Moving beyond MARCO

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Abstract

The use of imaging systems in protein crystallisation means that the experimental setups no longer require manual inspection to determine the outcome of the trials. However, it leads to the problem of how best to find images which contain useful information about the crystallisation experiments. The adoption of a deeplearning approach in 2018 enabled a four-class machine classification system of the images to exceed human accuracy for the first time. Underpinning this was the creation of a labelled training set which came from a consortium of several different laboratories. The MARCO classification model does not have the same accuracy on local data as it does on images from the original test set; this can be somewhat mitigated by retraining the ML model and including local images. We have characterized the image data used in the original MARCO model, and performed extensive experiments to identify training settings most likely to enhance the local performance of a MARCO-dataset based ML classification model.

Introduction

Structural biology, where knowledge of the atomic detail of biological macromolecules informs our understanding of their function, has profoundly changed our understanding of the living world [1]. The most successful technique used to generate the 3D atomic models is X-ray crystallography, where the molecule of interest is coaxed into a well-ordered lattice (a crystal) which is used as a diffraction grating for a monochromatic beam of hard X-rays [2]. The resulting diffraction pattern can be used to generate a model of the molecule of interest. The value of this technique can be seen not only in the number of Nobel Prizes associated with the technique [3], but also in the contribution that X-ray crystallography makes to human health [4,5].

Every technique has a bottleneck; in macromolecular crystallography the limiting factor is the production of well-ordered crystals which are suitable for diffraction experiments [6,7]. Diffraction quality crystals need to be sufficiently large (10-100 µm per side), single and well-ordered. The process of generating crystals has remained essentially unchanged for many decades - most often a small droplet of a pure and concentrated sample of the macromolecule is mixed with a similar volume of a chemical cocktail, and the system is left to equilibrate against a larger volume of the cocktail [8,9]. The outcome of the experiment is monitored either by observing the droplet directly under a microscope or by looking at an image of the droplet. Crystallisation requires firstly that that the solution being crystallized is or becomes supersaturated, secondly that nucleation (the creation of a transient assembly of molecules into a formation suitable for other molecules to join) occurs and thirdly that the nuclei continue to grow, rather than to dissolve [9]. All three of these processes can be slow and stochastic, thus each droplet/trial has to be monitored repeatedly over time (days to months) for crystal formation or other, less positive outcomes.

Although the fundamental approach to crystallisation has been unchanged for 50 years, the details of how the trials are set up have evolved, largely in response to the Structural Genomics initiatives of the early 2000s [10]. These high-throughput approaches have been adopted by many modern laboratories; particularly larger industrial and service laboratories. Firstly, the volumes of sample and cocktail have shrunk from many microlitres (µL) per trial to 50-1000 nanolitres (nL) per trial, as the

crystal size required for diffraction experiments has decreased. This has allowed for trials to be set up in parallel using laboratory automation - often 96-1536 droplets on a single SBS footprint plate (Standards ANSI/SLAS 1-2004 through ANSI/SLAS 4-2004) [11]. In turn, the high density has driven the development of automated incubation/imaging systems, where crystallisation plates with many trials are stored and periodically imaged - there are generally between 4 and 20 plate inspections (*i.e.* the process by which discrete set of images, one for each trial on a plate, is collected at a single timepoint) over the course of weeks to months. This removes the necessity for multiple manual inspections of the droplet-containing plate, but leads to the generation of large numbers of images that need to be scanned for interesting results [12]. Many of the images in a droplet timecourse will be essentially identical, as much of the droplet variation happens early in the lifetime of the crystallisation experiment [13]. There will still be slight differences between the timecourse images even if the droplet remains unchanged; these are due to subtle variations in lighting or positioning of the droplet under the imaging lens. Early attempts to find interesting results by looking for changes between images in a timecourse were too slow to be useful [14].

The problem of interpreting images was anticipated even before the first commercial imaging systems were available and there were a number of academic groups working on Structural Genomics projects who developed image interpretation applications based on Random Forest or other machine learning (ML) algorithms [15–18]. However, none of these were accurate enough to be adopted outside their home laboratory. More recently, an approach based on the Inception V3 deep learning architecture [19] showed that machine interpretation of crystallisation images can exceed the accuracy of human interpretation of the images [20]. This work (Machine Recognition of Crystallization Outcomes or "MARCO") required the compilation of a large training set of classified images, which were provided by a consortium of academic, government and industrial structural biology laboratories. As there are no standard classification schemes for describing a crystallisation trial [21], or an image of the trial droplet, each of the individual structural biology laboratories that contributed images used their own local classification system, which were collapsed down to four classes - 'Crystal', 'Precipitate', 'Clear' and 'Other'. The four classes were chosen as they cover much of the gross information in a crystallisation droplet. Although the most interesting result for a

crystallographer is 'Crystal', it is also the most rare; a paper from a decade ago ⁶⁴ estimated that harvestable crystals (*i.e.* those gauged by visual inspection to be ⁶⁵ potentially useful for diffraction experiments) are a small subset of all the crystals seen ⁶⁶ and make up 1-2% of the trial outcomes [22]. The classifications 'Clear' and 'Precipitate' ⁶⁷ help to identify trials which did not reach supersaturation and/or did not support ⁶⁸ nucleation ('Clear') or became over-supersaturated or disrupted the protein sample ⁶⁹ integrity ('Precipitate'). These non-crystal results can help locate an appropriate region ⁷⁰ of chemical space for further trials. The category 'Other' applies to all results that do ⁷¹ not fall into the other three categories, and includes badly set up and mis-imaged ⁷² experiments as well as trial outcomes like liquid-liquid phase separation and spherulites. ⁷³

The outcomes from crystallisation trials are complex, and the classification of images ⁷⁴ of the trials into four classes is a gross simplification of the outcome space. The ⁷⁵ boundaries between any classification are blurred [23] - for example, very small crystals ⁷⁶ (microcrystals) can be indistinguishable from precipitate, and the human interpretation ⁷⁷ of outcomes can be variable, see Fig 1. Many drops display more than one outcome - for ⁷⁸ example, a drop might contain both crystals and precipitate. Drops may also display ⁷⁹ outcomes not captured by any of the classes - e.g. wrinkled protein skin, container ⁸⁰ imperfections or fungal growth, Fig 2. Futher, humans also focus on the domain ⁸¹ relevant result: an image that contains a droplet with a dust fibre in it may well be ⁸² classified as 'Clear' as the dust is considered irrelevant to the crystallisation experiment. ⁸³

Fig 1. Blurred boundaries between MARCO classes. The three images here look similar, but have been classifed into three different MARCO classes by a crystallographer A: Other (phase separation). B: Precipitate. C. Crystal.

Any ML application is dependent on the quality of the data used to train the system ⁸⁴ - the MARCO model used half a million images from 5 different groups, and gives ⁸⁵ impressive results [24]. However, during the process of training the MARCO model it ⁸⁶ was noticed that the model's performance on images from our lab (The Collaborative ⁸⁷

Fig 2. Outcomes not captured in the MARCO closed set. Many crystallisation images display outcomes that are not easily captured in four classes A: Dust fibre in an otherwise clear drop. B: Precipitated protein, along with an artifact from a wrinkled skin over the droplet. C. Collapsed air bubbles, (introduced inadvertently during drop dispensing) against a background of precipitated protein.

Crystallisation Centre (C3) at CSIRO in Parkville, Australia) improved dramatically when a small number of images from C3 were included in the training set. This is a truism in ML; including local data into a general model invariably improves local performance [25]. Less obvious is how much local data is needed [26], and what quality [diversity, accuracy] of the local data is needed [27]. Another question is how best to utilise local data: *via* incorporation into an existing model (transfer learning [28]), or by building a new model using both the general data (here the MARCO training set) and local data [29]. Modern ML tools also allow for the re-examination of the approach of using local data only [**JN:** *cite julie here*] We were interested in understanding the dataset used to create the original ML model. Can we develop tools to measure the quality of these data? For example, we would like to be able to measure how much of the MARCO data is unique, and how representative of all crystallisation data the MARCO training data are.

In this current work we have contributed by:

- Characterising and identifing problems with the MARCO data set.

- Defining the parameters that allow the creation of models using currently available architectures that exceed the success of the original Inception V3 model.

- Developing guidelines for which local images to include to most effectively train a ML model using both MARCO and local images with the goal of producing an image classification model tuned to that local environment.

- Making code available to enable the facile development of a novel classification

model tuned to the local environment which which uses both the MARCO 109

training set and a limited amount of local data. [1] 110

- Creating a freely available protein crystallization dataset that contains both 111

visible and ultra-violet light images [2]. 112

1 Approach 113

1.1 Creation of a C3 validation set 114

The fundamental goal of this work was to investigate how the MARCO model and 115

dataset can be effectively adapted to a local environment. In order to evaluate methods 116

we developed a validation dataset. This dataset (C3 Test) uses the same meta-labels 117

that were chosen for the original MARCO dataset: Clear, Crystals, Precipitate and 118

Other. All of the images in C3 test are of vapour diffusion experiments, set up in 119

Innovadyne SD-2 plastic plates and imaged on a Formulatrix RockImager; thus similar 120

to many of the images in the MARCO dataset - by eye, the vapour diffusion images in 121

the C3 Test set and the vapour diffusion images from the MARCO training set are 122

indistinguishable. To ensure that there were no obviously redundant images in the C3 123

Test set only one image from any image timecourse was included. 124

1.2 Replicating the MARCO results 125

The model produced in the original MARCO paper used an InceptionV3 126

architecture [19]. Although this architecture achieved state of the art results upon its 127

release in 2015 [3], the Machine Learning community has since transitioned to using 128

ResNet architectures as the default [30]. In order to align our work with the community 129

we trained models using architectures from the ResNet family with the aim of 130

replicating the performance of the original MARCO model. 131

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[1] https://bitbucket.org/nelliottrosa/marco-local
[2] C3 Test dataset is available at https://zenodo.org/record/4635300
[3] https://paperswithcode.com/sota/image-classification-on-imagenet.

1.3 Image types in the MARCO dataset

During the visual inspection of the MARCO dataset we observed at least three different experimental setups [9]: under oil, vapour diffusion and a minor subset of Lipidic Cubic Phase (LCP) experiments, a type of liquid diffusion trial in which the concentrated protein sample is pre-mixed with a lipid such as monoolein [31], see Fig 3. To a crystallographer, images of LCP trials appear quite different from images of the other two types of crystallization experiment. Further, LCP experiments also show a greater range of outcomes than aqueous droplet trials as changes in the lipid phase contribute to image complexity along with the more commonly observed protein outcomes. There is no information about the number of LCP images in the MARCO dataset; it is not clear that the authors of the original MARCO work appreciated that any LCP images had been included in the dataset.

Fig 3. Different experimental setups found in the MARCO training set.
Images of three different crystallisation trial types are included in the MARCO dataset image A: Vapor Diffusion (the most abundant), images of these experiments were contributed by GSK, MERCK, C3 and BMS. B: Microbatch under oil. Most of the images from this type of experiment were from the HWI, with a small number from GSK. C. LCP bolus - there are few of these in the dataset, and were provided by MERCK.

We estimated the number of LCP images by training a mobileNetV2 [32] architecture as a simple LCP classifier. Further, we examined if the number of LCP images in the MARCO dataset would be enough to produce robust classification of these type of

images, using a small number of weakly labelled LCP images from the C3 laboratory. ₁₄₇

1.4 Redundancy of Images in the MARCO training set ₁₄₈

By observing the change in classification performance on the MARCO test data as ₁₄₉

increasing numbers of images are removed from the training data we can estimate the ₁₅₀

redundancy of the dataset. We used the approach of Birodkar [33] to estimate the ₁₅₁

redundancy of the dataset by removing semantically duplicate images in a stepwise ₁₅₂

manner. Semantic duplicates were determined by performing agglomerative

Fig 4. Examples of redundant images contained within the MARCO dataset. Each of these 16 images is found in the MARCO dataset, and are clustered into rows based on similarity as determined by the semantic redundancy process described in this Section. The similar images shown here are different timepoints of the same crystallisation experiment. Each image is so similar they cannot be distinguished apart by a human. There are other semantically redundant images that don't appear the same, but contribute the same information to the model (not shown).

₁₅₃

clustering [34], in which images with the lowest dissimilarity are grouped together into ₁₅₄

clusters. We use the euclidean distance between images in the representation space of a ₁₅₅

ResNet50 model [30] that was trained upon the entire MARCO dataset as our measure ₁₅₆

of dissimilarity. The image that was closest to the centre of the cluster was retained in ₁₅₇

the dataset whilst all others were removed. The number of clusters formed was chosen 158
to be the number of images that were to be retained in the reduced dataset; thus to 159
remove 10% of images in a hypothetical dataset containing 100 images, 90 clusters 160
would be chosen. We made one modification to the published method whereby we 161
performed the agglomerative clustering within pseudo-classes that were created by 162
K-means clustering [35] within each true class within the representation space. Each 163
pseudo-class contained approximately 10K images. This change was made to allow the 164
analysis to run within a reasonable amount of time. 165

1.5 Image and dataset characterisation 166

To characterize and understand the MARCO training set, we used dimension reduction 167
to probe for any inherent structure within the MARCO training data. 168

A SimCLR [36] model was trained to extract features from input images that were 169
useful for differentiating between semantically different images without imposing bias 170
from our preconceived notion of outcome class. SimCLR is an algorithm for training a 171
feature extractor in an self-supervised (*i.e.* label free) manner. 172

Uniform Manifold Approximation and Projection (UMAP) [37] was used to reduce 173
the high dimensional representations produced by SimCLR to two dimensions in order 174
to visualise the MARCO dataset. UMAP is a fast and scalable method for dimension 175
reduction that preserves global topological structure and is founded with provable 176
theoretical manifold learning. The reduced dimensional spaces learned by UMAP 177
generally provide a better view of the global structure of a dataset than alternative 178
approaches that seek to preserve local topological structure. We also visualised the 179
(labelled) MARCO test data using UMAP. Using an image representation from an 180
intermediate layer in the MARCO model, we used UMAP to identify a lower 181
dimensional manifold that preserves the local structure of the data. 182

To understand the distribution of sample positions relative to class boundaries we 183
used the methods described by Liu *et al* [38] to find the six binary class boundaries 184
between each class pair; then we plotted the distribution of distances between each 185
boundary and each sample in the MARCO and C3 test sets. 186

1.6 Adding local images to the MARCO training set

We explored different training setups where the original MARCO training dataset was
supplemented with local C3 images and observed what parameters created the model
that generalised best to the C3 setup. The three variables we tested were: image class
distribution, image selection method and the number of local images included. To
obtain images that could be added to the MARCO dataset we searched C3's collection
for images that had been scored by an expert. We we used the same criteria to select
images that was used in section Creation of a C3 validation set, however care was taken
to ensure that there was no overlap between the validation dataset (the C3 test set) and
this image pool. The score from the single crystallographer is taken as ground truth.

We also explored using both entropy and cross entropy as 'hardness' measures to
select local images for inclusion into the MARCO training data. 'Hard' local images
were found by using the entropy of model prediction as a proxy for hardness and
selecting images from the pool which had the largest entropy values for any given model.
We compared models where hard local images were included to models where the same
number and distribution of randomly selected images were included.

2 Experiments

2.1 Creation of a C3 validation set

To find images, C3's collection of experiments from January 2017 onwards was scraped
to find images that had already been labelled by at least one crystallographer (who was
not in the relabelling team, see below), this gave us a starting set of 9215 labelled
images. Since labelling crystallisation outcomes is known to be a hard problem, even for
experts (see for example [39]), we increased the robustness of our labels with a two-step
re-labelling procedure. In the first round of re-labelling all images were labelled by a
second crystallographer. A label was only considered robust if it was chosen by both the
two crystallographers (a majority score). In the second re-labelling round all images
that were yet to have a consistent label were scored again by third crystallographer. In
neither the two rounds of relabelling were the existing labels revealed, so every score
was assigned by simply observing the image. After the second round of additional

scoring, 347 of the original 9215 images remained ambiguous with each crystallographer choosing a different score; these images were not included in the final dataset. The breakdown of the scoring for each of the two validation rounds can be seen in Supplementary Information Tables **??** and **??**. The class breakdown of the final C3 Test dataset is shown in Table 1.

Both protein crystals and crystals of inorganic salts can grow in protein crystallisation trials. Images taken with ultra-violet (UV) light may be useful to distinguish between salt and protein crystals, as [most] protein crystals glow under UV illumination due to the intrinsic tryptophan fluorescence of protein crystals [40]. UV imaging is quite widely applied in crystallisation laboratories; and the orthogonal information these images provide could be helpful for classifying crystallisation experiment outcomes. Generally, a UV image of a crystallisation experiment is taken at the same time as a visible light image is collected, and we have included the associated UV image in the C3 Test dataset. To the best of our knowledge this is the first publicly available crystallisation outcome dataset to include labelled UV light images.

Table 1. Label distribution of different data sources

Data source	Clear	Precipitate	Crystal	Other
MARCO	33.6%	48%	12.8%	5.6%
C3 Test	23.5%	40.0%	28.3%	8.1%
Cinder	27%	50%	11%	12%

Class distributions found in the MARCO data and the C3 test data. These can be compared to the distributions found from the Cinder application [21], which may be an unbiased indication of the native distribution of outcomes.

2.2   Replicating the MARCO results

In order to replicate the performance of the MARCO model, we did a large exploration of training hyper-parameters using two sizes of the Resnet architecture, Resnet18 and Resnet50, along with multiple image resolutions. More information about the training procedure used can be found in the Appendix. We compared the accuracy of trained models upon both the MARCO test set and the C3 Test set. The best outcomes of our experiments are shown in Table 2. Due to the variance of individual training runs the reported accuracy for each model is the average of three trials which were run using

three different random seeds. More information about the training procedure used for these models can be found in the Appendix. We found high-resolution Resnet18 models were less prone to over-fitting on the MARCO data distribution so could be trained for longer, therefore we trained these models for 50 epochs rather than the 30 epochs used for the Resnet50 models.

Table 2. Performance of different ResNet models on the MARCO and C3 test datasets

Model	Image size	MARCO Acc.	C3 Acc.
MARCO	599	94.5*	82.03
Resnet18	256	93.21	79.51
Resnet18$^\diamond$	512	94.18	82.12
Resnet18$^\diamond$	600	94.21	**82.14**
Resnet50	256	94.24	76.46
Resnet50	512	**94.63**	81.5

The evaluation performance of several models trained upon the MARCO dataset. Best results are in **bold**. Results marked * were copied directly from [20]. Models marked ⋄ were trained for 50 epochs. The reported accuracy for each model is the average of three trials using different random seeds. More information about the training procedure used for these models can be found in the Appendix.

2.3   Image types in MARCO

There are images of three different types of crystallisation experiments in the MARCO data. Most common are images of vapor diffusion experiments; all the contributing groups - with the exception of HWI - provided images of this type of experiment. HWI provided images of crystallisation experiments which had been set up under a column of oil (microbatch experiments). Finally, some groups (neither CSIRO nor HWI) also included some images showing Lipidic Cubic Phase (LCP) experiments. Although the outcomes of the three different experiment types are broadly similar, the images themselves look significantly different, and LCP experiments can show features not seen in the other experiments. We created a binary classifier using PyTorch? [**JN:** *@Chris - Check!!!, The introduction says that we used mobileNetV2*], training the model with 1000 LCP images from CSIRO C3 and the same number and distribution of non-LCP images from the MARCO image dataset - the 1000 MARCO images were checked by hand to ensure that these images were of vapour diffusion or microbatch experiments. This model was used to scan the MARCO training data, and identified 26K images

which were classified as putative images of LCP experiments; human inspection showed ₂₅₉ about 1 in 10 of the 26K images thus identified were actually LCP images; we estimate ₂₆₀ about there are on the order of 2.5-3K LCP images in the entire dataset. A test of how ₂₆₁ well the MARCO model works with LCP images was done by selecting 960 labelled ₂₆₂ LCP images from C3, and looking at the confusion matrix generated from the MARCO ₂₆₃ scoring of this small set of images, see Figure 5. ₂₆₄

Fig 5. Confusion matrix for 96 LCP images from C3, classified using the MARCO model. The confusion matrix is normalized over the true conditions (e.g. rows), this shows that the MARCO model classifies most LCP images as crystals, overall accuracy 56%.

2.4   Redundancy of Images in the MARCO training set ₂₆₅

The accuracy on both the C3 Test set and the MARCO validation set was calculated ₂₆₆ using a Resnet50 model with images of 256×256 pixels for 5 subsets of the MARCO ₂₆₇ training set, with increasingly stringent requirements for distinctness of the images (i.e. ₂₆₈ with decreasing redundancy, as per the method of Brodikar, described above). Models ₂₆₉ were trained in triplicate, with the reported accuracies being the average across runs, ₂₇₀ and showed that even with 50% of the original data, less than 0.3% decrease in accuracy ₂₇₁ on the MARCO test data was observed, Figure 6. Interestingly, the accuracy of the ₂₇₂ classification of the both MARCO and C3 test data peaked whilst only using 80% of the ₂₇₃ MARCO training data, suggesting that the very redundant nature of the original ₂₇₄ MARCO data leads to a degree of overfitting on the redundant data, at the expense of ₂₇₅ accuracy. The reduced datasets used in this analysis can be found in the code repository. ₂₇₆

Fig 6. Performance of a set of Resnet50 models using 256×256 **pixel images on the C3 Test dataset and MARCO validation dataset when trained with increasingly strict redundancy cuts.** The blue line shows the accuracy of the reduced data models against the C3 Test data, and the orange line shows the accurancy of the reduced data models against the MARCO validation set. 10% of the data were removed stepwise from 100% to 40% of the original training data. The pale regions cover the area defined by the standard deviation of three runs. Insert shows detail of the MARCO validation data trials.

2.5   Image and dataset characterisation

Fig 7. UMAP projection of labelled data from the MARCO validation dataset. The left hand figure shows the projection coloured by classification, where clear, crystal, other and precipitate are coloured green, pink, blue and orange respectively. The right hand figure shows the same projection, but coloured by source of the images - Merck, GSK, BMS, HWI and C3 are coloured blue, green, purple, orange and red respectively. Only every nth image has been visualised for clarity.

UMAP was run using the penultimate layer of a ResNet50 trained from a random

Fig 8. UMAP projection of data from the MARCO validation dataset labelled using a simCLR unsupervised approach. The left hand figure shows the projection coloured by classification, where Precipitate, Crystal, Clear and Other are coloured green, orange, blue and pink respectively. The right hand figure shows the same projection, but coloured by source of the images - Merck, GSK, BMS, HWI and C3 are coloured blue, green, purple, orange and red respectively. Only every nth image has been visualised for clarity.

initialisation using the MARCO dataset to produce the plot shown in Fig **??**. Coloring the points by either class or by source, we can see a stronger clustering based on the MARCO classes, this makes sense MARCO was trained to differentiate images based on these classes.

Training an XX architecture using the SimCLR unsupervised learning approach we can produce the plots in Fig **??**. The points were coloured by either class or by source and one observes a strong clustering based on the images' origin. That is, the dominant feature SimCLR observes during a self-supervised training is the image source.

We used the weights from Resnet50 trained upon the MARCO dataset [**JN:** *Which model?*] and used the techniques in [38] to find the decision boundaries for each combination of pairs of classes. Then for each sample in both the C3 and MARCO test sets we find the distance to each decision boundary, see Figure 9.

2.6 Adding local images to the MARCO training set

We evaluated the local data performance with the addition of weakly labelled local data. For these experiments a Resnet50 was used with images of 256 resolution. The results are shown in Figure 10.

Fig 9. Histograms of distances of images from the decision boundaries.
Images from the C3 test set are shown in purple, and those from the MARCO validation set are shown in green. The four classes give 6 decision boundaries, which are shown as clear/grey boxes. The C3 images are closer to the decision boundaries as defined by the MARCO training set, and are are not symmetric around the decision boundaries.

Discussion

The early attempts to develop machine learning tools to interpret crystallisation images failed to come close to human scoring accuracy for (at least) two reasons: firstly, by not having access to a sufficiently large, diverse and well-classified training set and secondly, many of the attempts predated the breakthrough applications of Convolutional Neural Networks (CNNs) in 2012 [41]. The MARCO consortium brought together images from five different institutes, creating a larger and more diverse dataset than had been used before for training crystallisation image classification models, and also tapped into the established expertise in CNNs. The Inception V3 model produced from the MARCO dataset gave impressive results against a test set of data culled from the MARCO data, however, the performance of the model drops significantly when tested against local data.

Approximately 423K images of varying image sizes make up the MARCO training set. These images are either 5 megapixel (MP) images obtained from commercial

Fig 10. Performance of Resnet50 architecture using 256×256 pixel images on the MARCO validation dataset (Left hand graph) and C3 dataset (Right hand graph) when supplementing the training set with different numbers of images from a weakly labelled pool of local images. Trials varied both the number of images included and the class distribution of the included images - blue bars have a uniform class distribution and orange bars show the results when the class distribution of the included images matched the distribution of classes seen in the original MARCO training set (see Table 1). Error bars are the standard deviation of three runs. The dashed red line shows the accuracy of the unmodified training set for comparison. The scales on the two graphs are different: little change in the performance against the MARCO training set is seen irrespective of the number of added images; the accuracy of the model trained with MARCO data supplemented with 1200 local images increases by 6%.

imagers or 2 MP images collected from the custom built imagers at the HWI [42]. The images were labelled with one of four possible labels, 'Crystal', 'Clear', 'Precipitate', 'Other'. The images were initially classified using local classification systems, which totaled 38 different labels. These local categories were mapped to the four meta-classes described above during the collation of the dataset. Subtleties in classification which may have been captured with local classification systems were lost during the relabelling; for example, at CSIRO C3, there are six classes relating to 'Crystal' outcomes (Crystalline, Crystal*, Crystal**, Crystal***, 'shoot me', possible salt crystal, confirmed salt crystal) which all ended up with the same 'Crystal' classification. We were surprised to observe a small number of LCP images in the original MARCO data. Using a binary classification system we estimate that images of LCP droplets make up less than 1% of the images.

Although the MARCO training dataset is large there are still limitations to the both the accuracy of the labels and to the diversity of the data. For example, of the five

groups that contributed images to the training set, four used the same (or similar) type of crystallisation plate to set up the trial droplets - one of the five sitting drop plates manufactured by Swissci UK. There are many other crystallisation plate types on the market (see, for example, https://hamptonresearch.com/cat-187.html), each will have different imaging properties. For trials set up on these different plates to be classified well, the classification model would need to be robust to a domain change or image distribution shift. Interestingly, despite C3 currently using the same Swissci plate type and same Formulatrix imaging system as were used by at least two groups in the original MARCO data, there is a performance reduction for these C3 images. This suggests there are further factors affecting the domain beyond the obvious plate and imager types. Although C3 images were included in the original MARCO data, those images were produced from older equipment, supplied by a different vendor. The more recent C3 images are not represented in the original MARCO data, and thus mimic the situation found in most laboratories (that is, those which were not part of the MARCO consortium).

Within each organisation, more than one set of automation was used to create the images in the MARCO dataset - both the equipment used to set up crystallization droplets and to image them varied within each organisation. For example, the images that C3 contributed to the original MARCO dataset were collected on two different imaging systems, and the drops themselves were set up with three different dispensing robots. The images from Merck were collected on 6 different machines, and the images from GlaxoSmithKline also used 6 different imaging setups.

We suspect that much of the data used to train the MARCO model were not initially compiled with the goal of creating a training set, but were scraped from images that had already been scored for other reasons. If so, it will certainly have affected the quality of the MARCO data. Firstly, the re-purposed images may not be perfectly classified, as creating a "ground truth" dataset was not the goal of the original scoring process. Ensuring accurate classification is difficult, particularly given the range of outcomes seen in a crystallisation trial: experience looking at crystallisation trials is essential for correctly interpreting outcomes. It might be that interns or other early career researchers were hired to classify large numbers of images so that more senior crystallographers could be presented with a set of images which were enriched for

promising results. In this case the classifications would most likely be biased towards $_{355}$ over-estimating positive results, to ensure that any promising image was retained for $_{356}$ further examination. Another consequence might be that less effort would have been $_{357}$ put into classifying the non-crystal containing images correctly; thus non-crystal images $_{358}$ may be more weakly labelled than crystal images, or may simply be underrepresented in $_{359}$ the labelled data. Further, there would have been no reason to ensure that images of $_{360}$ the same drop (collected as part of a time course) were excluded from the dataset. $_{361}$ Figure 4 shows examples of images from timecourses that appear in the dataset. We $_{362}$ have shown that the MARCO data is highly redundant, adding weight to the idea of $_{363}$ MARCO dataset being composed of (at least some) repurposed images. $_{364}$

Including many images from a single timecourse of the same droplet will increase the $_{365}$ absolute size of the training set; however, because these images are likely near $_{366}$ duplicates of each other they can lead to bias in the measurement of the model's $_{367}$ performance [43]. Furthermore, the MARCO data were split into a training set and a $_{368}$ validation (test) set; given the high level of redundancy of the data the training and test $_{369}$ sets would almost certainly not be independent, thus the stated performance of the $_{370}$ model in the work published in 2018 [20] will have been overestimated. We show that $_{371}$ excluding the 20% most redundant images made no difference to the accuracy of the $_{372}$ model, and only a small drop in model performance was observed when 60% of the most $_{373}$ redundant images were excluded. $_{374}$

Redundant images can be images with similar information content, even if they are $_{375}$ not simply different timepoints of the same droplet. Much of the outcome space of a $_{376}$ crystallisation trial is a function of the protein sample in the trial, rather than the $_{377}$ chemical cocktail used. Thus if the same protein is set up against many different $_{378}$ cocktails, the same protein-dependent set of outcomes is likely, so that the diversity of $_{379}$ the image data depends not only on the number of trials, but the number of different $_{380}$ samples used to generate those trials. Industrial protein crystallographers are most $_{381}$ likely to work in large pharmaceutical companies, where they focus on developing SAR $_{382}$ (Structure Activity Relationship) for proteins which are targets for pharmaceutical $_{383}$ intervention. This process requires that a limited number of proteins are subjected to $_{384}$ extensive crystallisation trials, in turn limiting the possible outcome space. Ideally the $_{385}$ images collected for a MARCO type dataset should come from many different $_{386}$

laboratories, so to have as broad a sampling of crystallisation outcome space as possible. ³⁸⁷

The MARCO dataset contains a small (2.5-3K) number of images capturing LCP ³⁸⁸ setups, which would certainly have increased the diversity of the dataset, as LCP ³⁸⁹ experiments consist of a bolus of lipid/protein mesophase surrounded by a small volume ³⁹⁰ of precipitant. The possible outcomes from the LCP trials include breakdown of the ³⁹¹ mesophase or transformation of the phase [31] as well as the outcomes seen in other ³⁹² crystallisation droplets. However, LCP experiments may be sufficiently divergent from ³⁹³ more traditional crystallisation trials as to not expand the outcome space in a useful ³⁹⁴ manner. The MARCO model did not reliably classify LCP images from the C3 ³⁹⁵ laboratory, see Figure 5. This result needs to be tempered by understanding that there ³⁹⁶ were no C3 generated LCP images in the MARCO dataset, and this current work ³⁹⁷ highlights how important including local data in the training set is to the accuracy of ³⁹⁸ classification of local images. However, it could well be that the small number of LCP ³⁹⁹ images in the MARCO data simply do no provide enough coverage of possible LCP ⁴⁰⁰ outcome space to ever provide accurate classification of LCP images. Further, it could ⁴⁰¹ be that the simplistic 4 class classification scheme which only crudely covers soluble ⁴⁰² crystallisation trial outcomes is not granular enough to capture the results seen in LCP ⁴⁰³ experiments. ⁴⁰⁴

To get a more tactile representation of the MARCO data, we used a UMAP ⁴⁰⁵ projection to visualise the MARCO test dataset. UMAP reduces the high-dimensional ⁴⁰⁶ vector that MARCO uses to calculate probabilities into a two dimensional point we can ⁴⁰⁷ visualise in a scatter plot. This approach allows us to identify natural structure that ⁴⁰⁸ occurs in the MARCO data as seen by the MARCO model. Naturally, the ⁴⁰⁹ representations taken from intermediate MARCO layers will be coerced by the ⁴¹⁰ supervised learning process of the MARCO model. To overcome this, a model was ⁴¹¹ trained using SimCLR, a self-supervised method. SimCLR encourages a model to learn ⁴¹² representations it can use to differentiate between images of different things. We ⁴¹³ repeated the dimension reduction approach using the self-supervised model ⁴¹⁴ representations and observed the structures upon which the model learned to focus. As ⁴¹⁵ shown in section Image and dataset characterisation the largest axis of variation in ⁴¹⁶ crystallisation images can be attributed the source laboratory. This indicated a strong ⁴¹⁷ difference between images from different laboratories. ⁴¹⁸

When using an existing model to classify local data there is a domain shift [44], which causes a drop in performance. One way to circumvent this issue is to retrain the model with images from the new source included in the training data; this is the approach taken here. The local data used for retraining were scored (classified) by a single person. As classification of crystallisation images is a difficult problem - both this current work and previous work [23] show that single-person scoring is not robust - we refer to these local images as "weakly labelled". Weakly labelled images were used as these are most readily available in crystallography laboratories, as the procedure for producing a robustly labelled local dataset is expensive in both time and human resources.

In order to provide a retraining protocol that could be adopted by any crystallisation laboratory, we tested different combinations of parameters using recent data from the C3 laboratory. As the more recently acquired C3 images were not included in the MARCO dataset, this mimics the situation found in most crystallisation laboratories. Our results suggest that whilst model capacity (number of parameters) is important for performance on the MARCO test set, it does not help with generalization and may even harm performance at lower image resolutions. The most important factor for inclusion of local data from a laboratory - as suggested by our results using the C3 local data - is image resolution. This supports the assumption presented by Bruno *et al.* [20], that as crystallisation involves detailed, thin structures a larger input image may yield better outcomes. We hypothesize resolution has a smaller impact on the MARCO test set results because images from the MARCO test set share the same data distribution as the training set and the model is able to hallucinate high-resolution features from the low resolution images. However when evaluating the C3 test set where the data distribution is different the model cannot hallucinate these features.

When selecting images to supplement the original MARCO images, we firstly considered the class distribution as it is known to be an important variable [45]. A naïve choice is to randomly select images from a large pool, however, this would likely lead to a biased class distribution which could cause the model to favour any class which is sampled more in the added images. Other options are to match the the label distribution found in the original MARCO dataset; to use a uniform distribution to equally condition each class to the new source images or to match the label distribution

of the local images. In all cases, adding images with a label distribution that matches the original MARCO dataset outperforms adding an equal amount for each class, even when evaluated on the C3 test set which, as shown in Table 1, has a different class distribution. The MARCO test set performance slowly decreases as the number of local images are added which is as expected since the model is using capacity to learn about the local image type. Surprisingly the C3 test set performance increased when 600 and 1200 local images were added but then began to decay when more images were added. This is counter-intuitive as one would expect that adding further local images would continue to increase performance. We conjecture that this decay is due to the weakly labelled nature of the image pool. With a small number of images the model is conditioned to the local images. However, as the number of images increases the poor labels start to negatively impact the classification performance.

The label distribution seen in any laboratory does not necessarily reflect the underlying class distribution, as - most often - not every image is labelled. A human with limited time will tend to label only the more domain-interesting images ('Crystal'), and will often just skip labelling images if they fall into one of the other label classes. We estimate the true distribution of results (as opposed to the distribution of labels) using data from the Cinder mobile application [21]. In this app, images were presented to the viewer one by one, and the viewer uses the app to associate one of four labels to the image. The user can skip labelling an image, but there is a time penalty for doing so. We believe that the labelling statistics obtained from this app are possibly the best estimation available of the true (4-class) distribution in crystallisation. The MARCO and CINDER distributions are shown in Table 1. If we match the MARCO label distribution we will also approximately match the typical class distribution of images.

In other AI problems such as Object Detection, it has been shown that the process of *bootstrapping* or *hard negative mining* can improve performance [46]. Hard negative mining is a process in which examples that are 'easy' are removed from the training set and replaced by 'hard' examples. 'Easy' samples are ones which are correctly classified with a high confidence and 'hard' examples are ones which are incorrectly classified or classified with a low confidence. Such an image selection method could also benefit retraining to reduce performance drop for new images. Intuitively, this suggests that an 'easy' example is already within the original model's distribution - as it was well and

confidently scored by the model, whereas a 'hard' example violates the original model ₄₈₃ and thus would teach the model more. We explored using both entropy and cross ₄₈₄ entropy as hardness measures to select local images for inclusion into the MARCO ₄₈₅ training data. However, neither improved model performance. ₄₈₆

Another measure of hardness is distance from the decision boundary. Samples that ₄₈₇ are clearly distant from a class boundary are simple to classify, as they fall cleanly into ₄₈₈ a single class, but those which lie close to a decision boundary have some ambiguity ₄₈₉ about them, they must have some features that are similar to both classes. Figure 9 ₄₉₀ illustrates two features of the C3 test data as compared to the MARCO validation set. ₄₉₁ We consistently see the mean distance of the C3 Test set to the decision boundary is ₄₉₂ less than the distance from the decision boundary for the MARCO validation set. ₄₉₃ Furthermore, we can observe that a large fraction of the C3 test sample cross over the ₄₉₄ decision boundary, indicating that they are incorrectly classified by the reference model. ₄₉₅ These features are not only illustrative of the increased difficulty of the C3 dataset over ₄₉₆ the MARCO dataset for the MARCO model, but are also indicative of the distribution ₄₉₇ shift between datasets. The stark difference in distributions from C3 and MARCO in ₄₉₈ figure 9 demonstrate the necessity to retrain or fine-tune the MARCO model when ₄₉₉ moving between labs to better align decision boundaries. The greater distance between ₅₀₀ the boundaries and the MARCO images could come from two sources - firstly, it could ₅₀₁ simply show that the validation set is not independent of the training set, which is likely ₅₀₂ given the redundancy of the data, but also it could support the notion that the MARCO ₅₀₃ dataset simply does not contain many ambiguous or difficult-to-classify images. ₅₀₄

The UMAP projection of the unsupervised training using the MARCO data show ₅₀₅ that the axes of largest variation of crystallisation images in the SimCLR representation ₅₀₆ space are not equivalent to the labels associated by humans, but are dependant on the ₅₀₇ laboratory and equipment used to collect the images. ₅₀₈

Conclusion ₅₀₉

Although the MARCO model developed in 2018 has had a huge impact by producing ₅₁₀ reliable classifications of crystallisation images, it was believed that the success of the ₅₁₁ model was due to the extent of the training dataset. We have shown that the training ₅₁₂

data are redundant, and the true extent of the original MARCO data is probably less than 50% of the data that were used. The MARCO performance is significantly less impressive on local data than on test data derived from the same sources as the training data. We recommend including robustly labelled local data; further, our work suggests that there is an upper limit to the number of poorly labelled local images that can be included without decreasing model performance. For our local data we observe this value to be around 1000 local images.

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References

1. Burley SK, Berman HM, Bhikadiya C, Bi C, Chen L, Di Costanzo L, et al. RCSB Protein Data Bank: biological macromolecular structures enabling research and education in fundamental biology, biomedicine, biotechnology and energy. Nucleic Acids Research. 2019;47:D464–D474. doi:10.1093/nar/gky1004.

2. Calero G, Cohen AE, Luft JR, Newman J, Snell EH. Identifying, studying and making good use of macromolecular crystals. Acta Crystallographica Section F. 2014;70(8):993–1008. doi:10.1107/S2053230X14016574.

3. Galli S. X-ray Crystallography: One Century of Nobel Prizes. Journal of Chemical Education. 2014;91(12):2009–2012. doi:10.1021/ed500343x.

4. Bárcena M, Barnes CO, Beck M, Bjorkman PJ, Canard B, Gao GF, et al. Structural biology in the fight against COVID-19. Nature Structural & Molecular Biology. 2021;28(1):2–7. doi:10.1038/s41594-020-00544-8.

5. Lynch ML, Snell EH, Bowman SEJ. Structural biology in the time of COVID-19: perspectives on methods and milestones. IUCrJ. 2021;8(3):335–341. doi:10.1107/S2052252521003948.

6. McPherson A. Current approaches to macromolecular crystallization. European Journal of Biochemistry. 1990;189(1):1–23. doi:10.1111/j.1432-1033.1990.tb15454.x.

7. Chayen NE. Tackling the bottleneck of protein crystallization in the post-genomic era. Trends in Biotechnology. 2002;20(3):98.

8. Luft JR, Newman J, Snell EH. Crystallization screening: the influence of history on current practice. Acta Crystallographica Section F. 2014;70(7):835–853. doi:10.1107/S2053230X1401262X.

9. Bergfors T. Protein Crystallization. 2nd ed. IUL Biotechnology Series. International University Line; 2009.

10. Manjasetty BA, Turnbull AP, Panjikar S, Büssow K, Chance MR. Automated technologies and novel techniques to accelerate protein crystallography for structural genomics. PROTEOMICS. 2008;8(4):612–625. doi:10.1002/pmic.200700687.

11. McPherson A, Gavira JA. Introduction to protein crystallization. Acta Crystallographica Section F. 2013;70(1):2–20. doi:10.1107/S2053230X13033141.

12. Meijers R, Mueller-Dieckmann J. Advances in High-Throughput Crystallisation. In: John Wiley & Sons, Ltd, editor. eLS. John Wiley & Sons, Ltd; 2011. p. a0023171. Available from: http://doi.wiley.com/10.1002/9780470015902.a0023171.

13. Ng JT, Dekker C, Reardon P, von Delft F. Lessons from ten years of crystallization experiments at the SGC. Acta Crystallographica Section D. 2016;72(2):224–235. doi:10.1107/S2059798315024687.

14. Mele K, Lekamge BMT, Fazio VJ, Newman J. Using Time Courses To Enrich the Information Obtained from Images of Crystallization Trials. Crystal Growth & Design. 2013;doi:10.1021/cg4014569.

15. Cumbaa C, Jurisica I. Automatic Classification and Pattern Discovery in High-throughput Protein Crystallization Trials. Journal of Structural and Functional Genomics. 2005;6(2):195–202. doi:10.1007/s10969-005-5243-9.

16. Pan S, Shavit G, Penas-Centeno M, Xu DH, Shapiro L, Ladner R, et al. Automated classification of protein crystallization images using support vector machines with scale-invariant texture and Gabor features. Acta Crystallographica Section D. 2006;62(3):271–279.

17. Spraggon G, Lesley SA, Kreusch A, Priestle JP. Computational analysis of crystallization trials. Acta Crystallographica Section D. 2002;58(11):1915–1923.

18. Watts D, Cowtan K, Wilson J. Automated classification of crystallization experiments using wavelets and statistical texture characterization techniques. Journal of Applied Crystallography. 2008;41:8–17. doi:10.1107/S0021889807049308.

19. Szegedy C, Vanhoucke V, Ioffe S, Shlens J, Wojna Z. Rethinking the Inception Architecture for Computer Vision. arXiv:151200567 [cs]. 2015;.

20. Bruno AE, Charbonneau P, Newman J, Snell EH, So DR, Vanhoucke V, et al. Classification of crystallization outcomes using deep convolutional neural networks. PLoS ONE. 2018;13(6):e0198883. doi:10.1371/journal.pone.0198883.

21. Rosa N, Ristic M, Marshall B, Newman J. *Cinder* : keeping crystallographers app-y. Acta Crystallographica Section F. 2018;74(7):410–418. doi:10.1107/S2053230X18008038.

22. Newman J, Bolton EE, Müller-Dieckmann J, Fazio VJ, Gallagher DT, Lovell D, et al. On the need for an international effort to capture, share and use crystallization screening data. Acta Crystallographica Section F. 2012;68(3):253–258. doi:10.1107/S1744309112002618.

23. Wilson J. Automated classification of images from crystallisation experiments. In: Advances in data mining applications in medicine, web mining, marketing, image and signal mining: 6th Industrial Conference on Data Mining, ICDM 2006, Leipzig, Germany, July 14-15, 2006 : proceedings. Springer; 2006. p. 459–473. Available from: `http://site.ebrary.com/id/10530328`.

24. William Edwards II D, Dinc I. Classification of Protein Crystallization Images using EfficientNet with Data Augmentation. In: CSBio '20: Proceedings of the

Eleventh International Conference on Computational Systems-Biology and Bioinformatics. ACM; 2020. p. 54–60. Available from: https://dl.acm.org/doi/10.1145/3429210.3429220.

25. Pan SJ, Yang Q. A Survey on Transfer Learning. IEEE Transactions on Knowledge and Data Engineering. 2010;22:1345–1359.

26. Huh M, Agrawal P, Efros AA. What makes ImageNet good for transfer learning? ArXiv. 2016;abs/1608.08614.

27. Ding J, Li X. An Approach for Validating Quality of Datasets for Machine Learning. In: 2018 IEEE International Conference on Big Data (Big Data); 2018. p. 2795–2803.

28. Weiss K, Khoshgoftaar TM, Wang D. A survey of transfer learning. Journal of Big Data. 2016;3(1):9. doi:10.1186/s40537-016-0043-6.

29. Thompson NC, Greenewald K, Lee K, Manso GF. The Computational Limits of Deep Learning; 2020. Available from: http://arxiv.org/abs/2007.05558.

30. He K, Zhang X, Ren S, Sun J. Deep Residual Learning for Image Recognition. arXiv:151203385 [cs]. 2015;.

31. Caffrey M, Cherezov V. Crystallizing membrane proteins using lipidic mesophases. Nature Protocols. 2009;4(5):706–731. doi:10.1038/nprot.2009.31.

32. Sandler M, Howard AG, Zhu M, Zhmoginov A, Chen LC. MobileNetV2: Inverted Residuals and Linear Bottlenecks. 2018 IEEE/CVF Conference on Computer Vision and Pattern Recognition. 2018; p. 4510–4520.

33. Birodkar V, Mobahi H, Bengio S. Semantic Redundancies in Image-Classification Datasets: The 10

34. Defays D. An efficient algorithm for a complete link method. The Computer Journal. 1977;20(4):364–366. doi:10.1093/comjnl/20.4.364.

35. Lloyd S. Least squares quantization in PCM. IEEE Transactions on Information Theory. 1982;28(2):129–137. doi:10.1109/TIT.1982.1056489.

36. Chen T, Kornblith S, Norouzi M, Hinton G. A Simple Framework for Contrastive Learning of Visual Representations. In: III HD, Singh A, editors. Proceedings of the 37th International Conference on Machine Learning. vol. 119 of Proceedings of Machine Learning Research. PMLR; 2020. p. 1597–1607. Available from: https://proceedings.mlr.press/v119/chen20j.html.

37. McInnes L, Healy J, Saul N, Großberger L. UMAP: Uniform Manifold Approximation and Projection. Journal of Open Source Software. 2018;3(29):861. doi:10.21105/joss.00861.

38. Liu W, Wen Y, Yu Z, Li M, Raj B, Song L. SphereFace: Deep Hypersphere Embedding for Face Recognition. In: The IEEE Conference on Computer Vision and Pattern Recognition (CVPR); 2017.

39. Buchala S, Wilson JC. Improved classification of crystallization images using data fusion and multiple classifiers. Acta Crystallographica Section D. 2008;64(8):823–833.

40. Desbois S, Seabrook SA, Newman J. Some practical guidelines for UV imaging in the protein crystallization laboratory. Acta Crystallographica Section F. 2013;69(2):201–208. doi:10.1107/S1744309112048634.

41. Krizhevsky A, Sutskever I, Hinton GE. ImageNet classification with deep convolutional neural networks. Communications of the ACM. 2017;60(6):84–90. doi:10.1145/3065386.

42. Snell EH, Luft JR, Potter SA, Lauricella AM, Gulde SM, Malkowski MG, et al. Establishing a training set through the visual analysis of crystallization trials. Part I: ∼150 000 images. Acta Crystallographica Section D. 2008;64(11):1123–1130. doi:10.1107/S0907444908028047.

43. Birodkar V, Mobahi H, Bengio S. Semantic Redundancies in Image-Classification Datasets: The 10% You Don't Need. arXiv:190111409 [cs, stat]. 2019;.

44. Sun B, Feng J, Saenko K. Return of Frustratingly Easy Domain Adaptation. In: Proceedings of the Thirtieth AAAI Conference on Artificial Intelligence. AAAI'16. AAAI Press; 2016. p. 2058–2065.

45. Weiss GM, Provost F. The effect of class distribution on classifier learning: an empirical study. Rutgers University; 2001.

46. Shrivastava A, Gupta A, Girshick R. Training Region-Based Object Detectors with Online Hard Example Mining. In: 2016 IEEE Conference on Computer Vision and Pattern Recognition (CVPR); 2016. p. 761–769.