Inference of gene-environment interaction from heterogeneous case-parent trios

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ABSTRACT

In genetic epidemiology, log-linear models of population risk may be used to study the effect of genotypes and exposures on the relative risk of a disease. Such models may also include gene-environment interaction terms that allow the genotypes to modify the effect of the exposure, or equivalently, the exposure to modify the effect of genotypes on the relative risk. When a measured test locus is in linkage disequilibrium with a causal locus, exposure-related genetic population structure can lead to spurious gene-environment interaction; that is, to apparent gene-environment interaction at the test locus in the absence of true gene-environment interaction at the causal locus. Exposure-related genetic population structure occurs when the distributions of exposures and of haplotypes at the test and causal locus both differ across population strata (i.e., population substructure). In a case-parent trio design researchers collect genotypes and exposures on affected children and genotypes on their parents. The design permits inference of genetic main effects while avoiding confounding bias due to population stratification. Unfortunately, when there is exposure-related genetic population structure, the protection against confounding bias for the main effect does not extend to the interaction term. We show that current methods to reduce the bias in estimated gene-environment interactions can only account for simple population structure involving two strata. To fill this gap, we propose to directly accommodate multiple population strata by adjusting for genetic principal components. We evaluate our approach through simulation and illustrate it on data from a study of genetic modifiers of cleft palate.

Keywords: Gene-environment interaction, case-parent trios, population stratification, genome-wide association study, cleft palate, principal components

1 INTRODUCTION

This work is concerned with a log-linear model of population disease risk that includes main effects for genotypes G, environmental exposures E, and a gene-environment interaction term $G \times E$. The $G \times E$ term allows genotypes to modify the effect of the exposure or, equivalently, the exposure to modify the effect of genotypes on the relative risk of developing the disease. We suppose throughout that G is an unmeasured causal locus in linkage disequilibrium with a measured non-causal test locus G', and that the distribution of GG' haplotypes differs across population strata (i.e. population substructure). Subpopulation-specific differences in the GG' haplotype frequencies can lead to differences in G' risk across the population strata where none exist for G [24]. Exposure-related genetic population structure occurs when the distribution of E also differs across the population strata [22]. Without some adjustment for the population strata, E will

tag the stratum-specific differences in G' risk (Figure 1), suggesting that E modifies G' risk, even in the absence of $G \times E$ [16, 22]; we refer to this as spurious $G' \times E$.

Figure 1. Schematic of log-GRRs for a non-causal test locus versus exposure in a structured population with two sub-populations, S=0 and S=1. Dashed lines represent log-GRRs within each sub-population. Horizontal positioning of these dashed lines indicates the support of the respective E distributions. High values of E are associated with S=1, in which one of the alleles at the test locus is associated with increased disease risk. Low values of E are associated with S=0 in which this same allele at the test locus is associated with low disease risk. Ignoring S yields the linear log-GRR curve indicated by the solid line, which erroneously suggests that E modifies the disease risk at the test locus.

The case-parent trio design permits inference of genetic main effects while avoiding confounding bias due to population stratification [21]. In this design, investigators collect information on G' and E in children affected with a disease of interest as well as the genotypes, G'_p, of their parents. Assuming G' and E are independent within families, a log-linear model of disease risk leads to a conditional likelihood for the G' and $G' \times E$ effects, based on the child's genotype given their exposure, affection status and parental genotypes [17]. Unfortunately, when there is exposure-related genetic population stratification, the protection against confounding bias for the main effect does not extend to the interaction term [16, 22]. Thus, spurious $G' \times E$ may be inferred from case-parent trio data in the absence of true $G \times E$.

Methods to mitigate this bias may be classified as design- or data-based. For a binary environmental exposure, the *design*-based tetrad approach of [16] augments the case-parent trio by adding the exposure of an unaffected sibling. These authors control the bias by including the sibship-averaged exposure in the

log-linear model. They show that all information about the interaction in the tetrad design comes from the siblings, not the parents [22]. Accordingly, they propose a sibling-augmented case-only design and analysis. By contrast, [17] takes a *data*-based approach, replacing the sibship-averaged exposure of [16] with the *predicted* exposure given ancestry. Predictions are obtained from a regression of exposure on principal components (PCs) computed from genetic markers that are unlinked to the test locus. This data-based approach may be applied to arbitrary exposures, including continuous exposures, and does not require siblings. However, its properties have not been evaluated in the case of more than two population strata.

We use the GENEVA Oral Cleft Study to motivate a new approach to unbiased inference of $G' \times E$ in case-parent trios. The analysis of [2] found multiple single nucleotide polymorphisms (SNPs) that appeared to modify the effect of maternal smoking, maternal alcohol consumption or maternal multivitamin supplementation on the risk of CP. The self-reported ancestry of the study sample is primarily European or East Asian, and all three exposures are more common in self-reported Europeans than in self-reported East Asians [2, Table 2]. If the frequencies of haplotypes spanning causal SNPs also vary by ancestral groups, exposure-related population stratification may lead to spurious gene-environment interaction. [11] focused on the self-reported Europeans and East Asians in the GENEVA Oral Cleft Study data. Applying the approach of [17], they confirmed the gene-environment interaction found by [2], and concluded that the evidence for gene-environment interaction is predominantly from the data of self-reported Europeans. These authors also considered whether exposure-related genetic population stratification *within* self-reported Europeans could explain the apparent $G' \times E$. Their results were inconclusive, however, possibly owing to the methodology's limitation to just two ancestry groups. In modern datasets, the possibility of both inter- and intra-continental stratification necessitates methods that can more flexibly accommodate multiple population strata. In this work we propose such an approach which relies on direct use of the genetic PCs to adjust for population strata.

The manuscript is structured as follows. In Section 2 we develop our direct PC-adjustment method and compare it to the indirect PC-based approach of [17]. In Section 3 we present simulations to evaluate the statistical properties of both approaches. In Section 4 we re-analyze the GENEVA data. Section 5 includes a discussion and areas for future work.

2 MODELS AND METHODS

2.1 Overview

We start with a log-linear model of disease risk parametrized in terms of genotype relative risks (GRRs) at a causal locus G. Under this model, $G \times E$ is equivalent to GRRs that depend on the exposure E. We then derive the GRRs at a non-causal test locus G' in linkage disequilibrium with G and show that, in the absence of $G \times E$, the G'-GRRs can depend on E when there is dependence between E and GG' haplotypes in the population. Such dependence can lead to spurious inference of $G' \times E$ in the absence of $G \times E$. However, valid inference is obtained if we adjust the risk model for any variable X for which E and GG' haplotypes are conditionally independent given X [17]. We review the rationale for the adjustment used by [17] in this context, and propose an alternative adjustment based on inferred population structure. In particular, we use the method of [5] to select a parsimonious set of PCs with which to adjust the risk model. A key question is whether the PC-selection method yields a set of PCs that provide enough adjustment to maintain type 1 error in the absence of $G \times E$, but not so much that we compromise power in the presence of $G \times E$. The Models and Methods section concludes with a discussion of the simulation methods used to answer this question.

2.2 Risk model and likelihood

Let $G = 0$, 1 or 2 denote the number of copies of the variant allele at the causal locus and E denote the exposure variable. The disease-risk model of [17] can be obtained from a log-linear model of the GRRs

$$\log GRR_g(e) = \log \frac{P(D=1|G=g, E=e)}{P(D=1|G=g-1, E=e)} = \beta_g + f_g(e) \quad \text{for } g = 1, 2, \tag{1}$$

and the log-disease risk for carriers of the baseline genotype $G = 0$

$$\log P(D=1|G=0, E=e) \equiv \eta(e).$$

The parameters β_g and $f_g(\cdot)$ are, respectively, genotype-specific main effects and functions that allow for $G \times E$ interaction. We can also write disease risk in terms of the baseline risk $\eta(e)$ and the GRRs as follows. First define $GRR_0(e) \equiv 1$. Next, note that

$$\frac{P(D=1|G=1, E=e)}{P(D=1|G=0, E=e)} = GRR_1(e) = GRR_1(e)GRR_0(e)$$

and

$$\begin{aligned} \frac{P(D=1|G=2, E=e)}{P(D=1|G=0, E=e)} &= \frac{P(D=1|G=2, E=e)}{P(D=1|G=1, E=e)} \frac{P(D=1|G=1, E=e)}{P(D=1|G=0, E=e)} \\ &= GRR_2(e)GRR_1(e)GRR_0(e). \end{aligned}$$

It follows that

$$P(D=1|G=g, E=e) = \eta(e) \prod_{i=0}^{g} GRR_i(e) \quad \text{for } g\text{=0, 1 or 2.} \tag{2}$$

A likelihood for estimation of the GRR parameters β_g and $f_g(\cdot)$, $g = 1, 2$, from case-parent trio data can be derived under the assumption that G and E are conditionally independent given parental genotypes G_p. As shown in Appendix A.1, the likelihood is based on the conditional probability of the child's genotype given their exposure and parental genotypes.

2.3 GRRs at a non-causal test locus

Let G' denote genotypes at a non-causal test locus in linkage disequilibrium with the causal locus G. We assume D and G' are conditionally independent given G and E, so that

$$P(D=1|G=g, G'=g', E=e) = P(D=1|G=g, E=e).$$

Therefore, the risk of disease given G' and E can be written as

$$P(D=1|G'=g', E=e) = \sum_{g=0}^{2} P(D=1|G=g, E=e)P(G=g|G'=g', E=e). \tag{3}$$

Equation (3) is a latent-class model [23] with the unobserved causal locus G as the latent class having probabilities $P(G=g|G'=g', E=e)$. Equations (3) and (2) enable the log-GRRs at G' to be written in

terms of the latent-class probabilities and the GRRs at G as follows:

$$\begin{aligned}
\log GRR_{g'}(e) &\equiv \log \frac{P(D=1|G'=g',E=e)}{P(D=1|G'=g'-1,E=e)} \\
&= \log \frac{\sum_{g=0}^{2} P(D=1|G=g,E=e)P(G=g|G'=g',E=e)}{\sum_{g=0}^{2} P(D=1|G=g,E=e)P(G=g|G'=g'-1,E=e)} \\
&= \log \frac{\sum_{g=0}^{2}(\prod_{i=0}^{g} GRR_i(e))P(G=g|G'=g',E=e)}{\sum_{g=0}^{2}(\prod_{i=0}^{g} GRR_i(e))P(G=g|G'=g'-1,E=e)}. \qquad (4)
\end{aligned}$$

Without $G \times E$, GRRs at G do not depend on E. Importantly, though, the log-GRRs at G' *can* depend on E through the latent-class probabilities $P(G=g|G'=g',E=e)$. In fact, as shown in Appendix A.2, these latent-class probabilities will depend on E whenever GG' haplotypes and E are associated, as happens when the population has exposure-related genetic stratification. Since $G' \times E$ is equivalent to $GRR_{g'}$ varying with E, equation (4) gives insight into how exposure-related genetic stratification creates spurious $G' \times E$.

2.4 Augmented risk model

The development so far has considered a disease-risk model that depends only on E and a causal locus G. We now consider an augmented disease-risk model that depends on E, G and a third variable X:

$$\log GRR_g(e,x) \equiv \log \frac{P(D=1|G=g,E=e,X=x)}{P(D=1|G=g-1,E=e,X=x)} = \beta_g + f_g(e,x) \quad \text{for } g\text{=1,2},$$

where β_g and $f_g(\cdot,x)$ are, respectively, genotype specific main effects and functions that allow for $G \times E \times X$ interaction. Defining

$$GRR_0(e,x) \equiv 1,$$

an analogous development to Section 2.3 leads to the following X-adjusted log-GRRs at G':

$$\begin{aligned}
\log GRR_{g'}(e,x) &\equiv \log \frac{P(D=1|G'=g',E=e,X=x)}{P(D=1|G'=g'-1,E=e,X=x)} \\
&= \log \frac{\sum_{g=0}^{2}(\prod_{i=0}^{g} GRR_i(e,x))P(G=g|G'=g',E=e,X=x)}{\sum_{g=0}^{2}(\prod_{i=0}^{g} GRR_i(e,x))P(G=g|G'=g'-1,E=e,X=x)}. \qquad (5)
\end{aligned}$$

In the next section we discuss choices for X that eliminate E from the latent-class probabilities for G, and hence eliminate the spurious interaction due to exposure-related genetic stratification discussed in the previous section.

2.5 Removing dependence of the latent-class probabilities on E

The diagram in Figure 2 depicts the dependence between GG' haplotypes and E from exposure-related genetic stratification of the population. In the figure, S is a categorical variable that indicates population strata. The categorical variable X_E is a "coarsening" of S such that different levels of X_E correspond to different E distributions, and, similarly, $X_{GG'}$ is a coarsening of S such that different levels of $X_{GG'}$ correspond to different GG' haplotype distributions.

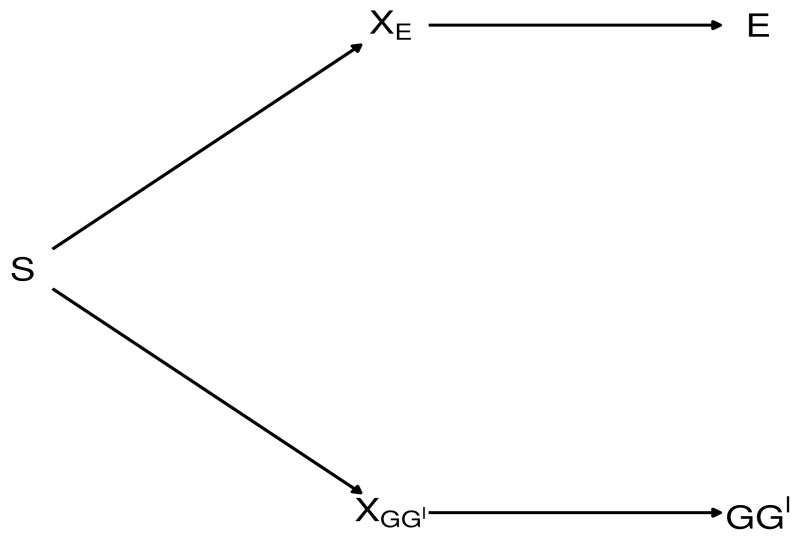

Figure 2. Diagram depicting exposure-related genetic stratification. The latent population strata S induce dependence beetween E and GG'. Latent factors X_E and $X_{GG'}$ encode different distributions of E and GG', respectively. E and GG' are conditionally independent given any of the three variables that lie on the path between them.

The path connecting E and GG' in Figure 2 is said to be *blocked* by each of the variables X_E, S and $X_{GG'}$ [10, Definition 1]. Therefore, E and GG' are conditionally independent given any of X_E, S or $X_{GG'}$ [9]. As shown in Appendix A.2, a consequence is that conditioning on any of these variables removes the dependence of the latent-class probabilities on E. That is, letting X denote any of X_E, S or $X_{GG'}$, $P(G = g|G' = g', E = e, X = x) = P(G = g|G' = g', X = x)$. Consequently, from equation (5),

$$\begin{aligned} \log GRR_{g'}(e,x) &\equiv \log \frac{P(D=1|G'=g', E=e, X=x)}{P(D=1|G'=g'-1, E=e, X=x)} \\ &= \log \frac{\sum_{g=0}^{2}(\prod_{i=0}^{g} GRR_i(e,x))P(G=g|G'=g', X=x)}{\sum_{g=0}^{2}(\prod_{i=0}^{g} GRR_i(e,x))P(G=g|G'=g'-1, X=x)}. \end{aligned} \tag{6}$$

GRRs at G' will thus depend on E if and only if GRRs at G do.

2.6 Linear approximation to the $\log GRR_{g'}$'s

From equation (6) we see that, for fixed g' and x, $\log GRR_{g'}(e,x)$ varies with e if and only if the $GRR_g(e,x)$ do. We can therefore test for $G \times E$ by fitting a model for $\log GRR_{g'}(e,x)$ that allows separate curves in e for each combination of g' and x [18]. We take these curves to be straight lines, and test whether any of these have non-zero slope.

2.7 Choice of X

Following [16], [17] set X to be the categorical variable X_E that distinguishes E distributions among the genetic strata of the population. Since X_E is unobserved, [17] consider the expectation of E given genetic markers (EEGM) as a surrogate $\hat{X}_E$. The idea behind their EEGM approach is to distinguish exposure distributions by their mean, which may vary across genetic strata, S. Though S is not known, it is

reflected in principal components (PCs), $\hat{S}$, computed from a set of genetic markers that are unlinked to G'. The expectation of E given $\hat{S}$ can be estimated by linear regression of E on $\hat{S}$ when E is continuous, or by logistic regression when E is binary. For EEGM adjustment, the expected exposure within genetic strata is estimated by $\hat{X}_E = \widehat{E(E|\hat{S})}$. [17] showed that EEGM adjustment works well where there are two population strata, but our simulation results (Section 3) indicate that it works poorly for more than two strata. We therefore propose to adjust for population strata directly; i.e., to take $X = S$.

Suppose the population has $K + 1$ genetic strata, indexed $0, \ldots, K$, and let S denote a vector of K dummy variables that distinguish these strata such that that the kth element $S_k = 1$ for trios in stratum $k > 0$ and 0 otherwise, for $k = 1, \ldots, K$. The log-linear GRR model for G' is

$$\log GRR_{g'}(e, s) = \beta_{g'} + \sum_{k=1}^{K} \beta_{g'k} s_k + \beta_{g'E} \times e + \sum_{k=1}^{K} \beta_{g'Ek} s_k \times e; \quad g' = 1, 2. \tag{7}$$

This model may be interpreted as follow. For trios with $G' = g'$ in genetic stratum 0, the log-GRR curve is linear with intercept $\beta_{g'}$ and slope $\beta_{g'E}$. For trios with $G' = g'$ in geneteic stratum k, $k = 1, \ldots, K$, the log-GRR curve is linear with intercept $\beta_{g'} + \beta_{g'k}$ and slope $\beta_{g'E} + \beta_{g'Ek}$.

To test for $G' \times E$ based on the model (7) we use a likelihood-ratio test of the null hypothesis that all curves have zero slope,

$$H_0 : \beta_{g'E} = \beta_{g'E1} = \cdots = \beta_{g'EK} = 0; \quad g' = 1, 2, \tag{8}$$

versus the alternative hypothesis that at least one of the slope parameters is non-zero.

2.8 Inferred population strata

The population stratum variable S reflects genetic ancestry and is not generally known. Since adjustment for self-reported ancestry can lead to bias [20] we use marker-based PCs, $\hat{S}$, to distinguish population strata in equation (7).

Standard PC adjustment in genetic association analyses relies on a relatively large set of PCs. From equation (8), the degrees of freedom of the test for $G' \times E$ is equal to $2(K + 1)$. Thus, using more PCs than are necessary reduces the power of the test for $G' \times E$. We seek methods to select a parsimonious set of PCs that provides enough adjustment to control type 1 error rate, without sacrificing power. We consider three PC-selection methods. The first [25] is an automated version of the graphical approach of looking for an "elbow" in the scree plot of variance explained by the PCs as a function of their number. The second [5] is an estimator of the rank of a matrix under a model in which the data matrix is a noisy version of a low-rank matrix. The third [8] is to select PCs corresponding to eigenvalues that exceed a significance threshold determined from the distribution of the largest eigenvalue of an unstructured random matrix.

2.9 Simulation methods

2.9.1 Simulating G, G' and E on case-parent trios

To study the statistical properties of our proposed approach and compare it to the method of [17], we generated 5000 data sets of 3000 informative case-parent trios. Trios were sampled from one of four population strata labelled $S = 0$, 1, 2 or 3. We assumed random mating within and no mixing between strata. We performed some simulations using equal-sized strata and others using unequal-sized strata. In

the case of unequal stratum sizes, the split was 60%, 40% for two strata; 50%, 30% and 20% for three strata; and 40%, 30%, 20% and 10% for four strata.

For a given stratum, informative trios were simulated following the methods proposed by [19, 18]. Briefly, GG' haplotypes are first simulated on parents in a random-mating population according to the stratum-specific GG' haplotype distributions in Table 1. Child haplotypes are then simulated following Mendel's laws and assuming no recombination between G and G'. The child's exposure E is also simulated according to the stratum-specific distributions described below. Finally, the child's disease status is simulated based on the disease-risk model (1). Trios with an affected child and at least one heterozygous parent at the test locus are retained. The data recorded on each trio are G'_p, G', and E, where G'_p is the pair of parental genotypes at the test locus.

Spurious $G' \times E$ is induced by specifying different distributions of E and GG' haplotypes in the four strata of Table 1. The GG' distributions for strata $S = 0$ and $S = 1$ are as in [17]. Alleles at G are denoted R (risk) and N (non-risk), while alleles at G' are denoted 1 and 0. We summarize the haplotype distributions by the implied allelic correlations between the index alleles R and 1. Under the GG' haplotype frequencies given in Table 1, these correlations are $r_0 = -1$ in stratum $S = 0$, $r_1 = 1$ in stratum $S = 1$, $r_2 = 0.5$ in stratum $S = 2$ and $r_3 = -0.5$ in stratum $S = 3$.

Table 1. GG' haplotype frequencies in four population strata.

	Stratum			
GG'	$S = 0$	$S = 1$	$S = 2$	$S = 3$
R1	0.0	0.5	0.375	0.125
R0	0.5	0.0	0.125	0.375
N1	0.5	0.0	0.125	0.375
N0	0.0	0.5	0.375	0.125

The stratum-specific distributions of E are chosen to be normal with common variance $\sigma^2 = 0.36$, and means $\mu_0 = -0.8$, $\mu_1 = 0.8$, $\mu_2 = 2.4$ and $\mu_3 = 4.0$ in strata 0, 1, 2 and 3, respectively. The E distributions for strata $S = 0$ and $S = 1$ are as in [17].

The disease-risk model is specified as follows. The genetic main effect is $\beta_g = log(3)/2$ for $g = 1, 2$, corresponding to a $\sqrt{3}$-fold increase in relative risk for each copy of the risk allele (R) in the absence of $G \times E$. To evaluate the type 1 error rate of the $G \times E$ test we set $f_g(e) = 0$ in our simulations. To investigate power we choose a linear interaction model for the $G \times E$ term, setting $f_g(e) = \beta_{gE} e$ with $\beta_{gE} = -0.10, -0.15, -0.20$ or -0.25.

2.9.2 Simulating markers for PC adjustment

A standard method of PC adjustment is to calculate PCs from a genomic region that is unlinked to the test locus. It is recommended that markers in this region be thinned, or LD pruned, to have pairwise correlations of $r^2 \leq 0.1$ [6]. We simulated such panels of markers based on data from the 1000 genomes project [4] using two East Asian (Chinese Dai in Xishuangbanna, China [CDX] and Han Chinese in Bejing China

[CHB]) and two European (Iberian population in Spain [IBS] and Finnish in Finland [FIN]) populations. From the initial download of the genome-wide data, we retained 6,929,035 diallelic, autosomal markers with minor allele frequency (MAF) 0.05 or greater in all four of the population groups.

Our initial approach to simulating markers for a given population population stratum was to fit a hidden Markov model (HMM) to the haplotypes in that stratum, chromosome by chromosome, using fastPHASE [14], and use this fitted model to simulate individual multilocus genotypes using SNPknock [15]. The simulated data are then LD pruned and principal components are computed from the thinned panel of markers. However, the computation involved in this approach proved to be prohibitive. For example, fitting the HMMs took up to 5 hours per chromosome. We therefore considered two computationally cheaper alternatives. In the first alternative, we started from an LD-pruned set of markers in the original data and fit HMMs to this set. In the second alternative, we used the same panel of pruned markers, but simulated genotypes *independently* based on the MAFs in the population strata. In what follows we refer to the first and second alternatives as *LD-based* and *independent* marker simulation, respectively.

Independent markers could contain more information about the population strata than markers in LD. As a result, PC adjustment with independent markers might control type 1 error more effectively than adjustment with markers in LD. To assess this possibility, we completed 100 preliminary simulation replicates using LD-based marker simulation and 5000 replicates using independent marker simulation. We simulated trios from four population strata under the null hypothesis of no $G \times E$, used the PC selection method of [5] to adjust the risk model and estimated the resulting type 1 error rates. Estimated type 1 error rates and their 95% confidence intervals under the LD-based and independent simulation methods were 0.04 (0.002, 0.078) and 0.0496 (0.044, 0.056), respectively, and consistent with similar type 1 error rates for the two approaches. We therefore used the faster simulation of independent markers for the simulation study.

In Sections 3.2 and 3.3 we present type I error and power results for two, three or four population strata. For two strata ($S = 0$ and $S = 1$), marker simulations were based on the CHB and IBS population groups. For three strata ($S = 0$, $S = 1$ and $S = 2$), simulations were based on the CHB, IBS and CDX population groups.

3 RESULTS

3.1 Selection of Principal Components

All PC selection methods performed well when the population stratum sizes are equal (results not shown), but not when the stratum sizes are unequal. We illustrate with simulation results involving datasets of 3000 trios sampled from four unequal-sized strata. For $K + 1 = 4$ populations we require $K = 3$ PCs. In 5000 simulation replicates, the method of [5] always selected three, the method of [25] always selected one, and the method of [8] selected three PCs 4942 times and four PCs 58 times. Other simulation results with unequal-sized strata (not shown) yielded similar results. Therefore, in what follows we use the method of [5] to select PCs.

3.2 Type I Error Rate

We compared the type I error rates of the test for $G' \times E$ using (i) adjustment with the true stratum membership S, (ii) the EEGM adjustment of [17], and (iii) PC adjustment. Results for simulated datasets with equal or unequal stratum sizes are shown in Table 2. For both equal and unequal stratum sizes, adjustment by S or PCs maintains the nominal 5% error rate regardless of the number of strata. By contrast, EEGM adjustment leads to an inflated type 1 error rate when there are more than two strata. In light of

the inflated type 1 error rate of the test, we do not consider EEGM adjustment in the following section on power.

Table 2. Estimated type 1 error rates (top entry) and corresponding 95% confidence intervals (bottom entry) when data are simulated from 2, 3 or 4 strata with equal (top three rows) or unequal (bottom three rows) stratum sizes

Equal stratum sizes

	Number of strata		
Adjustment	2	3	4
S	0.0556 (0.049, 0.062)	0.0524 (0.046, 0.0586)	0.0498 (0.044, 0.056)
EEGM	0.0538 (0.048, 0.060)	1.0000 NA	1.0000 NA
PC	0.0546 (0.048, 0.061)	0.0534 (0.047, 0.060)	0.0496 (0.044, 0.056)

Unequal stratum sizes

	2	3	4
S	0.0524 (0.046, 0.058)	0.0482 (0.042 0.054)	0.0536 (0.047,0.059)
EEGM	0.0536 (0.047, 0.060)	1.0000 NA	1.0000 NA
PC	0.0540 (0.048, 0.060)	0.0508 (0.045, 0.057)	0.0527 (0.046, 0.059)

3.3 Power

Table 3 provides a comparison of estimated power when data are simulated from two, three or four strata. Results are shown for simulations using both equal and unequal stratum sizes and for different values of the $G \times E$ effect. From these results we see that power increases with effect size, decreases with number of strata and tends to be slightly larger for unequal strata than equal strata. Importantly, the estimated power under PC adjustment is always within simulation error of that under adjustment for true stratum membership.

Table 3. Estimated power (top entry) and corresponding 95% confidence intervals (bottom entry) of different adjustment schemes for different $G \times E$ interaction effects β_{gE}, number of strata and stratum-size distributions.

Equal stratum sizes

		β_{gE}			
Num. Strata	Adjustment	-0.10	-0.15	-0.20	-0.25
2	S	0.2602	0.5660	0.8420	0.9558
		(0.248, 0.272)	(0.552, 0.580)	(0.832, 0.852)	(0.950, 0.961)
	PC	0.2580	0.5660	0.8404	0.9564
		(0.246, 0.270)	(0.552, 0.580)	(0.830, 0.850)	(0.951, 0.962)
3	S	0.1742	0.3844	0.6498	0.8288
		(0.164, 0.185)	(0.371, 0.398)	(0.636, 0.663)	(0.818, 0.839)
	PC	0.1788	0.3920	0.6616	0.8316
		(0.168, 0.189)	(0.378, 0.406)	(0.648, 0.675)	(0.821, 0.842)
4	S	0.1306	0.2766	0.5010	0.6970
		(0.121, 0.140)	(0.264, 0.289)	(0.487, 0.515)	(0.684, 0.710)
	PC	0.1396	0.2936	0.5088	0.6918
		(0.130, 0.149)	(0.281, 0.306)	(0.495, 0.523)	(0.679, 0.704)

Unequal stratum sizes

		β_{gE}			
		-0.10	-0.15	-0.20	-0.25
2	S	0.2636	0.5724	0.8328	0.9518
		(0.251, 0.276)	(0.559, 0.586)	(0.822, 0.843)	(0.946, 0.958)
	PC	0.2648	0.5722	0.8322	0.9514
		(0.252, 0.277)	(0.558, 0.586)	(0.822, 0.842)	(0.945, 0.957)
3	S	0.1950	0.4322	0.7082	0.8640
		(0.184, 0.206)	(0.418, 0.446)	(0.696, 0.721)	(0.854, 0.874)
	PC	0.1936	0.4334	0.7054	0.8632
		(0.183, 0.204)	(0.420, 0.447)	(0.693, 0.718)	(0.854, 0.873)
4	S	0.1614	0.3470	0.6028	0.7894
		(0.151, 0.172)	(0.334, 0.360)	(0.589, 0.616)	(0.778, 0.801)
	PC	0.1598	0.3380	0.5872	0.7820
		(0.150, 0.170)	(0.325, 0.351)	(0.574, 0.601)	(0.770, 0.794)

4 THE GENEVA ORAL CLEFT STUDY

4.1 Data and objectives

The GENEVA Oral Cleft study [1] is comprised of 550 case-parent trios from 13 different sites across the United States, Europe, Southeast and East Asia. Data were obtained through dbGAP at `https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000094.v1.p1` with accession number phs000094.v1.p1. Of the 550 trios, only 462 were available for analysis. Summaries of the trios by ancestry and gender of the affected child are shown in Table 4. From this table we see the ancestry of the sample is predominantly European (46%) and East Asian (51%).

Table 4. Gender of 462 affected children by self-reported ancestry

Ancestry	Males	Females	Total	%
European	103	111	214	46%
Asian	93	141	234	51%
Other/Afr	3	11	14	3%
Total	199	263	462	100%

The objective of the GENEVA study is to discover genetic contributions to orofacial clefts, the most common type of craniofacial birth defect in humans, and to assess whether these genes modify the effect of exposures known to be associated with cleft palate. Maternal exposure to multivitamins, alcohol and smoking were assessed through maternal interviews focused on the peri-conceptual period (3 months prior to conception through the first trimester), which includes the first 8-9 weeks of gestation when palatal development is completed. Exposure status is summarized in Table 5. From this table we see that the three dichotomous exposures are all more common in Europeans. In contrast to the continuous exposures of the simulation study, the exposures we consider in the GENEVA study are all dichotomous.

Table 5. Exposure rates for maternal alcohol consumption, maternal smoking and maternal vitamin supplementation by self-reported ancestry in affected trios.

	Percent exposed to Maternal			
Ancestry	Alcohol	Smoking	Vitamin Supp.	Affected children
European	41%	28%	57%	214
East Asian	4%	3%	21%	234
Other/Afr	14%	7%	71%	14
Total	21%	14%	39%	462

4.2 GENEVA data analysis

4.2.1 PC selection

LD pruning of the genome-wide panel of SNPs at an r^2 threshold of 0.1 yielded 63,694 markers. In a principal component analysis of these markers, the first PC explains 6.3% of the total variance and all others explain less than 0.4%. Not surprisingly, the method of [5] selects one PC. A plot of the projections of the data onto the first PC by self-reported ancestry is shown in Figure 3, with points colored by self-reported ancestry. We see that the first PC distinguishes those with self-reported East Asian ancestry from those with self-reported European ancestry.

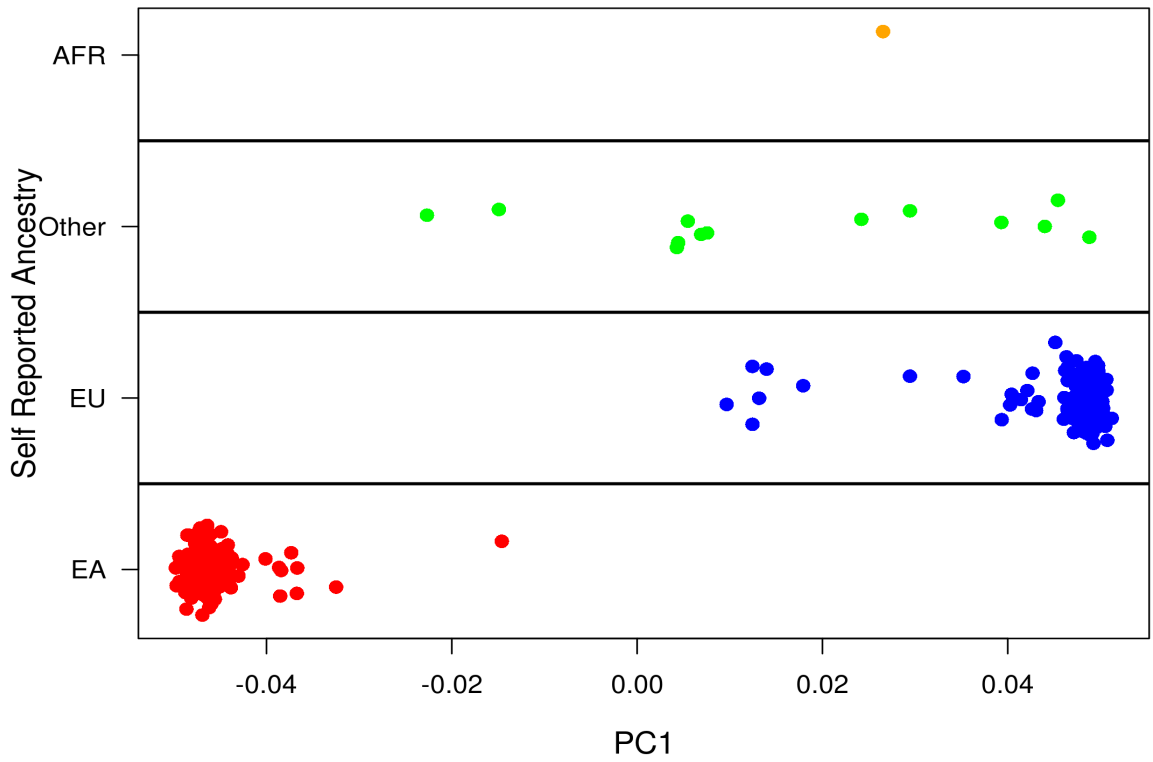

Figure 3. Projections of each affected child onto the first PC by self-reported ancestry: red=East Asian (234 trios), blue=European (214 trios), orange=African (one trio) and green=multiple ancestry/other (13 trios)

4.2.2 Inference of $G \times E$

The conditional-likelihood methods outlined in Appendix A.1 were applied to the data. We focused on inference of $G \times E$ between maternal alcohol consumption and the six SNPs in the *MLLT3* gene that had significant $G \times E$ at the 5% level in the analysis of [2]. Table 6 shows the results of the three different tests of $G' \times E$. Following [2], each test is based on an additive genetic model that specifies equal log-GRRs for genotypes $g' = 1$ or 2. Results based on fitting a more general co-dominant model (1) were similar (results not shown). The first test, as in [2], makes no adjustment for exposure-related genetic stratification

in the population, the second uses EEGM adjustment and the third uses PC adjustment. From the table we see that, for each test SNP, p-values are smallest when we make no adjustment. Comparing the EEGM and PC adjustment approaches we find that p-values from PC adjustment are similar to, but tend to be slightly smaller than, those from the EEGM adjustment. Of the six test SNPs show in the table, four retain significance at the 5% level after adjustment for exposure-related genetic stratification.

Table 6. P-values from the analysis of the GENEVA data at 6 SNPs in the MLLT3 gene (Chr 9) that showed significant interaction with maternal alcohol consumption in [2]. Tests are based on fitting an additive genetic model and use (i) no adjustment, (ii) EEGM adjustment or (iii) PC adjustment to control for exposure-related genetic stratification in the population.

SNP	no adj.	EEGM	PC
rs4621895	0.0006	0.0047	0.0037
rs4977433	0.0003	0.0036	0.0028
rs6475464	0.0104	0.0158	0.0139
rs668703	0.0008	0.0032	0.0025
rs623828	0.0481	0.1368	0.1384
rs2780841	0.0417	0.1613	0.1471

5 DISCUSSION

We consider a log-linear model of GRRs at a causal locus G. Under this model, $G \times E$ is equivalent to GRRs that vary with the exposure E. We show that exposure-related genetic stratification in the population can lead to spurious $G' \times E$ at a non-causal test locus G' in LD with G. However, valid inference of $G' \times E$ can be obtained by augmenting the GRR model with a blocking variable X, such that GG' haplotypes and E are conditionally independent given X. We discuss the choice of X for inference of $G' \times E$ when data are collected from a study of case-parent trios. The population strata S would be an ideal choice for X but is not known. We propose to use principal components (PCs) instead. In particular, we calculate PCs from a genomic region unlinked to the test locus and select a parsimonious subset using the method of [5]. Through simulations, we show that our PC adjustment maintains the nominal type-1 error rate and has nearly identical power to detect $G \times E$ as an oracle approach based directly on S. We illustrate our approach by applying it to an analysis of real data from case-parent trios in the GENEVA Oral Cleft Study.

To reduce bias from exposure-related genetic stratification, direct PC adjustment has advantages over the EEGM approach and design-based strategies such as the tetrad approach of [16] and the sibling-augmented case-only approach of [22]. Unlike the EEGM approach, PC adjustment controls the type 1 error when there are more than two population strata. Unlike the design-based strategies, PC adjustment does not require siblings nor assume binary exposures.

Development of alternative approaches based on propensity scores is an area for future work. The EEGM approach is attractive in that it reduces the genetic principal components to a single score, $E(E|\hat{S})$. For binary exposures, such as those in the GENEVA study, the EEGM is a propensity score [13]. For continuous exposures, such as those in the simulation study, the analog to the EEGM is a continuous-treatment propensity score [3]. With continuous exposures, we could predict E given the genetic markers and *then* convert the predictions to a Normal density score that takes low values for predictions far from their

observed value. These density scores could be used either as predictors [7] or weights [12] in subsequent analyses. It would be interesting to explore the use of propensity-score methods in inference of $G' \times E$ from case-parent trios with continuous exposures, particularly when there are more than two population strata.

CONFLICT OF INTEREST STATEMENT

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

AUTHOR CONTRIBUTIONS

PR developed the statistical methods, performed the simulations and data analyses, and wrote the initial draft of the manuscript. JG and BM conceptualized the study and revised the manuscript. All authors proofread and approved the final version of the manuscript.

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DATA AVAILABILITY STATEMENT

Datasets used for the analyses described in this manuscript were obtained from dbGaP at `https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000094.v1.p1` through dbGaP accession number phs000094.v1.p1.

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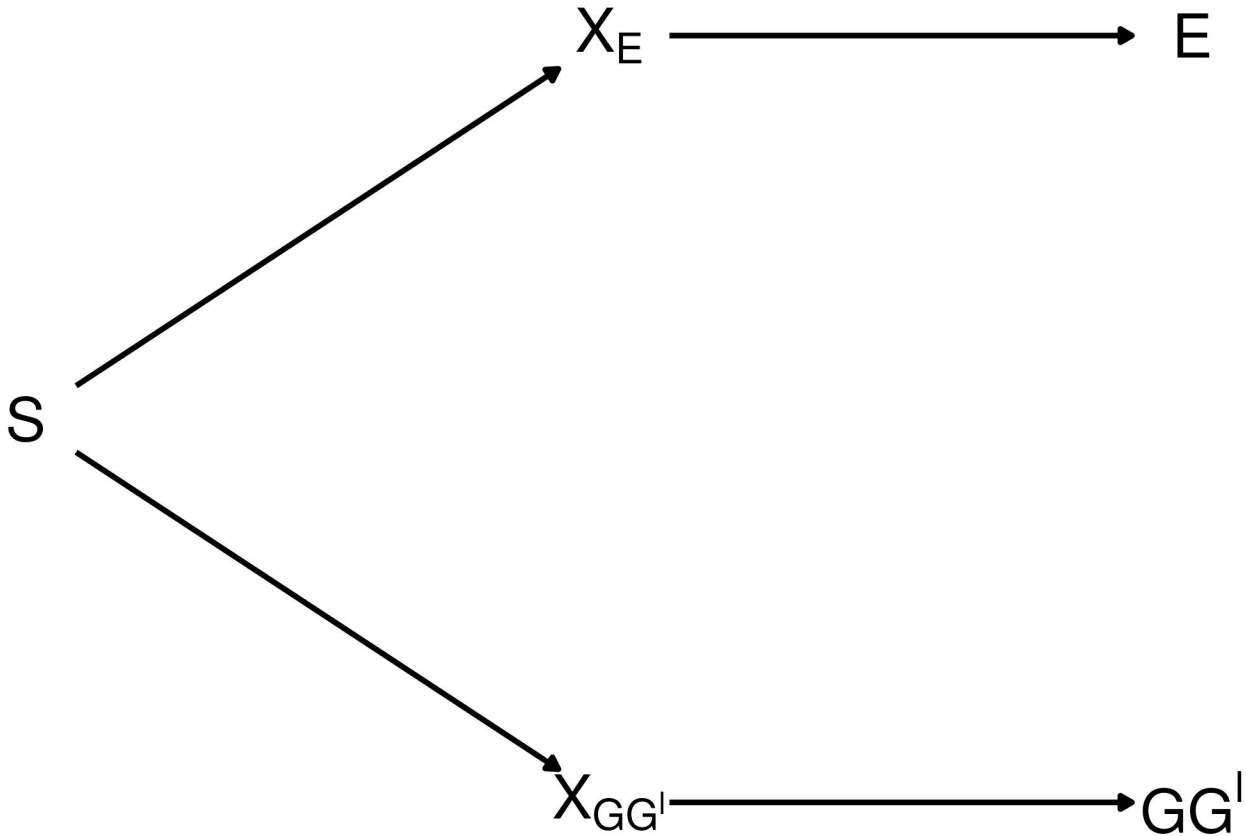
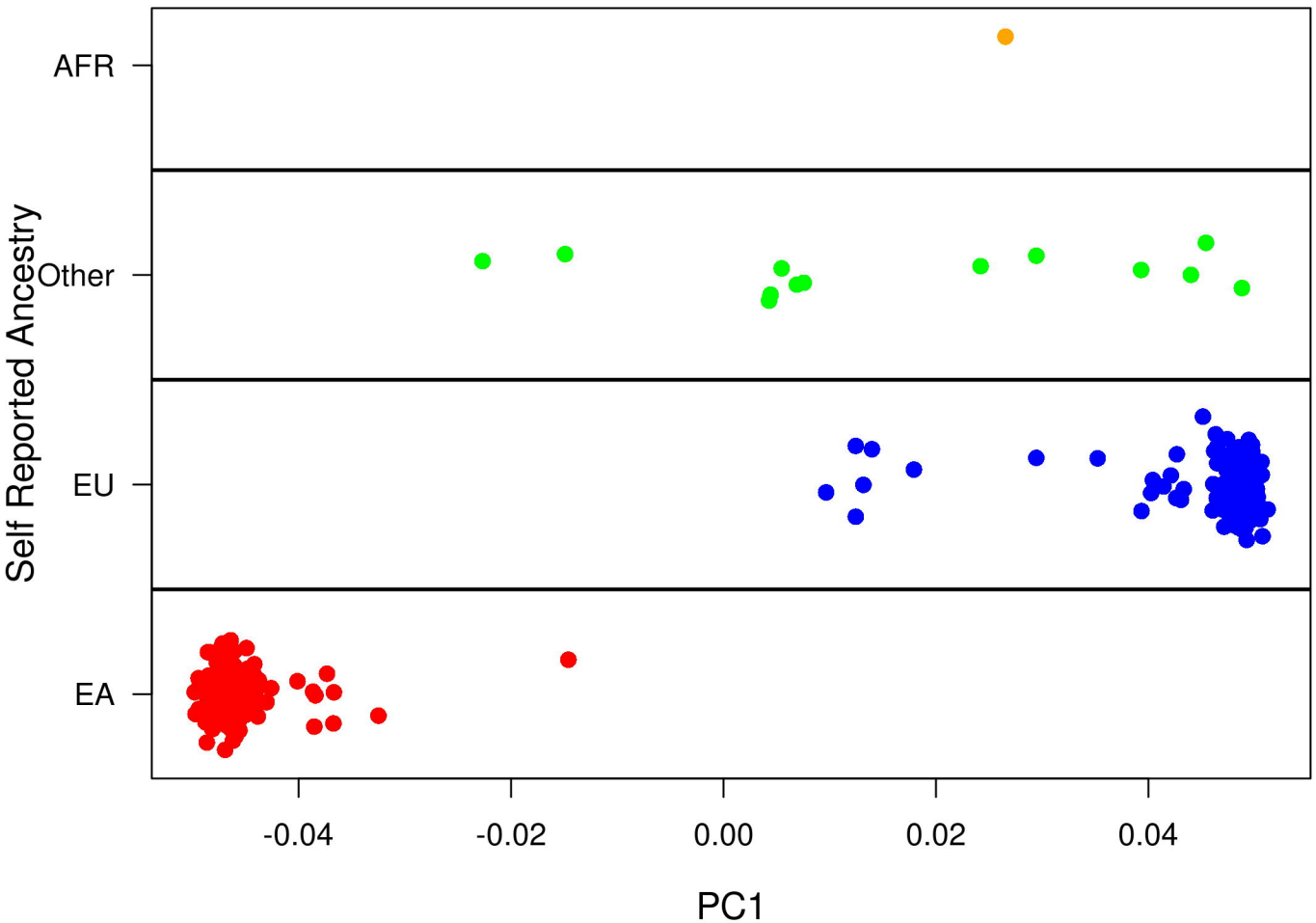
S=1
S=0
Log-GRR for Test Locus
Sub-population specific Exposure


X_E
E
S
$X_{GG'}$
GG'


AFR
Other
EU
EA
Self Reported Ancestry
-0.04
-0.02
0.00
0.02
0.04
PC1