal 'slow' stimulus. This excessive GR facilitation, however, was observed in both MC phases, indicating a constitutively elevated E/I ratio in the visual cortex in women with PMDD regardless of the effect of neurosteroid hormones.

Contrary to our expectations, neither GR frequency nor perceptual spatial suppression were affected in women with PMDD. In the discussion that follows, we will argue that in women with PMDD, neural excitability in the visual cortex is constitutively elevated and that the E/I balance shifts even more toward excitability during the luteal phase, possibly due to tolerance to ALLO.

4.1. Representativeness of the PMDD sample

It has been suggested that the often inconsistent results in PMDD may be explained, at least in part, by inconsistent diagnostic practices and varying symptom severity in patients (Amiel Castro et al., 2021; Eisenlohr-Moul et al., 2017; Munday et al., 1981). We therefore start the discussion with the characterization of our PMDD sample.

To confirm the presence of PMDD and assess its severity, we used a questionnaire in which participants reported their symptoms daily for two or three MCs. This approach provides a more reliable assessment of PMDD than questionnaires based on retrospective PMS reports (Bosman et al., 2018; Eisenlohr-Moul et al., 2017). Consistent with the previously described PMDD symptomatology (Eisenlohr-Moul et al.,

2017; Hammarbäck et al., 1989; Seippel & Bäckström, 1998), the severity of self-reported symptoms in participants of the clinical group increased from the ovulatory to the late luteal phase and decreased after the onset of menstruation (Fig. 2). In a similar vein, according to the STAI and BDI questionnaires, their state anxiety and depressive symptoms, which are diagnostic features of PMDD (American Psychiatric Association, 2013), also exacerbated during the luteal phase. The state anxiety scores of women with PMDD were higher than those of control participants specifically during the luteal phase (Table 2). In addition to MC-dependent mood changes, high scores on trait anxiety, depression, and bipolar disorder questionnaires (according to their cut-off scores) in the PMDD sample agree well with previous reports of a high frequency of respective comorbid disorders in this clinical group (de Carvalho et al., 2018).

A comparison of our participants' daily PMS scores with those in the Eisenlohr-Moul et al's study suggests that the majority of participants in our clinical sample had severe rather than mild form of PMDD (compare Figure 2 in the present paper and Figure 1 in (Eisenlohr-Moul et al., 2020). Abnormal luteal hormone levels in women with PMDD compared with age-matched control participants are also consistent with the presence of severe PMDD in the clinical group. Indeed, there is evidence that decreased luteal progesterone, together with an elevated luteal estradiol/progesterone ratio in the mid-luteal phase of the MC (Fig. 3f,g) characterize women with severe rather than mild PMS (Munday et al., 1981).

Thus, we discuss below the results obtained in a sample of women with severe PMDD and age-matched control women.

4.2. Altered modulation of MEG visual GR by excitatory drive in PMDD

Phase-dependent changes in gamma suppression in control women and in those with PMDD

In control women, changes in the visual GR power caused by the increase of excitatory drive were affected by the MC phase on the descending branch of the GR power modulation curve: during transition from the 'slow' to the 'medium' velocity (Fig. 6c). The suppression of GR with an increase in excitatory drive from the 'slow' to the 'medium' and then to the 'fast' drift rate most likely reflects desynchronization of gamma oscillations as a result of compensatory 'excessive' activation of inhibitory neurons (Adesnik, 2017; Börgers & Kopell, 2005; Pavlov et al., 2014; see Orekhova, Prokofyev et al. 2020 for a detailed discussion). Indeed, animal studies have shown that increasing excitatory drive to the visual cortex above a certain limit leads to a greater increase in the inhibitory than excitatory current on the principal neurons, so

the overall E/I ratio shifts toward inhibition (Adesnik, 2017). Hence, a steeper slope of GR suppression at stimulation intensities exceeding the 'optimal' one for gamma generation may indicate a transition to a stronger inhibition of the principal neurons.

Despite the presence of phase-related changes in GR suppression in women with a healthy MC, their absolute GR power did not differ in the follicular to the luteal phases. The later result is consistent with that of Sumner et al. (2018), and underscores the functional relevance of velocity-related *modulation* of power of gamma oscillations as a measure of the E/I ratio in the visual cortex. Considering the presumed role of strong inhibition in GR suppression, the increase in GR suppression in the luteal phase compared to the follicular phase of the MC in control women is consistent with enhanced inhibitory transmission in the luteal phase. The increase in inhibition associated with the luteal phase may result from potentiation of the tonic $GABA_A$ conductance in principal neurons by ALLO, the neuroactive metabolite of progesterone. ALLO concentration follows that of progesterone (Timby et al., 2006) and is elevated during the luteal phase. ALLO targets the $GABA_ARs$ family (MacKenzie & Maguire, 2014), but especially strongly potentiates $\delta GABA_ARs$ ($\delta GABA_ARs$) (Bäckström et al., 2014; Hantsoo & Epperson, 2020; Reddy, 2010). These receptors are located extrasynaptically and produce a tonic form of inhibition (Reddy, 2010), which plays an important role in controlling neuronal excitability (Stell et al., 2003) and gamma generation (Héja et al., 2012; Pavlov et al., 2014). Furthermore, the elevated concentration of ALLO during the luteal phase affects $GABA_ARs$ expression pattern: it potentiates the expression of $\delta GABA_ARs$ and therefore further augments the effect ALLO on tonic inhibitory neurotransmission (Maguire et al., 2005; X. Wu et al., 2013).

The putative enhancement of inhibitory neurotransmission from the follicular (i.e., low ALLO) to the luteal (i.e., high ALLO) phase of the MC is consistent with studies in rodents that demonstrate a potentiating effect of physiological levels of ALLO on inhibitory neurotransmission during the estrous cycle (Devall et al., 2015; Maguire et al., 2005; X. Wu et al., 2013). Evidence in favor of increased inhibitory tone during high-ALLO mid-luteal phase also comes from TMS studies in the motor cortex of naturally cycling women (Smith et al., 1999, 2002, 2003) and from reports of decreased seizure susceptibility during this phase of the MC (Finocchi & Ferrari, 2011). However, the effect of ALLO on net excitability may vary between brain regions and species (Engin & Treit, 2007; Lovick et al., 2005; Rossetti et al., 2016). As for the human visual cortex, the results by Epperson et al. (2002) are particularly relevant to our finding. In a magnetic resonance spectroscopy study, these authors showed a significant decrease of GABA concentration in the occipital cortex from the follicular to the mid-luteal phase of the healthy MC and interpreted this result in terms of

homeostatic regulation, which partially compensates for the increased sensitivity of GABA$_A$Rs to ALLO by depressing GABA synthesis.

Given the scarce and inconsistent literature on the role of neurosteroid hormones in the human visual cortex (Schloemer et al., 2020; Sy et al., 2016), our results pointing to increased inhibitory tone in gamma-generating visual circuitry during the luteal phase in healthy MC are particularly important. They strengthen the evidence linking the physiological effect of ALLO with increased tonic inhibition in the visual cortex of healthy naturally cycling women.

In contrast to control participants, women with PMDD did not show an increase in GR suppression from the follicular to the luteal phase of the MC (Fig. 6c). The absence of this normal change in the GR suppression suggests the lack of normal enhancement of neural inhibition in the visual cortex in women with PMDD during their symptomatic luteal phase. This result resembles that obtained by Smith and colleagues in the motor cortex of women with strong PMS (Smith et al., 2003). Using TMS, Smith et al. found in these women a luteal phase-specific deficit in cortical inhibition and explained it by the lack of the normal increase in inhibition during the luteal phase.

Since our results in women with PMDD suggest the lack of normal luteal increase in GR suppression (indicative of unchanged inhibition between the follicular and luteal phases) rather than luteal decrease in GR suppression (which would be indicative of reduced inhibition in the luteal phase), they are more compatible with developing tolerance to ALLO (Turkmen et al., 2011) than with its paradoxical excitatory effect (Bäckström et al., 2014). This tolerance may reflect an abnormally attenuated action of ALLO on δGABA$_A$Rs on principal neurons, possibly due to an abnormal or insufficient plasticity of the δGABA$_A$Rs in response to the postovulatory increase in ALLO (Hantsoo & Epperson, 2020; Turkmen et al., 2011). Indeed, reduced sensitivity to the sedative effects of GABA$_A$Rs agonists, such as benzodiazepines, ethanol, and GABA-active steroids (pregnanolone), was previously demonstrated in women with PMDD during the luteal phase, when ALLO concentration is high, but not during the follicular phase, when the levels of GABA-active steroids are low (Nyberg et al., 2004; Sundstrom et al., 1997; Sundström & Bäckström, 1998; Timby et al., 2016).

Although changes in the visual circuitry are unlikely to be directly related to the core symptoms of PMDD (i.e., depression, anxiety, irritability, and mood lability), the luteal phase-specific deficit in visual GR suppression in our PMDD participants was associated with more severe clinical symptoms in them estimated on the same day (Fig. 6d). This correlation can be explained by the broadly altered action of progesterone-derived neurosteroids, i.e., ALLO, on δGABA$_A$Rs, which affects gamma

oscillations via altering inhibitory neurotransmission in the visual cortex and is also manifested in clinical symptoms of PMDD via altering inhibitory neurotransmission in the brain regions involved in the control of mood and anxiety (see, e.g., Liu et al., 2015; Ossewaarde et al., 2010; Van Wingen et al., 2007).

To sum up, our results indicate that the GR suppression associated with increasing excitatory drive to the visual cortex reflects a luteal phase-specific and symptom severity-related deficit in neural inhibition in PMDD.

GR facilitation in control women and those with PMDD

The GR facilitation caused by a moderate increase in excitatory drive (when drift rate of the grating changes from 'static' to 'slow') most probably has a different mechanism than the GR suppression caused by strong increase in excitatory drive (when drift rate of the grating changes from 'slow' to 'medium' or 'fast') (see Manyukhina et al., 2021; Orekhova, Prokofyev, et al., 2020 for discussion). While GR suppression is thought to be caused by an 'excessive' inhibitory neurotransmission (Börgers & Kopell, 2005), GR facilitation reflects increased activation of the excitatory principal cells and their synchronization (Buzsáki & Wang, 2008; Vinck et al., 2013).

In contrast to gamma suppression, gamma facilitation did not show MC-related changes in our participants with or without PMDD. Yet, the enhanced GR facilitation differentiated women with PMDD from control participants in both phases of the MC. The dissociation between the GR facilitation and GR suppression in their sensitivity to cyclic changes in neurosteroids supports a difference in the mechanisms underlying the two phenomena. Although this result was not predicted, it suggests the existence of additional MC-independent mechanisms involved in PMDD.

The increase of the GR from the 'static' to the 'slow' condition is most likely mediated by increasing excitation of principal neurons, which in turn activate local inhibitory neurons through recurrent excitatory connections (PING mechanism of gamma generation, see Buzsáki & Wang, 2008; Vinck et al., 2013). Hence, the main factor determining the degree of the gamma power increase, i.e., gamma facilitation, is the phasic excitability of the principal neurons in the visual cortex, which is not affected by neurosteroid fluctuations in the MC (Mody & Maguire, 2012). Indeed, when stimulation strength increases to a certain level, more excited cortical circuitries produce stronger gamma oscillations in both animals (Towers et al., 2004; Yizhar et al., 2011) and humans (Hermes et al., 2015; Perry et al., 2013).

Both principal cells and inhibitory interneurons play pivotal roles in generation of gamma (Buzsáki & Wang, 2008; Vinck et al., 2013). However, normal peak frequency of the GR in women with PMDD (Table 4a) speaks against an apparent

deficit in functioning of the inhibitory neurons (see later in this Discussion). Therefore, the atypically high GR facilitation in PMDD is rather due to an elevated excitability of the principal neurons and enhanced propensity of these neurons to engage in synchronous oscillations in response to visual stimulation (i.e., 'increased gain'). A similar deficit in gain regulation was observed in the visual cortex of patients with idiopathic generalized epilepsy using the steady-state visually evoked potentials (SSVEP) paradigm. These patients demonstrated an abnormally strong increase of SSVEPs associated with increasing contrast of a flickering grating (Porciatti et al., 2000; Tsai et al., 2011; Won et al., 2017). The authors concluded that the altered regulation of gain control in these patients is the result of hyperexcitability of local neuronal ensembles and enhanced lateral spread of the excitation.

Since increased GR facilitation is observed in women with PMDD regardless of the MC phase, it is unlikely to depend on $\delta GABA_A Rs$-mediated tonic inhibition, and may reflect constitutively enhanced phasic excitability of principal neurons. Our finding of a neurofunctional abnormality that is not limited to the symptomatic luteal phase in women with PMDD is not unique. A number of studies that applied a range of different methods and paradigms reported in women with PMDD neurofunctional abnormalities during asymptomatic period of the MC (Bannbers et al., 2012; Comasco et al., 2014; Dan et al., 2020; Gingnell et al., 2012; Kask et al., 2008; Monteleone et al., 2000; Epperson et al., 2002; Nyberg et al., 2004; Sundstrom et al., 1997; Sundström & Bäckström, 1998; Timby et al., 2016). In particular, significantly decreased GABA concentration in the visual cortex was found in women with PMDD during the follicular phase (Epperson et al., 2002). There is also evidence on altered activation in the dorsolateral prefrontal-cingulate cortex (Comasco et al., 2014) and enhanced amygdala response to negative stimuli (Gingnell et al., 2012) in women with PMDD regardless of the MC phase. Thus, our results provide further support for the idea that pathophysiology of PMDD is not limited to the postovulatory luteal phase events, and that E/I imbalance in PMDD is present even during the asymptomatic follicular phase, but is further exacerbated during the premenstrual period, presumably due to altered sensitivity to GABA-active neurosteroids.

4.3. Frequency of GR and first scan effect

The neural mechanisms regulating peak frequency of gamma oscillations are substantially different from those regulating their power. While power of gamma oscillations reflects local excitatory-inhibitory interactions, their frequency is predominantly controlled by tonic excitability of parvalbumin-containing (PV+)

inhibitory neurons (Ferando & Mody, 2013, 2015; Mann & Mody, 2010; Towers et al., 2004). Difference in the mechanisms that regulate GR frequency and power may explain their different dynamics associated with increasing excitatory drive (Jia et al., 2013; Orekhova et al., 2018; Salelkar et al., 2018).

Similarly to Sumner et al. (2018), we found that in women with healthy MC, frequency of GR induced by static and slowly moving gratings increased from the follicular to the luteal phase (Fig. 4a). However, a closer investigation showed that this effect was exclusively driven by those subjects who first came for the MEG investigation during the luteal phase (Fig. 4c, right) and was absent in those who first came for the MEG investigation during the follicular phase (Fig. 4c, left). Although Sumner et al. counterbalanced phases of the first visit (luteal 1st vs. follicular 1st), they did not test the effect of the order of visits, which, in our study, appears to be the main source of the luteal phase-specific increase in gamma frequency.

Remarkably, control participants and those with PMDD showed the same dependency of GR frequency on the order of visits (Fig. 4). Furthermore, no differences were found between the PMDD and control groups in either the GR frequency or its modulation by the drift rate and MC phase. Thus, contrary to our prediction, frequency of visual gamma oscillations seems to be normal in PMDD.

The effect of the visit order on the GR frequency is interesting by itself. It shows that modulation of gamma frequency by a hormonal status depends on some situational factor. Changes in attention and/or learning are unlikely to explain this result, as neither reaction time, nor percent of omission and commission errors differed between the first and second visit in subjects first investigated during the luteal or follicular phase of the MC (all p's>0.11). On the other hand, according to the STAI questionnaire, the first visit to the MEG laboratory was associated in our participants with significantly higher situational anxiety than the second visit. We, therefore, suggest that a mild acute stress associated with being a study participant for the first time is a probable factor interfering with the hormone levels and contributing to our gamma frequency results. In line with this suggestion, several studies do show increased stress sensitivity in the luteal phase in both women without severe PMS (Hastrup & Light, 1984; Ossewaarde et al., 2010; Sato & Miyake, 2004) and women with PMDD (Beddig et al., 2019; Epperson et al., 2007; Hantsoo & Epperson, 2015, 2020; Logue & Moos, 1986; Parry et al., 2000).

The factors associated with the 1st visit led to an increase in GR frequency only if they coincided with the luteal phase of the MC. During this phase, the $\delta GABA_A Rs$-mediated inhibition is potentiated by ALLO (see the previous section). On the other hand, the $\delta GABA_A Rs$-mediated inhibition is further increased in response to acute stress (Bali & Jaggi, 2014; Biggio et al., 2014; Bortolato et al., 2022). The increase in

δGABA$_A$Rs-mediated tonic inhibition is expected to dampen excitability of PV+ inhibitory neurons, whose tonic inhibition is exclusively mediated by δGABA$_A$Rs (Ferando & Mody, 2014; Lee & Maguire, 2013). This dampened tonic activity of PV+ inhibitory neurons is expected to decrease GR frequency (Barth et al., 2015; Mann & Mody, 2010). Our results, however, strongly indicate the opposite.

Although speculative, the impact of mild acute stress on gamma frequency during the luteal phase may be explained by the paradoxical excitatory effect of GABA-mediated currents on inhibitory neurons. It has been demonstrated that experimentally increased tonic GABA$_A$Rs conductance, mimicking its natural increase during the luteal phase, has a depolarizing effect on adult hippocampal inhibitory neurons (Pavlov et al., 2014). While this excitatory effect, mediated by tonic GABA currents, may not be sufficient to cause a measurable increase in gamma frequency in the luteal phase, mild acute stress, e.g., associated with the first visit, may further increase interneuron excitability. Indeed, acute stress rapidly increases plasma and cortical ALLO levels, which, in turn, leads to upregulation of δGABA$_A$Rs (Barbaccia et al., 1998; Mody & Maguire, 2012; Purdy et al., 1991). That is, when acute stress coincides with the luteal phase, δGABA$_A$Rs reach their highest density. Apart from the potentiating effect of ALLO, other stress-derived neurosteriods have also been shown to increase the burst firing of inhibitory neurons (Mody & Maguire, 2012; Shin et al., 2011). The increased excitability of inhibitory neurons due to the additive effects of the luteal phase and mild acute stress could thus explain the increased frequency of gamma oscillations in women whose first visit to the laboratory coincided with the luteal phase of their MC.

Whatever the exact mechanism leading to the luteal phase-specific effect of the first MEG scanning on the visual GR frequency, it did not differ in women with and without PMDD. Overall, the absence of PMDD-specific differences in the frequency of visual GR as well as in its modulation by either excitatory drive (i.e., grating's drift rate), visit order, or MC phase suggests that activity of PV+ interneurons in the visual cortex is not altered in PMDD.

Although GR frequency was not affected in women with PMDD, we did find in this group an altered relationship between GR frequency and strength of GR suppression. In control women, we reproduced the previously reported correlation (Orekhova et al., 2018): higher gamma frequency correlated with greater gamma suppression caused by a strong excitatory drive to the visual cortex. This correlation suggests that in the healthy brain, the inhibitory down-regulation of the E/I balance evidenced by GR power suppression is proportional to excitation of inhibitory PV+ neurons that control frequency of gamma oscillations (see Orekhova et al., 2018 for

thorough discussion). In women with PMDD, GR frequency did not correlate with the GR suppression, and the difference in the correlation coefficients between PMDD and control groups was significant in the luteal phase. This finding suggests that in women with PMDD the activity of normally functioning presynaptic inhibitory neurons is not the main factor determining the E/I balance in gamma generating visual circuitry. Instead, as discussed above, the atypically strong excitability of principal cells may be the key factor contributing to interindividual variability of the local E/I ratio in the visual cortex in PMDD.

4.4. Perceptual spatial suppression in PMDD

Spatial suppression is a perceptual phenomenon, which depends on neural inhibition strength in the visual cortex (Tadin, 2015; Tadin et al., 2003, 2019). It manifests itself in a longer time needed to discriminate the direction of motion of large high-contrast gratings, compared to small ones. Reduced spatial suppression has been used as a behavioral indication of impaired neural surround inhibition in patients with neuropsychiatric disorders (Foss-Feig et al., 2013; Linares et al., 2020; Serrano-Pedraza et al., 2014; Sysoeva et al., 2017). Surround inhibition, however, is a complex phenomenon that relies on inhibition within primary visual cortex and its top-down regulation by higher-tier cortical areas (Angelucci et al., 2017; Angelucci & Bressloff, 2006; Nurminen et al., 2018). Animal studies suggest that somatostatin-containing interneurons, which subserve top-down contextual effects, are particularly important for surround inhibition and, most probably, for perceptual spatial suppression as well (Nienborg et al., 2013; Zhang et al., 2014).

During the follicular phase, our participants with PMDD tended to have even stronger spatial suppression than control women. This trend is opposite to the effects found in elderly individuals (Betts et al., 2005, 2012; Tadin et al., 2019), patients with depression (Golomb et al., 2009) or schizophrenia (Tadin et al., 2006), and is not consistent with a reduction of neural inhibition. This rather indicates preserved or even elevated top-down inhibition in the asymptomatic follicular phase in women with PMDD. However, the complex hierarchical organization of excitatory-inhibitory interactions underlying perceptual spatial suppression makes it difficult to separate initially elevated inhibition from homeostatic compensation for its deficit (see, e.g., Nelson & Valakh, 2015). Interestingly, such a putative compensatory mechanism may also present in patients with migraine, who are thought to have an elevated neural excitability in the visual cortex (Mulleners et al., 2001; Wickmann et al., 2015), but at

the same time demonstrate increased perceptual spatial suppression (Battista et al., 2010, 2011).

While in control participants perceptual spatial suppression was independent of the MC phase, in women with PMDD, it significantly decreased from the follicular to the luteal phase (Fig. 7b). This decrease may reflect diminished effectiveness of the top-down compensatory influences in women with PMDD during the symptomatic luteal phase. This suggestion is broadly consistent with MRI findings of decreased anterior cingulate cortex control over the amygdala response to negative social stimulation in women with PMDD during the luteal phase compared to the follicular phase (Gingnell et al., 2014; but see Comasco et al., 2014; Petersen et al., 2019), and with animal studies showing a facilitating effect of anterior cingulate cortex activation on surround inhibition (Zhang et al., 2014).

The atypical regulation of perceptual spatial suppression in women with PMDD is further supported by the lack of the neuro-behavioral correlation found in the control group. Consistent with our previous results in typically developing children (Orekhova, Rostovtseva, et al., 2020), visual GR suppression predicted spatial suppression in control women, with stronger gamma suppression corresponding to greater perceptual suppression (see Results section and Supplementary table S2 for additional analysis). This relationship was absent in women with PMDD, indicating that some atypical compensatory processes may result in apparently normal perceptual spatial suppression in this clinical group.

4.5. Limitations and future direction of research

One limitation of the present study is the relatively small sample size. Although being comparable to that in the majority of neurophysiological studies into PMS/PMDD pathogenesis, it did not allow us to investigate the subtypes, which likely exist within the PMDD nosological category (Eisenlohr-Moul et al., 2020) and which may be associated with distinct neuro-functional deficits. According to the literature, about 50% of women with PMDD have comorbid conditions (Cirillo et al., 2012; de Carvalho et al., 2018; Kim et al., 2004). In future, it would be important to investigate if the elevated cortical excitability characterizes all women with PMDD, or rather certain patients with specific symptom patterns and/or comorbidities.

Further, our current design contrasts the early-to-mid follicular phase with the mid-to-late luteal phase, while it misses events in the middle of the MC. The inclusion of the ovulatory period where many women with PMDD are already starting to develop

adverse symptoms, would help to better understand the relationship between MC-related changes in the E/I balance and PMS onset.

It should be also mentioned that in the future it would be interesting to model the predicted effects of excitatory drive, acute stress, top-down control and PMDD diagnosis on gamma parameters using computer modeling.

Finally, our work has been restricted to the occipital cortex, which is not usually involved in the pathogenesis of affective disorders. One cannot rule out the possibility that E/I balance in other, 'core' for PMDD, brain regions will be differently affected by neurosteroids and other factors.

5. Conclusion

Although the main symptoms of PMDD are emotional rather than sensory-related, the neuro-functional abnormalities associated with this disorder are present even in the visual cortex. Our results showing atypical modulation of gamma power as a function of excitatory drive suggest that neuronal excitability is constitutively elevated in women with PMDD, which may be related to the high frequency of comorbid neuropsychiatric conditions in this clinical group. The E/I balance in women with PMDD shifts even further to excitation during the luteal phase, possibly as a result of their atypical sensitivity to neurosteroids. The mechanisms of elevated neural excitability and E/I imbalance in PMDD remain to be elucidated, but they are rather unlikely to be caused by deficits in inhibitory neurons. Indeed, the peak frequency of oscillatory gamma response, which is primarily regulated by tonic excitability of parvalbumin-containing interneurons, was normal in PMDD. Moreover, women with PMDD and control women demonstrated the same luteal phase-specific effect of 'the first scan' on the gamma peak frequency. The presence of normal or even slightly increased (in the follicular phase) perceptual spatial suppression in PMDD also indicates that there is no major deficit in activity of inhibitory neurons, at least in the visual cortex. Therefore, our results suggest that the changes in neural E/I balance in the visual cortex in women with PMDD are most likely explained by increased tonic excitability of the principal cells and/or impaired regulation of their excitability at the synaptic levels. Overall, our results contribute to elucidating the mechanisms of PMDD and provide new insights into the functional correlates of the visual gamma rhythm.

ACKNOWLEDGMENTS

We sincerely thank all the participants who have participated in this study.

FUNDING INFORMATION

This work was supported by an internal grant from Moscow State University of Psychology and Education (MSUPE).

AUTHOR CONTRIBUTIONS

Study concept and design: Tatiana Stroganova, Elena Orekhova. *Data acquisition and preprocessing*: Viktoriya Manyukhina, Andrey Prokofyev, Tatiana Obukhova. *Data analysis*: Viktoriya Manyukhina, Elena Orekhova, Tatiana Stroganova. *Writing the first draft*: Viktoriya Manyukhina and Elena Orekhova. *Supervision study*: Elena Orekhova and Tatiana Stroganova. *Read and approve the final version*: all authors.

CONFLICT OF INTEREST

All authors declare no competing interests.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

REFERENCES

Adesnik, H. (2017). Synaptic Mechanisms of Feature Coding in the Visual Cortex of Awake Mice. *Neuron*, *95*(5), 1147-1159.e4. https://doi.org/10.1016/j.neuron.2017.08.014

Amiel Castro, R. T., Ehlert, U., & Fischer, S. (2021). Variation in genes and hormones of the hypothalamic-pituitary-ovarian axis in female mood disorders – A systematic review and meta-analysis. *Frontiers in Neuroendocrinology*, *62*, 100929. https://doi.org/10.1016/j.yfrne.2021.100929

Angelucci, A., Bijanzadeh, M., Nurminen, L., Federer, F., Merlin, S., & Bressloff, P. C. (2017). Circuits and Mechanisms for Surround Modulation in Visual Cortex. *Annual Review of Neuroscience*, *40*, 425–451. https://doi.org/10.1146/annurev-neuro-072116-031418

Angelucci, A., & Bressloff, P. C. (2006). Contribution of feedforward, lateral and feedback connections to the classical receptive field center and extra-classical receptive field surround of primate V1 neurons. *Progress in Brain Research*, *154*, 93–120. https://doi.org/10.1016/S0079-6123(06)54005-1

American Psychiatric Association (2013). *Diagnostic and statistical manual of mental disorders (5th ed.)*. https://doi.org/10.4103/0019-5545.117131

Avitabile, T., Longo, A., Caruso, S., Gagliano, C., Amato, R., Scollo, D., Lopes, R., Pulvirenti, L., Toto, L., Torrisi, B., & Agnello, C. (2007). Changes in visual evoked potentials during the menstrual cycle in young women. *Current Eye Research*, *32*(11), 999–1003. https://doi.org/10.1080/02713680701679006

Azcoitia, I., Yague, J. G., & Garcia-Segura, L. M. (2011). Estradiol synthesis within the human brain. *Neuroscience, 191*, 139–147. https://doi.org/10.1016/j.neuroscience.2011.02.012

Bäckström, T., Bixo, M., Johansson, M., Nyberg, S., Ossewaarde, L., Ragagnin, G., Savic, I., Strömberg, J., Timby, E., van Broekhoven, F., & van Wingen, G. (2014). Allopregnanolone and mood disorders. *Progress in Neurobiology*, *113*, 88–94. https://doi.org/10.1016/j.pneurobio.2013.07.005

Bäckström, T., Haage, D., Löfgren, M., Johansson, I. M., Strömberg, J., Nyberg, S., Andréen, L., Ossewaarde, L., van Wingen, G. A., Turkmen, S., & Bengtsson, S. K. (2011). Paradoxical effects of GABA-A modulators may explain sex steroid induced negative mood symptoms in some persons. *Neuroscience, 191*, 46–54. https://doi.org/10.1016/j.neuroscience.2011.03.061

Bali, A., & Jaggi, A. S. (2014). Multifunctional aspects of allopregnanolone in stress and related disorders. *Progress in Neuro-Psychopharmacology and Biological Psychiatry, 48*, 64–78. https://doi.org/10.1016/j.pnpbp.2013.09.005

Baller, E. B., Wei, S. M., Kohn, P. D., Rubinow, D. R., Alarcón, G., Schmidt, P. J., & Berman, K. F. (2013). Abnormalities of dorsolateral prefrontal function in women with premenstrual dysphoric disorder: A multimodal neuroimaging study. *American Journal of Psychiatry*, *170*(3), 305–314. https://doi.org/10.1176/appi.ajp.2012.12030385

Balthazart, J., & Ball, G. F. (2006). Is brain estradiol a hormone or a neurotransmitter? *Trends in Neurosciences*, *29*(5), 241–249. https://doi.org/10.1016/j.tins.2006.03.004

Bannbers, E., Gingnell, M., Engman, J., Morell, A., Comasco, E., Kask, K., Garavan, H., Wikström, J., & Sundström Poromaa, I. (2012). The effect of premenstrual dysphoric disorder and menstrual cycle phase on brain activity during response inhibition. *Journal of Affective Disorders*, *142*(1–3), 347–350. https://doi.org/10.1016/j.jad.2012.04.006

Barbaccia, M. L., Concas, A., Serra, M., & Biggio, G. (1998). Stress and neurosteroids in adult and aged rats. *Experimental Gerontology*, *33*(7–8), 697–712. https://doi.org/10.1016/S0531-5565(98)00042-4

Barth, A. M. I., Ferando, I., & Mody, I. (2014). Ovarian cycle-linked plasticity of δ-GABAA receptor subunits in hippocampal interneurons affects γ oscillations in vivo. *Frontiers in Cellular Neuroscience*, *8*(AUG). https://doi.org/10.3389/fncel.2014.00222

Barth, C., Villringer, A., & Sacher, J. (2015). Sex hormones affect neurotransmitters and shape the adult female brain during hormonal transition periods. *Frontiers in Neuroscience*, *9*, 1–20. https://doi.org/10.3389/fnins.2015.00037

Bartos, M., Vida, I., & Jonas, P. (2007). Synaptic mechanisms of synchronized gamma oscillations in inhibitory interneuron networks. *Nature Reviews Neuroscience*, *8*(1), 45–56. https://doi.org/10.1038/nrn2044

Battista, J., Badcock, D. R., & McKendrick, A. M. (2010). Center-surround visual motion processing in migraine. *Investigative Ophthalmology and Visual Science*, *51*(11), 6070–6076. https://doi.org/10.1167/iovs.10-5290

Battista, J., Badcock, D. R., & McKendrick, A. M. (2011). Migraine increases centre-surround suppression for drifting visual stimuli. *PLoS ONE*, *6*(4). https://doi.org/10.1371/journal.pone.0018211

Bazanova, O. M., Kondratenko, A. V., Kuzminova, O. I., Muravlyova, K. B., & Petrova, S. E. (2014). EEG alpha indices depending on the menstrual cycle phase and salivary progesterone level. *Human Physiology*, *40*(2), 140–148. https://doi.org/10.1134/S0362119714020030

Beck, A. T., Steer, R. A., & Carbin, M. G. (1988). Psychometric properties of the Beck Depression Inventory: Twenty-five years of evaluation. *Clinical Psychology Review*, *8*(1), 77–100. https://doi.org/10.1016/0272-7358(88)90050-5

Beddig, T., Reinhard, I., & Kuehner, C. (2019). Stress, mood, and cortisol during daily life in women with Premenstrual Dysphoric Disorder (PMDD). *Psychoneuroendocrinology*, *109*, 104372. https://doi.org/10.1016/j.psyneuen.2019.104372

Betts, L. R., Sekuler, A. B., & Bennett, P. J. (2012). Spatial characteristics of motion-sensitive mechanisms change with age and stimulus spatial frequency. *Vision Research*, *53*(1), 1–14. https://doi.org/10.1016/j.visres.2011.11.004

Betts, L. R., Taylor, C. P., Sekuler, A. B., & Bennett, P. J. (2005). Aging reduces center-surround antagonism in visual motion processing. *Neuron*, *45*(3), 361–366. https://doi.org/10.1016/j.neuron.2004.12.041

Biggio, G., Pisu, M. G., Biggio, F., & Serra, M. (2014). Allopregnanolone modulation of HPA axis function in the adult rat. *Psychopharmacology*, *231*(17), 3437–3444. https://doi.org/10.1007/s00213-014-3521-6

Biggs, W. S., & Demuth, R. H. (2011). Premenstrual syndrome and premenstrual dysphoric disorder. *Am Fam Physician*, *84*(8), 918–924. https://doi.org/10.1007/s10304-022-00451-9

Bixo, M., Backstrom, T., Winblad, B., Selstam, G., & Andersson, A. (1986). Comparison between pre- and postovulatory distributions of oestradiol and progesterone in the brain of the PMSG-treated rat. *Acta Physiologica Scandinavica*, *128*(2), 241–246. https://doi.org/10.1111/j.1748-1716.1986.tb07972.x

Börgers, C., & Kopell, N. (2005). Effects of noisy drive on rhythms in networks of excitatory and inhibitory neurons. *Neural Computation*, *17*(3), 557–608. https://doi.org/10.1162/0899766053019908

Bortolato, M., Coffey, B. J., Gabbay, V., & Scheggi, S. (2022). Allopregnanolone: The missing link to explain the effects of stress on tic exacerbation? *Journal of Neuroendocrinology*, *34*(2), 1–14. https://doi.org/10.1111/jne.13022

Bosman, R. C., Albers, C. J., De Jong, J., Batalas, N., & Aan Het Rot, M. (2018). No Menstrual Cyclicity in Mood and Interpersonal Behaviour in Nine Women with Self-Reported Premenstrual Syndrome. *Psychopathology*, *51*(4), 290–294. https://doi.org/10.1159/000489268

Brötzner, C. P., Klimesch, W., & Kerschbaum, H. H. (2015). Progesterone-associated increase in ERP amplitude correlates with an improvement in performance in a spatial attention paradigm. *Brain Research*, *1595*, 74–83. https://doi.org/10.1016/j.brainres.2014.11.004

Buzsáki, G., & Wang, X.-J. (2008). Mechanisms of Gamma Oscillations. *Annu Rev Neurosci*, *35*, 1–7. https://doi.org/10.1146/annurev-neuro-062111-150444.Mechanisms

Cannon, J., Mccarthy, M. M., Lee, S., Lee, J., Börgers, C., Whittington, M. A., & Kopell, N. (2014). Neurosystems: Brain rhythms and cognitive processing. *European Journal of Neuroscience*, *39*(5), 705–719. https://doi.org/10.1111/ejn.12453

Chan, J. H., Lo, C., Hsu, C. D., Chiu, C. C., Huang, M. C., Liao, S. C., Chen, I. M., Chen, W. Y., Chen, H. C., & Kuo, P. H. (2021). Premenstrual dysphoric symptoms and lifetime suicide experiences in patients with mood disorder. *General Hospital Psychiatry*, *71*(7), 82–87. https://doi.org/10.1016/j.genhosppsych.2021.04.009

Cirillo, P. C., Passos, R. B. F., Bevilaqua, M. C. do N., López, J. R. R. A., & Nardi, A. E. (2012). Bipolar disorder and premenstrual syndrome or premenstrual dysphoric disorder comorbidity: A systematic review. *Revista Brasileira de Psiquiatria*, *34*(4), 467–479. https://doi.org/10.1016/j.rbp.2012.04.010

Clemens, A. M., Lenschow, C., Beed, P., Li, L., Sammons, R., Naumann, R. K., Wang, H., Schmitz, D., & Brecht, M. (2019). Estrus-Cycle Regulation of Cortical Inhibition. *Current Biology*, *29*(4), 605-615.e6. https://doi.org/10.1016/j.cub.2019.01.045

Comasco, E., Hahn, A., Ganger, S., Gingnell, M., Bannbers, E., Oreland, L., Wikström, J., Epperson, C. N., Lanzenberger, R., & Sundström-Poromaa, I. (2014). Emotional fronto-

cingulate cortex activation and brain derived neurotrophic factor polymorphism in premenstrual dysphoric disorder. *Human Brain Mapping*, *35*(9), 4450–4458. https://doi.org/10.1002/hbm.22486

Dan, R., Reuveni, I., Canetti, L., Weinstock, M., Segman, R., Goelman, G., & Bonne, O. (2020). Trait-related changes in brain network topology in premenstrual dysphoric disorder. *Hormones and Behavior*, *124*, 104782. https://doi.org/10.1016/j.yhbeh.2020.104782

de Carvalho, A. B., Cardoso, T. de A., Mondin, T. C., da Silva, R. A., Souza, L. D. de M., Magalhães, P. V. da S., & Jansen, K. (2018). Prevalence and factors associated with Premenstrual Dysphoric Disorder: A community sample of young adult women. *Psychiatry Research*, *268*, 42–45. https://doi.org/10.1016/j.psychres.2018.06.005

De Lima Resende, L. A., Silva, M. D., Impemba, F., Achôa, N. B., & Schelp, A. O. (2000). Multimodal evoked potentials and the ovarian cycle in young ovulating women. *Arquivos de Neuro-Psiquiatria*, *58*(2-B), 418–423. https://doi.org/10.1590/s0004-282x2000000300004

Devall, A. J., Santos, J. M., Fry, J. P., Honour, J. W., Brandão, M. L., & Lovick, T. A. (2015). Elevation of brain allopregnanolone rather than 5-HT release by short term, low dose fluoxetine treatment prevents the estrous cycle-linked increase in stress sensitivity in female rats. *European Neuropsychopharmacology*, *25*(1), 113–123. https://doi.org/10.1016/j.euroneuro.2014.11.017

Direkvand-Moghadam, A., Sayehmiri, K., Delpisheh, A., & Satar, K. (2014). Epidemiology of premenstrual syndrome, a systematic review and meta-analysis study. *Journal of Clinical and Diagnostic Research*, *8*(2), 106–109. https://doi.org/10.7860/JCDR/2014/8024.4021

Dubey, N., Hoffman, J. F., Schuebel, K., Yuan, Q., Martinez, P. E., Nieman, L. K., Rubinow, D. R., Schmidt, P. J., & Goldman, D. (2017). The ESC/E(Z) complex, an effector of response to ovarian steroids, manifests an intrinsic difference in cells from women with premenstrual dysphoric disorder. *Molecular Psychiatry*, *22*(8), 1172–1184. https://doi.org/10.1038/mp.2016.229

Dubol, M., Epperson, C. N., Lanzenberger, R., Sundström-Poromaa, I., & Comasco, E. (2020). Neuroimaging premenstrual dysphoric disorder: A systematic and critical review. *Frontiers in Neuroendocrinology*, *57*, 100838. https://doi.org/10.1016/j.yfrne.2020.100838

Eisenlohr-Moul, T. A., Girdler, S. S., Schmalenberger, K. M., Dawson, D. N., Surana, P., Johnson, J. L., & Rubinow, D. R. (2017). Toward the reliable diagnosis of DSM-5 premenstrual dysphoric disorder: The Carolina premenstrual assessment scoring system (C-PASS). *American Journal of Psychiatry*, *174*(1), 51–59. https://doi.org/10.1176/appi.ajp.2016.15121510

Eisenlohr-Moul, T. A., Kaiser, G., Weise, C., Schmalenberger, K. M., Kiesner, J., Ditzen, B., & Kleinstäuber, M. (2020). Are there temporal subtypes of premenstrual dysphoric disorder?: Using group-based trajectory modeling to identify individual differences in symptom change. *Psychological Medicine*, *50*(6), 964–972. https://doi.org/10.1017/S0033291719000849

Engin, E., & Treit, D. (2007). The anxiolytic-like effects of allopregnanolone vary as a function of intracerebral microinfusion site: The amygdala, medial prefrontal cortex, or hippocampus.

Behavioural Pharmacology, *18*(5–6), 461–470.
https://doi.org/10.1097/FBP.0b013e3282d28f6f

Epperson, C. N., Pittman, B., Czarkowski, K. A., Stiklus, S., Krystal, J. H., & Grillon, C. (2007). Luteal-phase accentuation of acoustic startle response in women with premenstrual dysphoric disorder. *Neuropsychopharmacology*, *32*(10), 2190–2198. https://doi.org/10.1038/sj.npp.1301351

Epperson, N. C., Haga, K., Mason, G. F., Sellers, E., Gueorguieva, R., Zhang, W., Weiss, E., Rothman, D. L., & Krystal, J. H. (2002). Cortical γ-aminobutyric acid levels across the menstrual cycle in healthy women and those with premenstrual dysphoric disorder: A proton magnetic resonance spectroscopy study. *Archives of General Psychiatry*, *59*(9), 851–858. https://doi.org/10.1001/archpsyc.59.9.851

Farage, M. A., Osborn, T. W., & MacLean, A. B. (2008). Cognitive, sensory, and emotional changes associated with the menstrual cycle: A review. *Archives of Gynecology and Obstetrics*, *278*(4), 299–307. https://doi.org/10.1007/s00404-008-0708-2

Ferando, I., & Mody, I. (2013). Altered gamma oscillations during pregnancy through loss of δ subunit-containing GABAA receptors on parvalbumin interneurons. *Frontiers in Neural Circuits*, *7*(144), 1–14. https://doi.org/10.3389/fncir.2013.00144

Ferando, I., & Mody, I. (2014). Interneuronal GABAA receptors inside and outside of synapses. *Current Opinion in Neurobiology*, *26*, 57–63. https://doi.org/10.1016/j.conb.2013.12.001

Ferando, I., & Mody, I. (2015). In vitro gamma oscillations following partial and complete ablation of δ subunit-containing GABAA receptors from parvalbumin interneurons. *Neuropharmacology*, *88*, 91–98. https://doi.org/10.1016/j.neuropharm.2014.09.010

Finocchi, C., & Ferrari, M. (2011). Female reproductive steroids and neuronal excitability. *Neurological Sciences*, *32*(1), 31–35. https://doi.org/10.1007/s10072-011-0532-5

Foss-Feig, J. H., Tadin, D., Schauder, K. B., & Cascio, C. J. (2013). A substantial and unexpected enhancement of motion perception in autism. *Journal of Neuroscience*, *33*(19), 8243–8249. https://doi.org/10.1523/JNEUROSCI.1608-12.2013

Gaetz, W., Roberts, T. P. L., Singh, K. D., & Muthukumaraswamy, S. D. (2012). Functional and structural correlates of the aging brain: Relating visual cortex (V1) gamma band responses to age-related structural change. *Human Brain Mapping*, *33*(9), 2035–2046. https://doi.org/10.1002/hbm.21339

Gao, M., Gao, D., Sun, H., Cheng, X., An, L., & Qiao, M. (2021). Trends in Research Related to Premenstrual Syndrome and Premenstrual Dysphoric Disorder From 1945 to 2018: A Bibliometric Analysis. *Frontiers in Public Health*, *9*, 1–14. https://doi.org/10.3389/fpubh.2021.596128

Ghaemi, S. N., Miller, C. J., Berv, D. A., Klugman, J., Rosenquist, K. J., & Pies, R. W. (2005). *Sensitivity and specificity of a new bipolar spectrum diagnostic scale*. *84*, 273–277. https://doi.org/10.1016/S0165-0327(03)00196-4

Gingnell, M., Ahlstedt, V., Bannbers, E., Wikström, J., Sundström-Poromaa, I., & Fredrikson, M. (2014). Social stimulation and corticolimbic reactivity in premenstrual dysphoric disorder: A preliminary study. *Biology of Mood and Anxiety Disorders*, *4*(1), 1–10. https://doi.org/10.1186/2045-5380-4-3

Gingnell, M., Morell, A., Bannbers, E., Wikström, J., & Sundström Poromaa, I. (2012). Menstrual cycle effects on amygdala reactivity to emotional stimulation in premenstrual dysphoric disorder. *Hormones and Behavior*, *62*(4), 400–406. https://doi.org/10.1016/j.yhbeh.2012.07.005

Golomb, J. D., McDavitt, J. R. B., Ruf, B. M., Chen, J. I., Saricicek, A., Maloney, K. H., Hu, J., Chun, M. M., & Bhagwagar, Z. (2009). Enhanced visual motion perception in major depressive disorder. *Journal of Neuroscience*, *29*(28), 9072–9077. https://doi.org/10.1523/JNEUROSCI.1003-09.2009

Halbreich, U., Borenstein, J., Pearlstein, T., & Kahn, L. S. (2003). The prevalence, impairment, impact, and burden of premenstrual dysphoric disorder (PMS/PMDD). *Psychoneuroendocrinology*, *28*, 1–23. https://doi.org/10.1016/S0306-4530(03)00098-2

Hammarbäck, S., Damber, J. E., & Bäckström, T. (1989). Relationship between symptom severity and hormone changes in women with premenstrual syndrome. *Journal of Clinical Endocrinology and Metabolism*, *68*(1), 125–130. https://doi.org/10.1210/jcem-68-1-125

Hantsoo, L., & Epperson, C. N. (2015). Premenstrual Dysphoric Disorder: Epidemiology and Treatment. *Current Psychiatry Reports*, *17*(11). https://doi.org/10.1007/s11920-015-0628-3

Hantsoo, L., & Epperson, C. N. (2020). Allopregnanolone in premenstrual dysphoric disorder (PMDD): Evidence for dysregulated sensitivity to GABA-A receptor modulating neuroactive steroids across the menstrual cycle. *Neurobiology of Stress*, *12*, 100213. https://doi.org/10.1016/j.ynstr.2020.100213

Haraguchi, R., Hoshi, H., Ichikawa, S., Hanyu, M., Nakamura, K., Fukasawa, K., Poza, J., Rodríguez-González, V., Gómez, C., & Shigihara, Y. (2021). The Menstrual Cycle Alters Resting-State Cortical Activity: A Magnetoencephalography Study. *Frontiers in Human Neuroscience*, *15*, 1–16. https://doi.org/10.3389/fnhum.2021.652789

Harris, C. R., Millman, K. J., van der Walt, S. J., Gommers, R., Virtanen, P., Cournapeau, D., Wieser, E., Taylor, J., Berg, S., Smith, N. J., Kern, R., Picus, M., Hoyer, S., van Kerkwijk, M. H., Brett, M., Haldane, A., del Río, J. F., Wiebe, M., Peterson, P., … Oliphant, T. E. (2020). Array programming with NumPy. *Nature*, *585*(7825), 357–362. https://doi.org/10.1038/s41586-020-2649-2

Hastrup, J. L., & Light, K. C. (1984). Sex differences in cardiovascular stress responses: modulation as a function of menstrual cycle phases. *Journal of Psychosomatic Research*, *28*(6), 475–483.

Héja, L., Nyitrai, G., Kékesi, O., Dobolyi, Á., Szabó, P., Fiáth, R., Ulbert, I., Pál-Szenthe, B., Palkovits, M., & Kardos, J. (2012). Astrocytes convert network excitation to tonic inhibition of neurons. *BMC Biology*, *10*. https://doi.org/10.1186/1741-7007-10-26

Hermes, D., Miller, K. J., Wandell, B. A., & Winawer, J. (2015). Stimulus dependence of gamma oscillations in human visual cortex. *Cerebral Cortex*, *25*(9), 2951–2959. https://doi.org/10.1093/cercor/bhu091

Hirschfeld, R. M. A., Williams, J. B. W., Spitzer, R. L., Calabrese, J. R., Flynn, L., Keck, J., Lewis, L., McElroy, S. L., Post, R. M., Rapport, D. J., Russell, J. M., Sachs, G. S., & Zajecka, J. (2000). Development and validation of a screening instrument for bipolar spectrum disorder: The mood disorder questionnaire. *American Journal of Psychiatry*, *157*(11), 1873–1875. https://doi.org/10.1176/appi.ajp.157.11.1873

Hoogenboom, N., Schoffelen, J. M., Oostenveld, R., Parkes, L. M., & Fries, P. (2006). Localizing human visual gamma-band activity in frequency, time and space. *NeuroImage*, *29*(3), 764–773. https://doi.org/10.1016/j.neuroimage.2005.08.043

Huang, Y., Zhou, R., Wu, M., Wang, Q., & Zhao, Y. (2015). Premenstrual syndrome is associated with blunted cortisol reactivity to the TSST. *Stress*, *18*(2), 160–168. https://doi.org/10.3109/10253890.2014.999234

Jeong, J. K., Tremere, L. A., Burrows, K., Majewska, A. K., & Pinaud, R. (2011). The mouse primary visual cortex is a site of production and sensitivity to estrogens. *PLoS ONE*, *6*(5). https://doi.org/10.1371/journal.pone.0020400

Jia, X., Xing, D., & Kohn, A. (2013). No consistent relationship between gamma power and peak frequency in macaque primary visual cortex. *Journal of Neuroscience*, *33*(1), 17–25. https://doi.org/10.1523/JNEUROSCI.1687-12.2013

Kask, K., Gulinello, M., Bäckström, T., Geyer, M. A., & Sundström-Poromaa, I. (2008). Patients with premenstrual dysphoric disorder have increased startle response across both cycle phases and lower levels of prepulse inhibition during the late luteal phase of the menstrual cycle. *Neuropsychopharmacology*, *33*(9), 2283–2290. https://doi.org/10.1038/sj.npp.1301599

Kim, D. R., Gyulai, L., Freeman, E. W., Morrison, M. F., Baldassano, C., & Dubé, B. (2004). Premenstrual dysphoric disorder and psychiatric co-morbidity. *Archives of Women's Mental Health*, *7*(1), 37–47. https://doi.org/10.1007/s00737-003-0027-3

Kow, L. M., Easton, A., & Pfaff, D. W. (2005). Acute estrogen potentiates excitatory responses of neurons in rat hypothalamic ventromedial nucleus. *Brain Research*, *1043*(1–2), 124–131. https://doi.org/10.1016/j.brainres.2005.02.068

Lee, V., & Maguire, J. (2013). Impact of inhibitory constraint of interneurons on neuronal excitability. *Journal of Neurophysiology*, *110*(11), 2520–2535. https://doi.org/10.1152/jn.00047.2013

Lin, I. M., Tsai, Y. C., Peper, E., & Yen, C. F. (2013). Depressive mood and frontal alpha asymmetry during the luteal phase in premenstrual dysphoric disorder. *Journal of Obstetrics and Gynaecology Research*, *39*(5), 998–1006. https://doi.org/10.1111/jog.12020

Linares, D., Amoretti, S., Marin-Campos, R., Sousa, A., Prades, L., Dalmau, J., Bernardo, M., & Compte, A. (2020). Spatial suppression and sensitivity for motion in schizophrenia. *Schizophrenia Bulletin Open*, *1*(1), 1–76.

Liu, B., Wang, G., Gao, D., Gao, F., Zhao, B., Qiao, M., Yang, H., Yu, Y., Ren, F., Yang, P., Chen, W., & Rae, C. D. (2015). Alterations of GABA and glutamate-glutamine levels in premenstrual dysphoric disorder: A 3T proton magnetic resonance spectroscopy study. *Psychiatry Research - Neuroimaging, 231*(1), 64–70. https://doi.org/10.1016/j.pscychresns.2014.10.020

Logue, C. M., & Moos, R. H. (1986). Perimenstrual symptoms: Prevalence and risk factors. *Psychosomatic Medicine, 48*(6), 388–414. https://doi.org/10.1097/00006842-198607000-00002

Lovick, T. A. (2014). Sex determinants of experimental panic attacks. *Neuroscience and Biobehavioral Reviews, 46*, 465–471. https://doi.org/10.1016/j.neubiorev.2014.03.006

Lovick, T. A., Griffiths, J. L., Dunn, S. M. J., & Martin, I. L. (2005). Changes in GABAA receptor subunit expression in the midbrain during the oestrous cycle in Wistar rats. *Neuroscience, 131*(2), 397–405. https://doi.org/10.1016/j.neuroscience.2004.11.010

Lusk, B. R., Carr, A. R., Ranson, V. A., Bryant, R. A., & Felmingham, K. L. (2015). Early visual processing is enhanced in the midluteal phase of the menstrual cycle. *Psychoneuroendocrinology, 62*, 343–351. https://doi.org/10.1016/j.psyneuen.2015.08.022

MacKenzie, G., & Maguire, J. (2014). The role of ovarian hormone-derived neurosteroids on the regulation of GABAA receptors in affective disorders. *Psychopharmacology, 231*(17), 3333–3342. https://doi.org/10.1007/s00213-013-3423-z

Maguire, J. L., Stell, B. M., Rafizadeh, M., & Mody, I. (2005). Ovarian cycle-linked changes in GABAA receptors mediating tonic inhibition alter seizure susceptibility and anxiety. *Nature Neuroscience, 8*(6), 797–804. https://doi.org/10.1038/nn1469

Mann, E. O., & Mody, I. (2010). Control of hippocampal gamma oscillation frequency by tonic inhibition and excitation of interneurons. *Nature Neuroscience, 13*(2), 205–212. https://doi.org/10.1038/nn.2464

Manyukhina, V. O., Rostovtseva, E. N., Prokofyev, A. O., Obukhova, T. S., Schneiderman, J. F., Stroganova, T. A., & Orekhova, E. V. (2021). Visual gamma oscillations predict sensory sensitivity in females as they do in males. *Scientific Reports, 11*(1), 1–10. https://doi.org/10.1038/s41598-021-91381-2

Micevych, P., & Sinchak, K. (2008). Estradiol regulation of progesterone synthesis in the brain. *Molecular and Cellular Endocrinology, 290*, 44–50. https://doi.org/10.1016/j.mce.2008.04.016

Mody, I., & Maguire, J. (2012). The reciprocal regulation of stress hormones and GABA A receptors. *Frontiers in Cellular Neuroscience, 6*, 1–6. https://doi.org/10.3389/fncel.2012.00004

Monteleone, P., Luisi, S., Tonetti, A., Bernardi, F., Genazzani, A. D., Luisi, M., Petraglia, F., & Genazzani, A. R. (2000). Allopregnanolone concentrations and premenstrual syndrome. *European Journal of Endocrinology, 142*(3), 269–273. https://doi.org/10.1530/eje.0.1420269

Mulleners, W. M., Chronicle, E. P., Palmer, J. E., Koehler, P. J., & Vredeveld, J. W. (2001). Visual cortex excitability in migraine with and without aura. *Headache, 41*(6), 565–572. https://doi.org/10.1046/j.1526-4610.2001.041006565.x

Munday, M. R., Brush, M. G., & Taylor, R. W. (1981). Correlations between progesterone, oestradiol and aldosterone levels in the premenstrual syndrome. *Clinical Endocrinology*, *14*(1), 1–9. https://doi.org/10.1111/j.1365-2265.1981.tb00359.x

Muthukumaraswamy, S. D., & Singh, K. D. (2013). Visual gamma oscillations: The effects of stimulus type, visual field coverage and stimulus motion on MEG and EEG recordings. *NeuroImage*, *69*, 223–230. https://doi.org/10.1016/j.neuroimage.2012.12.038

Muthukumaraswamy, S. D., Singh, K. D., Swettenham, J. B., & Jones, D. K. (2010). Visual gamma oscillations and evoked responses: Variability, repeatability and structural MRI correlates. *NeuroImage*, *49*(4), 3349–3357. https://doi.org/10.1016/j.neuroimage.2009.11.045

Neill Epperson, C., Haga, K., Mason, G. F., Sellers, E., Gueorguieva, R., Zhang, W., Weiss, E., Rothman, D. L., & Krystal, J. H. (2002). Cortical γ-aminobutyric acid levels across the menstrual cycle in healthy women and those with premenstrual dysphoric disorder: A proton magnetic resonance spectroscopy study. *Archives of General Psychiatry*, *59*(9), 851–858. https://doi.org/10.1001/archpsyc.59.9.851

Nelson, S. B., & Valakh, V. (2015). Excitatory/Inhibitory Balance and Circuit Homeostasis in Autism Spectrum Disorders. *Neuron*, *87*(4), 684–698. https://doi.org/10.1016/j.neuron.2015.07.033

Niederhofer, H. (2009). Repetitive Transcranial Magnetic Stimulation for Premenstrual Dysphoric Disorder: A Case Report. *Physicians Postgraduate Press*, *11*(2), 84–85.

Nienborg, H., Hasenstaub, A., Nauhaus, I., Taniguchi, H., Josh Huang, Z., & Callaway, E. M. (2013). Contrast dependence and differential contributions from somatostatin- and parvalbumin-expressing neurons to spatial integration in mouse V1. *Journal of Neuroscience*, *33*(27), 11145–11154. https://doi.org/10.1523/JNEUROSCI.5320-12.2013

Nurminen, L., & Angelucci, A. (2014). Multiple components of surround modulation in primary visual cortex: Multiple neural circuits with multiple functions? *Vision Research*, *104*, 47–56. https://doi.org/10.1016/j.visres.2014.08.018

Nurminen, L., Merlin, S., Bijanzadeh, M., Federer, F., & Angelucci, A. (2018). Top-down feedback controls spatial summation and response amplitude in primate visual cortex. *Nature Communications*, *9*(1). https://doi.org/10.1038/s41467-018-04500-5

Nyberg, S., Wahlström, G., Bäckström, T., & Poromaa, I. S. (2004). Altered sensitivity to alcohol in the late luteal phase among patients with premenstrual dysphoric disorder. *Psychoneuroendocrinology*, *29*(6), 767–777. https://doi.org/10.1016/S0306-4530(03)00121-5

Orekhova, E. V., Butorina, A. V., Sysoeva, O. V., Prokofyev, A. O., Nikolaeva, A. Y., & Stroganova, T. A. (2015). Frequency of gamma oscillations in humans is modulated by velocity of visual motion. *Journal of Neurophysiology*, *114*(1), 244–255. https://doi.org/10.1152/jn.00232.2015

Orekhova, E. V., Prokofyev, A. O., Nikolaeva, A. Y., Schneiderman, J. F., & Stroganova, T. A. (2020). Additive effect of contrast and velocity suggests the role of strong excitatory drive in suppression of visual gamma response. *PLoS ONE*, *15*(2), 1–23. https://doi.org/10.1371/journal.pone.0228937

Orekhova, E. V., Rostovtseva, E. N., Manyukhina, V. O., Prokofiev, A. O., Obukhova, T. S., Nikolaeva, A. Y., Schneiderman, J. F., & Stroganova, T. A. (2020). Spatial suppression in visual motion perception is driven by inhibition: Evidence from MEG gamma oscillations. *NeuroImage*, *213*, 116753. https://doi.org/10.1016/j.neuroimage.2020.116753

Orekhova, E. V., Stroganova, T. A., Schneiderman, J. F., Lundström, S., Riaz, B., Sarovic, D., Sysoeva, O. V., Brant, G., Gillberg, C., & Hadjikhani, N. (2019). Neural gain control measured through cortical gamma oscillations is associated with sensory sensitivity. *Human Brain Mapping*, *40*(5), 1583–1593. https://doi.org/10.1002/hbm.24469

Orekhova, E. V., Sysoeva, O. V., Schneiderman, J. F., Lundström, S., Galuta, I. A., Goiaeva, D. E., Prokofyev, A. O., Riaz, B., Keeler, C., Hadjikhani, N., Gillberg, C., & Stroganova, T. A. (2018). Input-dependent modulation of MEG gamma oscillations reflects gain control in the visual cortex. *Scientific Reports*, *8*(1), 1–13. https://doi.org/10.1038/s41598-018-26779-6

Ossewaarde, L., Hermans, E. J., van Wingen, G. A., Kooijman, S. C., Johansson, I. M., Bäckström, T., & Fernández, G. (2010). Neural mechanisms underlying changes in stress-sensitivity across the menstrual cycle. *Psychoneuroendocrinology*, *35*(1), 47–55. https://doi.org/10.1016/j.psyneuen.2009.08.011

Parry, B. L., Javeed, S., Laughlin, G. A., Hauger, R., & Clopton, P. (2000). Cortisol circadian rhythms during the menstrual cycle and with sleep deprivation in premenstrual dysphoric disorder and normal control subjects. *Biological Psychiatry*, *48*(9), 920–931. https://doi.org/10.1016/S0006-3223(00)00876-3

Pavlov, I., Savtchenko, L. P., Song, I., Koo, J., Pimashkin, A., Rusakov, D. A., & Semyanov, A. (2014). Tonic GABAA conductance bidirectionally controls interneuron firing pattern and synchronization in the CA3 hippocampal network. *Proceedings of the National Academy of Sciences of the United States of America*, *111*(1), 504–509. https://doi.org/10.1073/pnas.1308388110

Perry, G., Hamandi, K., Brindley, L. M., Muthukumaraswamy, S. D., & Singh, K. D. (2013). The properties of induced gamma oscillations in human visual cortex show individual variability in their dependence on stimulus size. *NeuroImage*, *68*, 83–92. https://doi.org/10.1016/j.neuroimage.2012.11.043

Petersen, N., Ghahremani, D. G., Rapkin, A. J., Berman, S. M., Wijker, N., Liang, L., & London, E. D. (2019). Resting-state functional connectivity in women with PMDD. *Translational Psychiatry*, *9*(1). https://doi.org/10.1038/s41398-019-0670-8

Porciatti, V., Bonanni, P., Fiorentini, A., & Guerrini, R. (2000). Lack of cortical contrast gain control in human photosensitive epilepsy. *Nature Neuroscience*, *3*(3), 259–263. https://doi.org/10.1038/72972

Purdy, R. H., Morrow, A. L., Moore, P. H., & Paul, S. M. (1991). Stress-induced elevations of γ-aminobutyric acid type a receptor-active steroids in the rat brain. *Proceedings of the National Academy of Sciences of the United States of America*, *88*(10), 4553–4557. https://doi.org/10.1073/pnas.88.10.4553

R Core Team. (2013). R: A language and environment for statistical computing. *R Foundation for Statistical Computing, Vienna, Austria*. https://doi.org/10.5005/jp/books/11435_12

Ray, S., & Maunsell, J. H. R. (2015). Do gamma oscillations play a role in cerebral cortex? *Trends in Cognitive Sciences, 19*(2), 78–85. https://doi.org/10.1016/j.tics.2014.12.002

Reddy, D. S. (2010). Neurosteroids: Endogenous role in the human brain and therapeutic potentials. *Progress in Brain Research, 186*, 113–137. https://doi.org/10.1016/B978-0-444-53630-3.00008-7

Rodriguez, R., Kallenbach, U., Singer, W., & Munk, M. H. J. (2004). Short- and long-term effects of cholinergic modulation on gamma oscillations and response synchronization in the visual cortex. *Journal of Neuroscience, 24*(46), 10369–10378. https://doi.org/10.1523/JNEUROSCI.1839-04.2004

Rossetti, M. F., Cambiasso, M. J., Holschbach, M. A., & Cabrera, R. (2016). Oestrogens and Progestagens: Synthesis and Action in the Brain. *Journal of Neuroendocrinology, 28*(7), 1–11. https://doi.org/10.1111/jne.12402

Salelkar, S., Somasekhar, G. M., & Ray, S. (2018). Distinct frequency bands in the local field potential are differently tuned to stimulus drift rate. *Journal of Neurophysiology, 120*(2), 681–692. https://doi.org/10.1152/jn.00807.2017

Sato, N., & Miyake, S. (2004). Cardiovascular reactivity to mental stress: Relationship with menstrual cycle and gender. *Journal of Physiological Anthropology and Applied Human Science, 23*(6), 215–223. https://doi.org/10.2114/jpa.23.215

Schloemer, N., Lenz, M., Tegenthoff, M., Dinse, H. R., & Höffken, O. (2020). Parallel modulation of intracortical excitability of somatosensory and visual cortex by the gonadal hormones estradiol and progesterone. *Scientific Reports, 10*(1), 1–13. https://doi.org/10.1038/s41598-020-79389-6

Schmidt, P. J., Nieman, L. K., Danaceau, M. A., Adams, L. F., & Rubinow, D. R. (1998). With and in Those Without Premenstrual Syndrome. *The New England Journal of Medicine, 338*(4), 209–216.

Schroeder, A., Hudson, M., Du, X., Wu, Y. W. C., Nakamura, J., van den Buuse, M., Jones, N. C., & Hill, R. A. (2017). Estradiol and raloxifene modulate hippocampal gamma oscillations during a spatial memory task. *Psychoneuroendocrinology, 78*, 85–92. https://doi.org/10.1016/j.psyneuen.2017.01.022

Schwarz, J. M., Liang, S. L., Thompson, S. M., & McCarthy, M. M. (2008). Estradiol Induces Hypothalamic Dendritic Spines by Enhancing Glutamate Release: A Mechanism for Organizational Sex Differences. *Neuron, 58*(4), 584–598. https://doi.org/10.1016/j.neuron.2008.03.008

Seippel, L., & Bäckström, T. (1998). Luteal-phase estradiol relates to symptom severity in patients with premenstrual syndrome. *Journal of Clinical Endocrinology and Metabolism, 83*(6), 1988–1992. https://doi.org/10.1210/jc.83.6.1988

Sergeeva, E. G., Espinosa-Garcia, C., Atif, F., Pardue, M. T., & Stein, D. G. (2018). Neurosteroid allopregnanolone reduces ipsilateral visual cortex potentiation following unilateral optic nerve injury. *Experimental Neurology*, *306*, 138–148. https://doi.org/10.1016/j.expneurol.2018.05.005

Serrano-Pedraza, I., Romero-Ferreiro, V., Read, J. C. A., Diéguez-Risco, T., Bagney, A., Caballero-González, M., Rodríguez-Torresano, J., & Rodriguez-Jimenez, R. (2014). Reduced visual surround suppression in schizophrenia shown by measuring contrast detection thresholds. *Frontiers in Psychology*, *5*, 1–10. https://doi.org/10.3389/fpsyg.2014.01431

Shankara-Narayana, N., Zawada, S., Walters, K. A., Desai, R., Marren, A., & Handelsman, D. J. (2016). Accuracy of a Direct Progesterone Immunoassay. *The Journal of Applied Laboratory Medicine*, *1*(3), 294–299. https://doi.org/10.1373/jalm.2016.020123

Shin, S. Y., Han, T. H., Lee, S. Y., Han, S. K., Park, J. B., Erdelyi, F., Szabo, G., & Ryu, P. D. (2011). Direct corticosteroid modulation of GABAergic neurons in the anterior hypothalamic area of GAD65-eGFP mice. *Korean Journal of Physiology and Pharmacology*, *15*(3), 163–169. https://doi.org/10.4196/kjpp.2011.15.3.163

Smith, M. J., Adams, L. F., Schmidt, P. J., Rubinow, D. R., & Wassermann, E. M. (2002). Effects of ovarian hormones on human cortical excitability. *Annals of Neurology*, *51*(5), 599–603. https://doi.org/10.1002/ana.10180

Smith, M. J., Adams, L. F., Schmidt, P. J., Rubinow, D. R., & Wassermann, E. M. (2003). Abnormal luteal phase excitability of the motor cortex in women with premenstrual syndrome. *Biological Psychiatry*, *54*(7), 757–762. https://doi.org/10.1016/S0006-3223(02)01924-8

Smith, M. J., Keel, J. C., Greenberg, B. D., Adams, L. F., Schmidt, P. J., Rubinow, D. A., & Wassermann, E. M. (1999). Menstrual cycle effects on cortical excitability. *Neurology*, *53*(9), 2069–2072. https://doi.org/10.1212/wnl.53.9.2064

Spielberger, C. D. (1983). *State-Trait Anxiety Inventory for Adults*. Sampler set. Palo Alto, CA: Mind Garden. https://kristenbeckler.weebly.com/uploads/9/2/9/1/92910342/stai.pdf

Steiner, M., Peer, M., MacDougall, M., & Haskett, R. (2011). The premenstrual tension syndrome rating scales: An updated version. *Journal of Affective Disorders*, *135*(1–3), 82–88. https://doi.org/10.1016/j.jad.2011.06.058

Steiner, M., & Streiner, D. L. (2005). Validation of a revised visual analog scale for premenstrual mood symptoms: Results from prospective and retrospective trials. *Canadian Journal of Psychiatry*, *50*(6), 327–332. https://doi.org/10.1177/070674370505000607

Stell, B. M., Brickley, S. G., Tang, C. Y., Farrant, M., & Mody, I. (2003). Neuroactive steroids reduce neuronal excitability by selectively enhancing tonic inhibition mediated by δ subunit-containing GABAA receptors. *Proceedings of the National Academy of Sciences of the United States of America*, *100*(24), 14439–14444. https://doi.org/10.1073/pnas.2435457100

Sumner, R. L., McMillan, R. L., Shaw, A. D., Singh, K. D., Sundram, F., & Muthukumaraswamy, S. D. (2018). Peak visual gamma frequency is modified across the healthy menstrual cycle. *Human Brain Mapping*, *39*(8), 3187–3202. https://doi.org/10.1002/hbm.24069

Sundstrom, I., Ashbrook, D., & Backstrom, T. (1997). Reduced benzodiazepine sensitivity in patients with premenstrual syndrome: a pilot study. *Psychoneuroendocrinology*, *22*(1), 25–38.

Sundström, I., & Bäckström, T. (1998). Patients with premenstrual syndrome have decreased saccadic eye velocity compared to control subjects. *Biological Psychiatry, 44*(8), 755–764. https://doi.org/10.1016/S0006-3223(98)00012-2

Sy, J. L., Tomarken, A. J., Patel, V., & Blake, R. (2016). The time course of binocular rivalry during the phases of the menstrual cycle. *Journal of Vision, 16*(15), 1–19. https://doi.org/10.1167/16.15.22

Sysoeva, O. V., Galuta, I. A., Davletshina, M. S., Orekhova, E. V., & Stroganova, T. A. (2017). Abnormal size-dependent modulation of motion perception in children with Autism Spectrum Disorder (ASD). *Frontiers in Neuroscience, 11*, 1–13. https://doi.org/10.3389/fnins.2017.00164

Tadin, D. (2015). Suppressive mechanisms in visual motion processing: From perception to intelligence. *Vision Research*, *115*, 58–70. https://doi.org/10.1016/j.visres.2015.08.005

Tadin, D., Kim, J., Doop, M. L., Gibson, C., Lappin, J. S., Blake, R., & Park, S. (2006). Weakened center-surround interactions in visual motion processing in schizophrenia. *Journal of Neuroscience, 26*(44), 11403–11412. https://doi.org/10.1523/JNEUROSCI.2592-06.2006

Tadin, D., Lappin, J. S., Gilroy, L. A., & Blake, R. (2003). Perceptual consequences of centre-surround antagonism in visual motion processing. *Nature, 424*(6946), 312–315. https://doi.org/10.1038/nature01800

Tadin, D., Park, W. J., Dieter, K. C., Melnick, M. D., Lappin, J. S., & Blake, R. (2019). Spatial suppression promotes rapid figure-ground segmentation of moving objects. *Nature Communications*, *10*(1), 1–12. https://doi.org/10.1038/s41467-019-10653-8

Taulu, S., & Simola, J. (2006). Spatiotemporal signal space separation method for rejecting nearby interference in MEG measurements. *Physics in Medicine and Biology, 51*(7), 1759–1768. https://doi.org/10.1088/0031-9155/51/7/008

Timby, E., Bäckström, T., Nyberg, S., Stenlund, H., Wihlbäck, A. C. N., & Bixo, M. (2016). Women with premenstrual dysphoric disorder have altered sensitivity to allopregnanolone over the menstrual cycle compared to controls—a pilot study. *Psychopharmacology, 233*(11), 2109–2117. https://doi.org/10.1007/s00213-016-4258-1

Timby, E., Balgård, M., Nyberg, S., Spigset, O., Andersson, A., Porankiewicz-Asplund, J., Purdy, R. H., Zhu, D., Bäckström, T., & Poromaa, I. S. (2006). Pharmacokinetic and behavioral effects of allopregnanolone in healthy women. *Psychopharmacology, 186*(3), 414–424. https://doi.org/10.1007/s00213-005-0148-7

Towers, S. K., Gloveli, T., Traub, R. D., Driver, J. E., Engel, D., Fradley, R., Rosahl, T. W., Maubach, K., Buhl, E. H., & Whittington, M. A. (2004). α5 subunit-containing GABAA receptors affect the dynamic range of mouse hippocampal kainate-induced gamma frequency oscillations in vitro. *Journal of Physiology, 559*(3), 721–728. https://doi.org/10.1113/jphysiol.2004.071191

Tsai, J. J., Norcia, A. M., Ales, J. M., & Wade, A. R. (2011). Contrast gain control abnormalities in idiopathic generalized epilepsy. *Annals of Neurology*, *70*(4), 574–582. https://doi.org/10.1002/ana.22462

Tschudin, S., Bertea, P. C., & Zemp, E. (2010). Prevalence and predictors of premenstrual syndrome and premenstrual dysphoric disorder in a population-based sample. *Archives of Women's Mental Health*, *13*(6), 485–494. https://doi.org/10.1007/s00737-010-0165-3

Turkmen, S., Backstrom, T., Wahlstrom, G., Andreen, L., & Johansson, I. M. (2011). Tolerance to allopregnanolone with focus on the GABA-A receptor. *British Journal of Pharmacology*, *162*(2), 311–327. https://doi.org/10.1111/j.1476-5381.2010.01059.x

Van Veen, B. D., Van Drongelen, W., Yuchtman, M., & Suzuki, A. (1997). Localization of brain electrical activity via linearly constrained minimum variance spatial filtering. *IEEE Transactions on Biomedical Engineering*, *44*(9), 867–880. https://doi.org/10.1109/10.623056

Van Wingen, G., Van Broekhoven, F., Verkes, R. J., Petersson, K. M., Bäckström, T., Buitelaar, J., & Fernández, G. (2007). How progesterone impairs memory for biologically salient stimuli in healthy young women. *Journal of Neuroscience*, *27*(42), 11416–11423. https://doi.org/10.1523/JNEUROSCI.1715-07.2007

Vieta, E., Sánchez-Moreno, J., Bulbena, A., Chamorro, L., Ramos, J. L., Artal, J., Pérez, F., Oliveras, M. A., Valle, J., Lahuerta, J., & Angst, J. (2007). Cross validation with the mood disorder questionnaire (MDQ) of an instrument for the detection of hypomania in Spanish: The 32 item hypomania symptom check list (HCL-32). *Journal of Affective Disorders*, *101*(1–3), 43–55. https://doi.org/10.1016/j.jad.2006.09.040

Vinck, M., Womelsdorf, T., & Fries, P. (2013). Gamma-band synchronization and information transmission. *Principles of Neural Coding*, 449–470. https://doi.org/10.1201/b14756

Virtanen, P., Gommers, R., Oliphant, T. E., Haberland, M., Reddy, T., Cournapeau, D., Burovski, E., Peterson, P., Weckesser, W., Bright, J., van der Walt, S. J., Brett, M., Wilson, J., Millman, K. J., Mayorov, N., Nelson, A. R. J., Jones, E., Kern, R., Larson, E., … Vázquez-Baeza, Y. (2020). SciPy 1.0: fundamental algorithms for scientific computing in Python. *Nature Methods*, *17*(3), 261–272. https://doi.org/10.1038/s41592-019-0686-2

Wickmann, F., Stephani, C., Czesnik, D., Klinker, F., Timäus, C., Chaieb, L., Paulus, W., & Antal, A. (2015). Prophylactic treatment in menstrual migraine: A proof-of-concept study. *Journal of the Neurological Sciences*, *354*(1–2), 103–109. https://doi.org/10.1016/j.jns.2015.05.009

Wikman, A., Sacher, J., Bixo, M., Hirschberg, A. L., Kopp Kallner, H., Epperson, C. N., Comasco, E., & Sundström Poromaa, I. (2022). Prevalence and correlates of current suicidal ideation in women with premenstrual dysphoric disorder. *BMC Women's Health*, *22*(1), 1–7. https://doi.org/10.1186/s12905-022-01612-5

Wohlfarth, K. M., Bianchi, M. T., & Macdonald, R. L. (2002). Enhanced neurosteroid potentiation of ternary GABAA receptors containing the δ subunit. *Journal of Neuroscience*, *22*(5), 1541–1549. https://doi.org/10.1523/jneurosci.22-05-01541.2002

Won, D., Kim, W., Chaovalitwongse, W. A., & Tsai, J. J. (2017). Altered visual contrast gain control is sensitive for idiopathic generalized epilepsies. *Clinical Neurophysiology*, *128*(2), 340–348. https://doi.org/10.1016/j.clinph.2016.12.008

Wu, X., Gangisetty, O., Carver, C. M., & Reddy, D. S. (2013). Estrous cycle regulation of extrasynaptic d-containing GABAA receptor-mediated tonic inhibition and limbic epileptogenesis. *Journal of Pharmacology and Experimental Therapeutics*, *346*(1), 146–160. https://doi.org/10.1124/jpet.113.203653

Wu, Y. W. C., Du, X., Van den Buuse, M., & Hill, R. A. (2014). Sex differences in the adolescent developmental trajectory of parvalbumin interneurons in the hippocampus: A role for estradiol. *Psychoneuroendocrinology*, *45*, 167–178. https://doi.org/10.1016/j.psyneuen.2014.03.016

Yang, Q., Sjölander, A., Li, Y., Viktorin, A., Bertone-Johnson, E. R., Ye, W., Fang, F., Valdimarsdóttir, U. A., & Lu, D. (2021). Clinical indications of premenstrual disorders and subsequent risk of injury: a population-based cohort study in Sweden. *BMC Medicine*, *19*(1), 1–12. https://doi.org/10.1186/s12916-021-01989-4

Yilmaz, H., Erkin, E. F., Mavioğlu, H., & Sungurtekin, Ü. (1998). Changes in pattern reversal evoked potentials during menstrual cycle. *International Ophthalmology*, *22*(1), 27–30. https://doi.org/10.1023/A:1006165126702

Yizhar, O., Fenno, L. E., Prigge, M., Schneider, F., Davidson, T. J., Ogshea, D. J., Sohal, V. S., Goshen, I., Finkelstein, J., Paz, J. T., Stehfest, K., Fudim, R., Ramakrishnan, C., Huguenard, J. R., Hegemann, P., & Deisseroth, K. (2011). Neocortical excitation/inhibition balance in information processing and social dysfunction. *Nature*, *477*(7363), 171–178. https://doi.org/10.1038/nature10360

Zhang, S., Xu, M., Kamigaki, T., Do, J. P. H., Chang, W. C., Jenvay, S., Miyamichi, K., Luo, L., & Dan, Y. (2014). Long-range and local circuits for top-down modulation of visual cortex processing. *Science*, *345*(6197), 660–665. https://doi.org/10.1126/science.1254126

Zhuang, X., Chen, Y., Zhuang, X., Xing, T., Chen, T., Jiang, G., & Yang, X. (2017). Impaired center-surround suppression in patients with Alzheimer's disease. *Journal of Alzheimer's Disease*, *55*(3), 1101–1108. https://doi.org/10.3233/JAD-160603

Zhang, S., Xu, M., Kamigaki, T., Do, J. P. H., Chang, W. C., Jenvay, S., Miyamichi, K., Luo, L., & Dan, Y. (2014). Long-range and local circuits for top-down modulation of visual cortex processing. *Science*, *345*(6197), 660–665. https://doi.org/10.1126/science.1254126

Zhuang, X., Chen, Y., Zhuang, X., Xing, T., Chen, T., Jiang, G., & Yang, X. (2017). Impaired center-surround suppression in patients with Alzheimer's disease. *Journal of Alzheimer's Disease*, *55*(3), 1101–1108. https://doi.org/10.3233/JAD-160603

Zilles, K., Palomero-Gallagher, N., Grefkes, C., Scheperjans, F., Boy, C., Amunts, K., & Schleicher, A. (2002). Architectonics of the human cerebral cortex and transmitter receptor fingerprints: Reconciling functional neuroanatomy and neurochemistry. *European Neuropsychopharmacology*, *12*(6), 587–599. https://doi.org/10.1016/S0924-977X(02)00108-6