The evolution of the age of onset of resistance to infectious disease

Lydia J. Buckingham[1,2,*] and Ben Ashby[1,2,3]

1. Department of Mathematical Sciences, University of Bath, Bath, UK

2. Milner Centre for Evolution, University of Bath, Bath, UK

3. Department of Mathematics, Simon Fraser University, Burnaby, BC, Canada

*Corresponding author: ljb74@bath.ac.uk

Keywords: juvenile, adult, susceptibility, pathogen, parasite, resistance

ABSTRACT

Many organisms experience an increase in disease resistance as they age but the time of life

at which this change occurs varies. Increases in resistance are partially due to prior exposure

and physiological constraints but these cannot fully explain the observed patterns of age-

related resistance. An alternative explanation is that developing resistance at an earlier age

incurs costs to other life-history traits. Here, we explore how trade-offs with host

reproduction or mortality affect the evolution of the onset of resistance, depending on when

during the host's life-cycle the costs are paid (only when resistance is developing, only when

resistant or throughout the lifetime). We find that the timing of the costs is crucial to

determining evolutionary outcomes, often making the difference between resistance

developing at an early or late age. Accurate modelling of biological systems therefore relies

on knowing not only the shape of trade-offs but also when they take effect. We also find that

the evolution of the rate of onset of resistance can result in evolutionary branching. This

provides an alternative, possible evolutionary history of populations which are dimorphic in

disease resistance, where the rate of onset of resistance has diversified rather than the level

of resistance.

INTRODUCTION

Many organisms experience changes in their level of resistance to infectious disease as they age, with repercussions for both their own health and transmission to others (Altizer et al. 2004; Apolloni et al. 2013; Clark et al. 2017). Age-related differences in resistance have been recorded across many taxa, including plants (Miller 1983; Panter and Jones 2002; Develey-Rivière and Galiana 2007; Bruns et al. 2017), invertebrates (Sait et al. 1994; Kubi et al. 2006; Armitage and Boomsma 2010; Garbutt et al. 2014; Green et al. 2016; Klemme et al. 2022) and vertebrates (Duca 1948; Zuckerman and Yoeli 1954; Francis 1961), including humans (Baird 1998; Kurtis et al. 2001; Glynn and Moss 2020). In particular, adults are generally more resistant than juveniles (Duca 1948; Zuckerman and Yoeli 1954; Francis 1961; Sait et al. 1994; Baird 1998; Kurtis et al. 2001; Panter and Jones 2002; Kubi et al. 2006; Develey-Rivière and Galiana 2007; Armitage and Boomsma 2010; Green et al. 2016; Bruns et al. 2017) although there is variation in precisely when these changes in resistance occur. This variation in the timing of resistance onset has received little empirical or theoretical attention, with very few studies seeking to explain or even describe the time within a host's lifespan at which resistance changes. A key outstanding question is therefore, 'how should evolution shape the timing of the onset of resistance?'

As there is variation in the rate of onset of resistance within species and resistance traits are often heritable, it is reasonable to assume that the rate of onset of resistance is an evolvable trait. In this paper, we seek to generate predictions for how the rate of onset of resistance might evolve. Intuitively, an earlier onset of resistance will tend to lower disease prevalence and hence reduce the risk of infection, thereby acting as a negative feedback on selection if

resistance is costly. However, precisely how this phenomenon affects the evolution of the

onset of resistance remains to be determined.

Several factors are known to contribute towards the onset of disease resistance. Many

species' immune systems respond to prior exposure, meaning that resistance may increase

substantially following infection. This mechanism can explain some, but not all, observed

variation in the rate of onset of disease resistance, particularly in the case of species which

rely solely or predominantly on innate resistance. For instance, different species of plants

experience changes in resistance at different life stages (Develey-Rivière and Galiana 2007).

Alternatively, variation in the rate of onset of resistance may have evolved due to contrasting

selection pressures at different life-stages. For instance, if a pathogen is more harmful to older

hosts than younger hosts then the onset of resistance may be delayed if developing or

maintaining resistance is costly. In some cases, however, age-related changes in resistance

have been observed towards pathogens which have similar effects on hosts of all ages (Bruns

et al. 2017), suggesting that the timing of the onset of resistance is also affected by other

factors. Hence, none of these mechanisms can provide a full explanation for variation in the

rate of onset of disease resistance.

An alternative evolutionary explanation for increases in resistance with age is that heightened

resistance incurs costs to other life-history traits. For instance, hosts which invest in

developing resistance quickly may allocate resources away from growth or development of

other traits. In principle, such costs may be paid only while resistance is developing but may

also have long-term consequences which affect the reproduction or survival of the host later

in life (Chaplin and Mann 1978; Simons 1979; Tian et al. 2003). Evolutionary trade-offs may

therefore provide an explanation as to why hosts might develop resistance later in life rather

than being resistant from birth or from a very early age. Empirical studies reveal trade-offs

between reproduction (Biere and Antonovics 1996; Karasov et al. 2017) or growth (Biere and

Antonovics 1996; Susi and Laine 2015; Karasov et al. 2017; Bartlett et al. 2018) and disease

resistance in plants and insects, but little data is available on age-specific costs (Izhar and Ben-

Ami 2015).

Epidemiological models have considered disease spread in age-structured populations (e.g.

(Clark et al. 2017)) and evolutionary models have considered the evolution of resistance in

populations with no age structure (e.g. (Antonovics and Thrall 1994; Boots and Haraguchi

1999; Carlsson-Graner and Thrall 2006; Miller et al. 2007; Donnelly et al. 2015)) but, in

contrast, the evolution of age-structured resistance has received relatively little attention.

However, recent theory has explored the evolution of age-specific resistance in juveniles and

adults, showing that differences in resistance between life stages are most pronounced for

particularly long- or short-lived hosts (Ashby and Bruns 2018) and that adult resistance often

evolves to exceed juvenile resistance, depending on which traits trade off with resistance

(Buckingham et al. 2022). Here, we investigate a different characteristic of age-specific

resistance, namely the timing of the onset of resistance. We focus our analysis on trade-offs

with mortality and reproduction, which may affect the host during development of resistance,

during the resistant stage or throughout the lifetime of the host. We also consider variation

in pathogen traits, specifically transmissibility and the strength and type of virulence. We find

that the timing of the trade-offs (costs paid before or after the onset of resistance or

throughout the host's lifetime) has a significant effect on evolutionary outcomes, often

determining whether the host becomes resistant very early or very late in life. We also find

that evolutionary branching of the rate of onset of resistance can generate a dimorphic

population where some hosts are resistant throughout their lives and others never develop

resistance.

METHODS

MODEL DESCRIPTION

We consider a well-mixed, asexual host population in which hosts are born fully susceptible

to an infectious disease but develop full innate resistance to infection at a constant (but

evolvable) rate $\zeta > 0$ (see figure 1A for a model schematic and table 1 for a full list of

parameters and variables). Let S, I and R be the densities of susceptible, infected and

resistant hosts respectively, giving a total host population density of $N = S + I + R$. Hosts

reproduce at a maximum rate $a > 0$, subject to density-dependent competition given by $q >$

0, and die naturally at rate $b > 0$. Pathogen transmission is assumed to be density-

dependent, with transmission rate β, and infected hosts may either experience sterility

virulence equal to $1 - f$, where $0 \leq f \leq 1$ is the reduction in fecundity when infected, or

mortality virulence given by $\alpha > 0$, the relative increase in the mortality rate. We assume that

hosts do not recover from infection, as recovery would complicate the timing of the onset of

resistance (it is not clear how infected individuals would transition to the innately resistant

stage in this case) and would also require us to account for potential long-term effects of

sterility virulence.

In a monomorphic population and in the absence of any costs of resistance, the population

dynamics are described by the following set of ordinary differential equations:

$$\frac{dS}{dt} = a(1-qN)(S+fI+R) - \beta SI - bS - \zeta S \tag{1a}$$

$$\frac{dI}{dt} = \beta SI - b(1+\alpha)I \tag{1b}$$

$$\frac{dR}{dt} = -bR + \zeta S \tag{1c}$$

Intuitively, in the absence of any costs of resistance, selection will favour ever larger ζ so that

there is an immediate onset of resistance at the start of the host lifespan, which will in turn

drive the pathogen extinct. We therefore consider cases where being resistant or developing

resistance incurs a cost to the host, either through higher mortality or lower fecundity (e.g.,

due to resource allocation). Such costs might be paid only when the host is resistant (i.e., in

the R class), or prior to the onset of resistance (i.e., during development in classes S and I),

or throughout the lifetime of the host. The magnitude of any costs may therefore be constant,

or depend on the rate of onset of resistance, ζ. To account for costs of resistance, we modify

equation (1) so that the reproduction rate, $a(\zeta)$, and mortality rate, $b(\zeta)$, prior to the onset

of resistance may depend on its rate of onset, and to allow these rates to differ for resistant

individuals, $a_R(\zeta)$ and $b_R(\zeta)$:

$$\frac{dS}{dt} = a(\zeta)(1-qN)(S+fI) + a_R(\zeta)(1-qN)R - \beta SI - b(\zeta)S - \zeta S \tag{2a}$$

$$\frac{dI}{dt} = \beta SI - b(\zeta)(1+\alpha)I \tag{2b}$$

$$\frac{dR}{dt} = -b_R(\zeta)R + \zeta S \tag{2c}$$

132   We consider six possible scenarios:

133   (1) A constant fecundity cost, paid only when the host is resistant ($a_R(\zeta) = ah$, where h

134   is the relative reduction in fecundity when resistant, with $b_R(\zeta) = b$, $a = a_0$ and $b =$

135   b_0)

(2) A constant mortality cost, paid only when the host is resistant ($b_R(\zeta) = b(1 + \delta)$,

where δ is the relative increase in mortality when resistant, with $a_R(\zeta) = a$, $a = a_0$

and $b = b_0$)

(3) A whole-life fecundity cost ($a_R(\zeta) = a(\zeta)$ as defined in equation (3a), with $b_R(\zeta) = b$

and $b = b_0$)

(4) A whole-life mortality cost ($b_R(\zeta) = b(\zeta)$ as defined in equation (3b), with $a_R(\zeta) = a$

and $a = a_0$)

(5) A developmental fecundity cost ($a(\zeta)$ as defined in equation (3a), with $a_R(\zeta) =$

$a_0, b_R(\zeta) = b$ and $b = b_0$)

(6) A developmental mortality cost ($b(\zeta)$ as defined in equation (3b), with $a_R(\zeta) = a$,

$b_R(\zeta) = b_0$ and $a = a_0$).

where a_0 and b_0 are baseline reproduction and mortality rates and we use the following

trade-offs for the reproduction rate:

$$a(\zeta) = a_0 \left(1 - \frac{c_1^a \left(1 - e^{-c_2^a \zeta} \right)}{1 - e^{-c_2^a}} \right) \tag{3a}$$

150 and the mortality rate:

$$b(\zeta) = b_0 \left(1 + \frac{c_1^b \left(1 - e^{-c_2^b \zeta} \right)}{1 - e^{-c_2^b}} \right) \tag{3b}$$

151 where $c_1^i > 0$ determines the maximum strength of the trade-off (i.e. the maximum

152 proportional reduction or increase in the associated life-history trait) and c_2^i determines the

153 shape of the trade-off (larger absolute values correspond to greater deviations from linearity;

154 figure 1B & figure 1C). Note that it is possible to rescale the system of equations (2a) to (2c)

155 so that we can set $q = 1$ and $b_0 = 1$ without loss of generality (see *Supplementary Materials*).

EPIDEMIOLOGICAL DYNAMICS

158   The disease-free equilibrium of system (2) is given by:

$$S^* = \frac{b_R(ab_R + a_R\zeta - bb_R - b_R\zeta)}{q(b_R + \zeta)(ab_R + a_R\zeta)} \qquad (4a)$$

$$R^* = \frac{\zeta(ab_R + a_R\zeta - bb_R - b_R\zeta)}{q(b_R + \zeta)(ab_R + a_R\zeta)} \qquad (4b)$$

159   and is stable provided that $ab_R + a_R\zeta > bb_R + b_R\zeta$ and

$$R_0 = \frac{\beta b_R(ab_R + a_R\zeta - bb_R - b_R\zeta)}{q(b_R + \zeta)(ab_R + a_R\zeta)} - b(1 + \alpha) + 1 < 1 \qquad (5)$$

160   where R_0 is the basic reproductive ratio of the pathogen (see *Supplementary Materials* for

161   derivation). The pathogen can spread when $R_0 > 1$, in which case there is a stable, endemic

162   (non-trivial) equilibrium (see *Supplementary Materials*).

EVOLUTIONARY INVASION ANALYSIS

We use evolutionary invasion analysis (adaptive dynamics) to determine the evolutionary

dynamics of the rate of onset of resistance, ζ (Geritz et al. 1998). Specifically, we assume that

mutations are sufficiently rare that there is a separation of ecological and evolutionary

timescales (the ecological dynamics of the resident population reach equilibrium before the

next mutation occurs) and are sufficiently small that $\zeta_m \approx \zeta$, where ζ_m is the mutant trait and

ζ is the resident trait. The invasion dynamics of rare host mutants are analogous to equation

2 and so are given in the *Supplementary Materials*. Using the next generation method

(Hurford et al. 2010), one can derive the following expression for the invasion fitness:

$$w(\zeta_m, \zeta) = \frac{a(\zeta_m)(1 - qN^*)b(\zeta_m)(1 + \alpha)b_R(\zeta_m) + a(\zeta_m)(1 - qN^*)f\beta I^* b_R(\zeta_m) + a_R(\zeta_m)(1 - qN^*)b(\zeta_m)(1 + \alpha)\zeta_m}{(\beta I^* + b(\zeta_m) + \zeta_m)b(\zeta_m)(1 + \alpha)b_R(\zeta_m)} - 1 \quad (6)$$

where asterisks denote the endemic equilibrium of the resident population and a, a_R, b and

b_R may be functions of ζ_m, depending on the nature of the costs of resistance. In the general

case, the lengthy expression for the endemic equilibrium limits analytical progress. However,

in cases (1) and (2), where the costs of resistance are constant, we can proceed to determine

an expression for the singular value of the rate of onset of resistance.

SIMULATIONS

To simulate an evolutionary trajectory of the evolving trait, ζ, we first choose a resident value

for ζ and an initial composition for the host population. We then run the ecological dynamics

for a fixed length of time, T. At this stage, we introduce a small, second sub-population with

a different value of ζ (chosen at random to be either slightly higher or lower than the resident

value) to model the introduction of a rare mutant into the population. The ecological

dynamics are then run again for this new population for a fixed length of time, T. If the size

of the sub-population with a particular value of ζ falls below a threshold, then that sub-

population is removed (it has gone extinct). An additional, small sub-population with a new

value of ζ (chosen at random to be slightly higher or lower than one of the existing trait values

in the population) is introduced at each evolutionary timestep, the ecological dynamics are

run and sub-populations that fall below the extinction threshold are removed. This process is

repeated for many evolutionary timesteps to simulate the evolution of ζ.

Parameter/variable	Description	Default value or range
a, b	Reproduction rate/natural mortality rate of non-resistant hosts	n/a
a_R, b_R	Reproduction rate/natural mortality rate of resistant hosts	n/a
a_0	Baseline value of the reproduction rate	5
b_0	Baseline value of the natural mortality rate	0.1
c_1^a, c_1^b	Strength of reproduction/mortality trade-offs	0.1
c_2^a, c_2^b	Shape of reproduction/mortality trade-offs	± 2
$1 - f$	Sterility virulence	$0 \leq f \leq 1$
ζ	Rate of onset of resistance	$0 \leq \zeta$
S, I, R	Density of susceptible/infected/resistant hosts	n/a
N	Total host population density	n/a
q	Strength of host density dependence	1
t	Time, measured in arbitrary units	n/a
α	Mortality virulence (proportional increase in mortality rate)	$0 \leq \alpha$
β	Pathogen transmission rate	0.5
$1 - h$	Direct fecundity cost of resistance	$0 \leq h \leq 1$
δ	Direct mortality cost of resistance (proportional increase)	$0 \leq \delta$

192   Table 1: Model parameters and variables.

Figure 1: (A) Model schematic for a monomorphic population. (B) Examples of reproduction trade-off functions (with $a_0 = 5$). (C) Examples of mortality trade-off functions (with $b_0 = 1$). Trade-off strength is controlled by the parameter c_1^i; a relatively strong trade-off ($c_1^i = 0.5$, red) results in a much larger reduction in the birth rate for a given level of adult resistance than a relatively weak trade-off does ($c_1^i = 0.25$, blue). Trade-off shape is controlled by the parameter c_2^i; a positive value ($c_2^i = 2$, solid) means that the costs of resistance decelerate (increasing returns) as the rate of onset gets faster whereas a negative value ($c_2^i = -1$, dashed) leads to accelerating costs (diminishing returns).

RESULTS

CONSTANT COSTS

We begin by considering the effects of constant fecundity and mortality costs (cases 1 and 2

described in the methods) on the evolution of the onset of resistance, ζ. This means that we

have a constant fecundity cost when resistant $a_R = ah$, where the reproduction rate when

susceptible is $a = a_0$, and a constant mortality cost when resistant, $b_R = b(1 + \delta)$, where

the mortality rate when susceptible is $b = b_0$. In this case, we can differentiate the invasion

fitness with respect to the mutant trait ζ_m to calculate the fitness gradient (see

Supplementary Materials) and then find the root of the fitness gradient to give us the singular

strategy for the rate of onset of resistance:

$$\zeta^* = b_0(1 + \delta)\left(\frac{p - 1}{fp - (1 + \alpha)} + \frac{\beta(a_0 - b_0 p)}{a_0 q\, b_0(1 + \alpha)} - 1\right) \qquad (7)$$

where $p = \frac{1+\delta}{h}$ for notational convenience. Note that this expression can take a negative

value, in which case the singular strategy is effectively at $\zeta^* = 0$ (no onset of resistance).

We can see from this expression that the rate of onset of resistance rises linearly as the

pathogen transmissibility increases. This is because a more transmissible parasite will have a

higher disease prevalence and so will exert stronger selection for fast onset of resistance. The

singular strategy also rises as sterility virulence increases because more virulent pathogens

impose stronger selection for resistance (figure 2D). This pattern is also seen as mortality

virulence rises but only as long as mortality virulence is not too high. When mortality virulence

is high, infected hosts die quickly, reducing the infectious period and so the potential for

disease transmission. This reduces the pathogen density and so weakens selection for fast

onset of resistance, leading to a fall in the rate of onset of resistance (figure 2B).

We might also expect that the rate of onset of resistance would fall as the costs of being

resistant rise. This is indeed true when the costs of fast onset of resistance are fecundity costs

or when they are relatively large mortality costs (figure 2A, C). However, when low mortality

costs are paid, it is possible for the rate of onset to rise as the costs of being resistant increase

(figure 2A). This is because raising the mortality costs causes a significant increase in the

density of the pathogen; resistant individuals die faster and so there are fewer of them in the

population, yet reproduction is still high enough to maintain the population near to its

density-dependent limit, meaning that the density of non-resistant individuals rises. This

leads to a population with a reduced proportion of resistant individuals and an increased

proportion of infected individuals. This heightened pathogen density then imposes greater

selection for fast onset of resistance.

Analytical calculations reveal the invasion fitness of any mutant in a resident population at

the singular strategy is zero: $w(\zeta_m, \zeta^*) = 0$ (see *Supplementary Materials*). Hence

$\frac{\partial^2 w}{\partial \zeta_m^2}|_{\zeta_m = \zeta = \zeta^*} = 0$ always and so the singular strategy is not strictly evolutionarily stable. The

expression which determines convergence stability $\left(\frac{\partial^2 w}{\partial \zeta_m \partial \zeta}|_{\zeta_m = \zeta = \zeta^*}\right)$ is intractable and so it is

not possible to determine conditions for convergence stability analytically. However,

extensive numerical calculations suggest that the singular strategy is always convergence

stable (*see Supplementary* Materials). In order to classify the stability of the singular strategy

fully however, we also need to determine whether it appears to be evolutionarily stable or

unstable in practice. To do this, we use simulations. Evolutionary simulations (see

Supplementary Materials for description) reveal that the singular strategies behave like

continuously stable strategies (CSS's), for a wide range of parameter sets. Figure 3 depicts

graphically the simulated changes in the rate of onset of resistance over evolutionary time,

showing how it tends towards the analytically determined singular strategy and then remains

there.

Figure 2: Evolution of the rate of onset of resistance, ζ^*, in the case of constant costs of

resistance. Parameters used are as in table 1 except for $\beta = 1$, with (A) $1 - f = 0, 1 - h = 0$

and $\alpha = 1$, (B) $1 - f = 0, 1 - h = 0$ and $\delta = 0.25$, (C) $1 - f = 0.5, \alpha = 0$ and $\delta = 0$ and (D)

$1 - h = 0.25, \alpha = 0$ and $\delta = 0$.

Figure 3: Simulated trajectories of the evolution of the rate of onset of resistance over time.

The red, dashed lines show the analytically determined values of the singular strategies.

Parameters used are as in table 1 except for $\beta = 1$, with (A) $1 - f = 0, \alpha = 1, 1 - h = 0$ and

$\delta = 0.25$ in the case of scenario (2) where the host pays a constant mortality cost when

resistant and with (B) $1 - f = 0.5, \alpha = 0, 1 - h = 0.25$ and $\delta = 0$ in the case of scenario (1)

where the host pays a constant fecundity cost when resistant.

  NON-CONSTANT COSTS

We now consider non-constant costs for the onset of resistance (cases 3-6 described in the

methods). When costs to reproduction or mortality accelerate ($c_2^a, c_2^b < 0$), the rate of onset

of resistance always evolves to a single, continuously stable strategy (CSS). When the costs

decelerate, there are more possible evolutionary outcomes: a single CSS, repeller or

branching point, a repeller above a branching point, a branching point above a repeller or two

repellers with a branching point in between (parameter values for which these occur are

shown in figure S1 and figure S2). When the costs of having a faster onset of resistance are

paid throughout the host's lifetime (cases 3 & 4), we can show analytically that the singular

strategy (or the uppermost singular strategy if there is more than one) is always convergence

stable (see *Supplementary Materials*). When the costs of having a faster onset of resistance

are only paid before the onset of resistance (cases 5 & 6), this singular strategy is never

convergence stable (see *Supplementary Materials*). This means that, if the rate of onset of

resistance is initially sufficiently high, then it will increase indefinitely, in effect causing

newborn hosts to become resistant immediately. This is because the host is already paying a

significant cost for a reasonably fast onset of resistance. It is more beneficial for hosts to pay

a slightly higher cost for a shorter time (before it becomes resistant and stops paying the cost)

than to pay a slightly lower cost for a longer time.

Many factors influence whether the onset of resistance is fast or slow. However, certain

parameters have especially strong effects. We can see these effects most clearly if we

consider accelerating costs ($c_2^a, c_2^b < 0$), as each set of parameters produces a single

evolutionary endpoint which does not depend on the initial conditions. We find that the

virulence of the pathogen plays a particularly important role, with the fastest onset of

resistance evolving when the pathogen exerts weak mortality virulence and strong sterility

virulence (see figure 4; note that contours show the proportion of the host population which

is resistant to the pathogen). This is because strong mortality virulence reduces disease

prevalence by reducing the average duration of infection and so weakens selection for

resistance. Sterility virulence, in contrast, does not affect the average duration of infection

and so does not reduce disease prevalence but does greatly reduce host fitness. The fastest

onset of resistance therefore evolves when disease prevalence is high (low mortality

virulence) and when there is strong sterility virulence. Conversely, the rate of onset of

resistance tends to zero when both sterility and mortality virulence are negligible (because

the cost of infection is minimal) or when mortality virulence is very high (because disease

prevalence is so low that the host is unlikely to be exposed to the pathogen). The results

described above are qualitatively similar for decelerating trade-offs when the costs of

resistance are paid throughout the lifetime of the hosts and there is a single CSS.

Figure 4: Heatmaps showing the evolved rate of onset of resistance as mortality and sterility

virulence vary and for qualitatively different trade-offs. Contours show the proportion of the

host population that is resistant to the disease and so do not correspond to heatmap colours.

The rate of onset of resistance is highest (red) when mortality virulence is low and sterility

virulence is high (bottom right of each panel). Increasing sterility virulence (left to right within

each panel) always causes the rate of onset of resistance to increase. Increasing mortality

virulence (bottom to top within each panel) may also cause the rate of onset of resistance to

rise and then fall for sufficiently low values of sterility virulence. These patterns are broadly

consistent across all six trade-off scenarios (A-F). Parameters used are as described in table 1,

except $c_2^a = c_2^b = -2$ (accelerating costs), $h = 0.75$ and $\delta = 0.25$, where applicable.

In general, greater pathogen transmissibility leads to the evolution of faster onset of

resistance. However, as pathogen transmissibility gets very high, the rate of onset of

resistance begins to fall (figure 5). This is because higher transmissibility corresponds to a

lower average age on infection, and so it is more costly for a host to invest in a rapid onset of

resistance (which will have to be very fast to take effect before the host becomes infected by

a highly transmissible pathogen) than it is for the host to become infected with the pathogen

(having not paid the costs of fast onset of resistance). This is consistent for both fecundity and

mortality trade-offs (figure 5) of a variety of different strengths (values of c_1^a and c_1^b). As

expected, greater costs (higher values of c_1^a and c_1^b) lead to the evolution of slower onset of

resistance. Fecundity and mortality costs with the same proportional effects (e.g., a 50%

increase in mortality or a 50% decrease in reproduction) have similar quantitative impacts on

the rate of onset of resistance (figure 5).

Figure 5: The effect of disease transmissibility (β) on the rate of onset of resistance, when

either a fecundity cost (blue curves) or a mortality cost (red curves) is paid throughout the

lifetime of the host (scenarios 3 and 4). The qualitative effects of transmissibility are the same

whether the costs of fast onset of resistance are low (c_1^a or $c_1^b = 0.3$; solid) or high (c_1^a or

$c_1^b = 0.8$; dotted). Parameters used are as in table 1 with $c_2^a = -2$, $c_2^b = -2$, $f = 0.5$ and

$\alpha = 0$.

 DIVERSIFICATION

The host population may diversify through evolutionary branching when the costs are

decelerating ($c_2^a, c_2^b > 0$; see figure 6). Evolutionary branching leads to one sub-population

with a very high rate of onset of resistance (ζ large) that increases indefinitely (see

Supplementary Materials), until these individuals effectively develop resistance immediately

after birth, and another sub-population with a very low rate of onset of resistance ($\zeta \approx 0$),

with these individuals unlikely ever to become resistant given the typical lifespan of the host.

When costs are paid throughout the lifetime of the host, these costs must be strongly

decelerating ($c_2^i \gg 0$) for branching to occur (figure 7A). When costs are paid before the onset

of resistance, branching only occurs for weakly decelerating costs (c_2^i small but positive). The

latter branching points always sit below repellers and so the population will only branch if the

initial rate of onset of resistance is sufficiently low (otherwise, the rate of onset of resistance

will increase indefinitely; figure 7B).

When costs of having a faster onset of resistance are paid throughout the lifetime of the host,

branching is most commonly observed when the baseline reproduction rate (a_0), disease

transmissibility (β) and sterility virulence ($1 - f$) are high, baseline mortality (b_0) and the

strength of trade-offs (c_1^a or c_1^b) are low and mortality virulence (α) is intermediate (figure

S1). When costs of fast onset of resistance are paid only before the onset of resistance,

branching is most common when the baseline reproduction rate (a_0), disease transmissibility

(β), mortality virulence (α), baseline mortality rate (b_0) and strength of the mortality trade-

off (c_1^b) take intermediate values, when sterility virulence ($1 - f$) is high and when the

strength of the reproduction trade-off (c_1^a) is low (figure S2).

Figure 6: Simulated trajectories of the evolution of the rate of onset of resistance over time.

(A) When costs of fast onset of resistance are paid throughout the lifetime of the host, the

lower branch often tends towards a low, non-zero value whereas the upper branch goes to

infinity. (B) When costs are paid only before the onset of resistance, the lower branch goes to

zero whereas the upper branch goes to infinity. Parameters used are as in table 1 with $1 -$

$f = 0.5$ and $\alpha = 0$. We also use (a) $c_2^a = 5$ and (b) $c_2^a = 1$ to generate branching points.

Figure 7: The effect of the shape of trade-offs on the incidence of branching points for

different timings of costs. Continuously stable strategies are shown in green (solid), repellers

are shown in orange (dotted) and branching points are shown in purple (dashed). (A) When

costs of fast onset of resistance are paid throughout the host's lifetime, costs must be strongly

decelerating (high c_2^a) for branching to occur. (B) When costs are paid only before the onset

of resistance, they must be weakly decelerating for branching to occur. Parameters used are

as in table 1 with $1 - f = 0$ and $\alpha = 1$.

DISCUSSION

Recent theory has shown how trade-offs and life-history traits combine to shape the evolution of age-specific resistance (Ashby and Bruns 2018; Buckingham et al. 2022). Here, we have shown how the average age at which hosts become resistant $(1/\zeta)$ evolves, depending on how and when costs of resistance are paid (costs only paid before or after the onset of resistance, or throughout the host's lifetime). Our key findings are that (1) the rate of onset of resistance peaks for low mortality virulence, high sterility virulence and intermediate pathogen transmissibility; (2) when costs of resistance are paid throughout the lifetime of the host (or only after the onset of resistance) the onset of resistance typically evolves to a stable, intermediate rate whereas when costs are paid only before the onset of resistance, hosts may evolve no onset of resistance, extremely fast onset of resistance or an immediate rate of onset of resistance (depending on parameters and initial conditions); and (3) pathogens may drive host diversification in the rate of onset of resistance.

Hosts evolve the fastest onset of resistance when pathogens cause high sterility virulence, low mortality virulence and intermediate transmissibility. Such pathogens induce the highest selection for resistance because they are able to maintain a significant density within the host population but also cause a large fitness cost to their hosts. Given a fixed, low-to-intermediate level of sterility virulence, an intermediate level of mortality virulence leads to the fastest rate of onset of resistance, but we have found that the combination of high sterility virulence and low mortality virulence leads to the fastest rate of onset overall in our model. This could be tested by determining the level of sterility or mortality inflicted by pathogens against which hosts experience a particularly early onset of resistance. We tentatively hypothesise that such

pathogens would be more likely to inflict high levels of sterility virulence than high levels of

mortality virulence.

The life-stage at which trade-offs occur has a significant effect on the evolution of the rate of

onset of resistance. It is well established that the magnitude and shape of trade-offs, as well

as the life-history traits involved, are critical to the evolution of various traits, including

resistance (Kisdi 2006; Ashby and Bruns 2018; Buckingham et al. 2022). We have shown that

the life-stage at which trade-offs act also drastically alters the evolutionary outcome for the

rate of onset of resistance. An intermediate rate of onset of resistance is most likely to evolve

when costs are paid throughout the host's lifetime whereas very fast or very slow rates of

onset, or dimorphism, are more likely if the costs are paid predominantly during the

development of resistance. Very little empirical work has considered age-specific costs of

resistance. However, we tentatively predict that organisms with very fast, very slow or

polymorphic onset of resistance are most likely to experience costs which only act early in

life.

When the rate of onset of resistance diversifies, it results in a fully (or almost fully) susceptible

subpopulation coexisting with a fully resistant subpopulation. Such dimorphic populations,

where some individuals are effectively resistant throughout their lives and others are always

(or almost always) susceptible, are often seen in real, biological systems (Parker 1988; Jarosz

and Burdon 1990; Wayne et al. 1996; Turkington et al. 2019). This evolutionary endpoint is

very similar to classical models of host resistance evolution which result in dimorphic host

populations with resistant and susceptible phenotypes (Antonovics and Thrall 1994; Boots

and Haraguchi 1999). However, the mechanism for arriving at this end point is very different

in our model. We have shown that this evolutionary endpoint can also arise through evolution

of the rate of onset of resistance, where all hosts may transition from being susceptible to

resistant as they age, rather than resistance/susceptibility being fixed before birth. This

suggests that there may be an alternative evolutionary history of populations which are

dimorphic in disease resistance, with the rate of onset of resistance diverging rather than the

level of resistance. If this is indeed the case in some species, then one would expect

"susceptible" hosts typically to have resistance genes that are suppressed rather than absent.

However, it is possible that once the rate of onset of resistance approaches zero, resistance

genes are lost rather than remaining suppressed. Experimental evolution may offer another

path to testing this result by always selecting for hosts that have an earlier or later onset of

resistance, thus validating whether a similar evolutionary endpoint can be achieved through

the evolution of the timing of onset of resistance.

This paper constitutes an initial investigation into the theoretical consequences of the

evolution of the age of onset of disease resistance. As such, we have considered a simple

model which allows us to determine some possible outcomes of this evolutionary process.

There are many additions and adjustments which could make this a more realistic model for

specific biological systems. For example, we have only considered the case of a disease with

no recovery, as recovery would require differential handling of infection-acquired immunity

or individuals who experience the onset of resistance while infected. While this is a

reasonable assumption for some host-pathogen interactions where recovery is negligible,

such as systemic infections in plants, invertebrates and bacteria (Ebert 2008; Abatángelo et

al. 2017; Bruns et al. 2017), a more general development of the theory here would require

careful consideration of how recovery affects immunity and post-infection sequelae.

Recovery could be incorporated into our model in several different ways, depending on the

mechanisms underlying recovery and the onset of resistance. For example, one could allow

infected individuals to return to the susceptible class, but then the rate of onset of resistance

takes on a different interpretation, as the process of becoming resistant is effectively turned

off and on again as the host becomes infected and then later recovers. Alternatively, one

could allow infected individuals to move into the resistant class if the onset of resistance

occurred while they were infected, but would this process cause the host to clear the infection

or just become resistant to re-infection? These scenarios cover rather different biological

scenarios and so deserve careful consideration in future work.

For simplicity, we assumed that hosts experience a rapid onset of disease resistance (i.e.

transitioning from susceptible to resistant in a short space of time). Rapid changes from low

to high levels of resistance have been observed to occur in a number of biological systems.

For example, Kurtis *et al.* showed that malaria (*Plasmodium falciparum*) resistance increases

significantly in boys during puberty (between ages 15 and 20) which is relatively quick in

relation to their lifespan (Kurtis et al. 2001). Critchlow *et al.* found that flour beetles (*Tribolium*

castaneum) express more antimicrobial proteins as pupae than as larvae, resulting in a

sudden increase in resistance during metamorphosis (Critchlow et al. 2019). Bruns *et al.*

showed that alpine carnations (*Dianthus pavonius*) are far more resistant to anther-smut

disease (*Microbotryum*) as adults than during the seedling stage, suggesting a rapid onset of

resistance at maturation (Bruns et al. 2017). Farber & Mundt found that wheat plants

(*Triticum aestivum*) significantly increase their resistance to stripe rust (*Puccinia striiformis*)

between the ages of 3 and 5 weeks (with an average lifespan of 20 to 25 weeks) (Farber and

Mundt 2017). As none of these changes in resistance occur instantaneously, however, our

model could be extended to include not just completely susceptible and completely resistant

stages, but also an additional stage in which resistance is intermediate, to represent the

period during which these organisms are developing resistance. In cases where this stage is

very short, our model should provide a good approximation.

Our model also assumes that hosts are born completely susceptible and later in life become

fully resistant. Although large increases in the level of disease resistance may occur during a

host's lifetime, in many cases the host may shift between quantitative levels of resistance

rather than qualitative resistance (Kurtis et al. 2001; Bruns et al. 2017; Farber and Mundt

2017; Critchlow et al. 2019). In future, our model should be extended to explore the timing

of shifts between different levels of partial resistance. There are also empirical examples of

reductions in resistance with host age and so the timing of the loss of resistance could be

considered. For instance, Klemme *et al.* found that Atlantic salmon (*Salmo salar*) experience

a reduction in resistance to a trematode (*Diplostomum pseudospathaceum*) when they

mature from resident to migrant life stages (Klemme et al. 2022).

Overall, we have shown how the rate of onset of resistance evolves in response to variation

in the nature of associated trade-offs and disease characteristics, potentially leading to host

diversification and the same evolutionary endpoint as classical models of resistance

evolution. Our model therefore provides an alternative explanation for the origin of

dimorphism in host resistance.

DATA AVAILABILITY

Source code is available at github.com/ecoevotheory/Buckingham_and_Ashby_2022b

REFERENCES

Abatángelo, V., N. Peressutti Bacci, C. A. Boncompain, A. A. Amadio, S. Carrasco, C. A. Suárez,

and H. R. Morbidoni. 2017. Broad-range lytic bacteriophages that kill Staphylococcus aureus

local field strains. PLoS ONE 12.

Altizer, S., A. K. Davis, K. C. Cook, and J. J. Cherry. 2004. Age, sex, and season affect the risk of

mycoplasmal conjunctivitis in a southeastern house finch population. Canadian Journal of

Zoology 82:755–763.

Antonovics, J., and P. H. Thrall. 1994. The cost of resistance and the maintenance of genetic

polymorphism in host-pathogen systems. Proceedings of the Royal Society B 257:105–110.

Apolloni, A., C. Poletto, and V. Colizza. 2013. Age-specific contacts and travel patterns in the

spatial spread of 2009 H1N1 influenza pandemic. BMC Infectious Diseases 13.

Armitage, S. A. O., and J. J. Boomsma. 2010. The effects of age and social interactions on

innate immunity in a leaf-cutting ant. Journal of Insect Physiology 56:780–787.

Ashby, B., and E. Bruns. 2018. The evolution of juvenile susceptibility to infectious disease.

Proceedings of the Royal Society B 285.

Baird, J. K. 1998. Age-dependent characteristics of protection v. susceptibility to Plasmodium

falciparum. Annals of Tropical Medicine and Parasitology 92:367–390.

Bartlett, L. J., L. Wilfert, and M. Boots. 2018. A genotypic trade-off between constitutive

resistance to viral infection and host growth rate. Evolution 72:2749–2757.

Biere, A., and J. Antonovics. 1996. Sex-specific costs of resistance to the fungal pathogen

ustilago violacea (microbotryum violaceum) in silene alba. Evolution 50:1098–1110.

Boots, M., and Y. Haraguchi. 1999. The evolution of costly resistance in host-parasite systems.

American Naturalist 153.

Bruns, E. L., J. Antonovics, V. Carasso, and M. Hood. 2017. Transmission and temporal

dynamics of anther-smut disease (Microbotryum) on alpine carnation (Dianthus pavonius).

Journal of Ecology 105:1413–1424.

Buckingham, L. J., E. L. Bruns, and B. Ashby. 2022. The evolution of age-specific resistance to

infectious disease. bioRxiv.

Carlsson-Graner, U., and P. H. Thrall. 2006. The impact of host longevity on disease

transmission: host-pathogen dynamics and the evolution of resistance. Evolutionary Ecology

Research 8:659–675.

Chaplin, J. F., and T. J. Mann. 1978. Evaluation of tobacco mosaic resistance factor transferred

from burley to flue-cured tobacco. Journal of Heredity 69:175–178.

Clark, J., J. S. Garbutt, L. McNally, and T. J. Little. 2017. Disease spread in age structured

populations with maternal age effects. Ecology Letters 20:445–451.

Critchlow, J. T., A. Norris, and A. T. Tate. 2019. The legacy of larval infection on immunological

dynamics over metamorphosis. Philosophical Transactions of the Royal Society B 374.

Develey-Rivière, M. P., and E. Galiana. 2007. Resistance to pathogens and host developmental

stage: A multifaceted relationship within the plant kingdom. New Phytologist 175:405–416.

Donnelly, R., A. White, and M. Boots. 2015. The epidemiological feedbacks critical to the

evolution of host immunity. Journal of Evolutionary Biology 28:2042–2053.

Duca, C. J. 1948. Age specific susceptibility to tuberculosis; experiments on guinea pigs and

rats. American Review of Tuberculosis 57:389–399.

Ebert, D. 2008. Host-parasite coevolution: Insights from the Daphnia-parasite model system.

Current Opinion in Microbiology 11:290–301.

Farber, D. H., and C. C. Mundt. 2017. Effect of plant age and leaf position on susceptibility to

wheat stripe rust. Phytopathology 107:412–417.

Francis, J. 1961. The effect of age on the susceptibility of guinea pigs to tuberculosis. Tubercle

42:333–336.

Garbutt, J. S., A. J. P. O'Donoghue, S. J. McTaggart, P. J. Wilson, and T. J. Little. 2014. The

development of pathogen resistance in Daphnia magna: Implications for disease spread in

age-structured populations. Journal of Experimental Biology 217:3929–3934.

Geritz, S. A. H., E. Â. Kisdi, G. Mesze, Â. Na, and J. A. J. Metz. 1998. Evolutionarily singular

strategies and the adaptive growth and branching of the evolutionary tree. Evolutionary

Ecology 12:35–57.

Glynn, J. R., and P. A. H. Moss. 2020. Systematic analysis of infectious disease outcomes by

age shows lowest severity in school-age children. Scientific Data 7.

Green, T. J., A. Vergnes, C. Montagnani, and J. de Lorgeril. 2016. Distinct immune responses

of juvenile and adult oysters (Crassostrea gigas) to viral and bacterial infections. Veterinary

Research 47.

Hurford, A., D. Cownden, and T. Day. 2010. Next-generation tools for evolutionary invasion

analyses. Journal of the Royal Society Interface 7:561–571.

Izhar, R., and F. Ben-Ami. 2015. Host age modulates parasite infectivity, virulence and

reproduction. Journal of Animal Ecology 84:1018–1028.

Jarosz, A. M., and J. J. Burdon. 1990. Predominance of a single major gene for resistance to

Phakopsora pachyrhizi in a population of Glycine argyrea. Heredity 64:347–353.

Karasov, T. L., E. Chae, J. J. Herman, and J. Bergelson. 2017. Mechanisms to mitigate the trade-

off between growth and defense. Plant Cell 29:666–680.

Kisdi, É. 2006. Trade-off geometries and the adaptive dynamics of two co-evolving species.

Evolutionary Ecology Research 8:959–973.

Klemme, I., P. V. Debes, C. R. Primmer, L. S. Härkönen, J. Erkinaro, P. Hyvärinen, and A.

Karvonen. 2022. Host developmental stage effects on parasite resistance and tolerance.

American Naturalist.

Kubi, C., J. van den Abbeele, R. de Deken, T. Marcotty, P. Dorny, and P. van den Bossche. 2006.

The effect of starvation on the susceptibility of teneral and non-teneral tsetse flies to

trypanosome infection. Medical and Veterinary Entomology 20:388–392.

Kurtis, J. D., R. Mtalib, F. K. Onyango, and P. E. Duffy. 2001. Human resistance to Plasmodium

falciparum increases during puberty and is predicted by dehydroepiandrosterone sulfate

levels. Infection and Immunity 69:123–128.

Miller, M. E. 1983. Relationships between onion leaf age and susceptibility to Alternaria porri.

Plant Disease 67.

Miller, M. R., A. White, and M. Boots. 2007. Host life span and the evolution of resistance

characteristics. Evolution 61:2–14.

Panter, S. N., and D. A. Jones. 2002. Age-related resistance to plant pathogens. Advances in

Botanical Research 38:251–280.

Parker, M. A. 1988. Polymorphism for disease resistance in the annual legume Amphicarpaea

bracteata. Heredity 60:27–31.

Sait, S. M., M. Begon, and D. J. Thompson. 1994. The influence of larval age on the response

of Plodia interpunctella to a Granulosis virus. Journal of Invertebrate Pathology 63:107–110.

Simons, M. D. 1979. Influence of genes for resistance to Puccinia coronata from Avena sterilis

on yield and rust reaction of cultivated oats. Phytopathology 69:450–452.

Susi, H., and A. L. Laine. 2015. The effectiveness and costs of pathogen resistance strategies

in a perennial plant. Journal of Ecology 103:303–315.

Tian, D., M. B. Traw, J. Q. Chen, M. Kreitman, and J. Bergelson. 2003. Fitness costs of R-gene-

mediated resistance in Arabidopsis thaliana. Nature 423:74–77.

Turkington, C. J. R., A. Morozov, M. R. J. Clokie, and C. D. Bayliss. 2019. Phage-resistant phase-

variant sub-populations mediate herd immunity against bacteriophage invasion of bacterial

meta-populations. Frontiers in Microbiology 10.

Wayne, M. L., D. Contamine, and M. Kreitman. 1996. Molecular population genetics of ref(2)P,

a locus which confers viral resistance in Drosophila. Molecular Biology and Evolution 13:191–

199.

Zuckerman, A., and M. Yoeli. 1954. Age and sex as factors influencing Plasmodium berghei

infections in intact and splenectomized rats. Journal of Infectious Diseases 94:225–236.

590