Dual receptive fields underlying target and wide-field motion sensitivity in looming sensitive descending neurons

Short title: Dual receptive fields in descending neurons

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Keywords

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Abstract

Responding rapidly to visual stimuli is fundamental for many animals. For example, predatory birds and insects alike have amazing target detection abilities, with incredibly short neural and behavioral delays, enabling efficient prey capture. Similarly, looming objects need to be rapidly avoided to ensure immediate survival, as these could represent approaching predators. Male *Eristalis tenax* hoverflies are non-predatory, highly territorial insects, that perform high-speed pursuits of conspecifics and other territorial intruders. During the initial stages of the pursuit the retinal projection of the target is very small, but grows to a larger object before physical interaction. Supporting such behaviors, *E. tenax* and other insects have both target-tuned and loom-sensitive neurons in the optic lobes and the descending pathways. We here show that these visual stimuli are not necessarily encoded in parallel. Indeed, we describe a class of descending neurons that respond to small targets, to looming and to widefield stimuli. We show that these neurons have two distinct receptive fields where the dorsal receptive field is sensitive to the motion of small targets and the ventral receptive field responds to larger objects or widefield stimuli. Our data suggest that the two receptive fields have different pre-synaptic input, where the inputs are not linearly summed. This novel and unique arrangement could support different behaviors, including obstacle avoidance, flower landing, target pursuit or capture.

Significance Statement

If you are playing baseball, when the ball is far away, it appears as a very small object on your retina. However, as the ball gets closer, its image becomes a rapidly expanding object. Here, we show that within the hoverfly visual system, a single neuron could respond to both of these images. Indeed, we found a class of descending neurons with dual sensitivity, separated into two distinct parts of the visual field. The neurons have a more dorsal receptive field that is sensitive to small targets and a more ventral receptive field that is sensitive to larger objects.

Introduction

The ability to respond quickly to visual stimuli is vital for the survival of many animals. Indeed, visual input may be used for a variety of tasks, from navigating around obstacles, choosing a suitable surface to rest upon, and also for detecting predators or prey. For example, many predatory insects rely on vision to identify suitable prey and engage in pursuits, doing so with astonishing precision and accuracy (e.g. Olberg et al., 2007; Nityananda et al., 2016; Fabian et al., 2018). Some non-predatory insects, including hoverflies, also have superb target detecting capabilities, which they may use for territorial defense or courtship (Fitzpatrick and Wellington, 1983; Zeil, 1986). Small target motion detector (STMD) neurons in the hoverfly optic lobe, and their presumed post-synaptic targets, the target selective descending neurons (TSDNs), have size and velocity tuning properties that match the target image at the start of the pursuit, suggesting that they could support these behaviors (Nordström et al., 2006; Nicholas et al., 2020; Thyselius et al., 2023).

In addition to the precise responses which occur during pursuit, behaviors elicited by looming stimuli , such as the escape response, also need to be fast and accurate (e.g. Fotowat et al., 2009; Santer et al., 2012; von Reyn et al., 2017; Mancienne et al., 2021; Lenzi et al., 2022). Indeed, neurons that respond strongly to rapidly looming stimuli exist in a range of species and visual structures, including the cat superior colliculus (Liu et al., 2011), the bullfrog optic tectum (Nakagawa and Hongjian, 2010), and in zebrafish retinal ganglion cells (Temizer et al., 2015). In insects, there are looming neurons in the optic lobes as well as in the descending pathways. Examples of these include the locust LGMD/DCMD system (Santer et al., 2012), the *Drosophila* Foma-1 neurons (de Vries and Clandinin, 2012), and the descending giant fiber (Fotowat et al., 2009).

Historically, insect looming neurons have been studied in the context of predator avoidance (e.g. Fotowat et al., 2009; Santer et al., 2012; von Reyn et al., 2017). However, there is emerging evidence that looming neurons also play a key role in pursuit behaviors. For example, silencing *Drosophila* Foma-1 neurons not only affects the escape response (de Vries and Clandinin, 2012), but also the ability of male flies to follow females during courtship (Coen et al., 2016). This is interesting as many looming neurons also respond strongly to small moving targets. For example, the locust LGMD/DCMD pathway was originally thought to play a role in object tracking (Rind and Simmons, 1992), and several looming sensitive neurons in the locust central complex also respond to small moving targets (Rosner and Homberg, 2013). Conversely, some dragonfly TSDNs respond not only to targets but also to looming stimuli (Frye and Olberg, 1995; Gonzalez-Bellido et al., 2013).

Taken together, this suggests that some neurons classically defined as either target or looming selective (Santer et al., 2008; Gonzalez-Bellido et al., 2013) respond to both. Like locusts (Santer et al., 2008) and dragonflies (Gonzalez-Bellido et al., 2013), hoverflies have descending neurons that respond to both looming stimuli and to small moving targets (Nicholas et al., 2020). We here investigate this dual sensitivity and find that these descending neurons have two distinct receptive fields, one more dorsal that responds selectively to the motion of small targets, and one more ventral that responds to larger objects, including sinusoidal gratings and high-contrast edges. We show that when the center of the ventral grating receptive field is to the right of the visual midline, the local motion sensitivity is to rightward motion, and vice versa. However, the preferred direction of the dorsal target receptive field and the ventral grating receptive field are not always the same. We also show that the two receptive fields receive separate input, from the pre-synaptic target pathway and the pre-synaptic widefield motion pathway, respectively, and when stimulated

103 simultaneously the responses are not linearly summed. We hypothesize that the unique
104 response characteristics of these neurons could be used in different behaviors.

105 Materials and Methods

106 *Electrophysiology*

107 We recorded from 98 looming sensitive descending neurons (Nicholas et al., 2020) in 94
108 male *Eristalis tenax* hoverflies, reared and maintained in-house as described previously
109 (Nicholas et al., 2018a). At experimental time the hoverfly was immobilized ventral side up,
110 using a beeswax and resin mixture, before an opening was made in the thoracic cavity. A
111 small silver hook was used to elevate and support the cervical connective and a silver wire
112 inside the opening served as a reference electrode.

113 Recordings were made from the cervical connective using a sharp polyimide-insulated
114 tungsten microelectrode (2 MOhm, Microprobes, Gaithersburg, USA). Signals were
115 amplified at 100x gain and filtered through a 10 – 3000 Hz bandwidth filter using a DAM50
116 differential amplifier (World Precision Instruments, Sarasota, USA), with 50 Hz noise
117 removed with a HumBug (Quest Scientific, North Vancouver, Canada). The data were
118 digitized via a Powerlab 4/30 and recorded at 40 kHz with LabChart 7 Pro software
119 (ADInstruments, Sydney, Australia). Single units were discriminated by amplitude and half-
120 width using Spike Histogram software (ADInstruments).

121 *Visual stimuli*

122 Hoverflies were positioned perpendicular to and 6.5 cm away from the middle of a linearized
123 Asus LCD screen (Asus, Taipei, Taiwan) with a mean illuminance of 200 Lux, a refresh rate
124 of 165 Hz and a spatial resolution of 2,560 × 1,440 pixels (59.5 × 33.5 cm), giving a

projected screen size of 155 × 138°. Visual stimuli were displayed using custom software written in Matlab (R2019b, Mathworks) using the Psychophysics toolbox (Brainard, 1997; Pelli, 1997). The stimuli were not perspective corrected. When values are given in degrees, this corresponds to the retinal size in the center of the visual field. Velocities are given in pixels/s.

Potential looming sensitive descending neurons were initially identified based on their response to a small, black, moving target (left, Fig. 1A-D, and see Nicholas et al., 2020). Those neurons that responded stronger to a looming stimulus compared to an appearance control (Nicholas et al., 2020) were kept for further analysis (Fig. 1E, F). The looming stimulus was a black circle on a white background, expanding over 1 s from 1° diameter to 118° (right, Fig. 1A, C), with a 10 ms rate of expansion (Fotowat and Gabbiani, 2007), also referred to as $l/|v|$. The appearance control was a black circle with 118° diameter that appeared and remained on the screen for 1 s.

We mapped the target receptive field (Nicholas et al., 2020) of each neuron by scanning a target horizontally and vertically at 20 evenly spaced elevations and azimuths to create a 20 x 20 grid (Fig. S1A). The 15 x 15 pixel (3 x 3°) black, square target moved at a velocity of 900 pixels/s. There was a minimum 0.5 s interval between each stimulation.

We mapped the grating receptive field (Nicholas et al., 2020) using local sinusoidal gratings (400 x 400 pixels, 71 x 71° in the visual field center, Fig. S1B) where the internal pattern moved in a series of 8 different directions presented in a pseudorandom order for 0.36 s each. The gratings had a wavelength of 75 pixels (13° for the central patch, 0.08 cpd) and drifted at 5 Hz. The local gratings were placed in an overlapping tiling fashion so that 8 x 6 (width x height) squares covered the majority of the screen (Fig. S1B). There was a minimum 1 s interval between each stimulation.

To map the leading-edge receptive field, we scanned the entire height or width of the screen with an OFF-contrast edge moving left, right, down or up, at 900 pixels/s (Fig. S3A-D).

For size tuning experiments a bar drifted at 900 pixels/s, in the preferred direction of each neuron's target receptive field. The bar drifted either horizontally or vertically through the center of the target or grating receptive field, as specified. The bar side parallel to the direction of travel was maintained at a fixed size of 15 pixels (3°) whilst the perpendicular side varied from 0.2° to 155° (1 pixel to the full height of the screen). When presented simultaneously with a small target, the bar moved in the preferred horizontal direction through the grating receptive field only. In this case the bar height was varied between 5.7° and 106° (28 to 749 pixels), whilst a fixed size 3 x 3° target moved through the target receptive field.

To determine the input mechanism of each receptive field, an OFF edge, an ON edge and a discrete bar, with a width of 15 pixels (3°) drifted horizontally at 900 pixels/s across the entire width of the screen. The height of these objects was 3° (15 pixels) when drifted through the target receptive field, 84° (500 pixels) when drifted through the grating receptive field, or the height of the screen (138°) to cover both receptive fields.

All stimuli were presented in a random order, except for the receptive field stimuli, which were presented in a pseudo-random order.

Experimental Design and Statistical Analyses

All data analysis was performed in Matlab and GraphPad Prism 9.3.1 (GraphPad Software, USA). Statistical analysis was done using either GraphPad Prism 9.3.1 or the circular statistics toolbox (Berens, 2009) in Matlab, as appropriate. The sample size, statistical test and P-values are indicated in each figure legend, where *n* refers to the number of repetitions

172 within one neuron and *N* to the number of neurons. Neurons were initially identified based on
173 the response to a small target, with data from all neurons that subsequently passed our
174 definition of a looming neuron (Nicholas et al., 2020, and see above) included in the analysis.

175 For target receptive field mapping we used the resulting 20 x 20 grid (Fig. S1A) to calculate
176 the local preferred direction and local motion sensitivity (Fig. S1C), assuming a neural delay
177 of 20 ms. We calculated the local average response to the four directions of motion (dotted
178 line, Fig. S1C) after subtracting the spontaneous rate, calculated in the 485 ms prior to
179 stimulus presentation. We interpolated this to a 100 x 100 grid to generate receptive field
180 maps (Fig. S1E) using Matlab's *contour* function. We defined the center of the receptive field
181 (Fig. S1E) as the center of the 50% contour line using Matlab's *centroid* function. We fitted a
182 cosine function (Nicholas et al., 2020) to the response to the four directions of motion
183 (Nordström et al., 2006) and extracted its local preferred direction and amplitude (Fig. S1C).
184 We calculated the preferred direction for each neuron by averaging the local preferred
185 directions from the locations where the local motion sensitivity was above 50% of the
186 maximum (blue, Fig. S1G).

187 For the grating receptive fields we used the resulting 8 x 6 grid (Fig. S1B) to quantify the
188 local mean response for each direction of motion, after removing the first 100 ms of the
189 response, to avoid any initial onset transients (Nordström and O'Carroll, 2009). We
190 calculated the local mean response (dotted line, Fig. S1D) after subtracting the spontaneous
191 rate, calculated for 800 ms preceding stimulus presentation. We spatially interpolated this 10
192 times and calculated the center from the 50% contour line (Fig. S1F). For each spatial
193 location we fitted a cosine function (Fig. S1D) to the response to get the local preferred
194 direction and local motion sensitivity (Nicholas et al., 2020). We calculated the overall
195 direction selectivity using the top 50% of the local preferred directions (red, Fig. S1H).

We calculated the horizontal and vertical distance between the receptive field centers and the midline and equator. The distance between the two receptive field centers was calculated using the Euclidean distance.

For leading-edge receptive field mapping we first quantified the spike histogram for each neuron, after smoothing the response with a 100 ms square-wave filter with 40 kHz resolution (Fig. S3A-D). We identified the maximum response to any direction of motion (purple, Fig. S3D). 50% maximum response was used as a threshold to determine the limits of the leading-edge receptive field (cyan, Fig. S3A, C, D). If a neuron's response did not reach threshold to one direction of motion (e.g. Fig. S3B), the opposite direction of motion determined the receptive field outlines. If a neuron responded to both directions of motion (e.g. up and down) the outer thresholds were used to delineate the receptive field (Fig. S3E). From the resulting rectangular receptive field, we determined the center, and the proximity to the target and the grating receptive fields respectively (Fig. S3F), using the following proximity index:

$$(d1 - d2) / (d1 + d2)$$

where *d1* is the Euclidean distance between the leading edge and the target receptive field centers, and *d2* the Euclidean distance between the leading edge and the grating receptive field centers (Fig. 4B). Thus, if the leading-edge receptive field center was closer to the grating receptive field center, the proximity index was positive, but if it was closer to the target receptive field center the proximity index was negative.

For all stimuli other than receptive field mapping, quantification of responses was done by averaging the spike rate within a 0.56 s analysis window centered on each neuron's target or grating receptive field center, as specified.

Code Accessibility

All Matlab scripts used for data analysis in this paper can be found here:
https://doi.org/10.5281/zenodo.7227236

Data Accessibility

All raw and analyzed data presented here have been deposited to DataDryad:
https://doi.org/10.5061/dryad.6wwpzgn2p

Private link for peer review:
https://datadryad.org/stash/share/jZnw3f4ZNuytFZjEqDp2WNyAmKo32GGWTrWgTnywJaU

Results

Looming neurons have dorsal target receptive fields and ventral grating receptive fields

We recorded from 98 looming sensitive descending neurons in male *Eristalis tenax* hoverflies. The neurons described here responded both to small target motion (left, Fig. 1A-D) and to looming stimuli (right, Fig. 1A-D). The response to a looming stimulus (right, Fig. 1B, D) started well before the stimulus reached its full size (right, Fig. 1C), and was much stronger than the response to an appearance control (Fig. 1E, F, and see Nicholas et al., 2020).

To investigate this dual sensitivity (Fig. 1A-D) in more detail, we mapped the receptive fields using two different methods. For this purpose we either scanned the visual monitor with a small, black target (Nordström et al., 2006) moving in four different directions (Fig. S1A, C), or we used a local sinusoidal grating (Fig. S1B, D) where the internal pattern drifted in eight

240 different directions (Nicholas et al., 2020). The data from an example neuron show two
241 distinct receptive fields, with a dorsal target receptive field (blue, Fig. 2A, S1E), and a ventral
242 grating receptive field (red, Fig. 2A, S1F).

243 We used the 50% response contours to locate the two receptive field centers (blue and red
244 circle, Fig. 2A, S1E, F). Across the 98 neurons, the target receptive field centers cluster
245 above the visual equator (blue, Fig. 2B), whereas the grating receptive field centers cluster
246 below the equator (red, Fig. 2B), even if there are some exceptions (see also Fig. S2). We
247 quantified the vertical distance between each receptive field center and the visual equator
248 (44° and -50° in the example neuron, Fig. 2A), and found a bimodal distribution with target
249 receptive field centers peaking 36° dorsal (median value) and grating receptive field centers
250 peaking -36° (ventral, Fig. 2B, C).

251 We next quantified the horizontal distance between each receptive field center and the visual
252 midline (14° and 21° in the example neuron, Fig. 2A). Across neurons we found a bimodal
253 distribution with a gap along the visual midline (Fig. 2B, D). The grating receptive field
254 center medians were at -27° and 19°, whereas the target receptive field center medians were
255 at -25° and 20° (Fig. 2B, D). There was no significant difference between the target and the
256 grating receptive field center distributions (Mann-Whitney test, left visual field, $P = 0.31$,
257 right visual field, $P = 0.40$).

258 We noted that the target receptive field was often in the dorsal visual field, but that there
259 were some exceptions (Fig. 2B, C, S2). We next investigated if the target receptive field was
260 always dorsal to the grating receptive field and determined the Euclidean distance between
261 the two receptive field centers (black line, 82°, Fig. 2A). Across neurons we found that the
262 target receptive field was indeed most often dorsal to the grating receptive field (grey, Fig.
263 2E), and that the median distance between the two was 77°. When the grating receptive field

264 was more dorsal, the two receptive field centers were significantly closer to each other (black
265 data, Fig. 2E, median distance 26°, $P < 0.0001$, Mann-Whitney test).

266 *The grating receptive field is sensitive to motion away from the midline*

267 We next determined the local motion sensitivity and average preferred direction of each
268 neuron's target and grating receptive field (colored arrows, Fig. S1G, H, S2). In the example
269 neuron, the preferred direction of the target receptive field is toward the right (blue arrows,
270 Fig. 2A, S1G), similar to the preferred direction of the grating receptive field (red arrows,
271 Fig. 2A, S1H). For comparison across neurons we color coded the preferred direction into
272 four cardinal directions, and plotted them as a function of receptive field center location. This
273 analysis shows that the preferred direction of the target receptive fields depended on location
274 (Fig. 3A). We found a significantly non-uniform distribution ($P < 0.01$, Rayleigh test), with a
275 median direction preference up and away from the visual midline (vector lengths 0.37 and
276 0.37, insets, Fig. 3C).

277 The directionality of the grating receptive fields depended more strongly on center location
278 (Fig. 3B, D). Indeed, we found a significantly non-uniform distribution ($P < 0.0001$, Rayleigh
279 test), with median direction preferences slightly up and away from the visual midline (vector
280 lengths 0.82 and 0.66, inset, Fig. 3D).

281 We next quantified the difference between the preferred directions of the target and the
282 grating receptive fields. In the example neuron, this is 8° (bottom right pictogram, Fig. 2A,
283 see also Fig. S1G, H). Across neurons the median direction difference was 75° if they were
284 on opposite sides of the visual midline, and 44° if they were on the same side, albeit with
285 neurons encompassing the entire span of possible directionality differences (Fig. 3E, see also
286 Fig. S2). However, there was no distribution difference based on whether the two receptive

287 field centers were on the same side (grey, Fig. 3E) or opposite sides (black, Fig. 3E) of the
288 visual midline (Mann-Whitney test, P = 0.15).

289 *Leading-edge sensitivity matches grating receptive field*

290 These descending neurons thus have two receptive fields, one that responds to small target
291 motion (blue, Fig. 2, S1, S2) and one that responds to local sinusoidal gratings (red, Fig. 2,
292 S1, S2). Which one of these receptive fields is most likely to contribute to their looming
293 sensitivity (right, Fig. 1A-D)? Looming sensitive neurons in *Drosophila,* including the giant
294 fiber, also respond strongly to high-contrast bars and edges (Ache et al., 2019a). We thus
295 used full screen OFF edges to map the looming receptive field (Fig. S3). An example neuron
296 shows strong responses to an OFF edge sweeping either left (Fig. S3A), down (Fig. S3C), or
297 up (Fig. S3D), across the visual field, but not right (Fig. S3B). The resulting leading-edge
298 receptive field (cyan, Fig. S3E, F) overlaps substantially with the grating receptive field (red,
299 Fig. S3F), but not the target receptive field (blue, Fig. S3F).

300 Across neurons we compared the location of the leading-edge, the target, and the grating
301 receptive field centers (circles, Fig. 4A, S3F). A qualitative analysis shows that the leading-
302 edge receptive field centers tend to cluster below the visual equator, just like the grating
303 receptive field centers do (cyan and red, Fig. 4A). For quantification, we calculated a
304 proximity index (Fig. 4B). When the leading-edge receptive field center is closer to the
305 grating receptive field center the proximity index is positive, up to a maximum of 100%. In
306 the example neuron, the proximity index is 83% (Fig. 4B, S3). Across neurons, we found that
307 the leading-edge receptive field centers were more frequently closer to the grating receptive
308 field centers (11 vs 4 neurons, Fig. 4C). In those neurons where the leading-edge receptive
309 field center was closer to the target receptive field center, the proximity index was lower
310 (medians of -30% and 48%, Fig. 4C) and the difference was significant (Mann-Whitney test,

311 P = 0.0015). In summary, it is likely that these neurons get their looming sensitivity
312 predominantly within the ventral visual field, overlapping with the location of the grating
313 receptive field.

314 *Different size response function in the two receptive fields*

315 Our previous work showed that looming sensitive neurons have a peculiar size response
316 function, with one peak to bars of a few degrees height, similar to the size tuning of target
317 selective neurons, and a second peak to full-screen bars (Nicholas et al., 2020). To investigate
318 if this size sensitivity differs between the two receptive fields we scanned bars of fixed width
319 across the visual monitor, while varying the height. We used two different trajectories, one
320 centered on the target receptive field (blue, Fig. 5A, S4), and one on the grating receptive
321 field (red, Fig. 5A, S4). We found that the neurons responded strongly to small bars moving
322 through the target receptive field (blue, Fig. 5A, Fig. S4A-E), but not through the grating
323 receptive field (red, Fig. 5A, Fig. S4A-E). For the middle-sized bars, the neurons responded
324 weakly whether they traversed the target or the grating receptive field (Fig. 5A, S4F-I).

325 When the bars were extended to cover a large part of the visual monitor, they traversed both
326 receptive fields (Fig. S4J, K), making it hard to determine which receptive field the strong
327 response came from (Fig. 5A). To bypass this, we scanned the bars vertically instead of
328 horizontally, so that they traversed the grating receptive field and the target receptive field at
329 different points in time (pictogram, Fig. 5B, S5). We found that the neurons responded
330 strongly when small bars moved through the target analysis window (blue, Fig. 5B, S5A-E),
331 and strongly to large bars when they moved through the grating analysis window (red, Fig.
332 5B, S5G-L). This shows that the target receptive field is tuned to small targets, whereas the
333 grating receptive field responds better to full-screen bars.

334 *Separate inputs to the two receptive fields*

335 The data above show that looming neurons have two receptive fields (Fig. 2-4), with size
336 tuning suggesting that they receive separate input (Fig. 5). What happens when they are
337 stimulated simultaneously? To investigate this, we first determined the response when the
338 two receptive fields were stimulated separately, by scanning a small target through the target
339 receptive field (blue, Fig. 6A), and a series of bars through the grating receptive field (red,
340 Fig. 6A, consistent with the data in Fig. 5). We compared this to the response to simultaneous
341 stimulation (black, Fig. 6B). We found that the response to simultaneous presentation (black,
342 Fig. 6B) was smaller than the linear sum of the two independent presentations (purple, Fig.
343 6B). Importantly, this cannot be due to response saturation, as the linear sum to the smallest
344 bars (purple, Fig. 6B) is on par with the measured response to the largest bars (black, Fig.
345 6B).

346 We next compared the response to simultaneous presentation (black, Fig. 6C) with the
347 strongest response for each neuron (green, Fig. 6C). We found that while there was a
348 significant dependence on bar size, there was no significant difference between the two
349 conditions (compare green and black, Fig. 6C, 2-way ANOVA), suggesting non-linear
350 interactions. Similarly, the locust LGMD (Krapp and Gabbiani, 2005) displays non-linear
351 interactions when stimuli are placed in different parts of the visual field, suggesting that the
352 details of its receptive fields may be worth investigating in the future.

353 *Target receptive field is based on 1-point correlator input, whereas the grating receptive*
354 *field uses 2-point correlator input*

355 The data above show two independent receptive fields. What is the likely pre-synaptic input
356 to each? Both optic lobe and descending target tuned neurons (Wiederman et al., 2013;

357 Nicholas and Nordström, 2021) generate their target selectivity using 1-point correlators,
358 which are based on the comparison of an OFF contrast change immediately followed by an
359 ON contrast change at a single point in space (Wiederman et al., 2008). These correlators are
360 fundamentally different from 2-point correlators, such as Hassenstein-Reichard elementary
361 motion detectors (Hassenstein and Reichardt, 1956), in their response to high-contrast edges.
362 For example, a 1-point correlator will respond only weakly to either OFF or ON contrast
363 edges, compared with complete objects, whereas 2-point correlators respond equally well to
364 single edges and complete objects (inset, Fig. 7, data replotted from Wiederman et al., 2013).

365 To investigate the potential input to the two receptive fields we first scanned an OFF edge
366 through the grating receptive field, then an ON edge, followed by a complete bar (red, Fig.
367 7). We found that the responses to single edges were similar to the response to a complete bar
368 (ns, red, Fig. 7). The response was thus consistent with an underlying 2-point correlator input
369 (compare red data with inset, Fig. 7). In contrast, when we scanned edges or targets through
370 the target receptive field, the response to a complete target was much stronger than to either
371 OFF or ON edges on their own (one-way ANOVA, followed by Tukey's multiple
372 comparisons test, $P < 0.0001$, blue, Fig. 7). Indeed, the response was consistent with an
373 underlying 1-point correlator input (compare blue data with inset, Fig. 7). In response to full-
374 screen edges and bars, which cover both receptive fields, we found the strongest response to
375 the OFF edge ($P = 0.0049$ for OFF vs bar, ns for ON vs bar, black, Fig. 7).

376 Discussion

377 We have shown here a group of descending neurons in the hoverfly *Eristalis tenax* that are
378 sensitive to both small moving targets and to looming stimuli (Fig. 1). We show that the
379 neurons have two discrete receptive fields, with different locations (Fig. 2, S1E, F, S2) and
380 preferred directions (Fig. 3, S1G, H, S2). We show that the looming sensitivity is likely

381 associated with the ventral receptive field (Fig. 4, S3). The size tuning (Fig. 5, S4, S5) and
382 sensitivity to OFF and ON contrast edges (Fig. 7) supports independent input to the two
383 receptive fields, using two fundamentally different pre-synaptic pathways. The input from the
384 two pathways is not linearly summed (Fig. 6B).

385 *Dual receptive fields*

386 The neurons that we describe here were classified as looming sensitive based on a strong
387 response to a looming stimulus (Fig. 1E, F) compared with an appearance control (Nicholas
388 et al., 2020). However, as they also respond strongly to small moving targets (left, Fig. 1A-D,
389 Fig. 5), they could have been classified as target selective descending neurons (TSDNs).
390 Indeed, the dragonfly TSDN DIT3 responds strongly to both small targets and to looming
391 stimuli (Gonzalez-Bellido et al., 2013). In the locust, LGMD/DCMD neurons respond to both
392 targets and to looming stimuli (Santer et al., 2012), and some central complex looming
393 sensitive neurons also respond to small moving targets (Rosner and Homberg, 2013).
394 Similarly, in *Drosophila,* some optic lobe and descending looming sensitive neurons also
395 respond to smaller objects (e.g. de Vries and Clandinin, 2012; Klapoetke et al., 2017; Namiki
396 et al., 2018; Ache et al., 2019a).

397 However, as opposed to these examples (e.g. Santer et al., 2012; Gonzalez-Bellido et al.,
398 2013; Rosner and Homberg, 2013; Ache et al., 2019a), we show that the dual sensitivity to
399 small targets and to larger objects is associated with two discrete receptive fields (Fig. 2 – 4,
400 S1 – S3). It is currently unknown if the dual sensitivity described in other insects (e.g. Santer
401 et al., 2012; Gonzalez-Bellido et al., 2013; Rosner and Homberg, 2013; Ache et al., 2019a)
402 also comes from different receptive fields. In *Drosophila* Foma-1 target sensitivity was
403 specific to the dorsal visual field, similar to our data (blue, Fig. 2B, C), while the visual field
404 location of the looming sensitivity was not specified (de Vries and Clandinin, 2012).

405 Our recordings were done extracellularly (Fig. 1), meaning that neurons with no spontaneous
406 activity are difficult to discover without presenting a suitable stimulus. We used a small
407 moving target to initially identify visual neurons (left, Fig. 1A-D), thus biasing our results
408 towards those looming sensitive descending neurons that also responded to small targets.
409 However, it is likely that there are looming neurons that do not respond to small objects, such
410 as found in e.g. *Drosophila* (e.g. Klapoetke et al., 2017; Ache et al., 2019b) and crabs (see
411 e.g. Cámera et al., 2020). Additionally, our visual monitor was placed in front of the animal,
412 thus biasing our results to neurons with frontal sensitivity. It is likely that there are additional
413 looming sensitive descending neurons with dorsal receptive fields, which could be useful for
414 e.g. detecting predators approaching from above, or lateral receptive fields, which could be
415 useful for avoiding imminent collision. For example, in the crab there are 16 retinotopically
416 arranged looming sensitive MLGs that underlie directional escape behaviors (Medan et al.,
417 2015). While each receptive field is small, together the 16 neurons cover 360° of the visual
418 field (Medan et al., 2015), and are thus able to encode directional escape responses.

419 A further technical limitation of our work was that we recorded from immobile animals that
420 were placed upside down in front of the monitor. In this situation there is no feedback from
421 the motor system, which could affect neural responses (see e.g. Fujiwara et al., 2017; Fenk et
422 al., 2021).

423 *Neuronal input mechanisms*

424 We showed that the looming sensitive descending neurons likely receive distinct input to the
425 two receptive fields (Fig. 5-7). Indeed, the dorsal target receptive field is likely to use pre-
426 synaptic 1-point correlators (blue, Fig. 7), just like the TSDNs do (Nicholas and Nordström,
427 2021). Furthermore, the size tuning of the dorsal target receptive field (blue, Fig. 5, S4-5) is
428 similar to the size tuning of TSDNs (Nicholas et al., 2018b; Nicholas and Nordström, 2021),

429 and of the presumably pre-synaptic STMDs (Nordström, 2012). This suggests that the dorsal
430 target receptive field could share input with the TSDNs.

431 In contrast, the ventral grating receptive field responded better to larger bars than to small
432 targets (red, Fig. 5, S4-5), similar to optic flow sensitive descending neurons (Nicholas and
433 Nordström, 2021). In addition, the ventral grating receptive field is likely to use pre-synaptic
434 2-point correlators of the EMD-type (red, Fig. 7), similar to optic flow sensitive neurons
435 (Harris et al., 1999). Interestingly, the looming sensitive LPLC2 neurons, which are pre-
436 synaptic to the *Drosophila* giant fiber (Ache et al., 2019b), get their input from T4/T5
437 (Klapoetke et al., 2017), which is consistent with a 2-point, EMD-type, correlator input (see
438 e.g. Salazar-Gatzimas et al., 2016). As our leading-edge data suggests that looming
439 sensitivity could be associated with the grating receptive field (Fig. 4, S3), this indicates that
440 looming sensitivity might be generated by 2-point correlation. Indeed, in the housefly,
441 escapes can be triggered by widefield gratings, even if not as efficiently as by looming
442 stimuli (Holmqvist and Srinivasan, 1991).

443 We found that the directionality of the grating receptive field depended strongly on the
444 azimuthal location of the receptive field center (Fig. 3B, D). However, the directionality of
445 the target receptive field was less dependent on its visual field location (Fig. 3A, C). In
446 addition, we found that the direction preference differences of the two receptive fields
447 covered the full 180° of possible direction differences (Fig. 3E), further supporting
448 independent inputs.

449 *Behavioral role*

450 Previous work has shown that the same stimulus displayed in different parts of the visual
451 field can elicit different behavioral output. For example, when crabs living in mudflats see a

small dummy moved at ground level they initiate prey pursuit behavior, but when the same dummy is moved above the crab, they try to escape it (Tomsic et al., 2017). In flying *Drosophila*, a looming stimulus in the lateral visual field leads to an escape response, whereas a looming stimulus in the frontal visual field leads to landing attempts (Tammero and Dickinson, 2002). While we did not stimulate the lateral visual field in our set-ups, the strong responses to frontal looming stimuli (right, Fig. 1A-D), likely associated with the ventral receptive field (Fig. 4), suggests that this could be used during landing behaviors on e.g. flowers. Indeed, bees adjust their body angle when landing so the landing surface ends up in the ventral visual field (Evangelista et al., 2010).

Alternatively, the neurons that we described here could potentially be used in pursuit. Indeed, when a hoverfly is pursuing a target, during most of the pursuit it will be projected as a small object on the pursuer's eye (Thyselius et al., 2023). When the hoverfly is below the target, having a dorsal target receptive field would be appropriate (blue, Fig. 2A-C). This could thus be supported by either the neurons described here (blue, Fig. 2), or by TSDNs without looming sensitivity (Nicholas et al., 2018b; Nicholas et al., 2020; Nicholas and Nordström, 2021).

During later stages of the pursuit, when the hoverfly gets closer to the target (Thyselius et al., 2023), this will be seen as a looming object. It has been suggested that this part of the pursuit cannot be subserved by classic target tuned neurons, but instead requires neurons that respond to larger objects and looming stimuli (see e.g. Discussion in Bagheri et al., 2015), like in zebrafish larvae (Henriques et al., 2019). Furthermore, during the final stages before capture, the pursuer would need to orient itself to grab the target with its legs. During this stage the target would be seen as a larger object in the ventral visual field, which would make the more ventral receptive field useful (red, Fig. 2).

However, the behavioral output required during initial target detection and final capture, for predator avoidance and landing, are all quite different. The descending neurons play an important role in sensorimotor transformation (Namiki et al., 2018), but it is difficult to see how the same descending neuron could control such different behaviors. Future work investigating where the neurons described here project to, and which behaviors they are thus likely to contribute to, will help elucidate this.

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606

607 # Figure Legends

608 ***Figure 1. Looming sensitive descending neurons respond robustly to looming stimuli and***
609 ***to small targets.*** **A)** Pictograms of the 3° × 3° square target moving horizontally (*Left*) and
610 the looming stimulus with an l/|v| of 10 ms and a final size of 118° (*Right*), as projected on
611 the frontal visual monitor. **B)** Raw data trace from an extracellular recording of a looming
612 sensitive descending neuron in response to a small target (*Left*) or a looming stimulus (*Right*).
613 **C)** The position of the target on the visual monitor (*Left*) and the diameter of the looming
614 stimulus (*Right*), time aligned with the data in panel B. **D)** Spike histograms of the responses
615 in panel B using 20 ms bins. **E)** Example response from a single neuron (mean ± SEM, n = 4)
616 to the looming stimulus and the appearance of a stationary black disc with a diameter of
617 118°. **F)** The peak amplitude of the response to a looming stimulus was significantly stronger
618 than the peak response to the appearance control ($p < 0.0001$, paired t-test).

619 ***Figure 2. Two different receptive fields.*** **A)** The location of the two receptive fields of an
620 example neuron as projected onto the visual monitor. The outlines show the 50% response
621 and the small circle the center of the target receptive field (blue) and the grating receptive
622 field (red). Euclidean distance between the receptive field centers (black line and value) and
623 the distance of each receptive field center to the equator and the visual midline (colored lines
624 and values) are indicated. Bottom right pictogram indicates the preferred direction of the
625 target (blue arrow) and the grating receptive field (red arrow), and the difference between the
626 two (black number). **B)** Location of target (blue) and grating receptive field centers (red)
627 across 98 neurons. **C)** Vertical distance between target (blue) and grating receptive field
628 centers (red) and the visual equator (10° bins). The target receptive field center locations
629 were significantly different from the grating receptive field center locations ($p < 0.0001$,
630 Mann-Whitney test). **D)** Horizontal distance from the visual midline (10° bins). There was no

631 significant difference between the target receptive field center distance to the midline, and the
632 grating receptive field distance (Mann-Whitney test). **E)** Euclidean distance between each
633 neuron's two receptive field centers (10° bins). Grey data come from neurons where the
634 target receptive field was dorsal to the grating receptive field center (N = 83) and black data
635 from neurons where the grating receptive field was dorsal to the target receptive field center
636 (N = 15). When the grating receptive field center was dorsal (black), the distance between the
637 two was significantly smaller ($p < 0.0001$, Mann-Whitney test).

638 ***Figure 3. The preferred direction depends on the receptive field center location.*** **A)** The
639 target receptive field centers, color coded according to their preferred direction (pictogram
640 bottom right). **B)** The grating receptive field centers, color coded according to their preferred
641 direction. **C)** Preferred direction of target receptive fields with centers in either the left or the
642 right visual field (30° bins). The distribution for target receptive fields in the left visual field
643 was significantly non-uniform ($p < 0.001$, Rayleigh test) with median preferred direction up
644 and to the left, and a median vector length of 0.37 (polar plot, scale 0 to 1). The distribution
645 for target receptive fields in the left visual field was significantly non-uniform ($p = 0.0025$,
646 Rayleigh test) with median preferred direction up and to the right, and a median vector length
647 of 0.37. **D)** Preferred direction of grating receptive fields centers in either the left or the right
648 visual field (30° bins). The distribution for grating receptive fields in the left visual field was
649 significantly non-uniform ($p < 0.0001$, Rayleigh test) with median preferred direction slightly
650 up and to the left, and a median vector length of 0.83. The distribution for grating receptive
651 fields in the right visual field was significantly non-uniform ($p < 0.0001$, Rayleigh test) with
652 median preferred direction slightly up and to the right, and a median vector length of 0.66. **E)**
653 Preferred direction difference between the target and grating receptive field of each neuron
654 (10° bins). Grey data show neurons where the two receptive field centers were on the same
655 side of the visual midline (N = 87), and black data show neurons with receptive fields on

656 opposite sides of the visual midline (N = 11). These were not significantly different (p = 0.15,
657 Mann-Whitney test).

658 ***Figure 4. The leading-edge receptive field is closer to the grating receptive field.*** **A)** The
659 location of the target (blue), grating (red) and leading-edge (cyan) receptive field centers in
660 15 neurons. **B)** The receptive field centers for one example neuron (*Left*), with distances
661 (black lines) between the leading-edge and the target receptive field center (*d1*), or the
662 grating receptive field center (*d2*), used to calculate the proximity index (*Right*). **C)** Leading
663 edge proximity index across neurons (N = 15). The leading-edge receptive field was closer to
664 the grating receptive field cent (red) in more neurons (N = 11) than to the target receptive
665 field (blue, N = 4). The distribution was significantly different from 0 (P < 0.01, one sample t
666 and Wilcoxon signed rank test), and the two distributions were different from each other (P =
667 0.0015, Mann-Whitney test)

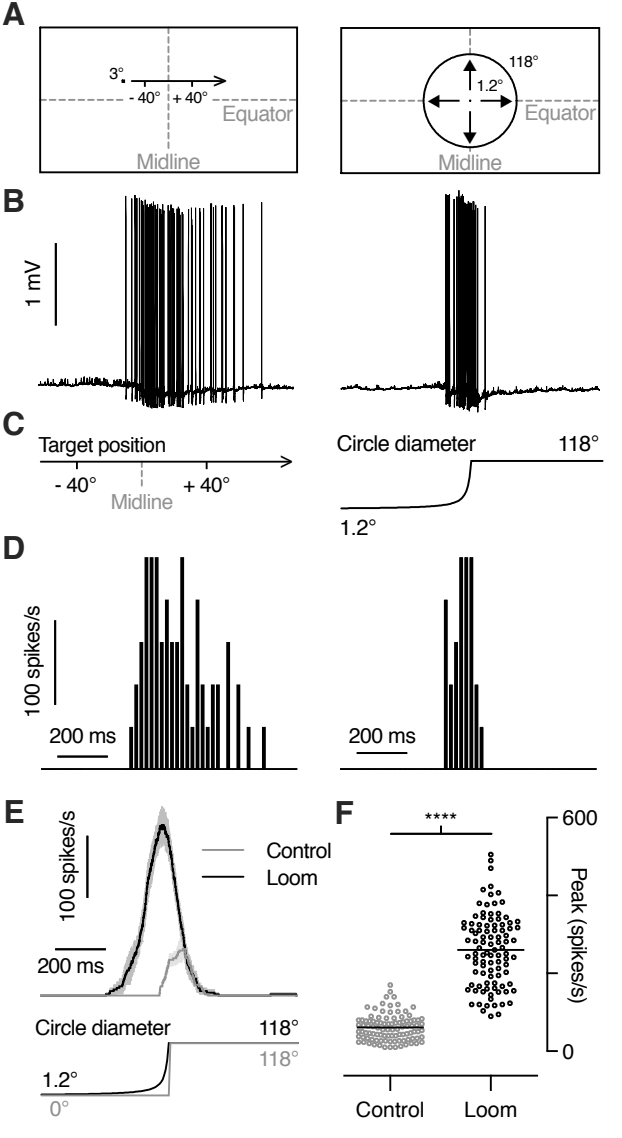
668 ***Figure 5. The two receptive fields have different size response functions.*** **A)** The pictograms
669 indicate the bar trajectory as it moved horizontally across the screen, subtending either the
670 target receptive field (*Left*, blue dashed line and arrow) or the grating receptive field (*Right*,
671 red dashed line and arrow, example bar height is 84°). Typical target and grating receptive
672 fields for an example neuron are shown. The grey shading shows the analysis window used to
673 calculate the mean response rate, which is the same for both trajectories. The graph shows
674 that responses to small bars are significantly stronger when passing through the target
675 receptive field (blue) compared to the grating receptive field (red, mean ± SEM, N = 8). **B)**
676 The pictogram indicates the analysis windows used to calculate the response to a bar of
677 varying width as it moved vertically along the screen (trajectory in black) subtending the
678 target receptive field (blue) or the grating receptive field (red). The graph shows that
679 responses to narrow bars are significantly stronger within the target analysis window (AW,

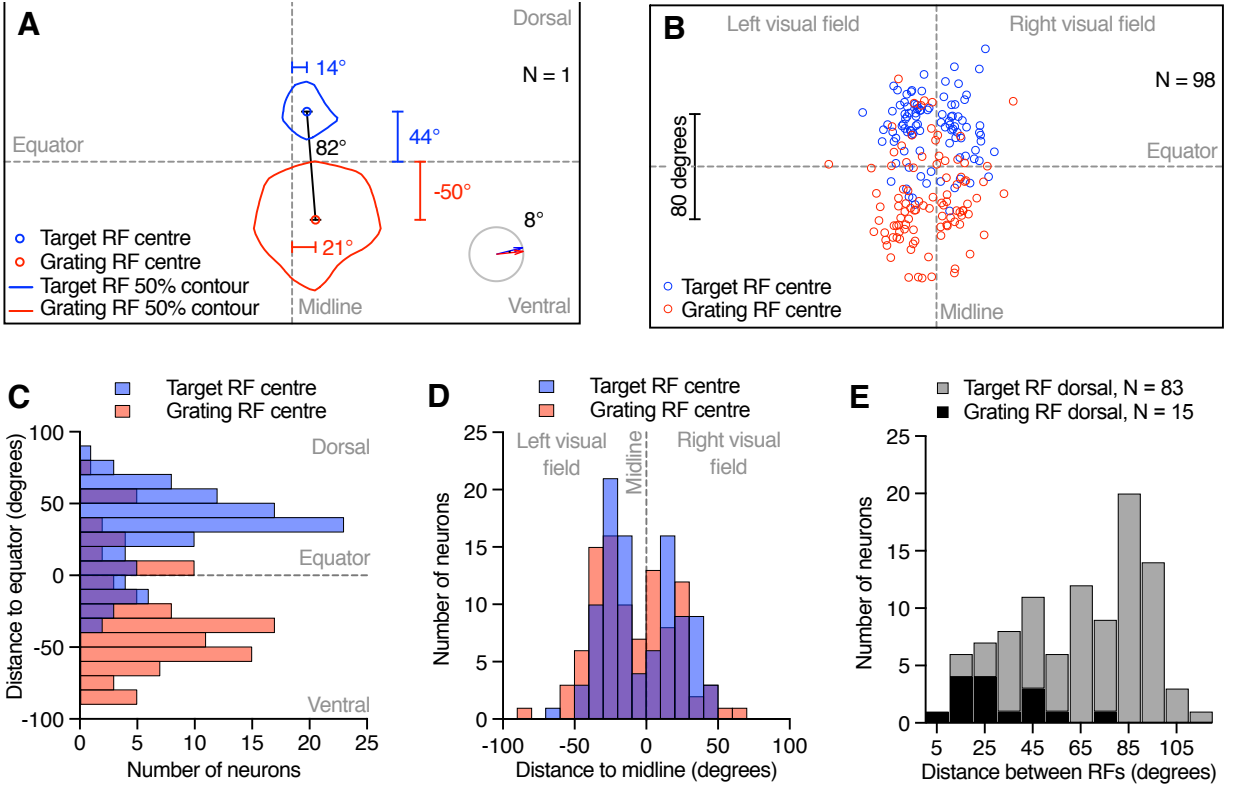
680 blue), while responses to wider bars are significantly stronger within the grating analysis
681 window (red, mean ± SEM, N = 10). Statistical test was a two-way ANOVA followed by
682 Sidak's multiple comparisons, with ****P < 0.0001, ***P < 0.001, **P < 0.01 and *P <
683 0.05.

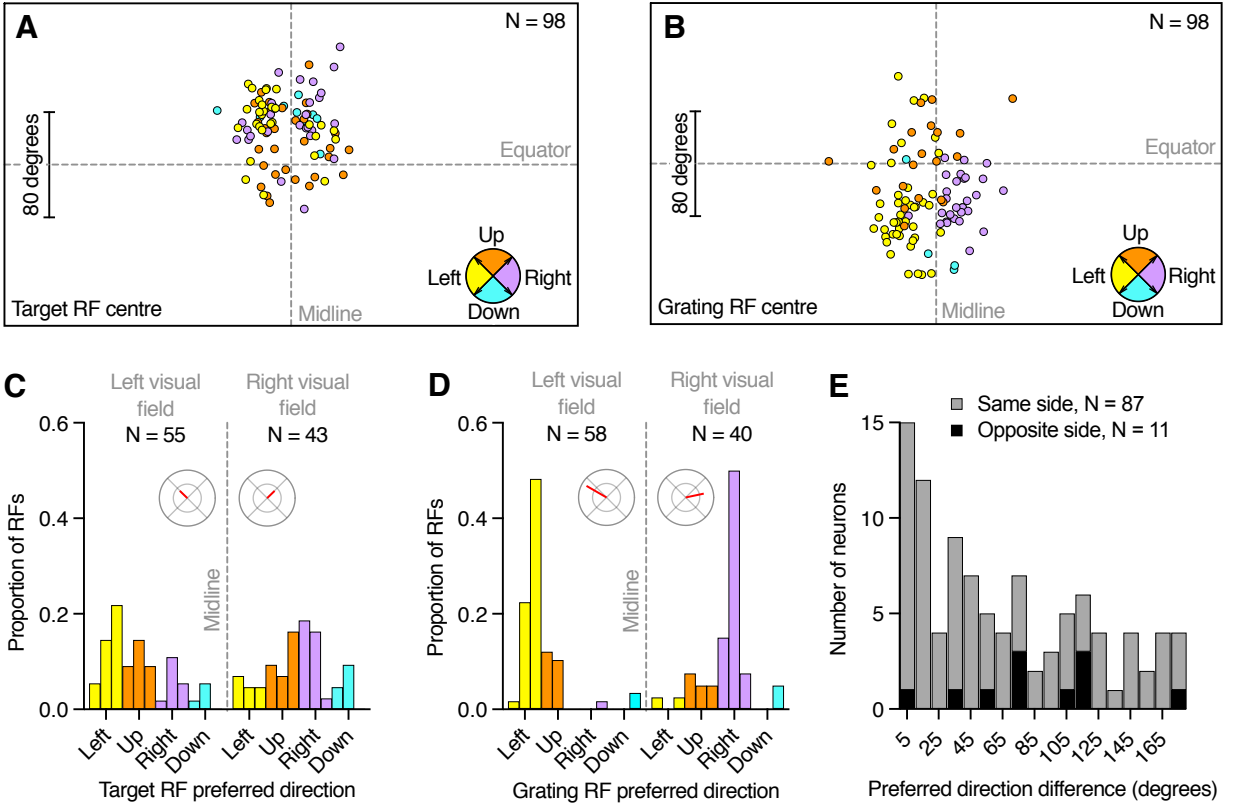
684 ***Figure 6. Response to simultaneous stimulation of the two receptive fields.*** **A)** Responses to
685 a small target traversing the target receptive field (blue), or bars of varying heights traversing
686 the grating receptive field (red). The pictograms at the top show the screen position of each
687 trajectory in relation to the receptive fields for an example neuron. Grey shading indicated the
688 analysis window used to calculate the mean response rate. **B)** Pictogram showing the screen
689 position of simultaneously presented target and bar traversing the target and grating receptive
690 fields. The graph shows that the responses to simultaneous presentation (black) are
691 significantly lower than the sum of the responses to the same stimuli presented on their own
692 (purple). **C)** The responses to simultaneous stimuli (black) are not significantly different from
693 the strongest response evoked by either the target or the bar on its own (green). For all panels
694 the data show mean ± SEM, for the same N = 5. Statistical analysis was done using two-way
695 ANOVA, with ****P < 0.0001, **P < 0.01 and ns indicating P > 0.05.

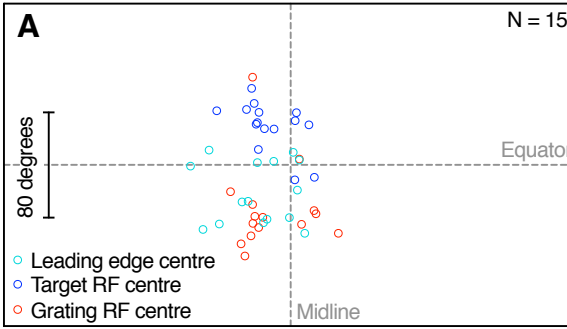
696 ***Figure 7. These looming neurons get input from both 1- and 2-point correlators.*** The
697 response to a leading OFF edge, a trailing ON edge, or a complete bar, all with a height of
698 84°, traversing the grating receptive field (red, N = 9), a height of 3° traversing the target
699 receptive field (blue, N = 10), or the full height of the screen (black, N = 7). The stimuli
700 moved horizontally at a velocity of 900 pixels/s. In all cases the response from each neuron
701 was normalized to the sum of the response to all three stimuli from the same trajectory (i.e.
702 OFF edge only, ON edge only, or complete bar). The inset shows the predicted response of a
703 motion detector that compares luminance changes over one point (also referred to as an

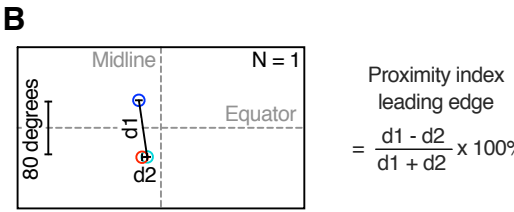
elementary STMD) or two points in space (often referred to as an EMD). The inset pictograms are replotted from Wiederman et al. (2013).

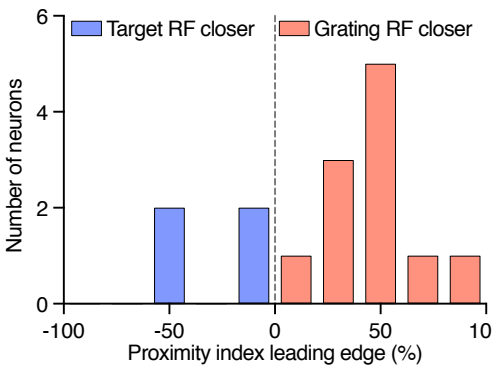

A
3°
- 40°
+ 40°
Equator
Midline
118°
1.2°
Equator
Midline
B
1 mV
C
Target position
- 40°
+ 40°
Midline
Circle diameter
118°
1.2°
D
100 spikes/s
200 ms
200 ms
E
100 spikes/s
200 ms
Control
Loom
Circle diameter
118°
118°
1.2°
0°
F
****
600
Peak (spikes/s)
0
Control
Loom

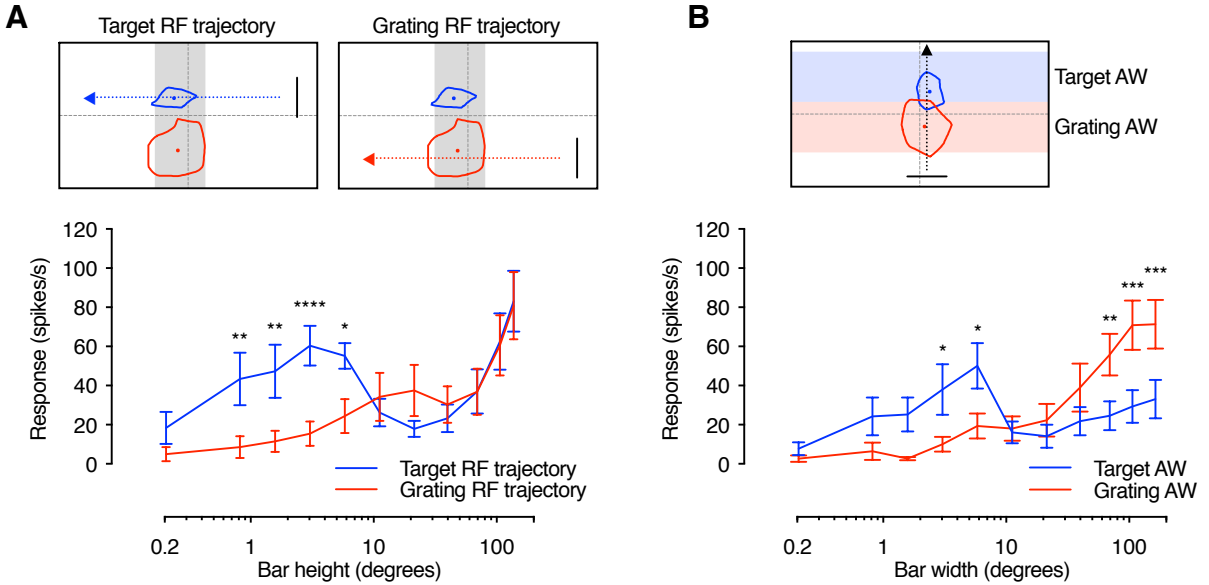

A
Dorsal
N = 1
14°
44°
82°
Equator
-50°
8°
21°
Target RF centre
Grating RF centre
Target RF 50% contour
Grating RF 50% contour
Midline
Ventral
B
Left visual field
Right visual field
N = 98
80 degrees
Equator
Target RF centre
Grating RF centre
Midline
C
Target RF centre
Grating RF centre
Dorsal
Equator
Ventral
Distance to equator (degrees)
Number of neurons
D
Target RF centre
Grating RF centre
Left visual field
Midline
Right visual field
Number of neurons
Distance to midline (degrees)
E
Target RF dorsal, N = 83
Grating RF dorsal, N = 15
Number of neurons
Distance between RFs (degrees)

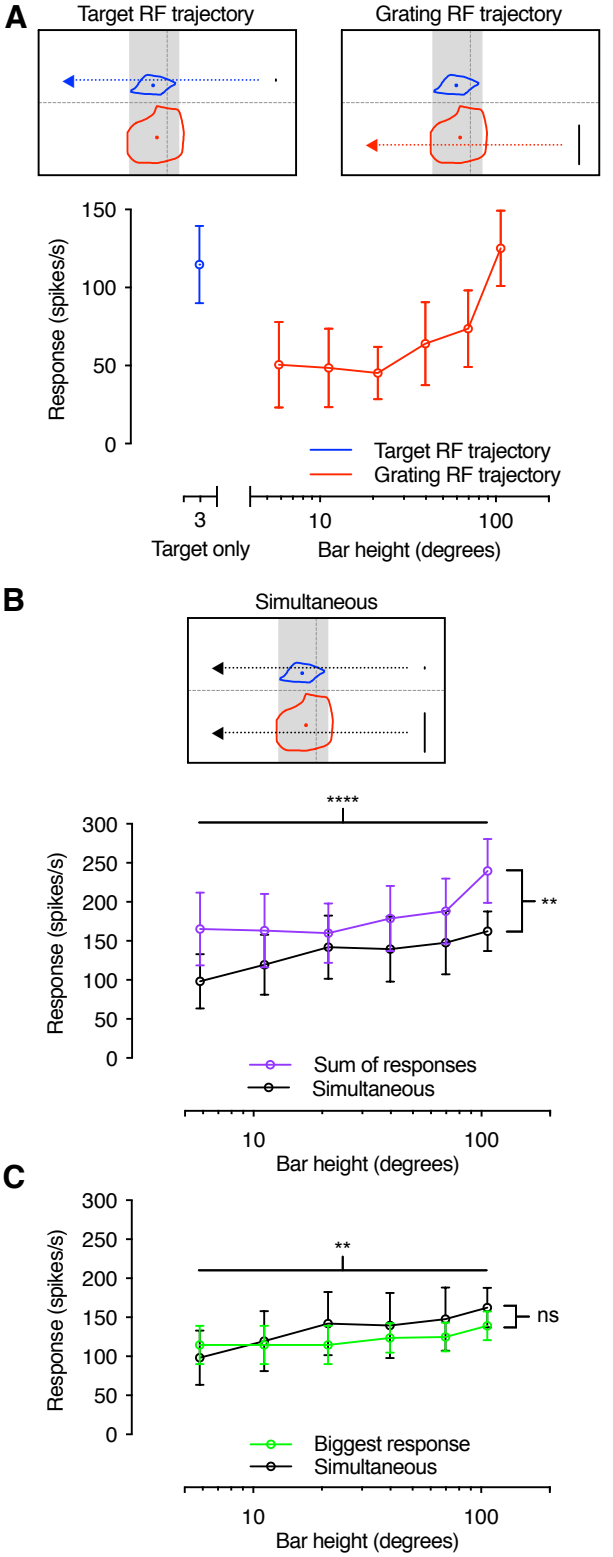

A
N = 98
80 degrees
Equator
Up
Left
Right
Down
Target RF centre
Midline
B
N = 98
80 degrees
Equator
Up
Left
Right
Down
Grating RF centre
Midline
C
Left visual field
N = 55
Right visual field
N = 43
Proportion of RFs
0.6
0.4
0.2
0.0
Midline
Left
Up
Right
Down
Left
Up
Right
Down
Target RF preferred direction
D
Left visual field
N = 58
Right visual field
N = 40
Proportion of RFs
0.6
0.4
0.2
0.0
Midline
Left
Up
Right
Down
Left
Up
Right
Down
Grating RF preferred direction
E
Same side, N = 87
Opposite side, N = 11
Number of neurons
15
10
5
0
5
25
45
65
85
105
125
145
165
Preferred direction difference (degrees)

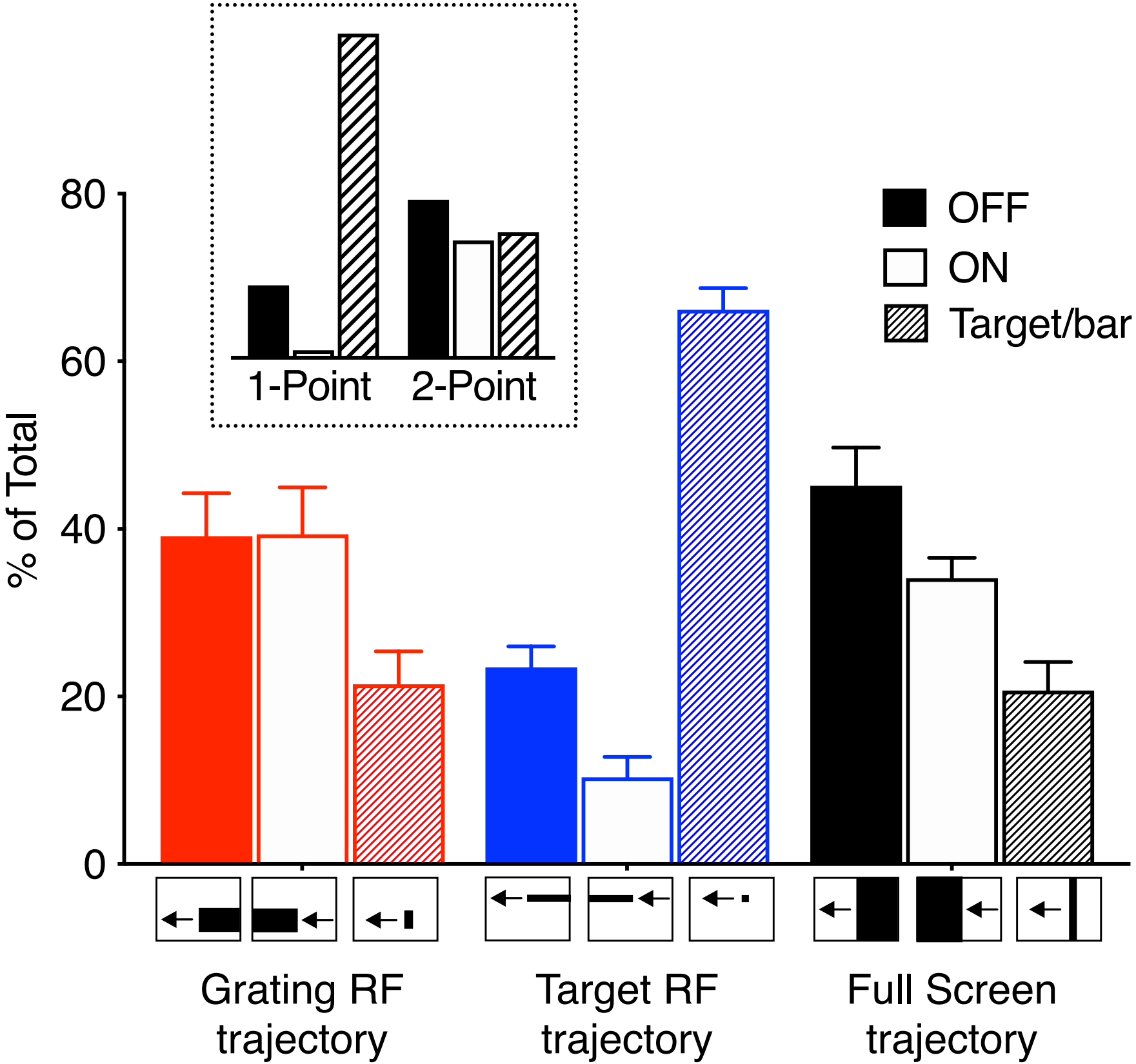

A
N = 15
80 degrees
Equator
Leading edge centre
Target RF centre
Grating RF centre
Midline


B
Midline
N = 1
80 degrees
Equator
d1
d2
Proximity index
leading edge
= (d1 - d2) / (d1 + d2) x 100%


C
Target RF closer
Grating RF closer
Number of neurons
0
2
4
6
-100
-50
0
50
100
Proximity index leading edge (%)


A
Target RF trajectory
Grating RF trajectory
Response (spikes/s)
120
100
80
60
40
20
0
**
**
****
*
Target RF trajectory
Grating RF trajectory
0.2
1
10
100
Bar height (degrees)
B
Target AW
Grating AW
Response (spikes/s)
120
100
80
60
40
20
0
*
*
**
***
***
Target AW
Grating AW
0.2
1
10
100
Bar width (degrees)


A
Target RF trajectory
Grating RF trajectory
Response (spikes/s)
150
100
50
0
Target RF trajectory
Grating RF trajectory
3
10
100
Target only
Bar height (degrees)
B
Simultaneous
Response (spikes/s)
300
250
200
150
100
50
0
****
**
Sum of responses
Simultaneous
10
100
Bar height (degrees)
C
Response (spikes/s)
300
250
200
150
100
50
0
**
ns
Biggest response
Simultaneous
10
100
Bar height (degrees)


80
60
40
20
0
% of Total
1-Point
2-Point
OFF
ON
Target/bar
Grating RF trajectory
Target RF trajectory
Full Screen trajectory