

# 1 An attempt at restoring original radiocarbon ages in collagen from bioapatite

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12

## 13 **Abstract**

14 In the literature there is a consensus that radiocarbon dating performed in bioapatite presents  
15 younger datings than those performed in collagen, thus, we propose a general regression that could  
16 be used to convert the radiocarbon dating performed in bioapatite to the original ones in collagen in  
17 fossil samples all of the world. This general regression presents several good indexes of quality,  
18 high correlation ( $R^2 = 0.98$ ), lower values of percent predicted error (%PE = 0.01), and the standard  
19 error of the estimate (%SEE = 25), showing that is a good tool, as the predicted values are similar to  
20 those observed. Using this regression we converted the radiocarbon datings in bioapatite to collagen  
21 made for several taxa from the Brazilian Intertropical Region, and suggest that these datings could  
22 be 1-7 Cal BP kyr older than previously thought.

23

24 **Keywords.** Reduced major axis regression; late Quaternary; <sup>14</sup>C AMS; mammals; Brazilian  
25 Intertropical Region.

26

## 27 **1. Introduction**

28

29 For late Quaternary researchers, known the age of their samples is very important to help  
30 them in interpretations about, for example, the paleoecology and extinction of the studied taxa (e.g.  
31 Barnosky & Lindsay, 2010; Dantas et al., 2020).

32 These researchers have as options for direct dating the use of radiocarbon dating (AMS) in  
33 collagen (<sup>14</sup>C<sub>collagen</sub>), which could date samples until ~60 kyr (Cook & Plicht, 2007). However, in  
34 Tropical regions they face one main problem, the lost of collagen due the diagenetic process  
35 (Hedges, 2002).

36 In the absence of collagen Cherkinsky (2009) presented an option in the absence of collagen,  
37 to perform the radiocarbon dating in bioapatite ( $^{14}\text{C}_{\text{bioapatite}}$ ), justifying that the mineral fraction  
38 survives much better than organic ones, suffering small changes through the diagenesis.

39 Since then, several papers dealing about the chronology and paleoecology of the meso-  
40 megamammals from the Brazilian Intertropical Region has been made using this technique, and  
41 presenting the occurrence of this fauna in the Late Pleistocene, between 9-32 Cal BP Kyr (e.g.  
42 Dantas et al., 2017; 2020; 2022).

43 However, Some authors (Zazzo & Saliège, 2011; Zazzo, 2014) suggests that during  
44 diagenesis bioapatite exchange carbon with a  $^{14}\text{C}$ -enriched (i.e. younger) carbon source, which  
45 promote in  $^{14}\text{C}_{\text{bioapatite}}$  younger datings than  $^{14}\text{C}_{\text{collagen}}$ , as older the dating, major was the difference  
46 between them. Thus, Zazzo (2014) recommended that  $^{14}\text{C}_{\text{bioapatite}}$  should be considered as minimum  
47 estimates.

48 Based on this observation, we propose, and test, regressions that could convert the  
49 radiocarbon dating in bioapatite to collagen in samples collected in different climatic zones (boreal,  
50 temperate, subtropical, and tropical).

51

## 52 **2. Material and methods**

53

54 In this paper we use several radiocarbon dating performed in collagen and bioapatite in the  
55 same samples, the collagen radiocarbon datings had expected C/N pattern ( $\sim 3$ ) and more than 5% of  
56 collagen in each samples. Another index was the presence of modern carbon (pMC) in the samples,  
57 were used samples with proportional lower pMC as older the samples (Cherkinsky, 2009; Zazzo,  
58 2014 and references therein; Cherkinsky et al, 2015; Table S1).

59 Reduced major axis (RMA, Model II) regressions were produced using the entire sample to  
60 create a general regression, and specific ones for each climatic zone (boreal, temperate, subtropical,  
61 and tropical; Table 1), because: (i) it deals better with extrapolation than ordinary least squares; (ii)  
62 incorporate an assumption that there is an error in  $X$ ; and (iii) is symmetric, meaning that the slope  
63 of the line do not differs depending upon which variable is identified as  $X$  and which is  $Y$  (OLS,  
64 Model I; Smith, 2009; Halenar, 2011 and references therein). This method use the slope ( $b_{\text{OLS}}$ ) find  
65 in OLS, the mean values of  $x$  and  $y$ , and the absolute value of the correlation of pearson ( $r$ ) to  
66 estimate a new slope ( $b_{\text{RMA}}$ ; equation 1) and intercept ( $a_{\text{RMA}}$ ; equation 2) (Harper, 2016).

67

$$68 \quad (1) b_{\text{RMA}} = b_{\text{OLS}}/|r|$$

$$69 \quad (2) a_{\text{RMA}} = \bar{Y} - b_{\text{RMA}} * \bar{X}$$

70

71 As the radiocarbon datings do not presented a normal distribution (Shapiro-Wilk test,  $p <$   
72 0.05), these data were transformed to logarithm values (at base 10) to approximate a log-normal  
73 distribution, due it assigns equal weight to all data points in a regression (e.g. Smith, 1993 and  
74 references therein).

75 In addition to the correlation of data log-transformed, as high correlation do not means that  
76 the regression is a good predictor (e.g. Smith, 1984), were calculated the percent predicted error  
77 (%PE) and the standard error of the estimate (%SEE).

78 The %PE of each sample is calculated using equation (3) (Van Valkenburgh, 1990 and  
79 references therein; Halenar, 2011), and then is made an average of the absolute %PE mean of the  
80 variables. This index provides a comparative value to see the predictive accuracy of the regressions.

81  
82 
$$(3) \%PE = (\text{observed} - \text{predicted}/\text{predicted}) * 100$$
  
83

84 To estimate the %SEE we use the equation (4), this index reflects the ability of the  
85 independent variable to predict the dependent variable (Van Valkenburgh, 1990 and references  
86 therein). SE is the standard error (= standard deviation/ $\sqrt{n}$ ).

87  
88 
$$(4) \%SEE = (10^{(2+SE)}) - 100$$
  
89

90 To test if are statistical differences between the proposed regressions was made by the  
91 analysis of variance, we used ANOVA (1 factor,  $\alpha = 0.05$ ) in PAST 3.11 software (Hammer et al.,  
92 2001).

93 The best estimated regression (results and discussion) was used to correct the radiocarbon  
94 dating in bioapatite to collagen of eight extinct meso- megamammals from the Brazilian  
95 Intertropical Region (BIR; sensu Cartelle et al., 1999; Table 2).

96  
97 **3. Results and discussion**

98  
99 *3.1. Converting  $^{14}C_{\text{bioapatite}}$  into  $^{14}C_{\text{collagen}}$*   
100

101 The radiocarbon dating samples (dated both in bioapatite and collagen) came from different  
102 countries located in boreal, temperate, subtropical, and tropical climatic zones (Table S1), in all  
103 those were observed that the diagenesis altered the bioapatite and provide, in general, younger  
104 radiocarbon dating in comparison with those found in collagen (Cherkinsky, 2009; Zazzo, 2014 and  
105 references therein; Cherkinsky et al, 2015).

106 Using these data were estimated regressions for each climatic zone, plus, a general one, and  
107 noted that they are similar (ANOVA,  $F_{obs} = 0.02$ ,  $p = 0.99$ ; Table 1), presenting strong correlations  
108 and similar slopes (m) values, however, presents different %PE and %SEE.

109 The slopes of these RMA regressions, created with the available data, allow us to interpret  
110 that the radiocarbon dating in bioapatite tend to be slightly lower than that in collagen in Boreal (m  
111 = 1.10), Temperate climate zones (m = 1.15), and in all world (m = 1.09). In Subtropical climate  
112 zones (m = 0.99) and Tropical climate zones the radiocarbon dating in bioapatite tend to be slightly  
113 higher than that in collagen (m = 0.97).

114 If was choose to use the regressions for each climatic zone there is a tendency to find different  
115 corrected collagen datings, being that for Temperate climatic zones higher than in the others zones.  
116 To avoid this, as all regressions are similar, the general regression must be used (Figure 1), as it  
117 presents a strong correlation ( $R^2 = 0.98$ ), lower mean %PE (= 0.01; Table 1), and average %SEE (=   
118 25.00; Table 1), combined.

119

$$\text{Log}_{10}^{14}\text{C}_{\text{collagen}} = 1.09 * \text{log}_{10}^{14}\text{C}_{\text{bioapatite}} - 0.31$$

120

121

122 The best regressions must have higher values of correlation, lower values of %PE (<15%) and  
123 %SEE (Delson, et al., 2000; Ruff, 2003), showing that the predicted values are similar to those  
124 observed, which this general equation reached.

125

### 126 3.2. Limit of conversion

127

128 The radiocarbon dating calibration curve could allow estimating the age of terrestrial samples  
129 to about 50 kyr, the limit of the method (Cook & Plicht, 2007; Wood, 2015). This limit depends on  
130 the pretreatment used, which could help to better purify the samples from contaminants, and allows  
131 older datings (Wood, 2015).

132 As stated before the radiocarbon dating in bioapatite is considered as minimum ages, and the  
133 proposed general regression can convert the  $^{14}\text{C}_{\text{bioapatite}}$  to  $^{14}\text{C}_{\text{collagen}}$ , however observing the limit of  
134 the method (50 kyr) this regression should be used to convert only  $^{14}\text{C}_{\text{bioapatite}} \sim 39,400$  yr. Older  
135 converted collagen dating could be not calibrated in CALIB 8.1 program (Reimer et al., 2020) due  
136 to the extrapolation of the limit of 50 kyr.

137

### 138 3.3. Study case: converting the $^{14}\text{C}_{\text{bioapatite}}$ of the meso- megamammals from the Brazilian 139 Intertropical Region

140

141 Using the new general regression, were converted the radiocarbon datings made in bioapatite  
142 to the collagen for eight extinct meso- megamammals taxa which lived in the BIR, and later  
143 calibrated into calendar ages before present, using the same standard error found in the  $^{14}\text{C}_{\text{bioapatite}}$ ,  
144 using CALIB 8.1 program (Reimer et al., 2020), SHCal20 curve (Hogg et al., 2020), and  $2\sigma$   
145 measured ages reported in Table 2.

146 The difference between the radiocarbon dating in bioapatite to the converted to collagen  
147 shows a variation between 1,141 to 7,187 years (Table 1), while the difference between the  
148 calibrated datings was 1,166 to 7,523 Cal BP yr older than previously thought (e.g. Cherkinsky et  
149 al., 2013; Dantas et al., 2017; Figure 2).

150 The diagenesis could promote small alterations in  $^{14}\text{C}/^{12}\text{C}$  in bioapatite carbonate, leading to  
151 younger datings, however, this alteration is non-significant in ratio stable isotopes of carbon  
152 ( $^{13}\text{C}/^{12}\text{C}$ ), at least, for the last 40 thousand years (Zazzo, 2014).

153 When the diagenesis affect the bioapatite, the substitutions are mainly in the hydroxyl  
154 position in the phosphate, and even with a carbonate substitution occurs, the isotope signature in  
155 stable and radioactive carbon maintain the original signature (Cherkinsky, 2009).

156 The available  $\delta^{13}\text{C}$  associated to the converted  $^{14}\text{C}_{\text{collagen}}$  for the megafauna of the BIR brings  
157 paleoecological information of a time span ranging ~12,700 to 42,100 years (Figure 2), and allow  
158 suggesting that these meso- megamammals lived in the BIR, at least, until 12 kyr, in the Late  
159 Pleistocene. Considering other dating techniques, as for example Electron Spin Resonance, this  
160 time span could be expanded to  $9\pm 2$  ky (Ribeiro et al., 2013).

161

#### 162 **4. Final remarks**

163

164 In this paper was proposed a regression to convert the radiocarbon dating performed in  
165 bioapatite to collagen, allowing facilitating the comparison of radiocarbon datings in all world.

166 Using this new tool were converted the radiocarbon dating performed in bioapatite in fossils  
167 of meso- megamammals from Brazil and suggest that these datings are 1-7 Cal BP kyr older than  
168 previously thought.

169

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171

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174

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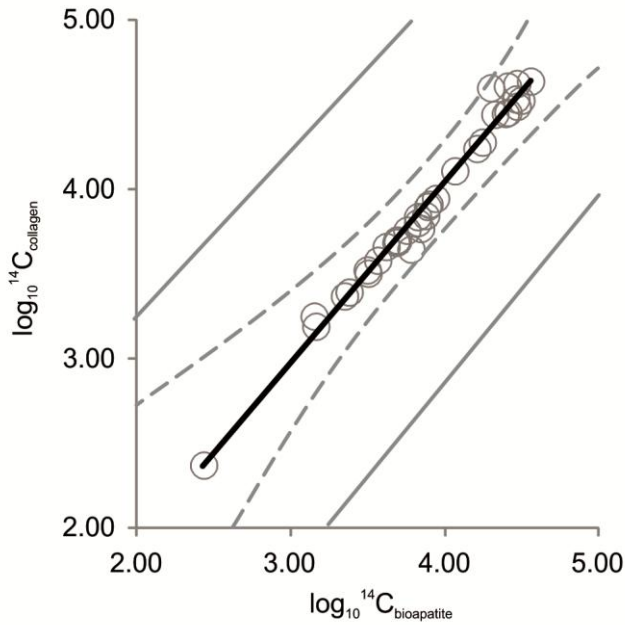
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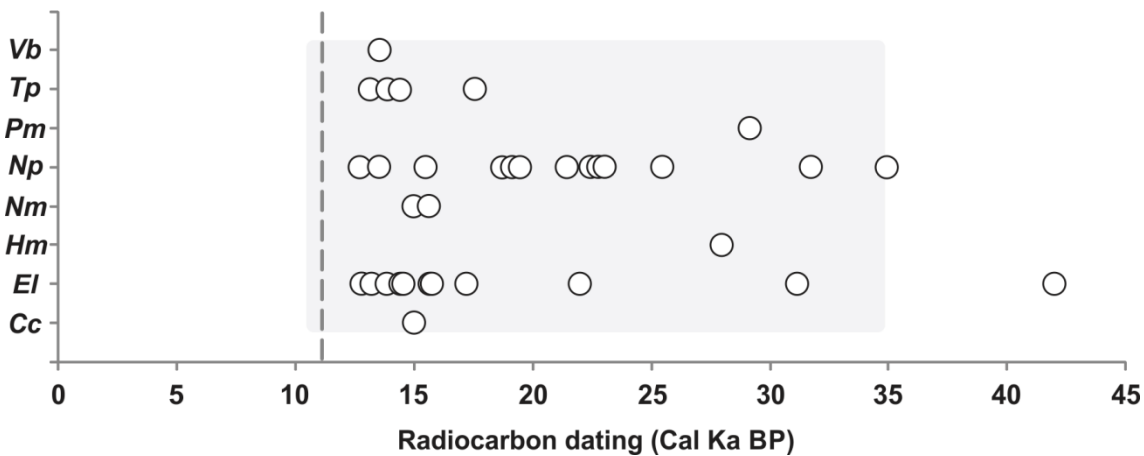
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244

245 **Figure 1.** Reduced major axis Regression of log Radiocarbon dating (bioapatite) and log  
 246 Radiocarbon dating (collagen) using 28 samples (Table S1). Regression line (Black solid line),  
 247 confidence intervals (Gray dotted lines), and prediction intervals (Gray solid lines).

248



249

250 **Figure 2.** Radiocarbon Chronology in collagen (white circles) of extinct meso- megamammals from  
 251 the Brazilian intertropical region. Gray shadow represents the interval found in Radiocarbon dating  
 252 performed in bioapatite. Doted Gray line represents the limit between Pleistocene-Holocene. **labels:**  
 253 *Cc* - *Catonyx cuvieri*; *El* - *Eremotherium laurillardi*; *Hm* - *Hemiauchenia mirim*; *Nm* -  
 254 *Nothrotherium maquinense*; *Np* - *Notiomastodon platensis*; *Pm* - *Palaeolama major*; *Tp* - *Toxodon*  
 255 *platensis*; *Vb* - *Valgipes bucklandi*.

256



257 **Table 1.** Values of the RMA regressions, coefficient of determination ( $R^2$ ), average percent  
258 prediction error (%PE) and standard error of the estimate (%SEE) obtained for each climatic zone  
259 (cz).

<b>cz</b>	<b>Samples</b>	<b>Slope</b>	<b>Intercept</b>	<b><math>R^2</math></b>	<b>%PE</b>	<b>%SEE</b>
Boreal	8	1.10	-0.41	0.97	0.02	10.90
Temperate	7	1.15	-0.50	0.98	0.05	89.32
Subtropical	8	0.99	0.05	0.99	0.06	21.26
Tropical	5	0.97	0.13	0.99	0.00	25.16
All	28	1.09	-0.31	0.98	0.01	25.00

260

261 **Table 2.** Radiocarbon datings in bioapatite ( $^{14}\text{C}_{\text{bioapatite}}$ ) converted to collagen ( $^{14}\text{C}_{\text{collagen}}$ ), presence of modern carbon (pMC), and calibrated ages  
 262 (SHCal20 curve) for extinct Late Pleistocene meso- megamammals taxa from Brazilian Intertropical Region.

Species	Sample number	Localities	$^{14}\text{C}_{\text{bioapatite}}$	pMC	$^{14}\text{C}_{\text{collagen}}$	$^{14}\Delta_{\text{bioapatite-collagen}}$	Age (Cal BP yr)
<i>C. cuvieri</i>	UGAMS 34121 <sup>(01)</sup>	Andaraí/BA	11,150±30	24.97	12634±30	1,484	14,839-15,165
<i>E. laurillardi</i>	UGAMS 09436 <sup>(01)</sup>	Barcelona/RN	10,050±35	28.62	11,281±35	1,231	13,096-13,192
	UGAMS 09435 <sup>(01)</sup>	Currais Novos/RN	15,490±40	-	18,078±40	2,588	21,851-22,115
	UGAMS 09431 <sup>(01)</sup>	Poço Redondo/SE	10,140±40	24.98	11,392±40	1,252	13,170-13,315
	UGAMS 09432 <sup>(01)</sup>	Poço Redondo/SE	22,440±50	-	27,078±50	4,638	31,064-31,206
	UGAMS 09433 <sup>(01)</sup>	Poço Redondo/SE	11,540±40	23.78	13,116±40	1,576	15,517-15,841
	UGAMS 13539 <sup>(01)</sup>	Poço Redondo/SE	10,990±30	25.45	12,436±30	1,446	14,250-14,644
	UGAMS 13540 <sup>(01)</sup>	Poço Redondo/SE	11,010±30	25.39	12,461±30	1,451	14,288-14,878
	UGAMS 13541 <sup>(01)</sup>	Poço Redondo/SE	9,720±30	29.82	10,878±30	1,158	12,736-12,786
	UGAMS 13542 <sup>(01)</sup>	Poço Redondo/SE	9,730±30	29.79	10,890±30	1,160	12,738-12,792
	UGAMS 13543 <sup>(01)</sup>	Poço Redondo/SE	11,580±30	23.65	13,166±30	1,586	15,610-15,898
	UGAMS 14017 <sup>(01)</sup>	Poço Redondo/SE	10,740±30	26.25	12,128±30	1,388	13,810-13,950
	UGAMS 09434 <sup>(01)</sup>	Gararu/SE	11,540±40	23.78	13,116±40	1,576	15,517-15,841
	UGAMS 42447 <sup>(02)</sup>	Ourolândia/BA	12,400±30	-	14,185±30	1,785	17,072-17,352
	UGAMS 34119 <sup>(02)</sup>	Iuiu/BA	30,080±90	2.37	37,267±90	7,187	41,839-42,141
<i>H. mirim</i>	UGAMS 36483 <sup>(03)</sup>	Campo Formoso/BA	20,010±65	8.28	23,898±65	3,888	27,764-28,177
<i>N. maquinense</i>	UGAMS 34123 <sup>(01)</sup>	Andaraí/BA	11,130±30	25.02	12,609±30	1,479	14,819-15,133
	UGAMS 34124 <sup>(01)</sup>	Andaraí/BA	11,520±35	23.84	13,091±35	1,571	15,483-15,801

*N. platensis* UGAMS 09440<sup>(01)</sup> Barcelona/RN 16,150±40 - 18,919 2,769 22,594-22,974

263 **References.** <sup>(1)</sup> Dantas et al. (2017 and references therein); <sup>(2)</sup> Dantas et al. (2020 and references therein); <sup>(3)</sup> Greco et al. (2022).

264 **Table 2 (continuation).**

Species	Sample number	Localities	<sup>14</sup> C <sub>bioapatite</sub>	pMC	<sup>14</sup> C <sub>collagen</sub>	<sup>14</sup> Δ <sub>bioapatite-collagen</sub>	Age (Cal BP yr)
<i>N. platensis</i>	UGAMS 09437 <sup>(01)</sup>	Poço Redondo/SE	13,950±40	17.61	16,128±40	2,178	19,261-19,544
	UGAMS 13535 <sup>(01)</sup>	Poço Redondo/SE	13,380±35	18.89	15,411±35	2,031	18,630-18,813
	UGAMS 13536 <sup>(01)</sup>	Poço Redondo/SE	16,370±40	13.03	19,201±40	2,831	22,947-23,164
	UGAMS 13537 <sup>(01)</sup>	Poço Redondo/SE	10,440±30	27.26	11,759±30	1,319	13,485-13,613
	UGAMS 13538 <sup>(01)</sup>	Poço Redondo/SE	13,760±35	18.04	15,889±35	2,129	18,966-19,252
	UGAMS 09439 <sup>(01)</sup>	Canhoba/SE	17,910±50	-	21,177±50	3,267	25,277-25,672
	UGAMS 09438 <sup>(01)</sup>	Coronel João Sá/BA	13,980±40	-	16,166±40	2,186	19,345-19,583
	UGAMS 09441 <sup>(01)</sup>	Coronel João Sá/BA	15,210±40	15.06	17,722±40	2,512	21,173-21,736
	UGAMS 34140 <sup>(04)</sup>	Coronel João Sá/BA	9,640±30	30.13	10,781±30	1,141	12,700-12,746
	UGAMS 42448 <sup>(02)</sup>	Ourolândia/BA	25,070±60	-	30,555±60	5,485	34,628-35,209
	UGAMS 39057 <sup>(05)</sup>	Caetanos/BA	11,450±30	24.05	13,005±30	1,555	15,327-15,662
	UGAMS 34116 <sup>(02)</sup>	Iuiu/BA	23,040±55	5.68	27,868±55	4,828	31,562-31,911
	UGAMS 34125 <sup>(04)</sup>	Vit. da Conquista/BA	15,890±40	13.83	18,588±40	2,698	22,346-22,535
<i>P. major</i>	LPRBUSP 0755 <sup>(01)</sup>	Iraquara/BA	20,850±50	-	24,993±50	4,143	29,077-29,248
<i>T. platensis</i>	UGAMS 09442 <sup>(01)</sup>	Rui Barbosa/RN	10,730±30	-	12,116±30	1,386	13,809-13,954
	UGAMS 09446 <sup>(01)</sup>	Poço Redondo/SE	10,050±30	-	11,281±30	1,231	13,098-13,188
	UGAMS 09444 <sup>(01)</sup>	Cel. João Sá/BA	12,580±40	20.88	14,410±40	1,830	17,340-17,789

	UGAMS 42449 <sup>(02)</sup>	Ourolândia/BA	10,740±30	-	12,128±30	1,388	13,810-13,950
	UGAMS 09445 <sup>(01)</sup>	Vit. da Conquista/BA	10,970±30	-	12,412±30	1,442	14,189-14,595
<i>V. bucklandi</i>	UGAMS 11763 <sup>(01)</sup>	Felipe Guerra/RN	10,440±35	27.25	11,759±35	1,319	13,482-13,617

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**References.** <sup>(1)</sup> Dantas et al. (2017 and references therein); <sup>(2)</sup> Dantas et al. (2020 and references therein); <sup>(4)</sup> Dantas et al. (2022); <sup>(5)</sup> Lessa et al. (2021)

265

266