A meta-analysis of the effect of protein synthesis inhibitors on rodent fear conditioning

Clarissa F. D. Carneiro[1,2], Felippe E. Amorim[1] and Olavo B. Amaral[1]

[1] Institute of Medical Biochemistry Leopoldo de Meis, Federal University of Rio de Janeiro, Rio de Janeiro, Brazil.

[2] Berlin Institute of Health (BIH) at Charité, BIH QUEST Center for Responsible Research, Berlin, Germany.

Abstract

Systematic reviews and meta-analyses have been increasingly recognized for their potential value in pre-clinical research, but their applications have not been extensively explored in behavioral neuroscience. In this work, we studied protein synthesis inhibition, a classic intervention used to disrupt fear learning, reconsolidation, and extinction in rodents, to explore how meta-analyses can identify potential moderators of its effect. We initially performed separate meta-analyses for different injection sites and target sessions to evaluate the effect of the intervention in various scenarios. Heterogeneity was further investigated by multilevel meta-regression models aggregating various sites, with article or research group as additional levels. We detected robust effects of protein synthesis inhibitors on training and reconsolidation, but not on extinction, possibly due to the lower number of studies on the latter. Our analyses identified some well-established moderators, such as intervention timing and reexposure duration. However, other factors proposed as boundary conditions for reconsolidation, such as memory age and training strength, were not associated with effect size. While our results point to the value of meta-analyses in consolidating findings from the literature, we believe that associations suggested by data synthesis should ideally be verified by well-powered, rigorous confirmatory experiments.

Introduction

Over the last decades, biomedical research has grown consistently; however, low reproducibility rates in some areas have shed doubt over its reliability[1–3]. Industry figures for the replicability of basic research have been as low as 11% and 21%[4,5]. In experimental neurology, only 30% of replications in spinal cord injury research were at least partly successful[6]. In basic cancer biology, a 6-year systematic effort reported that 46% of experiments from highly cited articles were reproduced, with effect sizes being 85% smaller in replications than in the original studies[2].

Systematic replication efforts, however, are cost- and labor-intensive endeavors and may not be feasible as a routine procedure in every area of science. Meta-analyses of the existing literature can thus be a useful and accessible tool to assess the reliability of effect estimates, investigate publication bias and explore sources of heterogeneity among studies[7]. This latter point is of particular interest to discovery science: as data synthesis is still rare in this field, conflicting results in the literature are usually not settled properly. Scientists tend to assume protocol discrepancies are responsible for divergences between studies, without considering the possibility that they might be due to random variation around statistical thresholds[8,9].

Methods for synthesizing data from basic biomedical research, however, are not as mature as in clinical research, where outcomes are typically fewer and more clearly defined. A single article in basic discovery science can include experiments exploring multiple questions, multiple methods addressing a single question, or multiple control

experiments testing boundary conditions (i.e. changing protocol parameters in order to establish the necessary conditions for an effect to be observed). This also leads to multiple experiments published in individual articles, as well as multiple articles by the same research groups, violating the assumption of independence between different results when synthesizing data.

While greater attention is present in other areas, meta-analysis of rodent behavioral studies is still incipient. Fear conditioning is a learning paradigm characterized by the association of a neutral conditioned stimulus (CS; usually a context or a tone) to a negative unconditioned stimulus (US; for example, a footshock)[10]. Conditioning protocols vary in many ways, but the outcome used to measure memory is usually the same – the animal's freezing response to the CS in a test session. The paradigm has been widely used to dissect the anatomical and biochemical pathways involved in the acquisition and consolidation of fear memories, as well as to study reconsolidation and extinction phenomena, where the fearful association is changed or replaced by a non-fearful one.

After a series of failed attempts to replicate findings on reconsolidation of fear conditioning in rodents[11,12], Schroyens *et al.* found evidence of publication bias in the field by reviewing the literature and contacting relevant authors[13]. In their sample, 80% of the published findings yielded statistically significant results, in contrast to only 20% of the unpublished findings shared with them[13]. The failed replications also raise the possibility that differences in protocols can determine the magnitude of the outcome. This suggests the need to determine the robustness of the effect of amnestic agents in different conditions, as well as to systematically investigate moderators that may explain the heterogeneity in results.

In this work, we use a classic intervention – the inhibition of protein synthesis to modulate fear learning, reconsolidation and extinction in mice and rats – as a model for exploring the possibilities of data synthesis in basic biomedical science. By doing this, we aim to investigate whether meta-analytic methods can detect well-established findings – such as the effect of protein synthesis inhibition on consolidation – and whether they can shed light on more controversial topics – such as the effect of reexposure duration, memory strength or memory age on reconsolidation. For this purpose, we explore different ways of aggregating studies within meta-analyses, as well as different approaches to deal with non-independence between experimental results.

Methods

The data collection and analysis protocol was posted prior to performing the work and is available online at https://osf.io/y8r2j. Details of the methods are presented below, highlighting additions to the prespecified protocol.

Study search and identification

PubMed, Web of Science and Scopus were searched for peer-reviewed publications using the terms "("protein synthesis" OR "anisomycin" OR "emetine" OR "cycloheximide" OR "protein synthesis inhibitor" OR "psi" OR "puromycin" OR "acetoxycycloheximide" OR "isocycloheximide" OR "chloramphenicol" OR "rapamycin") AND ("fear conditioning" OR "aversive conditioning" OR "conditioned fear" OR "fear memory" OR "fear learning" OR "aversive learning" OR "pavlovian conditioning" OR "fear-motivated learning" OR "threat conditioning" OR "threat learning" OR "contextual

conditioning" OR "auditory conditioning" OR "fear memory retrieval" OR "fear memory
extinction" OR "fear memory reconsolidation" OR "freezing behavior")". The specific
protein synthesis inhibitors included in the search were drawn from a pilot search using
"protein synthesis" and "memory". In all searches, publication date was restricted to
articles published until December 31st, 2018. The Scopus search was restricted to titles
and abstracts, and the Web of Science search excluded book chapters, editorials,
meeting abstracts and proceedings papers. Duplicates were excluded using Mendeley
(version 1.17.10, Mendeley Ltd).

Study selection

The first screening step considered only titles and abstracts and excluded (i)
articles not written in English, (ii) articles not presenting original results, such as
reviews, and (iii) articles not describing fear conditioning experiments in mice or rats.
Retracted articles were also excluded during this stage. This step was performed by
two reviewers, and at least one of them had to include the reference for it to be taken to
the next stage. If the title/abstract was not clear about the three criteria described
above, the article was still included for further screening.

The second screening stage considered the full text of the article, including any
supplementary material if available, and papers were included if they met the following
criteria: (i) described the effects of a single dose of a protein synthesis inhibitor,
administered either systemically or intracerebrally, (ii) administered protein synthesis
inhibitors from 1 h before up to 24 h after a behavioral session (e.g. fear conditioning
training or reexposure), (iii) assessed memory by a drug-free test using freezing
behavior as a measure of fear, (iv) had a clearly defined control group to which the
experimental group was compared, and (v) had available data on mean freezing and
standard deviation (SD) or standard error of the mean (SEM) in the test session for
each experimental group. In case multiple retrieval tests were performed, only the first
drug-free test session was included in the meta-analysis. Drugs were considered as
protein synthesis inhibitors only when directly affecting the process of translation itself.
Therefore, drugs interfering with transcription or mRNA synthesis (such as actinomycin
D) or affecting translation as a consequence of interference with an upstream signalling
pathway (such as rapamycin) were excluded.

In the second screening stage, each article was evaluated by one of two
investigators who indicated which experiments should be extracted for the meta-
analysis. Extraction of the data was performed by the other reviewer, so that all
included experiments were double-checked, and disagreements regarding inclusion
were discussed with a third investigator until consensus was reached. After the
completion of the screening phase, a random sample of 10% of excluded articles was
double-checked by both investigators to obtain agreement levels for exclusions.

Data extraction

For each included article, we extracted data regarding article- and experiment-
level features, as well as the quantitative results required for the meta-analyses. The
dataset used in all analyses is provided as **Supplementary File 1**. Additional data
collected but not used in the analyses presented here are described in the original
protocol and available at https://osf.io/agdte/.

For each article, we recorded the year of publication, impact factor (as per the
respective year's Journal Citation Reports), number of citations (obtained at the end of
data collection on June 25th 2020) and region of origin (defined by the corresponding
author's affiliation). As articles were published within a range of over 20 years, citations
were normalized as number of citations per year (number of citations/(2020 – year of
publication)).

As risk of bias indicators for each article, we assessed if the following items
were reported for any of the included experiments: randomization of animals to
experimental groups, blinding or automation of outcome assessment (i.e. freezing
measure), sample size calculation, statement regarding compliance with ethical
regulations, and statement on conflict of interest.

The following experiment-level features were extracted: type of conditioning,
training and testing protocols, reexposure protocols (when applicable), habituation and
handling procedures, species, sex, age and number of animals per cage. Habituation
and handling protocols were transformed to categorical dichotomous variables (i.e.
reported or not). When housing information was presented as a range, we recorded the
highest value. When age was presented as a range, we used the mean between
minimum and maximum values.

Fear conditioning protocols were classified as "contextual" or "tone" according
to the type of CS used in the training session. If contextual memory was evaluated by a
test in the same context after tone conditioning, this was labelled as "contextual
background". If presentation of the tone and footshock were separated by an interval,
this was labelled as "tone-trace". We also recorded the number and intensity of
footshocks presented at training and the time interval between sessions (training-
testing, training-reexposure and reexposure-testing).

For each intervention, we extracted the active drug, dose and site of injection.
These were later categorized as systemic, intra-amygdala, intra-hippocampal, intra-
cerebroventricular or other (for other brain structures). We also recorded the time
interval between the intervention and the behavioral session (training, reconsolidation
or extinction) at which it was targeted.

Mean and SD or SEM for freezing levels and sample size were extracted for
both the experimental and control groups. Numerical values were obtained from the
text or legends when available, or directly from graphs when necessary using Gsys
(version 2.4.6, Hokkaido University Nuclear Reaction Data Centre).

Missing Data

Corresponding authors were contacted for included articles in which data on the
sample size of individual groups or the meaning of error bars (SD or SEM) were
missing. Missing data on other protocol features were left blank.

Data exclusion

If authors did not respond to contact attempts, experiments with missing data on
sample size were excluded from the analyses. When error bars were not defined as SD
or SEM, data were included, but were assumed to represent SEM for a conservative
estimate of variance. Additional tests of the same type (tone, tone-trace or contextual)

on the same cohort of animals were excluded – i.e. meta-analyses only include the first
drug-free test after intervention session (training or reexposure). However, if the same
cohort was tested for tone and context memory (i.e. contextual background
conditioning), both tests were included and sample size was divided between them for
analysis. Similarly, when a group served as a control for more than one comparison,
its sample size was divided by the number of extracted comparisons to avoid
overrepresentation of this group in the meta-analyses.

During data extraction, we identified one article with duplicated data (same
figure presented for different experiments) and one article in which mean freezing
values in the text did not match the values in the figures. In both cases, the authors
were contacted but could not retrieve the original data, so these experiments were
excluded. After data extraction was completed, we also decided to exclude unclear or
unusual test protocols, such as re-exposure protocols including the US or those in
which the response to tone conditioning was not measured during the tone.

<u>Analysis</u>

All analyses were performed in R[14] (version 4.0.4). The complete code used is
provided as **Supplementary File 2**.

Article Features

Frequency distributions of region of origin, impact factor, number of citations
and year of publication are presented for the complete dataset. The percentage of risk
of bias items reported was also calculated for the whole sample of articles.

Identification of research groups

Research groups were identified based on co-authorship networks of included
articles[15]. Briefly, the list of authors was obtained for all articles included in our dataset
to create a co-authorship graph using the Louvain method[16]. Modularity classes were
then added to the dataset as an identification of research group. In case an article had
authors attributed to different research groups, we assigned the article to the group to
which most authors belonged.

Meta-Analyses

The random-effects model was chosen for all meta-analyses and fitted with the
restricted maximum-likelihood (REML) estimator (except for multivariable meta-
regressions, where the maximum-likelihood estimator was used due to computational
limitations). For three-level meta-analyses, we opted for a fully random model, where
random effects were used at both the article/group and experiment level. Main results
present effect sizes as absolute mean differences in freezing, but we also performed
sensitivity analyses using standardized mean differences (i.e. Hedges' *g*). All meta-
analyses are based on sample sizes adjusted for the number of comparisons per
control group.

As defined in the protocol, separate meta-analyses were performed for
interventions in different sessions (training, reconsolidation and extinction) and sites
(systemic, intra-amygdala, intra-hippocampus or intracerebroventricular). The
distinction between reconsolidation and extinction in this case was based on the

article's description of the experiment. We also included a meta-analysis for prefrontal
cortex interventions on training, as these were identified in 3 articles (the minimum
number specified in the protocol).

Also following the protocol, we performed a meta-analysis for memory
reactivation, including all studies with interventions in reexposure sessions – i.e. both
reconsolidation and extinction studies. Two experiments did not clearly state whether
their protocol was meant to cause reconsolidation or extinction and were included in
the reactivation analyses only. Of note, as most articles had more than one experiment
included, they could be included in multiple meta-analyses, or contribute multiple
experiments to a single meta-analysis.

Although this was not specified in the predefined protocol, we also performed
aggregated meta-analyses, combining data from all injection sites for each intervention
session and from all intervention sessions in each injection site. A meta-analysis of the
complete dataset, including all injection sites identified and all intervention sessions
was also included. This complete dataset, as well as the training and reactivation
datasets, were used in moderator analyses due to its larger sample size.

For each meta-analysis, I^2 is presented as a measure of heterogeneity
alongside the p-value for the Q-test for heterogeneity, which measures whether the
variability in the observed effect sizes is larger than would be expected based on
sampling variability alone. Low p values thus indicate that the observed differences in
effect sizes between experiments or studies would be unlikely to occur by chance
alone. Possible sources of variability were further explored through moderator
analyses.

Moderator analyses

Univariable models were built for aggregated meta-analyses (i.e. complete,
training and reactivation datasets) to investigate if individual methodological variables
or study features were associated with effect size across experiments. Categorical
variables used were conditioning type, reporting of habituation, reporting of handling
procedures, species, sex, active drug, site of injection, randomization, blinding, sample
size calculation, statement of compliance with regulatory requirements, presence of
conflict of interest and region of origin. Continuous variables used were shock intensity,
number of shocks, time between drug administration and training/reexposure, time
between training and reexposure, reexposure duration (not applicable for interventions
on initial conditioning), time between intervention session and test, mean age of
animals, number of animals housed together, drug dose, impact factor and citations per
year. Doses were z-scored by normalizing each value according to the mean value for
the respective drug and route of administration (systemic or intra-cerebral). Similarly,
reexposure duration was z-scored according to conditioning type using the mean
number of tones (for tone reexposure) or duration (for contextual reexposure) across
studies to allow inclusion of both types of study in the same analysis.

For each univariable model, we present the R^2 value and the p-value of the Q
test of moderators. In 3-level models, R^2 values were calculated as the difference in the
sum of σ^2 values for both levels between the model with no moderators and the

univariable model, divided by the sum of σ² values in the model with no moderators. In
case this yielded a negative value, R^2 was considered to be zero.

As defined in the protocol, we restricted the number of moderators included in
multivariable models to one variable for every 10 experiments. The following priority
order was defined *a priori* in the protocol: re-exposure duration (not applicable to
training meta-analyses), drug, dose, time between intervention session and test, time
to reexposure (not applicable to training meta-analyses), species, shock intensity,
number of shocks, sex and age of animals. Due to low prevalence of reporting, animal
age was not included among the variables tested in any of the datasets. In addition to
the predefined list, we added intervention session (applicable only to the complete
dataset), injection site, conditioning type and time between drug administration and
training/reexposure, which were selected on the basis of univariable models.

All combinations of variables from the selected list were tested in multivariable
models, and the best models were ranked by corrected Akaike Information Criteria
(AICc). For each best model selected (for complete, training and reactivation datasets
with 2- and 3-level analyses), we decomposed the R^2 value for each moderator
included. For this, we calculated the mean of the differences between R^2 from models
with and without the moderator in all possible orders of moderator inclusion.
Additionally, we performed a Q test of moderators for each variable (including all
dummy variables for each categorical moderator) to obtain p-values for individual
variables.

Publication Bias

Due to limitations in sample size, publication bias analyses were restricted to
systemic, intra-amygdala and intra-hippocampal interventions on training and
reconsolidation. This was performed using Egger's regression[17] and trim-and-fill
analyses[18,19]. As recommended by the authors of the method[20], both L_0 and R_0
estimators were used to estimate the number of missing studies and to correct the
effect estimate (and its p-value) after imputation of these studies.

As an additional exploratory measure of publication bias, excess significance
tests[21] were also performed for each meta-analysis. These were based on the meta-
analytic estimates using sample sizes as reported in the articles, with no adjustment for
control group or test session redundancies.

Finally, as planned in the protocol, statistical power for a two-sample two-sided
test was calculated for each experiment, considering its own sample size (without
adjustments) and standard deviation, along with the respective meta-analytical effect
size estimate expressed as absolute mean difference and an alpha of 0.05. We used
this to perform correlation analyses between statistical power and effect size with
article-level features. For this, we used the mean power and effect size of all
experiments per article and calculated Spearman's correlation with impact factor,
citations per year and risk of bias. In this case, risk of bias was used as a score
calculated as 5 minus the sum of reported items.

Results

Study selection

Our search initially retrieved 934 articles, of which 91 met our inclusion
criteria[22–112]. **Figure 1** presents a PRISMA flowchart for the inclusion and exclusion of
articles[113].

Figure 1 - Flowchart of screening stages. In the first stage of screening (n=934), two reviewers
assessed titles and abstracts. If one reviewer included the article, it was selected for full-text screening.
During this stage we did not record reasons for exclusion. The second screening stage (n=548) considered
the full-text and supplementary material, with each being assessed by a single reviewer (no disagreements
found in a double-screened sample of 36 articles). Additional exclusion of comparisons took place during
or after data extraction (see Methods for details), leaving 313 comparisons from 91 articles to be included
in the analyses. PSI: protein synthesis inhibitor.

Study description

Suppl. Figure 1 presents the frequency distribution of region of origin, year of
publication, impact factor and citations for the full set of articles. Included articles were
mostly from North America, published between 2005 and 2015 in journals with a
median impact factor of 4.3. Frequency of reporting of risk of bias factors is presented
in **Suppl. Table 1**. A fair number of articles reported blinded/automated assessment of
outcomes, while randomization and sample size calculation were reported very rarely.

A summary of protocol variables is presented in **Table 1** (full data in **Suppl.**
Table 2). Most comparisons studied the inhibition of protein synthesis in training or
reconsolidation sessions, while studies of extinction were less common. Systemic
injections accounted for about a third of all comparisons, while intracerebral injections
were mostly directed at the amygdala and hippocampus. Distribution between species

was balanced between rats and mice, with 94% of all comparisons including only male
animals, and 86% of studies using anisomycin to inhibit protein synthesis.

Table 1 – Summary of protocol features of comparisons included in the analyses. For categorical
variables, we show the number of comparisons per category, while for continuous variables we show the
median and range (minimum and maximum) and the number of comparisons in which the variable was
reported. Percentages refer to the total of 313 experiments, except for reexposure duration, where the total
is 58 experiments using tone-cued re-exposure protocols and 87 experiments using contextual reexposure
protocols. "Contextual background" refers to cases where conditioning was performed with tone cues, but
testing/reexposure used the context as the CS. Complete data is presented in Suppl. Table 2.

Protocol variable	Value	N (%)
Intervention session	Training	168 (53.7%)
	Reconsolidation	119 (38.0%)
	Extinction	23 (7.3%)
	Reactivation	3 (1.0%)
Site of injection	Systemic	97 (31.0%)
	Amygdala	94 (30.0%)
	Hippocampus	71 (22.7%)
	Cerebral ventricles	9 (2.9%)
	Prefrontal cortex	9 (2.9%)
	Other	33 (10.5%)
Type of conditioning	Contextual	138 (44.1%)
	Tone	103 (32.9%)
	Contextual background	59 (18.8%)
	Tone-trace	13 (4.2%)
Number of footshocks	2 (1 – 76)	313 (100%)
Footshock intensity	0.75 mA (0.3 – 2)	310 (99.0%)
Reexposure duration	Context: 3 min (0.5 – 30)	84 (96.5%)
	Tone: 1 tone (1 – 15)	58 (100%)
Species	Rats	167 (53.4%)
	Mice	146 (46.6%)
Sex	Male	293 (93.6%)
	Both	18 (5.8%)
	Not informed	2 (0.6%)
Age	10 weeks (4 – 22)	122 (39.0%)
Animals per cage	1 (1 – 6)	242 (77.3%)
Active drug	Anisomycin	270 (86.3%)
	Cycloheximide	41 (13.1%)
	4EGI-1	2 (0.6%)

Meta-analyses

As planned *a priori*, we initially performed meta-analyses separated by
intervention session (training, reconsolidation, extinction and reactivation) and site of
drug administration (systemic, amygdala, hippocampus and cerebral ventricles). Most
other intracerebral sites of drug administration were reported in less than 3 articles
(**Suppl. Table 3**); thus, the only additional meta-analysis performed was for
interventions on training with prefrontal cortex injections. In addition to these individual
meta-analyses, we performed an exploratory meta-analysis with the complete dataset
in order to explore predictors with greater power. **Table 2** summarizes these analyses,
and forest plots are presented as **Suppl. Figures 2-4**.

Table 2 - Summary of meta-analyses. Effect sizes are expressed as absolute mean differences in
freezing percentages between PSI-treated and control groups in the test session. Sample size is the
number of experiments. p values refer to main effect comparisons in random-effects meta-analyses (4th
column) and Q-tests for heterogeneity (6th column). i.c.v.: intracerebroventricular, CI: confidence interval.

Dataset	Sample size	Effect size (% freezing) [95% CI]	Meta-analysis p-value	I^2	Q-test p-value

Training, systemic	57	-20.2 [-24.0, -16.3]	6.4×10^{-25}	82.8%	1.5×10^{-38}
Training, amygdala	38	-20.2 [-26.1, -14.3]	1.5×10^{-11}	66.6%	1.5×10^{-10}
Training, hippocampus	41	-14.1 [-18.9, -9.3]	9.0×10^{-9}	87.3%	1.9×10^{-23}
Training, i.c.v.	3	-22.0 [-53.5, 9.6]	0.173	80.9%	0.004
Training, prefrontal cortex	6	-10.7 [-20.5, -0.9]	0.033	0%	0.713
Reconsolidation, systemic	36	-18.6 [-23.1, -14.1]	4.2×10^{-16}	83.4%	4.4×10^{-35}
Reconsolidation, amygdala	47	-20.1 [-25.2, -15.0]	1.4×10^{-14}	78.8%	5.5×10^{-26}
Reconsolidation, hippocampus	20	-19.3 [-27.8, -10.9]	7.8×10^{-6}	89.6%	1.9×10^{-34}
Reconsolidation, i.c.v.	4	-12.5 [-43.9, 18.9]	0.435	88.5%	9.0×10^{-4}
Extinction, systemic	4	28.0 [21.4, 34.6]	1.2×10^{-16}	0%	0.771
Extinction, amygdala	8	-1.1 [-11.6, 9.4]	0.831	69.9%	0.001
Extinction, hippocampus	8	6.3 [-7.3, 19.9]	0.364	79.2%	0.002
Extinction, i.c.v.	2	21.0 [-12.2, 54.2]	0.214	84.8%	0.010
Reactivation, systemic	40	-14.2 [-20.1, -8.2]	3.3×10^{-6}	91.7%	1.3×10^{-66}
Reactivation, amygdala	56	-18.1 [-23.2, -13.0]	2.9×10^{-12}	82.4%	2.3×10^{-43}
Reactivation, hippocampus	30	-13.2 [-21.3, -5.2]	0.001	90.7%	1.9×10^{-59}
Reactivation, i.c.v.	6	0.9 [-21.7, 23.5]	0.939	91.6%	2.8×10^{-5}
Complete	313	-15.4 [-17.4, -13.3]	6.0×10^{-48}	87.0%	6.5×10^{-284}

We found significant negative effects on test freezing after protein synthesis
inhibition in training and reconsolidation for almost all studied structures, with roughly
comparable effect sizes. For extinction, however, an effect in the opposite direction (i.e.
preventing freezing decline) was found only for systemic injections. That said, these
results should be interpreted with caution, as there were far fewer studies on extinction
than on training or reconsolidation. Since the reactivation dataset largely overlaps with
the reconsolidation one, results were similar between them, but with smaller effect
sizes for reactivation due to the inclusion of extinction experiments. Most analyses
showed a large amount of heterogeneity between experiments. Results of the same
analyses using standardized mean differences (Hedge's g) are shown as a robustness
check in **Suppl. Table 4**.

Publication bias

To evaluate the presence of publication bias in our sample, we performed trim-
and-fill analyses and Egger's regression for systemic, intra-amygdala and intra-
hippocampal interventions on training and reconsolidation (other meta-analyses were

not included due to their small sample size). **Figure 2** presents funnel plots with the
results of Egger's regression and trim-and-fill analyses using the L_0 method (results
with the R_0 method are presented in **Suppl. Table 5**). Unlike what was described by
Schroyens et al. (2021) for various drugs in contextual fear reconsolidation, we did not
find consistent evidence of publication bias for protein synthesis inhibitors: a significant
correlation between effect size and precision was found only for intra-hippocampal
interventions on reconsolidation, but in the opposite direction as would be expected
from publication bias.

Figure 2 – Funnel plots and Egger's regression. Each observed study is represented by the white circles and the red dashed line represents Egger's regression line. Black circles represent missing studies imputed by the trim-and-fill L_0 method. **(A)** Training, systemic. Egger's regression p-value = 0.39. **(B)** Training, amygdala. Egger's regression p-value = 0.75. **(C)** Training, hippocampus. Egger's regression p-value = 0.39. **(D)** Reconsolidation, systemic. Egger's regression p-value = 0.24. Estimated number of missing studies = 5. **(E)** Reconsolidation, amygdala. Egger's regression p-value = 0.98. **(F)** Reconsolidation, hippocampus. Egger's regression p-value = 0.009. S.E.; Standard error.

Another potential source of bias in a meta-analysis is lack of statistical power in
the original studies, which along with publication bias can lead to effect size inflation. In
Suppl. Figure 5, we show the distribution of sample sizes and statistical power for
training and reconsolidation experiments for systemic, intra-amygdala and intra-
hippocampus interventions, calculated using the respective meta-analytic estimate as
the reference effect size. Mean sample size (± SD) for all experiments combined was
9.1 ± 2.8 and mean statistical power for the corresponding meta-analytical effect size
estimate was 0.66 ± 0.26. While differences in power for individual meta-analyses are
heavily determined by differences between effect size estimates, experiments with
intracerebral injections also tended to have smaller sample sizes than those with
systemic interventions. No correlation was found between mean effect size or statistical
power and impact factor, citations or risk of bias score for articles (**Suppl. Figure 6**).

Consistent with what we found by funnel-plot analysis, excess significance test did not
detect evidence of publication bias (**Suppl. Table 6**).

Meta-regressions

404   A major challenge in performing meta-analyses of basic and pre-clinical
405 research is defining the boundaries of a research question – i.e. how similar
406 experiments should be for their data to be synthesized meaningfully. On one hand,
407 including experiments with diverse protocols adds heterogeneity to the analyses,
408 rendering their interpretation more difficult; on the other, this allows one to explore the
409 impact of protocol variables on effect size.

410   *A priori*, we determined that our analyses would be divided by the target session
of drug administration (training, reconsolidation or extinction) and by the site of drug
administration (hippocampus, amygdala, i.c.v. or systemic), as presented in **Table 2**
and **Suppl. Table 4**. However, to investigate whether aggregating data could provide
additional information, we also combined all experiments for each intervention session
(i.e. training, reconsolidation, extinction and reactivation), testing the injection site as a
moderator (**Table 3, Suppl. Table 7**), and combined all experiments for each site of
injection, testing the session as a moderator (**Table 3, Suppl. Table 8**). While the
variance explained by injection site was relatively small (and driven mostly by
structures other than the hippocampus and amygdala), the intervention session had a
larger impact on effect sizes, particularly due to the influence of extinction experiments,
in which the direction of effect was reversed. Nevertheless, a large amount of residual
heterogeneity persisted even after adding these moderators.

Table 3 – Meta-regression models for testing intervention site and target session. Grey rows show
the results of the models without moderators, while white lines show the results of meta-regressions. p-
values refer to the Q-test for residual heterogeneity (3rd column) or the test of significance for the
moderator (5th column). R^2 and p-values for multivariate meta-regression are presented in order, with the
first row showing the results of intervention session and the second row showing those of injection site.

Dataset and moderators	I^2	p-value	R^2	p-value
Training, no moderators	81.9%	9.7×10^{-79}		
Training, injection site as moderator	80.3%	3.3×10^{-69}	5.2%	0.058
Reconsolidation, no moderators	85.9%	2.3×10^{-123}		
Reconsolidation, injection site as moderator	83.2%	1.2×10^{-92}	17.6%	2.3×10^{-4}
Extinction, no moderators	84.4%	5.0×10^{-12}		
Extinction, injection site as moderator	70.0%	1.3×10^{-5}	37.1%	0.018
Systemic, no moderators	88.2%	3.7×10^{-104}		
Systemic, intervention session as moderator	82.4%	4.2×10^{-70}	36.6%	3.8×10^{-10}
Amygdala, no moderators	77.8%	9.2×10^{-51}		
Amygdala, intervention session as moderator	74.1%	6.4×10^{-36}	16.5%	0.002
Hippocampus, no moderators	90.5%	1.0×10^{-89}		
Hippocampus, intervention session as moderator	88.3%	1.6×10^{-55}	17.0%	0.004
i.c.v., no moderators	91.7%	1.8×10^{-9}		
i.c.v., intervention session as moderator	84.9%	6.6×10^{-6}	14.1%	0.198
Other, no moderators	70.0%	2.0×10^{-12}		
Other, intervention session as moderator	57.0%	8.1×10^{-6}	41.1%	4.9×10^{-4}
Complete, no moderators	86.9%	2.9×10^{-282}		
Complete, intervention session and injection site as moderators	81.9%	2.4×10^{-195}	20.9%	7.6×10^{-14}
			7.4%	9.0×10^{-5}

A final possibility to increase sample size for meta-regression is by combining
both intervention sessions and sites of drug administration using the complete dataset.
To assess the impact of this merging, we used a multivariable meta-regression model
considering both intervention session and site of injection (**Table 3, Suppl. Table 9**).
Although both moderators account for a relevant amount of heterogeneity, we
considered it reasonable to use the complete dataset to explore additional protocol
variables in meta-regression, due to the greater sample size achieved. Thus, from here
onward, we restrict our analyses to the complete dataset and to datasets with all
interventions (irrespectively of site) on training and on reactivation (including
reconsolidation, extinction and reexposure interventions).

Multilevel meta-analyses

The standard random-effects models in the meta-analyses presented above
consider two sources of variability: the real variance among different experiments and
the sampling error of each experiment. Both of these, however, are nested within at
least two additional sources of variability: that between articles from which experiments
are obtained and that between research groups publishing the articles. As multiple
experiments within a paper and/or by a research group are not completely independent
of each other, not taking these levels into account may bias the analyses towards
results of particular articles or research groups that are overrepresented in the sample.

To test the importance of including these additional levels of variability, we
applied an authorship network-based method[15] to identify research groups in the
complete set of articles included (see Methods for details). Forty-two groups were
identified by modularity analyses, as shown in **Suppl. Fig. 7**. The distribution of
experiments within articles and research groups can be visualized in **Suppl. Fig. 8**.
Although most articles and research groups contribute only a few experiments to the
sample, a few are distinctly overrepresented in the sample (with a single research
group contributing 58 experiments).

We then built additional models assuming random effects both within and
between either articles or research groups (**Table 4**). This led to minor changes in
estimates, and variance was larger between experiments than between either articles
or research groups for all models. Nevertheless, in the reactivation dataset, these
additional levels accounted for a reasonable fraction of the observed heterogeneity.
The amount of variance explained by articles and research groups can also be
observed by including articles and research groups as moderators (i.e. fixed factors) in
meta-regressions (**Suppl. Table 10**), although this does not allow correction of effect
estimates, as occurs when they are included as levels (i.e. random factors).

We also built a four-level model with articles and research groups as different
levels (**Suppl. Table 11**). However, variance estimations were much less precise in this
case, indicating a lack of clear distinction between each level's contribution to total
heterogeneity.

Table 4 – Three-level meta-analyses. Two 3-level models were built, considering nesting
within clusters   that represented either articles or research groups, which are modeled as
additional random factors. The two-level (i.e. standard random-effects) model is also included
for comparison. Effect sizes are in absolute mean differences. CI, confidence interval; Exp.,
experiments; cl., clusters.

Model	Sample size		Effect size (% freezing) [95% CI]	p-value	Variance [95% CI]		I^2	
	Exp.	cl			Between	Between	Exp.	Cluster

					experiments	clusters		
Complete dataset								
2-level	313	0	-15.35 [-17.4, -13.3]	7.9×10^{-48}	265.8 [215.7, 325.2]	-	85.8%	-
3-level (article)	313	91	-15.14 [-17.9, -12.4]	8.2×10^{-28}	202.3 [157.0, 260.5]	68.3 [25.2, 135.1]	65.2%	22.0%
3-level (group)	313	42	-15.27 [-18.6, -12.0]	1.2×10^{-19}	237.2 [188.9, 296.4]	40.4 [3.7, 125.8]	74.7%	12.7%
Training dataset								
2-level	168	0	-17.35 [-19.8, -14.9]	2.9×10^{-44}	174.6 [122.8, 230.2]	-	81.9%	-
3-level (article)	168	57	-17.19 [-19.8, -14.5]	4.5×10^{-37}	163.3 [114.8, 228.9]	11.4 [0, 58.1]	76.6%	5.4%
3-level (group)	168	29	-17.31 [-19.8, -14.9]	4.4×10^{-44}	174.6 [128.5, 236.9]	1.1×10^{-6} [0, >10.0]	81.9%	0%
Reactivation dataset								
2-level	145	0	-13.24 [-16.7, -9.8]	3.1×10^{-14}	361.6 [280.8, 497.8]	-	89.5%	-
3-level (article)	145	56	-13.06 [-17.5, -8.6]	6.8×10^{-9}	249.1 [171.2, 363.6]	119.8 [32.2, 265.5]	60.6%	29.1%
3-level (group)	145	28	-12.21 [-17.9, -6.5]	2.5×10^{-5}	296.9 [215.4, 413.6]	96.9 [6.6, 309.2]	68.0%	22.2%

Univariable meta-regressions

476   To explore the impact that individual features in the experiments might have on
477 effect sizes, we initially performed univariable meta-regressions using the two-level
478 model for all protocol and article variables collected, including risk of bias measures.
479 Results are summarized in **Figure 3A**, with a more extensive description available in
480 **Suppl. Tables 12-14**.

Figure 3 – Summary of univariable meta-regression models. Each column represents a dataset, while
lines correspond to each variable. Cell colors represent p-values for Q tests of the respective variable as a

moderator and numbers within cells are the R^2 value for the moderator in a univariable model. White cells
indicate that the variable was not applicable or unfeasible to include in the model due to lack of variability.
Reexposure duration is based on z-scored values for reexposure to tone (measured in number of tones)
and to context (measured in minutes), based on the mean and standard deviation for each type of
conditioning. Doses were also z-scored based on the mean and standard deviation of each drug and route
of administration. **(A)** Two-level models. **(B)** Three-level models, accounting for nesting of experiments
within articles. **(C)** Three-level models, accounting for nesting of experiments within research groups. Full
details for each model, including betas and directionality of effects, are shown on **Suppl. Tables S12-S21**.

For interventions in the training session, the time between drug administration
and training (i.e. intervention time) was found to be a significant moderator of effect
size. As can be observed in **Suppl. Figure 9**, this relationship is largely driven by
injections performed 24 hours after the training session, which yield near-null effects.

In the reactivation sample, reexposure duration and number of tones (which
were z-scored and included as a single variable) predictably influenced effect size. As
there was little variation in the number of tones across experiments, the moderator
effect can be attributed mainly to contextual fear conditioning experiments, in which
effects on freezing were negative with under 10 minutes of reexposure (suggesting
interference with reconsolidation), variable between 10 and 20 minutes, and mostly
positive with 30 minutes of reexposure (suggesting effects on extinction) (**Suppl.**
Figure 10A-B). This is confirmed by performing separate meta-regressions based on
the duration of reexposure to context or number of tones (**Suppl. Table 15**), which
show that the effect of reexposure duration on effect sizes is much more robust for
contextual conditioning than for tone conditioning, in which experiments exploring
larger number of tones were less common.

Additionally, conditioning type by itself also impacted effect sizes, with tone
conditioning leading to larger effects on reconsolidation. As can be observed in **Suppl.**
Figure 10C, the impact of conditioning type is probably due to the large number of
near-null effects for contextual fear reconsolidation, as well as to a greater number of
extinction experiments using contextual conditioning. A higher number of shocks was
also associated with larger effects on reconsolidation. That said, as can be seen from
Suppl. Figure 11A, this association is mostly driven by a few experiments with 60
shocks and large effects, while an opposite trend is observed for experiments with up
to 10 shocks.

Other proposed boundary conditions for reconsolidation, such as time between
training and reexposure, were not detected as moderators in the model. That said,
Suppl. Figure 11B shows that, for the few experiments with more than 20 days
between training and reexposure, effects seem to be consistently smaller than those
with shorter intervals, where there is greater variability in results. It is also worth noting
that extinction experiments generally used short intervals, which might have obscured
an effect of this variable on reconsolidation.

Next, we performed the same meta-regressions for three-level models,
accounting for the nesting of experiments within articles (**Figure 3B** and **Suppl. Tables**
16-18) and within research groups (**Figure 3C** and **Suppl. Tables 19-21**). In these
models, the amount of heterogeneity accounted for most moderators is lower,
suggesting that some of the moderator effects might be explained by the
article/research group of origin instead. However, the main findings persist – as in the
case of injection time in training, and of conditioning type and reexposure duration in
reactivation.

Multivariable meta-regressions

Lastly, we explored multivariable models to investigate whether the moderator
effects detected in univariable regressions could be due to confounding by other
variables. We started with the complete dataset and the original variables that
separated the analyses (injection site and intervention session), the two variables with
the largest effects in univariable models (conditioning type and intervention time) and 7
other variables selected *a priori* (active drug, dose, time to test, species, number of
shocks, shock intensity and sex). Missing data for some of these variables led to the
exclusion of 36 experiments, leaving 277 to be used in these analyses.

In the training dataset, including the same variables listed above (except for
intervention session) led to the exclusion of 11 experiments, leaving 157 available for
the analyses. In the reactivation dataset, the predetermined list of variables included
two additional variables: reexposure duration and time between test and reexposure.
This led to exclusion of 19 experiments, leaving 126 available.

After testing all possible variable combinations in the 2-level and 3-level models
(2,048 in the complete dataset, 1,024 in the training and reactivation datasets), we
ranked the resulting models by the Akaike Information Criteria (AICc). For each dataset
and clustering option, we present a summary of the best models in **Table** 7 and effect
sizes are presented in **Suppl. Tables 22-30**.

Table 7 – Best multivariable meta-regression models for each dataset and clustering option. Only
the model with lowest AICc is presented for each scenario. Weight refers to the Akaike weights. Q-test p-
values refer to the test for all included moderators. Effect sizes and p values for individual moderators in
each model are presented in **Suppl. Tables 22-30**.

Dataset	Clusters	Included variables	AICc	Weight	I^2	R^2	Q-test p-value
Complete	2-level	Intervention session Site of injection Conditioning type Active drug Number of shocks Intervention time	2328.1	0.060	76.4%	45.1%	9.6×10^{-24}
Complete	3-level (article)	Intervention session Site of injection Active drug Number of shocks Intervention time	2328.8	0.056	79.2%	44.5%	4.6×10^{-22}
Complete	3-level (group)	Intervention session Site of injection Active drug Number of shocks Intervention time	2326.7	0.081	79.7%	44.0%	2.1×10^{-22}
Training	2-level	Site of injection Intervention time	1299.2	0.037	73.1%	27.7%	1.9×10^{-6}
Training	3-level (article)	Site of injection Intervention time	1301.4	0.036	75.4%	30.7%	1.9×10^{-6}
Training	3-level (group)	Site of injection Intervention time	1301.4	0.037	75.4%	30.1%	1.2×10^{-6}
Reactivation	2-level	Site of injection Reexposure duration Number of shocks	1090.9	0.117	81.1%	45.2%	5.1×10^{-14}
Reactivation	3-level (article)	Site of injection Reexposure duration Number of shocks	1092.4	0.124	82.6%	43.6%	7.3×10^{-13}
Reactivation	3-level	Site of injection	1093.2	0.122	82.3%	47.4%	5.1×10^{-14}

(group)	Reexposure duration Number of shocks					

Notably, multivariable models did not include some article-level variables detected as significant by univariable models, such as presence of a regulatory requirements statement and region of origin, suggesting that these effects were likely due to confounding. Nevertheless, most variables related to the experimental protocol itself were preserved. These results can also be visualized by the importance of all variables in each set of models (**Suppl. Fig. 12**).

For each of these models we decomposed the R^2 value for each of the variables selected to investigate the contribution of each one (**Figure 4**). Once more, it is notable that R^2 values in the reactivation set are lower for most variables in multilevel models, suggesting that some of the correlation between these variables and effect sizes is driven by confounding from the use of specific reexposure protocols by particular articles or groups. Conditioning type in particular was not detected as a moderator when additional levels are considered, suggesting that this effect might have been driven by research groups who preferentially use tone or context conditioning. This does not seem to be the case in the training set, where including articles or groups as levels actually increased the effect of intervention time and site of injection.

Figure 4 – Summary of multivariable meta-regression models. Each column represents a dataset and lines are variables selected in the best model for each scenario. Cell colors represent the p value for the Q test for each moderator in the meta-regression and numbers within are the decomposed R^2 value for the moderator (see Methods). Reexposure duration is based on z-scored values for reexposure to tone (measured in numbers) and to context (measured in minutes), based on the mean and standard deviation for each type of conditioning. **(A)** Two-level models. **(B)** Three-level models, accounting for nesting of experiments within articles. **(C)** Three-level models, accounting for nesting of experiments within research groups.

Discussion

In this study, we explored different meta-analysis methods to analyze the effect of protein synthesis inhibitors on fear conditioning. Beyond the results, this effort also leads to some insights on how to apply existing meta-analysis methods to discovery science, which presents challenges to data synthesis.

One particular focus was to investigate different ways to aggregate studies.
Combining them by intervention session and by injection site showed a very robust
effect for interventions in the three most studied sites (systemic injections,
hippocampus and amygdala) on training and reconsolidation. This is unsurprising, as
the requirement of protein synthesis for consolidation has been well accepted for
decades[114]. More recently, protein synthesis inhibitors also became the paradigmatic
intervention to demonstrate the existence of reconsolidation[51,115].

On the other hand, the effect of protein synthesis inhibition on fear extinction
was not as robust. Findings on the role of protein synthesis in extinction, in fear
conditioning and beyond[116,117], were not consensual from the start[111]. Moreover, when
compared to reconsolidation, much less evidence on the topic is available. Thus, our
meta-analysis found a significant effect only for systemic interventions. That said, most
null effects on extinction from amygdala interventions come from a single study[88],
whereas the hippocampus analysis is heavily influenced by a study which, although
investigating extinction according to the authors, would be considered a reconsolidation
experiment by most standards[104].

Even though the training and reconsolidation data are robust on aggregate,
there is great heterogeneity among individual experiments for virtually all meta-
analyses performed. When articles were included as an additional level, most of this
heterogeneity was observed between different experiments within studies, perhaps
because behavioral neuroscience articles typically include experiments as tests of
boundary conditions or negative controls. This presents a challenge for meta-analysis,
as selection criteria will commonly lead to inclusion of many of these experiments,
which are likely to bias effect size estimates downwards.

Aggregating studies from all injection sites or from interventions in training and
reactivation did not cause much of an increase in heterogeneity, perhaps because it
was already large to start with. As the increased sample sizes obtained provided
greater power for identifying moderators, we used this approach for our meta-
regressions, in a decision that was not prespecified in the original protocol.

Some of the included moderators provide sanity checks of whether meta-
regression captures consensual knowledge. It is widely accepted, for instance, that the
effect of protein synthesis inhibitors is time-dependent, occurring when they are
administered up to a few hours after learning[109,118]. Accordingly, in the training dataset,
we found the time between training and drug administration to be a significant
moderator in all models tested.

Similarly, it is well accepted that the occurrence of reconsolidation or extinction
depends on the degree of mismatch between the training session and the reactivation,
which is classically modulated by varying the number of CS presentations[119,120] or the
duration of reexposure[105,121]. Our measure of reexposure duration, which incorporated
both of these manipulations, indeed was found to be a significant moderator in the
reactivation dataset. Interestingly, however, it had a much smaller effect size when
research group was included as a level in the analysis.

Other postulated boundary conditions for reconsolidation, however, were not
observed as clearly. Many studies suggest that training strength can determine the
occurrence of reconsolidation, extinction or neither[105,122,123]. This relation, however, is
not linear: while very low training strength favors extinction rather than
reconsolidation[122], very strong memories have been suggested to be more resistant to

destabilization[123]. Contrary to that, we found that a higher number of shocks led to a
greater effect of protein synthesis inhibitors; that said, this effect was largely driven by
a few experiments with a very large number of shocks. In contrast, shock intensity did
not appear to be a relevant moderator.

Another proposed boundary condition for reconsolidation is memory age[124], but
this has not been consistent across studies[46] and may depend on reexposure
conditions[105]. Our meta-analysis did not find an influence of memory age on the effect
of protein synthesis inhibitors on reactivation; nevertheless, it is worth pointing out that
our model does not include interactions between moderators; thus, an effect could
have been obscured by confounding. It is also possible that this effect is limited to very
remote memories: studies with memories over 20 days-old had smaller effects in our
sample, but this was offset by large effects for those between 1- and 20 days-old.

Type of conditioning (tone vs. contextual) was consistently detected as a
significant moderator for reconsolidation effects in our univariable models, mostly
driven by frequent null results in contextual conditioning experiments. Importantly,
however, the effect we found was nearly absent from multivariable models, perhaps
due to its interaction with site of injection, as both types of conditioning tend to be
studied with interventions in different brain structures.

The changes in the relative importance of different moderators when
multivariable and multilevel models are used underscore the importance of these
methods to minimize confounding. Interestingly, the effect of intervention time, perhaps
our best-established moderator, became stronger when multiple levels were added.
Conversely, seemingly spurious factors such as region of origin or presence of a
conflict-of-interest statement disappeared when research group was used as a level.

Even after moderators and additional levels were included, however, much
heterogeneity remained, suggesting that classic boundary conditions are not sufficient
to explain discrepancies in the literature. This echoes recent failures in replicating post-
retrieval amnesia studies, despite the use of similar protocols[11,125]. Minor differences in
reactivation protocols, animal traits[126], housing conditions[127] or interactions between
moderators could explain this additional heterogeneity. Nevertheless, it is also possible
that heterogeneity and replication failures are due to biases in the literature.

In this sense, the prevalence of randomization and sample size calculations in
our sample was low, in accordance with previous surveys of preclinical studies[128].
Encouragingly, however, we found a higher level of blinding than reported in other
areas[128], mostly due to the use of automated freezing detection. Another interesting
finding is the near absence of studies using female animals, which has also been
pointed out in previous studies of fear conditioning[129] and deviates markedly from
current recommendations[130,131].

Another explanation for divergences in the literature is random error, which can
lead to effect size vibration across statistical thresholds, particularly when power is low.
We found the average power of individual studies to detect the respective meta-
analytical effect size estimate to be 63%, which is in line with a previous estimate for
fear conditioning[129]. This means that around one third of studies should fail to find
significant differences between groups even when a real effect exists, suggesting that
lack of replication based on statistical significance criteria is expected to occur
frequently.

Interestingly, we found little evidence of publication bias, in contrast to what was
found by Schroyens et al (2021) for anisomycin, propranolol, MK-801 and midazolam in
contextual fear reconsolidation. The evidence for publication bias in their study is
weaker for anisomycin, however, suggesting that it may be a smaller problem in the
literature on protein synthesis inhibitors. Nevertheless, our ability to detect publication
bias may have been limited by experiments testing boundary conditions or negative
controls, as bias may act on the opposite direction in these cases (i.e. suppressing
statistically significant results). Publication bias assessments can also be limited by
true heterogeneity[20], which is quite large in our sample.

In conclusion, we have found that meta-analysis and meta-regression can
capture some well-established findings in the fear conditioning literature. This includes
a robust effect of protein synthesis inhibitors on consolidation and reconsolidation and
its dependence on moderators such as timing of post-training interventions and the
degree of reexposure in reactivation experiments. That said, not every boundary
condition commonly mentioned in the literature was confirmed. This may indicate that
the evidence for some of these is more controversial than suggested in the literature, or
alternatively that meta-regression might fail to capture real effects due to noise, non-
linearity or interactions between variables. This leads to a conundrum – when meta-
regression doesn't agree with individual studies that test a particular moderator, what
should we trust?

Unfortunately, there is no easy answer. On one hand, individual studies are
typically underpowered and subject to many biases. On the other, meta-regression
findings are observational and inevitably prone to confounding. The use of multilevel
models and multivariable regression can mitigate this problem but will not fully
eliminate it. Moderators found by meta-regression should thus be considered tentative
and ideally tested in empirical studies designed for this purpose. Although data
synthesis can be useful for evaluating the primary literature, it incorporates its biases,
and cannot fully substitute for confirmatory, high-powered replication studies[132].

A final advantage of empirically testing findings from meta-analysis is to
compare the predictions of different types of models in terms of their accuracy.
Although we argue in favor of using multilevel and multivariable models, these can be
unfeasible or inaccurate when the number of studies is low. It is thus important to
evaluate how much is lost by traditional two-level models, which in our work were in
general agreement with multilevel ones.

All of these issues will only be resolved when systematic iteration between data
synthesis and experimentation becomes more common in discovery science. This
requires engaging researchers in systematic reviews, as well as making these more
efficient through automated tools[133] and better structuring of primary data. It also
involves building up infrastructure for confirmatory research that can validate meta-
analytical findings empirically[132]. Ultimately, such interplay may produce a virtuous
cycle that can improve the robustness of basic science beyond what can be achieved
with the current model of isolated experiments and narrative synthesis.

Acknowledgments

We'd like to thank Daniel Menezes for assistance in the early stages of screening,
Thiago Moulin for assistance with co-authorship graphs, Kleber Neves for general
assistance with analyses in R and Jonathan Lee for opinions on the manuscript.

References

1.   Open Science Collaboration. PSYCHOLOGY. Estimating the reproducibility of
psychological science. *Science*. 2015;349(6251):aac4716. doi:10.1126/science.aac4716

2.   Errington TM, Mathur M, Soderberg CK, et al. Investigating the replicability of preclinical
cancer biology. *eLife*. 2021;10. doi:10.7554/eLife.71601

3.   Amaral OB, Neves K, Wasilewska-Sampaio AP, Carneiro CF. The brazilian reproducibility
initiative. *eLife*. 2019;8. doi:10.7554/eLife.41602

4.   Begley CG, Ellis LM. Drug development: Raise standards for preclinical cancer research.
Nature. 2012;483(7391):531-533. doi:10.1038/483531a

5.   Prinz F, Schlange T, Asadullah K. Believe it or not: how much can we rely on published
data on potential drug targets? *Nat Rev Drug Discov*. 2011;10(9):712.
doi:10.1038/nrd3439-c1

6.   Steward O, Popovich PG, Dietrich WD, Kleitman N. Replication and reproducibility in
spinal cord injury research. *Exp Neurol*. 2012;233(2):597-605.
doi:10.1016/j.expneurol.2011.06.017

7.   Vesterinen HM, Sena ES, Egan KJ, et al. Meta-analysis of data from animal studies: A
practical guide. *J Neurosci Methods*. 2014;221:92-102.
doi:10.1016/j.jneumeth.2013.09.010

8.   França TF, Monserrat JM. Reproducibility crisis in science or unrealistic expectations?
EMBO Rep. 2018;19(6):e46008. doi:10.15252/embr.201846008

9.   Stanley DJ, Spence JR. Expectations for Replications: Are Yours Realistic? *Perspect Psychol
Sci J Assoc Psychol Sci*. 2014;9(3):305-318. doi:10.1177/1745691614528518

10.   Wotjak CT. Sound check, stage design and screen plot – how to increase the
comparability of fear conditioning and fear extinction experiments. *Psychopharmacology
(Berl)*. 2019;236(1):33-48. doi:10.1007/s00213-018-5111-5

11.   Luyten L, Schnell AE, Schroyens N, Beckers T. Lack of drug-induced post-retrieval amnesia
for auditory fear memories in rats. *BMC Biol*. 2021;19(1):17. doi:10.1186/s12915-021-
00957-x

12.   Schroyens N, Alfei JM, Schnell AE, Luyten L, Beckers T. Limited replicability of drug-
induced amnesia after contextual fear memory retrieval in rats. *Neurobiol Learn Mem*.
2019;166:107105. doi:10.1016/j.nlm.2019.107105

13.   Schroyens N, Sigwald EL, Van Den Noortgate W, Beckers T, Luyten L. Reactivation-
dependent amnesia for contextual fear memories: Evidence for publication bias. *eNeuro*.
2021;8(1):1-16. doi:10.1523/ENEURO.0108-20.2020

14.   R Core Team. R: A Language and Environment for Statistical Computing. Published online
2022. https://www.r-project.org/

15.   Moulin TC, Amaral OB. Using collaboration networks to identify authorship dependence
in meta-analysis results. *Res Synth Methods*. 2020;11(5):jrsm.1430.
doi:10.1002/jrsm.1430

16.   Blondel VD, Guillaume JL, Lambiotte R, Lefebvre E. Fast unfolding of communities in large
networks. *J Stat Mech Theory Exp*. 2008;2008(10):P10008. doi:10.1088/1742-
5468/2008/10/P10008

17.   Egger M, Smith GD, Schneider M, Minder C. Bias in meta-analysis detected by a simple,
graphical test. *Br Med J*. 1997;315(7109):629-634. doi:10.1136/bmj.315.7109.629

18.   Duval S, Tweedie R. Trim and Fill: A Simple Funnel-Plot-Based Method of Testing and
Adjusting for Publication Bias in Meta-Analysis. *Biometrics*. 2000;56(2):455-463.
doi:10.1111/j.0006-341X.2000.00455.x

19.   Duval S, Tweedie R. A Nonparametric "Trim and Fill" Method of Accounting for
Publication Bias in Meta-Analysis. *J Am Stat Assoc*. 2000;95(449):89-98.
doi:10.1080/01621459.2000.10473905

20.   Duval S. The Trim and Fill Method. In: Rothstein H, Sutton A, Borenstein M, eds.
Publication Bias in Meta-Analysis: Prevention, Assessment and Adjustments. John Wiley &
Sons Ltd; 2005.

21.   Ioannidis JP, Trikalinos TA. An exploratory test for an excess of significant findings. *Clin
Trials*. 2007;4(3):245-253. doi:10.1177/1740774507079441

22.   Frankland PW, Josselyn SA, Anagnostaras SG, Kogan JH, Takahashi E, Silva AJ.
Consolidation of CS and US representations in associative fear conditioning.
Hippocampus. 2004;14(5):557-569. doi:10.1002/hipo.10208

23.   Runyan JD, Dash PK. Inhibition of hippocampal protein synthesis following recall disrupts
expression of episodic-like memory in trace conditioning. *Hippocampus*. 2005;15(3):333-
339. doi:10.1002/hipo.20055

24.   Motanis H, Maroun M. Differential involvement of protein synthesis and actin
rearrangement in the reacquisition of contextual fear conditioning. *Hippocampus*.
2012;22(3):494-500. doi:10.1002/hipo.20915

25.   XinChun J, XueLian Q, XiaoFei Y, BaoMing L. Protein synthesis inhibition in the basolateral
nucleus of amygdala facilitates extinction of auditory fear memory. *Chin Sci Bull*.
2007;52(18):2532-2542. doi:10.1007/s11434-007-0393-8

26.   Wanisch K, Tang J, Mederer A, Wotjak CT. Trace fear conditioning depends on NMDA
receptor activation and protein synthesis within the dorsal hippocampus of mice. *Behav
Brain Res*. 2005;157(1):63-69. doi:10.1016/j.bbr.2004.06.009

27.   Yim AJ, Moraes CRG, Ferreira TL, Oliveira MGM. Protein synthesis inhibition in the
basolateral amygdala following retrieval does not impair expression of morphine-
associated conditioned place preference. *Behav Brain Res*. 2006;171(1):162-169.
doi:10.1016/j.bbr.2006.03.031

28.   Patricio Casanova J, Madrid C, Contreras M, et al. A role for the interoceptive insular
cortex in the consolidation of learned fear. *Behav BRAIN Res*. 2016;296:70-77.
doi:10.1016/j.bbr.2015.08.032

29.   Abate G, Colazingari S, Accoto A, Conversi D, Bevilacqua A. Dendritic spine density and
EphrinB2 levels of hippocampal and anterior cingulate cortex neurons increase

sequentially during formation of recent and remote fear memory in the mouse. *Behav*
Brain Res. 2018;344:120-131. doi:10.1016/j.bbr.2018.02.011

30.   Rizzo V, Touzani K, Raveendra BL, et al. Encoding of Contextual Fear Memory Requires De
Novo Proteins in the Prelimbic Cortex. *Biol Psychiatry Cogn Neurosci Neuroimaging*.
2017;2(2):158-169. doi:10.1016/j.bpsc.2016.10.002

31.   Nakayama D, Baraki Z, Onoue K, Ikegaya Y, Matsuki N, Nomura H. Frontal association
cortex is engaged in stimulus integration during associative learning. *Curr Biol*.
2015;25(1):117-123. doi:10.1016/j.cub.2014.10.078

32.   Bekinschtein P, Cammarota M, Igaz LM, Bevilaqua LRM, Izquierdo I, Medina JH.
Persistence of Long-Term Memory Storage Requires a Late Protein Synthesis- and BDNF-
Dependent Phase in the Hippocampus. *Neuron*. 2007;53(2):261-277.
doi:10.1016/j.neuron.2006.11.025

33.   Blundell J, Kouser M, Powell CM. Systemic inhibition of mammalian target of rapamycin
inhibits fear memory reconsolidation. *Neurobiol Learn Mem*. 2008;90(1):28-35.
doi:10.1016/j.nlm.2007.12.004

34.   Mac Callum PE, Hebert M, Adamec RE, Blundell J. Systemic inhibition of mTOR kinase via
rapamycin disrupts consolidation and reconsolidation of auditory fear memory.
Neurobiol Learn Mem. 2014;112:176-185. doi:10.1016/j.nlm.2013.08.014

35.   Gould TJ, Wilkinson DS, Yildirim E, Poole RLF, Leach PT, Simmons SJ. Nicotine shifts the
temporal activation of hippocampal protein kinase A and extracellular signal-regulated
kinase 1/2 to enhance long-term, but not short-term, hippocampus-dependent memory.
Neurobiol Learn Mem. 2014;109:151-159. doi:10.1016/j.nlm.2014.01.009

36.   Kwapis JL, Jarome TJ, Lee JL, Helmstetter FJ. The retrosplenial cortex is involved in the
formation of memory for context and trace fear conditioning. *Neurobiol Learn Mem*.
2015;123:110-116. doi:10.1016/j.nlm.2015.06.007

37.   Jarome TJ, Ferrara NC, Kwapis JL, Helmstetter FJ. CaMKII regulates proteasome
phosphorylation and activity and promotes memory destabilization following retrieval.
Neurobiol Learn Mem. 2016;128:103-109. doi:10.1016/j.nlm.2016.01.001

38.   Signor C, Temp FR, Mello CF, et al. Intrahippocampal infusion of spermidine improves
memory persistence: Involvement of protein kinase A. *Neurobiol Learn Mem*.
2016;131:18-25. doi:10.1016/j.nlm.2016.03.003

39.   Bhattacharya S, Kimble W, Buabeid M, et al. Altered AMPA receptor expression plays an
important role in inducing bidirectional synaptic plasticity during contextual fear memory
reconsolidation. *Neurobiol Learn Mem*. 2017;139:98-108. doi:10.1016/j.nlm.2016.12.013

40.   Schmidt SD, Furini CRG, Zinn CG, et al. Modulation of the consolidation and
reconsolidation of fear memory by three different serotonin receptors in hippocampus.
Neurobiol Learn Mem. 2017;142:48-54. doi:10.1016/j.nlm.2016.12.017

41.   Kumar T, Jha SK. Influence of cued-fear conditioning and its impairment on NREM sleep.
Neurobiol Learn Mem. 2017;144:155-165. doi:10.1016/j.nlm.2017.07.008

42.   Song Z, Chen H, Xu W, Wu S, Zhu G. Basolateral amygdala calpain is required for
extinction of contextual fear-memory. *Neurobiol Learn Mem*. 2018;155:180-188.
doi:10.1016/j.nlm.2018.08.004

43.   Maciejak P, Szyndler J, Lehner M, et al. The differential effects of protein synthesis
inhibition on the expression and reconsolidation of pentylenetetrazole kindled seizures.
Epilepsy Behav. 2010;18(3):193-200. doi:10.1016/j.yebeh.2010.04.005

44.   Abel T, Nguyen PV, Barad M, Deuel TAS, Kandel ER, Bourtchouladze R. Genetic
demonstration of a role for PKA in the late phase of LTP and in hippocampus-based long-
term memory. *Cell*. 1997;88(5):615-626. doi:10.1016/S0092-8674(00)81904-2

45.   Barrientos RM, Higgins EA, Sprunger DB, Watkins LR, Rudy JW, Maier SF. Memory for
context is impaired by injecting anisomycin into dorsal hippocampus following context
exploration. *Behav Brain Res*. 2002;134(1-2):299-306. doi:10.1016/S0166-
4328(02)00045-1

46.   Debiec J, LeDoux JE, Nader K. Cellular and systems reconsolidation in the hippocampus.
Neuron. 2002;36(3):527-538. doi:10.1016/S0896-6273(02)01001-2

47.   Huff NC, Rudy JW. The amygdala modulates hippocampus-dependent context memory
formation and stores cue-shock associations. *Behav Neurosci*. 2004;118(1):53-62.
doi:10.1037/0735-7044.118.1.53

48.   Biedenkapp JC, Rudy JW. Context memories and reactivation: constraints on the
reconsolidation hypothesis. *Behav Neurosci*. 2004;118(5):956-964. doi:10.1037/0735-
7044.118.5.956

49.   Rudy JW, Matus-Amat P. The ventral hippocampus supports a memory representation of
context and contextual fear conditioning: implications for a unitary function of the
hippocampus. *Behav Neurosci*. 2005;119(1):154-163. doi:10.1037/0735-7044.119.1.154

50.   Kochli DE, Campbell TL, Hollingsworth EW, et al. Combined administration of MK-801 and
cycloheximide produces a delayed potentiation of fear discrimination memory extinction.
Behav Neurosci. 2018;132(2):99-105. doi:10.1037/bne0000232

51.   Nader K, Schafe GE, Le Doux JE. Fear memories require protein synthesis in the amygdala
for reconsolidation after retrieval. *Nature*. 2000;406(6797):722-726.
doi:10.1038/35021052

52.   Trent S, Barnes P, Hall J, Thomas KL. Rescue of long-term memory after reconsolidation
blockade. *Nat Commun*. 2015;6:1-7. doi:10.1038/ncomms8897

53.   Dębiec J, Díaz-Mataix L, Bush DEA, Doyère V, Ledoux JE. The amygdala encodes specific
sensory features of an aversive reinforcer. *Nat Neurosci*. 2010;13(5):536-537.
doi:10.1038/nn.2520

54.   Mamou CB, Gamache K, Nader K. NMDA receptors are critical for unleashing
consolidated auditory fear memories. *Nat Neurosci*. 2006;9(10):1237-1239.
doi:10.1038/nn1778

55.   Nomura H, Matsuki N. Ethanol enhances reactivated fear memories.
NEUROPSYCHOPHARMACOLOGY. 2008;33(12):2912-2921. doi:10.1038/npp.2008.13

56. Yang CH, Huang CC, Hsu KS. Generalization of fear inhibition by disrupting hippocampal
protein synthesis-dependent reconsolidation process. *Neuropsychopharmacol Off Publ*
Am Coll Neuropsychopharmacol. 2011;36(10):1992-2008. doi:10.1038/npp.2011.87

57. Haubrich J, Crestani AP, Cassini LF, et al. Reconsolidation Allows Fear Memory to Be
Updated to a Less Aversive Level through the Incorporation of Appetitive Information.
NEUROPSYCHOPHARMACOLOGY. 2015;40(2):315-326. doi:10.1038/npp.2014.174

58. Chai N, Liu JF, Xue YX, et al. Delayed Noradrenergic Activation in the Dorsal Hippocampus
Promotes the Long-Term Persistence of Extinguished Fear.
NEUROPSYCHOPHARMACOLOGY. 2014;39(8):1933-1945. doi:10.1038/npp.2014.42

59. Awad W, Ferreira G, Maroun M. Dissociation of the Role of Infralimbic Cortex in Learning
and Consolidation of Extinction of Recent and Remote Aversion Memory.
Neuropsychopharmacol Off Publ Am Coll Neuropsychopharmacol. 2015;40(11):2566-
2575. doi:10.1038/npp.2015.103

60. Jarome TJ, Ferrara NC, Kwapis JL, Helmstetter FJ. Contextual Information Drives the
Reconsolidation-Dependent Updating of Retrieved Fear Memories.
Neuropsychopharmacol Off Publ Am Coll Neuropsychopharmacol. 2015;40(13):3044-
3052. doi:10.1038/npp.2015.161

61. Kwapis JL, Jarome TJ, Ferrara NC, Helmstetter FJ. Updating Procedures can Reorganize
the Neural Circuit Supporting a Fear Memory. *Neuropsychopharmacol Off Publ Am Coll*
Neuropsychopharmacol. Published online January 2017. doi:10.1038/npp.2017.23

62. Haubrich J, Machado A, Boos FZ, et al. Enhancement of extinction memory by
pharmacological and behavioral interventions targeted to its reactivation. *Sci Rep.*
2017;7(1):1-11. doi:10.1038/s41598-017-11261-6

63. Lattal KM, Abel T. Behavioral impairments caused by injections of the protein synthesis
inhibitor anisomycin after contextual retrieval reverse with time. *Proc Natl Acad Sci U S*
A. 2004;101(13):4667-4672. doi:10.1073/pnas.0306546101

64. Dębiec J, Doyère V, Nader K, LeDoux JE. Directly reactivated, but not indirectly
reactivated, memories undergo reconsolidation in the amygdala. *Proc Natl Acad Sci U S*
A. 2006;103(9):3428-3433. doi:10.1073/pnas.0507168103

65. Ballarini F, Moncada D, Martinez MC, Alen N, Viola H. Behavioral tagging is a general
mechanism of long-term memory formation. *Proc Natl Acad Sci U S A.*
2009;106(34):14599-14604. doi:10.1073/pnas.0907078106

66. Poulos AM, Ponnusamy R, Dong HW, Fanselow MS. Compensation in the neural circuitry
of fear conditioning awakens learning circuits in the bed nuclei of the stria terminalis.
Proc Natl Acad Sci U S A. 2010;107(33):14881-14886. doi:10.1073/pnas.1005754107

67. Hoeffer CA, Cowansage KK, Arnold EC, et al. Inhibition of the interactions between
eukaryotic initiation factors 4E and 4G impairs long-term associative memory
consolidation but not reconsolidation. *Proc Natl Acad Sci U S A.* 2011;108(8):3383-3388.
doi:10.1073/pnas.1013063108

68.   Radwanska K, Medvedev NI, Pereira GS, et al. Mechanism for long-term memory
formation when synaptic strengthening is impaired. *Proc Natl Acad Sci U S A*.
2011;108(45):18471-18475. doi:10.1073/pnas.1109680108

69.   de Carvalho Myskiw J, Furini CRG, Schmidt B, Ferreira F, Izquierdo I. Extinction learning,
which consists of the inhibition of retrieval, can be learned without retrieval. *Proc Natl*
Acad Sci U S A. 2015;112(2):E230-3. doi:10.1073/pnas.1423465112

70.   Roy DS, Muralidhar S, Smith LM, Tonegawa S. Silent memory engrams as the basis for
retrograde amnesia. *Proc Natl Acad Sci U S A*. 2017;114(46):E9972-E9979.
doi:10.1073/pnas.1714248114

71.   Kwapis JL, Jarome TJ, Schiff JC, Helmstetter FJ. Memory consolidation in both trace and
delay fear conditioning is disrupted by intra-amygdala infusion of the protein synthesis
inhibitor anisomycin. *Learn Mem Cold Spring Harb N*. 2011;18(11):728-732.
doi:10.1101/lm.023945.111

72.   Jarome TJ, Kwapis JL, Werner CT, Parsons RG, Gafford GM, Helmstetter FJ. The timing of
multiple retrieval events can alter GluR1 phosphorylation and the requirement for
protein synthesis in fear memory reconsolidation. *Learn Mem*. 2012;19(7):300-306.
doi:10.1101/lm.024901.111

73.   Einarsson EÖ, Nader K. Involvement of the anterior cingulate cortex in formation,
consolidation, and reconsolidation of recent and remote contextual fear memory. *Learn*
Mem. 2012;19(10):449-452. doi:10.1101/lm.027227.112

74.   Nakayama D, Yamasaki Y, Matsuki N, Nomura H. Post-retrieval late process contributes
to persistence of reactivated fear memory. *Learn Mem*. 2013;20(6):307-310.
doi:10.1101/lm.029660.112

75.   Remaud J, Ceccom J, Carponcy J, et al. Anisomycin injection in area CA3 of the
hippocampus impairs both short-term and long-term memories of contextual fear. *Learn*
Mem. 2014;21(6):311-315. doi:10.1101/lm.033969.113

76.   Kochli DE, Thompson EC, Fricke EA, Postle AF, Quinn JJ. The amygdala is critical for trace,
delay, and contextual fear conditioning. *Learn Mem*. 2015;22(2):92-100.
doi:10.1101/lm.034918.114

77.   Almeida-Corrêa S, Moulin TC, Carneiro CFD, Gonçalves MMC, Junqueira LS, Amaral OB.
Calcineurin inhibition blocks within-, but not between-session fear extinction in mice.
Learn Mem. 2015;22(3):159-169. doi:10.1101/lm.037770.114

78.   Girardi BA, Ribeiro DA, Signor C, et al. Spermidine-induced improvement of
reconsolidation of memory involves calcium-dependent protein kinase in rats. *Learn*
Mem. 2016;23(1):21-28. doi:10.1101/lm.039396.115

79.   Ferrara NC, Cullen PK, Pullins SP, Rotondo EK, Helmstetter FJ. Input from the medial
geniculate nucleus modulates amygdala encoding of fear memory discrimination. *Learn*
Mem. 2017;24(9):414-421. doi:10.1101/lm.044131.116

80.   Viosca J, De Armentia ML, Jancic D, Barco A. Enhanced CREB-dependent gene expression
increases the excitability of neurons in the basal amygdala and primes the consolidation

of contextual and cued fear memory. *Learn Mem*. 2009;16(3):193-197.
doi:10.1101/lm.1254209

81.   Stafford JM, Lattal KM. Direct comparisons of the size and persistence of anisomycin-
induced consolidation and reconsolidation deficits. *Learn Mem*. 2009;16(8):494-503.
doi:10.1101/lm.1452209

82.   Ageta H, Ikegami S, Miura M, et al. Activin plays a key role in the maintenance of long-
term memory and late-LTP. *Learn Mem*. 2010;17(4):176-185. doi:10.1101/lm.16659010

83.   Frankland PW, Ding HK, Takahashi E, Suzuki A, Kida S, Silva AJ. Stability of recent and
remote contextual fear memory. *Learn Mem*. 2006;13(4):451-457.
doi:10.1101/lm.183406

84.   Suzuki A, Mukawa T, Tsukagoshi A, Frankland PW, Kida S. Activation of LVGCCs and CB1
receptors required for destabilization of reactivated contextual fear memories. *Learn*
Mem. 2008;15(6):426-433. doi:10.1101/lm.888808

85.   Bal NV, Rysakova MP, Vinarskaya AK, Ivanova V, Zuzina AB, Balaban PM. Cued memory
reconsolidation in rats requires nitric oxide. *Eur J Neurosci*. 2017;45(5):643-647.
doi:10.1111/ejn.13503

86.   Duvarci S, Nader K, LeDoux JE. Activation of extracellular signal-regulated kinase-
mitogen-activated protein kinase cascade in the amygdala is required for memory
reconsolidation of auditory fear conditioning. *Eur J Neurosci*. 2005;21(1):283-289.
doi:10.1111/j.1460-9568.2004.03824.x

87.   Parsons RG, Gafford GM, Baruch DE, Riedner BA, Helmstetter FJ. Long-term stability of
fear memory depends on the synthesis of protein but not mRNA in the amygdala. *Eur J*
Neurosci. 2006;23(7):1853-1859. doi:10.1111/j.1460-9568.2006.04723.x

88.   Duvarci S, Mamou C ben, Nader K. Extinction is not a sufficient condition to prevent fear
memories from undergoing reconsolidation in the basolateral amygdala. *Eur J Neurosci*.
2006;24(1):249-260. doi:10.1111/j.1460-9568.2006.04907.x

89.   Sacchetti B, Sacco T, Strata P. Reversible inactivation of amygdala and cerebellum but not
perirhinal cortex impairs reactivated fear memories. *Eur J Neurosci*. 2007;25(9):2875-
2884. doi:10.1111/j.1460-9568.2007.05508.x

90.   Lee SH, Choi JH, Lee N, et al. Synaptic protein degradation underlies destabilization of
retrieved fear memory. *Science*. 2008;319(5867):1253-1256.
doi:10.1126/science.1150541

91.   Blum S, Runyan JD, Dash PK. Inhibition of prefrontal protein synthesis following recall
does not disrupt memory for trace fear conditioning. *BMC Neurosci*. 2006;7:67.
doi:10.1186/1471-2202-7-67

92.   Kishioka A, Uemura T, Fukushima F, Mishina M. Consolidation of auditory fear memories
formed by weak unconditioned stimuli requires NMDA receptor activation and de novo
protein synthesis in the striatum. *Mol Brain*. 2013;6:17. doi:10.1186/1756-6606-6-17

93.   Barnes P, Thomas KL. Proteolysis of proBDNF Is a Key Regulator in the Formation of
Memory. Bartolomucci A, ed. *PLoS ONE*. 2008;3(9):e3248.
doi:10.1371/journal.pone.0003248

94.   Apergis-Schoute AM, Dębiec J, Doyère V, LeDoux JE, Schafe GE. Auditory fear
conditioning and long-term potentiation in the lateral amygdala require ERK/MAP kinase
signaling in the auditory thalamus: A role for presynaptic plasticity in the fear system. *J*
Neurosci. 2005;25(24):5730-5739. doi:10.1523/JNEUROSCI.0096-05.2005

95.   Lopez J, Gamache K, Schneider R, Nader K. Memory retrieval requires ongoing protein
synthesis and NMDA receptor activity-mediated AMPA receptor trafficking. *J Neurosci*.
2015;35(6):2465-2475. doi:10.1523/JNEUROSCI.0735-14.2015

96.   Santini E, Ge H, Ren K, Peña De Ortiz S, Quirk GJ. Consolidation of fear extinction requires
protein synthesis in the medial prefrontal cortex. *J Neurosci*. 2004;24(25):5704-5710.
doi:10.1523/JNEUROSCI.0786-04.2004

97.   Kim J, Song B, Hong I, et al. Reactivation of Fear Memory Renders Consolidated Amygdala
Synapses Labile. *J Neurosci*. 2010;30(28):9631-9640. doi:10.1523/JNEUROSCI.0940-
10.2010

98.   Holehonnur R, Phensy AJ, Kim LJ, et al. Increasing the GluN2A/GluN2B ratio in neurons of
the mouse basal and lateral amygdala inhibits the modification of an existing fear
memory trace. *J Neurosci*. 2016;36(36):9490-9504. doi:10.1523/JNEUROSCI.1743-
16.2016

99.   Cai WH, Blundell J, Han J, Greene RW, Powell CM. Postreactivation glucocorticoids impair
recall of established fear memory. *J Neurosci*. 2006;26(37):9560-9566.
doi:10.1523/JNEUROSCI.2397-06.2006

100.   Duvarci S, Nader K. Characterization of fear memory reconsolidation. In: *Journal of*
Neuroscience. Vol 24. Society for Neuroscience; 2004:9269-9275.
doi:10.1523/JNEUROSCI.2971-04.2004

101.   Wilensky AE, Schafe GE, Kristensen MP, LeDoux JE. Rethinking the fear circuit: The
central nucleus of the amygdala is required for the acquisition, consolidation, and
expression of pavlovian fear conditioning. *J Neurosci*. 2006;26(48):12387-12396.
doi:10.1523/JNEUROSCI.4316-06.2006

102.   Mamiya N, Fukushima H, Suzuki A, et al. Brain region-specific gene expression activation
required for reconsolidation and extinction of contextual fear memory. *J Neurosci*.
2009;29(2):402-413. doi:10.1523/JNEUROSCI.4639-08.2009

103.   von Hertzen LSJ. Memory Reconsolidation Engages Only a Subset of Immediate-Early
Genes Induced during Consolidation. *J Neurosci*. 2005;25(8):1935-1942.
doi:10.1523/JNEUROSCI.4707-04.2005

104.   Fischer A, Sananbenesi F, Schrick C, Spiess J, Radulovic J. Distinct Roles of Hippocampal
De Novo Protein Synthesis and Actin Rearrangement in Extinction of Contextual Fear. *J*
Neurosci. 2004;24(8):1962-1966. doi:10.1523/JNEUROSCI.5112-03.2004

105. Suzuki A, Josselyn SA, Frankland PW, Masushige S, Silva AJ, Kida S. Memory
reconsolidation and extinction have distinct temporal and biochemical signatures. *J*
Neurosci. 2004;24(20):4787-4795. doi:10.1523/JNEUROSCI.5491-03.2004

106. Reis DS, Jarome TJ, Helmstetter FJ. Memory formation for trace fear conditioning
requires ubiquitin-proteasome mediated protein degradation in the prefrontal cortex.
Front Behav Neurosci. 2013;7(OCT):150. doi:10.3389/fnbeh.2013.00150

107. Kwak C, Choi JH, Bakes JT, Lee K, Kaang BK. Effect of intensity of unconditional stimulus
on reconsolidation of contextual fear memory. *Korean J Physiol Pharmacol Off J Korean*
Physiol Soc Korean Soc Pharmacol. 2012;16(5):293-296. doi:10.4196/kjpp.2012.16.5.293

108. Stiedl O, Palve M, Radulovic J, Birkenfeld K, Spiess J. Differential impairment of auditory
and contextual fear conditioning by protein synthesis inhibition in C57BL/6N mice. *Behav*
Neurosci. 1999;113(3):496-506. doi:10.1037/0735-7044.113.3.496

109. Bourtchouladze R, Abel T, Berman N, Gordon R, Lapidus K, Kandel ER. Different training
procedures recruit either one or two critical periods for contextual memory
consolidation, each of which requires protein synthesis and PKA. *Learn Mem*. 1998;5(4-
5):365-374. doi:10.1101/lm.5.4.365

110. Schafe GE, LeDoux JE. Memory consolidation of auditory pavlovian fear conditioning
requires protein synthesis and protein kinase A in the amygdala. *J Neurosci Off J Soc*
Neurosci. 2000;20(18):RC96-RC96. doi:10.1523/jneurosci.20-18-j0003.2000

111. Lattal KM, Abel T. Different requirements for protein synthesis in acquisition and
extinction of spatial preferences and context-evoked fear. *J Neurosci*. 2001;21(15):5773-
5780. doi:10.1523/jneurosci.21-15-05773.2001

112. Scharf MT, Woo NH, Lattal KM, Young JZ, Nguyen PV, Abel T. Protein Synthesis Is
Required for the Enhancement of Long-Term Potentiation and Long-Term Memory by
Spaced Training. *J Neurophysiol*. 2002;87(6):2770-2777. doi:10.1152/jn.2002.87.6.2770

113. Moher D, Liberati A, Tetzlaff J, et al. Preferred reporting items for systematic reviews and
meta-analyses: The PRISMA statement. *PLoS Med*. 2009;6(7).
doi:10.1371/journal.pmed.1000097

114. Davis HP, Squire LR. Protein synthesis and memory: A review. *Psychol Bull*.
1984;96(3):518-559. doi:10.1037/0033-2909.96.3.518

115. Nader K. Memory traces unbound. *Trends Neurosci*. 2003;26(2):65-72.
doi:10.1016/S0166-2236(02)00042-5

116. Vianna MR, Szapiro G, McGaugh JL, Medina JH, Izquierdo I. Retrieval of memory for fear-
motivated training initiates extinction requiring protein synthesis in the rat hippocampus.
Proc Natl Acad Sci U S A. 2001;98(21):12251-12254. doi:10.1073/pnas.211433298

117. Berman DE, Dudai Y. Memory extinction, learning anew, and learning the new:
Dissociations in the molecular machinery of learning in cortex. *Science*.
2001;291(5512):2417-2419. doi:10.1126/science.1058165

1078 118. Tiunova AA, Anokhin KV, Rose SPR. Two critical periods of protein and glycoprotein
1079   synthesis in memory consolidation for visual categorization learning in chicks. *Learn*
1080   *Mem*. 1998;4(5):401-410. doi:10.1101/lm.4.5.401

1081 119. Pedreira ME, Pérez-Cuesta LM, Maldonado H. Mismatch between what is expected and
1082   what actually occurs triggers memory reconsolidation or extinction. *Learn Mem*.
1083   2004;11(5):579-585. doi:10.1101/lm.76904

1084 120. Sevenster D, Beckers T, Kindt M. Prediction error demarcates the transition from
1085   retrieval, to reconsolidation, to new learning. *Learn Mem*. 2014;21(11):580-584.
1086   doi:10.1101/lm.035493.114

1087 121. Alfei JM, Monti RIF, Molina VA, Bueno AM, Urcelay GP. Prediction error and trace
1088   dominance determine the fate of fear memories after post-training manipulations. *Learn*
1089   *Mem*. 2015;22(8):385-400. doi:10.1101/lm.038513.115

1090 122. Eisenberg M, Kobilo T, Berman DE, Dudai Y. Stability of retrieved memory: inverse
1091   correlation with trace dominance. *Science*. 2003;301(5636):1102-1104.
1092   doi:10.1126/science.1086881

1093 123. Wang SH, De Oliveira Alvares L, Nader K. Cellular and systems mechanisms of memory
1094   strength as a constraint on auditory fear reconsolidation. *Nat Neurosci*. 2009;12(7):905-
1095   912. doi:10.1038/nn.2350

1096 124. Milekic MH, Alberini CM. Temporally graded requirement for protein synthesis following
1097   memory reactivation. *Neuron*. 2002;36(3):521-525. doi:10.1016/S0896-6273(02)00976-5

1098 125. Rotondo F, Biddle K, Chen J, Ferencik J, d'Esneval M, Milton AL. Lack of Effect of
1099   Propranolol on the Reconsolidation of Conditioned Fear Memory due to a Failure to
1100   Engage Memory Destabilisation. *Neuroscience*. 2022;480:9-18.
1101   doi:10.1016/j.neuroscience.2021.11.008

1102 126. Soeter M, Kindt M. High Trait Anxiety: A Challenge for Disrupting Fear Memory
1103   Reconsolidation. *PLOS ONE*. 2013;8(11). doi:10.1371/journal.pone.0075239

1104 127. Schimanski LA, Nguyen PV. Multidisciplinary approaches for investigating the
1105   mechanisms of hippocampus-dependent memory: A focus on inbred mouse strains.
1106   *Neurosci Biobehav Rev*. 2004;28(5):463-483. doi:10.1016/j.neubiorev.2004.04.002

1107 128. Macleod MR, Lawson McLean A, Kyriakopoulou A, et al. Risk of Bias in Reports of In Vivo
1108   Research: A Focus for Improvement. *PLOS Biol*. 2015;13(10):e1002273.
1109   doi:10.1371/journal.pbio.1002273

1110 129. Carneiro CFD, Moulin TC, Macleod MR, Amaral OB. Effect size and statistical power in the
1111   rodent fear conditioning literature - a systematic review. *PLoS ONE*. 2018.

1112 130. Tannenbaum C, Ellis RP, Eyssel F, Zou J, Schiebinger L. Sex and gender analysis improves
1113   science and engineering. *Nature*. 2019;575(7781):137-146. doi:10.1038/s41586-019-
1114   1657-6

1115 131. Clayton JA, Collins FS. Policy: NIH to balance sex in cell and animal studies. *Nature*.
1116   2014;509(7500):282-283.

1117   132.   Amaral OB, Neves K. Reproducibility: expect less of the scientific paper. *Nature*.
1118   2021;597(7876):329-331. doi:10.1038/d41586-021-02486-7

1119   133.   Bannach-Brown A, Hair K, Bahor Z, Soliman N, MacLeod M, Liao J. Technological
1120   advances in preclinical meta-research. *BMJ Open Sci*. 2021;5(1):e100131.
1121   doi:10.1136/bmjos-2020-100131

1122