Deleterious effects of thermal and water stresses on life history and

physiology: a case study on woodlouse

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Abstract

We tested independently the influences of increasing temperature and decreasing moisture on life history and physiological traits in the arthropod *Armadillidium vulgare.* Both increasing temperature and decreasing moisture led individual body mass and reproductive success to decrease. While the density of immune cells decreased and the β-galactosidase activity increased with increasing temperature and decreasing moisture, which suggests a negative impact of these stressors on individual performance, increased temperature and decreased moisture affected differently the other biomarkers conjuring different underlying mechanisms depending on the stress applied. Our findings demonstrate overall a negative impact of high temperature and low moisture on woodlouse welfare. Changing temperature or moisture had slightly different effects, illustrating the need to test further the respective role of each of these key components of climate change on organisms to predict more reliably the future of our ecosystems.

Key words

Abiotic stresses, life history traits, physiological traits, arthropods, climate change

Conflict of interest disclosure

The authors declare they have no conflict of interest relating to the content of this article.

Introduction

The Intergovernmental Panel on Climate Change (IPCC) forecasts an average increase in temperature between +1.5°C and +4°C in 2100 (Masson-Delmotte et al., 2021). Not only will average temperatures and the frequency and intensity of precipitation change, but extreme events will increase in frequency. Although the link between global warming and drought is still highly debated and may not be direct (Trenberth et al., 2014), droughts due to a decrease in rainfall and an increase in evaporation are expected to take place in the coming decades (Dai, 2013), and should be much more intense than current droughts (Trenberth et al., 2014). As deterioration of environmental conditions are known to impact life history traits such as growth rate, reproductive success, or longevity (e.g. Chen et al., 2019; Johnson and Jones, 2016; Khadioli et al., 2014), identifying the potential implications of climate change for organisms is a research question of paramount importance.

Terrestrial arthropods are ectotherms that are particularly sensitive to temperature and moisture changes (Lister and Garcia, 2018; Maron et al., 2015). Global warming constitutes a threat for them (Johnson and Jones, 2016). In Lepidoptera, for example, too high temperatures prevent hatching (Khadioli et al., 2014). In both Lepidoptera and Hymenoptera, increasing temperature beyond the optimum can have detrimental effect on survival (Abou-Shaara et al., 2012; Khadioli et al., 2014). In some Coleoptera, egg viability decreases and hatching time increases for viable eggs when they are exposed to drought (Johnson et al., 2010). When facing the costs of increased temperature and drought frequency on life history traits, arthropods display different responses to resist or tolerate such changes (Strachan et al., 2015). For example, some arthropods can migrate to refugia, others can implement physiological resistance tactics (e.g. resistant eggs) and/or dormancy, and others are able to alter their life cycle and/or development (Strachan et al., 2015; Verberk et al., 2008). In organisms with limited movement capacity, increased temperature and decreased moisture are expected to induce pronounced physiological stresses. Studying how these stresses affect both life history and physiological traits would allow us to anticipate the effect of global warming on organisms with limited movement capacity.

The common woodlouse *Armadillidium vulgare* is a key soil decomposer naturally exposed to a wide range of environmental conditions (Souty-Grosset et al., 1988) that provides major ecosystem services (David and Handa, 2010), notably in agrosystems and grassland habitats

where it is used as an ecological indicator. In the course of its evolutionary history, the
common woodlouse had to adapt to terrestrial life. Consequently, it is still very sensitive to
moisture and temperature variations, which can induce water loss (Smigel and Gibbs, 2008)
and have major consequences in terms of distribution, behavior and survival (Hassall et al.,
2018; Paris and Pitelka, 1962). Moreover, their movement capacity is low (i.e. several
hundred meters during the entire lifetime at the best, Durand et al., 2019) to allow them to
migrate so to avoid the stress imposed by the environment. Our knowledge and ability to
measure woodlouse life history traits and the availability of molecular and cellular
biomarkers of individual quality (Depeux et al., 2020a) make this species a highly relevant
experimental model to study the influence of both temperature and moisture on life history
and physiological traits.

In this study, we tested independently the effects of increased temperature (experiment 1)
and of decreased moisture (experiment 2) on a selected set of key life history (i.e. growth,
reproductive success, and survival) and physiological (i.e. immune cell parameters (cell
viability, cell density and cell size) and β-galactosidase activity) traits in woodlouse. In
experiment 1 (i.e. testing the effect of increased temperature), we compared individuals
maintained at 20°C and 80% of moisture (i.e. the standard temperature and moisture
laboratory conditions) to individuals exposed at 28°C (simulating a temperature increase of
8°C) still at 80% of moisture. In experiment 2 (i.e. testing the effect of decreased moisture),
we compared animals in standard conditions to individuals exposed at 50% of moisture
(simulating a moisture loss of 30%) still at 20°C. We hypothesized that a rise in temperature
and a loss in moisture should be stressful and should induce changes in life history and
physiological traits.

MATERIALS & METHODS

Biological Material – Routine Breeding

All specimens of *A. vulgare* used in this study descend from individuals sampled in Denmark
(Helsingör) in 1982. Since then, animals were reared under laboratory conditions under the
natural photoperiod of Poitiers (France) (46°35'N; 0°20'E), at 20°C and about 80-85% of
moisture, in plastic boxes (length × width × height: 26.5 × 13.5 × 7.5 cm) containing humid
loam, and fed ad libitum with carrot slices and dried linden leaves. Controlled breeding, for

the maintenance of the lineage over years, is performed in individual boxes (diameter x

height: 9,8cm x 4,9cm), with reproductive pairs selected from their pedigree to avoid

inbreeding. One month after mating, offspring exit the female *marsupium* (i.e. female

ventral pouch on which the eggs develop) (Suzuki and Ziegler, 2005). We transferred these

offspring a few days after birth into a bigger box (length × width × height: 26.5 × 13.5 × 7.5

cm) with loam and food. After 3 months, once sexual characters have appeared but before

sexual maturity, we placed young males and females in separate boxes (length × width ×

height: 26.5 × 13.5 × 7.5 cm) in laboratory conditions described above, enabling us to obtain

virgin adults. For the maintenance of the lineage, about 40 crosses have been performed

following this protocol each year. Each of the 40 broods provides at least one breeder for the

next generation. The animals used in the experiments of this study came from this controlled

lineage.

Experimental Design

Experiment 1: effect of increased temperature on life history and physiological traits

The experiment 1 performed in January 2019 involved the comparison between two groups

of animals aged of 7 months old maintained at different temperatures in two climatic

chambers (Memmert HPP 256L with LED Light module cold white 6500K for HPP260 (15%)

and Interior IP68 socket (for temperature restriction)) during two months after standard

conditions of maintenance:

(i)   The "Control Temperature" group (CT) of animals maintained in standard
conditions (i.e. at 20°C and 80% of moisture) in one of our two climatic chambers.
(ii)   The "High Temperature" (HT) group of animals exposed at 28°C (simulating
increased temperature by 8°C (i.e. thermal stress condition)) and at 80% of
moisture in the second climatic chamber.

Eight degrees (i.e. difference in temperature between the two groups) corresponds to a

temperature increase close to daily variations observed in Poitiers during some summers,

which could chronically induce stress. Moreover, we have observed the stressful effect of

this temperature increase in a preliminary experiment in which we did not control the

moisture variation (Depeux et al., 2019).

In each group, animals were fed *ad libitum* in 3 boxes (length × width × height: 26.5 × 13.5 ×

7.5 cm; standard laboratory density conditions) of 30 females and 3 boxes of 30 males from

15 different clutches (i.e. all treatments included animals with the same genetic background

(i.e. issued from 15 same clutches) to be comparable). For each condition, one box was used

to monitor survival and growth (mass gain over time) of animals from the beginning to the

end of the experiment, another was used to evaluate reproductive success and the last box

served to quantify physiological traits (i.e. immune cell parameters: cell viability, cell density,

and cell size) and β-galactosidase activity, see below). In this last box, the animals had to be

sacrificed (see 'Ethical statement' section below) because of the protein extraction on nerve

chains that was required to measure the β-galactosidase activity.

Experiment 2: effect of moisture loss on life history and physiological traits

The experiment 2 performed in January 2021 involved the comparison between two groups

of 7 months old animals maintained under different conditions in our two climatic chambers

during two months after standard conditions of maintenance:

(i)   The "Control Moisture" (CM) group of animals maintained in standard conditions

(i.e. at 20°C and 80% of moisture) in one of our two climatic chambers

(ii)   The "Loss of Moisture" (LM) group of animals exposed at 50% of moisture

(simulating a moisture loss of 30% (i.e. water stress condition)) and at 20°C in the

second climatic chamber.

Similar to the experiment 1, in each group, animals were fed *ad libitum* in 3 boxes of 30

females and 3 boxes (length × width × height: 26.5 × 13.5 × 7.5 cm; standard laboratory

density conditions) of 30 males from 15 different clutches (i.e. all boxes to compare included

animals with the same genetic background (i.e. issued from 15 same clutches)). For each

condition, one box was used to monitor individual survival and growth from the beginning to

the end of the experiment, another was used to evaluate reproductive success and the last

box served to quantify physiological traits (see below).

In our two experiments, we aimed to compare individuals of the same age because age

negatively impacts both reproductive success (Depeux et al., 2020b) and physiological traits

(Depeux et al., 2020a). Having initially only two climatic chambers, we had to perform our

experiments 1 and 2 in different years (i.e. experiment 1 in 2019 and experiment 2 in 2021).

Thus, we systematically compared the effect of each stress against its own control condition

group (i.e. CT for HT and CM for LM). Moreover, at the beginning of each experiment (1 and

2), we selected individuals of the same size and we checked, at the end of the experiments,

potential statistical differences between the two control groups (CT and CM) on measures of

life history and physiological traits (Supp. File1).

Ethical statement

The Decree n°2003-768 from 01/08/2003 and the European Directive 2010/63/EU regulating

animal research does not require any ethical evaluation prior to research on arthropods.

However, we complied with the ethical 3R rules (Replace/Reduce/Refine). Even though it

was impossible to replace the use of animals in our study, we reduced the number of used

animals, optimizing this number to a minimum to ensure a reliable assessment of the effect

of the different stressors on life history and physiological traits. Although individuals were

obviously stressed during the experiments, we made sure that they were provided with

optimal living conditions throughout the experiments. In addition, when the tissue sampling

required the death of individuals (i.e. to measure physiological traits such as β-galactosidase

activity), the animals to be euthanized were frozen before protein extractions to take into

account animal welfare as much as possible.

Life history traits

Survival and growth

One box of males and one box of females from each group (i.e. for the groups CT, HT, CM,

LM) were used to monitor and compare changes of survival and body mass over time. All

individuals in these boxes were monitored for 124 days (i.e. about 4 months). We sampled

individuals at 14, 28, 42, 69, and 124 days (i.e. 5 sampling points per box) and assessed

survivorship and change in body mass (in grams) of all surviving animals in each box (body

mass was measured with a precision balance 650g|1mg Sartorius™ BCE653-1S Entris™ II

Essential). Then, we compared these traits over time and between groups (CT vs. HT groups

and CM vs. LM groups) to test independently the effect of temperature and moisture

changes on these traits (see section on Statistical analyses). Due to regular moults, individual

identification among the 30 animals sharing in given box cannot be performed, leading our

measures to be average survival and growth instead of individual trajectories.

186 **Reproductive success**

187 At the end of the exposure to different conditions, one box of males and one box of females
188 were collected from each group (i.e. for the groups CT, HT, CM, and LM). We formed 20
189 breeding pairs composed of one male and one female per group. Each breeding pair was
190 placed in a box, at 20°C, with food provided *ad libitum* and in a photoperiod of 16:8 (L/D)
191 stimulating the reproduction (Mocquard et al., 1989). We followed all these pairs for 5
192 months during which each clutch produced was recorded. At the end of this period, the
193 ability to produce a clutch (i.e. the probability that one clutch or more is produced by a given
194 pair) for the 80 pairs (i.e. 40 pairs for experiment 1 and 40 pairs for experiment 2) was
195 compared independently between CT and HT groups and between CM and LM groups to test
196 the effect of temperature and moisture changes on breeding success. As we created groups
197 from similar clutches to have the same genetic background among boxes, we cannot exclude
198 that some crosses were composed of related individuals although we expect this event to be
199 rare. However, the probability of forming sibling pairs (8%) was similar among groups that
200 were exposed either at 20°C vs. 28°C or at 80% vs. 50% of moisture.

201 *Physiological traits*

202 At the end of the experimental treatments (i.e. after two months in our experimental
203 conditions), one box of males and one box of females were taken from each group (i.e. for
204 the groups CT, HT, CM, and LM) for measuring the level of our set of physiological traits (i.e.
205 immune cells parameters and β-galactosidase activity) developed in Depeux et al. (2020a).
206 These physiological traits were firstly described as senescence biomarkers because they
207 allow predicting the amount of cellular senescence in different organisms and are strongly
208 age-dependent in *A. vulgare* (i.e. older the individual, higher the decline of these
209 biomarkers, Depeux et al. 2020a). We performed these measures on each remaining animals
210 (Table 1) and we compared these metrics independently between CT and HT groups for
211 experiment 1 and between CM and LM groups for experiment 2 (see section Statistical
212 analyses).

213 Table 1. Numbers of individuals on which we measured quality biomarkers.

Groups	CT (Control temperature)	HT (High temperature)	CM (Control Moisture)	LM (Loss of Moisture)

Numbers of females	13	12	9	15
Numbers of males	17	17	15	15

Immune cells

As immune cells are free-circulating, they can inform about a potential premature biological
aging. When an individual *A. vulgare* ages, its immune cells decrease in density and viability
while increasing in size (Depeux et al., 2020a). To measure these parameters, we collected
3µL of haemolymph per individual and placed it in 15µL of MAS-EDTA (EDTA 9 mM,
Trisodium citrate 27 mM, NaCl 336 mM, Glucose 115 mM, pH 7, (Rodriguez et al., 1995)). We
then added 6µL of Trypan Blue at 0.4% (Invitrogen) to discriminate live and dead cells. After,
10µL of this solution was put in Invitrogen Coutness® counting slide and put in an automated
Cell Counter (Invitrogen) to quantify cell density (measured as the number of cells per mL of
haemolymph), viability (measured as the proportion of live cells) as well as cell size (in µm).
These three parameters of the immune cells are physiological traits that were found to be
reliable biomarkers of cellular senescence in *A. vulgare* (Depeux et al., 2020a).

β-galactosidase activity

The β-galactosidase activity is a physiological trait commonly used as a marker of cellular
senescence (Lee et al., 2006). Its indirect activity in regards to the process of cellular
senescence increases with age in *A. vulgare* (Depeux et al., 2020a). To measure this
enzymatic activity, we dissected and removed the nerve cord of each individual after having
collected haemolymph for assessing the immune parameters. We put individual nerve cords
in 300µL of Lyse Buffer 1X (CHAPS 5 mM, Citric acid 40 mM, Sodium Phosphate 40 mM,
Benzamidine 0.5 mM, PMSF 0.25 mM, pH = 6) (Gary and Kindell, 2005). We centrifuged the
sample at 15 000g for 30 minutes at 4°C and then we collected and kept the supernatant at -
80°C. We quantified the protein concentration thanks to the BCA Assay and we
homogenized all samples at the 0.1mg/mL protein concentration. Then, 100µL of these
protein extracts were added to 100µL of reactive 4-methylumbelliferyl-D-galactopyranoside
(MUG) solution. The synthesis of the fluorescent 4-methylumbelliferone (4-MU), the result
of the contact of MUG reactive with β-galactosidase, was measured using the multimode
microplate reader Mithras LB940 133 HTS III, Berthold; excitation filter: 120 nm, emission

filter 460 nm, for 120 minutes. We included two technical replicates for each sample to
obtain the measures.

Statistical analyses

All statistical analyses were performed using the software R 4.2.1 (R core Team 2022).

The effects of the stress condition (control vs. high temperature, or control vs. low moisture)
on life history and physiological traits were tested using the following models. *(i)* Life history
traits. For the survival data, Cox proportional hazard models were fitted with stress
condition, sex and their interaction term as fixed variables, using the 'survival' package
(Therneau, 2022). For the growth data, the body mass was modelled using linear models
with Gaussian distribution with stress condition, sex, time (i.e. time after placing in climatic
chamber, in days) and their two-by-two interaction term as fixed variables. The female
reproductive success was modelled as binary data (*presence of at least one clutch or absence*
of clutch) using linear regression with a binomial distribution, with stress condition as fixed
variables. *(ii)* Physiological traits. The cell density, cell size, cell viability, and the β-
galactosidase activity were modelled using linear models with Gaussian distribution with
stress condition, sex, and their interaction term as fixed variables.

We proceeded to model selection starting with full (saturated) model. We ranked all nested
models according to their AICc using 'MuMIn' package (Barton, 2022). We selected the most
parsimonious models among the top ranked models (ΔAICc < 2) (Galipaud et al., 2017). The
tables summarizing the model selection procedure was presented in Supp. File2. To
represent the effect of the two environmental stresses in each variable, we presented our
results with indices of size effect. The effect of each stress on each measure of life history
traits and biomarkers of individual quality were measured using the standardized slopes
(Schielzeth et al 2010) and their SE calculated by rescaling the variable of the selected
model. For survival data, the effect size was the hazard ratio, calculated as the exponential
of the regression parameter (Collett 2003). When the selected model did not include the
effect of the stress, we took the model with the variable stress as fixed factor to obtain a size
effect as done in Depeux et al. 2020a.

Data, script and code availability

All datasets and source code are available as electronic supplementary materials on public

repository: https://gitlab.com/fxdm/armadillidium_stress

RESULTS

As said previously, we checked, at the end of the experiments, potential statistical

differences between the two control groups (CT and CM) on measures of life history and

physiological traits (Supp. File1). Although β-galactosidase activity and cell density were

higher in the CT group than in the CM group (Supp. File1), we observed the same dynamics

in these measures in the face of their stressful condition (HT and LM, respectively) (see

Results part). The body mass at day 14 was higher in the CM group than in the CT group

(Supp. File1). Whatever the differences observed between the two control groups (CT group

used in 2019 and CM group used in 2021), we compared the effect of each stress (HT in 2019

and LM in 2021, respectively) against its own control group (CT group in 2019 and CM group

in 2021, respectively) for testing the effect of each stress.

Life history traits

Survival was not impacted by an increased temperature (χ^2_1=2.16, P=0.14, Fig.1A, Supp.

File2 Table S2a, Supp. File3-1.A.1) although mortality risk was almost twice as lower at low

compared to high temperature. The hazard ratio was 1.78, with a 95%CI including 1 [0.81;

3.88]. By contrast, individuals exposed to a water stress had a 2.5 times higher mortality risk

(χ^2_1=4.54, P=0.03, Fig. 1B, Supp. File2 Table S2b, Supp. File3-1.A.2). The hazard ratio was

2.69, with a 95%CI excluding 1 [1.03; 7.01]. As a result, 90% of individuals placed at control

moisture were still alive at the end of the follow-up, whereas only 75% of individuals placed

in water stress condition survived at the end of the experiment.

In both thermal and water stresses, body mass increased during the entire experiment

duration (Fig. 1C and 1D, Supp. File3-1.B.1 and 1.B.2), as expected in an indeterminate

grower as *A. vulgare*, but there was no detectable interaction between day and sex (Supp.

File2 Table S2c and Table S2d). For the temperature experiment, interactions between sex

and stress ($F_{1,528}$=6.90, P=0.0088, Supp. File3), and between day and thermal stress

($F_{1,528}$=14.6, P=0.00015, Supp. File2 Table S2c) showed up, illustrating the impact of an

increasing temperature on growth. By contrast, in the moisture experiment, the body mass

was not affected by the sex ($F_{1,522}$=0.35, P=0.55), the water stress ($F_{1,522}$=0.31, P=0.58), or

any interaction between the variables (all P > 0.10).

The reproductive success markedly decreased in both experiments for the stressful
condition: a fourfold increase of reproductive failure in presence of thermal stress (χ^2_1=5.02,
p=0.025, Odd-ratio=0.23, 95%CI=[0.057;0.83], Fig. 1E, Supp. File2 Table S2e, Supp. File3-
1.C.1), and a fivefold increase of reproductive failure in presence of water stress (χ^2_1=5.38,
p=0.02, Odd-ratio=0.20, 95%CI=[0.045;0.79], Fig. 1F, Supp. File2 Table S2f, Supp. File 3-
1.C.2). In both cases, it corresponds to halving the reproductive success in the stress groups
(water stress: 55% in the control group vs. 20% in the stressed group; thermal stress: 65% in
the control group vs. 30% in the stressed group).

Figure 1. Effect of the two environmental stressors (Temperature (A, C and E) and Moisture (B, D and F) on Survival (A and B), Body mass (C and D) and Reproductive success (E and F). Blue colour: control groups, orange colour: stress groups. CT: Control Temperature (20°C), HT: High Temperature (28°C), CM: Control Moisture (80%), LM: Loss of Moisture (50%). NS: No significant; * p<0.05.

Physiological traits

Immune cells

Immune cell viability was not affected by the thermal stress ($F_{1,56}$=0.92, p=0.34, standardized

slope β=-0.25; 95%CI=[-0.79;0.27], Fig. 2A, Supp. File2 Table S2g and Supp. File3-2.A.1) but

decreased during the water stress ($F_{1,50}$=4.17, p=0.046, standardized slope β=0.55;

95%CI=[0.01;1.09], Fig. 2B, Supp. File2 Table S2h and Supp. File3-2.A.2).

Immune cell size decreased during the thermal stress ($F_{1,55}$=5.72, p=0.02, standardized slope

β=-0.60; 95%CI=[-0.1;-1.1], Fig. 2C, Supp. File2 Table S2i and Supp. File 3-2.B.1) but not under

the water stress ($F_{1,50}$=3.79, p=0.057, standardized slope β=-0.55; 95%CI=[-1.07;0.02], Fig.

2D, Supp. File2 Table S2j and Supp. File3-2.B.2).

Immune cell density decreased during the thermal stress ($F_{1,56}$=38.2, P<0.001, standardized

slope β=-1.26; 95%CI=[-1.67;-0.85], Fig. 2E, Supp. File2 Table S2k and Supp. File3-2.C.1)) and

the water stress ($F_{1,50}$=7.64, p=0.008, standardized slope β=0.72; 95%CI=[0.19;1.25], Fig. 2F,

Supp. File2 Table S2l and Supp. File3-2.C.2).

β-galactosidase activity

The *β*-galactosidase activity increased with the thermal stress ($F_{1,54}$=11.32, P=0.0014,

standardized slope β=0.82; 95%CI=[0.33;1.32], Fig.2G, Supp. File2 Table S2m, and Supp.

File3-2.D.1), but also with the water stress ($F_{1,50}$=10.50, P=0.002, standardized slope β=-0.83;

95%CI=[-1.31;-0.32], Fig. 2H, Supp. File2 Table S2n and Supp. File3-2.D.2).

Figure 2. Effect of the two environmental stressors (Temperature (A, C, E, and G) and Moisture (B, D, F and H) on immune cell viability (A and B), immune cell size (C and D), immune cell density (E and F) and β-galactosidase activity (G and H). Blue colour: control groups, orange colour: stress groups. CT: Control Temperature (20°C), HT: High Temperature (28°C), CM: Control Moisture (80%), LM: Loss of Moisture (50%). NS: No significant; * p<0.05, ** p<0.01, *** p<0.001.

DISCUSSION

Our results highlight that life history traits were negatively impacted by the two environmental stressors (thermal and water stresses) considered in this study. Moreover, the detrimental effects of these stressors on our set of biomarkers of individual quality are consistent with an overall premature ageing of stressed animals compared to unstressed ones. To briefly summarize, an increase in temperature (thermal stress) negatively affects both the body mass trajectory over time and the reproductive success of individuals. A decrease in moisture (water stress) resulted in a decrease of both survival and reproductive success. Concerning our physiological traits: (1) the density of immune cells decreases under both stresses, (2) immune cell size decreases under thermal stress, but is not impacted under water stress, (3) the viability of the cells decreases under water stress (but not under thermal stress) and (4) finally, the β-galactosidase activity increases for the two stressed groups. In this context, our results globally support marked negative effects of thermal and water stresses on woodlouse performance, with some minor differences between the two stressors in their effects on life history and physiological traits.

About the life history traits, if the thermal stress has no detectable effect on survival in *A. vulgare*, contrary to what has been previously reported in arthropods studied so far such as *Antestiopsis thunbergii*, *Calliphora stygia* and *Margaritifera margaritifera* (Azrag et al., 2017; Hassall et al., 2017; Kelly et al., 2013), this stressor leads to a slowdown in woodlouse growth, in line to what has been reported in three other isopods (Angilletta et al., 2004). In parallel, the water stress leads to a decrease in reproductive success, as previously reported in females of *Antestiopsis thunbergii* (Azrag et al., 2017). In *A. vulgare*, individual body size is positively correlated with fecundity (Durand et al., 2018; Lawlor, 1976), meaning that the slowdown in growth could explain, at least partly, the decrease in reproductive success for stressed animals compared to non-stressed ones. Concerning the water stress, if the loss of moisture has no detectable effect on woodlouse growth, it causes a decrease in both survival and reproductive success. These findings suggest a high cost of drought on individual fitness in *A. vulgare*.

About the physiological traits, our results of the thermal stress experiment show that although cell density is negatively impacted by increased temperature, cell viability is not affected. Moreover, contrary to the expectation when individuals are senescent, cell size

decreases instead of increasing. This last result supports our previous finding that cells
decrease in size when the temperature raises (without controlling moisture level, Depeux et
al., 2019). That smaller cells are associated with increased cell renewal in stressed animals
might explain this pattern. On the other hand, the increase of β-galactosidase activity seems
to indicate premature ageing (and thus a decrease in quality) in individuals exposed to
thermal stress. Concerning the water stress, the biomarkers of individual quality indicate a
decrease in cell density and viability, associated with an increase in β-galactosidase activity,
which suggests an acceleration of biological ageing in the individuals exposed to a water
stress (Depeux et al., 2020a).

We reported a global negative effect of the thermal stress in *A. vulgare* in our study, but our
results seem to show an even higher and clearer effect of the water stress on both life
history traits and biomarkers of individual quality. Although the woodlouse has become
terrestrial for a long time, the individuals of that species are still dependent on and require a
substantial water supply (Smigel and Gibbs, 2008). Thus, behaviours like aggregation that
allow individuals to resist to desiccation have been set up and thereby to maintain the rate
of moisture required for survival (Broly et al., 2013; Smigel and Gibbs, 2008). This can explain
the strong effect of water stress in our study. Under natural conditions, increase in
temperature and loss of moisture generally positively covary, leading to even higher
negative consequences on the woodlouse performance. Further work will be required to test
the influence of more extreme and maybe more realistic conditions by simultaneously
increasing temperature and decreasing moisture on life history and physiological traits. A
study in the wild comparing life history and physiological traits on *A. vulgare* collected across
areas with different temperature and drought gradient would also allow a better assessment
of the combined effects of these stressors.

Unlike what happens in nature, our experimental study on a laboratory line of woodlouse allowed us
to test the effect of the thermal and water stresses while controlling for potentially confounding
factors such as individual age. Indeed, it is highly challenging to control for individual age in the wild.
In this context, the use of our controlled laboratory line on which we developed our physiological
markers allowed us to account for the exact age of the animals (which is itself linked to life history
and physiological traits (Depeux et al., 2020b)) and for their genetic origin. We compared groups of
the same origin (and our controlled crosses guarantee the genetic diversity of our line) and of the

same age. This allows us to limit confounding effects as much as possible and to quantify the effects
of the two tested stressors independently.

Thanks to our experimental design that allowed us to test independently the influence of
stressors that organisms are likely to face in the wild, we showed that thermal and water
stress do not have the same impact. Although simulations based on mathematical models
have predicted that both temperature and drought changes overall affect arthropods,
experimental approaches such as reported in this work are required to quantify reliably the
influence of changing conditions on life history and physiological traits (Johnson et al. 2010).
Drought can have serious physiological consequences on invertebrates, involving e.g. protein
denaturation and undesirable macromolecular interactions (Sano et al., 1999; Tang and
Pikal, 2005) or oxidative damage (Lopez-Martinez et al., 2008), which are known to be
associated with cellular senescence (Gilca et al., 2007) and thus in the decreased
performance observed in stressed organisms. Due to the role of arthropods in services to
many ecosystems (e.g. biochemical balance of ecosystems, agriculture, pest management…),
and in the context of global warming, it is crucial to understand the effects of temperature
and moisture changes on these organisms (Santos et al., 2021). As temperature increase is
not the only environmental change expected to take place in the coming years, it is of
paramount importance to assess also the impact of other stressors. Although many
predictive models have been proposed so far, getting more accurate information on the
expected responses of organisms facing with different kinds of stress would provide the
required information to test these model predictions.

To conclude, *A. vulgare* is an important actor that delivers ecosystem services in many
ecosystems because it actively impacts soil fertility (Souty-Grosset and Faberi, 2018) and it is
also used as an ecological indicator of grassland habitats (Paoletti and Hassall, 1999; Souty-
Grosset et al., 2005). This detritivorous species facilitates decomposition processes and
nutrient cycling on which agricultural productivity and sustainability depend (Bredon et al.,
2018; Paoletti and Hassall, 1999), and plays thereby a key role in ecosystem services (David
and Handa, 2010). Extending knowledge in the response of soil biodiversity facing current
global changes could promote sustainability by helping to the development of new tools and
strategies for more efficient management of soils and associated crops, through more
effective and targeted recolonisation and/or restoration of soil biodiversity. Also, to better

understand what the future of the animal communities in the current context of global
warming will be, it is necessary to perform studies on models presenting particular
ecological requirements, such as woodlouse.

Supplementary files

All supplementary files are available on public repository:
https://www.biorxiv.org/content/10.1101/2022.09.26.509512v1.supplementary-material
Supp. File1 Comparison of the two control groups
Supp. File2 Model selection
Supp. File3 Graphical representations of results per sex

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Supplementary File 1: Comparison of the two control groups

Table 1: Comparison between the two control groups (CT (Control Temperature) and CM (Control Moisture)) of the two
experiments for each tested variable (in bold the variables with significant statistical differences with graphical associated
figures (Fig. 1, Fig. 2 and Fig. 3))

Traits	Statistical value	P-value
Life history traits measures		
Survival	$\chi_1^2 = 1.25$	P = 0.26
Body mass **(day 14)**	**$F_{1,111} = 13.00$,**	**P < 0.001**
Reproduction	$\chi_1^2 = 0.417$	P = 0.52
Physiological traits measures		
Immune cells parameters		
Density	**$F_{1,50} = 6.21$**	**P= 0.016**
Viability	$F_{1,50} = 2.49$	P= 0.12
Size	$F_{1,50} = 0.029$	P = 0.86
β-galactosidase activity	$F_{1,49} = 17.0$	**P < 0.001**

589 *Figure 1: Body mass comparison between the two control groups*
590 *(CT (Control Temperature) and CM (Control Moisture)) P-value < 0.001*

Figure 2: Immune cells density comparison between the two control groups
(CT (Control Temperature) and CM (Control Moisture)) P-value=0.02

Figure 3: β-galactosidase activity comparison between the two control groups
(CT (Control Temperature) and CM (Control Moisture)) P-value<0.001

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601 **Supplementary File 2: Model selection**
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Life history trait

Survival

Table S2a. Effect of the thermal stress condition and sex on the survival. For each model, we
reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood
(LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc),
change in AICc (ΔAICc) from the best model, and model weight. The presence of the categorial
variable (sex, stress condition, and their interaction term sex:stress) in the model is indicated by a "+"
symbol.

Intercept	sex	stress	sex:stress	adj.R²	df	logLik	AICc	ΔAICc	weight
+		+		0,08	1	-125,01	252,18	0,00	0,31
+				**0,00**	**0**	**-126,09**	**252,19**	**0,01**	**0,31**
+	+			0,02	1	-125,82	253,81	1,62	0,14
+	+	+		0,10	2	-124,70	253,89	1,71	0,13
+	+	+	+	0,17	3	-123,65	254,34	2,16	0,11

Table S2b. Effect of the water stress condition and sex on the survival. For each model, we reported
intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood (LogLik)
values, Akaike information criteria values with a correction for small sample sizes (AICc), change in
AICc (ΔAICc) from the best model, and model weight. The presence of the categorial variable (sex,
stress condition, and their interaction term sex:stress) in the model is indicated by a "+" symbol. The
most parsimonious model is highlighted in bold font.

Intercept	sex	stress	sex:stress	adj.R²	df	logLik	AICc	ΔAICc	weight
+		**+**		**0,20**	**1**	**-91,25**	**184,72**	**0,00**	**0,53**
+	+	+		0,22	2	-91,02	186,75	2,02	0,19
+				0,00	0	-93,52	187,04	2,32	0,17
+	+			0,02	1	-93,32	188,87	4,15	0,07
+	+	+	+	0,23	3	-90,94	189,39	4,67	0,05

Body mass

Table S2c. Effect of the thermal stress condition, sex and day on body mass. For each model, we reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood (LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc), change in AICc (ΔAICc) from the best model, and model weight. The presence of the categorial variable (sex, stress condition, and their two-by-two interaction terms) in the model is indicated by a "+" symbol. The regression parameter is only given for the corresponding continuous variable (day) when this variable is present in the model. The most parsimonious model is highlighted in bold font.

Intercept	day	sex	stress	day:sex	day:stress	sex:stress	adj.R²	df	logLik	AICc	ΔAICc	weight
0,02	0,00	+	+	+	+	+	-0,01	8	1444,54	2872,80	0,00	0,63
0,02	**0,00**	**+**	**+**		**+**	**+**	**-0,01**	**7**	**1442,55**	**2870,90**	**1,91**	**0,24**
0,02	0,00	+	+	+	+		-0,01	7	1441,20	2868,18	4,62	0,06
0,02	0,00		+		+		-0,01	5	1438,73	2867,35	5,45	0,04
0,02	0,00	+	+		+		-0,01	6	1439,08	2866,01	6,79	0,02
0,02	0,00	+	+	+		+	-0,01	7	1436,82	2859,43	13,37	0,00
0,03	0,00	+	+			+	-0,01	6	1435,27	2858,39	14,41	0,00
0,02	0,00	+	+	+			-0,01	6	1433,45	2854,75	18,05	0,00
0,03	0,00		+				-0,01	4	1431,36	2854,64	18,16	0,00
0,02	0,00	+	+				-0,01	5	1431,79	2853,47	19,33	0,00
0,02	0,00	+		+			-0,01	5	1431,45	2852,78	20,02	0,00
0,02	0,00						-0,01	3	1429,38	2852,71	20,09	0,00
0,02	0,00	+					-0,01	4	1429,89	2851,71	21,09	0,00
0,05			+				0,00	3	1179,76	2353,47	519,33	0,00
0,05							0,00	2	1178,27	2352,52	520,28	0,00
0,05		+	+				0,00	4	1179,85	2351,62	521,18	0,00
0,06		+	+			+	0,00	5	1180,51	2350,91	521,90	0,00
0,05		+					0,00	3	1178,39	2350,74	522,06	0,00

Table S2d. Effect of the water stress condition, sex and day on body mass. For each model, we reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood (LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc), change in AICc (△AICc) from the best model, and model weight. The presence of the categorial variable (sex, stress condition, and their two-by-two interaction terms) in the model is indicated by a "+" symbol. The regression parameter is only given for the corresponding continuous variable (day) when this variable is present in the model. The most parsimonious model is highlighted in bold font.

Intercept	day	sex	stress	day:sex	day:stress	sex:stress	adj.R²	df	logLik	AICc	△AICc	weight
0,04	**0,00**						**0,00**	**3**	**1309,82**	**-2613,60**	**0,00**	**0,29**
0,04	0,00		+		+		0,00	5	1311,34	-2612,57	1,03	0,17
0,04	0,00	+					0,00	4	1310,00	-2611,93	1,68	0,12
0,04	0,00		+				0,00	4	1309,98	-2611,89	1,72	0,12
0,04	0,00	+	+		+		0,00	6	1311,50	-2610,85	2,76	0,07
0,04	0,00	+	+				0,00	5	1310,18	-2610,24	3,37	0,05
0,04	0,00	+		+			0,00	5	1310,01	-2609,90	3,70	0,04
0,04	0,00	+	+		+	+	0,00	7	1311,79	-2609,37	4,24	0,03
0,04	0,00	+	+	+	+		0,00	7	1311,50	-2608,79	4,81	0,03
0,04	0,00	+	+			+	0,00	6	1310,48	-2608,79	4,82	0,03
0,04	0,00	+	+	+			0,00	6	1310,18	-2608,21	5,40	0,02
0,04	0,00	+	+	+	+	+	0,00	8	1311,79	-2607,30	6,30	0,01
0,04	0,00	+	+	+		+	0,00	7	1310,49	-2606,75	6,85	0,01
0,05							0,00	2	1272,47	-2540,92	72,68	0,00
0,05		+					0,00	3	1272,59	-2539,13	74,47	0,00
0,05			+				0,00	3	1272,57	-2539,10	74,50	0,00
0,05		+	+				0,00	4	1272,70	-2537,32	76,28	0,00
0,05		+	+			+	0,00	5	1272,79	-2535,47	78,13	0,00

Reproductive success

Table S2e. Effect of the thermal stress condition on the reproductive success. For each model, we
reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood
(LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc),
change in AICc (ΔAICc) from the best model, and model weight. The presence of the categorial
variable (stress condition) in the model is indicated by a "+" symbol. The value of regression
parameter is only given for the intercept. The most parsimonious model is highlighted in bold font.

Intercept	stress	adj.R²	df	logLik	AICc	ΔAICc	weight
0,62	**+**	**0,16**	**2**	**-25,17**	**54,66**	**0,00**	**0,80**
-0,10		0,00	1	-27,68	57,46	2,80	0,20

Table S2f. Effect of the water stress condition on the reproductive success. For each model, we
reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood
(LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc),
change in AICc (ΔAICc) from the best model, and model weight. The presence of the categorial
variable (stress condition) in the model is indicated by a "+" symbol. The value of regression
parameter is only given for the intercept. The most parsimonious model is highlighted in bold font.

Intercept	stress	adj.R²	df	logLik	AICc	ΔAICc	weight
-1,39	**+**	**0,17**	**2**	**-23,77**	**51,87**	**0,00**	**0,83**
-0,51		0,00	1	-26,46	55,03	3,16	0,17

Individual physiological traits

Immune cell viability

Table S2g. Effect of the thermal stress condition and sex on immune cell viability. For each model, we reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood (LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc), change in AICc (△AICc) from the best model, and model weight. The presence of the categorial variable (sex, stress condition, and their interaction term sex:stress) in the model is indicated by a "+" symbol. The value of regression parameter is only given for the intercept. The most parsimonious model is highlighted in bold font.

Intercept	sex	stress	sex:stress	adj.R²	df	logLik	AICc	△AICc	weight
60,31				**0,00**	**2**	**-231,04**	**466,30**	**0,00**	**0,47**
61,97		+		0,02	3	-230,57	467,58	1,28	0,25
61,00	+			0,00	3	-230,97	468,39	2,08	0,17
62,43	+	+		0,02	4	-230,53	469,81	3,51	0,08
63,67	+	+	+	0,03	5	-230,24	471,63	5,32	0,03

Table S2h. Effect of the water stress condition and sex on immune cell viability. For each model, we reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood (LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc), change in AICc (△AICc) from the best model, and model weight. The presence of the categorial variable (sex, stress condition, and their interaction term sex:stress) in the model is indicated by a "+" symbol. The value of regression parameter is only given for the intercept. The most parsimonious model is highlighted in bold font.

Intercept	sex	stress	sex:stress	adj.R²	df	logLik	AICc	△AICc	weight
51,82	+	+		0,15	4	-212,23	433,32	0,00	0,48
47,45		**+**		**0,08**	**3**	**-214,36**	**435,21**	**1,90**	**0,19**
52,29	+	+	+	0,15	5	-212,20	435,70	2,39	0,15
55,52	+			0,06	3	-214,86	436,23	2,91	0,11
51,29				0,00	2	-216,44	437,13	3,81	0,07

Immune cell size

Table S2i. Effect of the thermal stress condition and sex on immune cell size. For each model, we
reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood
(LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc),
change in AICc (ΔAICc) from the best model, and model weight. The presence of the categorial
variable (sex, stress condition, and their interaction term sex:stress) in the model is indicated by a "+"
symbol. The value of regression parameter is only given for the intercept. The most parsimonious
model is highlighted in bold font.

Intercept	sex	stress	sex:stress	adj.R²	df	logLik	AICc	ΔAICc	weight
7,91	+	+	+	0,25	5	-29,17	69,50	0,00	0,32
7,99	+	+		0,20	4	-30,45	69,66	0,17	0,30
7,91		**+**		**0,15**	**3**	**-31,64**	**69,73**	**0,23**	**0,29**
7,87	+			0,07	3	-33,33	73,09	3,60	0,05
7,77				0,00	2	-34,79	73,80	4,30	0,04

Table S2j. Effect of the water stress condition and sex on immune cell size. For each model, we
reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood
(LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc),
change in AICc (ΔAICc) from the best model, and model weight. The presence of the categorial
variable (sex, stress condition, and their interaction term sex:stress) in the model is indicated by a "+"
symbol. The value of regression parameter is only given for the intercept. The most parsimonious
model is highlighted in bold font.

Intercept	sex	stress	sex:stress	adj.R²	df	logLik	AICc	ΔAICc	weight
8,24		+		0,08	3	-44,46	95,42	0,00	0,45
8,10				**0,00**	**2**	**-46,36**	**96,97**	**1,54**	**0,21**
8,17	+	+		0,10	4	-44,10	97,06	1,63	0,20
8,04	+			0,01	3	-46,16	98,81	3,39	0,08
8,19	+	+	+	0,10	5	-44,06	99,42	4,00	0,06

Immune cell density

Table S2k. Effect of the thermal stress condition and sex on immune cell density. For each model, we
reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood
(LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc),
change in AICc (△AICc) from the best model, and model weight. The presence of the categorial
variable (sex, stress condition, and their interaction term sex:stress) in the model is indicated by a "+"
symbol. The value of regression parameter is only given for the intercept. The most parsimonious
model is highlighted in bold font.

Intercept	sex	stress	sex:stress	adj.R²	df	logLik	AICc	△AICc	weight
3538275,86		**+**		**0,41**	**3**	**-898,33**	**1803,10**	**0,00**	**0,66**
3644762,62	+	+		0,41	4	-898,12	1804,99	1,89	0,26
3600666,67	+	+	+	0,41	5	-898,08	1807,31	4,22	0,08
2472758,62				0,00	2	-913,43	1831,07	27,98	0,00
2707777,78	+			0,02	3	-912,92	1832,29	29,20	0,00

Table S2l. Effect of the water stress condition and sex on immune cell density. For each model, we
reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood
(LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc),
change in AICc (△AICc) from the best model, and model weight. The presence of the categorial
variable (sex, stress condition, and their interaction term sex:stress) in the model is indicated by a "+"
symbol. The value of regression parameter is only given for the intercept. The most parsimonious
model is highlighted in bold font.

Intercept	sex	stress	sex:stress	adj.R²	df	logLik	AICc	△AICc	weight
1465517,24		**+**		**0,13**	**3**	**-802,79**	**1612,09**	**0,00**	**0,52**
1827142,86	+	+	+	0,18	5	-801,23	1613,77	1,68	0,23
1575106,08	+	+		0,14	4	-802,60	1614,06	1,97	0,20
1892500,00				0,00	2	-806,49	1617,23	5,14	0,04
1960434,78	+			0,00	3	-806,44	1619,38	7,29	0,01

β-Galactosidase activity

Table S2m. Effect of the thermal stress condition and sex on β-Galactosidase activity. For each
model, we reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log
likelihood (LogLik) values, Akaike information criteria values with a correction for small sample sizes
(AICc), change in AICc (△AICc) from the best model, and model weight. The presence of the
categorial variable (sex, stress condition, and their interaction term sex:stress) in the model is
indicated by a "+" symbol. The value of regression parameter is only given for the intercept. The
most parsimonious model is highlighted in bold font.

Intercept	sex	stress	sex:stress	adj.R²	df	logLik	AICc	△AICc	weight
12399,83		**+**		**0,17**	**3**	**-556,53**	**1119,51**	**0,00**	**0,60**
12988,70	+	+		0,18	4	-556,14	1121,06	1,55	0,28
12487,80	+	+	+	0,19	5	-555,81	1122,82	3,30	0,11
14534,95				0,00	2	-561,86	1127,94	8,42	0,01
15052,70	+			0,01	3	-561,62	1129,71	10,20	0,00

Table S2n. Effect of the water stress condition and sex on β-Galactosidase activity. For each model,
we reported intercept of the regression, adjusted R² (adj.R²), degree of freedom (df), Log likelihood
(LogLik) values, Akaike information criteria values with a correction for small sample sizes (AICc),
change in AICc (△AICc) from the best model, and model weight. The presence of the categorial
variable (sex, stress condition, and their interaction term sex:stress) in the model is indicated by a "+"
symbol. The value of regression parameter is only given for the intercept. The most parsimonious
model is highlighted in bold font.
730

Intercept	sex	stress	sex:stress	adj.R²	df	logLik	AICc	△AICc	weight
10694,60		**+**		**0,17**	**3**	**-477,96**	**962,43**	**0,00**	**0,67**
10867,90	+	+		0,18	4	-477,84	964,53	2,10	0,23
10955,89	+	+	+	0,18	5	-477,79	966,89	4,47	0,07
9725,79				0,00	2	-482,92	970,08	7,65	0,01
10087,84	+			0,01	3	-482,55	971,60	9,17	0,01

733   **Supplementary file 3: Graphical representations of results per sex**

735   1. **Life history traits**

736   1.A. Survival

738   *Figure 1.A.1: Effect of thermal stress on survival*
739   *CT females: control females in Control Temperature (20°C), HT females: stressed females in High Temperature (28°C), CT*
740   *males: control males in Control Temperature (20°C), HT males: stressed males in High Temperature (28°C)*

742   *Figure 1.A.2.: Effect of water stress on survival*
743   *CM females: control females in Control Moisture (moisture 80%), LM females: stressed females in Loss of Moisture (moisture*
744   *50%), CM males: control males in Control Moisture (moisture 80%), LM males: stressed males in Loss of Moisture (moisture*
745   *50%)*

1.B. Body mass across time

Figure 1.B.1.: Boxplot of the effect of thermal stress on body mass (measured in grams) over time
CT females: control females in Control Temperature (20°C), HT females: stressed females in High Temperature (28°C), CT
males: control males in Control Temperature (20°C), HT males: stressed males in High Temperature (28°C)

Figure 1.B.2.: Boxplot of the effect of water stress on body mass (measured in grams) over time
CM females: control females in Control Moisture (moisture 80%), LM females: stressed females in Loss of Moisture (moisture
50%), CM males: control males in Control Moisture (moisture 80%), LM males: stressed males in Loss of Moisture (moisture
50%)

            1.C. Reproduction success

763   *Figure 1.C.1.: Effect of temperature on breeding success* *(0 = pairs that did not produce offspring; 1 = pairs that produced*
764   *offspring; CT: control individuals in Control Temperature (20°C), HT: Stressed individuals in High Temperature (28°C))*
765
766

768   *Figure 1.C.2.: Effect of moisture on breeding success* *(0 = pairs that did not produce offspring; 1 = pairs that produced*
769   *offspring; CM: control individuals in Control Moisture (moisture 80%), LM females: stressed individuals in Loss of Moisture*
770   *(moisture 50%))*
771

    2. Individual physiological traits

    2.A. Immune cells viability

    Figure 2.A.1.: Effect of thermal stress on immune cell viability (% of live cells)
    CT females: control females in Control Temperature (20°C), HT females: stressed females in High Temperature
    (28°C), CT males: control males in Control Temperature (20°C), HT males: stressed males in High Temperature
    (28°C)

    Figure 2.A.2.: Effect of water stress on immune cell viability (% of live cells)
    CM females: control females in Control Moisture (moisture 80%), LM females: stressed females in Loss of Moisture
    (moisture 50%), CM males: control males in Control Moisture (moisture 80%), LM males: stressed males in Loss of
    Moisture (moisture 50%)

800   **2.B.** Immune cells size

802   *Figure 2.B.1.: Effect of thermal stress on immune cells size (in μm)*
803   *CT females: control females in Control Temperature (20°C), HT females: stressed females in High Temperature (28°C), CT*
804   *males: control males in Control Temperature (20°C), HT males: stressed males in High Temperature (28°C)*

807   *Figure 2.B.2.: Effect of water stress on immune cells size (in μm)*
808   *CM females: control females in Control Moisture (moisture 80%), LM females: stressed females in Loss of Moisture (moisture*
809   *50%), CM males: control males in Control Moisture (moisture 80%), LM males: stressed males in Loss of Moisture (moisture*
810   *50%)*

    2.C. Immune cells density

Figure 2.C.1.: Effect of thermal stress on immune cells density (number of cells per mL of haemolymph)
CT females: control females in Control Temperature (20°C), HT females: stressed females in High Temperature (28°C), CT
males: control males in Control Temperature (20°C), HT males: stressed males in High Temperature (28°C)

Figure 2.C.2.: Effect of water stress on immune cells density (number of cells per mL of haemolymph)
CM females: control females in Control Moisture (moisture 80%), LM females: stressed females in Loss of Moisture (moisture
50%), CM males: control males in Control Moisture (moisture 80%), LM males: stressed males in Loss of Moisture (moisture
50%)

828   **2.D**. β-galactosidase activity

Figure 2.D.1.: Effect of thermal stress on β-galactosidase activity

CT females: control females in Control Temperature (20°C), HT females: stressed females in High Temperature (28°C), CT males: control males in Control Temperature (20°C), HT males: stressed males in High Temperature (28°C)

Figure 2.D.2.: Effect of water stress on β-galactosidase activity

CM females: control females in Control Moisture (moisture 80%), LM females: stressed females in Loss of Moisture (moisture 50%), CM males: control males in Control Moisture (moisture 80%), LM males: stressed males in Loss of Moisture (moisture 50%)

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