Variability of visual field maps in human early extrastriate cortex challenges
the canonical model of organization of V2 and V3

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Abstract

Visual field maps in human early extrastriate areas (V2 and V3) are traditionally

thought to form mirror-image representations which surround the primary visual cortex

(V1). According to this scheme, V2 and V3 form nearly symmetrical halves with

respect to the calcarine sulcus, with the dorsal halves representing lower contralateral

quadrants, and the ventral halves representing upper contralateral quadrants. This

arrangement is considered to be consistent across individuals, and thus predictable

with reasonable accuracy using templates. However, data that deviate from this

expected pattern have been observed, but mainly treated as artifactual. Here we

systematically investigate individual variability in the visual field maps of human early

visual cortex using the 7T Human Connectome Project (HCP) retinotopy dataset. Our

results demonstrate substantial and principled inter-individual variability. Visual field

representation in the dorsal portions of V2 and V3 was more variable than in their

ventral counterparts, including substantial departures from the expected mirror-

symmetrical patterns. In addition, left hemisphere retinotopic maps were more variable

than those in the right hemisphere. Surprisingly, only one-third of individuals had maps

that conformed to the expected pattern in the left hemisphere. Visual field sign analysis

further revealed that in many individuals the area conventionally identified as dorsal

V3 shows a discontinuity in the mirror-image representation of the retina, associated

with a Y-shaped lower vertical representation. Our findings challenge the current view

that inter-individual variability in early extrastriate cortex is negligible, and that the

dorsal portions of V2 and V3 are roughly mirror images of their ventral counterparts.

Keywords

human connectome project, retinotopy, high-resolution fMRI, 7T, vision, hemispheric

differences, V3

Introduction

Non-invasive imaging has been instrumental in mapping the topographic organization of human visual cortex (Wandell and Winawer, 2010). The visual field maps in early visual areas (V1, V2, and V3) have been reported to be remarkably consistent across people, and predictable with reasonable accuracy using a template (Benson et al., 2014, 2012; Schira et al., 2010). While V1 contains a complete, first-order (continuous) representation of the contralateral visual hemifield, areas V2 and V3 form second-order (discontinuous) representations (Rosa, 2002). In these areas, a field discontinuity near the horizontal meridian splits the maps into upper and lower field representations that are only connected at the foveal confluence (Figure 1a,b). Accordingly, in parcellation schemes (Glasser et al., 2016; Wang et al., 2015), early visual areas form concentric bands, arranged in nearly symmetrical halves with respect to the calcarine sulcus. These bands, each containing the representation of a contralateral visual field quadrant, are referred to as the dorsal and ventral portions of V2 and V3 (Figure 1a). However, observations originating in several laboratories have indicated departures from this pattern, particularly in the dorsal region (Allen et al., 2021; Arcaro and Kastner, 2015; Benson and Winawer, 2018; Van Essen and Glasser, 2018). Even so, small-sized datasets, variability in acquisition sites and protocols, and methodological constraints have limited the investigation of this variability. As a result, no consensus exists about deviations from the canonical mirror-symmetrical organization of V2 and V3.

Figure 1 – Visual field mapping in the human early visual cortex. a, Coarse scale visual field mapping in the early visual cortex. The left (L) hemisphere maps the right visual field, and the right (R) hemisphere maps the left visual field. The dorsal portion of early visual areas maps the lower hemifield, and the ventral portion the upper field. **b,** Fine scale visual field mapping with visual field maps represented in polar angles (0-360º). In this model, the vertical (90º or 270º) and horizontal meridians (0º for the left and 180º for the right hemispheres) delineate boundaries between visual areas. **c,** Three "classical" polar angle maps, obtained from the left hemispheres of three individuals in the HCP retinotopy dataset, which conform to the traditional model. **d,** Three polar angle maps that deviate from this pattern, obtained from left hemispheres of three other individuals in the HCP retinotopy dataset. In the latter, the isopolar bands representing the anterior borders of dorsal V3 (V3d) and dorsal V2 (V2d) do not follow the proposed borders of V2 and V3 (dashed lines).

In humans, empirical visual field mapping using functional MRI (fMRI) is the primary means of delineating precise visual area boundaries in individuals. Visual field maps are typically defined in polar coordinates, resulting in two maps: one representing polar angle (or clock position) and the other eccentricity (or distance away from the fixation point) (Wandell and Winawer, 2010). In primates, isoangle bands representing the vertical and the horizontal meridians are thought to delineate boundaries between V1 and V2, V2 and V3, and V3 and higher-order visual areas (Figure 1b). Particularly, in human probabilistic maps, boundaries between the dorsal portions of early visual areas are roughly mirror images of their ventral counterparts (Figure 1c). Nevertheless, boundaries that deviate from the expected ones exist, but these have been mainly treated as artifactual, with researchers often overlooking the irregularities by simply drawing the boundaries to resemble that of a canonical map as best as possible (e.g. Figure 1d). Here, it may be important to remark that the border between the dorsal parts of V2 and V3 is well known to be variable in other mammals, and that it typically does not coincide with the representation of the horizontal meridian (see Rosa and Manger, 2005 for review).

Although previous reports of individual variability in the dorsal portion of human early visual cortex were primarily anecdotal (Allen et al., 2021; Arcaro and Kastner, 2015; Benson and Winawer, 2018; Van Essen and Glasser, 2018), a recently developed deep learning model predicts that individual variability in retinotopy exists, and that this is correlated with variations in gross anatomy (e.g., the pattern of sulci and gyri) (Ribeiro et al., 2021). Moreover, studies modelling the formation of retinotopic maps in non-human primates also indicate that different variants could develop based on the application of similar developmental rules (Yu et al., 2020).

Motivated by these findings, here we systematically investigate individual variability in

visual field maps of human early visual cortex using a recently released, large-scale

dataset: the 181 participants, 7T Human Connectome Project (HCP) retinotopy

dataset (Benson et al., 2018). Our aims were to quantify the level of individual

variability throughout early visual cortex (V1-V3) and to determine whether there are

common modes of retinotopic organization that differ from the established view (i.e.,

whether individual retinotopic maps differ from a template in similar ways). Our results

demonstrate that the dorsal portions of human early visual areas are more

heterogeneous than previously acknowledged and challenge the current view that

individual differences in retinotopic organization reflect experimental artifacts that may

be dismissed for practical purposes.

Materials and Methods

Dataset

We used the Human Connectome Project (HCP) 7T Retinotopy dataset (Benson et

al., 2018) to investigate individual variability in retinotopic maps of human early visual

cortex. This dataset consists of high-resolution functional retinotopic mapping and

structural data from 181 participants (109 females, age 22-35) with normal or

corrected-to-normal visual acuity. Participant recruitment and data collection were led

by Washington University and the University of Minnesota. The Institutional Review

Board (IRB) at Washington University approved all experimental procedures (IRB

number 201204036; "Mapping the Human Connectome: Structure, Function, and

Heritability"), and all participants provided written informed consent before data

collection (Van Essen et al., 2013). Additionally, the acquisition protocol has been

described in previous work (Benson et al., 2018; Van Essen et al., 2013).

Structural data were acquired at 0.7 mm isotropic resolution in a customized Siemens

3T Connectome scanner (Van Essen et al., 2013). Briefly, cortical surfaces were

reconstructed from T1w structural images using FreeSurfer and aligned to the 32k

fs_LR standard surface space. This standard 32k fs_LR cortical surface consists of

32,492 vertices sparsely connected, forming triangular faces. Functional data were

later aligned with this standard surface space.

Functional retinotopic mapping data were acquired using a Siemens 7T Magnetom

scanner at 1.6 mm isotropic resolution and 1 s TR. Data were preprocessed following

the HCP pipeline (Glasser et al., 2013), which included correction for head motion and

EPI spatial distortion, alignment of the fMRI data with the HCP standard surface space,

and denoising for spatially specific structured noise. Retinotopic mapping stimuli

comprised rotating wedges, expanding and contracting rings, and bars of different

orientations moving across different directions in the visual field. A population

receptive field (pRF) modeling procedure was then used to reconstruct visual field

maps (Benson et al., 2018; Dumoulin and Wandell, 2008; Kay et al., 2013), which

encompasses estimating the spatial preference of cortical surface vertices to different

locations of the visual field (i.e., its receptive field) defined in polar coordinates – for

more, see Benson et al, (2018). Hence, polar angle maps are retinotopic maps

reflecting the polar angle (angle relative to the horizontal vertical meridian) in the visual

field to which a vertex is most responsive, while eccentricity maps reflect the distance

from the center of the visual field (i.e., the fixation point). The combination of a polar

angle map and an eccentricity map completely specifies a map of the visual field, from

the center of the visual field to at least 8° of eccentricity (Benson et al., 2018).

Region of Interest

Early visual areas were defined by a surface-based probabilistic atlas (Wang et al.,

2015). This probabilistic atlas includes the dorsal and ventral portions of V1, V2 and

V3, not including the foveal confluence. For the clustering analysis, we slightly

modified the atlas by extending the dorsal border of V3 and including V1/V2/V3 foveal

confluence (Schira et al., 2009), in line with our previous work (Ribeiro et al., 2021).

Individual variability

We determined individual variability in visual field maps to quantify how variable these

maps were across visual areas (V1, V2, and V3), portions (dorsal and ventral), and

hemispheres (left and right) in human early visual cortex. First, we computed the

average retinotopic maps across all 181 individuals from the HCP retinotopy dataset

for both left and right hemispheres (Supplementary Figure 1). Then, we iteratively

calculated the vertex-wise difference between an individual's retinotopic map and the

average map. The difference between two angles is given by:

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$$\text{MIN} \left(\left| \hat{\theta} - \theta \right|, \left| \hat{\theta} - \theta + 2\pi \right|, \left| \hat{\theta} - \theta - 2\pi \right| \right) \tag{1}$$

for $0 < \theta < 2\pi$.

Finally, vertex-wise difference scores were averaged over vertices in the range of 1-

8° of eccentricity within the dorsal and ventral portions of early visual areas, resulting

in one scalar value per individual per visual area, which we refer to as the individual

variability. The eccentricity mask was defined using the group-average eccentricity

map. This range of eccentricity values was chosen because, in the original population

receptive field mapping experiment of the HCP, the visual stimulus extended to 8° of

eccentricity (Benson et al., 2018). Additionally, due to the inherent difficulty in mapping

the foveal confluence (Schira et al., 2009), we constrained our comparison to

eccentricity values above 1°. According to studies in non-human primates, this

corresponds approximately to half of the expected extent of V1, V2 and V3 (Gattass

et al., 1988, 1981).

Linear mixed-effects model

We determined whether there were main effects and interactions of hemispheres (left,

right), visual areas (V1, V2, V3), and portions (dorsal, ventral) on individual variability

of retinotopic maps using linear mixed-effect (LME) models. Standard ANOVAs and t-

tests assume statistical independence of individuals' data (Yu et al., 2022), which is

often not the case. For example, the 7T HCP retinotopy dataset includes data from 50

monozygotic and 34 dizygotic twins, totaling 168 individuals out of 181. Therefore, to

meet the statistical independence criterion, many data points would have to be

disregarded for standard statistical inference. However, LME models allow us to take

full advantage of the dataset by explicitly modeling cluster-specific means (random

intercepts). Indeed, individual variability from different visual areas is naturally

clustered by individuals (Magezi, 2015). Therefore, using this statistical model, we can

appropriately model individual-specific effects (Magezi, 2015; Yu et al., 2022).

In our linear mixed effect model, the dependent variable is the individual variability (Y),

which is modeled as a function of the fixed effects (β) of three factors (x) and their

interactions. These three factors are: hemisphere, visual area, and portion.

Additionally, we also consider the random effects (γ_i) associated with the individual (i

= 1, …, 181), and the random effects of each factor nested within the individual (γ_{ij},

with j = 1, 2, and 3). This model is expressed as:

$$Y_i = \beta_0 + \sum_{j=1}^{3} \beta_j x_j + \beta_{12} x_1 x_2 + \beta_{13} x_1 x_3 + \beta_{23} x_2 x_3 + \beta_{123} x_1 x_2 x_3 + \sum_{j=1}^{3} \gamma_{ij} + \gamma_i + \varepsilon_i \quad (2)$$

where β_0 is the intercept and ε is the residual random error. We built two separate

models for individual variability associated with polar angle and eccentricity maps

using Jamovi ("The jamovi project (2021)," n.d.).

Individual variability in retinotopic maps and potential confounds

Other potential sources of individual variability in retinotopic maps can be readily

included as covariates in LME models after their appropriate standardization by

computing z-scores. Effectively, adding a covariate to an LME model adds a βx term

to equation 2, where β is the estimated effect and x is the covariate. Here, we consider

the effect of potential confounders, including curvature, normalized mean BOLD

signal, and intra-individual variability in pRF estimates. These factors are known to be

correlated with retinotopy and could explain part of the variability found across visual

areas. Accordingly, we calculated pair-wise correlations among polar angle,

eccentricity, curvature, and normalized mean BOLD signal for both V1-3 and each

visual area separately. Given a region of interest, each topographic map was

vectorized, and data were concatenated across all participants (n=181) for each

modality. Polar angle maps were converted such that 0º corresponds to the horizontal

meridian and 90º corresponds to the upper and lower vertical meridians (Kurzawski et

al., 2022). Finally, pair-wise correlations were determined using these concatenated

sets of vectorized maps. Below we provide further motivation for considering each of

these covariates.

Curvature maps highlight the geometry of folding patterns of individuals' cortical

surfaces, with negative values representing sulci and positive gyri (as per the HCP

dataset). In V1, the horizontal and vertical meridian representations of the visual field

correlate with underlying sulcal patterns (Hinds et al., 2009; Holmes and Lister, 1916;

Horton and Hoyt, 1991; Inouye, 1909; Rajimehr and Tootell, 2009). This tight structure-

function relationship was further leveraged for individual-level predictions of retinotopy

from underlying anatomy, using atlas-fitting algorithms (Benson et al., 2014, 2012), a

Bayesian model (Benson and Winawer, 2018), and deep learning (Ribeiro et al.,

2021). Therefore, although curvature is a good predictor of retinotopy, it is unclear if it

is sufficient to explain the systematic differences in individual variability across visual

areas. Here, we consider the individual variability in curvature maps as a covariate.

Individual variability in curvature was operationalized as the mean absolute difference

between an individual's curvature map and the corresponding average map over

vertices in the range of 1-8º of eccentricity within the dorsal and ventral portions of

early visual areas, resulting in one scalar value per individual per visual area.

The mean pre-processed BOLD signal was used as a proxy for the location of large

veins (Boyd Taylor et al., 2019; Kurzawski et al., 2022), which are known to affect pRF

estimates. Voxels near large veins show lower mean BOLD signal, a phenomenon

known as the venous eclipse, which affects pRF estimates, for example, in area hV4

(Boyd Taylor et al., 2019; Winawer et al., 2010). As such, deviations in the expected

mean BOLD signal could lead to deviations from the expected retinotopic organization.

Therefore, we considered individual variability in the mean BOLD signal as another

covariate. To do so, we first normalized the mean BOLD signal (over the time course)

by dividing the value of each vertex by the maximum intensity (Boyd Taylor et al.,

2019). Then, we computed the individual variability as previously described for

curvature maps.

Finally, different levels of individual variability in retinotopic maps could also reflect

variations in their reliability across visual areas. For example, in the HCP dataset,

despite pRF estimates being highly consistent between two model fits (Benson et al.,

2018), there was still some intra-individual (between-fit) variability. Thus, we

considered intra-individual variability in pRF estimates as a covariate. Intra-individual

variability was operationalized as the difference in pRF estimates from two pRF model

fits of half splits of the retinotopic mapping data (fit 2 and 3; Supplementary Table 1),

which were provided and determined using the first and second half of the temporal

data from each run (Benson et al., 2018). By using these two other fits, it is possible

to assess the reliability of the pRF parameter estimates (Supplementary Figure 2);

hence, the difference in pRF estimates is a proxy for the reliability of individuals'

retinotopic maps and can be included as a covariate in the linear mixed effects model.

Clusters of spatial organization

We performed an exploratory clustering analysis to determine whether retinotopic

maps differ from the average map in similar ways, particularly in the dorsal portion of

early visual cortex. Specifically, we investigated the spatial overlap between

retinotopic maps as an unambiguous indicator of the similarity between two maps.

First, to obtain such a measure of the spatial overlap, the continuous polar angle maps

were converted into discrete maps, such that each vertex was categorized into one

out of four possible labels:

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$$\theta_{discrete} = \begin{cases} 0°, for\ 0° \leq \theta_{continuous} \leq 45° \\ 90°, for\ 45° < \theta_{continuous} \leq 180° \\ 270°, for\ 180° \leq \theta_{continuous} < 315° \\ 360°, for\ 315° \leq \theta_{continuous} < 360° \end{cases}$$

262 these categories were chosen because they highlight the location of visual area

263 boundaries. Discrete eccentricity maps were determined by:

$$\theta_{discrete} = \begin{cases} 0°, for\ 0° \leq \theta_{continuous} \leq 2° \\ 2°, for\ 2° < \theta_{continuous} \leq 4° \\ 4°, for\ 4° < \theta_{continuous} \leq 6° \\ 6°, for\ 6° < \theta_{continuous} \end{cases}$$

Next, the spatial overlap between discrete maps from all possible pairs of individuals

was estimated using the Jaccard similarity coefficient (Levandowsky and Winter, 1971;

Taha and Hanbury, 2015). The Jaccard index estimates similarity between two maps

by taking the size of the intersection (in number of vertices) divided by the size of the

union of two label sets. Hence, the Jaccard score ranges from 0 to 1; the closer to 1

the score is, the more similar the two maps are. For our data and each pair of

individuals, the Jaccard index is determined from the two possible individuals'

combinations (i.e., individual 1 *vs.* individual 2 and individual 2 *vs.* individual 1) since

the order of the maps determines which map is the reference. For each combination,

we estimated the Jaccard index for each label, and their weighted average was

determined using the number of labels' instances in the reference map to account for

label imbalance. Then, these two estimates were averaged, resulting in one estimate

of the spatial overlap between two individuals' discrete retinotopic maps.

To assess whether inter-individual differences fell into stereotyped patterns, we

applied a spectral clustering algorithm from Scikit-learn (Abraham et al., 2014;

Pedregosa et al., 2011). This algorithm operates on the low-dimensional embedding

of the affinity matrix (our Jaccard index-based similarity matrix), followed by K-means

clustering of the components of the eigenvectors in the low-dimensional space. This

low dimensional space is determined by selecting the most relevant eigenvectors of

the graph Laplacian of the affinity matrix, of which corresponding eigenvalues reflect

important properties of the affinity matrix that can be used to partition it (Luxburg,

2007). In implementing the spectral clustering algorithm, we set the number of clusters

to 6 and fixed the random state for replication purposes. We selected this number of

clusters as there are at least five different models of third-tier visual cortex organization

in non-human primates (Angelucci and Rosa, 2015), with a sixth cluster intended to

capture noisy or unclear retinotopic organization. Note, however, that this selection is

simply a speculation of the possibility space and do not reflect known inter-individual

differences in non-human primates. After clustering, we computed each cluster's mean

map by averaging the continuous retinotopic maps across individuals within each

cluster.

Visual field sign analysis

Lastly, we further examined unusual retinotopic maps to elucidate the kind of deviation

from the canonical maps they represent. We performed a visual field sign analysis

(Sereno et al., 1995, 1994), which combines both polar angle and eccentricity maps

into a unique representation of the visual field as either a non-mirror-image (like V2)

or a mirror-image representation of the retina (like V1). Since the left hemisphere

represents the right visual field, which in polar angle includes 0º-90º (upper right visual

field) and 270º- 360º (lower right visual field), we shifted the polar angle values so the

point of wrap-around (from 360º to 0º) was positioned at the horizontal meridian in the

contralateral hemifield, avoiding the discontinuous representation between 360º and

0º. Then, we interpolated the sparse and flattened polar angle and eccentricity maps

onto a regular x-y grid using SciPy (Virtanen et al., 2020). Next, we determined the

gradient of polar angle and eccentricity maps, mathematically expressed as:

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$$\Delta f_\theta = \frac{\partial f_\theta}{\partial x}\hat{\imath} + \frac{\partial f_\theta}{\partial y}\hat{\jmath} \text{ and } \Delta f_r = \frac{\partial f_r}{\partial x}\hat{\imath} + \frac{\partial f_r}{\partial y}\hat{\jmath}$$

where f_θ is the polar angle map and f_r the eccentricity, using the NumPy gradient

numerical method (Harris et al., 2020). Finally, the angle between the polar angle and

eccentricity maps' gradient vectors was determined at each x-y coordinate. If the angle

between the gradient vectors is between 0 and π, by convention, the cortical patch is

a mirror-image representation of the retina; otherwise, it is a non-mirror-image. After

binarizing the angle projection, we can conveniently infer borders between visual

areas because adjacent areas often have the opposite visual field sign (Sereno et al.,

1995; but see Yu et al. 2020 for caveat).

Data and code availability

The data used in this study is publicly available at BALSA

(https://balsa.wustl.edu/study/show/9Zkk). In addition, all accompanying Python and

MATLAB source codes are available on GitHub

(https://github.com/felenitaribeiro/VariabilityEarlyVisualCortex). On our GitHub

repository, executable code is available on interactive computational notebooks

(Jupyter notebooks) that allow anyone to execute interactive plotting functions with

dropdown menus to visualize an individual's polar angle and visual field sign maps,

given their cluster assignment. Note that our documentation provides instructions for

running them on Neurodesk (Renton et al., 2022). Finally, the intermediate files for

fitting the linear mixed effect models using Jamovi are available on the Open Science

Framework (https://osf.io/tdkuj/).

Results

Individual variability in retinotopy

We defined an individual variability metric to quantify how variable visual field maps

are across visual areas (V1, V2, and V3), portions (dorsal and ventral), and

hemispheres (left and right) in human early visual cortex. First, we computed the

average visual field maps across all 181 individuals from the HCP retinotopy dataset

for both left and right hemispheres. Then, we iteratively calculated the difference

between an individual's visual field map and the average map. Finally, these

differences were averaged over all vertices within the dorsal and ventral portions of

early visual areas, resulting in one scalar value per individual per visual area, which is

our individual variability metric. Therefore, this metric is a proxy for large-scale

deviations in visual field mapping. Figure 2 shows the distribution of individual

variability scores across all participants.

Figure 2 – Individual variability in visual field maps of early visual areas. a,
Hypothetical diagram of symmetrical distributions of individual variability across visual
areas. The centre and right columns illustrate empirical distributions of individual
variability of polar angle (**b** and **c**) and eccentricity (**d** and **e**) maps for both dorsal (dark
shades) and ventral (lighter shades) portions of early visual areas in left (purple) and
right (green) hemispheres.

We built a linear mixed effect model (Yu et al., 2022) to test the fixed effects of

hemispheres, visual areas, and portions on large-scale individual variability of polar

angle (Table 1) and eccentricity (Table 2) maps. Table 1 shows statistically significant

main effects of all factors on individual variability of polar angle maps. Specifically,

polar angle maps of the left hemisphere show higher individual variability than those

found in the right hemisphere (mean difference = 3.35, p<.001). The dorsal portions

of early visual areas are also more variable than the ventral portions (mean difference

= 3.30, p<.001). Finally, post-hoc comparisons of visual areas indicated that V3 has

higher individual variability than V2 (mean difference = 1.60, p<.001) and V1 (mean

difference = 3.99, p<.001); V2 also has higher individual variability than V1 (mean

difference = 2.38, p<.001). For brevity, we only show the main effects in Table 1,

although we also found statistically significant interactions. Briefly, each visual area in

the left hemisphere has significantly higher individual variability than its analogous

area in the right hemisphere. In addition, the dorsal portion of each visual area of the

left hemisphere is significantly more variable than its dorsal analogue in the right

hemisphere and the ventral analogue of both the left and right hemispheres (for more,

see the Supplementary Material). These findings suggest that individual variability in

polar angle representations varies across hemispheres, visual areas, and according

to dorsal/ventral locations.

Table 1 - Fixed effects parameter estimates for the linear mixed effect model of
individual variability of polar angle maps. SE – standard error; CI – confidence
interval.

		Polar angle						
				95% CI				
Names	**Effect**	**Estimate**	**SE**	**Lower**	**Upper**	**df**	**t**	**p**
Intercept	Intercept	18.58	0.30	17.99	19.17	180	61.86	<.001
Hemisphere	RH-LH	-3.35	0.32	-3.97	-2.72	181	-10.47	<.001
Visual area (1)	V2-V1	2.38	0.29	1.81	2.95	210	8.21	<.001
Visual area (2)	V3-V1	3.99	0.32	3.36	4.61	187	12.54	<.001
Portion	ventral - dorsal	-3.30	0.29	-3.86	-2.74	181	-11.50	<.001

Moreover, Table 2 shows statistically significant main effects of the hemisphere, visual

area, and the visual area portion on individual variability of eccentricity maps. Like

polar angle maps, eccentricity maps of the left hemisphere show higher individual

variability than those in the right hemisphere (mean difference = 0.14, p<.001). The

dorsal portion of early visual areas is also more variable than the ventral portion (mean

difference = 0.13, p<.001). For visual areas, post-hoc comparisons indicated that the

only statistically significant difference was that of V3 versus V1, with V3 having higher

individual variability than V1 (mean difference = 0.05, p<.004). In addition, statistically

significant interactions were also found (Supplementary Material). Each visual area in

the left hemisphere has significantly higher individual variability than analogous areas

in the right hemisphere, except for V3. Eccentricity maps of each visual area's dorsal

portion in the left hemisphere are significantly more variable than the dorsal

384   counterpart in the right hemisphere, and the ventral analogues in both the left and the

385   right hemispheres.

386   **Table 2 – Fixed effects parameters estimates for the linear mixed model of**
387   **individual variability of eccentricity maps.** SE – standard error; CI – confidence
388   interval.

		Eccentricity						
				95% CI				
Names	Effect	Estimate	SE	Lower	Upper	df	t	p
Intercept	Intercept	0.81	0.02	0.77	0.85	180	41.86	<.001
Hemisphere	RH-LH	-0.14	0.01	-0.16	-0.11	181	-10.91	<.001
Visual area (1)	V2-V1	0.01	0.01	-0.01	0.04	402	0.98	0.326
Visual area (2)	V3-V1	0.05	0.01	0.02	0.08	182	3.24	0.001
Portion	ventral - dorsal	-0.13	0.03	-0.18	-0.07	180	-4.64	<.001

Influence of covariates on individual variability of retinotopic maps

Other potential sources of individual variability in retinotopic maps include covariates,

such as cortical curvature and mean BOLD signal. As mentioned in the Materials and

Methods section, these factors are known to be correlated with retinotopy and could

explain part of the variability found across visual areas. In Figure 3, we show pair-wise

correlations among polar angle, eccentricity, curvature, and normalized mean BOLD

signal for both V1-3 (main plots) and each visual area separately (inset plots). Across

V1-3 (Figure 3, main plots), polar angle was significantly correlated with curvature and

the mean BOLD signal in both the right and left hemispheres; eccentricity was

correlated with the mean BOLD signal in both hemispheres and curvature in the right

hemisphere. Moreover, the correlation between these maps varied across visual

areas. Specifically, correlations between polar angle and curvature, polar angle and

mean BOLD signal, and curvature and mean BOLD signal decreased from V1 to V2

to V3.

Figure 3 – Retinotopic maps correlation with covariates. Pair-wise correlations
among polar angle, eccentricity, curvature, and normalized mean BOLD signal for
early visual areas (V1-3; main plot) and each visual area separately (inset plots). Polar
angle maps were converted such that 0° corresponds to the horizontal meridian and
90° corresponds to the upper and lower vertical meridians (Kurzawski et al., 2022).
Finally, data were concatenated across all participants (n=181). * p < .001.

Given the meaningful correlation between some of these variables and their varying

degree of association across visual areas, we further include individual variability in

curvature and mean BOLD signal maps and the intra-individual variability in pRF

estimates (a proxy for the reliability of the retinotopic maps) as covariates in the LME

model (Tables 3 and 4). Our findings indicate that the main effects found here were

not a mere reflection of variation in the reliability of the individual maps and other

covariates, at least at large-scale. For example, we found that the main effects of all

factors (hemispheres, visual areas, and portions) on individual variability of polar angle

maps persisted, but the estimated effects were slightly reduced (Table 3). To

determine whether other properties would predict individual variability in polar angle

maps, we included age, gender (as cofactor), and gray matter volume as additional

covariates, but did not find significant effects of any of these variables (not shown here,

but available as supplementary material). For eccentricity, we found the main effects

of hemisphere and portion but not visual area after accounting for these covariates

(Table 4). Crucially, these findings suggest that individual variability in polar angle

representations varies as a function of hemispheres, visual areas, and dorsal/ventral

locations. These effects are not only due to trivial intra-individual variability in pRF

estimates or individual variability in curvature and the mean BOLD signal.

Table 3 - Fixed effects parameter estimates for the linear mixed effects model of

individual variability of polar angle maps using covariates. The covariates

included in the model were intra-individual variability in pRF estimates, and individual

variability in curvature and the mean BOLD signal. SE – standard error; CI –

confidence interval.

				Polar angle				
				95% CI				
Names	**Effect**	**Estimate**	**SE**	**Lower**	**Upper**	**df**	**t**	**p**
Intercept	Intercept	18.58	0.27	18.06	19.11	179	69.58	<.001
Hemisphere	RH-LH	-2.44	0.28	-2.99	-1.89	189	-8.71	<.001
Visual area (1)	V2-V1	1.93	0.33	1.29	2.57	323	5.90	<.001
Visual area (2)	V3-V1	3.54	0.32	2.92	4.16	288	11.22	<.001
Portion	ventral - dorsal	-2.12	0.29	-2.69	-1.54	207	-7.20	<.001
Intra-individual variability	Intra-individual variability	2.90	0.16	2.58	3.22	1364	17.81	<.001

Names	Effect	Estimate	SE	Lower	Upper	df	t	p
Individual variability in curvature	Individual variability in curvature	0.40	0.14	0.12	0.68	1812	2.82	0.005
Individual variability in mean BOLD signal	Individual variability in mean BOLD signal	0.21	0.19	-0.16	0.57	597	1.11	0.268

Table 4 - Fixed effects parameter estimates for the linear mixed effects model of individual variability of eccentricity maps using covariates. The covariates included in the model were intra-individual variability in pRF estimates, and individual variability in curvature and the mean BOLD signal. SE – standard error; CI – confidence interval.

	Polar angle							
				95% CI				
Names	**Effect**	**Estimate**	**SE**	**Lower**	**Upper**	**df**	**t**	**p**
Intercept	Intercept	0.81	0.01	0.78	0.84	179	54.64	<.001
Hemisphere	RH-LH	-0.07	0.01	-0.09	-0.05	201	-6.40	<.001
Visual area (1)	V2-V1	-0.02	0.01	-0.05	0.01	506	-1.52	0.128
Visual area (2)	V3-V1	-0.02	0.02	-0.05	0.01	262	-1.20	0.231
Portion	ventral - dorsal	-0.07	0.02	-0.11	-0.03	186	-3.24	.001
Intra-individual variability	Intra-individual variability	0.21	0.01	0.20	0.23	1982	26.51	<.001
Individual variability in curvature	Individual variability in curvature	0.01	0.01	-0.00	0.02	1829	1.52	0.128
Individual variability in mean BOLD signal	Individual variability in mean BOLD signal	0.02	0.01	0.00	0.03	489	2.34	0.020

Common modes of variability

Next, we performed an exploratory analysis to determine whether polar angle maps

differ from the average map in similar ways, particularly in the dorsal portion of early

visual cortex of the left hemisphere. Note that an analogous analysis was performed

for eccentricity maps, but in agreement with our statistical models we did not find

meaningful differences across eccentricity map clusters (Supplementary Figure 3). We

computed the extent of overlap between discrete polar angle maps from all possible

pairs of individuals using the Jaccard index, resulting in a similarity matrix (Figure 4a).

Next, we applied a spectral clustering algorithm with a fixed number of clusters equal

to 6 (Figure 4b). Finally, we averaged the continuous polar angle maps across

individuals within each cluster to visualize common patterns of retinotopic organization

in the dorsal portion of early visual cortex (Figure 4c; see Supplementary Figure 4 for

the complete set of average maps based on the Figure 4c clustering assignment).

Figure 4 - Clusters of retinotopic organization in the dorsal portion of early visual cortex. a, Continuous polar angle maps were converted into discrete maps, such that each vertex would be categorized into one out of four possible labels. Spatial overlap between discrete maps was estimated using the Jaccard similarity coefficient from all possible pairs of individuals, resulting in a 181×181 similarity matrix. **b,** Then, we applied a spectral clustering algorithm – setting the number of clusters to 6. **c,** An average map (discrete and continuous) was calculated for each cluster by averaging the continuous polar angle maps across all individuals within each cluster.

Our findings clearly indicate shared patterns of retinotopic organization that deviate from the canonical polar angle representation in the dorsal portion of early visual

cortex (Figure 1c). Specifically, average maps from clusters 1 and 5 capture nearly a

third of individuals and show canonical polar angle representations, with clear

boundaries between V1/V2 and V2/V3 (Figure 1c and Figure 4c). However, clusters

2, 3, and 4 capture nearly two thirds of individuals and deviate from this canonical

polar angle representation (Figure 4c). The average map from cluster 2 shows that the

boundaries between V1 and V2, and the most anterior portion of V3 and higher-order

visual areas, merge to form a Y-shaped (or forked) lower vertical representation.

Clusters 3 and 4 show a truncated V3 boundary, indicating that dorsal V3 does not

cover the entire quarter visual field (i.e., from 360º to 270º) either throughout its length

or only in its most anterior portion. Finally, cluster 6 reflects unclear retinotopic

organization, with a handful of individuals' retinotopic maps showing overall low

correspondence with the canonical retinotopic organization.

Qualitatively, individual maps seem to agree with their corresponding average cluster

map, but there are some exceptions (Figure 5, Supplementary Figure 5-10). Figure 5

shows the average cluster maps from each cluster and examples of individuals' maps

that are qualitatively similar and dissimilar to their corresponding average cluster map.

While most polar angle maps correspond well with their average cluster maps (as seen

in the middle row of Figure 5), there is also an apparent mismatch between a few maps

and their corresponding cluster average (bottom row in Figure 5). For example,

individual #132118 was assigned to Cluster 4, but their polar angle map is qualitatively

more similar to Cluster 5. These mismatches are likely due to the extensive overlap

between within-cluster and between-clusters distributions of pairwise Jaccard scores

(Figure 6). Note in Figure 6 that the within-cluster distributions highlighted in grey are

generally shifted to the right compared to the between-clusters distributions, indicating

their higher Jaccard scores. However, the overlap between these distributions is

substantial. For example, the between cluster 1 and 5 distribution overlaps with within-

cluster 1 distribution throughout its entirety, which is justified by the significant

similarity between their average maps. Despite this, we found that the average within-

cluster Jaccard score is 0.54 (SD = 0.07), while the average between-clusters score

is 0.46 (SD = .08), showing that pairs of maps within a cluster are, on average, more

similar than between-clusters.

Figure 5 - Qualitative evaluation of clusters. Average cluster maps are shown in

the top row. The middle row shows examples of maps from each cluster with a similar

retinotopic organization to the corresponding average map. Finally, in the bottom row,

examples of those with dissimilar organizations are shown.

Figure 6 - Distributions of pairwise Jaccard scores. Within and between-clusters distribution of Jaccard scores across all pairs of individuals. Within-cluster distributions are highlighted in grey. Between-clusters distributions are the same regardless of the order of the clusters, i.e., the Jaccard score distribution between cluster 1 and cluster 2 ('between 1 and 2') is the same as the one between cluster 2 and 1. Black vertical lines indicate distributions' means.

Visual field sign analysis for delineating visual area boundaries

Finally, we further examined retinotopic maps with a Y-shaped lower vertical representation, i.e., those primarily assigned to cluster 2, to elucidate the kind of deviation from the canonical maps they represent (Figure 7 and Supplementary Figure 11). To do so, we performed a visual field sign analysis (Sereno et al., 1995, 1994),

which combines both polar angle and eccentricity maps into a unique representation

of the visual field as either a non-mirror-image or a mirror-image representation of the

retina (Figure 7a; see Materials and Methods). With such representation, we can

directly infer visual area parcellation.

Figure 7b shows polar angle gradients in a 'streamline' representation and their

respective visual field sign representation for two participants with canonical and four

with Y-shaped lower vertical representations. While visual area boundaries in early

visual cortex are conventionally identified by reversals in the progression of the polar

angle values – or changes in the direction of polar angle gradients, it is unclear how to

delineate boundaries in the dorsal portion of polar angle maps in those participants

with non-canonical maps (note that their respective ventral portion followed the

classical representation), but not on those with canonical maps. However, with the

visual field sign representation, the boundaries delineating dorsal V2 in those

participants with non-canonical maps are more explicit, and it reveals that the area

identified as dorsal V3 shows a discontinuity in the expected mirror-image

representation. Such representation has been proposed as the 'incomplete-V3' model

of the third-tier cortex for the macaque (Angelucci and Rosa, 2015) and other similar

models for the owl monkey (Sereno et al., 2015) and the marmoset monkey (Rosa

and Tweedale, 2000). Supplementary Figure 11 shows five more examples of polar

angle maps with unusual Y-shaped lower vertical representations and five other

examples with a truncated V3 boundary. While individuals with unusual Y-shaped

lower vertical representations have a discontinuity in the canonical mirror-image

representation of the retina in dorsal V3, individuals with a truncated V3 do not show

such a discontinuity. Altogether, our findings may suggest that, at least in humans, the

canonical model does not oppose other models established for non-human primates;

these models coexist and reflect common modes of variability in the retinotopic

mapping of early visual areas.

Figure 7 - Visual field sign analysis for delineating visual areas. a, The visual field
sign analysis (Sereno et al., 1995, 1994) combines polar angle and eccentricity maps
(not shown) into a unique representation of the visual field as either a non-mirror-
image (like V2) or a mirror-image (like V1) representation of the retina. This analysis
consists of determining the angle between the polar angle and eccentricity maps'
gradient vectors, respectively, the green and purple vectors, at each cortical
coordinate. If the angle between the gradient vectors is between 0 and π, by
convention, the cortical patch is a mirror-image representation of the retina; otherwise,
it is a non-mirror-image. **b,** Six examples of left hemisphere polar angle maps with
canonical (on the left) and non-canonical (on the right) representations in the dorsal
portion of early visual cortex are shown (top row). Polar angle gradients are shown in
a 'streamline' representation to highlight reversals in the progression of the polar angle
values. Their respective visual field sign representation (bottom row) is also shown.
While it was unclear how to delineate boundaries in the dorsal portion of polar angle

maps in those participants with non-canonical maps, the visual field sign

representation reveals that the area identified as dorsal V3 shows a discontinuity in

the canonical mirror-image representation (solid white circles).

Discussion

In this study, we systematically investigated individual variability in visual field

representation of human early visual cortex using the HCP 7T retinotopy dataset. We

found that retinotopic maps in the left hemisphere were more variable than those in

the right hemisphere. Moreover, in the left hemisphere the dorsal portions of early

visual areas were more variable than their ventral counterparts, and these effects were

not merely due to trivial large-scale intra-individual variability in pRF estimates or

individual variability in curvature and the mean BOLD signal. Thus, we investigated

whether there were common motifs in the observed individual variability in retinotopic

maps of left hemispheres. This analysis showed that deviations from the canonical

model of continuous, alternating bands of vertical and horizontal meridian

representation in V2 and V3 exist in the majority of individuals. Specifically, the visual

field sign analysis revealed that the area identified as dorsal V3 shows a discontinuity

in the expected mirror-image representation of the retina in individuals with a Y-shaped

lower vertical representation in the dorsal portion of early visual cortex. Overall, our

findings challenge the current view that the dorsal portions of early visual areas form

retinotopic maps which are consistent between individuals and are roughly mirror

images of their ventral counterparts.

Individual variability in retinotopy

Although previous evidence for the variability seen across dorsal early visual cortex in

humans has been mostly anecdotal, a number of studies have indicated a complex,

retinotopic organization of dorsal early visual areas in non-human primates, using both

electrophysiological recordings and high-resolution fMRI (Angelucci and Rosa, 2015;

Gattass et al., 1988; Sereno et al., 2015; Zhu and Vanduffel, 2019). Accordingly, there

is a long-standing debate about the number of visual areas – and their boundaries –

in the third-tier visual cortex of New and Old-World monkeys (Angelucci and Rosa,

2015; Hadjidimitrakis et al., 2019). However, the question of whether the areal

boundaries in this region show significant individual variability has not been studied

systematically in non-human primates. Only Gattass et al. (1988) reported, in the

macaque monkey, that the representation of the lower vertical meridian in dorsal V3

varied across individuals, but firm conclusions could not be drawn due to the small

sample. These authors indicated that some animals showed a continuous

representation of this meridian along the rostral border of this area, whereas in others

additional field discontinuities created a discontinuous representation. Notably, the

same discontinuities in the anterior border of dorsal V3 were also found in our

systematic investigation of individual variability in human polar angle maps. It is also

significant that the same pattern of variation (relatively simple and reproducible

representations of the upper contralateral quadrant, and complex and variable

representations of the lower quadrant) characterize V2 and V3 in at least one non-

primate, the cat (Rosa and Manger, 2005; Tusa et al., 1979). Overall, our findings in

humans demonstrate that the organization of dorsal early visual areas is more

heterogeneous than previously acknowledged and suggest that this may be a common

feature of mammals with developed vision.

Our results also indicate that the variability in retinotopic organization increases

between V1 and V2, and between V2 and V3. These findings are compatible with a

model whereby retinotopic maps develop sequentially from "core" areas like V1, where

maps are determined by genetically-encoded molecular gradients, towards late-

maturing areas, where their formation occurs interactively through a wire-length

minimization rule, allowing progressively greater degrees of freedom (Yu et al., 2020).

The fact that the organization of ventral maps tends to be more reproducible than that

of dorsal maps may be related to the presence of the middle temporal area (MT, or

V5) as a second "anchor" node for the formation of dorsal maps in the proximity of the

dorsal cortex, resulting in merging maturational gradients in the region of dorsal V3

and V4 (Rosa and Tweedale, 2005).

Although different models of third-tier visual cortex organization in non-human

primates (Angelucci and Rosa, 2015) also suggest unusual eccentricity mapping, we

did not find meaningful differences in clusters of eccentricity maps (Supplementary

Figure 3) nor in the average eccentricity maps based on clusters from Figure 4c

(Supplementary Figure 4). This may be associated with the limited extent of the visual

stimulus (up to 8° of eccentricity) (Benson et al., 2018) and remains to be further

investigated. Indeed, much of the controversy regarding the organization of dorsal

areas in non-human primates refers to regions anterior to the peripheral

representations in V2 (Angelucci and Rosa, 2015). Another alternative is having a

complex pattern of polar angle representation coexisting with a preserved eccentricity

gradient, as demonstrated by previous work in areas V2 and V3 of cats (Tusa et al.,

1979), flying foxes (Rosa, 1999), ferrets (Manger et al., 2002) and tree shrews

(Sedigh-Sarvestani et al., 2021).

Our investigation provides firm evidence for individual variability in the retinotopic

organization across parts of early visual areas in the human visual cortex. Moreover,

the exploratory analysis indicates the presence of shared patterns of retinotopic

organization that deviate from the canonical polar angle representation in the dorsal

portion of early visual cortex. Future work could extend these insights through

additional analyses – for example, by employing different similarity metrics, using

different features, or changing the number of clusters. Here, we limited our analysis to

the spatial overlap of discrete polar angle maps, which means that a pair of

qualitatively similar but spatially misaligned polar angle maps, for example, might have

a low Jaccard score. If another more suitable metric can consider the topographic

organization of polar angle maps regardless of the spatial location, it would be possible

to increase the consistency between an individual's map and their cluster average

map. It would also be possible to estimate the similarity between two individuals'

retinotopic maps from specific features extracted from the maps, such as linear

magnification along isoeccentricity lines, to provide insights into changes in these

properties as a function of cortical location (Schira et al., 2010). Another option would

be to use the visual field sign representation directly for clustering individuals based

on the non-mirror-image and mirror-image representation of the retina across early

visual cortex. Finally, it is important to note that selecting the ideal number of clusters

depends on the similarity metric employed, prior knowledge, and the clustering

algorithm. Therefore, future work could be performed to explore the effect of the

number of clusters on clustering quality (perhaps as indicated by within- vs. between-

cluster similarity measures).

Potential sources of individual variability in retinotopy

Given the presence of individual variability across early visual cortex in humans,

another potential line of investigation involves the origin of this variability. Pertinently,

we recently developed a deep learning model of retinotopy able to predict this

individual variability from anatomical information (including curvature) (Ribeiro et al.,

2021; Ribeiro et al., 2022), suggesting that it is, to some extent, a structure-related

variation. Accordingly, here we modelled individual variability in curvature as an

anatomy-related covariate, which significantly affected the individual variability of polar

angle maps. Additionally, to determine if our findings were not merely due to other

covariates, we included the normalized mean BOLD signal and intra-individual

variability in pRF estimates as additional covariates. The former is an important proxy

for the location of large veins (Boyd Taylor et al., 2019; Kurzawski et al., 2022), which

are known to affect pRF estimates (Boyd Taylor et al., 2019; Winawer et al., 2010).

The latter is a proxy for the reliability of individuals' retinotopic maps, which could vary

across visual areas. While we found a significant effect of intra-individual variability,

the main effects of all factors (hemispheres, visual areas, and portions) on individual

variability of polar angle maps persisted. This finding indicates that these large-scale

effects were not merely due to the covariates included in the model. Note, however,

that as we only consider the effect of large-scale deviations (averaged over a region

of interest) of each of these covariates on large-scale variability of the retinotopic

maps, it could be the case they still have a crucial role in local variability, which we

further discuss in the Limitations section below.

Retinotopic maps could also vary as a function of data resolution and cortical depth.

For example, signal-to-noise ratio (SNR) and partial volume artifacts are directly

affected by data resolution (or voxel size); that is, both reduce with voxel size

(Hoffmann et al., 2009). While lower SNR might lead to noisier (or less smooth)

retinotopic maps (Hoffmann et al., 2009), the gain from the reduced susceptibility to

partial volume artifacts will likely result in increased validity of the observed maps.

Partial volume artifacts may arise from patches of opposite walls of a sulcus running

across a single voxel or even small vessels, leading to inaccurate signals from the

combination of different brain regions and tissues. With increasing magnetic field

strength, it should be easier to strike the right balance between high SNR and low

partial volume artifacts, which is crucial for determining the impact of registration errors

due to partial volume artifacts on the variability of retinotopic maps. Moreover, previous

studies have also shown that the hemodynamic response function (Puckett et al.,

2016) and a spatial pattern of activation (Polimeni et al., 2010) varied across depths

in V1. Specifically, Polimeni and colleagues found that a spatial pattern of activation

(an 'M') becomes clearer from the white matter surface to the mid-thickness surfaces

and then deteriorates once again near the pial surface. Altogether, these studies

motivate a more thorough investigation of how retinotopy, as measured by fMRI, varies

as a function of data resolution and cortical depth and its implication on individual

variability in retinotopy. However, it is also important to note that studies of the

columnar organization of non-human primates using single-cell recordings have not

found any evidence that the receptive field location varies with cortical depth, although

the receptive field size changes, being smallest in the middle layers (Hubel and

Wiesel, 1974; Rosa et al., 1997).

Another potential research direction is determining the extent to which eye movement

underlies some of the variability found in retinotopic maps of early visual areas. For

example, one could systematically evaluate pRF modeling accuracy as a function of

gaze position change (and other eye-tracking signal derivatives). We performed a

preliminary analysis of the deviation in gaze position at each time point and averaged

across runs of retinotopic mapping stimuli and individuals assigned to each cluster.

Supplementary Table 2 summarizes the average deviation of gaze position from the

fixation point along the X and Y axes for each cluster. In brief, we did not find consistent

results across clusters. However, we do not rule out the effect of eye movement on

the variability of the retinotopic maps because the eye-tracking data quality is variable

and unavailable for some individuals in the HCP retinotopy dataset (Benson et al.,

2018), and the clustering quality could be improved (e.g., through manual clustering

or the other approaches previously discussed). Another possibility is determining the

reliability of these retinotopic maps through connective field modeling (Haak et al.,

2013) with unconstrained eye movement data (Tangtartharakul et al., 2023). Yet, we

observe that unusual maps in the dorsal portion of the early visual cortex coincide with

canonical representations in the ventral portion (Figure 7 and Supplementary Figures

4 and 11), which is unlikely to be the case for noisy data driven by massive eye

movements.

Limitations

Although we demonstrate that common modes of deviation from the canonical dorsal

V2 and V3 organization exist in the left hemisphere, further analyses are necessary to

fully ascertain if such variability is indeed neurogenic or if it could be a result of

measurement errors. Amongst potential sources of measurement error is the presence

of large veins running adjacent to regions of interest. Accordingly, by using the

normalized mean BOLD signal as a proxy for the location of large veins (Boyd Taylor

et al., 2019; Kurzawski et al., 2022), studies have shown that voxels near these veins

show lower mean BOLD signal, a phenomenon known as the venous eclipse, which

affects pRF estimates, for example, in area hV4 (Boyd Taylor et al., 2019; Winawer et

al., 2010). As such, deviations in the expected mean BOLD signal could result in

deviations from the expected retinotopic organization. Thus, to better understand the

potential effects of the presence of large veins on the different levels of variability in

visual field representation across early visual areas, we considered both the pair-wise

correlations between retinotopic maps and the normalized mean BOLD signal (Figure

3) and the large-scale deviation in the normalized mean BOLD signal as covariates in

the LME model (Tables 3 and 4).

In the pair-wise correlation analysis, we found that the polar angle representation is

indeed correlated with the normalized mean BOLD signal, and the magnitude of such

association varies across visual areas. Specifically, we found a higher correlation (in

magnitude) between the polar angle and normalized BOLD signal in ventral visual

areas than in dorsal areas. This difference could be explained by ventral areas'

proximity to the dural venous sinuses (Winawer et al., 2010), i.e., the transverse sinus,

the superior sagittal sinus, and the straight sinus. These venous sinuses are known to

introduce artifacts to the BOLD signal, which might lead to changes in retinotopic

maps. Importantly, none of these sinuses run near the dorsal V2 and V3 (see

Supplementary Figure 4, in which it is possible to observe the likely confluence of

sinuses), and both of which show the lowest correlation between polar angle and

normalized BOLD signal maps.

Moreover, we modeled large-scale deviation in visual field maps using the large-scale

deviation of the normalized mean BOLD signal as a covariate. We did find a significant

effect of the normalized mean BOLD signal on the individual variability of eccentricity

(Table 4) but not of polar angle maps (Table 3). Thus, to determine the effectiveness

of the LME model for uncovering the effects of covariates, we similarly modeled the

large-scale variability in polar angle maps of hV4 (not shown in the manuscript, but

available as supplementary material). We did not find an effect of the normalized mean

BOLD signal on large-scale deviation in polar angle maps in hV4. However, we found

a weak correlation between the polar angle and normalized mean BOLD signal (LH: r

= -0.06; RH: r = -0.07). This finding does not reconcile with previous reports, i.e., that

venous artifact impacts retinotopy. However, a fine-grained analysis of polar angle

maps in hV4 indicated the inconsistent effect of the venous artifact on polar angle

mapping (Boyd Taylor et al., 2019). In our analysis, though, these results might reflect

an inability to appropriately parcellate hV4 at the individual level when using an atlas-

based parcellation; while early visual areas are more consistently found at specific

spatial locations, this is not the case for other visual areas of which spatial location

and extent seem to vary across participants. Therefore, although we did not find a

significant effect of the normalized mean BOLD signal in our LME model, it does not

mean that the macro- and microvasculature do not affect retinotopy. For the former,

future research might consider a fine-grained analysis of topographic deviations, such

as the one reported by Boyd Taylor et al. (2019). For the latter, given that the mean

BOLD signal is only used as a proxy for the location of large veins, a more detailed

analysis of the microvasculature might require other imaging data, such as high-

resolution time-of-flight magnetic resonance angiography data (Bollmann et al.,

2022).

Future Directions

Our findings raise questions about if and how cortical atlases should be revisited to

accommodate deviations from the canonical model of retinotopic organization,

especially for dorsal V3. Here, by using the visual field sign representation, we could

better understand what kind of deviation from the canonical model atypical maps

represent. Therefore, using such data representation could be helpful for the manual

segmentation of atypical maps. Alternatively, combining deep learning models

(Ribeiro et al., 2021) for generating a retinotopic prior that accommodates more

variability with a Bayesian framework (Benson and Winawer, 2018) for boundary

delineation might prove fruitful to support the need for automated individual-level

parcellation methods. It would also be desirable to use functional characteristics of

areas, such as the responses to specific types of visual stimulation, to increase the

confidence in assignment of boundaries. Whereas this may be possible in some

situations (e.g. using motion selectivity as a localizer for area MT; Pitzalis et al., 2010),

attempts to segregate V3 from adjacent areas on this basis may be more challenging,

due to the physiological similarity between this area and the adjacent V2 and V3a

(Gegenfurtner et al., 1997; Levitt et al., 1994; Zeki, 1978). Differences in pRF size (Zhu

and Vanduffel, 2019) could offer some insight, although the wide overlap in the

distributions of single-unit receptive field sizes in adjacent areas (Rosa, 1997)

suggests that obtaining clear-cut boundaries on this basis remains unlikely.

Importantly, accommodating individual variability for the automatic parcellation of

visual areas is also crucial for understanding the functional properties of the human

visual cortex. These analyses require the precise delineation of boundaries between

visual areas, either by manually tracing transitions in visual field maps or by using an

automatic segmentation method (Benson et al., 2014; Benson and Winawer, 2018;

Dougherty et al., 2003). In both cases, a spatially consistent mapping (i.e., a canonical

representation) of continuous, alternating bands of vertical and horizontal meridian

representation in V2 and V3 is often assumed. However, we demonstrate that

deviations from the canonical model exist, especially in the left hemisphere and dorsal

V3. This finding could have several implications for post hoc analyses requiring visual

area delineation, suggesting that previous studies may have mischaracterized

differences between these visual areas due to misidentification. Therefore, an

important future direction of this work is determining how functional selectivity varies

across visual areas parcellated according to the canonical model compared to using

new parcels estimated with the visual field sign analysis.

In addition, the present findings highlight the need for a more comprehensive

assessment of the degree of variability in visuotopic maps in non-human primates,

where a higher degree of precision can be achieved with invasive methods including

single neuron recordings and optical imaging of intrinsic signals. To date, variability

has only been reported in macaque monkeys (Gattass et al. 1988), but the available

data in marmoset and owl monkeys indicate a reproducible organization that does not

fully agree with the canonical model of dorsal V3 (Angelucci and Rosa, 2015; Rosa

and Schmid, 1995; Rosa and Tweedale, 2000; Sereno et al., 2015). Whether this

simply reflects the small number of individuals explored, or a truly more stable

configuration (perhaps associated with the larger brain size in humans; Angelucci and

Rosa, 2015; Rosa and Tweedale, 2005) remains to be determined.

Another finding that requires consideration is the interhemispheric difference revealed

in our data: retinotopic maps in the left hemisphere showed more variation than those

in the right hemisphere. To date, there has been no report of interhemispheric

differences in early visual cortex of other mammals, including non-human primates. In

part, this may be traced to the relatively small samples in these studies compared to

those possible using human fMRI. However, another possibility is that such differences

may arise more frequently in human brains due to the scaling of callosal connections

with brain size (Rilling and Insel, 1999), which may promote a higher degree of

connectional independence during development.

Finally, these results raise many questions regarding how retinotopic maps develop.

For example, studies modelling the formation of retinotopic maps in development have

suggested that multistable solutions may occur depending on factors such as the

degree of elongation of the area (Sedigh-Sarvestani et al., 2021; Wolf et al., 1994) and

adjacency with other areas (Yu et al., 2020), which do not violate the need to minimize

the length of connections (Durbin and Mitchison, 1990; Swindale, 1996). Therefore,

future work could evaluate whether there is an overlap between function- and

anatomy-based clusters to help elucidate the developmental mechanisms underlying

the variability of human dorsal extrastriate cortex.

In conclusion, using a large-scale brain imaging dataset, we provide new insights into

the variability in the topographical organization of human visual cortex. These insights

may prove crucial in guiding further experimental investigations and theories about

retinotopic organization differentiation across species, development, and individuals.

Acknowledgments

This work was supported by the Australian Research Council (DE180100433 and

DP210101042) and National Health and Medical Research Council (APP1194206).

Data were provided by the Human Connectome Project, WU-Minn Consortium

(Principal Investigators: David Van Essen and Kamil Ugurbil; 1U54MH091657) funded

by the 16 NIH Institutes and Centers that support the NIH Blueprint for Neuroscience

Research; and by the McDonnell Center for Systems Neuroscience at Washington

University. In addition, FLR acknowledges support through the Australian Government

Research Training Program Scholarship. We thank the reviewers (Elisha Merriam,

Tomas Knapen, and Noah C. Benson) for helpful comments on early versions of our

manuscript and Mark Schira for helpful discussions.

Competing financial interests

The authors declare no competing financial interests.

References

Abraham, A., Pedregosa, F., Eickenberg, M., Gervais, P., Mueller, A., Kossaifi, J.,

Gramfort, A., Thirion, B., Varoquaux, G., 2014. Machine learning for neuroimaging

with   scikit-learn.   Front.   Neuroinform.   8,   1–10.

https://doi.org/10.3389/fninf.2014.00014

Allen, E.J., St-Yves, G., Wu, Y., Breedlove, J.L., Prince, J.S., Dowdle, L.T., Nau, M.,

Caron, B., Pestilli, F., Charest, I., Hutchinson, J.B., Naselaris, T., Kay, K., 2021.

A massive 7T fMRI dataset to bridge cognitive neuroscience and artificial

intelligence. Nat. Neurosci. https://doi.org/10.1038/s41593-021-00962-x

Angelucci, A., Rosa, M.G.P., 2015. Resolving the organization of the third tier visual

cortex in primates: A hypothesis-based approach. Vis. Neurosci. 32.

https://doi.org/10.1017/S0952523815000073

Arcaro, M.J., Kastner, S., 2015. Topographic organization of areas V3 and V4 and its

relation to supra-areal organization of the primate visual system. Vis. Neurosci.

https://doi.org/doi:10.1017/S0952523815000115

Benson, N.C., Butt, O.H., Brainard, D.H., Aguirre, G.K., 2014. Correction of Distortion

in Flattened Representations of the Cortical Surface Allows Prediction of V1-V3

Functional   Organization   from   Anatomy.   PLoS   Comput.   Biol.   10.

https://doi.org/10.1371/journal.pcbi.1003538

Benson, N.C., Butt, O.H., Datta, R., Radoeva, P.D., Brainard, D.H., Aguirre, G.K.,

2012. The retinotopic organization of striate cortex is well predicted by surface

topology. Curr. Biol. 22, 2081–2085. https://doi.org/10.1016/j.cub.2012.09.014

Benson, N.C., Jamison, K.W., Arcaro, M.J., Vu, A.T., Glasser, M.F., Coalson, T.S.,

869 Van Essen, D.C., Yacoub, E., Ugurbil, K., Winawer, J., Kay, K., 2018. The Human

870 Connectome Project 7 Tesla retinotopy dataset : Description and population

871 receptive field analysis. J. Vis. 18, 1–22.

872 Benson, N.C., Winawer, J., 2018. Bayesian analysis of retinotopic maps. Elife 7, 1–

873 29. https://doi.org/10.7554/elife.40224

874 Bollmann, S., Mattern, H., Bernier, M., Robinson, S.D., Park, D., Speck, O., Polimeni,

875 J.R., 2022. Imaging of the pial arterial vasculature of the human brain in vivo using

876 highresolution 7T time-of-flight angiography. Elife 11, 1–35.

877 https://doi.org/10.7554/eLife.71186

878 Boyd Taylor, H.C., Puckett, A.M., Isherwood, Z.J., Schira, M.M., 2019. Vascular

879 effects on the BOLD response and the retinotopic mapping of hV4. PLoS One 14.

880 Dougherty, R.F., Koch, V.M., Brewer, A.A., Fischer, B., Modersitzki, J., Wandell, B.A.,

881 2003. Visual field representations and locations of visual areas V1/2/3 in human

882 visual cortex. J. Vis. 3, 586–598. https://doi.org/10.1167/3.10.1

883 Dumoulin, S.O., Wandell, B.A., 2008. Population receptive field estimates in human

884 visual cortex. Neuroimage 39, 647–660.

885 https://doi.org/10.1016/j.neuroimage.2007.09.034

886 Durbin, R., Mitchison, G., 1990. A dimension reduction framework. Nature 343, 644–

887 647.

888 Gattass, R., Gross, C.G., Sandell, J.H., 1981. Visual topography of V2 in the macaque.

889 J. Comp. Neurol. 201, 519–539. https://doi.org/10.1002/cne.902010405

890 Gattass, R., Sousa, A.P.B., Gross, C.G., 1988. Visuotopic organization and extent of

891 V3 and V4 of the macaque. J. Neurosci. 8, 1831–1845.

https://doi.org/10.1523/jneurosci.08-06-01831.1988

Gegenfurtner, K.R., Kiper, D.C., Levitt, J.B., 1997. Functional properties of neurons in

macaque area V3. J. Neurophysiol. 77, 1906–1923.

https://doi.org/10.1152/jn.1997.77.4.1906

Glasser, M.F., Coalson, T.S., Robinson, E.C., Hacker, C.D., Harwell, J., Yacoub, E.,

2016. A multi-modal parcellation of human cerebral cortex. Nature 536, 171–178.

https://doi.org/10.1038/nature18933

Glasser, M.F., Sotiropoulos, S.N., Wilson, J.A., Coalson, T.S., Fischl, B., Andersson,

J.L., Xu, J., Jbabdi, S., Webster, M., Polimeni, J.R., Van Essen, D.C., Jenkinson,

M., 2013. The minimal preprocessing pipelines for the Human Connectome

Project. Neuroimage 80, 105–124.

https://doi.org/10.1016/j.neuroimage.2013.04.127

Haak, K. V., Winawer, J., Harvey, B.M., Renken, R., Dumoulin, S.O., Wandell, B.A.,

Cornelissen, F.W., 2013. Connective field modeling. Neuroimage 66, 376–384.

https://doi.org/doi:10.1016/j.neuroimage.2012.10.037

Hadjidimitrakis, K., Bakola, S., Chaplin, T.A., Yu, H.H., Alanazi, O., Chan, J.M.,

Worthy, K.H., Rosa, M.G.P., 2019. Topographic organization of the 'third-tier'

dorsomedial visual cortex in the macaque. J. Neurosci. 39, 5311–5325.

https://doi.org/10.1523/JNEUROSCI.0085-19.2019

Harris, C.R., Millman, K.J., van der Walt, S.J., Gommers, R., Virtanen, P.,

Cournapeau, D., Wieser, E., Taylor, J., Berg, S., Smith, N.J., Kern, R., Picus, M.,

Hoyer, S., van Kerkwijk, M.H., Brett, M., Haldane, A., del Río, J.F., Wiebe, M.,

Peterson, P., Gérard-Marchant, P., Sheppard, K., Reddy, T., Weckesser, W.,

Abbasi, H., Gohlke, C., Oliphant, T.E., 2020. Array programming with NumPy.

Nature 585, 357–362. https://doi.org/10.1038/s41586-020-2649-2

Hinds, O.P., Rajendran, N., Polimeni, J.R., Augustinack, J.C., Wiggins, G., Wald, L.L.,

Rosas, H.D., Potthast, A., Schwarts, E.L., Fischl, B., 2009. Accurate prediction of

V1 location from cortical folds in a surface coordinate system. Neuroimage 39,

1585–1599.

Hoffmann, M.B., Stadler, J., Kanowski, M., Speck, O., 2009. Retinotopic mapping of

the human visual cortex at a magnetic field strength of 7 T. Clin. Neurophysiol.

120, 108–116. https://doi.org/10.1016/j.clinph.2008.10.153

Holmes, G., Lister, W.T., 1916. Disturbances of Vision from Cerebral Lesions, with

Special Reference to the Cortical Representation of the Macula. Brain 39, 34–73.

Horton, J.C., Hoyt, W.F., 1991. The Representation of the Visual Field in Human

Striate Cortex: A Revision of the Classic Holmes Map. Arch. Ophthalmol. 109,

816–824. https://doi.org/10.1001/archopht.1991.01080060080030

Hubel, D.H., Wiesel, T.N., 1974. Uniformity of monkey striate cortex: A parallel

relationship between field size, scatter, and magnification factor. J. Comp. Neurol.

158, 295–305. https://doi.org/10.1002/cne.901580305

Inouye, T., 1909. Die Sehstörungen bei Schussverletzungen der kortikalen

Sehsphäre: nach Beobachtungen an Verwundeten der letzten japanischen

Kriege. W. Engelmann.

Kay, K.N., Winawer, J., Mezer, A., Wandell, B.A., 2013. Compressive spatial

summation in human visual cortex. J. Neurophysiol. 110, 481–494.

https://doi.org/10.1152/jn.00105.2013

Kurzawski, J.W., Gulban, O.F., Jamison, K., 2022. Non-Neural Factors Influencing

BOLD Response Magnitudes within Individual Subjects. J. Neurosci. 42, 7256–

7266.

Levandowsky, M., Winter, D., 1971. Distance between sets. Nature 234, 34–35.

https://doi.org/10.1038/234034a0

Levitt, J.B., Kiper, D.C., Movshon, J.A., 1994. Receptive fields and functional

architecture   of   macaque   V2.   J.   Neurophysiol.   71,   2517–2542.

https://doi.org/10.1152/jn.1994.71.6.2517

Luxburg, U. Von, 2007. A tutorial on spectral clustering. Stat. Comput. 17, 395–416.

https://doi.org/10.1007/s11222-007-9033-z

Magezi, D.A., 2015. Linear mixed-effects models for within-participant psychology

experiments: An introductory tutorial and free, graphical user interface (LMMgui).

Front. Psychol. 6, 1–7. https://doi.org/10.3389/fpsyg.2015.00002

Manger, P.R., Kiper, D., Masiello, I., Murillo, L., Tettoni, L., Hunyadi, Z., Innocenti,

G.M., 2002. The Representation of the Visual Field in Three Extrastriate Areas of

the Ferret (Mustela putorius) and the Relationship of Retinotopy and Field

Boundaries   to   Callosal   Connectivity.   Cereb.   Cortex   12,   423–437.

https://doi.org/10.1093/cercor/12.4.423

Pedregosa, F., Varoquaux, G., Gramfort, A., Michel, V., Thirion, B., Grisel, O., Blondel,

M., Prettenhofer, P., Weiss, R., Dubourg, V., Vanderplas, J., Passos, A.,

Cournapeau, D., Brucher, M., Perrot, M., Duchesnay, E., 2011. Scikit-learn:

Machine Learning in Python. J. Mach. Learn. Res. 12, 2825–2830.

Pitzalis, S., Sereno, M.I., Committeri, G., Fattori, P., Galati, G., Patria, F., Galletti, C.,

2010. Human V6: The medial motion area. Cereb. Cortex 20, 411–424.

https://doi.org/10.1093/cercor/bhp112

Polimeni, J.R., Fischl, B., Greve, D.N., Wald, L.L., 2010. Laminar analysis of 7T BOLD

964   using an imposed spatial activation pattern in human V1. Neuroimage 52, 1334–

965   1346. https://doi.org/10.1016/j.neuroimage.2010.05.005

966   Puckett, A.M., Aquino, K.M., Robinson, P.A., Breakspear, M., Schira, M.M., 2016. The

967   spatiotemporal hemodynamic response function for depth-dependent functional

968   imaging of human cortex. Neuroimage 139, 240–248.

969   https://doi.org/10.1016/j.neuroimage.2016.06.019

970   Rajimehr, R., Tootell, R.B.H., 2009. Does retinotopy influence cortical folding in

971   primate visual cortex? J. Neurosci. 29, 11149–11152.

972   https://doi.org/10.1523/JNEUROSCI.1835-09.2009

973   Renton, A.I., Dao, T.T., Johnstone, T., Civier, O., Sullivan, R.P., White, D.J., Lyons,

974   P., Slade, B.M., Abbott, D.F., Amos, T.J., Bollmann, Saskia, Botting, A.,

975   Campbell, M.E.J., Chang, J., Close, T.G., Eckstein, K., Egan, G.F., Evas, S.,

976   Flandin, G., Garner, K.G., Garrido, M.I., Ghosh, S.S., Grignard, M., Hannan, A.J.,

977   Huber, R., Kaczmarzyk, J.R., Kasper, L., Kuhlmann, L., Lou, K., Mantilla-Ramos,

978   Y.-J., Mattingley, J.B., Morris, J., Narayanan, Akshaiy, Pestilli, F., Puce, A.,

979   Ribeiro, F.L., Rogasch, N.C., Rorden, C., Schira, M., Shaw, T.B., Sowman, P.F.,

980   Spitz, G., Stewart, A., Ye, X., Zhu, J.D., Hughes, M.E., Narayanan, Aswin,

981   Bollmann, Steffen, 2022. Neurodesk: An accessible, flexible, and portable data

982   analysis environment for reproducible neuroimaging. bioRxiv 1–45.

983   https://doi.org/https://doi.org/10.1101/2022.12.23.521691

984   Ribeiro, F.L., Bollmann, S., Puckett, A.M., 2021. Predicting the retinotopic organization

985   of human visual cortex from anatomy using geometric deep learning. Neuroimage

244, 118624. https://doi.org/https://doi.org/10.1016/j.neuroimage.2021.118624

Rilling, J.K., Insel, T.R., 1999. Differential expansion of neural projection systems in

primate brain evolution. Neuroreport 10, 1453–1459.

https://doi.org/10.1097/00001756-199905140-00012

Rosa, M.G.P., 2002. Visual maps in the adult primate cerebral cortex: some

implications for brain development and evolution. Brazilian J. Med. Biol. Res. 35,

1485–1498. https://doi.org/10.1590/S0100-879X2002001200008

Rosa, M.G.P., 1999. Topographic organisation of extrastriate areas in the flying fox:

Implications for the evolution of mammalian visual cortex. J. Comp. Neurol. 411,

503–523. https://doi.org/10.1002/(SICI)1096-9861(19990830)411:3<503::AID-

CNE12>3.0.CO;2-6

Rosa, M.G.P., 1997. Visuotopic Organization of Primate Extrastriate Cortex, in:

Rockland, K.S., Kaas, J.H., Peters, A. (Eds.), Extrastriate Cortex in Primates.

Springer US, Boston, MA, pp. 127–203. https://doi.org/10.1007/978-1-4757-

9625-4_4

Rosa, M.G.P., Fritsches, K.A., Elston, G.N., 1997. The second visual area in the

marmoset monkey: Visuotopic organisation, magnification factors, architectonical

boundaries, and modularity. J. Comp. Neurol. 387, 547–567.

https://doi.org/10.1002/(SICI)1096-9861(19971103)387:4<547::AID-

CNE6>3.0.CO;2-2

Rosa, M.G.P., Manger, P.R., 2005. Clarifying homologies in the mammalian cerebral

cortex: The case of the third visual area (V3). Clin. Exp. Pharmacol. Physiol. 32,

327–339. https://doi.org/10.1111/j.1440-1681.2005.04192.x

Rosa, M.G.P., Schmid, L.M., 1995. Visual areas in the dorsal and medial extrastriate

cortices of the marmoset. J. Comp. Neurol. 359, 272–299.

https://doi.org/10.1002/cne.903590207

Rosa, M.G.P., Tweedale, R., 2005. Brain maps, great and small: Lessons from

comparative studies of primate visual cortical organization. Philos. Trans. R. Soc.

B Biol. Sci. 360, 665–691. https://doi.org/10.1098/rstb.2005.1626

Rosa, M.G.P., Tweedale, R., 2000. Visual areas in lateral and ventral extrastriate

cortices of the marmoset monkey. J. Comp. Neurol. 422, 621–651.

https://doi.org/10.1002/1096-9861(20000710)422:4<621::AID-CNE10>3.0.CO;2-

E

Schira, M.M., Tyler, C.W., Breakspear, M., Spehar, B., 2009. The foveal confluence in

human visual cortex. J. Neurosci. 29, 9050–9058.

https://doi.org/10.1523/JNEUROSCI.1760-09.2009

Schira, M.M., Tyler, C.W., Spehar, B., Breakspear, M., 2010. Modeling magnification

and anisotropy in the primate foveal confluence. PLoS Comput. Biol. 6, 1–10.

https://doi.org/10.1371/journal.pcbi.1000651

Sedigh-Sarvestani, M., Lee, K.-S., Jaepel, J., Satterfield, R., Shultz, N., Fitzpatrick, D.,

2021. A sinusoidal transformation of the visual field is the basis for periodic maps

in area V2. Neuron 109, 4068-4079.e6.

https://doi.org/10.1016/j.neuron.2021.09.053

Sereno, M., Dale, A., Reppas, J., Kwong, K., Belliveau, J., Brady, T., Rosen, B.,

Tootell, R., 1995. Borders of multiple visual areas in humans revealed by

functional magnetic resonance imaging. Science (80-.). 268, 889–893.

Sereno, M.I., Mcdonald, C.T., Allman, J.M., 1994. Analysis of retinotopic maps in extrastriate cortex. Cereb. Cortex 4, 601–620. https://doi.org/10.1093/cercor/4.6.601

Sereno, M.I., McDonald, C.T., Allman, J.M., 2015. Retinotopic organization of extrastriate cortex in the owl monkey—dorsal and lateral areas. Vis. Neurosci. 32, E021. https://doi.org/10.1017/S0952523815000206

Swindale, N. V, 1996. The development of topography in the visual cortex: a review of models. Network 7, 161–247. https://doi.org/10.1088/0954-898X/7/2/002

Taha, A.A., Hanbury, A., 2015. Metrics for evaluating 3D medical image segmentation: Analysis, selection, and tool. BMC Med. Imaging 15. https://doi.org/10.1186/s12880-015-0068-x

Tangtartharakul, G.T., Morgan, C.A., Rushton, S.K., Schwarzkopf, D.S., 2023. Retinotopic connectivity maps of human visual cortex with unconstrained eye movements. bioRxiv 1–32. https://doi.org/https://doi.org/10.1101/2023.03.16.533037

The jamovi project (2021), n.d.

Tusa, R.J., Rosenquist, A.C., Palmer, L.A., 1979. Retinotopic organization of areas 18 and 19 in the cat. J. Comp. Neurol. 185, 657–678. https://doi.org/10.1002/cne.901850405

Van Essen, D.C., Glasser, M.F., 2018. Parcellating Cerebral Cortex: How Invasive Animal Studies Inform Noninvasive Mapmaking in Humans. Neuron 99, 640–663. https://doi.org/10.1016/j.neuron.2018.07.002

Van Essen, D.C., Smith, S.M., Barch, D.M., Behrens, T.E.J., Yacoub, E., Ugurbil, K.,

2013. The WU-Minn Human Connectome Project: An overview. Neuroimage 80,

62–79. https://doi.org/10.1016/j.neuroimage.2013.05.041

Virtanen, P., Gommers, R., Oliphant, T.E., Haberland, M., Reddy, T., Cournapeau, D.,

Burovski, E., Peterson, P., Weckesser, W., Bright, J., van der Walt, S.J., Brett,

M., Wilson, J., Millman, K.J., Mayorov, N., Nelson, A.R.J., Jones, E., Kern, R.,

Larson, E., Carey, C.J., Polat, İ., Feng, Y., Moore, E.W., VanderPlas, J., Laxalde,

D., Perktold, J., Cimrman, R., Henriksen, I., Quintero, E.A., Harris, C.R.,

Archibald, A.M., Ribeiro, A.H., Pedregosa, F., van Mulbregt, P., Vijaykumar, A.,

Bardelli, A. Pietro, Rothberg, A., Hilboll, A., Kloeckner, A., Scopatz, A., Lee, A.,

Rokem, A., Woods, C.N., Fulton, C., Masson, C., Häggström, C., Fitzgerald, C.,

Nicholson, D.A., Hagen, D.R., Pasechnik, D. V., Olivetti, E., Martin, E., Wieser,

E., Silva, F., Lenders, F., Wilhelm, F., Young, G., Price, G.A., Ingold, G.L., Allen,

G.E., Lee, G.R., Audren, H., Probst, I., Dietrich, J.P., Silterra, J., Webber, J.T.,

Slavič, J., Nothman, J., Buchner, J., Kulick, J., Schönberger, J.L., de Miranda

Cardoso, J.V., Reimer, J., Harrington, J., Rodríguez, J.L.C., Nunez-Iglesias, J.,

Kuczynski, J., Tritz, K., Thoma, M., Newville, M., Kümmerer, M., Bolingbroke, M.,

Tartre, M., Pak, M., Smith, N.J., Nowaczyk, N., Shebanov, N., Pavlyk, O.,

Brodtkorb, P.A., Lee, P., McGibbon, R.T., Feldbauer, R., Lewis, S., Tygier, S.,

Sievert, S., Vigna, S., Peterson, S., More, S., Pudlik, T., Oshima, T., Pingel, T.J.,

Robitaille, T.P., Spura, T., Jones, T.R., Cera, T., Leslie, T., Zito, T., Krauss, T.,

Upadhyay, U., Halchenko, Y.O., Vázquez-Baeza, Y., 2020. SciPy 1.0:

fundamental algorithms for scientific computing in Python. Nat. Methods 17, 261–

272. https://doi.org/10.1038/s41592-019-0686-2

Wandell, B.A., Winawer, J., 2010. Imaging retinotopic maps in the human brain. Vision

Res. 51, 718–737. https://doi.org/10.1016/j.visres.2010.08.004.Imaging

Wang, L., Mruczek, R.E.B., Arcaro, M.J., Kastner, S., 2015. Probabilistic maps of
visual topography in human cortex. Cereb. Cortex 25, 3911–3931.
https://doi.org/10.1093/cercor/bhu277

Winawer, J., Horiguchi, H., Sayres, R.A., Amano, K., Wandell, B.A., 2010. Mapping
hV4 and ventral occipital cortex: The venous eclipse. J. Vis. 10, 1–22.
https://doi.org/10.1167/10.5.1

Wolf, F., Bauer, H.U., Geisel, T., 1994. Formation of field discontinuities and islands
in visual cortical maps. Biol. Cybern. 70, 525–531.
https://doi.org/10.1007/BF00198805

Yu, H.H., Rowley, D.P., Price, N.S.C., Rosa, M.G.P., Zavitz, E., 2020. A twisted visual
field map in the primate cortex predicted by topographic continuity. Sci. Adv.
https://doi.org/10.1101/682187

Yu, Z., Guindani, M., Grieco, S.F., Chen, L., Holmes, T.C., Xu, X., 2022. Primer
Beyond t test and ANOVA : applications of mixed-effects models for more rigorous
statistical analysis in neuroscience research. Neuron 110, 21–35.
https://doi.org/10.1016/j.neuron.2021.10.030

Zeki, S.M., 1978. Uniformity and diversity of structure and function in rhesus monkey
prestriate visual cortex. J. Physiol. 277, 273–290.
https://doi.org/https://doi.org/10.1113/jphysiol.1978.sp012272

Zhu, Q., Vanduffel, W., 2019. Submillimeter fMRI reveals a layout of dorsal visual
cortex in macaques, remarkably similar to New World monkeys. Proc. Natl. Acad.
Sci. U. S. A. 116, 2306–2311. https://doi.org/10.1073/pnas.1805561116

1102