Temperature as a likely driver shaping global patterns in

mineralogical composition in bryozoans:

Implications for marine calcifiers under Global Change

Running title: Temperature patterns of bryozoan mineralogy

Blanca Figuerola[1], Huw J. Griffiths[2], Malgorzata Krzeminska[3], Anna Piwoni-

Piorewicz[4], Anna Iglikowska[5], Piotr Kuklinski[3]

1. Institute of Marine Sciences (ICM-CSIC), Barcelona, Spain

2. British Antarctic Survey, Cambridge, United Kingdom

3. Institute of Oceanology, Polish Academy of Sciences, Sopot, Poland

4. Institute of Oceanography, University of Gdańsk, Gdynia, Poland

5. Faculty of Biology, University of Gdańsk, Wita Stwosza 59, 80-308 Gdańsk,

Poland

*Corresponding author: bfiguerola@gmail.com / figuerola@icm.csic.es

Abstract

The Southern Ocean is showing one of the most rapid responses to human-induced

global change, thus acting as a sentinel of the effects on marine species and

ecosystems. Ocean warming and acidification are already impacting benthic species

with carbonate skeletons, but the magnitude of these changes to species and

ecosystems remains largely unknown. Here we provide the largest carbonate

mineralogical dataset to date for Southern Ocean bryozoans, which are diverse,

abundant and important as carbonate producers, thus making them excellent for

monitoring the effects of ocean warming and acidification. To improve our

understanding of how bryozoans might respond to ocean warming and acidification,

we assess latitudinal and seafloor temperature patterns of skeletal mineralogy using

bryozoan species occurrences together with temperature data for the first time. Our

findings, combining new mineralogical data with published data from warmer

regions, show that the proportions of high-Mg calcite and bimineralic species

increase significantly towards lower latitudes and with increasing seawater

temperature. These patterns are consistent with the hypothesis that seawater

temperature is likely a significant driver of variations in bryozoan mineralogy at a

global scale.

Keywords: calcifying organisms, aragonite, Mg-calcite, ocean warming, acidification,

Antarctic

1 INTRODUCTION

Increased levels of atmospheric carbon dioxide (CO_2) as a result of human activities are driving rising temperatures, ocean acidification and consequent impacts on marine ecosystems (Kroeker et al., 2013). Climate change models predict a global increase in mean sea surface temperature of 1–4ºC and a decrease of ~0.3–0.4 pH by the year 2100 (IPCC, 2014, 2019). The Southern Ocean, the water masses extending south of the Polar Front to the coasts of the Antarctic continent, is predicted to be among the first and most highly affected regions by ocean acidification due to increased solubility of CO_2 at low temperatures leading to low carbonate ion concentrations ($[CO_3^{2-}]$) (Fabry et al., 2009; Orr et al., 2005). Ocean acidification is also shallowing the saturation horizon, which is the depth below which calcium carbonate ($CaCO_3$) minerals (such as aragonite and calcite) can dissolve. It is anticipated that surface waters in some Southern Ocean regions will become undersaturated with respect to aragonite and calcite by 2050 and 2095, respectively (McNeil & Matear, 2008; Orr et al., 2005), and that 70% of the Southern Ocean could be aragonite undersaturated by 2100 (Hauri et al., 2016). Due to the low interannual variability in surface ocean $[CO_3^{2-}]$ of this region (Conrad & Lovenduski, 2015), many organisms may be unable to adapt rapidly to these changing conditions. Some parts of the continent, such as the Western Antarctic Peninsula, are also among the fastest warming regions globally (Steig et al., 2009). The Southern Ocean acts as a sentinel region for evaluating the impacts of ocean warming and acidification on marine organisms and for making predictions about future impacts at lower latitudes where ocean acidification is expected to occur later (Andersson et al., 2008; Fabry et al., 2009; Figuerola et al., 2021).

Understanding how marine organisms can cope with these environmental changes is

essential for predicting the future of marine ecosystems. Marine calcifiers, organisms

which build their skeletons from $CaCO_3$, may be particularly sensitive to these

threats. Increased pCO$_2$ and decreased pH, caused by ocean acidification, increase

the solubility of $CaCO_3$ skeletons (Andersson et al., 2008; Fabry, 2008; Figuerola et

al., 2021). The ability of marine calcifiers to respond to these rapid environmental

stresses may, therefore, be compromised, especially for Southern Ocean taxa,

which are often characterized by having lower metabolic, development and growth

rates than related species from warmer latitudes (Peck, 2018; Peck, 2001, 2005).

Southern Ocean taxa that deposit more soluble $CaCO_3$ mineral phases such as

aragonite and high Mg-calcite could be at greater risk to changes in ocean carbonate

chemistry than species secreting a mixture of calcite and aragonite (bimineralic) or

lower Mg-calcite levels (Andersson et al., 2008; Figuerola et al., 2021; Lebrato et al.,

2013; McClintock et al., 2011, 2017). For instance, common habitat-forming or key

predator taxa, which have important roles in Antarctic ecosystems, have aragonitic

(e.g. scleractinian cold-water corals) or high Mg-calcite skeletons (e.g. echinoderms

and bryozoans) (Bax & Cairns, 2014; Figuerola et al., 2019; McClintock et al., 2011;

Turley et al., 2007). The negative ocean acidification impact on these taxa could thus

lead to the loss of habitat complexity and resilience in these ecosystems.

Ocean warming may even accelerate skeletal solubility as a positive correlation

between Mg-calcite and/or aragonite and seawater temperature has been found in

diverse phyla such as molluscs, serpulid annelids, foraminifera, corals, echinoderms,

bryozoans and crustaceans (Chang et al., 2004; Chave, 1954; Cohen & Branch,

1992; Lowenstam, 1954). Seawater temperature seems to affect the spatial

distribution of marine calcifiers, with higher abundances of species depositing

aragonite and/or higher Mg-calcite levels in the tropics, although this pattern has yet

to be rigorously tested in some groups such as bryozoans.

In addition, there remain uncertainties in the predicted effects of ocean warming on

marine calcifiers because diverse environmental factors (e.g. salinity, pH, $CaCO_3$

saturation state, seawater Mg/Ca ratio and diet) and biological (e.g. skeletal growth

rate) may interfere in the calcification process, especially on a local scale (Amsler et

al., 2010; Borremans et al., 2009; Duquette, Vohra, et al., 2018; Figuerola et al.,

2015; Krzeminska et al., 2016; Kołbuk et al. 2019; Krzemińska et al., 2022; Lebrato

et al., 2016; Lowenstam & Weiner, 1989; Loxton, Kuklinski, Barnes, et al., 2014;

Ries, 2010). For instance, variability in seawater Mg/Ca ratios (Lebrato et al., 2020)

is known to affect the skeletal Mg-calcite and mineralogy of a range of species (Ries,

2010; Kołbuk et al. 2021). Among biological processes, "vital effects" may distort or

mask any signal of environmental effects on mineralogy (Weiner & Dove, 2003).

Plasticity is also a mechanism by which calcifiers such as molluscs may adapt to

environmental changes, as observed when organisms are exposed to different

experimental conditions (Figuerola et al., 2021; Hancock et al., 2020). Therefore,

more studies are needed on a wider range of phyla and species with broad

geographical ranges, skeletal mineralogies, biological traits and physiological

processes.

Among calcifying taxa, bryozoans are abundant and speciose suspension-feeding

aquatic invertebrates worldwide (>6000 extant species; Bock and Gordon, 2013),

being present from the intertidal to the deep sea (e.g. Barnes and Kuklinski, 2010;

Figuerola et al., 2018). They are dominant members of many benthic communities,

including the Antarctic region (Barnes & Kuklinski, 2010; Figuerola et al., 2012).

Many species are major carbonate sediment producers, thus being well represented

in the fossil record (Taylor, 2020) and good model systems for understanding

ecological and evolutionary processes (Orr et al., 2022). The calcareous skeletons of

bryozoans can be entirely calcitic, aragonitic or bimineralic, and incorporate varying

amounts of Mg in their calcite, ranging from low Mg-calcite (LMC; <4 wt% $MgCO_3$),

through intermediate Mg-calcite (IMC; 4–8%), to high Mg-calcite (HMC; >8%)

following the categories proposed by Rucker and Carver (1969) (see Taylor et al.,

2015).

Early studies exploring marine invertebrate mineralogy proved cheilostome

mineralogy to be highly variable, unlike other taxa, thus having great potential for

reconstruction changes (Lowenstam, 1954; Poluzzi & Sartori, 1975; Rucker &

Carver, 1969). The mineralogical studies have persisted and proliferated even to

today (e.g., Bone and James, 1993; Borisenko and Gontar, 1991; Smith et al., 1998).

In recent years significant efforts have been put into increasing data on skeletal

mineralogy in a wide range of bryozoan species from Arctic (Iglikowska et al., 2020;

Krzemińska et al., 2022; Kuklinski & Taylor, 2009; Piwoni-Piórewicz et al., 2019),

temperate (e.g., Loxton et al., 2014b; Smith and Clark, 2010; Steger and Smith,

2005), tropical (Taylor et al., 2016; Taylor & Di Martino, 2014), and Antarctic

(Figuerola et al., 2015, 2019; Krzeminska et al., 2016; Loxton et al., 2013; Loxton,

Kuklinski, Barnes, et al., 2014) regions. However, several reviews have pointed out

that mineralogy is known only for a small proportion of living species at all latitudes

(Smith et al., 2006; Taylor et al., 2009).

Southern Ocean bryozoans, which comprise over 300 species, exhibit interspecific variations in the Mg-calcite of their skeletons, and wide geographic and depth distributions (Barnes & Kuklinski, 2010; De Broyer et al., 2011; Figuerola et al., 2012, 2017; Hayward, 1995; Krzeminska et al., 2016), making them suitable indicators to monitor the effects of ocean warming and acidification in a hotspot for global change. As ecosystem engineers, this group also creates habitat heterogeneity and refugia for many associated macro- and micro-invertebrates and thus are likely to boost ecological resilience (Santagata et al., 2018). In this study, we present the largest carbonate mineralogical dataset for Southern Ocean bryozoans to date. To improve our understanding of how bryozoans might respond to ocean warming and acidification globally, we then employ rigorous correlative statistical analyses to assess latitudinal and seafloor temperature patterns of skeletal mineralogy. We hypothesized that seawater temperature is the main driver on influencing mineralogical composition in bryozoans at a global scale as maintaining aragonitic and HMC skeletons is more energetically demanding in cold waters. Thus, we expect that the seawater temperature affects spatial distribution of bryozoans. To our knowledge, this is the first study to test this hypothesis using a seafloor temperature data set and species occurrences combined with new and published mineralogical data.

2 MATERIAL AND METHODS

2.1 Study area

Bryozoan specimens were collected from East Antarctica using beam trawl during the CEAMARC cruise on the RSV 'Aurora Australis' (2007 to 2009). The study area

covers part of the region Terre Adélie (141–142° E) and George V Land (142–145°

E). Collection depths ranged from 138–662 m. Sampling sites were geo-referenced

using GPS and depth was recorded at each site (Fig. 1; Table S1; Tables S1–4 are

given   in   the   Dryad   Digital   Repository:

https://datadryad.org/stash/share/3ZheuFuQESDCw-

AQUt10tE7f2ol5uIDSe0Vv5laUtBQ). On the research vessel, all bryozoan samples

were preserved in 95% ethanol. Bryozoan colonies were identified to the lowest

possible taxonomic rank in the laboratory using an optical microscope and a

taxonomic reference guide (Hayward, 1995). To confirm identification, a subset of

specimens was bleached for 1–4 hours in a 10% solution of sodium hypochlorite

(domestic bleach, diluted using tap water), rinsed in freshwater, air-dried, and

examined using a LEO SEM at the Natural History Museum in London (Fig. 2).

2.2 Mineralogical analyses

Specimens were cleaned carefully of epibionts using a scalpel blade to avoid

mineralogical contamination. If possible a piece was cut from the growing edge of

each specimen and air dried as described in previous studies (Figuerola et al., 2015;

Kuklinski & Taylor, 2009). Otherwise any available part of colony was used for

analysis. The pieces were powdered using an agate pestle and mortar. Mineralogical

analyses were carried out at the Natural History Museum using a high-precision

Enraf-Nonius X-ray diffractometer (XRD) equipped with an INEL CPS-120 Curved

Position Sensitive detector (–120°, 2-theta) and a cobalt X-ray source. Operating

conditions of the cobalt source were 40 kV and 40 mA. The tilt angle between source

and sample was 5.9°. The horizontal slit system was set to 0.14 mm to confine the

X-ray beam to pure cobalt Kα1. An internal standard (NaCl) was added to the

samples to correct for sample displacement. The samples were rotated during the

measurements to improve the randomness of grain orientations in the X-ray beam.

The 2-theta linearity of the detector was calibrated using Y_2O_3 (yttrium (III) oxide)

(99.9% BDH Laboratory Chemicals) and SRM 640 silicon powder (NIST) and the

calibration curve was fitted using a least-squares cubic spline function. Peak

positions were calibrated on Y_2O_3 data from Mitric et al. (1997). Bryozoan skeleton

was compared with external standard of calcite (Iceland spar) by fitting peak

intensities of the sample patterns and the standard patterns. The wt% $MgCO_3$ in

calcite was calculated by measuring the position of the d_{104} peak, assuming a linear

interpolation between $CaCO_3$ and $MgCO_3$ (Mackenzie et al., 1983). A linear trend of

d_{104} vs. mol% $MgCO_3$ can be observed in the range between 0 and 17 mol% $MgCO_3$

(Mackenzie et al. 1983); all analyses from the present study fall within this range.

This composition information is accurate to within 2% on a well-calibrated instrument

(Kuklinski & Taylor, 2009).

2.3 Temperature data

The seafloor temperature data set employed was that presented in Clarke et al.

(2009). Seawater temperature values were taken from the World Ocean Atlas 2005,

a data product of the Ocean Climate Laboratory of the U.S. National Oceanographic

Data Centre (http://www.nodc.noaa.gov/OC5/WOA05/pr_woa05.html). Global mean

annual ocean *in situ* temperature data interpolated into 33 standard vertical intervals

(depth ranges) at a 1° spatial resolution were overlain onto a bathymetry grid

(ETOPO1) (Amante & Eakins, 2009). Seawater temperature data were then

extracted when the depth range coincided with the bathymetry of the seafloor to

create a seawater temperature data set. Potential temperature was calculated

according to the UNESCO equation of state for seawater and the International Temperature Scale of 1990 (ITS-90) specified by the International Committee of Weights and Measures. A nearest neighbor interpolation was then used to create a seafloor temperature grid for both *in situ* and potential temperature in ArcGIS. Temperatures were extrapolated for each bryozoan occurrence record (latitude and longitude) to assign a temperature value.

2.4 Literature search and bryozoan species distributions

We integrated new data from the current study with recent (e.g., Krzeminska et al., 2016; Loxton et al., 2018) and older mineralogy data from cheilostome and cyclostome bryozoans worldwide previously summarized by Smith et al. (2006) and Taylor et al. (2009). Available mineralogical data from genera with undetermined species, and also known species from the White Sea which has unusual conditions that may mask temperature influence (Krzemińska et al., 2022), were removed from the analyses (see Tables S2–S4). The global dataset comprised 549 species (102 Southern Ocean species, excluding six genera with indeterminate species). Calcite composition was classified as LMC, IMC and HMC. Data previously reported as mol% $MgCO_3$ were converted to weight% (wt%) $MgCO_3$ to facilitate comparisons.

To generate the mineral type and skeletal Mg-calcite distribution maps and graphs, we downloaded occurrence data for all living bryozoan species from the Global Biodiversity Information Facility (GBIF, n.d.) (GBIF.org (28 January 2022) GBIF Occurrence Download https://doi.org/10.15468/dl.59nzhf). We used our list of 549 species with available mineralogical data (with species names verified using the World Register of Marine Species Taxon Match tool -

) to filter the downloaded GBIF

data. Duplicate occurrence records (repeated combinations of latitude, longitude,

and scientific name) and any occurrences on land were removed. The filtered GBIF

dataset comprised 96,241 occurrences for the 493 selected species that had

occurrence records within GBIF (Table S5).

2.5 Statistical analysis

Coefficients of determination (R^2) for the regressions and p values were calculated to

evaluate correlative relationships between latitude and temperature and

mineralogical composition. Data were analysed and figures constructed in: R v.

4.1.0/Rstudio v. 1.4.1717, using the R packages Tidyverse (ggplot2) and Ggally v.

2.1.2 (Schloerke et al., 2021; Wickham et al., 2019). Statistical significance was set

at $p < 0.05$.

3 RESULTS

3.1 Skeletal mineralogy of the Southern Ocean and warmer regions

Among the new samples (n = 126) analysed in the present study, 37 Antarctic

bryozoan species had skeletons consisting of calcite and were characterized by a

wide range of Mg-calcite in the skeleton (1.52–7.55 wt% $MgCO_3$; Fig. 3; Table S2).

The mean Mg-calcite levels for all analysed samples was 4.15 ± 1.28 wt% $MgCO_3$.

Fifty-seven percent of the species we examined were categorized as IMC (21

species), whereas the remaining 43% were categorized as LMC (16 species; Fig. 3).

The resulting data set, when combined similar data from the literature, included 102

Southern Ocean species (n = 1415; Table S3). The average Mg-calcite from all

analysed samples was 4.54 ± 1.32 wt% $MgCO_3$. Most species fell into the IMC

category (67%; 68 species) while the remaining were in the LMC category (32%; 33

species) apart from *Beania erecta* Waters, 1904 which was in the HMC category

(1%; 8.03 ± 1.56 wt% $MgCO_3$) (Fig. 4A; Table S3).

Globally, 72% (397 species) of a total of 549 species were calcitic, 22% (122

species) bimineralic and 6% (30 species) aragonitic (Fig. 5A). Most bryozoan

species had IMC (65%; 322 species) while the remaining had LMC (27%; 133

species) or HMC (8%; 41 species) (Fig.6A).

3.2 Global-scale latitudinal patterns of skeletal mineralogy

The proportions of occurrence records of different mineral types remained stable

south of the Polar Front, with aragonite almost completely absent, bimineralic

occurrence records less than 3% with most occurrence records (97%) dominated by

calcite. North of the Polar Front was also dominated by calcite but to a lower level

(~62–97%), higher levels of bimineralic skeletons (~3–32% and 100% for a single

occurrence record south of 60º S) (Table S5). There was also a trend of increasing

aragonite (from zero up to a maximum of ~9.5%) with decreasing latitude (Fig. 4C).

There was a significant increase in the percentage of bimineralic occurrence records

(R^2 = 0.16; *p* = 0.017) and decrease of calcitic occurrence records (R^2 = 0.19; *p* =

0.01) with an increase in seafloor water temperature (Fig. 5E). Aragonitic occurrence

records were absent below 0ºC and very low below 1ºC (0.05%), with no significant

trend in waters above that temperature (*p* = 0.11) with proportions ranging between

1% and 11% of the total number of occurrences.

The proportions of LMC, IMC and HMC occurrence records did not vary south of the Polar Front either (Fig. 4C). IMC dominated occurrence records (60–68%), 30% to 36% of occurrence records were LMC and HMC was only present in very low proportions (1.5–5%). However, HMC species significantly increased in prevalence from the Polar Front towards lower latitudes (R^2 = 0.58; p = 0.0025; Fig. 4C) and with increasing seawater temperatures (R^2 = 0.48; p = $3.6e^{-06}$; Fig. 6E). IMC species had the opposite trend, with a decrease towards lower latitudes (R^2 = 0.26; p = 0.077; Fig. 4C) and a significant decrease with increasing seawater temperatures (R^2 = 0.45; p = $1.1e^{-06}$; Fig. 6E). No significant trend was found in LMC species north of the Polar Front with latitude or with temperature.

4. DISCUSSION

4.1 Seawater temperature as a likely driver influencing mineralogical composition at a global scale

In the present study we have compiled, and in some cases analysed, the largest carbonate mineralogical and distribution dataset for Southern Ocean bryozoans to date, including data of approximately one-third of the known extant bryozoan species from Antarctica based on those listed in the Register of Antarctic Marine Species database (102 of 335 species) (De Broyer et al., 2022). This dataset thus increases our knowledge of the potential effects of global change on Antarctic bryozoans, an important phylum of invertebrates in the Antarctic benthos which creates framework habitats that enhance biodiversity and produces large quantities of carbonate sediments, thus making up significant blue carbon sinks (Barnes, 2018). These data provide a useful baseline for understanding the vulnerability of different bryozoan

species and make predictions of the effects of ocean acidification on benthic habitats and sediments.

Our results confirm that all Southern Ocean species have monomineralic calcite skeletons, as previously revealed by several studies on bryozoan skeletal mineralogy (Figs. 3 and 4) (Borisenko & Gontar, 1991; Figuerola et al., 2015, 2019; Krzeminska et al., 2016; Loxton et al., 2013; Loxton, Kuklinski, Barnes, et al., 2014; Taylor et al., 2009). While occasional analyses of specimens belonging to the bryozoan species *Chondriovelum* [*Labioporella*] *adeliensis* (Livingstone, 1928) and *Isosecuriflustra angusta* (Kluge, 1914) detected aragonite (Borisenko & Gontar, 1991; Loxton et al., 2013), recent analyses including the current study contradict these findings and provide evidence of their monomineralic calcitic nature (Krzeminska et al., 2016; Taylor et al., 2009). We suggest that these detections of aragonite are likely due to sample contamination by epibionts or sediment (Taylor et al., 2015).

Our data illustrate that the skeletons of most Southern Ocean species of bryozoans are formed of IMC, some of LMC and a few of HMC, consistent with previous global data (Figs. 3 and 4; Table S3) (Smith et al., 2006; Taylor et al., 2009). However, we found higher IMC (67 vs 65–58%) and LMC (32 vs 29–27%) percentages and a lower HMC (1 vs 13–8%) percentage compared to those from our own and previous global data this was as expected due to the higher solubility of HMC in cold waters (Fig. 6A) (Davis et al., 2000; Mucci, 1983; Smith et al., 2006; Taylor et al., 2009).

Our findings also reveal that the proportions of skeletal aragonite, bimineralic

mixtures and calcite remained stable south of the Polar Front consistent with the low

and nearly constant temperatures found in the Southern Ocean (Fig. 4 C) (Moore et

al., 1999). From the Polar Front boundary towards lower latitudes, we did not detect

any clear general latitudinal trend (Fig. 4 C) as previously found in studies that did

not extrapolate values onto wider occurrence data (e.g. Taylor et al., 2009).

However, when we plotted our mineralogical findings against the seafloor

temperature, there were an increase in the proportions of bimineralic occurrence

records (R^2 = 0.16; p = 0.017) and a decrease in the proportion of calcitic occurrence

records (R^2 = 0.19; p = 0.01) with increasing temperature consistent with decreased

costs of skeletal production (Fig. 5E). The lack of a significant increase in aragonite

occurrence records with increasing temperature (R^2 = 0.079; p = 0.11) is potentially

due to a lack of published mineralogical studies of recent bryozoans from the tropics,

where most species with aragonitic skeletons occur (Taylor et al., 2009, 2016).

However, species with aragonitic skeletons are absent from seas with temperatures

of zero degrees centigrade or below, indicating that this is some sort of temperature

threshold or limit for aragonitic species.

In addition, two cosmopolitan aragonitic species, *Membranipora membranacea*

(Linnaeus, 1767), which had high occurrence records (6759), and *Hippothoa*

divaricata Lamouroux, 1821, which had some Antarctic occurrence records (n = 21),

could mask the general trend (Table S5). These species could also switch in

mineralogy in higher latitudes, although they were not found below zero degrees

Celsius. Some mineralogical analyses of *M. membranacea* recovered calcite (Taylor

et al., 2009), although it is not clear if it is due to taxonomic misidentification.

Interestingly, the only Antarctic occurrence records of *H. divaricata* are from the

Western Antarctic Peninsula, where the water is warmer than other Antarctic

regions. We thus suggest that zero degrees Celsius could be a better cut off point,

above which aragonite does not show a significant trend towards the equator.

However, further taxonomic studies are needed to confirm the identification of this

species. It should be also taken into account that our analyses make the assumption

that species do not switch mineralogy, although it has been found in some bryozoan

species (e.g. Smith and Girvan, 2010), and might impact some of our findings.

Mg-calcite remained low and constant south of the Polar Front and significantly

increased towards lower latitudes (HMC R^2 = 0.58; *p* = 0.003) and with increasing

seafloor water temperatures (HMC R^2 = 0.48; *p* < 0.001), consistent with our

hypothesis that seawater temperature influence on bimineralization seems to affect

the spatial distribution of bryozoans. For the first time we thus provide evidence that

seafloor water temperature is likely a significant driver influencing mineralogical

composition in bryozoans at a global scale, with an increase of HMC and bimineralic

species in warmer waters. Previous studies, lacking an assessment of temperature,

revealed an inverse correlation between Mg-calcite and latitude in different calcifying

marine invertebrates (Andersson et al., 2008; Duquette, Halanych, et al., 2018;

McClintock et al., 2011). For instance, this general pattern was observed in

echinoderm species with LMC skeletons only found in Antarctica although most

species have HMC skeletons (Duquette, Halanych, et al., 2018; Figuerola et al.,

2021). Smith et al. (2016) also found a similar trend in different life stages and nine

skeletal elements of echinoids. Other taxa also have LMC shells at higher latitudes

such as Antarctic brachiopods (Barnes & Peck, 1996). In the case of bryozoans, a

latitudinal gradient in cheilostome mineralogy, in which aragonite and Mg-calcite

increase towards the tropics, was also observed (Krzeminska et al., 2016; Piwoni-

Piórewicz et al., 2019; Taylor et al., 2009). Kuklinski & Taylor (2008, 2009) also

compared congeneric species across a latitudinal transect and corroborated a shift

towards aragonitic and bimineralic skeletons from the Arctic into lower latitudes. Our

assessment of latitudinal patterns also provide support for the viability of using

latitudinal gradients to infer bryozoan species' responses to seawater temperature

although more tropical species should be analysed mineralogically to confirm this

hypothesis.

Our data may reflect several adaptive advantages of secreting monomineralic calcite

and less soluble $CaCO_3$ mineral phases (e.g. LMC skeletons) in cold waters

bryozoans such as more rapid secretion of large zooids/colonies, more energetically

efficient skeleton secretion and reduction in skeleton dissolution as previously

suggested for Arctic bryozoans (Kuklinski & Taylor, 2009). Consistent with the high

energetic costs to maintain calcified skeletons, especially those with HMC, in cold

waters, a decrease of skeletal investment with latitude and decreasing temperature

has been also noted in several taxa such as gastropods and echinoids (Watson et

al., 2017). Globally, an increase of bryozoans depositing stable carbonate minerals

such as calcite and LMC is thus expected as previously suggested by Andersson et

al. (2008).

The energetic cost of calcification due to the increasing solubility of $CaCO_3$ with

decreasing temperature may also lead to compensatory mechanisms in diverse

species, likely improving their resilience to global change. For instance, a recent

study showed mussels produce thinner shells with a higher organic content in the polar region compared to temperate latitudes (Telesca et al., 2019). Some brachiopod and coral species can also have the ability to form thicker shells and denser skeletons, respectively, under acidified conditions (Cross et al., 2019; Teixidó et al., 2020). However, these changes in the calcification may not be sufficient to overcome impacts of both warming and acidification.

At a local scale, it remains unclear what the effect of seawater temperature and other environmental (e.g. salinity, pH and the seawater Mg/Ca ratio) or biotic factors have on the skeleton of bryozoans. For instance, the expected positive trends of increasing aragonite or Mg-calcite with seawater temperature were not found in some polar and temperate bryozoan species although the authors argued that the existing variability in seawater temperature might be too low in these instances to significantly affect skeletal Mg-calcite (Figuerola et al., 2015; Iglikowska et al., 2020; Loxton, Kuklinski, Barnes, et al., 2014; Loxton, Kuklinski, Najorka, et al., 2014). In addition, local intraspecific and/or interspecific variation was previously found in a range of bryozoan species, which were not related with temperature changes (Figuerola et al., 2015, 2019; Loxton, Kuklinski, Barnes, et al., 2014; Loxton, Kuklinski, Najorka, et al., 2014) suggesting other biological and/or local environmental factors remain important. This could be the case of some Antarctic and temperate bryozoan species, which exhibited a decrease in skeletal Mg-calcite at lower salinities (Figuerola et al., 2015; Loxton, Kuklinski, Najorka, et al., 2014). In contrast, Loxton et al. (2014a) did not found any relationship between the skeletal Mg-calcite and salinity in four Antarctic bryozoans. Several studies investigating the effect of temperature and salinity on Mg/Ca ratios in other calcifying groups also

revealed interspecific variation. While a positive correlation between salinity and

skeletal Mg/Ca ratio was found in some foraminifer and echinoderm species –

although Mg/Ca responses to changes in temperature are more strongly (Borremans

et al., 2009; Dissard et al., 2010; Hönisch et al., 2013), no correlation with salinity

was showed in other foraminifer and echinoderm species (Hermans et al., 2010;

Geerken et al., 2018). Further laboratory and field research should thus attempt to

tease apart the influence of seawater temperature and other factors such as salinity

on bryozoan mineralogy.

4.2 Southern Ocean bellwethers for global change impacts on marine

calcifiers

Southern Ocean bryozoans are likely to be particularly vulnerable to global change

impacts as most cyclostomes and cheilostomes release lecithotrophic short-lived

larvae (Jackson, 1986) and may thus not be able to shift their distribution to more

favourable habitats. Our findings suggest that the combined effects of ocean

warming and acidification may be seen first in HMC species (e.g. *Beania erecta*)

inhabiting in Antarctic coastal areas that have rapidly warmed in recent years such

as the Western Antarctic Peninsula.

Bryozoan species secreting IMC and even LMC skeletons may also be at increased

risk to global change in the near-future, as their skeletons could show an increase in

Mg-calcite with warmer seawater temperatures (Figuerola et al., 2019). Bryozoan

species of the genus *Pentapora* showed an increase of Mg-calcite in parts of the

skeleton formed during the summer (Lombardi et al., 2008). Colonies of two common

Mediterranean bryozoans, *Myriapora truncata* (Pallas, 1766) and *Pentapora fascialis*

(Pallas, 1766), also exhibited higher Mg-calcite values when exposed to experimental high temperatures compared to those of control conditions. These differences only approached significance perhaps due to the short time period of the controlled temperature exposure period (Pagès-Escolà et al., 2018). Skeleton dissolution was also observed in the bryozoan *Jellyella tuberculata* (Bosc, 1802), when colonies were exposed to a combination of reduced pH and elevated temperature, likely due to an increase in levels of skeletal Mg-calcite with increased temperature, making skeletons more susceptible to dissolution at low pH (Swezey et al., 2017). However, these findings contrast with those of Duquette et al. (2018b) who found that a chronic exposure to elevated temperature resulted in a significant reduction in the Mg-calcite of the sea urchin *Lytechinus variegatus* (Lamarck, 1816). Clearly, in a big picture, more experimental and *in-situ* studies that combine near-future predicted elevated temperature and reduced pH (Cummings et al., 2019; Stark et al., 2018) are needed to predict the bryozoans that will be "winners" and "losers" a determination that will be particularly important for those species that are key ecosystem engineers, whose loss could lead to a decline of resilience in benthic Antarctic ecosystems.

ACKNOWLEDGMENTS

We would like to acknowledge the voyage leader, Martin Riddle, the crew and the captain of the RV Aurora Australis. The CAML-CEAMARC cruise of RV Aurora Australis (IPY project n°53) were supported by the Australian Antarctic Division, the Japanese Science Foundation, the French polar institute IPEV and the Muséum National d'Histoire Naturelle). We thank Dr Steve J. Roberts for helping with the figures 6 and 7, Jen Najorka for help with mineralogical analysis and Dr Paul D.

Taylor and three anonymous reviewers for their valuable comments and edits. The study was supported by National Science Centre Poland (project contracts 2016/23/B/ST10/01936 and 2021/40/C/ST10/00225). BF has received funding from the postdoctoral fellowships programme *Beatriu de Pinós* funded by the Secretary of Universities and Research (Government of Catalonia) and by the Horizon 2020 programme of research and innovation of the European Union under the Marie Skłodowska-Curie grant agreement No 801370 (Incorporation grant 2019 BP 00183). This work has been developed in the framework of MedCalRes project Grant PID2021-125323OA-I00 funded by MCIN/AEI/ 10.13039/501100011033 and by "ERDF A way of making Europe". With the institutional support of the 'Severo Ochoa Centre of Excellence' accreditation (CEX2019-000928-S).

References

Amante, C., & Eakins, B. (2009). ETOPO1 arc-minute global relief model: Procedures, data sources and analysis. Boulder, Colo, U.S. Dept. of Commerce, National Oceanic and Atmospheric Administration, National Environmental Satellite, Data, and Information Service, National Geophysical Data Center, Marine Geology and Geophysics Division. https://purl.fdlp.gov/GPO/gpo441.

Amsler, C., McClintock, J., Baker, B., & Zamzow, J. (2010). Habitat choice and predator avoidance by Antarctic amphipods: The roles of algal chemistry and morphology. Marine Ecology Progress Series, 400, 155–163. https://doi.org/10.3354/meps08399

Andersson, A. J., Mackenzie, F. T., & Bates, N. R. (2008). Life on the margin: Implications of ocean acidification on Mg-calcite, high latitude and cold-water marine calcifiers. Marine Ecology Progress Series, 373, 265–273. https://doi.org/10.3354/meps07639

Barnes, D. K. A. (2018). Blue carbon on polar and subpolar seabeds. In Carbon Capture, Utilization and Sequestration. IntechOpen. https://doi.org/10.5772/intechopen.78237

Barnes, D. K. A., & Kuklinski, P. (2010). Bryozoans of the Weddell Sea continental shelf, slope and abyss: Did marine life colonize the Antarctic shelf from deep water, outlying islands or in situ refugia following glaciations? Journal of Biogeography, 37(9), 1648–1656. https://doi.org/10.1111/j.1365-2699.2010.02320.x

Barnes, D. K. A., & Peck, L. S. (1996). Epibiota and attachment substrata of deep-water brachiopods from

Antarctica and New Zealand. Philosophical Transactions of the Royal Society of London. Series B:

Biological Sciences, 351(1340), 677–687. https://doi.org/10.1098/rstb.1996.0064

Bax, N. N., & Cairns, S. D. (2014). Stylasteridae (Cnidaria; Hydrozoa). In De Broyer C et al. Biogeographic Atlas of

the Southern Ocean. Cambridge: SCAR 107-112 (DeBroyer C). SCAR.

http://repository.si.edu/xmlui/handle/10088/22593

Bock, P. E., & Gordon, D. P. (2013). Phylum Bryozoa Ehrenberg, 1831. Zootaxa, 3703(1), 67–74.

https://doi.org/10.11646/zootaxa.3703.1.14

Bone, Y., & James, N. P. (1993). Bryozoans as carbonate sediment producers on the cool-water Lacepede Shelf,

southern Australia. Sedimentary Geology, 86(3–4), 247–271. https://doi.org/10.1016/0037-

0738(93)90025-Z

Borisenko, Y., & Gontar, V. I. (1991). Biogeochemistry of skeletons of coldwater Bryozoa. Biol Morya, 1, 80–90.

Borremans, C., Hermans, J., Baillon, S., André, L., & Dubois, P. (2009). Salinity effects on the Mg/Ca and Sr/Ca in

starfish skeletons and the echinoderm relevance for paleoenvironmental reconstructions. Geology, 37,

351–354. https://doi.org/10.1130/G25411A.1

Chang, V. T.-C., Williams, R. J. P., Makishima, A., Belshawl, N. S., & O'Nions, R. K. (2004). Mg and Ca isotope

fractionation during $CaCO_3$ biomineralisation. Biochemical and Biophysical Research Communications,

323(1), 79–85. https://doi.org/10.1016/j.bbrc.2004.08.053

Chave, K. E. (1954). Aspects of the biogeochemistry of magnesium 1. Calcareous marine organisms. The Journal

of Geology, 62(3), 266–283. https://doi.org/10.1086/626162

Clarke, A., Griffiths, H. J., Barnes, D. K. A., Meredith, M. P., & Grant, S. M. (2009). Spatial variation in seabed

temperatures in the Southern Ocean: Implications for benthic ecology and biogeography. Journal of

Geophysical Research: Biogeosciences, 114(G3). https://doi.org/10.1029/2008JG000886

Cohen, A. L., & Branch, G. M. (1992). Environmentally controlled variation in the structure and mineralogy of

Patella granularis shells from the coast of southern Africa: Implications for palaeotemperature

assessments. Palaeogeography, Palaeoclimatology, Palaeoecology, 91(1), 49–57.

https://doi.org/10.1016/0031-0182(92)90031-Y

Conrad, C. J., & Lovenduski, N. S. (2015). Climate-Driven variability in the Southern Ocean carbonate system.

Journal of Climate, 28(13), 5335–5350. https://doi.org/10.1175/JCLI-D-14-00481.1

Cross, E. L., Harper, E. M., & Peck, L. S. (2019). Thicker shells compensate extensive dissolution in brachiopods

under future ocean acidification. Environmental Science & Technology, 53(9), 5016–5026.

https://doi.org/10.1021/acs.est.9b00714

Cummings, V. J., Barr, N. G., Budd, R. G., Marriott, P. M., Safi, K. A., & Lohrer, A. M. (2019). In situ response of

Antarctic under-ice primary producers to experimentally altered pH. Scientific Reports, 9(1), Article 1.

https://doi.org/10.1038/s41598-019-42329-0

Davis, K. J., Dove, P. M., & De Yoreo, J. J. (2000). The role of Mg^{2+} as an impurity in calcite growth. Science, 290,

1134–1137. https://doi.org/10.1126/science.290.5494.1134

De Broyer, C. D., Danis, B., & Editors, S. T. (2011). How many species in the Southern Ocean? Towards a

dynamic inventory of the Antarctic marine species. Deep-Sea Research Part II, 58(1–2), 5–17.

https://doi.org/10.1016/j.dsr2.2010.10.007

De Broyer, D., Clarke, A., Koubbi, P., Pakhomov, E., Scott, F., Vanden Berghe, E., & Danis, B. (2022). Register of

Antarctic Marine Species. Accessed at https://www.marinespecies.org/rams on 2022-01-26.

Dissard, D., Nehrke, G., Reichart, G. J., & Bijma, J. (2010). The impact of salinity on the Mg/Ca and Sr/Ca ratio in

the benthic foraminifera Ammonia tepida: Results from culture experiments. Geochimica et

Cosmochimica Acta, 74(3), 928–940. https://doi.org/10.1016/j.gca.2009.10.040

Duquette, A., Halanych, K. M., Angus, R. A., & McClintock, J. B. (2018). Inter and intraspecific comparisons of the

skeletal Mg/Ca ratios of high latitude Antarctic echinoderms. Antarctic Science, 30(3), 160–169.

https://doi.org/10.1017/S0954102017000566

Duquette, A., Vohra, Y. K., McClintock, J. B., & Angus, R. A. (2018). Near-future temperature reduces Mg/Ca

ratios in the major skeletal components of the common subtropical sea urchin Lytechinus variegatus.

Journal of Experimental Marine Biology and Ecology, 509, 1–7.

https://doi.org/10.1016/j.jembe.2018.08.007

Fabry, V. J. (2008). Marine calcifiers in a high-CO_2 ocean. Science, 320, 1020–1022.

https://doi.org/10.1126/science.1157130

Fabry, V., McClintock, J., Mathis, J., & Grebmeier, J. (2009). Ocean acidification at high latitudes: The bellwether.

Oceanography, 22(4), 160–171. https://doi.org/10.5670/oceanog.2009.105

Figuerola, B., Ballesteros, M., Monleón-Getino, T., & Avila, C. (2012). Spatial patterns and diversity of bryozoan

communities from the Southern Ocean: South Shetland Islands, Bouvet Island and Eastern Weddell

Sea. Systematics and Biodiversity, 10(1), 109–123. https://doi.org/10.1080/14772000.2012.668972

Figuerola, B., Barnes, D. K. A., Brickle, P., & Brewin, P. E. (2017). Bryozoan diversity around the Falkland and

South Georgia Islands: Overcoming Antarctic barriers. Marine Environmental Research, 126, 81–94

https://doi.org/10.1016/j.marenvres.2017.02.005

Figuerola, B., Gordon, D. P., & Cristobo, J. (2018). New deep Cheilostomata (Bryozoa) species from the

Southwestern Atlantic: Shedding light in the dark. Zootaxa, 4375(2), 211–249.

https://doi.org/10.11646/zootaxa.4375.2.3

Figuerola, B., Gore, D. B., Johnstone, G., & Stark, J. S. (2019). Spatio-temporal variation of skeletal Mg-calcite in

Antarctic marine calcifiers. PLoS ONE, 14(5). https://doi.org/10.1371/journal.pone.0210231

Figuerola, B., Hancock, A. M., Bax, N., Cummings, V. J., Downey, R., Griffiths, H. J., Smith, J., & Stark, J. S.

(2021). A review and meta-analysis of potential impacts of ocean acidification on marine calcifiers from

the Southern Ocean. Frontiers in Marine Science, 8, 584445. https://doi.org/10.3389/fmars.2021.584445

Figuerola, B., Kuklinski, P., & Taylor, P. D. (2015). Depth patterns in Antarctic bryozoan skeletal Mg-calcite: Can

they provide an analogue for future environmental changes? Marine Ecology Progress Series, 540, 109–

120. https://doi.org/10.3354/meps11515

GBIF. (n.d.). GBIF.org (28 January 2022) GBIF Occurrence Download https://doi.org/10.15468/dl.pkvtkg.

Geerken, E., de Nooijer, L. J., van Dijk, I., & Reichart, G.-J. (2018). Impact of salinity on element incorporation in

two benthic foraminiferal species with contrasting magnesium contents. Biogeosciences, 15(7), 2205–

2218. https://doi.org/10.5194/bg-15-2205-2018

Hancock, A. M., King, C. K., Stark, J. S., McMinn, A., & Davidson, A. T. (2020). Effects of ocean acidification on

Antarctic marine organisms: A meta-analysis. Ecology and Evolution, 00, 1–20.

https://doi.org/10.1002/ece3.6205

Hauri, C., Friedrich, T., & Timmermann, A. (2016). Abrupt onset and prolongation of aragonite undersaturation

events in the Southern Ocean. Nature Climate Change, 6(2), 172–176.

https://doi.org/10.1038/nclimate2844

Hayward, P. (1995). Antarctic Cheilostomatous Bryozoa. Oxford University Press, Oxford.

Hermans, J., Borremans, C., Willenz, P., André, L., & Dubois, P. (2010). Temperature, salinity and growth rate

dependences of Mg/Ca and Sr/Ca ratios of the skeleton of the sea urchin *Paracentrotus lividus*

(Lamarck): An experimental approach. Marine Biology, 157(6), 1293–1300.

https://doi.org/10.1007/s00227-010-1409-5

Hönisch, B., Allen, K. A., Lea, D. W., Spero, H. J., Eggins, S. M., Arbuszewski, J., deMenocal, P., Rosenthal, Y.,

Russell, A. D., & Elderfield, H. (2013). The influence of salinity on Mg/Ca in planktic foraminifers –

Evidence from cultures, core-top sediments and complementary δ^{18}O. Geochimica et Cosmochimica

Acta, 121, 196–213. https://doi.org/10.1016/j.gca.2013.07.028

Iglikowska, A., Krzemińska, M., Renaud, P. E., Berge, J., Hop, H., & Kukliński, P. (2020). Summer and winter

$MgCO_3$ levels in the skeletons of Arctic bryozoans. Marine Environmental Research, 162, 105166.

https://doi.org/10.1016/j.marenvres.2020.105166

IPCC. (2014). Synthesis report in climate change 2014: Contribution of working groups I, II and III to the fifth

assessment report of the Intergovernmental Panel on Climate Change.

IPCC. (2019). Summary for Policymakers. In: Special report on the ocean and cryosphere in a changing climate

[H.-O. Pörtner, D.C. Roberts, V. Masson-Delmotte, P. Zhai, M. Tignor, E. Poloczanska, K. Mintenbeck, A.

Alegría, M. Nicolai, A. Okem, J. Petzold, B. Rama, N.M. Weyer (eds.)].

Jackson, J. B. C. (1986). Modes of dispersal of clonal benthic invertebrates: Consequences for species'

distributions and genetic structure of local populations. Bulletin of Marine Science, 39(2), 588–606.

Kołbuk, D., Dubois, P., Stolarski, J.. & Gorzelaka, P. (2019). Effects of seawater chemistry (Mg^{2+}/Ca^{2+} ratio) and

diet on the skeletal Mg/Ca ratio in the common sea urchin Paracentrotus lividus. Marine Environmental

Research. 145, 22–26.

Kołbuk, D., Dubois, P., Stolarski, J., & Gorzelaka, P. (2021). Impact of seawater Mg^{2+}/Ca^{2+} on Mg/Ca of

asterozoan skeleton – Evidence from culturing and the fossil record. Chemical Geology, 584, 120557.

Kroeker, K. J., Kordas, R. L., Crim, R., Hendriks, I. E., Ramajo, L., Singh, G. S., Duarte, C. M., & Gattuso, J.-P.

(2013). Impacts of ocean acidification on marine organisms: Quantifying sensitivities and interaction with

warming. Global Change Biology, 19(6), 1884–1896. https://doi.org/10.1111/gcb.12179

Krzeminska, M., Kuklinski, P., Najorka, J., & Iglikowska, A. (2016). Skeletal mineralogy patterns of Antarctic

bryozoa. The Journal of Geology, 124, 411–422. https://doi.org/10.1086/685507

Krzemińska, M., Piwoni-Piórewicz, A., Shunatova, N., Duczmal-Czernikiewicz, A., Muszyński, A., Kubiak, M., &

Kukliński, P. (2022). Bryozoan carbonate skeletal geochemical composition in the White Sea compared

with neighbouring seas. Marine Environmental Research, 173, 105542.

https://doi.org/10.1016/j.marenvres.2021.105542

Kuklinski, P., & Taylor, P. D. (2008). Are bryozoans adapted for living in the Arctic? In: Hageman SJ, Key MM,

Winston JE (eds) Bryozoan Studies 2007, Proceedings of the 14th International Bryozoology Association

Conference, Boone, North Carolina, 1–8 July 2007. Virginia Museum of Natural History, Memoir, Special

Publication Number, 15, 101–110.

Kuklinski, P., & Taylor, P. D. (2009). Mineralogy of Arctic bryozoan skeletons in a global context. Facies, 55, 489–

500. https://doi.org/10.1007/s10347-009-0179-3

Lebrato, M., Andersson, A. J., Ries, J. B., Aronson, R. B., Lamare, M. D., Koeve, W., Oschlies, A.,

Iglesias‐Rodriguez, M. D., Thatje, S., Amsler, M., Vos, S. C., Jones, D. O. B., Ruhl, H. A., Gates, A. R., &

McClintock, J. B. (2016). Benthic marine calcifiers coexist with $CaCO_3$-undersaturated seawater

worldwide. Global Biogeochemical Cycles, 30(7), 1038–1053. https://doi.org/10.1002/2015GB005260

Lebrato, M., Garbe-Schönberg, D., Müller, M. N., Blanco-Ameijeiras, S., Feely, R. A., Lorenzoni, L., Molinero, J.-C.,

Bremer, K., Jones, D. O. B., Iglesias-Rodriguez, D., Greeley, D., Lamare, M. D., Paulmier, A., Graco, M.,

Cartes, J., Barcelos e Ramos, J., de Lara, A., Sanchez-Leal, R., Jimenez, P., … Oschlies, A. (2020).

Global variability in seawater Mg:Ca and Sr:Ca ratios in the modern ocean. Proceedings of the National

Academy of Sciences, 117(36), 22281–22292. https://doi.org/10.1073/pnas.1918943117

Lebrato, M., McClintock, J. B., Amsler, M. O., Ries, J. B., Egilsdottir, H., Lamare, M., Amsler, C. D., Challener, R.

C., Schram, J. B., Mah, C. L., Cuce, J., & Baker, B. J. (2013). From the Arctic to the Antarctic: The major,

minor, and trace elemental composition of echinoderm skeletons. Ecology, 94(6), 1434–1434.

https://doi.org/10.1890/12-1950.1

Lombardi, C., Cocito, S., Hiscock, K., Occhipinti-Ambrogi, A., Setti, M., & Taylor, P. D. (2008). Influence of

seawater temperature on growth bands, mineralogy and carbonate production in a bioconstructional

bryozoan. Facies, 54(3), 333–342. https://doi.org/10.1007/s10347-008-0143-7

Lowenstam, H. A. (1954). Environmental relations of modification compositions of certain carbonate secreting

marine invertebrates. Proceedings of the National Academy of Sciences, 40(1), 39–48.

https://doi.org/10.1073/pnas.40.1.39

Lowenstam, H. A., & Weiner, S. (1989). On biomineralization. Oxford University Press.

https://doi.org/10.1093/oso/9780195049770.001.0001

Loxton, J., Jones, M. S., Najorka, J., Smith, A. M., & Porter, J. S. (2018). Skeletal carbonate mineralogy of Scottish

bryozoans. PLOS ONE, 13(6), e0197533. https://doi.org/10.1371/journal.pone.0197533

Loxton, J., Kuklinski, P., Barnes, D. K. A., Najorka, J., Jones, M. S., & Porter, J. S. (2014). Variability of Mg-calcite

in Antarctic bryozoan skeletons across spatial scales. Marine Ecology Progress Series, 507, 169–180.

https://doi.org/10.3354/meps10826

Loxton, J., Kuklinski, P., Mair, J. M., Jones, M. S., & Porter, J. S. (2013). Patterns of magnesium-calcite distribution

in the skeleton of some polar bryozoan species mineralogy of polar bryozoan skeletons. In: Bryozoan

Studies 2010, 143, 169–185. https://doi.org/10.1007/978-3-642-16411-8

Loxton, J., Kuklinski, P., Najorka, J., Jones, M. S., & Porter, J. S. (2014). Variability in the skeletal mineralogy of

temperate bryozoans: The relative influence of environmental and biological factors. 510, 45–57.

https://doi.org/10.3354/meps10889

Mackenzie, F. T., Bischoff, W. D., Bishop, F. C., Loijens, M., Schoonmaker, J., & Wollast, R. (1983). Magnesian

calcites: Low-temperature occurrence, solubility and solid-solution behavior. In R. Reeder (Ed.),

Carbonates: Mineralogy and Chemistry. Reviews in Mineralogy (pp. 97–143). Mineralogical Society of

America, Washington, DC.

McClintock, J. B., Amsler, M. O., Angus, R. A., Challener, R. C., Schram, J. B., Amsler, C. D., Mah, C. L., Cuce, J.,

Baker, B. J., The, S., & September, N. (2011). The Mg-Calcite composition of Antarctic echinoderms:

Important implications for predicting the impacts of ocean acidification. The Journal of Geology, 119(5),

457–466. https://doi.org/10.1086/660890

McClintock, J. B., Amsler, R., Angus, R., Mcclintock, F., Geneviève, A.-M., & Lebrato, M. (2017). A comparison of

$MgCO_3$ in individual sub-components of major skeletal structures of the common temperate sea urchin

Paracentrotus lividus. Cahiers Marine Biologie, 58, 99–106. https://doi.org/10.21411/CBM.A.B27402D4

McNeil, B. I., & Matear, R. J. (2008). Southern Ocean acidification: A tipping point at 450-ppm atmospheric CO_2.

Proceedings of the National Academy of Sciences of the United States of America, 105(48), 18860–

18864.

Mitric, M., Kremenovic, A., Dimitrijevic, R., & Rodic, D. (1997). Structure and microstructure of $Sm_xY_{2-x}O_3$. Solid

State Ionics, 101–103, 495–501. https://doi.org/10.1016/S0167-2738(97)84074-X

Moore, J. K., Abbott, M. R., & Richman, J. G. (1999). Location and dynamics of the Antarctic Polar Front from

satellite sea surface temperature data. Journal of Geophysical Research: Oceans, 104(C2), 3059–3073.

https://doi.org/10.1029/1998JC900032

Mucci, A. (1983). The solubility of calcite and aragonite in seawater at various salinities, temperatures, and one

atmosphere total pressure. American Journal of Science, 283(7), 780–799.

https://doi.org/10.2475/ajs.283.7.780

Orr, J. C., Fabry, V. J., Aumont, O., Bopp, L., Doney, S. C., Feely, R. a, Gnanadesikan, A., Gruber, N., Ishida, A.,

Joos, F., Key, R. M., Lindsay, K., Maier-Reimer, E., Matear, R., Monfray, P., Mouchet, A., Najjar, R. G.,

Plattner, G.-K., Rodgers, K. B., … Yool, A. (2005). Anthropogenic ocean acidification over the twenty-first

century and its impact on calcifying organisms. Nature, 437(7059), 681–686.

https://doi.org/10.1038/nature04095

Orr, R. J. S., Martino, E. D., Ramsfjell, M. H., Gordon, D. P., Berning, B., Chowdhury, I., Craig, S., Cumming, R. L.,

Figuerola, B., Florence, W., Harmelin, J.-G., Hirose, M., Huang, D., Jain, S. S., Jenkins, H. L., Kotenko, O.

N., Kuklinski, P., Lee, H. E., Madurell, T., … Liow, L. H. (2022). Paleozoic origins of cheilostome

bryozoans and their parental care inferred by a new genome-skimmed phylogeny. Science Advances, 8,

eabm7452. https://doi.org/10.1126/sciadv.abm7452

Pagès-Escolà, M., Hereu, B., Garrabou, J., Montero-Serra, I., Gori, A., Gómez-Gras, D., Figuerola, B., & Linares,

C. (2018). Divergent responses to warming of two common co-occurring Mediterranean bryozoans.

Scientific Reports, 8(1), 17455. https://doi.org/10.1038/s41598-018-36094-9

Peck, L. (2018). Antarctic marine biodiversity: Adaptations, environments and responses to change. In S. J.

Hawkins, A. J. Evans, A. C. Dale, L. B. Firth, & I. P. Smith (Eds.), Oceanography and Marine Biology: An

annual review (No. 56; Vol. 56, Issue 56, pp. 105–236). Taylor and Francis.

Peck, L. S. (2001). Ecophysiology of Antarctic marine ectotherms: Limits to life. Polar Biology, 25, 31–40.

Peck, L. S. (2005). Prospects for survival in the Southern Ocean: Vulnerability of benthic species to temperature

change. Antarctic Science, 17(4), 497–507. https://doi.org/10.1017/S0954102005002920

Piwoni-Piórewicz, A., Krzemińska, M., Iglikowska, A., Jens, N., & Kukliński, P. (2019). Skeletal mineralogy patterns

of Bryozoa from the Aleutian Islands in the context of revealing a global pattern in bryozoan skeletal

mineralogy. Bryozoan Studies, 17.

Poluzzi, A., & Sartori, R. (1975). Report on the carbonate mineralogy of Bryozoa. Documents des Laboratoires de

Geologie de la Faculte des Sciences de Lyon. Hors Serie, 3, 193–210.

Ries, J. B. (2010). Review: Geological and experimental evidence for secular variation in seawater Mg / Ca (

calcite-aragonite seas) and its effects on marine biological calcification. 2795–2849.

https://doi.org/10.5194/bg-7-2795-2010

Rucker, J. B., & Carver, R. E. (1969). A survey of the carbonate mineralogy of cheilostome Bryozoa. Journal of

Paleontology, 43(3), 791–799. JSTOR.

Santagata, S., Ade, V., Mahon, A. R., Wisocki, P. A., & Halanych, K. M. (2018). Compositional differences in the

habitat-forming bryozoan communities of the Antarctic shelf. Frontiers in Ecology and Evolution, 6.

https://www.frontiersin.org/article/10.3389/fevo.2018.00116

Schloerke, B., Briatte, F., Joseph, bigbeardesktop, elbamos, Crowley, J., Cook, D., Ibanez, E., Sievert, C., Ogden,

K., Ross, Wallace, E., Gilligan, J., Spiller, T., Thoen, E., Richter, J., Toomet, O., Beck, M. W.,

Sergushichev, A., … ewenharrison. (2021). ggobi/ggally: V2.1.2. Zenodo.

https://doi.org/10.5281/zenodo.5009047

Smith, A. M., & Clark, D. E. (2010). Skeletal carbonate mineralogy of bryozoans from Chile: An independent check

of phylogenetic patterns. PALAIOS, 25(4), 229–233. https://doi.org/10.2110/palo.2009.p09-106r

Smith, A. M., Clark, D. E., Lamare, M. D., Winter, D. J., & Byrne, M. (2016). Risk and resilience: Variations in

magnesium in echinoid skeletal calcite. Marine Ecology Progress Series, 561, 1–16.

https://doi.org/10.3354/meps11908

Smith, A. M., & Girvan, E. (2010). Understanding a bimineralic bryozoan: Skeletal structure and carbonate

mineralogy of Odontionella cyclops (Foveolariidae: Cheilostomata: Bryozoa) in New Zealand.

Palaeogeography, Palaeoclimatology, Palaeoecology, 289(1–4), 113–122. Scopus.

https://doi.org/10.1016/j.palaeo.2010.02.022

Smith, A. M., Key, M. M., & Gordon, D. P. (2006). Skeletal mineralogy of bryozoans: Taxonomic and temporal

patterns. Earth-Science Reviews, 78, 287–306. https://doi.org/10.1016/j.earscirev.2006.06.001

Smith, A. M., Nelson, C. S., & Spencer, H. G. (1998). Skeletal carbonate mineralogy of New Zealand bryozoans.

Marine Geology, 151, 27–46.

Stark, J. S., Roden, N. P., Johnstone, G. J., Milnes, M., Black, J. G., Whiteside, S., Kirkwood, W., Newbery, K.,

Stark, S., Van Ooijen, E., Tilbrook, B., Peltzer, E. T., Berry, K., & Roberts, D. (2018). Carbonate chemistry

of an in-situ free-ocean CO_2 enrichment experiment (antFOCE) in comparison to short term variation in

Antarctic coastal waters. Scientific Reports, 8(1), 1–16. https://doi.org/10.1038/s41598-018-21029-1

Steger, K. K., & Smith, A. M. (2005). Carbonate mineralogy of free-living bryozoans (Bryozoa: Otionellidae), Otago

shelf, southern New Zealand. Palaeogeography, Palaeoclimatology, Palaeoecology, 218(3–4), 195–203.

https://doi.org/10.1016/j.palaeo.2004.12.013

Steig, E. J., Schneider, D. P., Rutherford, S. D., Mann, M. E., Comiso, J. C., & Shindell, D. T. (2009). Warming of

the Antarctic ice-sheet surface since the 1957 International Geophysical Year. Nature, 457(7228), Article

7228. https://doi.org/10.1038/nature07669

Swezey, D. S., Bean, J. R., Ninokawa, A. T., Hill, T. M., Gaylord, B., & Sanford, E. (2017). Interactive effects of

temperature, food and skeletal mineralogy mediate biological responses to ocean acidification in a widely

distributed bryozoan. Proceedings of the Royal Society B: Biological Sciences, 284, 20162349.

https://doi.org/10.1098/rspb.2016.2349

Taylor, P. D. (2020). Bryozoan Paleobiology | Wiley. https://www.wiley.com/en-us/Bryozoan+Paleobiology-p-

9781118455005

Taylor, P. D., & Di Martino, E. (2014). Why is the tropical Cenozoic fossil record so poor for bryozoans? Studi

Trentini Di Scienze Naturali, 94, 249–257.

Taylor, P. D., James, N. P., Bone, Y., Kuklinski, P., & Kyser, T. K. (2009). Evolving mineralogy of cheilostome

bryozoans. Palaios, 24, 440–452. https://doi.org/10.2110/palo.2008.p08

Taylor, P. D., Lombardi, C., & Cocito, S. (2015). Biomineralization in bryozoans: Present, past and future.

Biological Reviews, 90, 1118–1150. https://doi.org/10.1111/brv.12148

Taylor, P. D., Tan Shau-Hwai, A., Kudryavstev, A. B., & Schopf, J. W. (2016). Carbonate mineralogy of a tropical

bryozoan biota and its vulnerability to ocean acidification. Marine Biology Research, 12(7), 776–780.

https://doi.org/10.1080/17451000.2016.1203951

Teixidó, N., Caroselli, E., Alliouane, S., Ceccarelli, C., Comeau, S., Gattuso, J., Fici, P., Micheli, F., Mirasole, A.,

Monismith, S. G., Munari, M., Palumbi, S. R., Sheets, E., Urbini, L., De Vittor, C., Goffredo, S., & Gambi,

M. C. (2020). Ocean acidification causes variable trait-shifts in a coral species. Global Change Biology,

26(12), 6813–6830. https://doi.org/10.1111/gcb.15372

Telesca, L., Peck, L. S., Sanders, T., Thyrring, J., Sejr, M. K., & Harper, E. M. (2019). Biomineralization plasticity

and environmental heterogeneity predict geographical resilience patterns of foundation species to future

change. Global Change Biology, 25(12), 4179–4193. https://doi.org/10.1111/gcb.14758

Turley, C. M., Roberts, J. M., & Guinotte, J. M. (2007). Corals in deep-water: Will the unseen hand of ocean

acidification destroy cold-water ecosystems? Coral Reefs, 26(3), 445–448.

https://doi.org/10.1007/s00338-007-0247-5

Watson, S.-A., Morley, S. A., & Peck, L. S. (2017). Latitudinal trends in shell production cost from the tropics to the

poles. Science Advances, 3(9), e1701362. https://doi.org/10.1126/sciadv.1701362

Weiner, S., & Dove, P. M. (2003). An overview of biomineralization processes and the problem of the vital effect.

Reviews in Mineralogy and Geochemistry, 54(1), 1–29. https://doi.org/10.2113/0540001

Wickham, H., Averick, M., Bryan, J., Chang, W., McGowan, L., François, R., Grolemund, G., Hayes, A., Henry, L.,

Hester, J., Kuhn, M., Pedersen, T., Miller, E., Bache, S., Müller, K., Ooms, J., Robinson, D., Seidel, D.,

Spinu, V., … Yutani, H. (2019). Welcome to the Tidyverse. Journal of Open Source Software, 4(43),

1686. https://doi.org/10.21105/joss.01686

Figures

Fig. 1. Map of the study area (Terre Adélie and George V Land) in East Antarctica
and the sampling stations.

789
790
791

Fig. 2. SEM images of 6 selected bryozoan species studied from East Antarctica. (A)
Systenopora contracta, zooids on branch surface (scale bar = 200 µm); (B)
Fasciculipora ramosa, colony (scale bar = 1 mm); (C) *Lageneschara lyrulata*, group
of zooids (scale bar = 200 µm); (D) *Melicerita obliqua*, zooids from near centre of
branch (scale bar = 100 µm); (E) *Chondriovelum adeliense*, group of zooids (scale
bar = 200 µm); (F) *Astochoporella cassidula*, group of zooids (scale bar = 200 µm).

798
799

Fig. 4. Distribution patterns of the mineral type and of the skeletal Mg-calcite of
bryozoans in the Southern Hemisphere. (A) Maps of the Southern Ocean and
neighbouring regions showing the Polar Front (PF; black line) and the distribution
patterns of Mg-calcite in bryozoan skeletons; Frequency distributions for mineral type
(B) and (C) skeletal Mg-calcite in 1415 specimens belonging to 102 species from the
Southern Ocean; Relationships between latitude (D) and (E) mineralogical
composition in the Southern Hemisphere. LMC: low Mg-calcite; IMC: intermediate
Mg-calcite; HMC: high Mg-calcite. South of PF (squares): Calcite = R^2 = 0.0166, p =
0.81, y = 98 - 0.009X; Bimineralic = R^2 = 0.0334, p = 0.73, y = 2.2 + 0.013X;
Aragonite = R^2 = 0.1543, p = 0.44, y = -0.24 - 0.0041X. North of PF (circles): Calcite
= R^2 = 0.0743, p = 0.37, y = 81 + 0.33X; Bimineralic = R^2 = 0.0957, p = 0.3, y = 13 -
0.38X; Aragonite = R^2 = 0.0831, p = 0.34, y = 6.1 + 0.051X. South of PF (squares):
LMC = R^2 = 0.0387, p = 0.71, y = 37 + 0.045X; IMC = R^2 = 0.0698, p = 0.62, y = 58 -
0.081X; HMC = R^2 = 0.0634, p = 0.63, y = 5.2 + 0.036X. North of PF (circles): LMC =
R^2 = 0.0004, p = 0.95, y = 21 - 0.012X; IMC = R^2 = 0.2571, p = 0.077, y = 50 - 0.41X;
HMC = R^2 = 0.5802, p = 0.0025, y = 29 + 0.42X.
823

Fig. 5. Global distribution patterns of the mineral type of bryozoans. (A) Global
frequency distributions for mineral type; (B) Global distribution patterns of calcite
(blue), (C) bimineralic (yellow) and (D) aragonite (red); E) Relationships between
seawater temperature and mineral type in the Southern Hemisphere. Shading bands
represent standard errors on linear regressions. Calcite = R^2 = 0.1882, p = 0.01, y =
85 - 0.49X; Bimineralic = R^2 = 0.1648, p = 0.017, y = 13 + 0.4X; Aragonite = R^2 =
0.079, p = 0.11, y = 2.3 + 0.092X.

833
834

Fig. 6. Global distribution patterns of the skeletal Mg-calcite of bryozoans. (A) Global
frequency distributions for skeletal Mg-calcite; (B) Global distribution patterns of low
Mg-calcite (LMC; blue), (C) intermediate Mg-calcite (IMC; yellow) and (D) high Mg-
calcite (HMC; red); (E) Relationships between seawater temperature and
mineralogical composition in the Southern Hemisphere. Shading bands represent
standard errors on linear regressions. LMC = R² = 0.0067, p = 0.62, y = 30 + 0.094X;
IMC = R² = 0.4488, p = 1.1e-05, y = 67 - 0.9X; HMC = R² = 0.4828, p = 3.6e-06, y =
3.5 + 0.8X.

844