

# 1 **Determinants of the microbiome spatial variability in** 2 **chronic rhinosinusitis**

3 **Running title:** Microbiome spatial variability in CRS

4

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## 28 Abstract

29 **Background.** The sinus microbiome in patients with chronic rhinosinusitis (CRS) is  
30 considered homogenous across the sinonasal cavity. The middle nasal meatus is the  
31 recommended sampling site for 16S rRNA sequencing. However, individuals with unusually  
32 high between-site variability between the middle meatus and the sinuses were identified in  
33 previous studies. This study aimed to identify which factors determine increased microbial  
34 heterogeneity between sampling sites in the sinuses.

35 **Methodology.** In this cross-sectional study samples for 16S rRNA sequencing were obtained  
36 from the middle meatus, the maxillary and the frontal sinus in 50 patients with CRS. The  
37 microbiome diversity between sampling sites was analysed in relation to the size of the sinus  
38 ostia and clinical metadata.

39 **Results.** In approximately 15% of study participants, the differences between sampling sites  
40 within one patient were greater than between the patient and other individuals. Contrary to a  
41 popular hypothesis, obstruction of the sinus ostium resulted in decreased dissimilarity  
42 between the sinus and the middle meatus. The dissimilarity between the sampling sites was  
43 patient-specific: greater between-sinus differences were associated with greater meatus-sinus  
44 differences, regardless of the drainage pathway patency. Decreased spatial variability was  
45 observed in patients with nasal polyps and extensive mucosal changes in the sinuses.

46 **Conclusions.** Sampling from the middle meatus is not universally representative of the sinus  
47 microbiome. The differences between sites cannot be predicted from the patency of  
48 communication pathways between them.

49

50

## 51 Introduction

52 The significance of the microbiome for the development of chronic rhinosinusitis (CRS) is  
53 unclear. CRS is a complex inflammatory disease and treatment outcomes are influenced by  
54 multiple factors (1, 2), while bacteria are believed to cause exacerbations and contribute to the

55 recalcitrance of CRS (1, 3, 4). The effectiveness of antimicrobial treatment and reliability of  
56 research depends on representative sampling of the sinonasal microbiome.

57 It is generally assumed that microbiome samples from the middle nasal meatus are  
58 representative of the sinuses (1, 5). In CRS, however, the communication between the sinuses  
59 and the middle meatus is often impaired. Previously, we showed that the results of the middle  
60 meatus culture in patients with CRS were discordant with maxillary sinus culture in 20% of  
61 cases and frontal sinus culture in 34% of cases (6).

62 In most studies on the sinonasal microbiome that were conducted using 16S rRNA  
63 sequencing, the authors observed only small or insignificant differences between the sinuses  
64 and the middle meatus (7-10). Nevertheless, several researchers noted larger dissimilarity  
65 between sampling sites in some patients (7, 11). The causes of increased between-site  
66 differences in certain individuals have not been studied before.

67 This study aimed to identify the factors that determine the heterogeneity of the sinonasal  
68 microbiome. Samples for 16S rRNA sequencing were obtained from the middle meatus, the  
69 maxillary sinus, and the frontal sinus. To evaluate the impact of the anatomical separation of  
70 the subsites, we compared three groups of patients: (a) with narrow sinus ostia, (b) with  
71 blocked sinus ostia, and (c) with wide sinus ostia after previous surgery. Subsequently, we  
72 evaluated the relationship between other clinical metadata and microbiome diversity.  
73 Obstruction of the sinus ostia did not result in increased differences between the sinuses and  
74 the middle meatus. Decreased between-site microbiome variability was associated with  
75 certain clinical characteristics of the patients (nasal polyps, extensive opacification of the  
76 sinuses).

77

## 78 **Materials and methods**

### 79 **Sample collection**

80 In this cross-sectional observational study, samples were collected from patients with CRS  
81 during endoscopic sinus surgery between October 2018 and June 2019 at the University

82 Hospital in Krakow. CRS was defined according to the EPOS 2012 guidelines (12). All of the  
83 participants had not improved after medical treatment and were scheduled for surgery.  
84 Patients who received antibiotics one month before the surgery were excluded. Clinical data  
85 collected for the patients included: age, time since CRS onset, nasal polyps, comorbidities  
86 (asthma, aspirin-exacerbated respiratory disease, gastroesophageal reflux, allergy), recent  
87 steroid use, history of recurrent exacerbations and previous sinus surgeries, radiological  
88 staging (Lund-Mackay score (13)) and self-evaluation of the symptoms on the visual analogue  
89 scale.

90 The swabs were collected under endoscopic guidance from 3 sites (the middle nasal meatus,  
91 the maxillary sinus and the frontal sinus on the same side) in 50 patients which provided a  
92 total of 150 samples. If the sinus was blocked, the swab was collected immediately after the  
93 surgical opening of its ostium. Contact with the nasal vestibule or other sites was strictly  
94 avoided. The samples were transported on ice to the laboratory where they were stored at  
95  $-80^{\circ}\text{C}$ . The study was approved by the Jagiellonian University Bioethics Committee  
96 (1072.6120.78.2018, 20.04.2018).

97

## 98 **DNA isolation, library preparation and sequencing**

99 The swabs were thawed and vigorously shaken in 1 mL of saline. Afterwards, the samples  
100 were treated with lysozyme ( $1\ \mu\text{mg/mL}$ ) and lysostaphin ( $0.1\ \mu\text{mg/mL}$ ) enzymes (Sigma-  
101 Aldrich, Poznań, Poland) at  $37^{\circ}\text{C}$  for 20 min to digest the bacterial cell walls. Further, the  
102 samples were subjected to DNA extraction using a Mini Genomic DNA isolation kit (A&A  
103 Biotechnology, Gdynia, Poland). The concentration and purity of DNA isolates were  
104 determined spectrophotometrically for A260 nm and A260nm / 280nm ratio using NanoDrop  
105 (ThermoFisher, Waltham, MA USA).

106 Libraries were prepared strictly according to Illumina's protocol (San Diego, CA, USA) and  
107 Kowalska-Duplaga *et al.* (14, 15).

108 Primers (Genomed, Warsaw, Poland) specific to the V3 and V4 16S rRNA sequences of  
109 bacteria were used: (F)

110 5'TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGCCTACGGGNGGCWGCAG3'

111 (R)5'

112 GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGGACTACHVGGGTATCTAATCC  
113 3'

114 After purification and concentration measurement, libraries were pooled and sequenced using  
115 the MiSeq sequencer (Illumina, San Diego, CA, USA).

116 In one sample from the middle meatus, amplifying the library was impossible due to too low  
117 DNA concentration - it was excluded from further analysis.

118

## 119 **Data and statistical analysis**

120 The sequencing results were processed using a pipeline available within QIIME 2 (16).  
121 Truncation was performed at 290 bp length for forward reads and at 250 bp length for reverse  
122 reads to avoid the technical quality drop with the increased base position. Paired-end  
123 sequences were denoised using DADA2 via q2-dada2 and merged with minimal overlap of 12  
124 base pairs. The phylogenetic tree was constructed using q2-fragment-insertion with the SEPP  
125 reference database based on SILVA 128. After analysis of rarefaction curves, the sampling  
126 depth of 20,504 reads per sample was chosen. The rarefaction procedure resulted in the  
127 exclusion of additional 14 samples from the analysis (7 from the middle meatus, 5 from the  
128 maxillary sinus, and 2 from the frontal sinus).

129 The measure of microbiome diversity within each sample (alpha diversity) used in this study  
130 was Faith's phylogenetic diversity which incorporates phylogenetic differences between the  
131 taxa identified in the sample.

132 The disparities in microbiome composition between pairs of samples (beta diversity) were  
133 measured using two dissimilarity metrics: Bray-Curtis and weighted UniFrac (wUniFrac).  
134 Bray-Curtis dissimilarity quantifies compositional differences between biological  
135 communities based on counts at each site. The values of the metrics range from 0 to 1, where  
136 0 indicates that the samples are identical and 1 means that the two samples do not share any  
137 species. The wUniFrac distance additionally incorporates information on the phylogenetic  
138 distances between organisms. The lowest wUniFrac value is 0 if the communities do not  
139 differ. Larger values indicate greater differences and the maximal value may exceed 1 (17).

140 Phylogenetic Investigation of Communities by Reconstruction of Unobserved States  
141 (PICRUSt2) was used for the prediction of the functional potential of the microbial  
142 communities. This computational tool allows for the prediction of metagenomic functional  
143 profiles from 16 rRNA sequencing data via hidden state prediction on a constructed  
144 phylogenetic tree. Although prediction accuracy is varied depending on the microbial  
145 environment, PICRUSt2 was shown to be highly efficient within the human organism (18).

146 The normality of distribution was assessed using the Shapiro-Wilk test. The t-Student test and  
147 Pearson correlation were used for biodiversity measures that had a normal distribution.  
148 Wilcoxon signed-rank test, Mann-Whitney-Wilcoxon test, Kruskal-Wallis test and  
149 Spearman's rank correlation were used for biodiversity measures that did not have a normal  
150 distribution.

151 The differences in microbiome composition between subgroups of samples were further  
152 explored using permutational multivariate analysis of variance (PERMANOVA) using the  
153 adonis function.

154 Data is available at the European Nucleotide Archive (ENA):  
155 <https://www.ebi.ac.uk/ena/browser/view/PRJEB55924>

156 The Strengthening The Organization and Reporting of Microbiome Studies (STORMS)  
157 Checklist can be found at <https://zenodo.org/record/7092029#.YzF6CuxBx6o>

158 The code is available at: [https://github.com/bioinf-mcb/spatial\\_sinus\\_microbiome](https://github.com/bioinf-mcb/spatial_sinus_microbiome)

159

160

## 161 **Results**

### 162 **Clinical characteristics of the study group**

163 Fifty patients with CRS were enrolled in the study. Samples were collected during endoscopic  
164 sinus surgery from the middle meatus, maxillary sinus and frontal sinus. The study  
165 participants' demographics and clinical characteristics are shown in Table 1.

166

167

168 Table 1. Demographic and clinical characteristics of the 50 study participants.

169

Age	19-83 (mean 49)
Gender	female: 25, male: 25
Previous endoscopic sinus surgery	24
CRS with nasal polyps	21
Computed tomography – Lund-Mackay score (0-24, greater values indicate greater extension of the sinus opacification)	2-24 (mean 13)
Comorbidities	asthma: 20 aspirin-exacerbated respiratory disease: 9 allergy: 24 gastroesophageal reflux: 15

170

171

## 172 **Variability between sampling sites**

173 In every patient, the Bray-Curtis and weighted UniFrac distances between the following pairs  
174 of samples were calculated:

175 (a) middle meatus-maxillary sinus distance,

176 (b) middle meatus-frontal sinus distance,

177 (c) maxillary sinus-frontal sinus distance.

178 The distribution of the beta-diversity indexes was not normal. Therefore, nonparametric tests  
179 were used in the analysis. The values of the weighted UniFrac and Bray-Curtis dissimilarity  
180 are shown in Table 2. Although the median values of dissimilarity measures were 0.28-0.34  
181 for the weighted UniFrac and 0.22-0.26 for the Bray-Curtis metrics, in some patients we  
182 observed much higher values of the indexes. For example, the maximal values of the Bray-  
183 Curtis metric reached 0.75 for the middle meatus-maxillary sinus distance and 0.92 for the  
184 middle meatus-frontal sinus.

185 Table 2. The values of the weighted UniFrac and Bray-Curtis dissimilarity between sampling  
186 sites.

187

	distance	median	min	max	interquartile range
weighted UniFrac	middle meatus-frontal sinus	0.34	0.05	1.05	0.40
	middle meatus-maxillary sinus	0.28	0.07	1.16	0.34
	frontal sinus-maxillary sinus	0.31	0.04	1.63	0.47
Bray Curtis	middle meatus-frontal sinus	0.24	0.09	0.92	0.26



	middle meatus-maxillary sinus	0.22	0.10	0.74	0.22
	frontal sinus-maxillary sinus	0.26	0.07	0.96	0.31

188

189

190 We observed statistically significant positive correlations between all intra-individual  
191 distances within a patient. The greater disparity between the middle meatus and the maxillary  
192 sinus correlated with a greater disparity between the middle meatus and the frontal sinus and  
193 between both sinus cavities in the same individual (Bray-Curtis: Spearman's rho 0.55-0.71,  
194 weighted UniFrac: Spearman's rho 0.58-0.66;  $p < 0.05$ ).

195

## 196 **The effect of the maxillary ostium size on the microbiome** 197 **continuity**

198 To assess whether the large middle meatus-maxillary sinus differences noted in some patients  
199 were caused by the anatomical separation between the middle meatus and the maxillary sinus,  
200 we divided the patients into three groups according to the size of the maxillary ostium. We  
201 identified 23 patients with blocked ostium, 14 patients with narrow ostium and 12 patients  
202 with wide ostium created during previous surgery.

203 We found that obstruction of the drainage pathway did not cause larger differences between  
204 the microbial communities in the maxillary sinus and the middle meatus. On the contrary, we  
205 found that the middle meatus-maxillary sinus distances were smaller in patients with blocked  
206 ostia than in patients with narrow or wide ostia. The differences were not statistically  
207 significant. However, the difference between the Bray-Curtis metrics in the groups with  
208 narrow and blocked ostia was close to the significance cutoff (Mann-Whitney-Wilcoxon test;

209  $p = 0.06$ ) with closer middle meatus-maxillary sinus similarity if the ostium was blocked than  
210 in case of patent narrow ostium (Fig. 1A).

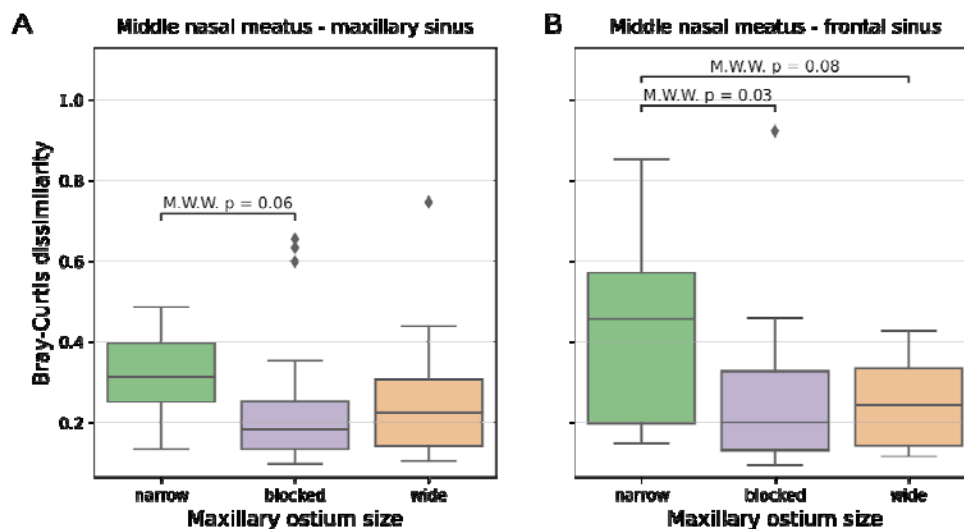
211 A similar but stronger relationship with the maxillary ostium size was noted for the middle  
212 meatus-frontal sinus beta diversity measures (Fig. 1B). The middle meatus-frontal sinus Bray-  
213 Curtis distance in patients with narrow maxillary ostium was significantly greater than in  
214 patients with blocked maxillary ostium (Mann-Whitney-Wilcoxon test;  $p = 0.03$ ) and  
215 insignificantly greater than in patients with wide ostium ( $p = 0.08$ ).

216 Due to the fact, that only 3 patients in the study group had a patent frontal ostium, analogical  
217 computations for frontal ostium did not yield statistically significant results.

218

219 **Fig. 1.** The Bray-Curtis distances between the middle meatus and (A) the maxillary sinus, (B)  
220 the frontal sinus in the three subgroups of patients (with wide, narrow and blocked maxillary  
221 ostium). Larger metrics values mean greater differences between the sampling sites. M.W.W.  
222 - Mann-Whitney-Wilcoxon test.

223



224

225

226

## 227 **Relationships between intra-individual beta diversity and** 228 **clinical metadata**

229 Subsequently, we investigated the relationships between the clinical characteristics of the  
230 patients and intra-patient beta diversity measures. The distance between the middle meatus  
231 and the frontal sinus was significantly smaller in patients with nasal polyps than in patients  
232 without nasal polyps (Mann-Whitney-Wilcoxon test; Bray-Curtis distance:  $p = 0.016$ ,  
233 weighted UniFrac distance:  $p = 0.025$ ). The middle meatus-frontal sinus dissimilarity was also  
234 less pronounced in patients with more extensive sinus opacification in the Lund-Mackay score  
235 (Spearman's  $\rho -0.35$ ;  $p < 0.05$ ). Analogical correlations were not noted for the middle  
236 meatus-maxillary sinus distance. Other clinical metadata did not correlate significantly with  
237 the beta diversity measures between sampling sites.

238

## 239 **Intra-individual versus inter-individual variability**

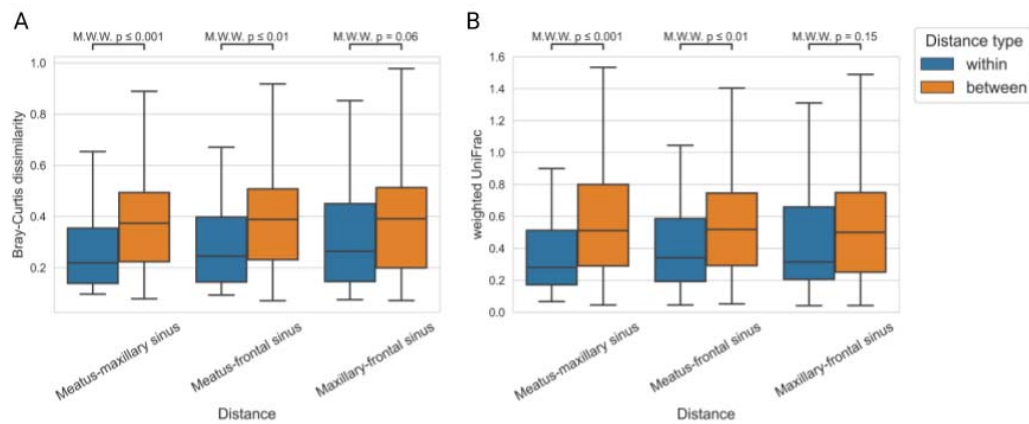
240 Despite the high values of beta diversity noted in some patients, PERMANOVA tests (adonis)  
241 indicated that in the whole study group the variation between the community structure in  
242 different sampling sites was not statistically significant (Bray-Curtis:  $p = 0.596$ , weighted  
243 UniFrac:  $p = 0.281$ ). On the contrary, variation caused by differences between patients was  
244 significant in both metrics (Bray-Curtis:  $p = 0.041$ , weighted UniFrac:  $p = 0.03$ ). This result  
245 indicates that overall the highly pronounced variability between subjects outweighs intra-  
246 individual variation of the sinonasal microbiota.

247 Figure 2 shows that the middle meatus-maxillary sinus and middle meatus-frontal sinus  
248 distances within each patient were significantly smaller than distances between the same sites  
249 among the study participants. However, the difference between intra-individual maxillary  
250 sinus-frontal sinus distances and inter-individual distances was insignificant.

251 **Fig. 2.** Comparisons of beta-diversity distances within and between patients. The “within  
252 patient” distances were calculated between two sites for each study participant (50 distances).  
253 The “between patient” distances were calculated between one site in each patient and the

254 second site in all other patients ( $50 \times 49 = 2450$  distances). A. Bray-Curtis distances. B.  
255 weighted UniFrac distances. M.W.W. - Mann-Whitney-Wilcoxon test.

256



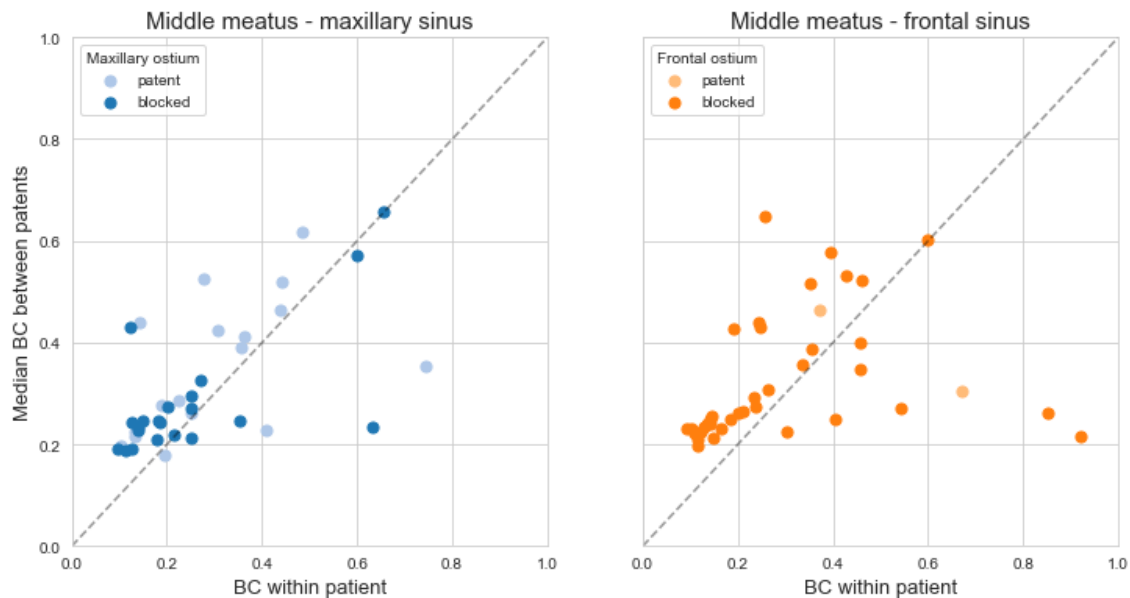
257

258 In several patients, between-patient Bray-Curtis distances were greater than within-patient  
259 distances (Fig. 3). The middle meatus-maxillary sinus differences within a patient were  
260 greater than distances between the patient's middle meatus and the maxillary sinuses of other  
261 participants in 7 individuals. The same relationship was observed in 8 patients for middle  
262 meatus-frontal sinus distances. A trend towards increased intra-individual variability was  
263 observed in patients who also presented higher beta diversity distances from other study  
264 participants.

265

266 **Fig. 3.** Bray-Curtis beta-diversity distances within and between patients presented for each  
267 patient separately. The “within patient” distances were calculated between two sites for each  
268 study participant. The “between patient” distances were calculated between one site in each  
269 patient and the second site in all other patients. The points below the dotted line indicate  
270 patients who presented higher variability between sites than between the patient and other  
271 study participants.

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## 276 Inter-individual functional variability

277 Phylogenetic Investigation of Communities by Reconstruction of Unobserved States  
278 (PICRUST2) was used to predict the functional potential of the bacterial communities in the  
279 samples (18). PERMANOVA analysis of the results revealed statistically significant  
280 differences between sampling sites (Bray Curtis dissimilarity based on KO predictions  
281  $F=2.86$ ,  $p=0.01$ ; similar values were observed for EC and MetaCyc predictions). The impact  
282 of the sinus ostium size on the functional predictions was not significant.

283

## 284 Alpha diversity in the sinonasal samples

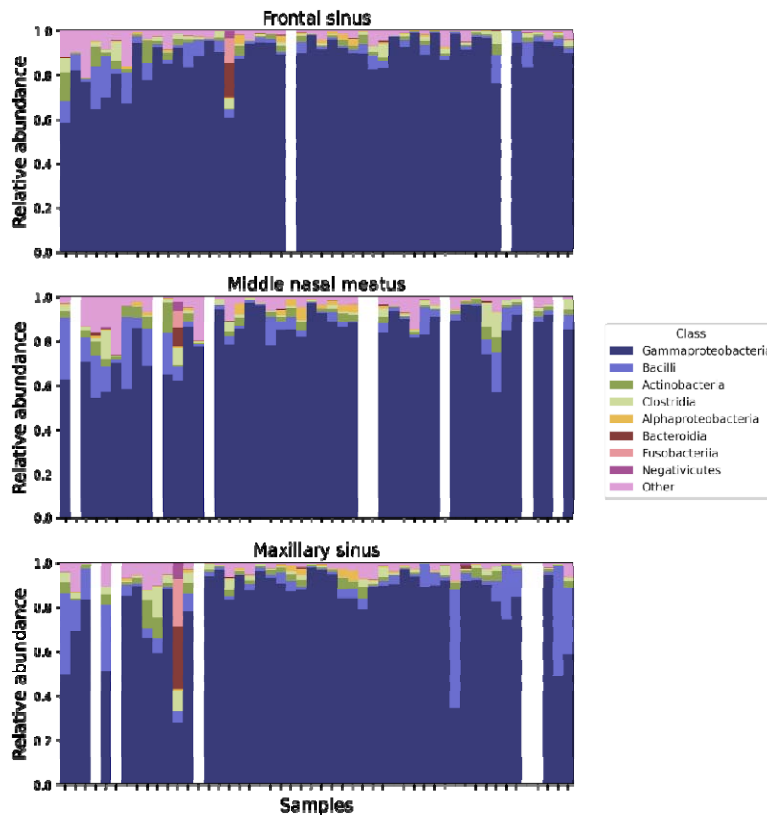
285 We found no significant differences in alpha diversity (Faith's phylogenetic diversity)  
286 between samples from the middle meatus, the maxillary sinus and the frontal sinus (data not  
287 shown). Moreover, there were no differences between alpha diversity in the samples from the

288 maxillary sinuses with wide, narrow and blocked ostia. The taxonomic composition of the  
289 samples can be found in the supplementary material (Fig. S1 and Fig. S2).

290 The relationships between alpha diversity and clinical metadata were not statistically  
291 significant. Still, we observed decreased alpha diversity in patients with nasal polyps, a  
292 history of recurrent exacerbations or comorbidities such as aspirin-exacerbated respiratory  
293 disease, gastroesophageal reflux or asthma. Recent steroid use was almost significantly  
294 associated with lower alpha diversity (t-Student test,  $p = 0.052$ ).

295

296 **Fig. S1.** The taxonomic composition of the samples from the frontal sinus, middle nasal  
297 meatus and maxillary sinus.

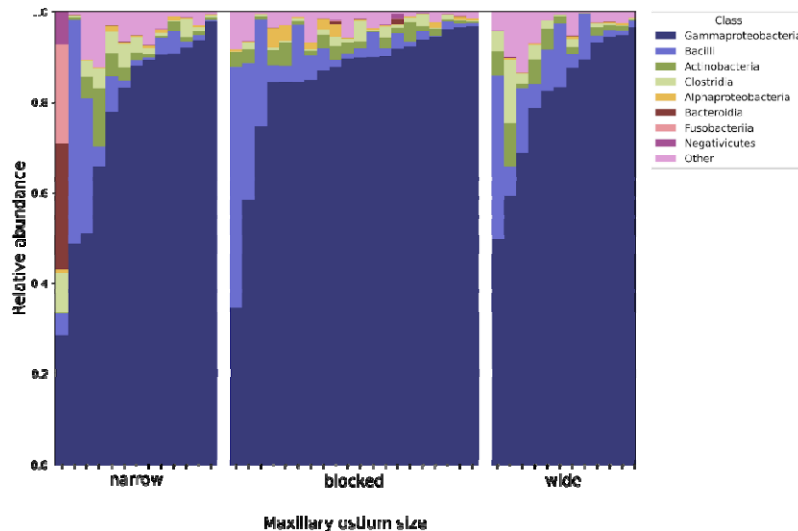


298

299

300 **Fig. S2.** The taxonomic composition of the samples from the maxillary sinus in the groups  
301 with wide, narrow and blocked maxillary ostium.

302



303

304

305

## 306 Discussion

307 In this study, we explored the topographical differences between the microbial communities  
308 in several locations of the sinonasal cavity. We investigated whether the sinonasal cavity  
309 could be considered a homogenous habitat or a system of communicating but distinct  
310 microenvironments.

311 Most studies on the subject suggested that the samples from the sinus cavities and the middle  
312 nasal meatus were similar (7, 8) and the samples from the middle nasal meatus were  
313 representative of the sinuses (1). We decided to question this paradigm because in CRS the  
314 drainage into the middle meatus is frequently impaired. It has also been postulated that the  
315 hypoxic microenvironment inside an occluded sinus may promote the development of a  
316 distinct microbiome while the wide opening of the ostium may decrease the concentration of  
317 nitric oxide that controls bacterial growth (7, 19-21). Moreover, our previous study that  
318 utilized microbial cultures showed that the middle meatus swabs missed pathogens found in  
319 the sinuses in 24% of patients (6). We also found it striking that in several studies of the

320 sinonasal microbiome some patients presented significant differences between sampling sites  
321 (7, 11).

322 These controversies motivated us to explore whether marked heterogeneity within the  
323 sinonasal microbiome is frequently encountered and whether it is associated with obstruction  
324 of the sinus ostia or driven by other factors. For this purpose, we conducted a detailed analysis  
325 of the sequencing data results in the context of the individual patterns of drainage pathway  
326 disruption and clinical metadata.

327

### 328 **The microbiome is not universally homogenous across the sinonasal cavity**

329 Differences between samples are quantified by beta diversity measures. However, there is no  
330 consensus on what values of these metrics indicate truly meaningful differences. Joss *et al.*  
331 assumed that if the weighted UniFrac distance between samples obtained from two sampling  
332 sites in the sinuses did not exceed 0.2 then sampling from one site could be considered  
333 representative of the other (11). In our study, values of weighted UniFrac above 0.2 were  
334 noted in 66% of patients for the middle meatus-maxillary sinus distances and 73% of patients  
335 for the middle meatus-frontal sinus distances. Therefore, according to the definition suggested  
336 above, the middle meatus samples were not representative of the sinus samples in the majority  
337 of patients. The maximal Bray-Curtis value noted in our study reached 0.75 for the middle  
338 meatus-maxillary sinus distance and 0.92 for the middle meatus-frontal sinus distance. Joss *et*  
339 *al.* noted between-site weighted UniFrac distances that exceeded 0.2 in 38% of patients while  
340 De Boeck *et al.* reported the median Bray-Curtis distance of 0.27 between the maxillary sinus  
341 and the ethmoid cells (10). These results show that specimens from a single site may not be  
342 representative of the whole sinonasal cavity.

343 Even though beta diversity metrics are stable within a single experiment and saturate quickly  
344 with sequencing depth, they still retain variance dependent upon experiment design (i.e.  
345 primer selection) (22), sequencing parameters (i.e. sequencing depth) (23) and data  
346 preprocessing pipeline (i.e. denoising method, trimming and rarefaction parameters) (24). The  
347 additional layer of complexity is added by a nonlinear nature of common beta diversity  
348 measures. All these factors restrict the ability to interpret raw numbers. To alleviate this  
349 problem, in our analysis we compare beta diversity between different sampling sites within



350 one patient and beta diversity of the same sampling sites across all patients, utilizing the latter  
351 as a point of reference.

352

353 **In a minority of patients, the intrapersonal differences outweigh the interpersonal**  
354 **variation**

355 Although the values of beta-diversity measures suggested large between-site dissimilarity in  
356 many patients, PERMANOVA analysis did not demonstrate statistically significant  
357 differences between sampling sites. The adonis test indicated that interpersonal differences  
358 were a significant source of variation, but not the sampling site. There were also no significant  
359 differences in alpha diversity between the three locations. The observation that interpersonal  
360 variation significantly outweighs intrapersonal variation remains in agreement with the  
361 observations of other authors (7, 9-11, 25). Therefore, for certain types of studies, sampling  
362 from the middle meatus can be sufficient to represent the composition of the whole sinonasal  
363 microbial community (7). De Boeck, who studied a large group of 190 patients with CRS,  
364 observed that the sampling site explained a small (2.2%) but statistically significant  
365 proportion of microbial variation (10). However, differences were observed between the  
366 sinuses and the nasopharynx or the anterior nares. The maxillary sinus and the ethmoid sinus  
367 were not significantly different.

368 Nevertheless, in our study, we showed that in approximately 15% of patients the intrapatient  
369 middle meatus-sinus distances were greater than the differences between the patient's middle  
370 meatus and the sinuses of other study participants. (Fig. 3) These observations prove that the  
371 apparent intrapersonal variation in some individuals is not captured by statistical analyses that  
372 comprise the whole group of CRS patients.

373 Moreover, PERMANOVA analysis of PICRUSt2 results proved that the functional potential  
374 of the bacterial communities differed significantly between various locations within one  
375 patient. This observation sheds new light on the sinonasal microbial ecology. It indicates that  
376 the microenvironments of the sinuses are heterogeneous and the metabolic activity of bacteria  
377 in various niches is distinct. These differences could be explored in detail by deep shotgun  
378 sequencing.

379

380 **Differences between sampling sites are not related to their anatomical separation**

381 To investigate whether the microbiome heterogeneity was caused by the anatomical  
382 separation of the sampling sites, we analysed the configuration of ostia occlusion in each  
383 patient (Fig. 1). We expected that blockage of the drainage pathway between the sinus and the  
384 middle meatus would result in increased differences between the two sites. Our observations  
385 did not support these predictions.

386 The largest beta diversity distances between the maxillary sinus and the middle meatus were  
387 noted in patients with a narrow maxillary ostium. Blocked maxillary ostia were associated  
388 with smaller beta diversity between the maxillary sinus and the middle meatus (Fig. 1).  
389 Narrow maxillary ostia were also associated with greater distances between the frontal sinus  
390 and the middle meatus than blocked or wide maxillary ostia. It suggests that a more divergent  
391 sinonasal microbiome was observed in patients whose sinus anatomy was not significantly  
392 altered by inflammatory changes (blocked ostia) or by previous surgery (wide ostia). This  
393 result was unexpected because other authors reported greater meatus-sinus differences in CRS  
394 patients than in healthy individuals and concluded that microbial communities in the sinuses  
395 diverge during CRS (9, 25). However, the differences noted in these studies were not  
396 statistically significant and require verification in larger study groups.

397 The results of our study did not support the hypothesis that the microenvironment in an  
398 occluded sinus stimulates the development of a distinct microbial community. The analysis of  
399 alpha-diversity and PICRUSt2 predictions of the metabolic potential of the microbiome did  
400 not reveal any significant microbiome differences between blocked and well-ventilated  
401 sinuses. However, the predictions provided by PICRUSt2 are only an approximation of  
402 metagenomic functional potential and this phenomenon requires further investigation with the  
403 use of deep shotgun sequencing and untargeted metabolomics.

404

405 **The degree of sinonasal microbiome variability is characteristic of an individual and not**  
406 **of the topography of the sinuses**

407 Our study showed that between-site beta diversity distances within a patient were positively  
408 correlated (patients with greater meatus-maxillary distances also had greater meatus-frontal  
409 distances, regardless of the size of the ostia). This observation indicates that a tendency for an

410 increased or decreased between-site variability is a feature that characterizes an individual's  
411 sinonasal microbiome rather than local drainage pathway obstruction between sites.

412 Patients with nasal polyps or extensive mucosal changes in the sinuses had less dissimilar  
413 microbial communities in the middle meatus and the frontal sinus than patients without nasal  
414 polyps. Massive sinus opacification and nasal polyps are frequently associated with type 2  
415 inflammation (1). The alpha diversity was not significantly associated with the sampling site  
416 or the patency of the sinus ostium. Still, it tended to be lower in patients who presented certain  
417 clinical features (recent steroid use, nasal polyps, comorbidities such as asthma, aspirin-  
418 exacerbated respiratory disease, and gastroesophageal reflux). Therefore, it is probable that  
419 the spatial heterogeneity and composition of the microbiome depend on the individual  
420 characteristics of the patient and the type of inflammatory reaction rather than the pattern of  
421 ostia occlusion.

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428

### 429 **Authorship contribution**

430 Joanna Szaleniec – conceptualization, data curation, formal analysis, investigation,  
431 methodology, project administration, resources, supervision, visualization, writing – original  
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443

#### 444 **Conflict of interest**

445 The authors declare no conflict of interest.

446

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