# Longitudinal map of transcriptome changes in the Lyme pathogen *Borrelia burgdorferi* during tick-borne transmission

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**ABSTRACT**

*Borrelia burgdorferi* (*Bb*), the causative agent of Lyme disease, must adapt to vastly different environments as the bacterium cycles between the tick vector and a vertebrate host. During a bloodmeal, *Bb* migrates from the tick midgut to the salivary glands and changes its gene expression, priming *Bb* for vertebrate infection. These tick-specific transmission events are dependent on key transcriptional regulators; however, the full range of transcriptional changes that occur over several days inside of the tick are technically challenging to capture. We developed an experimental approach to enrich *Bb* cells from *Ixodes* ticks during a transmitting bloodmeal to longitudinally define their global transcriptomic landscape. We identified 192 genes that substantially change expression over the course of the tick bloodmeal, most of which were located on plasmids of the *Bb* genome. The majority of upregulated genes encode proteins found at the cell envelope or proteins of unknown function, including 45 outer surface lipoproteins embedded in the unusual protein-rich coat of *Bb*. As these proteins may facilitate *Bb* interactions with host tissues or immune systems, we also utilized mass spectrometry to identify candidate tick proteins that physically associate with *Bb*. The *ex vivo Bb* transcriptomes and candidate tick interacting proteins presented here provide an important roadmap for investigating key determinants of *Bb* priming and transmission during the tick stage of its unique transmission cycle.

## INTRODUCTION

Vector-borne microbial pathogens are transmitted by the bite of arthropods and have evolved sophisticated ways to adapt to vastly different environments as they move between vector and host. Uncovering their adaptive mechanisms can not only open avenues for disrupting pathogen transmission but also provide fundamental insights into vector physiology and microbial symbioses (Shaw and Catteruccia, 2018). Lyme disease, the most reported vector-borne disease in North America, is caused by the bacterial pathogen *Borrelia burgdorferi* (*Bb*) (Rosenberg et al., 2018; Steere et al., 2016). Its primary vector, the blacklegged tick *Ixodes scapularis,* acquires and transmits *Bb* through two separate multi-day bloodmeals, one in which *Bb* is acquired from an infected vertebrate host and a second, during the subsequent life stage, in which *Bb* is transmitted to a new host (Tilly et al., 2008). During the transmission bloodmeal, *Bb* proliferate in the tick midgut before a subset of these cells disseminate to the salivary glands (Dunham-Ems et al., 2009). *Bb* is deposited into the new host via the tick saliva extruded into the bite site (Spielman et al., 1987). Given the prolonged nature of *I. scapularis* feeding, the high specificity of vector-pathogen relationships, and the complicated array of events needed for successful *Bb* transmission, the tick bloodmeal provides an opportune intervention point for preventing pathogen spread. However, we do not currently have a clear understanding of the molecular mechanisms involved in this process.

*Bb* must adapt to dramatically different environments as it cycles from tick to vertebrate host, and understanding the genes involved in this process will enable the identification of key interactions to target to prevent transmission. When an infected tick feeds, *Bb* responds to bloodmeal-induced environmental changes and undergoes cellular modifications driven by key transcriptional circuits, including the RpoN-RpoS sigma factor cascade and the Hk1/Rrp1 two-component system (Radolf et al., 2012). *In vitro* analyses of *Bb* cells cultured in tick- or mammalian-like growth conditions have pointed to additional genetic determinants of tick-borne transmission by revealing widespread transcriptome remodeling during host switching (reviewed in Samuels et al., 2021). However, it is not fully clear how these *in vitro* expression changes correspond to complex *in vivo* changes over the course of a transmission bloodmeal, which comprises several physiologically distinct stages for the tick vector. Capturing comprehensive, longitudinal data on *Bb* expression from inside its vector has been hampered by technical challenges given the dynamic nature of the bloodmeal and the general low abundance of bacterial cells relative to the tick (Samuels et al., 2021). Although microarray approaches have been used to identify large scale changes in *Bb* gene expression between the first and second tick bloodmeals (Iyer et al., 2015), we have limited

temporal resolution into the molecular transitions that happen across key steps of tick feeding. This problem necessitates novel approaches to capture the transcriptomic changes of *Bb* within the natural tick environment.

While the full landscape of *Bb* transmission determinants is not yet known, we do have a growing knowledge of functional processes that are critical during the tick stage, such as motility, metabolism, and immune evasion (Kurokawa et al., 2020; Phelan et al., 2019). These functions often rely on the unique protein-rich *Bb* outer surface. Notably, several specific tick–*Bb* protein–protein interactions are important for *Bb* survival, migration, or transmission to the next host. *Bb* encodes an extensive outer surface protein (Osp) family with members that are differentially expressed during host switching. One of these proteins, OspA, binds a tick cell surface protein, tick receptor for OspA (TROSPA), which is required for successful *Bb* colonization of the tick midgut during the first acquisition bloodmeal (Pal et al., 2004). Several other proteins have also been linked to *Bb* migration within the tick (Pal et al., 2022). For example, BBE31 binds a tick protein TRE31, and disruption of this interaction decreases the number of *Bb* cells that successfully migrate from the tick gut to salivary glands (Zhang et al., 2011). However, these interactions alone are not sufficient to block *Bb* growth or migration in ticks, suggesting there are likely additional molecular factors from *Bb* and ticks at play during tick-borne transmission.

To provide a more comprehensive set of *Bb* determinants driving tick-borne transmission of this important human pathogen, we longitudinally mapped *Bb* genome-wide expression changes for bacterial cells isolated from inside ticks during the transmission bloodmeal. We developed a novel sequencing-based strategy for *ex vivo* transcriptomic profiling of *Bb* populations within feeding ticks. We identified 192 highly differentially expressed genes, including both genes previously implicated in *Bb* transmission as well as many others. Genes upregulated during tick transmission were enriched for outer surface lipoproteins, suggesting *Bb* dramatically remodels its cell envelope as it migrates through the tick. Mass spectrometry analyses revealed dramatic changes in the tick environment over feeding, identifying new potential determinants of a more extensive and diverse set of tick–microbe molecular interactions than previously appreciated.

## RESULTS

### A two-step enrichment process facilitates robust transcriptional profiling of *Bb* during the tick bloodmeal

To gain a more comprehensive understanding of *Bb* gene expression throughout the tick phase of the transmission cycle, we developed an experimental approach to characterize the *Bb* transcriptome of spirochetes isolated from ticks during a bloodmeal. We aimed to establish a longitudinal transcriptional profile encompassing key pathogen transmission events each day of feeding after ticks attached to their mouse bloodmeal hosts (**Figure 1A**). The major bottleneck for such an RNA sequencing (RNA-seq) approach is capturing sufficient quantities of *Bb* transcripts from complex multi-organism samples in which pathogen transcripts represent a very small minority. Our initial attempts to uncover *Bb* mRNA by simply removing tick mRNA with polyA-depletion and removing tick rRNA sequences using Depletion of Abundant Sequences by Hybridization (DASH) (Dynerman et al., 2020; Gu et al., 2016) were unsuccessful. This approach resulted in an average of only 0.09% of RNA-seq reads mapping to *Bb* mRNA – approximately 10-fold less than we estimated would be needed to feasibly obtain robust transcriptome-wide gene expression analysis (Haas et al., 2012).

To dramatically increase *Bb* transcript representation in our libraries, we physically enriched *Bb* cells in tick lysates prior to library preparation by adding an initial step of immunomagnetic separation (**Figure 1B**). We took advantage of a commercial antibody previously generated against whole *Bb* cells (α*Bb*, RRID: AB_1016668). By western blot analysis, we confirmed that α*Bb* specifically recognized several *Bb* proteins, including surface protein OspA, which is highly prevalent on the *Bb* surface in ticks (**Figure S1A**). In addition, immunofluorescence microscopy with α*Bb* showed clear recognition of *Bb* cells from within the tick at each day of feeding (**Figure S1B**). After collecting infected ticks from mice one, two, three, and four days post-attachment, we used α*Bb* and magnetic beads to enrich *Bb* cells away from the majority of the tick material in the lysates. We tracked relative *Bb* enrichment through RT-qPCR of *Bb flaB* RNA and tick *gapdh* RNA in the separated samples. Measuring *Bb flaB* RNA from both *Bb*-enriched samples and their

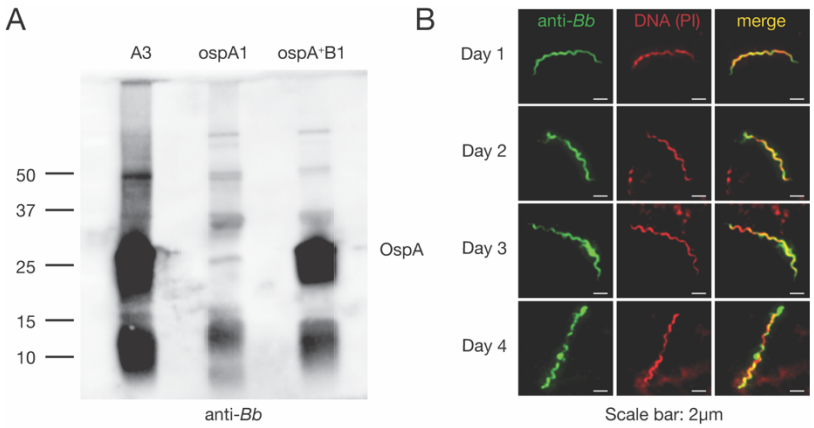


**Figure S1. Anti-*Bb* antibody recognizes ospA and binds *Bb* in the tick throughout the bloodmeal.** (**A**) Western blot with anti-*Bb* on lysate from cultured *Bb*: wildtype (A3, left), a mutant lacking ospA (ospA1), and the mutant with ospA restored (ospA+B1). Anti-*Bb* recognizes ospA among other proteins. (**B**) Immunofluorescence microscopy with anti-*Bb* (green) and propidium iodide (PI) (DNA, red) on each day of feeding (yellow is merge). Anti-*Bb* antibody recognizes *Bb* in the tick across the bloodmeal.

137 matched *Bb*-depleted fractions, we found over 95% of total *Bb flaB* RNA was present in enriched
138 fractions (**Figure 1C**), suggesting our approach captured the vast majority of *Bb* transcripts from
139 the tick.

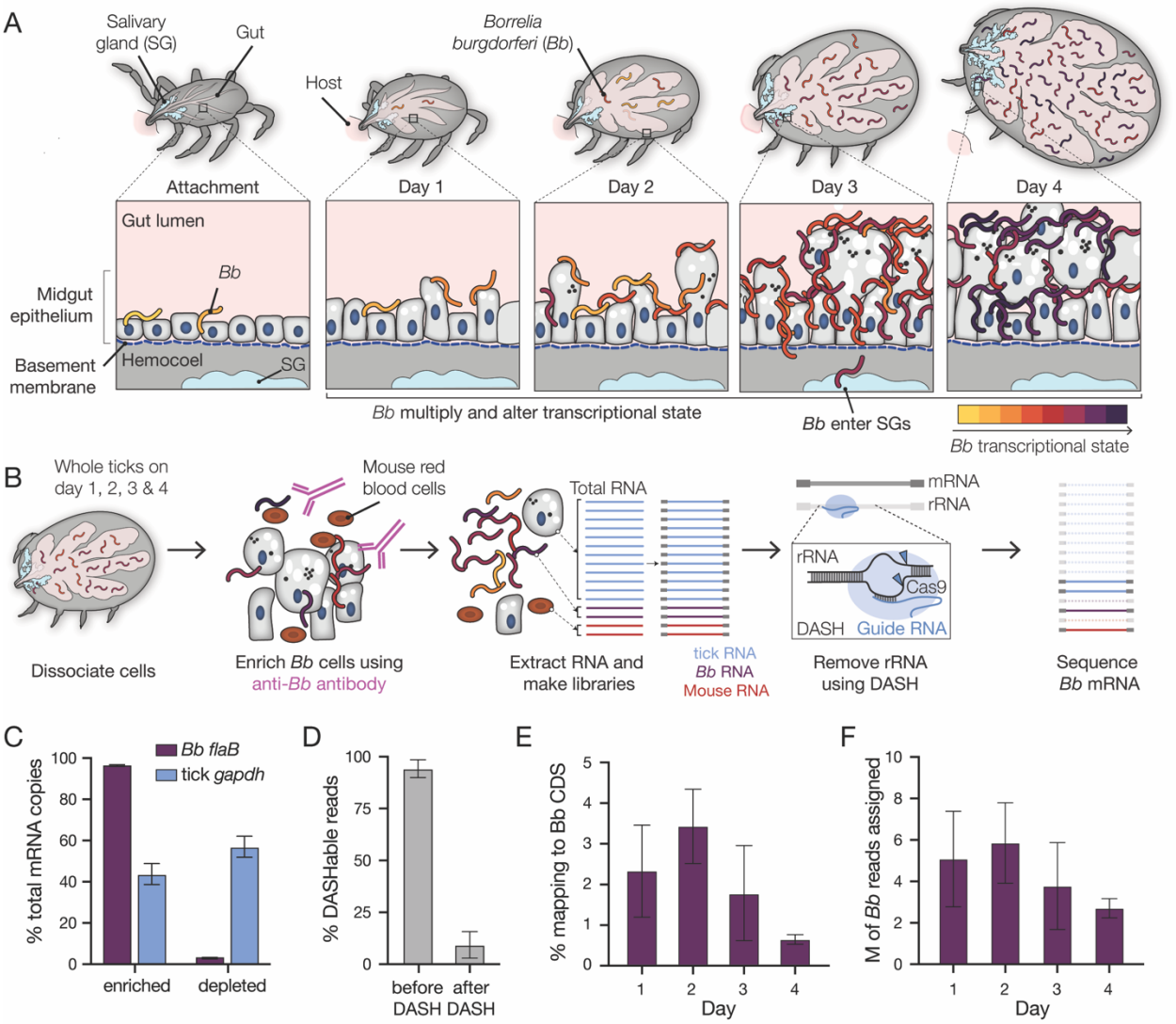


**Figure 1. A two-step enrichment process facilitates robust transcriptional profiling of *Bb* during the tick bloodmeal.** (**A**) Schematic of *Bb* during tick feeding. *Bb* in the nymphal tick midgut respond to the bloodmeal by multiplying and changing their transcriptional state. At the same time, the tick gut undergoes numerous changes to digest the bloodmeal. After two to three days of feeding, a small number of *Bb* leave the midgut and enter the salivary glands (blue), while the majority are left behind in the gut after engorgement. (**B**) Schematic of *Bb* enrichment process from feeding ticks. Whole ticks are dissociated, anti-*Bb* antibodies are added to lysates, and antibodies and *Bb* are captured magnetically. RNA is extracted and RNA-seq libraries are prepared. DASH is then used to remove rRNA before sequencing. This process increases *Bb* reads in the resulting sequencing. (**C**) RT-qPCR results showing the percentage of *Bb flaB* and tick *gapdh* RNA in the enriched versus depleted fractions after the enrichment process. Data come from 4 replicates each from day 2, day 3, and day 4, mean +/- SE. Nearly all *Bb flaB* RNA was found in the enriched fraction, while the majority of tick *gapdh* was found in the depleted fraction. (**D**) The percentage of reads mapping to rRNA before and after DASH. Data are shown as mean +/- SD. rRNA reads are drastically reduced after DASH. (**E**) The percentage of reads in RNA-seq libraries mapping to *Bb*. *Bb* mRNA reads make up a small but sufficient proportion of libraries for transcriptomics. Data are shown as mean +/- SD. (**F**) The number of reads in millions (M) mapped to *Bb* for each day. Data are shown as mean +/- SD. An average of 4.3 million reads per sample mapped to Bb genes, covering 92% of genes with at least 10 reads.

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141 Total RNA recovered from the enrichment process was used to create RNA-seq libraries that
142 subsequently underwent depletion of rRNA sequences from ticks, mouse, and *Bb* using DASH,
143 which targets unwanted sequences for degradation by Cas9 (Gu et al., 2016; Ring et al., 2022).
144 DASH reduced unwanted sequences from 94% to 9% of our total libraries, greatly increasing the
145 relative abundance of *Bb* transcripts (**Figure 1D**). For resulting libraries generated across feeding
146 (**Table S1**), between 0.6% and 3.4% of reads mapped to *Bb*
147 coding regions (**Figure 1E**), which translated to an average
148 of 4.3 million *Bb* reads per sample (**Figure 1F**). Read
149 coverage was sufficient for downstream differential
150 expression analyses for the vast majority of *Bb* genes, with
151 at least 10 reads mapping to 92% of *Bb* genes. To evaluate
152 whether our approach introduced any major artifacts, we
153 sequenced and compared RNA-seq libraries from *in vitro*
154 cultured *Bb* cells before and after immunomagnetic
155 enrichment. We found minimal expression differences (29
156 genes with $p<0.05$, fold changes between 0.83-1.12)
157 (**Figure S2, Table S2**), suggesting experimental enrichment
158 did not significantly alter global transcriptome profiles for *Bb*.
159 Thus, we developed an approach to enable genome-wide
160 analysis of *Bb* population-level expression changes that
161 occur within the feeding tick during transmission.

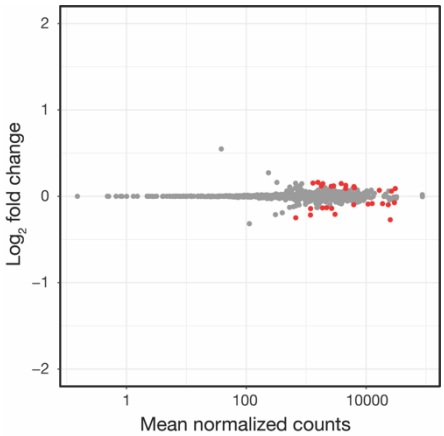


**Figure S2. Enrichment process does not induce large scale gene expression changes in *in vitro* cultured *Bb*.** $Log_2$ fold changes vs. mean normalized number of counts comparing cultured *Bb* input and samples after enrichment with anti-*Bb*. n=3. Red dots, p-value < 0.05, Wald tests. The gene expression changes induced during processing are much smaller than those observed between days of feeding.

162

163 **Global *ex vivo* profiling of *Bb* reveals extent and kinetics of transcriptional changes**

164

165 To provide a broad overview of *Bb* expression changes during feeding, we performed principal
166 component analysis (PCA) on the *Bb* transcriptome data from one, two, three, and four days post-
167 attachment (n=4). We reasoned that if many longitudinal expression changes were occurring
168 across *Bb* populations, we would observe greater data variability between time points than
169 between biological replicates. Indeed, we found replicates from each day grouped together,
170 whereas distinct time points were largely non-overlapping. The first principal component, which
171 explained 64% of the variance in our data, correlated well with day of feeding (**Figure 2A**). The
172 global pattern suggested that *Bb* gene expression changes generally trended in the same

173 direction over the course of feeding with the most dramatic differences between flanking
174 timepoints on day 1 and day 4.
175
176 Using day 1 (early attachment) as a baseline, we performed differential expression analysis for
177 all *Bb* genes at subsequent time points and examined changes above a twofold threshold (**Figure**
178 **2B, Table S3**). These analyses mirrored the global longitudinal expression pattern predicted by
179 our PCA. The total number of differentially expressed (DE) genes increased steadily over time
180 and peaked at day 4, which when compared to day 1 had 186 DE genes, including 153
181 upregulated and 33 downregulated (**Figure 2C-E**). Across time points, DE genes were highly
182 overlapping and largely changed in the same directions. In all three comparisons, we found 192
183 DE genes in total (**Table S4**). However, there were some notable differences between genes,
184 such as the overall timing and kinetics of expression changes. Transcript levels for some DE

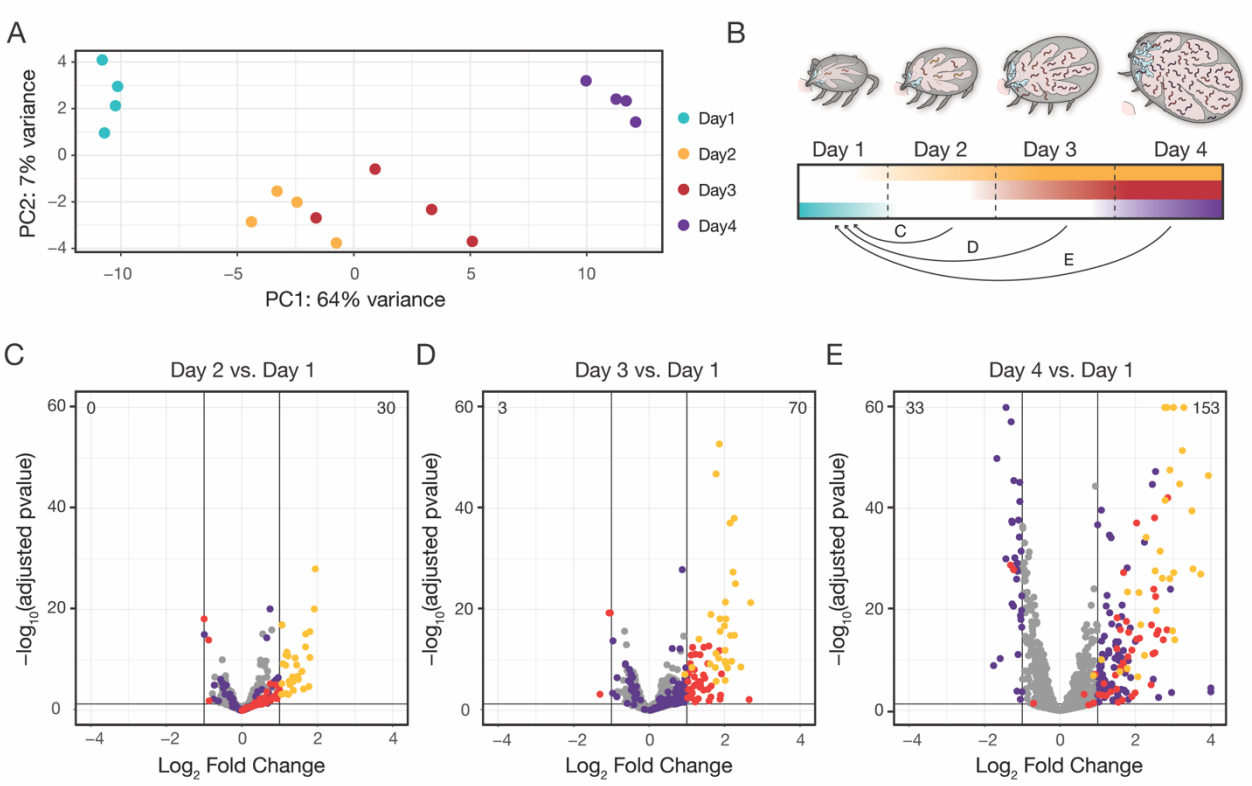


**Figure 2. Global *ex vivo* profiling of *Bb* reveals extent and kinetics of transcriptional changes.** (**A**) Principal component analysis of samples from across feeding. PC1 correlates strongly with day of feeding. (**B**) Schematic depicting how data was analyzed, as pairwise comparisons between the first day after attachment and all other days. (**C-E**) Volcano plots of differentially expressed genes of day 2 versus day 1 after attachment (**C**), day 3 versus day 1 (**D**), and day 4 versus day 1 (**E**). The total number of upregulated genes is shown in the top right and downregulated genes is shown in the top left. Yellow dots are genes that change expression between day 1 and day 2, red dots are genes that change expression between day 1 and day 3, and purple dots are genes that change expression between day 1 and day 4. Two genes with $\log_2$ fold changes >4 are shown at 4, and five genes with -$\log_{10}$(padj) >60 are shown at 60. Only genes with at least a 2-fold change are highlighted. p-value < 0.05, Wald tests. By day 4 of feeding, 153 genes are upregulated and 33 genes are downregulated from day 1 baseline levels.

genes changed suddenly over the course of feeding, while others were more gradual. To our knowledge, this is the first comprehensive report of global *Bb* expression changes within feeding ticks.

To help assess the integrity of our dataset, we examined expression profiles for previously characterized targets of major transcriptional programs activated at the onset of the bloodmeal, the Hk1/Rrp1 two-component system and the RpoN/RpoS sigma factor cascade (Caimano et al., 2019, 2015). Between day 1 and day 4, the majority of genes previously reported to be activated or repressed by Rrp1 *in vitro* (Caimano et al., 2015) trended significantly in the expected direction *ex vivo* (111/148 upregulated, 37/57 downregulated, p<0.05, Wald tests, **Figure S3A**). The majority of the genes previously reported to be activated by RpoS in a mammalian context, including canonical targets *ospC* and *dbpA*, were also significantly upregulated during tick feeding (33/55, p<0.05, Wald tests) (**Figure S3B**), along with *rpoS* itself (**Figure S3C**), as expected (Hübner et al., 2001). However, very few genes repressed by RpoS in mammals were similarly repressed in ticks (2/33, p<0.05, Wald tests). These data support the hypothesis that while RpoS-dependent gene activation occurs across both vertebrate and vector stages of *Bb* transmission, RpoS-dependent repression is specific to spirochetes infecting a mammalian host (Caimano et

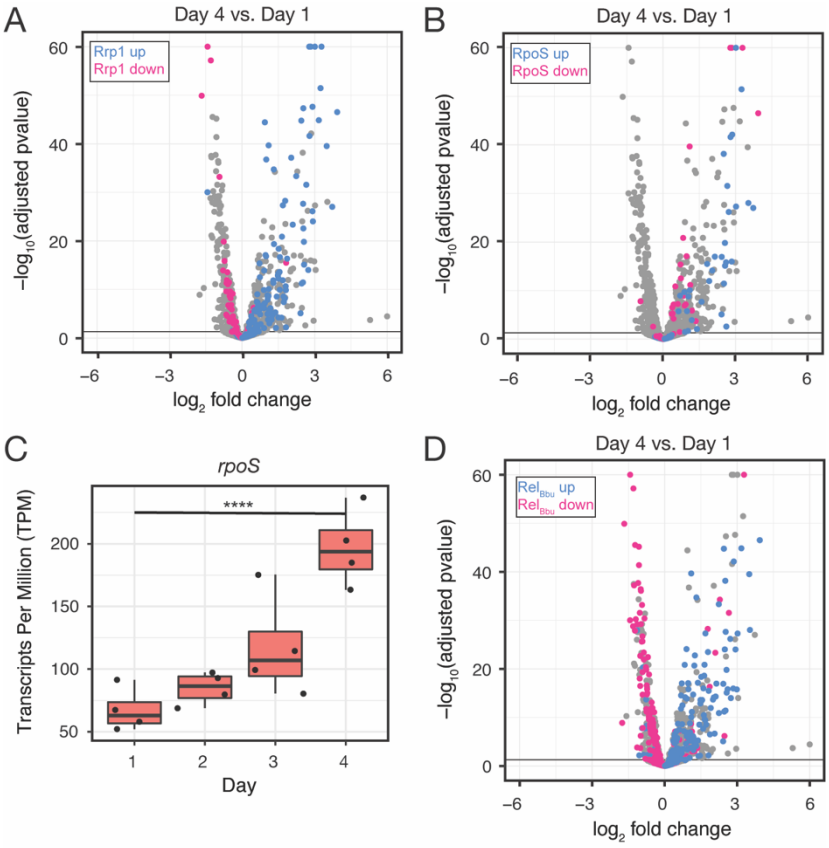


**Figure S3. *Ex vivo* RNA-seq corroborates key transcriptional programs in the tick.** (**A**) Volcano plot of DE genes comparing day 4 to day 1, with Rrp1-upregulated (blue) and downregulated (pink) genes. Rrp1-regulated genes correlate well with up and downregulated genes during feeding. (**B**) Volcano plot of DE genes comparing day 4 to day 1, with RpoS-upregulated (blue) and downregulated (pink) genes. Genes upregulated by RpoS in mammals increase during feeding, but genes repressed by RpoS in mammals are not repressed in the tick. (**C**) Tukey style boxplot of Transcripts Per Million (TPM) on each day for *rpoS*. Black dots represent replicates. ****p-value < 0.00001, Wald test. *rpoS* expression increases over the course of feeding. (**D**) Volcano plot of DE genes comparing day 4 to day 1, with $Rel_{Bbu}$-upregulated (blue) and downregulated (pink) genes. About half of $Rel_{Bbu}$ genes change in the expected direction over feeding.

202 al., 2019, 2007). We also compared our DE genes to those regulated by $Rel_{Bbu}$ as part of the
203 stringent response, another major transcriptional program active in *Bb* in the tick during nutrient
204 starvation (Drecktrah et al., 2015). About half of $Rel_{Bbu}$-regulated genes changed in the direction
205 expected if the stringent response was active during this time (**Figure S3D**); however, more of
206 the genes that were downregulated over feeding have been reported to be regulated by $Rel_{Bbu}$
207 than either RpoS or Rrp1, suggesting it may play some role during feeding.
208
209 By looking at these previously reported transcriptional regulons in our data, we were able to verify
210 that our data captured expected transcriptional programs active during tick feeding. Nevertheless,
211 25% of the twofold DE genes were not previously identified through these RNA-seq studies as
212 dependent upon RpoS, Rrp1, or $Rel_{Bbu}$ (Caimano et al., 2019, 2015; Drecktrah et al., 2015), which
213 are the three main *Bb* regulatory programs active in the tick (Samuels et al., 2021). These
214 additional genes highlight the necessity of measuring transcription in the tick environment and
215 suggest we uncovered gene expression changes specific to the tick-stage of the *Bb* enzootic
216 cycle. The nature and dynamics of these changes provide insights into potential genetic
217 determinants of *Bb* survival, proliferation, and dissemination in the tick during transmission.
218
219 ***Bb* genes upregulated during feeding are found predominantly on plasmids**
220
221 *Bb* has a complex, highly fragmented genome (Barbour, 1988) (**Figure 3A**), including numerous
222 plasmids that are necessary during specific stages of the enzootic cycle (reviewed in Schwartz et
223 al., 2021) suggesting they contain genes that are crucial for pathogen transmission and survival.
224 In fact, many genes found on the plasmids have been previously shown to alter expression upon
225 environmental changes or in different host environments (Iyer et al., 2015; Ojaimi et al., 2005;
226 Revel et al., 2002; Tokarz et al., 2004). Thus, we reasoned that many of the 192 *Bb* genes that
227 change expression over the course of the tick bloodmeal would reside on the plasmids, and we
228 examined their distribution throughout the genome. Consistent with these previous reports, we
229 found that most of the upregulated genes were located on the plasmids (143/158; 90%), while
230 fewer were found on the chromosome (15/158; 10%) (**Figure 3B**), which is home to the majority
231 of metabolic and other housekeeping genes. In contrast, the majority of the downregulated genes
232 were found on the chromosome (27/34, 79%) (**Fig 3C**).
233
234 Several plasmid-encoded genes that were longitudinally upregulated in our datasets have known
235 roles during the tick bloodmeal or in mammalian infection. For example, linear plasmid 54 (lp54),

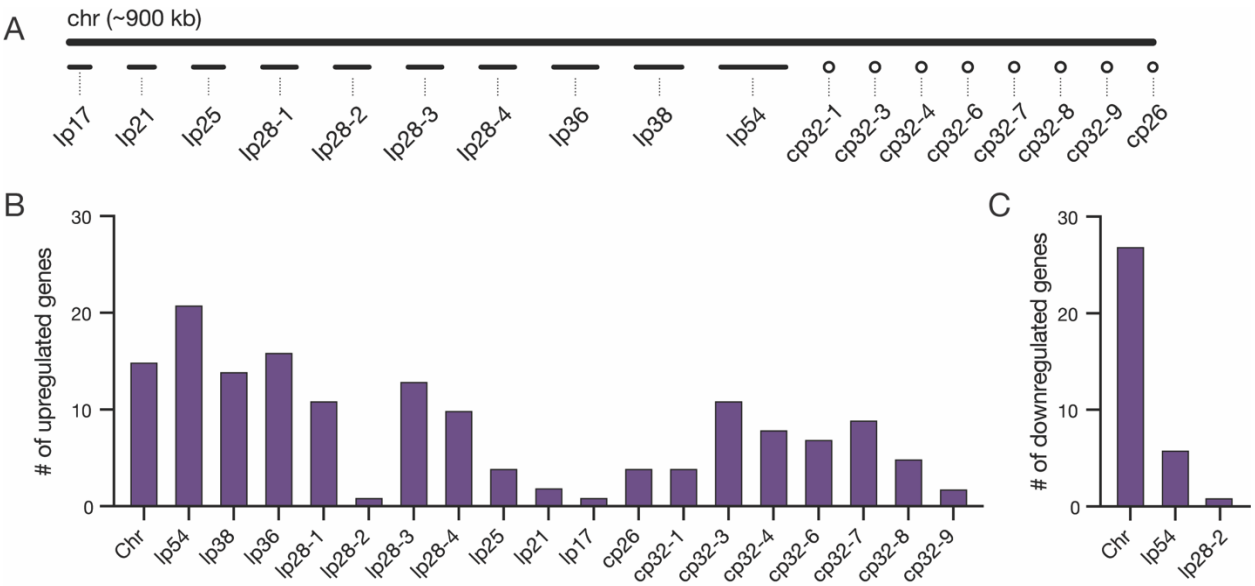


**Figure 3. *Bb* genes upregulated during feeding are found predominantly on plasmids.** (**A**) Schematic of the chromosome and plasmids in the *Bb* B31-S9 genome. (**B-C**) The number of genes from each chromosome or plasmid that increased (**B**) or decreased expression (**C**) twofold during feeding. Upregulated genes are distributed across plasmids, while downregulated genes are found on the chromosome and lp54.

which is an essential plasmid present in all *Bb* isolates (Casjens et al., 2012), contained the largest number of upregulated genes. Many of the genes on lp54 are known to be regulated by RpoS during feeding, including those encoding adhesins DbpA and DbpB, which are important for infectivity in the host (Blevins et al., 2008). This set also included five members of a paralogous family of outer surface lipoproteins BBA64, BBA65, BBA66, BBA71, and BBA73. BBA64 and BBA66 are necessary for optimal transmission via the tick bite (Gilmore et al., 2010; Patton et al., 2013). These findings indicate our dataset captures key *Bb* transcriptional responses known to be important for survival inside the tick during a bloodmeal.

Many upregulated genes were encoded by cp32 plasmid prophages. *Bb* strain B31-S9 harbors seven cp32 isoforms that are highly similar to each other (Casjens et al., 2012). When cp32 prophages are induced, phage virions called ϕBB1 are produced (Eggers and Samuels, 1999). In addition to phage structural genes, cp32 contain loci that encode various families of paralogous outer surface proteins (Stevenson et al., 2000). Amongst the cp32 genes that increased over feeding were members of the RevA, Erp, and Mlp families, which are known to increase expression during the bloodmeal (Gilmore et al., 2001). We also found several phage genes that were upregulated, including those encoding proteins annotated as phage terminases on cp32-3, cp32-4, and cp32-7 (BBS45, BBR45, and BBO44). Some cp32 genes have been shown to change expression in response to temperature changes (Tokarz et al., 2004) and as a part of the stringent response regulated by $Rel_{Bbu}$ (Drecktrah et al., 2015). Our data suggest that some

256 prophage genes are upregulated over the course of tick feeding, raising the possibility that cp32
257 prophage are induced towards the end of feeding. Overall, our data support the long-held idea
258 that the *Bb* plasmids, which house many genes encoding cell envelope proteins, proteins of
259 unknown function, and prophage genes, play a critical role in the enzootic cycle during the key
260 transition period of tick feeding.
261
262 ***Bb* genes encoding outer surface proteins are enriched among upregulated genes**
263
264 To gain a better overall sense of the types of genes that changed over feeding and the timing of
265 those changes, we grouped DE genes into functional categories. Since a high proportion of
266 plasmid genes encode lipoproteins within the unique protein-rich outer surface of *Bb*, genes of
267 unknown function, and predicted prophage genes (Casjens et al., 2000; Fraser et al., 1997), we
268 expected that many of the DE genes would fall into these categories. We classified the genes as
269 related to either: cell envelope, bacteriophage, cell division, DNA replication and repair,
270 chemotaxis and motility, metabolism, transporter proteins, transcription, translation, stress
271 response, protein degradation, or unknown (as in Drecktrah et al., 2015)(see **Table S4**).
272
273 Of the genes that increased at least twofold over feeding, the clear majority each day of feeding
274 fell into two broad categories, cell envelope (55 of 158 total genes) and proteins of unknown
275 function (69 of 158 total genes), with fewer genes related to metabolism, chemotaxis and motility,
276 transporters, bacteriophage, cell division, and transcription (**Figure 4A**). In contrast to the
277 upregulated genes, genes downregulated during feeding were more evenly distributed among the
278 functional categories – including translation, protein degradation, transcription, and metabolism –
279 consistent with many of them being located on the chromosome.
280
281 When looking at changes across these functional categories, the overrepresentation of cell
282 envelope proteins was striking, while not unexpected. The *Bb* outer surface is covered with
283 lipoproteins (**Figure 4B**), and these proteins are critical determinants in *Bb* interactions with the
284 various environments encountered during the enzootic cycle (Kurokawa et al., 2020). We found
285 that more than half of these outer surface lipoproteins changed expression during this time, with
286 46 of 83 total outer surface lipoproteins (Dowdell et al., 2017) differentially expressed twofold over
287 feeding (**Figure 4C**). These data suggested widespread changes may be occurring on the *Bb*
288 outer surface during feeding.
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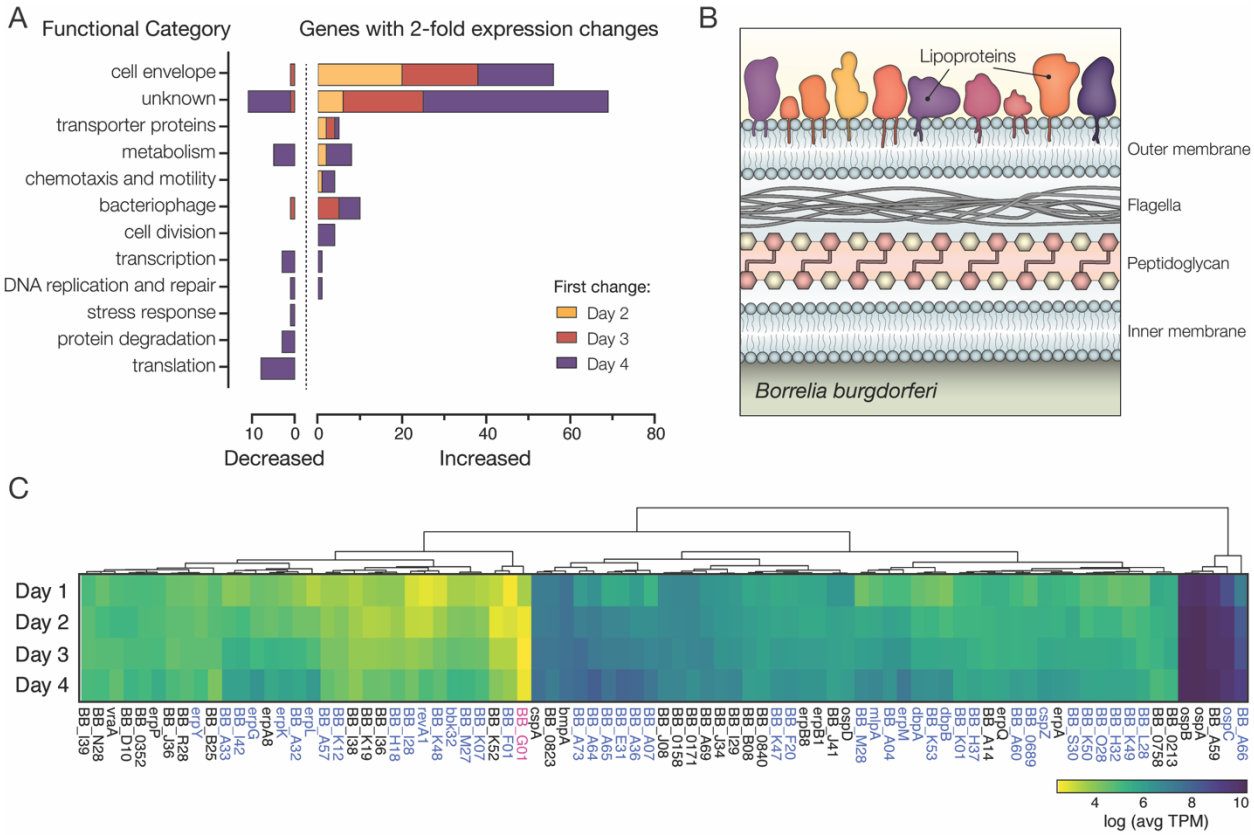


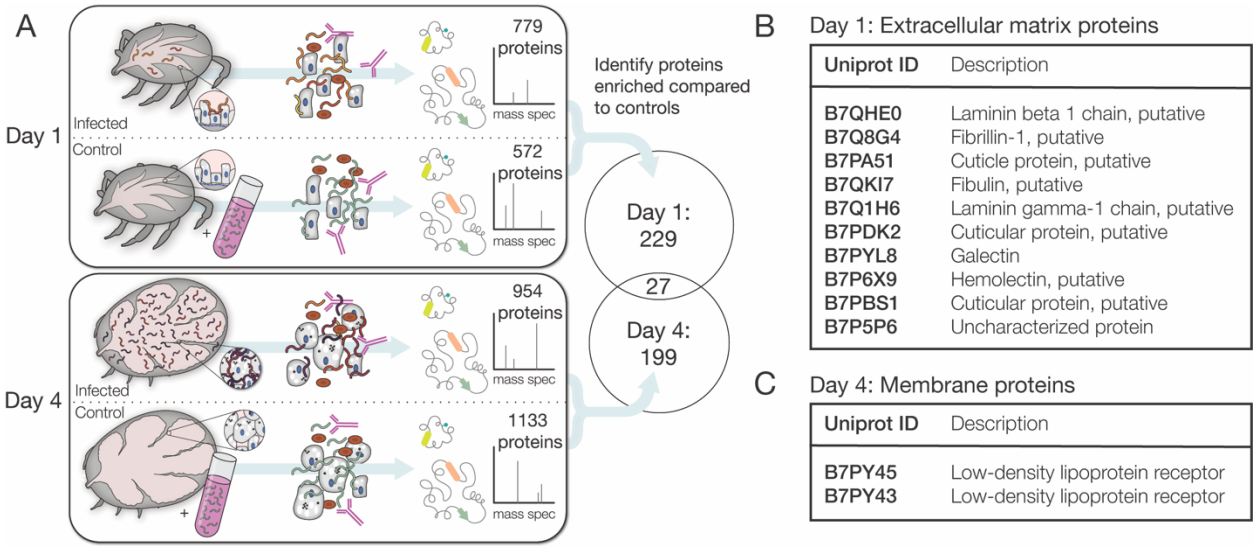

**Figure 4. *Bb* genes encoding outer surface proteins are enriched among upregulated genes.** (**A**) The number of *Bb* genes that change over the course of tick feeding sorted into functional categories. Genes that first change 2 days after attachment are shown in yellow, 3 days after attachment in red, and 4 days after attachment in purple. A majority of upregulated genes fall into cell envelope and unknown categories. (**B**) Schematic of the outer membrane of *Bb* showing the location of outer surface lipoproteins. (**C**) Heat map of the average Transcripts Per Million (TPM) of all genes encoding outer surface lipoproteins across the 4 days of tick feeding. Genes in blue were twofold upregulated and genes in pink twofold downregulated over feeding. A majority of genes encoding outer surface proteins increased in expression throughout feeding, while having different magnitudes of expression.

To understand the functional implications of expression changes in a majority of outer surface lipoproteins, we also compared their relative expression. The magnitude of expression varied greatly, with *ospA*, *ospB*, *ospC*, and *bba59* being the most highly expressed outer surface protein transcripts. Many of the outer surface protein genes that we found increased expression over feeding were much less abundantly expressed (**Table S5**). However, even the genes that appeared to have low expression in these population level measurements could play important roles in transmission if they are highly expressed in a small number of crucial cells, such as those that ultimately escape the midgut. While bulk RNA-seq cannot distinguish what is happening at the single cell level, our data suggest that during the bloodmeal, *Bb* are undergoing a complex outer surface transformation driven by increases in a majority of the genes encoding these crucial lipoproteins.

**Identification of candidate tick interaction partners of *Bb* cells *ex vivo***

Our RNA-seq data suggested that the outer surface of *Bb* transforms over the course of feeding as *Bb* are primed for transmission to a vertebrate host. At the same time, the tick midgut environment is changing as the tick begins to digest its bloodmeal. Tick-*Bb* interactions are likely crucial at the beginning of the tick bloodmeal, as *Bb* adhere to the tick gut epithelium before becoming motile and migrating out of the midgut and into the salivary glands (Dunham-Ems et al., 2009). Some *Bb* outer surface proteins, such as BBE31 and BBA52 play key roles in pathogen migration through interactions with the tick environment (Kurokawa et al., 2020). We wanted to explore the changing tick environment to identify tick proteins with which *Bb* could interact throughout the tick bloodmeal. Since our *Bb* enrichment process retained some tick material, we reasoned that tick proteins that interact with *Bb* would be present in these samples.

To outline the changes occurring in the tick during feeding and to identify candidate *Bb*-interacting tick proteins, we used mass spectrometry to survey the content of the tick material that was enriched along with the *Bb* cells we sequenced at early and late stages during feeding. We purified proteins from the α*Bb*-enriched fraction of crushed infected ticks on day 1 after attachment and day 4 after attachment in triplicate. As controls for each day, we also performed the *Bb* enrichment process on lysate from uninfected ticks mixed with *in vitro* cultured *Bb* to help rule out proteins




| Uniprot ID | Description |
|---|---|
| B7QHE0 | Laminin beta 1 chain, putative |
| B7Q8G4 | Fibrillin-1, putative |
| B7PA51 | Cuticle protein, putative |
| B7QKI7 | Fibulin, putative |
| B7Q1H6 | Laminin gamma-1 chain, putative |
| B7PDK2 | Cuticular protein, putative |
| B7PYL8 | Galectin |
| B7P6X9 | Hemolectin, putative |
| B7PBS1 | Cuticular protein, putative |
| B7P5P6 | Uncharacterized protein |

| Uniprot ID | Description |
|---|---|
| B7PY45 | Low-density lipoprotein receptor |
| B7PY43 | Low-density lipoprotein receptor |

**Figure 5. Identification of candidate tick interaction partners of *Bb* cells *ex vivo*.** (**A**) Schematic of experiment to determine candidate tick proteins interacting with *Bb* over the course of feeding. Ticks were collected 1 day after attachment and 4 days after attachment. Uninfected ticks at the same time points were mixed with cultured *Bb* as controls. *Bb* was enriched with anti-*Bb* antibody as in RNA-seq experiments and then subjected to mass spectrometry to identify tick proteins present in the samples. Venn diagram depicts the proteins enriched in day 1 and day 4 samples over controls. Tick proteins that are enriched with *Bb* vary greatly over the course of feeding. (**B**) Tick proteins uniquely identified one day after attachment that are annotated as extracellular matrix (ECM) proteins. (**C**) Tick proteins uniquely identified four days after attachment that are annotated as low-density lipoprotein receptors. ECM and membrane proteins may be good candidates for *Bb*-interacting proteins.

321 that were not pulled down through *in vivo* tick-*Bb* interactions (**Figure 5A**). We identified between
322 414 and 2240 protein groups per sample replicate. To identify proteins of interest, we looked for
323 those that were detected in at least two of three replicates with infected ticks and had a mean
324 average coverage twice that of uninfected ticks mixed with cultured *Bb*. We found 256 proteins
325 that were enriched with *Bb* from infected ticks one day after attachment and 226 proteins that
326 were enriched with *Bb* from infected ticks four days after attachment (**Table S6**). Of these proteins,
327 only 27 were detectable on both days, suggesting the tick proteins present upon *Bb* enrichment
328 change dramatically over the course of feeding. The vast majority of all detected proteins were
329 from ticks (98%). Amongst the small number of *Bb* proteins, we identified OspC in the samples
330 from day 4 after attachment but not day 1 after attachment, confirming the expression change we
331 saw in our RNA-seq. The distinct sets of tick proteins we identified at each timepoint suggest
332 dramatic changes occur in the *Bb*-infected tick midgut environment during feeding that may alter
333 the landscape of tick-*Bb* interactions.
334
335 Some of the proteins enriched with *Bb* from the tick may be good candidates for key *Bb*-interacting
336 partners during feeding, especially if they are localized to the surface of tick cells where they may
337 encounter *Bb*. The scarcity of both predicted and experimentally validated functions and
338 localizations for tick proteins makes it difficult to fully assess the potential for tick protein
339 interactions with extracellular *Bb*. Nevertheless, of the proteins found exclusively one day after
340 attachment, 10 were categorized as extracellular matrix proteins using the PANTHER gene
341 database (Thomas et al., 2022), and this category was statistically enriched (Fisher's exact test,
342 FDR=0.000093) (**Figure 5B, Table S7**). 30 additional proteins were annotated with a cellular
343 component as plasma membrane. Four days after attachment, we did not detect any annotated
344 extracellular matrix proteins; however, we identified 31 proteins that are likely to be found at the
345 membrane, including two proteins annotated as putative low-density lipoprotein receptors (**Figure
346 5C, Table S8**). These extracellular matrix and membrane proteins may be the most likely to
347 directly interact with *Bb* during this timeframe and are candidates for tick proteins important in the
348 *Bb* dissemination process. The proteins present in the changing tick environment may be key
349 determinants of pathogen transmission as *Bb* remodels its outer surface while preparing to
350 migrate through the tick to a new host.
351
352 **DISCUSSION**
353 Vector-borne pathogens must adapt to distinct environments as they are transmitted by
354 arthropods to colonize new bloodmeal hosts. The tick-borne Lyme disease pathogen *Bb*

355 undergoes transcriptional changes inside its vector over the course of a single bloodmeal that are
356 important for a number of key transmission events. For example, expression changes enable
357 survival in the feeding tick (He et al., 2011), facilitate transmission across several internal
358 compartments into the bloodmeal host (Kurokawa et al., 2020), and prime bacteria for successful
359 infection of the next host (Kasumba et al., 2016). Capturing these changes as they occur *in vivo*
360 has been challenging due to the low relative abundance of *Bb* material inside of the rapidly
361 growing, blood-filled tick. To date, many advances in our understanding of *Bb* expression during
362 transmission have come from tracking changes in small subsets of genes during tick feeding
363 (Gilmore et al., 2001), leveraging *Bb* culture conditions that approximate environmental changes
364 across its lifecycle (Ojaimi et al., 2005; Revel et al., 2002; Tokarz et al., 2004), and defining
365 transcriptional regulons outside of the tick (Caimano et al., 2019, 2015; Drecktrah et al., 2015).
366 Here, we developed an experimental RNA-seq-based strategy to more directly and longitudinally
367 profile gene expression for *Bb* populations isolated from ticks over the course of a transmitting
368 bloodmeal.
369
370 Our longitudinal collection of transcriptomes for *Bb* cells isolated from ticks serves as a resource
371 and starting point for delineating the functional determinants of *Bb* adaptation and transmission.
372 Our transcriptome profiles reveal more extensive expression changes for *Bb* outer surface
373 proteins than previously appreciated. These results raise the possibility that *Bb* is actively
374 remodeling its outer coat to navigate the dynamic tick environment during feeding, akin to
375 wardrobe changes across seasons. These outer surface changes may play roles in the
376 persistence of *Bb* in the tick, cell adhesion, or in immune evasion, either inside of the tick or later
377 in the vertebrate host (Kenedy et al., 2012). It has long been observed that molecular interactions
378 between pathogens and the midgut of their vectors are key determinants of transmission (Barillas-
379 Mury et al., 2022). Modifications to the *Bb* surface could facilitate different tick–pathogen
380 interactions that are critical for its physical movement through tick compartments. Notably, we
381 also identified numerous genes of unknown function that change expression during feeding.
382 Although challenging to address, efforts focused on uncovering the contribution of these genes
383 to the *Bb* life cycle could be groundbreaking for understanding the unique aspects of tick-borne
384 transmission.
385
386 More comprehensive knowledge about *Bb* outer surface proteins and their functional
387 consequences could lead to new avenues for curtailing the spread of Lyme disease. Protective
388 vaccines against vector-borne pathogens have often targeted surface proteins encoded by

389 pathogens (Kovacs-Simon et al., 2010). While previous Lyme disease vaccine efforts effectively
390 targeted the highly expressed *Bb* outer surface protein OspA (Steere et al., 1998), more targets
391 could increase the likelihood of a successful vaccine. Our study provides a catalog of new *Bb*
392 candidates that could be explored. In addition, our biochemical pull-downs using tick-isolated *Bb*
393 cells as bait unearthed a preliminary list of potential tick proteins that could be involved in tick–*Bb*
394 molecular interactions. Blocking molecular interactions that are functionally critical for
395 transmission could also be explored as a therapeutic strategy (Barillas-Mury et al., 2022; Manning
396 and Cantaert, 2019).
397
398 Our transcriptomic resource provides critical insights into *Bb* population-level changes during the
399 vector stage of its lifecycle, an important starting point for understanding the primary drivers of
400 tick-borne transmission. Strikingly few *Bb* cells out of the total pathogen population in ticks are
401 ultimately transmitted to the next host during feeding. There are several major bottlenecks to
402 infection, including dissemination to the tick salivary glands and survival upon inoculation of the
403 host (Dunham-Ems et al., 2009; Rego et al., 2014). There may be important physiological or
404 molecular variations across *Bb* cells within the population residing in the tick that contribute to
405 these differential outcomes. An important clue is the fact that *Bb* proteins are indeed
406 heterogeneously expressed across cells within the feeding tick midgut (Ohnishi et al., 2001).
407 Conducting transcriptomic analyses at the single-cell level will be key. Given that only a small
408 number of *Bb* escape the tick midgut, genes that appear unaltered or show low expression levels
409 across the bulk population could still play an outsized role in infection for a minority of the cells.
410 Our work provides a foundational methodology that can be leveraged to greatly improve the
411 resolution of tick–microbe studies. Technological advances stemming from this work will provide
412 molecular details that potentiate new questions and may unearth surprising mechanistic insights
413 into the unique lifestyles of tick-borne pathogens.
414
415 **METHODS**
416 ***B. burgdorferi* culture**
417 *Bb* strain B31-S9 (Rego et al., 2011) was provided by Dr. Patricia Rosa (NIAID, NIH, RML) and
418 cultured in BSK II media at 35°C, 2.5% $CO_2$.
419
420 **Tick feeding experiments**
421 *I. scapularis* larvae were purchased from the Tick Lab at Oklahoma State University (OSU) (for
422 RNA-seq experiments) or provided by BEI Resources, a division of the Center for Disease Control

423 (for mass spectrometry experiments). Before and after feeding, ticks were maintained in glass
424 jars with a relative humidity of 95% (saturated solution of potassium nitrate) in a sealed incubator
425 at 22°C with a light cycle of 16h/8h (light/dark). Animal experiments were conducted in
426 accordance with the approval of the Institutional Animal Care and Use Committee (IACUC) at
427 UCSF. Ticks were fed on young (4–6-week-old) female C3H/HeJ mice acquired from Jackson
428 Laboratories. Mice were anesthetized with ketamine/xylazine before placement of ≤ 100 larval or
429 ≤ 30 nymphal ticks. Ticks were either pulled off isoflurane anesthetized mice at various times
430 during feeding (1-3 days after placement) or allowed to feed to repletion and collected from mouse
431 cages. Replete larval ticks were placed in the incubator to molt before being used as nymphs in
432 experiments.
433
434 **Western blot with anti-*Bb* antibody**
435 To determine whether the anti-*Bb* antibody targeted ospA, we cultured wildtype *Bb* (B31-A3),
436 ospA1-mutants (ospA1) and ospA-restored *Bb* (ospA$^{+}$B1) (Battisti et al., 2008) to approximately
437 5x10$^{7}$ *Bb*/mL and centrifuged 3 mLs of culture for 7 minutes at 8000 x g, washing twice with PBS.
438 Pelleted cells were lysed in 50 µL of water, and 25 ng per sample was mixed with 5X loading dye
439 (0.25% Bromophenol Blue, 50% Glycerol, 10% Sodium Dodecyl Sulfate, 0.25M Tris-Cl pH 6.8,
440 10% B-Mercaptoethanol), run on a Mini-PROTEAN TGX 4-15% gel (Biorad), and transferred
441 using the Trans-Blot Turbo Transfer System (Biorad). After transfer, the blot was blocked for 30
442 minutes at 4°C in TBST (Tris buffered saline with .1% tween) with 5% milk, then treated with anti-
443 *Bb* antibody (Invitrogen: PA1-73004; RRID: AB_1016668) diluted 1:10,000 for 1 hour at room
444 temperature, followed by anti-rabbit HRP secondary antibody (Advansta) diluted 1:5,000 for 45
445 minutes at room temperature with short PBST washes between each step. Blots were exposed
446 using Clarity Western ECL Substrate (Biorad) and imaged using the Azure C400 imaging system
447 (Azure Biosystems). This experiment was repeated three times.
448
449 **Enrichment of *Bb* from feeding ticks**
450 To sequence RNA from *Bb* from feeding ticks, we enriched *Bb* to increase the ratio of *Bb* to tick
451 material. Larval ticks were fed to repletion on three mice that were infected with *Bb* through
452 intraperitoneal and subcutaneous injection with 10$^{4}$ total *Bb*. Approximately five months later, the
453 molted nymphal ticks were fed on eight mice. We estimated that 83% of the ticks were infected
454 with *Bb* by crushing 12 unfed nymphs in BSK II media and checking for viable *Bb* days later. Ticks
455 were pulled from all mice and pooled into four replicates 1 day after placement (14 ticks per
456 replicate), 2 days after placement (12 ticks per replicate), and 3 days after placement (6 ticks per

457 replicate) and collected from cages 4 days after placement (7 ticks per replicate). Shortly after
458 collection, ticks were washed with water and placed in a 2 mL glass dounce grinder (Kimble) in
459 500 µL of phosphate-buffered saline (PBS). Ticks were homogenized first with the large clearance
460 pestle and then the small clearance pestle. The homogenate was transferred to a 1.5 mL
461 Eppendorf tube and 500 µL of PBS was added to total 1 mL. At this stage, 50 µL of homogenate
462 was removed as an input sample and mixed with 500 µL of TRIzol (Invitrogen) for RNA extraction.
463 2 µL of anti-*Bb* antibody (Invitrogen: PA1-73004; RRID: AB_1016668) was added to the
464 homogenate, which was then placed on a nutator at 4°C for 30 minutes. During incubation, 50 µL
465 of Dynabeads Protein G (Invitrogen) per sample were washed twice in PBS. After incubation with
466 the antibody, the homogenate and antibody mixture were added to the beads. This mixture was
467 placed on a nutator at 4°C for 30 minutes. Tubes were then placed on a magnet to secure beads,
468 and the homogenate was removed and saved to create depleted samples. The depleted
469 homogenate was centrifuged at 8,000 x g for 7 minutes, 900 µL of supernatant was removed, and
470 500 µL of TRIzol was added to the pellet to create depleted samples. The beads were washed
471 twice with 1 mL of PBS, resuspending the beads each time. The second wash was removed and
472 500 µL of TRIzol was added to the beads to create enriched samples. RNA was extracted from
473 all input, enriched, and depleted samples using the Zymo Direct-zol RNA Microprep Kit with on-
474 column DNase treatment (Zymo Research). The step-by-step *Bb* enrichment protocol is available
475 at: https://www.protocols.io/view/b-burgdorferi-enrichment-from-feeding-ticks-36wgqjrbovk5/v1
476
477 **Enrichment of *Bb* from culture**
478 To test whether the *Bb* enrichment process altered gene expression levels, we performed the
479 enrichment protocol on cultured *Bb*. We grew three tubes of *Bb* in BSK II media to $9x10^4$ *Bb*/mL
480 at 35°C. 1 mL of culture was spun down at 8,000 x g for 7 minutes, media was removed, *Bb* were
481 washed in 1 mL of PBS and spun again. Pelleted *Bb* were resuspended in 1 mL of fresh PBS.
482 These samples were used as starting homogenate for the *Bb* enrichment protocol and input,
483 enriched, and depleted fractions were collected as above. RNA-seq libraries from these samples
484 were prepared and sequenced as below.
485
486 **RNA-seq library preparation and sequencing**
487 To make RNA-seq libraries from enriched *Bb* RNA, we used 50 ng of total RNA as input into the
488 NEBNext Ultra II Directional RNA Library Prep Kit for Illumina (New England BioLabs). We
489 prepared libraries following the manufacturer's protocol for use of the kit with purified mRNA or

490 rRNA-depleted RNA, despite starting with total RNA. Libraries were barcoded using NEBNext
491 Multiplex Oligos for Illumina Dual Index (New England BioLabs).
492
493 To deplete reads from the libraries that were from tick, *Bb*, or mouse rRNA, we used Depletion of
494 Abundant Sequences by Hybridization (DASH) (Gu et al., 2016), which targets Cas9 to unwanted
495 reads in RNA-seq libraries using custom dual-guide RNAs (dgRNAs). Our dgRNAs targeted short
496 sequences within tick rRNA, mouse rRNA, and *Bb* rRNA that were designed using DASHit
497 software (Dynerman et al., 2020). We ordered crRNAs that targeted our sequences of interest
498 (Ring et al., 2022). To transcribe the dgRNAs, we followed our protocol for In Vitro Transcription
499 for dgRNA V2 (Lyden et al., 2019b). Both tracrRNA and pooled crRNA DNA templates were
500 annealed to equimolar amounts of T7 primer by heating to 95°C for 2 minutes and slowly cooling
501 to room temperature. Annealed templates were used in 1 mL in vitro transcription reactions with:
502 120 µL10X T7 buffer (400 mM Tris pH 7.9, 200 mM MgCl2, 50 mM DTT, 20 mM spermidine
503 (Sigma)), 100 µL of T7 enzyme (custom prepped enzyme from E. Crawford, diluted 1:100 in T7
504 buffer, final concentration: 100 µg/mL), 300 µl NTPs (25 mM each, Thermo Fisher Scientific), 4
505 µg of annealed crRNA template or 8 µg of annealed tracrRNA template, and water to 1 mL. In
506 vitro transcription was performed for 2 hours at 37°C. Reactions were purified twice with the Zymo
507 RNA Clean & Concentrator-5 Kit (Zymo Research). To form the dgRNA complex for DASH, crRNA
508 and tracrRNAs were diluted to 80 µM, mixed in equimolar amounts and annealed by heating to
509 95ºC for 30 seconds and cooling slowly to room temperature.

510 After transcription of dgRNAs, we performed DASH Protocol Version 4 (Lyden et al., 2019a) on
511 each library individually. Cas9 and transcribed dgRNAs were prepped by mixing: 2.5 µL 10X Cas9
512 buffer, 5 µL Cas9 (New England BioLabs), and 5 µL of 40 µM transcribed dgRNAs. The mixture
513 was incubated at 37°C for 5 minutes before 7.5 µL of RNA-seq library (2.8 nM) was added. The
514 mixture was incubated at 37°C for 1 hour and then purified with Zymo DNA Clean & Concentrator-
515 5 (Zymo Research) following the PCR product protocol and eluting DNA into 10.5 µL of water.
516 During cleanup, Cas9 was again mixed with buffer and dgRNAs and incubated at 37°C for 5
517 minutes. Following cleanup, eluted DNA was added to the second Cas9-dgRNA mixture and
518 incubated at 37°C for 1 hour for a second time. Then, 1 µL of proteinase K (New England Biolabs)
519 was added, and the mixture was incubated at 50°C for 15 min. The libraries were then purified
520 with 0.9x volume of sparQ PureMag Beads (QuantaBio) following the standard protocol, eluting
521 in 24 µL of water. rRNA-depleted RNA-seq libraries were then amplified in a BioRad CFX96 using
522 the Kapa HiFi Real-Time Amplification Kit in a 50 µL reaction with 25 µL master mix, 23 µL of the

523 DASHed library pool, and 2 µL of 25 µM mix of Illumina P5 (5'-
524 AATGATACGGCGACCACCGAGATCT) and P7 (5'- CAAGCAGAAGACGGCATACGAGAT)
525 primers. The qPCR program for amplification was as follows: 98°C for 45 sec (1 cycle), (98°C for
526 15 sec, 63°C for 30 sec, 72°C for 45 sec, plate read, 72°C for 20 sec) for 10 cycles (day 3 and
527 day 4 samples) or 11 cycles (day 1 and day 2 samples). The libraries were removed from cycling
528 conditions before leaving the exponential phase of amplification and then purified with 0.9X
529 volume of sparQ PureMag Beads according to the standard protocol.

530 Following DASH, RNA-seq libraries were sequenced on an Illumina NovaSeq S2 (2 lanes) with
531 paired-end 100 base pair reads. Libraries from *in vitro* cultured control experiment were
532 sequenced on an Illumina NextSeq with paired-end 75 base pair reads. FASTQ files and raw *Bb*
533 read counts for *in vitro* control experiment (GSE217146) and *ex vivo* experiment (GSE216261)
534 have been deposited in NCBI's Gene Expression Omnibus (Edgar et al., 2002) under SuperSeries
535 accession number GSE217236.

536 **RNA-seq data analysis**

537 To measure the success of our rRNA depletion through DASH, we used DASHit software
538 (Dynerman et al., 2020) to determine the percentage of reads that would be DASHable by our
539 guide RNAs (Ring et al., 2022). For pre-DASH data, we sequenced the input of each of our RNA-
540 seq libraries before performing DASH on a MiSeq V2 Micro (Illumina). We tested DASHability on
541 a random subset of 200,000 paired-end reads chosen by seqtk (https://github.com/lh3/seqtk) from
542 each pre- and post-DASH library.
543
544 To map our RNA-seq data to *Bb*, we wanted to optimize for mapping reads that came from the
545 many paralogous gene families found across the plasmids of the genome. We used the
546 pseudoalignment tool Salmon v1.2.1 (Patro et al., 2017), which is used to accurately map reads
547 coming from different isoforms of the same gene, for this reason. While using Salmon to map to
548 CDS sequences may improve mapping to paralogous genes, it may also have a tradeoff of
549 reduced mapping of reads that fall on the ends of genes that reside in operons. Nevertheless, all
550 samples should be similarly affected, and any undercounting should not change differential
551 expression results. Reads were first trimmed of bases with quality scores less than 20 using Trim
552 Galore v0.6.5 (https://github.com/FelixKrueger/TrimGalore). We mapped our reads to *Bb* CDS
553 sequences as a reference transcriptome: NCBI Genbank GCA_000008685.2 ASM868v2 (with
554 plasmids lp5, cp9, and lp56 removed as they are not present in B31-S9) using Salmon with the
555 following parameters: --validateMappings --seqBias --gcBias. Before mapping, the transcriptome

556 was first indexed using the Salmon index command with the whole genome as decoys and the
557 parameter --keepDuplicates to keep all duplicate genes.
558
559 Read counts from Salmon were used as input into DESeq2 v1.24.0 (Love et al., 2014) for
560 differential expression analysis in R version 3.6.1. DESeq2 function PlotPCA() was used to create
561 a PCA plot from read counts after running the varianceStabilizingTransformation() function. For
562 differential expression analysis between days, a DESeq object was created from count data using
563 the DESeq() function. The lfcShrink() function with the apeglm method (Zhu et al., 2018) was
564 used to calculate fold changes between days. Code used for differential expression analysis is
565 available at https://github.com/annesapiro/Bb-tick-feeding.
566
567 **Gene classification**
568 To classify genes into functional groups, functional categories were sourced from Drecktrah et
569 al., 2015 where available. Other gene functions were sourced from Fraser et al., 1997. Genes
570 found within the co-transcribed "late" bacteriophage operon (Zhang and Marconi, 2005) were
571 considered "bacteriophage" even if their function is unknown. Outer surface lipoproteins were
572 those found in Dowdell et al., 2017 plus additional lipoproteins listed in Iyer et al., 2015 that were
573 found in Dowdell et al. 2017 categories SpII and SpI and evidence of outer surface localization.
574 Outer surface and periplasmic lipoproteins were classified as "cell envelope" in the absence of
575 other classifications. Gene family information from Casjens et al., 2000 was considered to aid in
576 classification.
577
578 **RT-qPCR measuring *Bb* enrichment**
579 To test the efficacy of our enrichment protocol, we used RT-qPCR to quantify *Bb flaB* and *I.*
580 *scapularis gapdh* transcript levels in enriched and depleted fractions. We synthesized cDNA from
581 8 µL of RNA extracted from *Bb* enrichment samples and their matched depleted samples from
582 feeding day 2, day 3, and day 4 using the qScript cDNA Synthesis Kit (Quantabio). cDNA was
583 diluted 2X before use in qPCR. To measure *flaB* copies, we created standards of known
584 concentration from purified PCR products. These standards were made from PCR with primers
585 with the following sequences: 5'-CACATATTCAGATGCAGACAGAGGTTCTA and 5'-
586 GAAGGTGCTGTAGCAGGTGCTGGCTGT. We ran a dilution series with ten-fold dilutions
587 between $10^6$ copies and $10^1$ copies of this PCR template alongside our enriched and depleted
588 samples. We performed qPCR using Taqman Universal PCR Master Mix (Applied Biosystems).
589 The primers used to amplify *flaB* were: 5'- TCTTTTCTCTGGTGAGGGAGCT and 5'-

590 TCCTTCCTGTTGAACACCCTCT (used at 900 nM) and the probe was /56-
591 FAM/AAACTGCTCAGGCTGCACCGGTTC/36-TAMSp (used at 250 nM). For tick *gapdh* RT-
592 qPCR, the cDNA samples were diluted an additional 2X. Standards of known concentration were
593 created using the qPCR primer sequences. Primer sequences were: 5'-
594 TTCATTGGAGACACCCACAG and 5'-CGTTGTCGTACCACGAGATAA (used at 900 nM). qPCR
595 was performed using PowerUp SYBR Green Master Mix (Applied Biosystems). For both *flaB* and
596 *gapdh*, the number of copies in each sample was calculated based on the standards of known
597 concentration. Three technical replicates were averaged from each of four biological replicates at
598 each time point tested. We totaled the number of copies in each matched enriched and depleted
599 fraction to calculate the fraction of *flaB* or *gapdh* that was found in either sample. All qPCR was
600 performed on the QuantStudio3 Real-Time PCR System (Applied Biosystems).
601
602 **Immunofluorescence microscopy**
603 To test whether the anti-*Bb* antibody recognized *Bb* inside of the tick, ticks at each day of feeding
604 were crushed in 50 µL of PBS. 10 µL of lysate was spotted onto slides and allowed to air dry
605 before slides were heated briefly three times over a flame. Heat fixed slides were then treated
606 with acetone for one hour. Slides were incubated with anti-*Bb* primary antibody (1:100 diluted in
607 PBS + 0.75% BSA) for 30 minutes at 37°C in a humid chamber. A control without primary antibody
608 was also used for each day. Slides were washed once in PBS for 15 minutes at room temperature,
609 then rinsed in distilled water and air dried. Anti-rabbit IgG Alexa 488 (Invitrogen) diluted 1:100 in
610 PBS + 0.75% BSA was added for 30 minutes at 37°C in a humid chamber. Slides were washed
611 in PBS for 15 minutes at room temperature three times, adding 1:100 Propidium Iodide
612 (Invitrogen) during the second wash. Slides were then rinsed with distilled water and air dried
613 before the addition of mounting media (Fluoromount-G, SouthernBiotech) and cover slips.
614 Fluorescence imaging was performed on a Nikon Ti2 inverted microscope for widefield
615 epifluorescence using a 100x/1.40 objective. Images were captured with NIS-Elements AR View
616 5.20 and then processed with ImageJ software (Schneider et al., 2012). No strong florescence
617 signal was observed on the control slides without primary antibody.
618
619 **Mass spectrometry of *Bb*-enriched samples**
620 To identify which tick proteins were found in samples after *Bb* enrichment across feeding, we fed
621 both uninfected and infected ticks on mice. We collected three replicates of uninfected ticks one
622 day after attachment (11 ticks per replicate), infected ticks one day after attachment (27 ticks per
623 replicate), uninfected ticks four days after attachment (8 ticks per replicate), and infected ticks

624 four days after attachment (16 ticks per replicate). Before anti-*Bb* enrichment, the uninfected tick
625 samples were mixed with *Bb* grown in culture that was washed with PBS (3 x$10^4$ *Bb* one day after
626 attachment and 3x$10^6$ *Bb* four days after attachment) and mixed lysates were rotated at room
627 temperature for 30 minutes. Infected tick samples underwent the *Bb* enrichment process
628 immediately. The enrichment process followed the same protocol used for RNA-seq. Sample
629 volumes were increased to 1 mL as needed, and then 2 µL of anti-*Bb* antibody (Invitrogen: PA1-
630 73004; RRID: AB_1016668) was added, and samples were rotated at 4℃ for 30 minutes. 50 µL
631 of Dynabeads Protein G per sample were washed in PBS during this incubation and added to the
632 lysates, which were rotated at 4°C for 30 minutes. The beads were washed twice with 1 mL of
633 PBS, and then placed into 50 µL of lysis buffer (iST LYSE, PreOmics). Samples were boiled at
634 95°C for 5 minutes, and lysates were removed from beads and frozen for mass spectrometry
635 preparation.
636
637 For mass spectrometry, a nanoElute was attached in line to a timsTOF Pro equipped with a
638 CaptiveSpray Source (Bruker). Chromatography was conducted at 40°C through a 25 cm
639 reversed-phase C18 column (PepSep) at a constant flowrate of 0.5 µL min−1. Mobile phase A
640 was 98/2/0.1% water/MeCN/formic acid (v/v/v) and phase B was MeCN with 0.1% formic acid
641 (v/v). During a 108 min method, peptides were separated by a 3-step linear gradient (5% to 30%
642 B over 90 min, 30% to 35% B over 10 min, 35% to 95% B over 4 min) followed by a 4 min isocratic
643 flush at 95% for 4 min before washing and a return to low organic conditions. Experiments were
644 run as data-dependent acquisitions with ion mobility activated in PASEF mode. MS and MS/MS
645 spectra were collected with m/z 100 to 1700 and ions with z = +1 were excluded.
646
647 Raw data files were searched using PEAKS Online Xpro 1.6 (Bioinformatics Solutions Inc.). The
648 precursor mass error tolerance and fragment mass error tolerance were set to 20 ppm and 0.03
649 respectively. The trypsin digest mode was set to semi-specific and missed cleavages was set to
650 2. The *I. scapularis* reference proteome (Proteome ID UP000001555, taxon 6945) and *Bb*
651 reference proteome (Proteome ID UP000001807, strain ATCC 35210/B31) was downloaded from
652 Uniprot, totaling 21,774 entries. The *I. scapularis* proteome was the primary search reference and
653 the *Bb* was used as a secondary to identify any bacterial proteins present. Carbamidomethylation
654 was selected as a fixed modification. Oxidation (M) and Deamidation (NQ) was selected as a
655 variable modification.
656

657 Experiments were performed in biological triplicate, with samples being a single run on the
658 instrument. Proteins present in a database search (−10 log(p-value) ≥ 20, 1% peptide and protein
659 FDR) were subjected to the following filtration process. We first filtered to include proteins found
660 in 2 out of 3 biological replicates, within each respective day (one or four days after attachment).
661 We calculated the mean area of proteins found in uninfected and infected samples. Proteins with
662 missing values (i.e. not identified in a sample) were set to 1. We calculated the ratio of mean area
663 for each protein as infected/uninfected, and enriched proteins were identified by having a
664 infected:uninfected ratio greater than 2 within their respective feeding day (one or four days after
665 attachment).
666
667 To classify the proteins we identified into functional groups, we used a PANTHER
668 Overrepresentation Test (released 07/12/2022) (Mi et al., 2019) with PANTHER version 17.0
669 (released 02/22/2022) (Thomas et al., 2022). We used all *Ixodes scapularis* genes in the database
670 as our reference list and analyzed the lists of proteins enriched on each day for their PANTHER
671 Protein Class. The test type used was Fisher's Exact, calculating a false discovery rate as the
672 correction.
673
674 Raw data files and searched datasets are available on the Mass Spectrometry Interactive Virtual
675 Environment (MassIVE), a full member of the Proteome Xchange consortium under the identifier:
676 MSV000090560.
677
678 **SUPPLEMENTAL TABLES**
679 **Table S1.** Overview of mapping statistics from 16 *Bb* sequencing samples.
680 **Table S2.** Differential expression analysis results between *in vitro* cultured *Bb* before and after
681 *Bb* enrichment.
682 **Table S3.** Transcriptome-wide differential expression analysis results from *Bb* across tick
683 feeding timepoints.
684 **Table S4.** Twofold differentially expressed *Bb* genes from across tick feeding timepoints.
685 **Table S5.** Transcripts per million (TPM) for all *Bb* genes across feeding timepoints.
686 **Table S6.** Mass spectrometry analysis for *Bb*-enriched samples.
687 **Table S7.** Annotation and GO term enrichment for tick proteins enriched on feeding day 1.
688 **Table S8.** Annotation and GO term enrichment for tick proteins enriched on feeding day 4.
689
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## COMPETING INTERESTS

Seemay Chou is president and CEO of Arcadia Biosciences.

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