# Neurogenomic landscape associated with status-dependent cooperative behavior

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**Abstract**

The neurogenomic mechanisms mediating male-male reproductive cooperative behaviors are unknown. We leveraged extensive transcriptomic and behavioral data on a neotropical bird (*Pipra filicauda*) that perform cooperative courtship displays to understand these mechanisms. Cooperative display is modulated by testosterone, it promotes cooperation in non-territorial birds, but suppresses cooperation in territory holders. We sought to understand the neurogenomic underpinnings of these three related traits: social status, cooperative behavior, and testosterone phenotype. To do this, we profiled gene expression in 10 brain nuclei spanning the social decision-making network (SDMN), and two key endocrine tissues that regulate social behavior. We associated gene expression with each bird's behavioral and endocrine profile derived from 3 years of repeated measures taken from free-living birds in the Ecuadorian Amazon. We found

distinct landscapes of constitutive gene expression were associated with social status,

testosterone phenotype, and cooperation, reflecting the modular organization and engagement of

neuroendocrine tissues. Sex-steroid and neuropeptide signaling appeared to be important in

mediating status-specific relationships between testosterone and cooperation, suggesting shared

regulatory mechanisms with male aggressive and sexual behaviors. We also identified

differentially regulated genes involved in cellular activity and synaptic potentiation, suggesting

multiple mechanisms underpin these genomic states. Finally, we identified SDMN-wide gene

expression differences between territorial and floater males that could form the basis of "status-

specific" neurophysiological phenotypes, potentially mediated by testosterone and growth

hormone. Overall, our findings provide new, systems-level insights into the mechanisms of

cooperative behavior and suggest that differences in neurogenomic state are the basis for

individual differences in social behavior.

**Introduction**

Cooperation is the synchronized behavior(s) of individuals to produce a net fitness benefit for one or more individuals (Taborsky & Taborsky, 2015). An individual's decisions about cooperation are based on immediate social stimuli, past experiences, developmental effects, and genetics. These effects contribute to repeatable, among-individual variation in behaviors – a behavioral phenotype – where some individuals are more cooperative than others. To understand the mechanisms of cooperation, it is important to understand not only the elements of neurophysiological signal processing, but also the genetic and neurophysiological components that underpin behavioral phenotypes. Understanding these mechanisms enables us to link how genetic and neurophysiological architecture can constrain and facilitate the evolution of cooperation (Hofmann et al., 2014; Soares et al., 2010; Taborsky & Taborsky, 2015). However, the mechanisms underlying the diversity of behaviors related to cooperation remain unclear (Díaz-Muñoz et al., 2014; Kasper et al., 2017). In particular, our understanding of the neuroendocrine mechanisms underlying seemingly paradoxical male-male reproductive cooperative behaviors is only emerging (Díaz-Muñoz et al., 2014; DuVal & Goymann, 2011; Jones & DuVal, 2021; Loveland et al., 2021; Ryder et al., 2020; Vernasco et al., 2020).

Differences in cooperative behavioral phenotype are partly achieved through pleiotropic yet modular effects of hormone signaling systems (Cox, 2020; Ketterson et al., 2009). Hormonal pleiotropy mediates phenotypic integration, whereby multiple traits covary with a hormonal signal (Cox, 2020; Ketterson et al., 2009). Trait independence is facilitated by variation in hormone receptor expression, where tissues across the body are modular in their response to hormone signals, enabling diverse and flexible responses to hormonal and social signals (Ketterson et al., 2009; Lipshutz et al., 2019). In particular, sex steroid concentration and receptor distribution in neural tissues are important mediators of social behaviors, including cooperation (Kasper et al., 2017; Soares et al., 2010). The neural substrates modulating social behaviors are conserved across vertebrates, in the social decision making network (SDMN) (O'Connell & Hofmann, 2011). The SDMN comprises reciprocally interconnected brain nuclei rich in steroid receptors, steroidogenic enzymes, and steroid-modulated neuropeptide systems (Goodson, 2005; Newman, 1999; O'Connell & Hofmann, 2011). This modular and dynamic network activates different combinations of brain regions in response to different social contexts (Goodson, 2005; Newman, 1999). Therefore, the neurological response in a single nucleus is not

reflective of all systems engaged in modulating a given behavior. Consistent variation among
individuals in neuroendocrine gene expression across the SDMN likely underlies repeatable
individual differences in social behavior and behavioral phenotype. For example, variation in the
expression of genes involved in sex-steroid signaling pathways across SDMN nuclei can modify
the male brain's sensitivity to testosterone, resulting in distinct behavioral responses to a specific
hormone signal. Indeed, recent research suggests that constitutive gene expression or
neurogenomic state, characterized by co-expressed gene suites in behaviorally relevant brain
regions, is associated with behavioral phenotypes (Antunes et al., 2021; Kabelik et al., 2021;
Lattin et al., 2021).

For the first time, we characterize constitutive gene expression across the neuroendocrine
system in relation to variation in a male-male cooperative behavior. The wire-tailed manakin
(*Pipra filicauda*) is a neotropical lek breeding bird in which unrelated males perform cooperative
displays to attract females (Figure 1A). Territorial and subordinate non-territorial (floater) males
form long-term display partnerships and form complex social networks (Figure 1A, B) (Ryder et
al., 2008; Ryder, Blake, et al., 2011). Status and cooperation are directly linked to fitness,
whereby territory holders with more display partners sire more offspring, and floaters with more
partners are more likely to ascend to territorial status (Ryder et al., 2008, 2009). Individuals have
a repeatable 'testosterone phenotype' where territorial males tend to have higher testosterone than
floaters (Figure 1D), and circulating testosterone has a status-specific effect on the cooperative
display behavior (Ryder et al., 2020; Ryder, Horton, et al., 2011). Experimental and
observational evidence show that higher testosterone levels are antagonistic to cooperation in
territorial males but promote cooperation in floater males (Ryder et al., 2020; Vernasco et al.,
2020) (Figure 1C).

To explore the neurogenomic basis of the status-specific androgenic modulation of
cooperative behavior in these birds, we quantify transcriptome-level gene expression in the
SDMN, pituitary and testes of free-living male wire-tailed manakins in the Ecuadorian Amazon.
Captive studies can obscure ecologically and evolutionary relevant variation in environmental
and behavioral responses (Griffith et al., 2017; Kasper et al., 2017), and we demonstrate the
exciting applicability of neuro-transcriptional methods in challenging rainforest conditions. We
aim to characterize the genes and neuroendocrine pathways that modulate individual variation in

cooperative behavior, social status, and testosterone phenotype as a modifier of both social status

and cooperative behavior. Given the androgenic regulation of cooperative display (Ryder et al.,

2020; Vernasco et al., 2020), we hypothesize that differences in gene regulation will manifest in

SDNM nuclei with high density of sex-steroid receptors and endocrine tissues (Goodson, 2005;

Newman, 1999), and will involve expression of candidate genes relating to sex-steroid signaling,

metabolism, and neuropeptide systems (Goodson & Thompson, 2010; Kingsbury & Wilson,

2016). We anticipate brain region-specific gene expression patterns within the modular SDMN

(Figure 1G), as well as systemic effects of testosterone on gene expression across the brain (Cox,

2020; Ketterson et al., 2009). We discuss our results in the context of modes of phenotypic

evolution and similarities with mechanisms of other cooperative and social behaviors.

## Materials and Methods

All methods described here were approved by Smithsonian Animal Care and Use Committee

(protocols 12-12,14-25,17-11), as well as the Ecuadorian Ministry of the Environment (MAE-

DNB-CM-2015-0008), and samples were transported with export permit (006-016-EXP-1C-

FAU-DNB/MA) and import permit (USDA APHIS 126133). Detailed methods can be found in

the supplementary material, and full code from analyses in R v 4.0.2 are available on GitHub

(https://periperipatus.github.io/PIFI_brain_transcriptome/) and FigShare

(10.25573/data.22186516).

### *Field Methods*

We sampled 16 wild territorial and floater wire-tailed manakin males for RNA sequencing from

Tiputini Biodiversity Station in the lowland Amazon rainforest of Ecuador (Table S1.1). Males

were categorized by social status: where the territorial birds (n=9) were males in definitive

plumage that held a consistent display territory over the study period, while floaters (n=7) had

either definitive or pre-definitive plumage and did not occupy consistent territories within leks

(Ryder et al., 2008). We used repeated testosterone sampling on 12 of these males to characterize

their testosterone phenotype (Figure 1D, Supplement 1). A male's testosterone phenotype is

represented by mean of the residuals of a linear regression from repeated sampling of

testosterone concentration, and this measure was previously shown to be repeatable within

individuals (Supplement 1 for more detail, see also (Ryder et al., 2020)). Previous research has

shown that territorial males have, on average, higher circulating testosterone than floaters (Ryder
et al., 2020; Ryder, Horton, et al., 2011), and is reflected in our sampled males (Figure 1D).

Social network structure and individual variation in cooperative display were characterized using
autonomous radio-telemetry and tag proximity detections within territories on a total population
of 180 male manakins over multiple years (Dakin & Ryder, 2018; Ryder et al., 2012, 2020). We
used this tagging data to calculate social network strength for the same 12 males with repeated
testosterone measures. Social network strength is a proxy for cooperative tendency and is the
time spent interacting with other males averaged over the study period (Figure 1E), and was the
most repeatable within individuals (Ryder et al., 2020). This particular proxy for cooperation
showed the strongest status-specific relationship with an individual's testosterone phenotype
(Ryder et al., 2020) (Figure 1C), but the overall frequency of cooperative interactions did not
differ between territorial and floater males (Figure 1E).

***Microdissection and RNAseq***
Methods for brain extraction, preservation, cryosectioning, microdissection, and RNA
extraction followed those published previously (Horton, Ryder, et al., 2020). Pituitaries were
preserved in RNAlater™ (Invitrogen) at ambient temperatures until imported into the United
States. We excised tissue from 10 different nuclei involved in social behavior – nine of which are
from the SDMN. The SDMN reflects the interconnectivity of two networks involved in
regulating social behavior (O'Connell & Hofmann, 2011): Social Behavior Network (SBN)
(Goodson, 2005; Newman, 1999) and the Mesolimbic Reward System (MRS) (O'Connell &
Hofmann, 2011; Olds & Milner, 1954). Figure 1F indicates the nomenclature used and the
relationships of these nuclei to the SBN and MRS. To clarify bird specific nomenclature: the
arcopallium intermedium (AI) is a region that may be, in part, homologous the basolateral
amygdala of mammals, a node of the MRS (O'Connell & Hofmann, 2011), which is
hypothesized to play a role in androgen-dependent manakin display behavior (Fusani et al.,
2014); and the nucleus taenia (TnA) is homologous to the mammalian medial amygdala
(O'Connell & Hofmann, 2011). The paraventricular nucleus (PVN) is not yet formally
recognized as part of the SDMN, despite its neuropeptide projections to nodes of the SDMN and
established role as a major regulator of vertebrate social behavior (Goodson & Kingsbury, 2013;
Goodson & Thompson, 2010). We additionally sampled the pituitary gland and testes
immediately after the brains. Testes were handled the same as brains (Horton, Ryder, et al.,

2020) and pituitaries were preserved in RNAlater™ (Invitrogen) at ambient temperatures until

imported into the United States.

Microdissected tissues from both hemispheres were combined for sequencing. RNA was

extracted and sequenced according to previously described methods (Horton, Ryder, et al.,

2020). RNA libraries for the various tissues and field seasons were randomized amongst

sequencing batches (Supplement 1, Table S1.1 & S1.2). All samples analyzed are associated

with BioProject PRJNA437157, and SRR12660169-198, SRR19521260-271, SRR19521432-

575.

***Gene expression analyses***

Reads were aligned to the annotated *Pipra filicauda* genome (GCA_003945595.1) using

splice-aware aligner STAR v2.7.5 (Dobin et al., 2013), and gene-level counts were obtained

from featureCounts v2.0.1 (Liao et al., 2014). A total of 170 libraries were retained after quality

filtering in each tissue (Supplement 1 & 3), but the number of individuals used in each tissue

varies (Table S1.1 & S1.2). To further characterize tissue and brain nucleus function in this

species (Horton, Ryder, et al., 2020), we characterized differences in gene expression among

tissues and brain nuclei using principal components analysis in PCAtools v2.8.0 and Weighted

Gene Co-Expression Analysis (WGCNA v1.71) (Blighe & Lun, 2020; Langfelder & Horvath,

2008) (Supplement 2). All analyses were conducted in R v 4.0.2 (R Core Team, 2020).

To describe the role of tissues and brain nuclei in relation to hormonal and behavioral

phenotypes we analyzed differential expression using DESeq2 v1.36 (Love et al., 2014) and co-

expression using WGCNA (Langfelder & Horvath, 2008) in each tissue and nucleus separately.

For a subset of tissues, we had an extra four individuals characterized for social status from a

previous study (Horton, Ryder, et al., 2020), for which behavioral and hormone sampling data

were unavailable. Owing to this, and to not overparameterize the models that account for batch

effects, we identified differentially expressed genes (DEGs) associated with each of our traits of

interest (social status, testosterone phenotype and social network strength) separately. These

were run as separate negative binomial generalized linear models for each trait that accounted for

batch effects where necessary using DESeq2 (Love et al., 2014) (For detailed description see

Supplement 3) Testosterone and social network strength were treated as continuous variables.

Significant DEGs were considered to be correlated with the predictor variable, and $Log_2$ Fold

Change (LFC) value represents the LFC of gene expression per unit of the predictor variable. In

all cases, significant DEGs were identified using the Wald Test and the default FDR corrected p-

value (q<0.1). The identity of many hub genes identified by WGCNA were similar to DEGs

identified byDESeq2, so for brevity and simplicity we focus mostly on the differential expression

results, but the full results are presented in Supplement 5 (WGCNA) and Supplement 3

(differential expression).

Due to the key role testosterone plays in modulating behavior in this species (Ryder et al.,

2020), we further analyzed nine candidate genes involved in sex-hormone metabolism and

signaling (Figure S4.1), as well as nine steroid sensitive neuropeptides and their receptors

(Figure S4.2; Table S1.3). Candidate genes showing differential gene expression with an

uncorrected $p < 0.05$ was considered "significant" and examined further. In addition to analyzing

models of status, mean testosterone and social network strength separately, we also explored

whether candidate genes were involved in mediating status-specific relationships between mean

testosterone and cooperative behavior (Ryder et al., 2020) by extracting "significant" candidate

genes from a final DESeq2 interaction analysis (model: expression ~ status x mean testosterone).

We primarily used this model to identify candidate genes with status-specific expression rather

than discovering new DEGs due to overfitting concerns.  The resulting putative gene-trait

relationships were plotted (Supplement 4), and those with highly influential observations or $R2 \leq$

0.2 were excluded. For candidates derived from the interaction effect models or genes that were

associated with multiple traits, the most convincing relationship was determined using a

stepdown AIC procedure (Supplement 4). Considering the smaller set of candidate genes,

uncorrected p-values were deemed significant ($p<0.05$), but we also report the FDR corrected

values.

To assess neuroanatomical constraints on gene expression patterns, we examined overlap

among brain nuclei by comparing ranked gene-lists (Supplement 6). The gene-lists were ranked

using -log10 transformed p-values multiplied by the expression direction. Similarity between

tissue pairs was measured using Pearson correlation and rank-rank hypergeometric overlap

(RRHO) test (Cahill et al., 2018).

Testosterone has pleiotropic effects, influencing gene expression in similar ways in

multiple tissues (Cox, 2020; Ketterson et al., 2009). We reasoned that these effects would be

reflected in shared (systemic) signatures of differential expression across brain nuclei. To
explore systemic gene expression patterns, we calculated median p-values from the previous
analysis and identified genes consistently differentially expressed across the entire brain (p-value
$< 0.05$, transformed value $> |1.3|$). Further, we correlated eigenvalues of each principal
component (PC) from the whole-brain PCA analysis (Figure 1G inset) with our variables of
interest using pcatools::eigencorplot() (Blighe & Lun, 2020) (Supplement 1). Correlations with
social status were validated by a linear mixed model in the nlme package v3.1 (Pinheiro et al.,
2016), which included individual as a random effect and accounted for correlated batch variables
(Supplement 1).

We used clusterProfiler v3.16.1(Yu et al., 2012) to determine patterns of functionality in
DEGs and WGCNA modules with Gene Ontology enrichment analyses. If the total number of
FDR corrected DEGs was less than 40, we used gene set enrichment analysis instead which
leverages the rank order of p-values and direction of expression (Supplement 3).

## Results

237 As expected, gene expression analyses revealed substantial differences among tissues and
238 brain regions (Figure 1G; Figure S2.7). These expression patterns were more similar among
239 adjacent nuclei, forming major brain-region groupings of the amygdala, hypothalamus, and
240 midbrain (Figure 1G). Additionally, our 18 candidate neuroendocrine genes (Table S1.3)
241 displayed tissue and brain region-specific expression (Table 2.2, Figures S4.1-4.2).

***Social status and testosterone phenotype are associated with tissue and brain region-specific***
***patterns of gene expression***

We observed a landscape of gene expression in association with testosterone phenotype
(Figure 2A) and social status (Figure 2B), with a varying number of differentially expressed
genes (DEGs) or co-expression modules identified across different tissues. Similarly, across
brain regions, we identified relationships between many of our 18 candidate genes and status,
testosterone phenotype, or their interaction (Figure 2C). Mean testosterone was a continuous
predictor in DESeq2, as such DEGs indicate up-or downregulation with a higher testosterone
phenotype (Figure 2A), while DEGs identified in social status indicated up or down regulation in
territorial males (Figure 2B). Here, we primarily focus on the tissues with large numbers of

DEGs and significant candidate genes for the main manuscript, but the co-expression results
were similar to our differential expression results, and all results are shown in Supplements 3 and
5. The number of DEGs in each tissue showed a similar pattern between testosterone phenotype
and status compared to other trait comparisons ($R = 0.5$, $p = 0.08$, Figure S3.7), which was
consistent across co-expression modules (Supplement 5) and candidate gene analysis (Figure 2C
& Figure 3B). This similarity was expected due to the correlation between testosterone and status
(Figure 1D) (Ryder et al., 2020; Ryder, Horton, et al., 2011).

The testes and TnA each had over 1,000 genes differentially expressed (at uncorrected p-
value $< 0.05$) in association with both testosterone phenotype and status (144 and 252 genes after
FDR correction $q<0.01$ in association with mean testosterone; 2 and 13 after FDR correction in
association with social status). DEGs identified in association with testosterone phenotype in the
testes were enriched for genes involved with response to corticosterone (GO:0051412) and
synaptic transmission and organization (e.g., GO:0007268, GO:0050808) among others (Figure
S3.20). Notable DEGs included steroidogenic acute regulatory protein (STAR, LFC = -1.1, p =
$2\times10^{-7}$, $q = 0.0003$, $R^2 = 0.6$) and candidate gene AVPR1A (LFC = 0.8, p = $5\times10^{-5}$, q = 0.01, $R^2 =$
0.7; Figure S4.4). Note that $Log_2$ Fold Change (LFC) represents a change in gene expression per
unit of the predictor variable. These transcriptional differences do not correspond to significant
differences in testis size in relation to testosterone phenotype (Table S3.2), nor social status
(Table S3.1 & S3.2).

Among the 252 DEGs associated with testosterone phenotype in TnA, there was non-
significant enrichment for genes involved in neural crest migration (GO:0001755), learning and
memory (GO:0007613, GO:0007612), and long-term synaptic potentiation (e.g., GO:0060291)
(Figure S3.28). Notably, the candidate gene estrogen receptor 2 (ESR2; aka Erβ) was
upregulated in TnA of males with higher testosterone (Figure 2C; LFC = 0.8, p = 0.001, q =
0.08, $R^2 = 0.4$). Other notable DEGs in TnA included upregulation of immediate early gene JUN,
also a 'coral1' hub gene (module membership (MM) = 0.93; LFC = 0.61, p = 0.0004, q = 0.05)
and corticotropin releasing hormone receptor 2 (CRHR2, LFC = 1, q = 0.006, $R^2 = 0.5$).

Not all nuclei showed similar patterns between status and testosterone phenotype. For
example, ICo of the midbrain showed 161 DEGs associated with testosterone phenotype after
FDR correction but did not have substantial association with social status (3 DEGs). Enriched

GO terms in ICo were primarily related to motor proteins and cilia (e.g., GO:0003341, Figure S3.26). Intriguingly, although the POM did not reveal a high number of DEGs in association with testosterone phenotype (45), our analysis revealed differential expression for compelling gene ontology categories and candidate neuroendocrine genes (Figure S3.25, Figure S4.10, Figure S5.7). Notably, males with higher testosterone exhibited upregulation of candidate gene androgen receptor (AR) in POM (Figure 2D). The 'floralwhite' module, which comprised many upregulated DEGs, was significantly associated with both testosterone phenotype and, to a lesser extent, social status (Figure S5.6). This module showed enrichment for biological processes related to G protein-coupled receptor signaling pathways (GO:0007186) and neuropeptide signaling pathway (GO:0007218), with statistically non-significant enrichment for steroid receptor signaling (GO:0030518), learning and memory (GO:0007611), and nervous system development (GO:0007399) (Figure S3.7). Indeed, many of the shared hub-genes and DEGs interact with candidate sex hormone pathways, such as prolactin releasing hormone (PRLH, MM = 0.91, LFC = 2.4, q = $5\times10^{-5}$, $R^2$ = 0.6, Figure S3.25A), growth regulating estrogen receptor binding 1 (GREB1, MM = 0.86, LFC = 0.6, q = $3\times10^{-5}$, $R^2$ = 0.7, Figure S3.25A), and gonadotropin releasing hormone 1 (GNRH1, MM = 0.76, LFC = 0.4, p = 0.006, q = 0.2, $R^2$ = 0.4). Other notable 'floralwhite' hub genes include the candidate gene progesterone receptor (PGR, MM = 0.81) which was also associated with social status (Figure 2C; LFC = 0.4 p = 0.006, q = 0.7) and serotonin receptor 5A (HTR5A, MM = 0.87, LFC = 0.4, p = 0.04, q = 0.6). Candidate gene aromatase (LOC113993669/CYP19A1) was also upregulated in males with higher testosterone phenotype (Figure 2C, LFC = 0.6, p = 0.02, q = 0.4, $R^2$ = 0.5), but was found in the 'lightcoral' co-expression module (MM= 0.86).

Despite overall similarity in gene expression associations with testosterone phenotype and status, certain brain regions exhibited stronger associations with either status or the interaction between status and testosterone phenotype (Figure 2B). Likewise, the 18 candidate neuroendocrine genes also showed varying associations with status or the interaction across different nuclei (Figure 2C). PVN of the hypothalamus showed the most DEGs associated with social status after FDR correction (42). These DEGs were enriched for cytoskeletal movement genes involved in intracellular transport and cilia (e.g., GO:0003341, GO:0061640), and non-significant enrichment for cholesterol biosynthesis (GO:0006695, GO:0045540) (Figure S3.12B). Candidate gene expression also revealed status-specific associations with testosterone

phenotype in the PVN, that is a statistical interaction between status and testosterone phenotype

(Figure 2C). In particular, AR expression was positively correlated with testosterone phenotype

in floater males, but not in territorial males (Figure 2F; LFC = -0.7, p = 0.008, q = 0.9). The PVN

of territorial males also showed higher VIPR1 (Figure S4.8; LFC = 0.3, p = 0.02, q = 0.4) and

lower ESR2 expression (LFC = -0.6, p = 0.006, q = 0.2). In BSTm, a region that bridges the

SDMN and MRS, there were more than 1000 DEGs, but only nine after FDR correction. Among

these, territorial males showed higher AR expression compared to floater males (Figure 2E; LFC

= 0.6, p = 0.0007, q =0.2). Further, testosterone metabolism gene 5-alpha reductase (SRD5A2)

was upregulated in the LS of territorial males (Figure S4.20; LFC = 0.7, p = 0.0005, q = 0.4). In

the TnA, the VIP system was implicated in status-specific responses to testosterone, with VIPR1

upregulated in association with testosterone in territorial males (Figure S4.17; LFC = 1, p = 0.04

q = 0.7).

325            GCt was a hotbed for gene expression associated with social status and status ×

testosterone phenotype interaction (Figure 1B, Figure S3.6). In relation to social status, gene set

enrichment analysis revealed a significant downregulation of genes involved in synapse

assembly (GO:0051965), response to estradiol (GO:0032355), as well as upregulation of genes

related to RNA processing (e.g., GO:0000184, GO:0006396), and translation (e.g., GO:0006413,

GO:0006412) (Figure S3.15). Candidate neuroendocrine genes showed status specific expression

in association with testosterone phenotype in this region including progesterone receptor (PGR,

Figure S4.15; LFC = 1.9, p= 0.003, q = 0.1), estrogen receptor 1 (ESR1; aka Erα; LFC = 1.1 p =

0.04, q = 0.4), prolactin receptor (PRLR; LFC = 0.7, p = 0.01, q = 0.2), and vasoactive intestinal

peptide (VIP; Figure S4.15; LFC = 1.6, q = 0.005, q = 0.1). Further, aromatase was

downregulated with higher testosterone phenotype in the GCt (LFC = -1.3, p = 0.002, q = 0.3, $R^2$

= 0.4).

***Cooperative behavior is associated with a distinct gene expression landscape***

338            Social network strength (the number of cooperative interactions between individuals)

varies widely among territorial and floater males (Figure 1E), representing a key dimension of

social behavior in this species. We predicted that the associations between gene expression and

cooperative behavior would be different from those observed for social status and testosterone

phenotype. Indeed, across tissues we found a distinct landscape of constitutive gene expression

in relation to cooperative behavior (Figure 3A, Figure S3.5), as well as among candidate genes

(Figure 3B). AH had the highest number of DEGs associated with a male's network strength,

with non-significant enrichment for genes involved in RNA metabolism (e.g., GO:0043928) and

regulation of translation (e.g., GO:0017148), development (e.g., GO:0061061, GO:0042472) and

synaptic potentiation (e.g., GO:0007274) (Figure S3.35). The pituitary also showed an

abundance of differentially expressed genes associated with male's network strength (109 genes

after FDR correction q < 0.1), with non-significant enrichment for actin filament organization

(GO:0007015), neural differentiation (e.g., GO:0031103, GO:0045664) and the cell cycle (e.g.,

GO:0043065, GO:0071158.) (Figure S3.33B). Within the pituitary, candidate genes prolactin

(PRL) (Figure 3C; LFC = -0.6, p=$5 \times 10^{-5}$ q = 0.03, $R^2$ = 0.7), ESR2 (Figure S4.5; LFC = -0.7, p =

0.04, q = 0.5, $R^2$ = 0.5) and VIPR2 (Figure S4.5; LFC = -0.4, p = 0.008, q = 0.3, $R^2$ = 0.4) were

downregulated in highly cooperative males. Intriguingly, in the PVN there was a weak negative

correlation between the expression of the candidate gene oxytocin (OXT) and network strength

(Figure 3D; LFC = -0.3, p = 0.04, q = 0.6, $R^2$ = 0.2). Furthermore, although there was less

representation of sex-steroid related pathways in the candidate gene analysis (Figure 2B), PGR

was downregulated in males with higher social network strength in BSTm (Figure 2E; LFC = -

0.3, p = $5 \times 10^{-4}$, q = 0.2, $R^2$ = 0.5).

Many genes in the LS were differentially expressed in association with an individual's

social network strength (98 genes after FDR correction q < 0.1). Among these, the top non-

significant terms were broadly involved in development (e.g., GO:0048538, GO:0048593), and

neural organization (e.g., GO:0099175, GO:0050771) (Figure S3.42). No candidate genes were

linked to social network strength in LS. Although few genes were differentially expressed in

BSTm related to social network strength, RRHO analysis showed that LS and BSTm shared

similar expression associations with strength (Cahill et al., 2018) (Figure 3G, F). In contrast, LS

and BSTm showed limited similarity in gene expression related to testosterone phenotype

(Figure 4H, I) or social status (Figure S6.1A), suggesting their unique role in modulating

cooperative behavior. Pairwise correlations of gene expression associated with testosterone

phenotype aligned with neuroanatomical similarity (Figure 3H) possibly reflecting similarities in

pathways related to sex-steroid signaling (Figure 2C), but LS/BSTm similarity was the only

neuroanatomical signal in pairwise gene expression associated with strength.

***Testosterone phenotype and social status are associated with systemic gene expression***
We reasoned that testosterone, with its pleiotropic effects and role in social status in
manakins (Ryder et al., 2020) would have systemic impacts on gene expression, resulting in
similar expression patterns across multiple regions. Social status and testosterone phenotype
showed the highest number of consistently differentially expressed genes across tissues (60 and
66 genes with median uncorrected $p < 0.05$ respectively), while social network strength had only
10 consistently differentially expressed genes . Further, PC5 of the whole brain analysis was
significantly associated with social status in a linear mixed model, accounting for batch
confounds and repeated sampling of individuals (Figure S2.5). There was overlap between genes
and GO categories among the consistently differentially regulated genes with social status and
testosterone phenotype, as well as with genes associated with PC5. But hereon we focus on
genes related to social status for simplicity (see Supplement 6).

The top two consistently differentially regulated genes were upregulation of the immune
system gene CLEC2B in the brains of territorial males (Figure 4A) and downregulation of the
development gene ROBO4 (Figure 4B). Enrichment analysis reflected the functional categories
of these genes, although no significant terms were found after FDR correction (Figure 4C). There
was also evidence for global differential regulation of telomere maintenance genes (Figure 4C),
with downregulation of POLD-1-like (LOC114000574) and upregulation of RAD51 in territorial
males. Further, there was evidence for global regulation of cholesterol biosynthesis (Figure 4C),
and glutathione transferases (Figure S2.6, Table S6.2), both of which are involved in steroid
hormone biosynthesis. These findings of global expression differences among status classes
could be partially explained by the upregulation of growth hormone (GH1) in the pituitary of
territorial males (Figure 4D; $LFC = 0.8$, $p = 3 \times 10^{-5}$, $q = 0.01$), which remained consistent in a
model accounting for the interaction between social status and mean T ($LFC = 0.9$, $p = 6 \times 10^{-5}$, $q$
$= 0.05$).

**Discussion**
Using the most comprehensive transcriptome-level sampling of the SDMN to-date
(Antunes et al., 2021; Bentz, George, et al., 2021; Bentz, Niederhuth, et al., 2021; Kabelik et al.,
2021; Lopes & König, 2020), our findings highlight both modular and integrative neurogenomic
organization underlying individual variation in cooperative behavior. The relative roles of each

are central to the development and expression of any complex trait (Cox, 2020; Ketterson et al.,
2009; Lipshutz et al., 2019).

**Different landscapes of gene expression with each trait reflect modular organization of the**
**neuroendocrine system**
Modularity is reflected in how different brain nuclei, endocrine tissues, and genetic
pathways are associated with the expression of different traits. These neurogenomic landscapes,
characterized by the upregulation of neuroendocrine genes in some brain regions and
downregulation in others, enabling differential sensitivity to testosterone and other signaling
molecules among SDMN nuclei. These neurogenomic states can be likened to the activation of
immediate early genes (IEGs) across the SDMN after a social stimulus (Goodson, 2005;
Newman, 1999). In that 'landscape' model, it is the pattern of IEG expression across multiple
brain nuclei that correlates with behavioral responses, rather than the up or downregulation of
particular genes within a single nucleus. This original model considered this landscape of IEG
expression (and later neuropeptides) in the context of proximate responses to social stimuli. Our
findings extend this model in two ways: by considering transcriptomic variation and how long-
term neurogenomic states underlie individual variation in social behavior.

In the landscape of constitutive gene expression in male wire-tailed manakins, most brain
regions exhibit differences in gene expression but the brain regions with higher numbers of
DEGs or candidate gene expression may be more important for mediating certain traits. Some
brain regions showed associations between gene expression and both social status and
testosterone phenotype (e.g., POM) or only testosterone phenotype (e.g., TnA, testes). This may
be because these regions are involved in testosterone mediated traits irrespective of social status
(O'Connell & Hofmann, 2012). Meanwhile, others appeared to be more related to social status
and status specific relationships with testosterone phenotype (e.g., PVN, GCt), or an individual's
strength of cooperative behavior (e.g., PIT, LS). Across brain regions, there were variations in
the identity and function of genes, but certain functions consistently emerged, including neural
development, synaptic potentiation or organization, and genes related to cellular metabolism or
transcription/translation regulation. These results suggest that individual variation is mediated by
neurogenomic states that encode synaptic modifications and connectivity (mechanism of
memory formation), larger scale shifts in cellular activity, and neural structure (Clayton et al.,

2019; George et al., 2020). These mechanisms may interact with variation in elements of sex-
steroid signaling that we investigated in detail (*sensu "neuroendocrine action potential"* (Clayton
et al., 2019)).

Our findings provide strong evidence of modularity in the expression of steroid-related
genes, particularly in relation to social status, testosterone phenotype, or their interaction. We
found evidence that status-specific regulation of behavior could be modulated directly by
testosterone through AR in multiple nuclei (e.g., POM, PVN, BSTm), which is consistent with
previous findings of AR expression linked to display performance in golden-collared manakins
(B. A. Schlinger et al., 2013). Further, the conversion of testosterone to estradiol by the enzyme
aromatase and subsequent binding to estrogen receptors is a key pathway in the regulation of
testosterone-dependent male behaviors (B. Schlinger & Balthazart, 2013), and we found
evidence for the role of aromatase and estrogen signaling role in multiple nuclei.

The co-upregulation of steroid related genes in the POM of males with high testosterone
phenotype (and territorial males) point to this region's role as a potent steroid regulator. The
POM is involved in regulating male aggression, sexual behavior and parental care (O'Connell &
Hofmann, 2011), and stimulates gonadal testosterone production through gonadotropin releasing
hormone (*GnRH*). Interestingly, we observed increase in GNRH1 transcription, which suggest
enhanced steroid production in the testes. Our finding of co-upregulation of AR and GNRH1
may seem contradictory due to the negative regulatory effects of androgens and AR on *GnRH*
(Brayman et al., 2012). However, these may not interact at the cell population level or may show
different temporal patterns of translation. The activation of POM and its steroid receptor
concentrations have been associated with social status differences and social status ascension
cichlid fish (Maruska et al., 2013), and mice (Lee et al., 2022). In the same cichlid species,
different preoptic cell types, including homologs for PVN, showed higher activation in one of the
two partners in cooperative territorial defense (Weitekamp & Hofmann, 2017). Testosterone can
link the POM to the Hypothalamic Pituitary Adrenal (HPA) axis via the PVN (Williamson et al.,
2010) – another important endocrine regulator of social status (DuVal & Goymann, 2011;
Goymann & Wingfield, 2004; Jones & DuVal, 2021). In our study, the PVN showed status-
specific gene expression many genes, including AR, and could be an important mediator of
status-specific regulation of cooperative display. Thus, close linkages between the PVN and

POM (Williamson et al., 2010), suggest these nuclei could be involved in multiple elements of
manakin behavior depending on cell types, social contexts and timescales investigated.

Our results also highlight the importance of neural VIP expression in modulating multiple
aspects of social behavior (Horton, Michael, et al., 2020; Kingsbury & Wilson, 2016), as this
neuropeptide system exhibited widespread differential expression patterns across all traits
examined. An intriguing finding in our study is the potential involvement of VIP and other
steroid receptors in mediating status-specific responses to testosterone in GCt. Although tyrosine
hydroxylase (TH) neurons in GCt have been associated with courtship behaviors (Ben-Tov et al.,
2021; Goodson et al., 2009), our novel findings point to the association between VIP and
courtship behaviors in the GCt (Kingsbury & Wilson, 2016). GCt coordinates social stimuli and
downstream motor processes that control behavior. For example, in songbirds, GCt links POM to
the song control system and thereby mediates androgen-dependent singing behavior (Haakenson
et al., 2020). Given that cooperative display in wire-tailed manakins requires coordinated
physical (and vocal displays in cooperative *Chiroxiphia* manakins (Trainer et al., 2002)), the
status-specific differential expression of genes for steroid receptors and steroid-sensitive
neuropeptides in GCt suggest that this SDMN region may play a unique role in mediating status-
specific relationships between testosterone and cooperative behavior.

The differential gene expression observed in TnA suggests its role as a hub for regulating
testosterone-mediated traits. TnA, the avian homolog of the medial amygdala (O'Connell &
Hofmann, 2011), integrates sensory information and regulates diverse social behaviors (Raam &
Hong, 2021). Our results may reflect persistent activation of this region, based on the
upregulation of IEGs such as JUN and SYT7 in males with high testosterone phenotype, as well
as differential regulation of genes involved in long-term synaptic potentiation (Clayton, 2000;
Clayton et al., 2019; Marrone et al., 2008; O'Connell et al., 2012). Further, our results implicate
potential estrogenic regulation in this region via upregulation of ESR2 in males with higher
testosterone phenotype. Estrogenic pathways through ESR1 have been shown to regulate status-
specific aggression in birds (Horton et al., 2014; Merritt et al., 2020) and prosocial behavior in
prairie voles (Cushing et al., 2008). Although ESR2 is not well studied in birds, it may function
in the medial amygdala of mammals to regulate social recognition (Lymer et al., 2018), and
sexual regulation (Nakata et al., 2016), and is associated with higher social rank in the forebrains

of cichlid fish (Burmeister et al., 2007). Finally, the fish homolog of TnA appears to mediate

aggression and cooperation in cooperative territory defense in an actor-specific manner

(Weitekamp et al., 2017; Weitekamp & Hofmann, 2017), supporting a role for modulating

cooperation in wire-tailed manakins.

There were distinct patterns of gene expression across sampled tissues in association with

social network strength (time spent interacting with other males), and some of these signals

suggest derivation of cooperative display from aggressive behaviors (Díaz-Muñoz et al., 2014).

Many genes were differentially expressed in AH in association with strength, a region primarily

linked to regulation of aggression through AVP and VIP (Horton, Michael, et al., 2020;

O'Connell & Hofmann, 2011). Although these were not differentially expressed in AH, we

found a weak negative relationship between OXT expression and social network strength in

PVN. This negative relationship is contradictory to oxytocin's role in promoting sociality (as in

humans (Feldman, 2012). The relationship observed in the wire-tailed manakin suggests that

oxytocin inhibits cooperation, further supporting the hypothesis that cooperation is derived from

aggressive or dominance displays in this species. However, nonapeptides (like OXT and AVP)

have complex and sometimes species-specific relationships with affiliative behavior and social

status (Goodson & Thompson, 2010; Lee et al., 2022; Maruska et al., 2022). Thus, further

research is needed to clarify the role of nonapeptides in this species.

LS showed many differentially expressed genes associated with cooperation, and highly

cooperative individuals may show differences in neural structure based on the enriched GO

terms. Patterns of expression in BSTm were similar, but weaker based on fewer statistically

significant DEGs. BSTm has neural projections into LS, and vasopressin (AVP) and VIP

neurons in the BSTm and receptors for these neuropeptides in the LS are associated with

affiliative behavior and gregariousness in birds (Kelly et al., 2011; Kelly & Goodson, 2013;

Kingsbury & Wilson, 2016). This suggests cooperative display is regulated by some of the same

conserved brain regions as other types of affiliative behavior. Those studies focused on the acute

processing of social information in these nuclei. In contrast, our study suggests that constitutive

differences in gene expression and neural structure in these regions may underlie individual

variation in cooperative tendencies, potentially affecting their excitability in response to social

information.

Our results reveal a variety of mechanisms that could contribute to individual variation in
cooperative behavior, and its status-specific modulation by testosterone. We identified multiple
forms of neuroplasticity are involved, through steroid receptor variation, synaptic potentiation,
neural development, as well as cellular activity (Clayton et al., 2019). Indeed, testosterone
manipulations are known to induce changes in neuroanatomy in behaviorally relevant brain
nuclei (Kabelik et al., 2008), suggesting a direct role for testosterone-mediated neural growth and
flexibility in modulating male-male cooperation (Soares et al., 2010). Our results provide
promising avenues for further study on the mechanisms underlying cooperative behavior, such as
examination of neuroanatomical features and the use of hormonal or other pharmaceutical
manipulations and RNAi technologies to examine causal relationships between neural gene
expression and behavior (e.g., (Merritt et al., 2020)).

**Brain-wide expression patterns reflect the role of testosterone as a phenotypic**
**integrator.**

The pleiotropic effects of testosterone are predicted to lead to similarities in gene
expression among tissues, supporting its role as a mediator of phenotypic integration. The extent
of gene expression similarities and differences among tissues may reflect the degree of
integration and independence (or modularity), that can either constrain or facilitate adaptive
evolution (Ketterson et al., 2009; Lipshutz et al., 2019). We identified similarities in testosterone
associated gene expression between brain regions that broadly recapitulated tissue ontology.
Additionally, we identified consistent (systemic) gene expression differences throughout the
brain, predominantly associated with social status and testosterone phenotype. However,
systemic results did not explain as much variation in overall neural gene expression compared
with certain personality traits (Kabelik et al., 2021; Lattin et al., 2021), suggesting that the brain
region-specific patterns may be more important in modulating male wire-tailed manakin
behavior. The association between social status and metabolic, immune, and telomere gene
ontology categories reflect differences in allostatic load among status classes in male-male
cooperative systems (Goymann & Wingfield, 2004; Jones & DuVal, 2021; Vernasco et al.,
2021). The differential global regulation of telomeres may explain the relative stability of
telomere length in territorial wire-tailed manakin males (Vernasco et al., 2021), suggesting
metabolic profiles associated with territoriality. Cooperation itself may also have metabolic
costs, as more cooperative male manakins have been shown to have shorter telomeres (Vernasco

et al., 2021), and social bond strength has been associated with signatures of energy metabolism
and stress (Anderson et al., 2022; Simons et al., 2022).

Our results highlight two additional hormones, prolactin and growth hormone, that may
have a system-wide role in the status-specific modulation of cooperative display behavior.
Transcripts for these hormones were differentially expressed in the pituitary, and may reflect
organism wide metabolic profiles. The anterior pituitary is the production site for both hormones,
and thus gene expression patterns are likely good predictors for circulating levels of those
hormones and thus signal strength. PRL transcript was downregulated in more cooperative
males. Prolactin is best known for its role in promoting parental care behaviors, including in
cooperative breeders (Schoech et al., 1996), but emerging studies suggest other functions such as
inhibiting aggression in cooperative breeders (Gilbert et al., 2022; Medger et al., 2019) and
promoting nest defense behavior (Mohamed et al., 2016). The negative relationship observed
appears contradictory to the role of prolactin in promoting prosocial behavior, and again, may
reflect the origin of these cooperative displays from aggressive interactions (Díaz-Muñoz et al.,
2014). In addition, growth hormone transcript (GH1) showed higher expression in territorial
males. This was surprising because ascension in social rank in the cooperatively displaying
manakins is associated with age (DuVal, 2007; McDonald, 1989; Ryder et al., 2008) and GH1
expression is higher in older territorial males. However, in *Anolis* lizards, stimulation of the
growth hormone axis is associated with increased testosterone that enables phenotypic
divergence between males and females (Cox et al., 2017). In manakins growth-hormone could
thus be involved in testosterone production and have subtle effects on multiple systems enabling
the status-specific regulation of behaviors. Status differences in GH1 expression may provide an
explanation for the consistent patterns of neural development and protein synthesis observed in
our results (Sonksen, 2006). These expression differences could reflect the heightened metabolic
demands placed on territorial males during cooperative displays, involving enhanced muscle
performance and the regulation of oxidative stress (Hu et al., 2019). The neuroethological role of
growth hormone is poorly understood, but it may be linked to stress (Greenwood & Landon,
1966; McCormick et al., 1998), and has been shown to increase aggression (Johansson et al.,
2004; Jönsson et al., 1998; Matte, 1981). While the growth hormone inhibition system (via
somatostatin) is an androgen-independent modulator of aggression and social status in cichlid
fish (Hofmann & Fernald, 2000; Trainor & Hofmann, 2006), and natal androgen treatment

increased somatostatin receptor expression in zebra finches (Bentz, Niederhuth, et al., 2021).
Thus, our results suggest that the growth hormone axis may play a role in the status-specific
regulation of wire-tailed manakin cooperative displays, where increased GH and androgens in
territorial males potentially inhibit cooperation by promoting aggression. Future work on social
behavior should consider the interplay between growth hormone, somatostatin, sex steroids, and
glucocorticoids to gain new insights into their roles in behavioral regulation.

## Conclusion

Our study reveals correlations between gene regulation, social status, cooperation, and
testosterone phenotype across the male wire-tailed manakin's SDMN and HPG axis, involving a
plurality of gene functions, including sex-steroid and neuropeptide signaling. This landscape of
gene expression speaks to the modular nature of behavioral mechanisms (Goodson, 2005;
Lipshutz et al., 2019; Newman, 1999) and highlights the continuum between phenotypic
integration and modularity (Ketterson et al., 2009). Unlike previous studies focusing on limited
candidate genes or specific brain regions, our work is the most comprehensive examination of
patterns across multiple nuclei in the SDMN. While our findings are correlative, they provide
valuable insights into the nuclei, genomic pathways and endocrine mechanisms underlying
understudied male-male cooperative behaviors, and its status-specific regulation in this species.
Although questions remain on the acute mediators of cooperative display behavior in the wire-
tailed manakin, we demonstrate the power and flexibility of a system-wide approach, and our
findings generate hypotheses for further experiments to implicate causality. We also contribute
to the emerging concept that constitutive gene expression landscapes across the brain are
associated with consistent behavioral phenotypes (Antunes et al., 2021; Kabelik et al., 2021;
Lattin et al., 2021). We propose that landscapes of constitutive gene expression, comprising
multiple genes across many tissues and brain regions, form the foundation of consistent inter-
individual differences in behavior, upon which flexible and acute responses to social stimuli are
built.

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**Data Accessibility:** Raw read data are available associated with BioProject PRJNA437157, and

all code used to conduct analyses is available at (doi:10.25573/data.22186516) or

https://github.com/periperipatus/PIFI_brain_transcriptome

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# References

Anderson, J. A., Lea, A. J., Voyles, T. N., Akinyi, M. Y., Nyakundi, R., Ochola, L., Omondi, M., Nyundo, F., Zhang, Y., Campos, F. A., Alberts, S. C., Archie, E. A., & Tung, J. (2022). Distinct gene regulatory signatures of dominance rank and social bond strength in wild baboons. *Philosophical Transactions of the Royal Society B*, *377*(1845). https://doi.org/10.1098/RSTB.2020.0441

Antunes, D. F., Teles, M. C., Zuelling, M., Friesen, C. N., Oliveira, R. F., Aubin-Horth, N., & Taborsky, B. (2021). Early social deprivation shapes neuronal programming of the social decision-making network in a cooperatively breeding fish. *Molecular Ecology*, *30*(16), 4118–4132. https://doi.org/10.1111/MEC.16019

Ben-Tov, M., Duarte, F., & Mooney, R. (2021). A neural hub that coordinates learned and innate courtship behaviors. *BioRxiv*, 2021.09.09.459618. https://doi.org/10.1101/2021.09.09.459618

Bentz, A. B., George, E. M., Wolf, S. E., Rusch, D. B., & Podicheti, R. (2021). Experimental competition induces immediate and lasting effects on the neurogenome in free-living female birds. *Proceedings of the National Academy of Sciences of the USA*, *118*(13), e2016154118. https://doi.org/10.1073/pnas.2016154118/-/DCSupplemental.Published

Bentz, A. B., Niederhuth, C. E., Carruth, L. L., & Navara, K. J. (2021). Prenatal testosterone triggers long-term behavioral changes in male zebra finches: unravelling the neurogenomic mechanisms. *BMC Genomics*, *22*(1), 158. https://doi.org/10.1186/s12864-021-07466-9

Blighe, K., & Lun, A. (2020). *PCAtools: Everything Principal Components Analysis* (R package version 2.0.0).

Brayman, M. J., Pepa, P. A., & Mellon, P. L. (2012). Androgen Receptor Repression of Gonadotropin-Releasing Hormone Gene Transcription via Enhancer 1. *Molecular and Cellular Endocrinology*, *363*(1–2), 92. https://doi.org/10.1016/J.MCE.2012.07.012

Burmeister, S. S., Kailasanath, V., & Fernald, R. D. (2007). Social dominance regulates androen and estrogen receptor gene expression. *Bone*, *51*(1), 164–170. https://doi.org/10.1038/jid.2014.371

Cahill, K., Huo, Z., Tseng, G. C., Logan, R. W., & Seney, M. L. (2018). Improved identification of concordant and discordant gene expression signatures using an updated rank-rank hypergeometric overlap approach. *Scientific Reports*, *8*, 9588. https://doi.org/10.1038/s41598-018-27903-2

Clayton, D. F. (2000). The genomic action potential. *Neurobiology of Learning and Memory*, *74*(3), 185–216. https://doi.org/10.1006/nlme.2000.3967

Clayton, D. F., Anreiter, I., Aristizabal, M., Frankland, P. W., Binder, E. B., & Citri, A. (2019). The role of the genome in experience-dependent plasticity: Extending the analogy of the genomic action potential. *Proceedings of the National Academy of Sciences*, *117*(38), 201820837. https://doi.org/10.1073/pnas.1820837116

Cox, R. M. (2020). Sex steroids as mediators of phenotypic integration, genetic correlations, and evolutionary transitions. *Molecular and Cellular Endocrinology*, *502*(December 2019), 110668. https://doi.org/10.1016/j.mce.2019.110668

Cox, R. M., Cox, C. L., McGlothlin, J. W., Card, D. C., Andrew, A. L., & Castoe, T. A. (2017). Hormonally Mediated Increases in Sex-Biased Gene Expression Accompany the Breakdown of Between-Sex Genetic Correlations in a Sexually Dimorphic Lizard. *Https://Doi.Org/10.1086/690105*, *189*(3), 315–332. https://doi.org/10.1086/690105

Cushing, B. S., Perry, A., Musatov, S., Ogawa, S., & Papademetriou, E. (2008). Estrogen Receptors in the Medial Amygdala Inhibit the Expression of Male Prosocial Behavior. *Journal of Neuroscience*, *28*(41), 10399–10403. https://doi.org/10.1523/JNEUROSCI.1928-08.2008

Dakin, R., & Ryder, T. B. (2018). Dynamic network partnerships and social contagion drive cooperation. *Proceedings of the Royal Society B: Biological Sciences*, *285*(1893), 20181973. https://doi.org/10.1098/rspb.2018.1973

Díaz-Muñoz, S. L., DuVal, E. H., Krakauer, A. H., & Lacey, E. A. (2014). Cooperating to compete: Altruism, sexual selection and causes of male reproductive cooperation. *Animal Behaviour*, *88*, 67–78. https://doi.org/10.1016/j.anbehav.2013.11.008

Dobin, A., Davis, C. A., Schlesinger, F., Drenkow, J., Zaleski, C., Jha, S., Batut, P., Chaisson, M., & Gingeras, T. R. (2013). STAR: ultrafast universal RNA-seq aligner. *Bioinformatics*, *29*, 15–21.

DuVal, E. H. (2007). Adaptive advantages of cooperative courtship for subordinate male lance-tailed manakins. *American Naturalist*, *169*(4), 423–432. https://doi.org/10.1086/512137

DuVal, E. H., & Goymann, W. (2011). Hormonal correlates of social status and courtship display in the cooperatively lekking lance-tailed manakin. *Hormones and Behavior*, *59*(1), 44–50. https://doi.org/10.1016/j.yhbeh.2010.10.004

Feldman, R. (2012). Oxytocin and social affiliation in humans. *Hormones and Behavior*, *61*(3), 380–391. https://doi.org/10.1016/J.YHBEH.2012.01.008

Fusani, L., Donaldson, Z., London, S. E., Fuxjager, M. J., & Schlinger, B. A. (2014). Expression of androgen receptor in the brain of a sub-oscine bird with an elaborate courtship display. *Neuroscience Letters*, *578*, 61–65. https://doi.org/10.1016/j.neulet.2014.06.028

George, J. M., Bell, Z. W., Condliffe, D., Dohrer, K., Abaurrea, T., Spencer, K., Leitão, A., Gahr, M., Hurd, P. J., & Clayton, D. F. (2020). Acute social isolation alters neurogenomic state in songbird forebrain. *Proceedings of the National Academy of Sciences*, *117*(38), 201820841. https://doi.org/10.1073/pnas.1820841116

Gilbert, J. D., Rossiter, S. J., Bennett, N. C., & Faulkes, C. G. (2022). The elusive role of prolactin in the sociality of the naked mole-rat. *Hormones and Behavior*, *143*, 105196. https://doi.org/10.1016/J.YHBEH.2022.105196

Goodson, J. L. (2005). The vertebrate social behaviour network: evolutionary themes and variations. *Hormones and Behavior*, *48*(1), 11–22. https://doi.org/10.1038/mp.2011.182.doi

Goodson, J. L., Kabelik, D., Kelly, A. M., Rinaldi, J., & Klatt, J. D. (2009). Midbrain dopamine neurons reflect affiliation phenotypes in finches and are tightly coupled to courtship. *Proceedings of the National Academy of Sciences*, *106*(21), 8737–8742. https://doi.org/10.1073/PNAS.0811821106

Goodson, J. L., & Kingsbury, M. A. (2013). What's in a name? Considerations of homologies and nomenclature for vertebrate social behavior networks. *Hormones and Behavior*, *64*(1), 103–112. https://doi.org/10.1016/J.YHBEH.2013.05.006

Goodson, J. L., & Thompson, R. R. (2010). Nonapeptide mechanisms of social cognition, behavior and species-specific social systems. *Current Opinion in Neurobiology*, *20*(6), 784–794. https://doi.org/10.1016/J.CONB.2010.08.020

Goymann, W., & Wingfield, J. C. (2004). Allostatic load, social status and stress hormones: the costs of social status matter. *Animal Behaviour*, *67*(3), 591–602. https://doi.org/10.1016/J.ANBEHAV.2003.08.007

Greenwood, F. C., & Landon, J. (1966). Growth hormone secretion in response to stress in man. *Nature*, *210*, 540–541.

Griffith, S. C., Crino, O. L., Andrew, S. C., Nomano, F. Y., Adkins-Regan, E., Alonso-Alvarez, C., Bailey, I. E., Bittner, S. S., Bolton, P. E., Boner, W., Boogert, N., Boucaud, I. C. A., Briga, M., Buchanan, K. L., Caspers, B. A., Cichoń, M., Clayton, D. F., Derégnaucourt, S., Forstmeier, W., … Williams, T. D. (2017). Variation in Reproductive Success Across Captive Populations: Methodological Differences, Potential Biases and Opportunities. *Ethology*, *123*(1), 1–29. https://doi.org/10.1111/eth.12576

Haakenson, C. M., Balthazart, J., & Ball, G. F. (2020). Cognition and Behavior Effects of Inactivation of the Periaqueductal Gray on Song Production in Testosterone-Treated Male Canaries (*Serinus canaria*). *ENeuro*, *7*, ENEURO.0048-20.2020. https://doi.org/10.1523/ENEURO.0048-20.2020

Hofmann, H. A., Beery, A. K., Blumstein, D. T., Couzin, I. D., Earley, R. L., Hayes, L. D., Hurd, P. L., Lacey, E. a., Phelps, S. M., Solomon, N. G., Taborsky, M., Young, L. J., & Rubenstein, D. R. (2014). An evolutionary framework for studying mechanisms of social behavior. *Trends in Ecology and Evolution*, *29*(July 2015), 1–9. https://doi.org/10.1016/j.tree.2014.07.008

Hofmann, H. A., & Fernald, R. D. (2000). Social Status Controls Somatostatin Neuron Size and Growth. *Journal of Neuroscience*, *20*(12), 4740–4744. https://doi.org/10.1523/JNEUROSCI.20-12-04740.2000

Horton, B. M., Hudson, W. H., Ortlund, E. A., Shirk, S., Thomas, J. W., Young, E. R., Zinzow-Kramer, W. M., & Maney, D. L. (2014). Estrogen receptor α polymorphism in a species with alternative behavioral phenotypes. *Proceedings of the National Academy of Sciences of the United States of America*, *111*(4), 1443–1448. https://doi.org/10.1073/pnas.1317165111

Horton, B. M., Michael, C., Mackenzie, P., & Maney, D. L. (2020). Vasoactive intestinal peptide as a mediator of the effects of a supergene on social behavior. *Proceedings of the Royal Society B: Biological Sciences*, *287*, 20200196.

Horton, B. M., Ryder, T. B., Moore, I. T., & Balakrishnan, C. N. (2020). Gene expression in the social behavior network of the wire-tailed manakin (*Pipra filicauda*) brain. *Genes, Brain and Behavior*, *October 2018*, e12560. https://doi.org/10.1111/gbb.12560

Hu, B., Hu, S., Yang, M., Liao, Z., Zhang, D., Luo, Q., Zhang, X., & Li, H. (2019). Growth hormone receptor gene is essential for chicken mitochondrial function in vivo and in vitro. *International Journal of Molecular Sciences*, *20*(7), 1–13. https://doi.org/10.3390/ijms20071608

Johansson, V., Winberg, S., Jönsson, E., Hall, D., & Björnsson, B. (2004). Peripherally administered growth hormone increases brain dopaminergic activity and swimming in rainbow trout. *Hormones and Behavior*, *46*, 436–443.

Jones, B., & DuVal, E. H. (2021). Glucocorticoids correlate with and predict social status in the cooperatively breeding lance-tailed manakin (*Chiroxiphia lanceolata*). *Integrative and Comparative Biology*, *61*, E428-429.

Jönsson, E., Johnsson, J., & Björnsson, B. (1998). Growth hormone increases aggressive behavior in juvenile rainbow trout. *Hormones and Behavior*, *33*, 9–15.

Kabelik, D., Julien, A. R., Ramirez, D., & O'Connell, L. A. (2021). Social boldness correlates with brain gene expression in male green anoles. *Hormones and Behavior*, *133*, 105007. https://doi.org/10.1016/j.yhbeh.2021.105007

Kabelik, D., Weiss, S. L., & Moore, M. C. (2008). Steroid hormones alter neuroanatomy and aggression independently in the tree lizard. *Physiology & Behavior*, *93*(3), 492. https://doi.org/10.1016/J.PHYSBEH.2007.10.008

Kasper, C., Vierbuchen, M., Ernst, U., Fischer, S., Radersma, R., Raulo, A., Cunha-Saraiva, F., Wu, M., Mobley, K. B., & Taborsky, B. (2017). Genetics and developmental biology of cooperation. *Molecular Ecology*, *26*(17), 4364–4377. https://doi.org/10.1111/MEC.14208

Kelly, A. M., & Goodson, J. L. (2013). Behavioral relevance of species-specific vasotocin anatomy in gregarious finches. *Frontiers in Neuroscience*, *7*, 242. https://doi.org/10.3389/FNINS.2013.00242

Kelly, A. M., Kingsbury, M. A., Hoffbuhr, K., Schrock, S. E., Waxman, B., Kabelik, D., Thompson, R. R., & Goodson, J. L. (2011). Vasotocin neurons and septal V1a-like receptors potently modulate songbird flocking and responses to novelty. *Hormones and Behavior*, *60*(1), 12–21. https://doi.org/10.1016/J.YHBEH.2011.01.012

Ketterson, E. D., Atwell, J. W., & McGlothlin, J. W. (2009). Phenotypic integration and independence: Hormones, performance, and response to environmental change. *Integrative and Comparative Biology*, *49*(4), 365–379. https://doi.org/10.1093/icb/icp057

Kingsbury, M. A., & Wilson, L. C. (2016). The Role of VIP in Social Behavior: Neural Hotspots for the Modulation of Affiliation, Aggression, and Parental Care. *Integrative and Comparative Biology*, *56*(6), 1238–1249. https://doi.org/10.1093/ICB/ICW122

Langfelder, P., & Horvath, S. (2008). WGCNA: An R package for weighted correlation network analysis. *BMC Bioinformatics*, *9*, 559. https://doi.org/https://doi.org/10.1186/1471-2105-9-559

Lattin, C. R., Kelly, T. R., Kelly, M. W., & Johnson, K. M. (2021). Constitutive gene expression differs in three brain regions important for cognition in neophobic and non-neophobic house sparrows (*Passer domesticus*). *BioRxiv*, 2021.07.06.451290. https://doi.org/10.1101/2021.07.06.451290

Lee, W., Dwortz, M. F., Milewski, T. M., Champagne, F. A., & Curley, J. P. (2022). Social status mediated variation in hypothalamic transcriptional profiles of male mice. *Hormones and Behavior*, *142*, 105176. https://doi.org/10.1016/J.YHBEH.2022.105176

Liao, Y., Smyth, G. K., & Shi, W. (2014). featureCounts: an efficient general purpose program for assigning sequence reads to genomic features. *Bioinformatics*, *30*(7), 923–930.

Lipshutz, S. E., George, E. M., Bentz, A. B., & Rosvall, K. A. (2019). Evaluating testosterone as a phenotypic integrator: From tissues to individuals to species. *Molecular and Cellular Endocrinology*, *496*(June), 110531. https://doi.org/10.1016/j.mce.2019.110531

Lopes, P. C., & König, B. (2020). Wild mice with different social network sizes vary in brain gene expression. *BMC Genomics*, *21*(1), 1–14. https://doi.org/10.1186/s12864-020-06911-5

Love, M. I., Huber, W., & Anders, S. (2014). Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biology*, *15*, 550.

Loveland, J. L., Giraldo-Deck, L. M., Lank, D. B., Goymann, W., Gahr, M., & Küpper, C. (2021). Functional differences in the hypothalamic-pituitary-gonadal axis are associated with alternative reproductive tactics based on an inversion polymorphism. *Hormones and Behavior*, *127*(November 2020), 104877. https://doi.org/10.1016/j.yhbeh.2020.104877

Lymer, J. M., Sheppard, P. A. S., Kuun, T., Blackman, A., Jani, N., Mahbub, S., & Choleris, E. (2018). Estrogens and their receptors in the medial amygdala rapidly facilitate social recognition in female mice. *Psychoneuroendocrinology*, *89*, 30–38. https://doi.org/10.1016/J.PSYNEUEN.2017.12.021

Marrone, D. F., Schaner, M. J., Mcnaughton, B. L., Worley, P. F., & Barnes, C. A. (2008). Immediate-Early Gene expression at rest recapitulates recent experience. *The Journal of Neuroscience*, *28*, 1030–1033. https://doi.org/10.1523/JNEUROSCI.4235-07.2008

Maruska, K. P., Anselmo, C. M., King, T., Mobley, R. B., Ray, E. J., & Wayne, R. (2022). Endocrine and neuroendocrine regulation of social status in cichlid fishes. *Hormones and Behavior*, *139*, 105110. https://doi.org/10.1016/J.YHBEH.2022.105110

Maruska, K. P., Zhang, A., Neboori, A., & Fernald, R. D. (2013). Social Opportunity Causes Rapid Transcriptional Changes in the Social Behaviour Network of the Brain in an African Cichlid Fish. *Journal of Neuroendocrinology*, *25*(2), 145–157. https://doi.org/10.1111/J.1365-2826.2012.02382.X

Matte, A. C. (1981). Growth hormone and isolation-induced aggression in wild male mice. *Pharmacology, Biochemistry, and Behavior*, *14 Suppl 1*, 85–87. https://doi.org/10.1016/S0091-3057(81)80014-7

McCormick, S. D., Shrimpton, J. M., Carey, J. B., O'Dea, M. F., Sloan, K. E., Moriyama, S., & Björnsson, B. T. (1998). Repeated acute stress reduces growth rate of Atlantic salmon parr and alters plasma levels of growth hormone, insulin-like growth factor I and cortisol.

*Aquaculture*, *168*(1–4), 221–235. https://doi.org/10.1016/S0044-8486(98)00351-2

McDonald, D. B. (1989). Cooperation Under Sexual Selection : Age-Graded Changes in a Lekking Bird. *The American Naturalist*, *134*(5), 709–730.

Medger, K., Bennett, N. C., Ganswindt, S. B., Ganswindt, A., & Hart, D. W. (2019). Changes in prolactin, cortisol and testosterone concentrations during queen succession in a colony of naked mole-rats (Heterocephalus glaber): a case study. *Die Naturwissenschaften*, *106*(5–6). https://doi.org/10.1007/S00114-019-1621-1

Merritt, J. R., Grogan, K. E., Zinzow-kramer, W. M., Sun, D., & Ortlund, E. A. (2020). *A supergene-linked estrogen receptor drives alternative phenotypes in a polymorphic songbird*. 1–8. https://doi.org/10.1073/pnas.2011347117

Mohamed, R. A., Shukry, M., Mousa-balabel, T. M., & Elbassiouny, A. A. (2016). Assessment of Plasma Prolactin and Nest Defense Behaviour During Breeding Cycle of Pigeon ( Columba livia domestica ). *Journal of Environmental & Agricultural Sciences*, *7*(19), 19–22.

Nakata, M., Sano, K., Musatov, S., Yamaguchi, N., Sakamoto, T., & Ogawa, S. (2016). Effects of Prepubertal or Adult Site-Specific Knockdown of Estrogen Receptor β in the Medial Preoptic Area and Medial Amygdala on Social Behaviors in Male Mice. *ENeuro*, *3*(2), 579–588. https://doi.org/10.1523/ENEURO.0155-15.2016

Newman, S. W. (1999). The medial extended amygdala in male reproductive behavior: A node in the mammalian social behavior network. *Annals of the New York Academy of Sciences*, *877*, 242–257. https://doi.org/10.1111/j.1749-6632.1999.tb09271.x

O'Connell, L. A., & Hofmann, H. A. (2011). The Vertebrate mesolimbic reward system and social behavior network: A comparative synthesis. *Journal of Comparative Neurology*, *519*(18), 3599–3639. https://doi.org/10.1002/cne.22735

O'Connell, L. A., & Hofmann, H. A. (2012). Social Status Predicts How Sex Steroid Receptors Regulate Complex Behavior across Levels of Biological Organization. *Endocrinology*, *153*(3), 1341–1351. https://doi.org/10.1210/EN.2011-1663

O'Connell, L. A., Matthews, B. J., & Hofmann, H. A. (2012). Isotocin regulates paternal care in a monogamous cichlid fish. *Hormones and Behavior*, *61*(5), 725–733. https://doi.org/10.1016/J.YHBEH.2012.03.009

Olds, J., & Milner, P. (1954). Positive reinforcement produced by electrical stimulation of septal area and other regions of rat brain. *Journal of Comparative and Physiological Psychology*, *47*(6), 419–427. https://doi.org/10.1037/H0058775

Pinheiro, J., Bates, D., DebRoy, S., & Sarkar, D. (2016). *Linear and Nonlinear Mixed Effects Models. R. package v 3.1-128*.

R Core Team. (2020). *R: A language and environment for statistical computing* (3.5.2). R Foundation for Statistical Computing. https://www.r-project.org/

Raam, T., & Hong, W. (2021). Organization of neural circuits underlying social behavior: a consideration of the medial amygdala. *Current Opinion in Neurobiology*, *68*, 124.

https://doi.org/10.1016/J.CONB.2021.02.008

Ryder, T. B., Blake, J. G., Parker, P. G., & Loiselle, B. A. (2011). The composition, stability, and kinship of reproductive coalitions in a lekking bird. *Behavioral Ecology*, *22*(2), 282–290. https://doi.org/10.1093/beheco/arq213

Ryder, T. B., Dakin, R., Vernasco, B. J., Evans, B. S., Horton, B. M., & Moore, I. T. (2020). Testosterone modulates status-specific patterns of cooperation in a social network. *The American Naturalist*, *195*(1), 82–94. https://doi.org/10.1086/706236

Ryder, T. B., Horton, B. M., & Moore, I. T. (2011). Understanding testosterone variation in a tropical lek-breeding bird. *Biology Letters*, *7*(4), 506–509. https://doi.org/10.1098/rsbl.2010.1219

Ryder, T. B., Horton, B. M., Van Den Tillaart, M., De Dios Morales, J., & Moore, I. T. (2012). Proximity data-loggers increase the quantity and quality of social network data. *Biology Letters*, *8*(6), 917–920. https://doi.org/10.1098/rsbl.2012.0536

Ryder, T. B., McDonald, D. B., Blake, J. G., Parker, P. G., & Loiselle, B. A. (2008). Social networks in the lek-mating wire-tailed manakin (*Pipra filicauda*). *Proceedings of the Royal Society B: Biological Sciences*, *275*(1641), 1367–1374. https://doi.org/10.1098/rspb.2008.0205

Ryder, T. B., Parker, P. G., Blake, J. G., & Loiselle, B. A. (2009). It takes two to tango: Reproductive skew and social correlates of male mating success in a lek-breeding bird. *Proceedings of the Royal Society B: Biological Sciences*, *276*(1666), 2377–2384. https://doi.org/10.1098/rspb.2009.0208

Schlinger, B. A., Barske, J., Day, L., Fusani, L., & Fuxjager, M. J. (2013). Hormones and the neuromuscular control of courtship in the golden-collared manakin (*Manacus vitellinus*). *Frontiers in Neuroendocrinology*, *34*(3), 143. https://doi.org/10.1016/J.YFRNE.2013.04.001

Schlinger, B., & Balthazart, J. (2013). Aromatase and Behavior: Concepts Gained from Studies of Aromatase in the Avian Brain. In *Brain Aromatase, Estrogens, and Behavior*. https://doi.org/10.1093/acprof:oso/9780199841196.003.0010

Schoech, S. J., Mumme L, R., & Wingfield.ast C, J. (1996). Prolactin and helping behaviour in the cooperatively breeding Florida scrub-jay, *Aphelocoma c. coerulescens*. *Animal Behaviour*, *52*(3), 445–456. https://doi.org/10.1006/ANBE.1996.0189

Simons, N. D., Michopoulos, V., Wilson, M., Barreiro, L. B., & Tung, J. (2022). Agonism and grooming behaviour explain social status effects on physiology and gene regulation in rhesus macaques. *Philosophical Transactions of the Royal Society B: Biological Sciences*, *377*(1845). https://doi.org/10.1098/RSTB.2021.0132

Soares, M. C., Bshary, R., Fusani, L., Goymann, W., Hau, M., Hirschenhauser, K., & Oliveira, R. F. (2010). Hormonal mechanisms of cooperative behaviour. *Philosophical Transactions of the Royal Society B: Biological Sciences*, *365*(1553), 2737–2750. https://doi.org/10.1098/rstb.2010.0151

Sonksen, P. (2006). Growth Hormone Replacement Therapy and Life Quality. Future

Perspectives. In F. Nyberg (Ed.), *The Somatotrophic Axis in Brain Function* (pp. 327–333). Elsevier Inc. https://doi.org/10.1016/B978-012088484-1/50028-8

Taborsky, M., & Taborsky, B. (2015). Evolution of genetic and physiological mechanisms of cooperative behaviour. *Current Opinion in Behavioral Sciences*, *6*, 132–138. https://doi.org/10.1016/J.COBEHA.2015.11.001

Trainer, J. M., McDonald, D. B., & Learn, W. A. (2002). The development of coordinated singing in cooperatively displaying long-tailed manakins. *Behavioral Ecology*, *13*(1), 65–69. https://doi.org/10.1093/beheco/13.1.65

Trainor, B. C., & Hofmann, H. A. (2006). Somatostatin Regulates Aggressive Behavior in an African Cichlid Fish. *Endocrinology*, *147*(11), 5119–5125. https://doi.org/10.1210/EN.2006-0511

Vernasco, B. J., Dakin, R., Majer, A. D., Haussmann, M. F., Ryder, T. B., & Moore, I. T. (2021). Longitudinal dynamics and behavioural correlates of telomeres in male wire-tailed manakins. *Functional Ecology*, *35*(2), 450–462. https://doi.org/10.1111/1365-2435.13715

Vernasco, B. J., Horton, B. M., Moore, I. T., & Ryder, T. B. (2020). Reduced cooperative behavior as a cost of high testosterone in a lekking passerine bird. *Behavioral Ecology*, *31*, 401–410. https://doi.org/10.1093/beheco/arz201

Weitekamp, C. A., & Hofmann, H. A. (2017). Neuromolecular correlates of cooperation and conflict during territory defense in a cichlid fish. *Hormones and Behavior*, *89*, 145–156. https://doi.org/10.1016/j.yhbeh.2017.01.001

Weitekamp, C. A., Nguyen, J., & Hofmann, H. A. (2017). Neuromolecular regulation of aggression differs by social role during joint territory defense. *Integrative and Comparative Biology*, *57*(3), 631–639. https://doi.org/10.1093/icb/icx009

Williamson, M., Bingham, B., Gray, M., Innala, L., & Viau, V. (2010). The Medial Preoptic Nucleus Integrates the Central Influences of Testosterone on the Paraventricular Nucleus of the Hypothalamus and Its Extended Circuitries. *The Journal of Neuroscience*, *30*(35), 11762. https://doi.org/10.1523/JNEUROSCI.2852-10.2010

Yu, G., Wang, L.-G., Han, Y., & He, Q.-Y. (2012). clusterProfiler: an R package for comparing biological themes among gene clusters. *OMICS: A Journal of Integrative Biology*, *16*, 284–287. https://doi.org/10.1089/omi.2011.0118

Fig. 1. Testosterone and neuroendocrine tissues are involved in the cooperative behavior of wire-tailed manakins.

A) Males engage in cooperative displays forming complex social networks. B) Example network shows single lek over one week, where node size indicates a male's average social network strength (measure of overall time spent interacting), and edge width represents interaction frequency. Node color denotes floater (red) and territorial (gray) status, and darker colors were included in the study. C) Testosterone phenotype (mean testosterone (corrected)) influences cooperative tendency in territorial and floater males differently. D) Repeated measures of testosterone for 12 individuals (points) from a larger population (density). E) as D but proximity-data logging measures of social network strength.. F) Brain and endocrine tissues sampled for RNAseq. Structures not to scale. Colors represent functional groups (O'Connell & Hofmann, 2011). G & inset) Principal component analysis reveals distinct gene expression profiles among brain regions and endocrine tissues (colors and shapes as F). (Photo: Alice Boyle). Note: sample size N=16 for social status, N=12 for testosterone and network strength.

Fig. 2. Neurogenomic landscape of social status and hormone phenotype.
A) Landscape of gene expression in male wire-tailed manakins with high testosterone phenotype (mean testosterone (corrected)) relative to lower testosterone phenotype. Red bars indicate number genes upregulated in males with higher testosterone phenotype, blue indicates genes downregulated in males with higher testosterone. Darker colors and text indicate number of differentially expressed genes after FDR correction. B) Landscape of gene expression in relation to social status, where red indicates genes upregulated in territorial males. C) Summary of candidate gene analysis and filtering, where colors indicate direction of expression, intensity indicates which trait is associated, and "i" indicates there was an interaction effect, and black boxes indicate result was significant after FDR correction. D) Upregulation of AR in POM of males with higher testosterone, where point colors indicate social status (red = floater; black = territorial. Fitted line from linear regression. Subheader includes results from DESeq2: Log$_2$ Fold Change (LFC), uncorrected p-value (p), and correction for multiple testing (q) and R$^2$ calculated separately. E) Upregulation of AR in BSTm of territorial males. F) Status-specific expression of AR in PVN in relation to testosterone phenotype. Note: Sample sizes vary in each tissue due to quality filtering. Maximum sample size for analysis of social status was N=16, while only 12 individuals were sampled for testosterone and behavior.

Fig. 3. A distinct landscape of gene expression in relation to cooperative behavior.
A) Gene expression landscape in male wire-tailed manakins with higher social network strength. Legend as fig. 2A. B) Summary of candidate gene analysis and filtering, where colors indicate direction of expression in in individuals with higher social network strength. Black boxes indicates significant results after FDR correction. C) Downregulation of prolactin (PRL) in the pituitary of males with higher social network strengths D) Downregulation of oxytocin (OXT) in PVN of males with higher social network strength E) Downregulation of progesterone receptor (PGR) in BSTm of males with higher social network strength. F) RRHO2 analysis shows similar gene expression profiles associated with social network strength in the LS and BSTm, G) while network strength related gene expression similarity is low among other tissues. H) Gene expression similarity among tissues in relation to testosterone phenotype follows the ontogeny of brain regions, I) No correlation between testosterone-related gene expression profiles in LS and BSTm. G,H) Pearson correlation coefficients of gene expression between tissues, represented by cell color and number. Gene expression p-values were $\log_{10}$ transformed and multiplied by direction of expression. Black boxes outline significant tissue correlation clusters based on 1000 bootstrap resamples.

**Fig. 4: Global neural gene expression differences between social classes**

A) Immune gene CLEC2B-like was consistently upregulated in the brains of territorial individuals. B) Developmental gene ROBO4 was consistently downregulated in the brains of territorial males. C) Enriched GO categories among genes that were consistently up- or downregulated between territorial and floater males, where bar color indicates uncorrected p-value. D) Upregulation of growth hormone transcript in the pituitary of territorial males could explain differences between status classes.