TITLE

Alternative designs lead to similar performance when traits and performance vary on different

axes

AUTHORS

Kristen M. Nolting[1,*] and Kent E. Holsinger[2]

[1]Department of Plant Biology, University of Georgia, Athens, GA 30602, USA

[2]Department of Ecology and Evolutionary Biology, University of Connecticut, Storrs, CT 06269,

USA

* For correspondence. Email: kristen.nolting@uga.edu; Phone: 706-542-3732

KEYWORDS

alternative designs, plant functional traits, trait covariation, trait-performance relationships,

functional equivalence

RUNNING TITLE

Alternative designs give similar performance

TYPE OF ARTICLE

Synthesis

WORD COUNTS

Abstract: 172

Main Text: 4,036

Text Boxes: NA

NUMBER OF REFERENCES: 70

NUMBER OF TABLES: 1

NUMBER OF FIGURES: 3

STATEMENT OF AUTHORSHIP

K.M.N. and K.E.H. conceived the study. K.E.H. developed the generalized model with input

from K.M.N. K.M.N. performed all analyses, simulations, and generated all figures with input

from K.E.H. K.M.N. and K.E.H. developed the biological interpretation of the simulated,

mathematical, and empirical results. K.M.N. wrote the first manuscript draft with input from

K.E.H.

DATA AND CODE AVAILABILITY

The data, R scripts, and model fit files underlying all analyses, figures, and results have been

deposited in a ZENODO repository (10.5281/zenodo.8083856).

ABSTRACT

Plants differ from one another in size, architecture, water relations, and resource uptake, and

these differences often lead to differences in performance. Yet within a community species that

differ markedly in these traits often have similar performance. Here we use a simple model to

show that when the major axes of trait covariation do not align with the axis of performance

variation, large differences among species in structural traits may have similar performance, i.e.,

'alternative designs.' We further illustrate this phenomenon using trait and performance data

from co-occurring *Protea* species in the Cape Floristic Region, South Africa. Long-term

coexistence of species within a community requires both similar levels of performance, so that

some species are not excluded by competition, and niche differentiation, so that multiple species

can coexist. Thus, misalignment between the axis of performance variation and the major axes of

trait variation may be common, just as genetic variation may be maintained within a population

when the selection gradient does not align with the major axes of the genetic variance-covariance

matrix.

INTRODUCTION

Hutchinson noted in his seminal paper – *"An Homage to Santa Rosalia"* – that there seem to be

more species on the planet than niches to hold them (Hutchinson 1959). Although a few authors

have argued that trait differences among species do not lead to competitive differences and that

species diversity can be largely accounted for by neutral processes (Bell 2000, 2001; Hubbell

2001, 2005; Chave 2004; Alonso *et al.* 2006), most ecologists hold that niche differentiation is

required for the maintenance of species diversity within communities (Macarthur & Levins 1967;

Chesson 2000; Chase & Leibold 2003; HilleRisLambers *et al.* 2012). Community ecologists

often use trait differences among co-occurring species as indices of niche differentiation

(Schoener 1968; Brown & Lieberman 1973; Cody 1986; Gillespie 2004; Treurnicht *et al.* 2020),

and more recently functional ecologists have identified structural traits, i.e., morphological and

allocation-related traits that link an organism's phenotype with its performance (Violle *et al.*

2007), as important for niche differentiation in diverse community assemblages (Mcgill *et al.*

2006; Cavender-Bares *et al.* 2009; Funk *et al.* 2017).

75   Global surveys of structural trait variation in land plants reveal large differences among

species (Reich *et al.* 1997; Wright *et al.* 2004; Díaz *et al.* 2016), and studies of trait differences

among species within diverse clades such as *Helianthus* demonstrate that the magnitude of

interspecific trait variation within a clade can rival that seen in global surveys (Mason &

Donovan 2015). Even within local plant communities and assemblages traits may differ by

orders of magnitude among species (Wright *et al.* 2004; Baraloto *et al.* 2010; Aiello-Lammens *et*

al. 2017). The variation within communities may even be greater than expected given regional

species pools (Swenson *et al.* 2012). In short, trait variation is abundant at almost any scale, even

among individuals within a species growing at a single site (Messier *et al.* 2010; Umaña &

Swenson 2019; Nolting *et al.* 2021; Taseski *et al.* 2021). In contrast, substantial performance

differences among co-occurring species are less commonly documented (Becker *et al.* 1999;

Worthy *et al.* 2020; Nolting *et al.* 2021). Why do we fail to see large differences in performance

when we see large differences in structural traits that are related to performance?

Nearly twenty years ago Marks and Lechowicz (2006) hinted at a solution to this

paradox. They showed that selection on combinations of traits within a single environment can

produce different functional forms ('alternative designs') with roughly equivalent performance.

Their result suggests that while individual ***trait*** differences among species may reflect

differential resource use (and niche partitioning), ***performance*** differences among species within

a community are likely to be minimal, a conclusion consistent with coexistence theory (Chesson

2000; Clark *et al.* 2007). Marks and Lechowicz (2006) argue that while trait variation in natural

communities is often assumed to reflect species' optimization to microenvironments, it could be

that "…aside from major environmental heterogeneity, alternative designs are a major, if not the

main, cause of functional diversity in nature" (Marks & Lechowicz 2006). In spite of their

provocative suggestion, few studies have investigated this possibility, and the mechanism that

leads to 'alternative designs' has not been described.

Here we demonstrate how a particular pattern of trait covariation among 'alternative

designs' leads to similar performance. First, we use a simple, two trait simulation of performance

variation under different scenarios of trait correlations and trait-performance associations, to

illustrate that 'alternative designs' lead to similar performance only when the major axis of trait

variation is not aligned with the axis of performance variation. We then develop a

straightforward mathematical model showing that this result holds for an arbitrary number of

traits.

We conclude by illustrating the phenomenon in a recently analyzed empirical dataset.

Specifically, we show that multivariate combinations of leaf and wood structural traits differ

markedly among species but lead to similar physiological performance among co-occurring

*Protea s*pecies in the Cape Floristic Region, South Africa (data originally published in Nolting et

al. 2021). Taken together, the model and the empirical results suggest the presence of persistent

'alternative designs' (i.e., large variation among species in individual structural traits that

converge on similar performance, (Marks & Lechowicz 2006; Marks 2007)) within natural

communities could be the result of processes that minimize performance differences among

species. If 'alternative designs' are common, these results have important implications for

understanding the ecological and evolutionary mechanisms contributing to persistence of

species-diverse assemblages world-wide.

EXPLORING ALTERNATIVE DESIGNS WITH A SIMPLE MODEL

We begin by developing our intuition with a straightforward model in which individual

performance is related to two traits. These traits may be either positively correlated, negatively

correlated, or uncorrelated, and the axis along which performance varies may be either aligned or

misaligned with the major axis of trait variation.

Simulating Variance in Performance in a Two-Trait Model

129 Assume that a single performance trait is linearly predicted by two structural traits:

$$y_i = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2} + \varepsilon_i \, ,$$

133 where y_i is the performance of individual i, x_{i1} and x_{i2} are the traits of individual i, and ϵ_i is the

134 residual error. β_1 and β_2 reflect the associations between traits x_1 and x_2 with y, and β_0 is the

135 intercept. For simplicity, we will assume that x_1 and x_2 are centered and scaled to have a mean of

136 zero and standard deviation of 1. Traits x_1 and x_2 may be correlated, and the correlation

137 coefficient between them is given by ρ.

138  For the positive trait correlation simulations, we assume $\rho = 0.8$ (i.e., a strong positive

139 trait correlation), and we explore two functional axis scenarios: one in which the regression

140 coefficients (β_1 and β_2) describing the relationship between our two traits and performance are

141 both positive and of the same magnitude, i.e., the axis of performance variation is aligned with

142 the major axis of trait variation, and a second scenario in which the regression coefficients are of

143 the same magnitude but opposite in sign, i.e., the axis of performance variation is orthogonal to

144 the major axis of trait variation (Figure 1, 'Positive Trait Correlation' Box):

146     **Scenario 1:** $\beta_1 = 0.5, \beta_2 = 0.5.$

147     **Scenario 2:** $\beta_1 = 0.5, \beta_2 = -0.5.$

We also assume $\beta_0 = 0$ (i.e., the expected intercept is zero) in our models under each scenario,

which is a reasonable approximation if we centered and scaled the response variable as well as

the covariates. Finally, we display the outcomes under these two scenarios, by simulating trait

data under each scenario for 1000 individuals, sampled from a multivariate normal distribution.

The performance value, y_i, for each individual in each scenario was determined from the

simulated trait data and the linear model described for each scenario, assuming a residual

variance of 0.02. Figure 1c ('Positive Trait Correlation' box) reflects the outcome of a single

realization of this process, under our two scenarios (scenario 1 above, scenario 2 below). We also

report the variance in our performance trait across 1000 realizations for each scenario, and we

summarize the distribution of reported variance across realizations in Figure 1d (scenario 1

above, scenario 2 below). For the negative correlation $(\rho = -0.8)$ and no correlation $(\rho = 0)$

simulations we use the same two functional axis scenarios.

Panel c of Figure 1 shows the distribution of simulated trait data for our three different

patterns of trait correlation: no correlation (the top box), positive correlation (the middle box),

and negative correlation (the bottom box). Within panel c (Figure 1) for each correlation

structure, we show the results from one realization of the trait simulation. The color of each point

corresponds to the performance predicted for that individual from the linear regression

depending on the scenario chosen, scenario 1 in the top plot and scenario 2 in the bottom plot.

The magenta dashed line in each plot represents the regression vector associated with

performance in each scenario (what we refer to as the 'functional axis' of trait variation), and the

gray arrows are the principal components associated with the trait data (with PC1 reflecting the

main axis of trait variation and PC2 reflecting the minor axis of trait variation). Panel d of Figure

1 shows the distribution of performance variation from 1000 independent simulations of each

scenario and each pattern of trait correlation.

The top box in Figure 1 ('No Trait Correlation' box) illustrates that when traits are

uncorrelated, the relationship between the major axis of trait variation and the axis of functional

variation has no impact on the amount of performance variation observed. In contrast, the middle

box ('Positive Trait Correlation' box) and bottom box ('Negative Trait Correlation' box) both

show that when traits are associated with one another the amount of performance variation

observed depends on whether the major axis of trait variation is aligned with the axis of

functional variation. Specifically, whether traits are positively associated (middle box) or

negatively associated (bottom box), there is substantially more performance variation when the

functional axis, i.e., the regression vector, is aligned with the major axis of trait variation, PC 1,

than when it is aligned with the minor axis of trait variation PC 2. In short, whether we find

'alternative designs' depends not on whether traits are negatively correlated – the classic

'tradeoff' example often invoked to explain such a phenomenon (Agrawal 2019) – but on ***both***

the pattern of trait correlation ***and*** the pattern of functional variation.

A GENERAL MODEL FOR ALTERNATIVE DESIGNS

Here we provide a straightforward multivariate extension of the two-trait model we just

illustrated:

$$193 \qquad y_i = \beta_0 + \sum_{k=1}^{K} \beta_k x_{ik} + \varepsilon_i$$

$$194 \qquad = \beta_0 + x_i^T \beta \qquad ,$$

where y_i is the observed physiological performance of individual i, β_0 is the intercept, x_{ik} is the

trait value for trait k in individual i, β_k is the regression coefficient for trait k, and ϵ_i is the

residual error (with mean 0 and variance σ^2). x_i is the vector of traits in individual i and

$\beta = (\beta_1,...,\beta_k)$, $k = 1,...,K$ is the regression vector. We assume that traits are drawn

independently from a multivariate distribution with mean μ and covariance Σ_W. We will take

expectations with respect to the distribution of x_{ik}.

The variance of y is

$$203 \qquad Var(y_i) = E(y_i - \mu)^2$$

$$204 \qquad = E\left((\beta_0 + x_i^T \beta + \epsilon_i) - (\beta_0 + \mu^T \beta) \right)^2$$

$$205 \qquad = E(((x_i - \mu)^T \beta)^2 + x_i^T \beta \epsilon_i + \epsilon_i^2)$$

$$206 \qquad = \beta^T \Sigma \beta + \sigma^2$$

208   Using the spectral decomposition theorem (Szabo 2015) we can rewrite Σ as

$$210 \qquad \Sigma = \lambda_1 u_1 u_1^T + \cdots + \lambda_K u_K u_K^T$$

212   where the eigenvalues λ_i are numbered from the largest to the smallest, and the corresponding

213   eigenvalues have unit length. This allows us to rewrite $Var(y_i)$ as

$$Var(y_i) = \beta^T (\lambda_1 u_1 u_1^T + \cdots + \lambda_K u_K u_K^T)\beta + \sigma^2$$

217   Suppose that the vector of regression coefficients is parallel with the ith principal component. It

218   is, by definition, orthogonal to all of the remaining components, and $Var(y_i)$ reduces to

$$Var(y_i) = \lambda_i \beta^T u_i u_i^T \beta + \sigma^2$$

If the ith principal component is one of the minor axes of trait variation, the variation in

performance will be much smaller than if it is parallel to one of the major axes of trait variation.

More generally, if the regression vector, i.e., the functional axis along which individuals differ in

performance, is mostly orthogonal to the major axes of trait variation, there will be little

variation in performance.

EXAMPLE: PROTEAS WITH DIFFERENT TRAITS HAVE SIMILAR PERFORMANCE

Alignment of the Functional Axis with Axes of Trait Variation

The models above show that 'alternative designs' may arise when the major axes of trait

variation do not align with the axis of functional variation. To determine whether we can detect

this phenomenon in empirical data, we re-examine a subset of a previously published dataset that

measured many structural traits (e.g., leaf area, wood density, stomatal size) and several aspects

of physiological performance (e.g., photosynthetic rate, stem hydraulic conductance) on naturally

co-occurring *Protea* species in the Cape Floristic Region in southwestern South Africa (Nolting

et al. 2021; see supplement for additional information about the species included and trait

sampling). Nolting et al. (2021) reported that (a) combinations of structural trait values explain

much of the variance in physiological performance traits (here our full model R^2 values range

from 0.26 to 0.44, Table 1; Figure S1), (b) species differ markedly in structural traits (with

estimated among-species variation in structural traits ranging from 54.9 to 95.1%; Figure S2a),

but (c) species largely overlap with respect to physiological performance traits (estimated

among-species variation in performance traits ranging from 3.2 to 31.5%; Table 1; Figure S2b).

See supplemental materials for more information about model fits and variance partitioning.

Using the princomp function in R (R Core Team 2022; version 4.2.1) we performed a

principal components analysis on the centered and scaled structural trait data in our *Protea*

dataset, yielding eight principal components. Then for each of the six physiological performance

measures we evaluated the alignment of the structural trait PC axes with the vector of regression

coefficients describing the association between traits and physiological performance, i.e., the

functional axis. Nolting et al. (2021) included site and species random effects in their analyses of

trait-performance relationships, but we exclude them here to facilitate comparison of the

empirical results with the model expectations developed above.

We quantified the degree of alignment for each performance trait model by calculating

the angle between the vector of regression coefficients for each response and each of the eight

principal components (Figure 2). An angle close to 90 degrees indicates that the functional axis

is almost completely misaligned with that principal component axis. An angle close to zero

degrees indicates that the functional axis is almost completely aligned with that principal

component (Figure 2). While the degree to which principal component axes align with the

functional axis differs among physiological performance traits, for every performance trait the

major axes of trait variation (i.e., PC axes 1 and 2) are generally the axes most misaligned with

the functional axis. Instead, the minor axes of trait variation (often the axes excluded from

analyses that use dimension reduction to estimate functional associations from trait matrices and

performance) are the axes most parallel with the regression vector reflecting the functional axis.

We see little differentiation in physiological performance among *Protea* species despite

considerable trait differences because the axis along which trait differences are associated with

differences in physiological performance is largely orthogonal to the axes that account for most

structural trait variation. An important implication of this result is that reducing trait variation to

the first few PCs and attempting to associate those with functional differences may fail to find

real associations of traits with functional differences.

Different Structural Trait Combinations Lead to Similar Performance within and across Species

Our results show that when the axis of performance variation does not align with the major axes

of trait variation, different trait combinations may lead to similar measures of performance.

Moreover, these 'alternative designs' (Marks & Lechowicz 2006; Marks 2007) might exist not

only across taxa but also *within* species that demonstrate substantial trait variation.

To illustrate how combinations of structural traits can lead to similar performance both

within and across species in our dataset, we plot our sampled *Protea* individuals in multivariate

structural trait space, but color the individuals based on their relative performance values. Each

point in each panel in the top row of Figure 3 (Figure 3a,b,c: using the first two PC axes) reflect

the position of individuals in the first two dimensions of structural trait space, i.e., the two most

important axes of structural trait variation. In each panel the points are in the same position, but

their coloring reflects the relative performance of each individual for one of three physiological

performance traits. For example, in Figure 3a the ***position*** of each sampled individual is

determined by its position in the structural trait PCA, but the ***coloring*** of each individual is based

on the quintile of K_s (sapwood-specific hydraulic conductivity) into which it falls. Confidence

ellipses in this top panel of Figure 3 (Figure 3a,b,c) delimit the traits within a 95 percent

confidence interval of each species mean. Notice that in the part of trait space accounting for

most variation, species are well differentiated from one another. In the bottom panel of Figure 3

(Figure 3d,e,f), we color the points in the same way but plot individuals in structural trait space

using the PCs that are most aligned with performance (i.e., the two PCs that were closest to

parallel with the regression vector, from Figure 2). Notice that in the part of trait space most

closely aligned with performance, species boundaries overlap broadly.

Closer examination of Figure 3 also shows both that individuals with different

combinations of structural traits (i.e., different positions in PCA space) can have similar

performance (i.e., fall into the same performance quintile - the same color) and that many

combinations of traits lead to performance in the highest (or lowest) quintile. In addition, not

only may individuals within species fall into different performance quintiles, but individuals

belonging to different species may fall into the same performance quintile. In fact, all species

(other than *P. nitida*) have some individuals with sapwood-specific hydraulic conductivity, K_s, in

the highest performance quintile and others in the lowest performance quintile. The same pattern

is repeated across all six physiological performance traits measured (Figure S3).

DISCUSSION

Given the well-established relationships between plant traits and plant performance, the

magnitude of species differences in performance is often much less than might be expected. We

demonstrate that this outcome is expected when the axis of functional variation does not align

with the major axes of trait variation. Using a recently published dataset of plant structural traits

and physiological performance traits measured in several natural populations of *Protea* from the

Cape Floristic Region, South Africa (Nolting et al. 2021), we show that large among-species

differences in structural traits related to plant performance do not lead to noticeable differences

in physiological performance. Consistent with our mathematical model, we show that the axis of

observed functional variation is not aligned with the major axes of observed trait variation. These

results illustrate that a full understanding of multivariate trait associations requires not only that

we evaluate axes of trait variation, but also that we determine whether major axes of trait

variation align with axes of performance variation.

Implications for Trait-Mediated Species Coexistence and Community Ecology

Plants or animals that live in the same place typically experience roughly the same

macroenvironment. As a result, we might expect them to have similar traits (Keddy 1992), but

basic principles of community ecology suggest that species can coexist long term only if they

differ along at least one niche axis (Chesson 2000; HilleRisLambers *et al.* 2012). To put it

another way, environmental filtering may explain an association between community-weighted

mean trait values and environments (Ackerly & Cornwell 2007) and phylogenetic

underdispersion in local community assemblages (Webb 2000; Webb *et al.* 2002; Mayfield &

Levine 2010), but within-community trait differences may promote coexistence (Cavender-Bares

et al. 2004a, b; Kraft *et al.* 2008; Swenson & Enquist 2009).

Marks and Lechowicz (2006) illustrated that 'alternative designs' may lead to roughly

equivalent functional performance and may resolve this apparent contradiction. As we show in

greater detail, many different trait combinations can produce similar performance. This suggests

trait differences promote coexistence through niche differentiation, but if the axis of functional

diversity is largely orthogonal to the axis of trait differentiation then performance differences are

minimized, an observation that has been observed in previous empirical systems (Adler *et al.*

2007; Umaña *et al.* 2016). Given that many structural traits not only influence differences in any

one performance trait, but that any one structural trait influences differences in many

performance traits (Worthy *et al.* 2020; Funk *et al.* 2021), the trait space where performance is

roughly equivalent may be extremely large (Marks & Lechowicz 2006; Laughlin 2014; Laughlin

& Messier 2015).

Should 'Alternative Designs' be Common?

Although we provide only one empirical example, we suspect that 'alternative designs' are

common in natural systems. Here we focused on structural trait and physiological performance

trait variation at the level of individual plants and plant communities, but similar phenomena

arise in quantitative genetics. It has long been known that in spite of strong stabilizing selection,

individual traits under selection can retain substantial genetic variation (Arnold 1981; Falconer

1981; Mousseau & Roff 1987). This putative inconsistency can be understood by considering the

complex interaction of selection operating on many covarying traits (Walsh & Blows 2009).

Lande and Arnold (1983) showed how to use the multivariate breeder's equation, $\Delta \mathbf{z} = \mathbf{G}\boldsymbol{\beta}$, to

identify the selection gradient on traits in a population. The selection gradient, $\boldsymbol{\beta}$, is the set of

regression coefficients relating trait values to fitness – the direct analog of the functional axis of

trait variation we have been discussing. Just as we demonstrated above, Walsh and Blows (2009)

show that if the selection gradient is not aligned with major axes of the G matrix (**G**, the genetic

variance-covariance matrix), then there may be substantial genetic variation for traits that are

under stabilizing selection.

Far from being an exception, misalignment between multivariate axes of genetic variation

and the selection gradient may actually be expected. It is often thought that the direction of

population divergence will correspond with 'genetic lines of least resistance', i.e., divergence

along the major axes of genetic variation (Schluter 1996), and there have been many examples of

this phenomenon presented (Mitchell-Olds 1996; Chenoweth *et al.* 2010). Recent work in *Anolis*

lizards suggests that the alignment between major axes of genetic variation and axes of

divergence can be maintained over millions of years, and despite evolution of the G matrix itself

(McGlothlin *et al.* 2018). Nonetheless, there are many other examples in which divergence

seems to be constrained due to misalignment between the major axes of genetic variation and the

selection gradient either within or among populations (Etterson & Shaw 2001; Henry &

Stinchcombe 2023). Taken together these results indicate that an evolutionary response might be

limited not only if a population lacks genetic variation, but also if the selection gradient is not

aligned with major axes of the G matrix (Donovan *et al.* 2011; Agrawal 2019). Interestingly,

Engen and Saether (2021) suggest that the evolutionary process itself may lead to misalignment

of the selection gradient and major axes of the G matrix. Specifically, they show that at an

evolutionary equilibrium the G matrix itself evolves so that the selection gradient is parallel to

the eigenvector associated with the smallest eigenvalue, i.e., the axis of the G matrix associated

with the least variation. We conjecture that the process of interspecific competition will lead to a

similar misalignment between the axis of functional variation and the major axes of structural

trait variation in natural communities.

CONCLUSION

Functional ecologists have often focused on broad-scale geographic patterns in which trait values

are seen as characteristic of, and perhaps even diagnostic of, particular environments (Diaz *et al.*

1998; Cavender-Bares *et al.* 2004b; Cornwell & Ackerly 2009; Anderegg *et al.* 2021).

Community ecologists, on the other hand, have typically focused on trait differences as key

features that allow long-term coexistence (Lack 1947; Cody 1986; Silvertown 2004). When these

perspectives are combined, we find that there are often large differences between species in

structural traits but that performance differences among co-occurring species are minimal, even

when those structural traits are related to performance. Marks and Lechowicz (2006)

demonstrated that 'alternative designs' can evolve within a homogenous environment, and they

suggested that 'alternative designs' may be a major cause of among-species trait variation

observed in plant communities (Marks and Lechowicz, 2006). Here we show how this pattern

can emerge. When the major axes of trait variation are largely orthogonal to the functional axis

describing the association between traits and performance, there will be little variation in

functional performance. Because long-term co-existence among species requires similar levels of

performance within a given habitat, we expect the axis of functional performance will often be

orthogonal to major axes of trait variation.

The pattern we describe is directly analogous to phenomena that are well described in

quantitative genetics. Just as the response to selection is largely the outcome of the alignment

between major axes of the G matrix and the selection gradient, so the extent of performance

variation across species depends on whether the major axes of trait variation are aligned with the

axis of functional variation. While it is tempting to describe 'alternative designs' as an example

of 'trade-offs', i.e., negative covariance between two traits important for performance, we show

that 'alternative designs' can arise even when traits positively covary. What matters is not

whether traits covary positively or negatively, but whether the axis of trait variation is aligned

with the axis of functional variation.

ACKNOWLEDGEMENTS

We are very grateful to R. Bagchi who initially gave us the idea to explore this question with

respect to trait variances and covariances. We also thank C. Jones, R. Bagchi, and members of

the Donovan and Burke Labs for very helpful feedback on earlier versions of this manuscript.

This work was supported in part by the National Science Foundation (DEB-1046328).

REFERENCES

Ackerly, D.D. & Cornwell, W.K. (2007). A trait-based approach to community assembly:

partitioning of species trait values into within- and among-community components. *Ecol.*

Lett., 10, 135–145.

Adler, P.B., Hille Ris Lambers, J. & Levine, J.M. (2007). A niche for neutrality. *Ecol. Lett.*, 10,

95–104.

Agrawal, A.A. (2019). A scale-dependent framework for trade-offs, syndromes, and

specialization in organismal biology. *Ecology*, e02924.

Aiello-Lammens, M.E., Slingsby, J.A., Merow, C., Mollmann, H.K., Euston-Brown, D., Jones,

C.S., *et al.* (2017). Processes of community assembly in an environmentally

heterogeneous, high biodiversity region. *Ecography* , 40, 561–576.

Alonso, D., Etienne, R.S. & McKane, A.J. (2006). The merits of neutral theory. *Trends Ecol.*

Evol., 21, 451–457.

Anderegg, L.D.L., Loy, X., Markham, I.P., Elmer, C.M., Hovenden, M.J., Hille Ris Lambers, J.,

et al. (2021). Aridity drives coordinated trait shifts but not decreased trait variance across

the geographic range of eight Australian trees. *New Phytol.*, 229, 1375–1387.

Arnold, S.J. (1981). Behavioral Variation in Natural Populations. I. Phenotypic, Genetic and

Environmental Correlations Between Chemoreceptive Responses to Prey in the Garter

Snake, Thamnophis elegans. *Evolution*, 35, 489–509.

Baraloto, C., Timothy Paine, C.E., Patiño, S., Bonal, D., Hérault, B. & Chave, J. (2010).

Functional trait variation and sampling strategies in species-rich plant communities.

Funct. Ecol., 24, 208–216.

Becker, P., Tyree, M.T. & Tsuda, M. (1999). Hydraulic conductances of angiosperms versus

conifers: similar transport sufficiency at the whole-plant level. *Tree Physiol.*, 19, 445–

452.

Bell, G. (2000). The Distribution of Abundance in Neutral Communities. *Am. Nat.*, 155, 606–

617.

Bell, G. (2001). Neutral macroecology. *Science*, 293, 2413–2418.

Brown, J.H. & Lieberman, G.A. (1973). Resource Utilization and Coexistence of Seed-Eating

Desert Rodents in Sand Dune Habitats. *Ecology*, 54, 788–797.

Cavender-Bares, J., Ackerly, D.D., Baum, D.A. & Bazzaz, F.A. (2004a). Phylogenetic

overdispersion in Floridian oak communities. *Am. Nat.*, 163, 823–843.

Cavender-Bares, J., Kitajima, K. & Bazzaz, F.A. (2004b). Multiple trait associations in relation

to habitat differentiation among 17 Floridian Oak species. *Ecological Monographs*, 74,

635–662.

Cavender-Bares, J., Kozak, K.H., Fine, P.V.A. & Kembel, S.W. (2009). The merging of

community ecology and phylogenetic biology. *Ecol. Lett.*, 12, 693–715.

Chase, J.M. & Leibold, M.A. (2003). *Ecological Niches: Linking Classical and Contemporary*

Approaches. University of Chicago Press, Chicago and London.

Chave, J. (2004). Neutral theory and community ecology. *Ecol. Lett.*, 7, 241–253.

Chenoweth, S.F., Rundle, H.D. & Blows, M.W. (2010). The contribution of selection and genetic

constraints to phenotypic divergence. *Am. Nat.*, 175, 186–196.

Chesson, P. (2000). Mechanisms of Maintenance of Species Diversity. *Annu. Rev. Ecol. Syst.*,

31, 343–366.

Clark, J.S., Dietze, M., Chakraborty, S., Agarwal, P.K., Ibanez, I., LaDeau, S., *et al.* (2007).

Resolving the biodiversity paradox. *Ecol. Lett.*, 10, 647–59; discussion 659-62.

Cody, M.L. (1986). Structural niches in plant communities. In: *Community ecology*. Harper &

Row, New York, NY, pp. 381–405.

Cornwell, W.K. & Ackerly, D.D. (2009). Community assembly and shifts in plant trait

distributions across an environmental gradient in coastal California. *Ecological*

Monographs, 79, 109–126.

Díaz, S., Cabido, M. & Casanoves, F. (1998). Plant functional traits and environmental filters at

a regional scale. *J. Veg. Sci.*, 9, 113–122.

Díaz, S., Kattge, J., Cornelissen, J.H.C., Wright, I.J., Lavorel, S., Dray, S., *et al.* (2016). The

global spectrum of plant form and function. *Nature*, 529, 167–171.

Donovan, L.A., Maherali, H., Caruso, C.M., Huber, H. & de Kroon, H. (2011). The evolution of

the worldwide leaf economics spectrum. *Trends Ecol. Evol.*, 26, 88–95.

Engen, S. & Sæther, B.-E. (2021). Structure of the G-matrix in relation to phenotypic

contributions to fitness. *Theor. Popul. Biol.*, 138, 43–56.

Etterson, J.R. & Shaw, R.G. (2001). Constraint to adaptive evolution in response to global

warming. *Science*, 294, 151–154.

Falconer, D.S. (1981). *Introduction to Quantitative Genetics. 2 Ed*. Longman, London.

Funk, J.L., Larson, J.E., Ames, G.M., Butterfield, B.J., Cavender-Bares, J., Firn, J., *et al.* (2017).

Revisiting the Holy Grail: using plant functional traits to understand ecological processes.

Biol. Rev. Camb. Philos. Soc., 92, 1156–1173.

Funk, J.L., Larson, J.E. & Ricks-Oddie, J. (2021). Plant traits are differentially linked to

performance in a semiarid ecosystem. *Ecology*, 102, e03318.

Gillespie, R. (2004). Community Assembly Through Adaptive Radiationin Hawaiian Spiders.

Science, 303.

Henry, G.A. & Stinchcombe, J.R. (2023). G-matrix stability in clinally diverging populations of

an annual weed. *Evolution*, 77, 49–62.

HilleRisLambers, J., Adler, P.B., Harpole, W.S., Levine, J.M. & Mayfield, M.M. (2012).

Rethinking Community Assembly through the Lens of Coexistence Theory. *Annu. Rev.*

Ecol. Evol. Syst., 43, 227–248.

Hubbell, S.P. (2001). *The Unified Neutral Theory of Biodiversity and Biogeography*. Princeton

University Press, Princeton, NJ.

Hubbell, S.P. (2005). Neutral theory in community ecology and the hypothesis of functional

equivalence. *Functional Ecology*.

Hutchinson, G.E. (1959). Homage to Santa Rosalia or Why Are There So Many Kinds of

Animals? *Am. Nat.*, 93, 145–159.

Keddy, P.A. (1992). Assembly and Response Rules: Two Goals for Predictive Community

Ecology. *J. Veg. Sci.*, 3, 157–164.

Kraft, N.J.B., Valencia, R. & Ackerly, D.D. (2008). Functional Traits and Niche-Based Tree

Community Assembly in an Amazonian Forest. *Science*, 322, 580–582.

Lack, D. (1947). *Darwin's Finches*. Cambridge University Press, Cambridge.

Lande, R. & Arnold, S.J. (1983). The measurement of selection on correlated characters.

Evolution, 37, 1210–1226.

Laughlin, D.C. (2014). The intrinsic dimensionality of plant traits and its relevance to

community assembly. *J. Ecol.*, 102, 186–193.

Laughlin, D.C. & Messier, J. (2015). Fitness of multidimensional phenotypes in dynamic

adaptive landscapes. *Trends Ecol. Evol.*, 30, 487–496.

Macarthur, R. & Levins, R. (1967). The Limiting Similarity, Convergence, and Divergence of

Coexisting Species. *Am. Nat.*, 101, 377–385.

Marks, C.O. (2007). The causes of variation in tree seedling traits: the roles of environmental

selection versus chance. *Evolution*, 61, 455–469.

Marks, C.O. & Lechowicz, M.J. (2006). Alternative designs and the evolution of functional

diversity. *Am. Nat.*, 167, 55–66.

Mason, C.M. & Donovan, L.A. (2015). Evolution of the leaf economics spectrum in herbs:

Evidence from environmental divergences in leaf physiology across Helianthus

(Asteraceae). *Evolution*, 69, 2705–2720.

Mayfield, M.M. & Levine, J.M. (2010). Opposing effects of competitive exclusion on the

phylogenetic structure of communities. *Ecol. Lett.*, 13, 1085–1093.

Mcgill, B., Enquist, B., Weiher, E. & Westoby, M. (2006). Rebuilding community ecology from

functional traits. *Trends Ecol. Evol.*, 21, 178–185.

McGlothlin, J.W., Kobiela, M.E., Wright, H.V., Mahler, D.L., Kolbe, J.J., Losos, J.B., *et al.*

(2018). Adaptive radiation along a deeply conserved genetic line of least resistance in

Anolis lizards. *Evol Lett*, 2, 310–322.

Messier, J., McGill, B.J. & Lechowicz, M.J. (2010). How do traits vary across ecological scales?

A case for trait-based ecology. *Ecol. Lett.*, 13, 838–848.

Mitchell-Olds, T. (1996). Pleiotropy causes long-term genetic constraints on life-history

evolution in Brassica rapa. *Evolution*, 50, 1849–1858.

Mousseau, T.A. & Roff, D.A. (1987). Natural selection and the heritability of fitness

components. *Heredity* , 59 (Pt 2), 181–197.

Nolting, K.M., Prunier, R., Midgley, G.F. & Holsinger, K.E. (2021). Intraspecific trait variation

influences physiological performance and fitness in the South Africa shrub genus Protea

(Proteaceae). *Ann. Bot.*, 127, 519–531.

Reich, P.B., Walters, M.B. & Ellsworth, D.S. (1997). From tropics to tundra: global convergence

in plant functioning. *Proc. Natl. Acad. Sci. U. S. A.*, 94, 13730–13734.

Schluter, D. (1996). Adaptive radiation along genetic lines of least resistance. *Evolution*, 50,

1766–1774.

Schoener, T.W. (1968). The Anolis Lizards of Bimini: Resource Partitioning in a Complex

Fauna. *Ecology*, 49, 704–726.

Silvertown, J. (2004). Plant coexistence and the niche. *Trends Ecol. Evol.*, 19, 605–611.

Swenson, N.G. & Enquist, B.J. (2009). Opposing assembly mechanisms in a neotropical dry

forest: implications for phylogenetic and functional community ecology. *Ecology*, 90,

2161–2170.

Swenson, N.G., Enquist, B.J., Pither, J., Kerkhoff, A.J., Boyle, B., Weiser, M.D., *et al.* (2012).

The biogeography and filtering of woody plant functional diversity in North and South

America. *Glob. Ecol. Biogeogr.*, 21, 798–808.

Szabo, F.E. (2015). *The linear algebra survival guide*. Academic Press, New York.

Taseski, G.M., Keith, D.A., Dalrymple, R.L. & Cornwell, W.K. (2021). Shifts in fine root traits

within and among species along a fine-scale hydrological gradient. *Ann. Bot.*, 127, 473–

481.

Treurnicht, M., Pagel, J., Tonnabel, J., Esler, K.J., Slingsby, J.A. & Schurr, F.M. (2020).

Functional traits explain the Hutchinsonian niches of plant species. *Glob. Ecol.*

Biogeogr., 29, 534–545.

Umaña, M.N., Forero-Montaña, J., Muscarella, R., Nytch, C.J., Thompson, J., Uriarte, M., *et al.*

(2016). Interspecific Functional Convergence and Divergence and Intraspecific Negative

Density Dependence Underlie the Seed-to-Seedling Transition in Tropical Trees. *Am.*

Nat., 187, 99–109.

Umaña, M.N. & Swenson, N.G. (2019). Does trait variation within broadly distributed species

mirror patterns across species? A case study in Puerto Rico. *Ecology*, 100, e02745.

Violle, C., Navas, M.-L., Vile, D., Kazakou, E., Fortunel, C., Hummel, I., *et al.* (2007). Let the

concept of trait be functional! *Oikos*, 116, 882–892.

Walsh, B. & Blows, M.W. (2009). Abundant Genetic Variation + Strong Selection =

Multivariate Genetic Constraints: A Geometric View of Adaptation. *Annu. Rev. Ecol.*

Evol. Syst., 40, 41–59.

Webb, C.O. (2000). Exploring the Phylogenetic Structure of Ecological Communities: An

Example for Rain Forest Trees. *Am. Nat.*, 156, 145–155.

Webb, C.O., Ackerly, D.D., McPeek, M.A. & Donoghue, M.J. (2002). Phylogenies and

Community Ecology. *Annu. Rev. Ecol. Syst.*, 33, 475–505.

Worthy, S.J., Laughlin, D.C., Zambrano, J., Umaña, M.N., Zhang, C., Lin, L., *et al.* (2020).

Alternative designs and tropical tree seedling growth performance landscapes. *Ecology*,

101, e03007.

Wright, I.J., Reich, P.B., Westoby, M., Ackerly, D.D., Baruch, Z., Bongers, F., *et al.* (2004). The

worldwide leaf economics spectrum. *Nature*, 428, 821–827.

TABLES WITH CAPTIONS

Table 1. Model fits and estimated among-species variance for each physiological performance

response trait explored in the *Protea* dataset. The mean estimated Bayesian R^2 for the full model

and for the fixed effects only (the marginal R^2) are presented for each model, as well as the mean

estimated among species variance for each physiological performance trait, relative to the

residual variance (i.e., within species variance and residual error).

Physiological Performance Measure	Full Model R^2 (Marginal R^2 in parentheses)	Estimated Among Species Variance (%)
Sapwood-Specific Hydraulic Conductivity	0.36 (0.20)	27.2
Leaf-Specific Conductivity	0.30 (0.22)	26.6
Photosynthetic Rate	0.44 (0.27)	31.5
Leaf-Specific Photosynthetic Rate	0.33 (0.36)	18.0
Stomatal Conductance	0.36 (0.18)	7.3
Instantaneous Water-Use Efficiency	0.26 (0.20)	3.2

FIGURE CAPTIONS

Figure 1. The simple two-trait models and the simulated trait data and distribution of

performance variation. Three types of trait correlations are presented: Top Box, no trait

correlation between the two traits; Middle Box, a positive trait correlation ($\rho = 0.8$); Bottom

Box, a negative trait correlation ($\rho = -0.8$). For each trait correlation, we simulate trait data

under two different scenarios: Scenario 1, in which both traits are positively associated with

performance ($\beta_1 = 0.5$ and $\beta_2 = 0.5$), and Scenario 2, in which one trait is positively associated

with performance ($\beta_1 = 0.5$) and one trait is negatively associated with performance ($\beta_2 =$

-0.5). For each trait correlation pattern and each scenario, we simulate trait data following the

linear models described in the main text. In panel C we present a single realization of the

simulation, where the color of each point reflects the predicted performance given the

combination of trait values and the linear model defined. The black dashed ellipse around each

cloud of points is the 95% confidence interval, the magenta dashed line reflects the functional

axis describing the association between the traits and performance, and the gray arrows represent

the principal components describing the main axis of trait variation (long arrow) and the minor

axis of trait variation (short arrow). We repeated each simulation 1000 times and present the

distribution of performance variances under each scenario and trait correlation in panel D. In

panel D the dashed vertical black lines reflect the 95% cut-offs for each distribution and the solid

black line is the mean variance given 1000 iterations of a simulation.

Figure 2: Alignment of each principal component with the regression vector describing the

functional relationship between structural traits and each of six performance traits in the *Protea*

dataset. (A) Sapwood-specific conductivity, (B) Leaf-specific conductivity, (C) Photosynthetic rate, (D) Leaf-specific photosynthetic rate, (E) Stomatal conductance, and (F) Instantaneous water-use efficiency. The horizontal axis is the angle (0-90°) between each of the eight principal components for the structural trait data measured on 132 *Protea* individuals and the regression vector associated with each physiological performance trait. Angles closer to 90° indicate that the functional axis (i.e., the vector of regression coefficients) and the PC axis are largely orthogonal and angles closer to 0° indicate they are more aligned. The gray histograms reflect the posterior distribution of angles, the black circles represent the mean of the posterior, the thick black lines reflect the 50 percent credible intervals, and the thin black line reflects the 95 percent credible intervals.

Figure 3: Visualizing variation in relative performance arising from different combinations of structural traits, for a subset of performance traits (Sapwood-specific hydraulic conductivity: A and D; Photosynthetic rate: B and E; Stomatal conductance: C and F). For each panel, individuals from our empirical dataset are plotted with respect to their position in multivariate structural trait space. For A, B, and C individuals are plotted using the first two principal components (PC1 and PC2). For D, E, and F individuals are plotted in the structural trait space that is most aligned with performance (i.e., the PC axes that are most parallel with the regression vector for each performance trait, from Figure 2). The color of the points reflects the relative performance of each individual, corresponding to one of five ordered categories (Low, Medium Low, Medium, Medium High, and High). The confidence ellipses correspond to species groupings. PRLA: *P. laurifolia*, PRNI: *P. nitida*, PREX: *P. eximia*, PRPU: *P. punctata*, and PRRE: *P. repens*.

Figure 1.

Figure 2.

PRRE

PRPU

| PC 4 (6.3%) | PC 5 (3.9%) | PC 5 (3.9%) |

Relative Performance   ● Low   ● Medium low   ● Medium   ● Medium high   ● High

32