Title

Fish Aggregating Devices could enhance the effectiveness of blue water MPAs

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ABSTRACT

In the past two decades, drifting fish aggregation devices (FADs) have revolutionised pelagic fisheries, and are now responsible for the majority of tuna purse seine harvests. Here, we argue that by taking advantage of the same proven aggregative properties, FADs could be used to enhance the benefits provided by blue water Marine Protected Areas (MPAs). Using models of harvested fish populations, we explore the potential benefits that could be achieved if semi-stationary conservation FADs were positioned within blue water MPAs. Our results suggest that conservation FADs could deliver benefits, both to target species and the broader ecosystem. By increasing the residence time of exploited species, conservation FADs will reduce average mortality rates inside MPAs. By increasing the local density of species whose populations are depressed by exploitation, FADs can also improve the function of ecosystems in blue water MPAs. Conservation FADs could therefore amplify the benefits of blue water MPAs. We find this amplification is largest in those contexts where blue water MPAs have attracted the most criticism - when their area is small compared to both the open ocean and the distribution of fish stocks that move through them.

INTRODUCTION

The open-ocean is under unprecedented threat from human activities. The creation of large, blue water marine protected areas (MPAs; Figure 1) has been an important part of the response to this threat (1). These blue water MPAs primarily protect open-ocean, pelagic ecosystems, and are often centred upon oceanic islands, reefs, or seamounts. The number and extent of blue water MPAs has accelerated rapidly in recent years and is likely to continue to accelerate in support of international agreements such as the Convention on Biological Diversity's post-2020 Global Biodiversity Framework.

The anticipated ecological benefits of blue water MPAs are uncertain for two reasons. First, despite being among the world's largest protected areas, blue water MPAs still only cover a tiny fraction (<5%) of the ocean's vast area (2). Their ecological benefits may be commensurately small. Second, a substantial proportion of pelagic biodiversity is highly mobile or migratory and can easily move across reserve boundaries. This is especially true for species targeted by commercial fisheries: including tuna and swordfish (3,4), whose migration patterns may encompass whole oceans. Even the largest blue water MPAs could not protect individuals of these species from harvest mortality throughout their lives, a key design objective of coastal and

reef MPAs (5). As a consequence, the ability of blue water MPAs to conserve pelagic species and ecosystems is a question of intense debate (6–10).

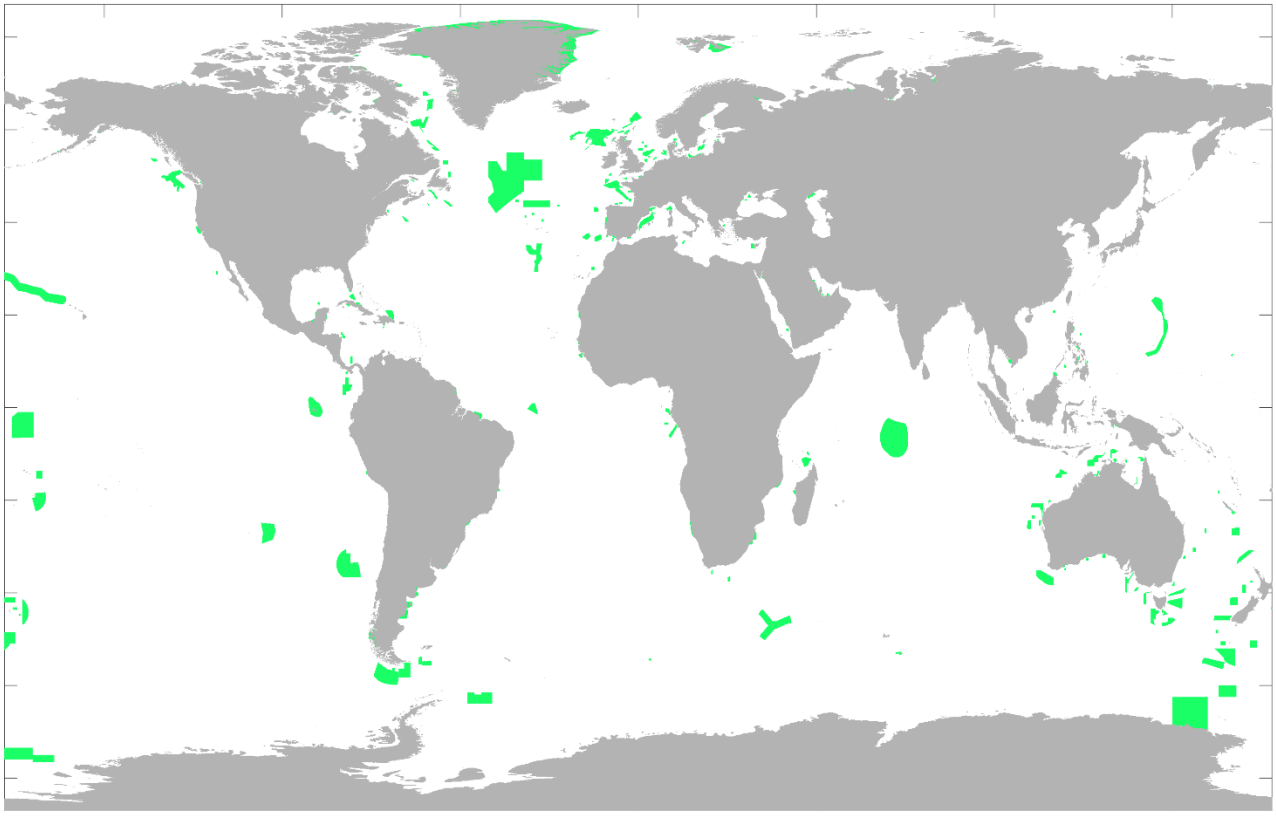

Figure 1: Distribution of the world's blue water MPAs. Data on MPA boundaries are sourced from the World Database of Protected Areas (11).

Drifting Fish Aggregation Devices (dFADs) are floating objects deployed by fishers, to exploit the natural propensity of many pelagic species to congregate under such objects. By increasing the local density and consistency of target species, dFADs increase fishers' expected catch-per-unit-effort (12). Although dFADs attract a variety of pelagic species, their effectiveness at aggregating commercially important tropical tuna species has made them the mainstay of the world's purse seine tuna fisheries (13). Drifting FADs increase local pelagic biomass density, and are increasingly fitted with echosounder-equipped, satellite-linked buoys. These provide real-time information on the biomass around them, greatly increasing the efficiency of fisheries production (14,15). The number of active dFADs deployed in the world's oceans is notoriously difficult to estimate, but likely exceeds 100,000 per year, and is increasing (16). A number of possible negative ecological impacts have been suggested as a result of extensive dFAD use, including bycatch of aggregating species such as silky sharks (17), detrimental effects on tuna behaviour (18), and damage caused by abandoned dFADs

colliding with vulnerable habitat such as coral reefs (19). Drifting FADs may also pose a challenge for the effectiveness of blue water MPAs because they cross into and out of the protected areas, taking apex pelagic species with them (20).

FADs also have potential uses in marine conservation. For more than 20 years, anchored FADs (aFADs) have been deployed near to fishing communities by development and fisheries organizations, to redistribute fishing pressure from coastal ecosystems to pelagic resources. Here, we propose that by deploying stationary FADs inside blue water MPAs, conservation organisations could directly and positively enhance their benefits. For the purpose of this paper, we refer to these as conservation FADs (cFADs), although we acknowledge that this term could also describe the anchored fishing FADs referred to above.

cFADs could deliver benefits to species that experience direct fishing mortality, and also to the broader ecosystem. By increasing the residence time of highly-mobile and migratory species inside the protected area, cFADs decrease the exposure of these species to fishing mortality. Despite the temporary nature of this protection, it could nevertheless enhance stock levels, via the same mechanism as temporary closures. An increased residence time could deliver density-dependent ecosystem benefits, which are lost when harvesting reduces the abundance of target or bycatch species. For instance, bird species often rely on high tuna densities for effective foraging (21,22).

The deployment of cFADs inside blue water MPAs could therefore help to counterbalance the extensive use of dFADs by fisheries in the open ocean, allowing MPAs to enhance their benefit without increasing their size. However, as far as we are aware, the potential for cFADs to amplify the benefits of blue water MPAs has not previously been explored, theoretically or empirically. The goal of this study is to explore the logic of this proposal, by introducing cFADs to standard theoretical models of exploited fish species with MPAs.

NONSPATIAL MODEL

Nonspatial model

The first model describes the effects of cFADs as simple, spatially-implicit redistributions of stock numbers. Following ideas from optimal foraging theory (23), we assume that each FAD attracts fish by offering them

greater benefits than the surrounding seascape. As a consequence, all FADs simply concentrate existing animals from the surrounding seascape.

A stock of fish has population X, which is distributed across an area N, of which an area M has been placed into a blue water MPA. The area contains f_F fishing dFADs, which move at random across the entire stock distribution (including some time spent inside the MPA). The area also contains f_C stationary cFADs, all of which are permanently inside the MPA.

Fish gain some density-dependent benefit from being in proximity to any FAD, and the population therefore aggregates in increasing density at each FAD until the marginal benefit of joining a FAD aggregation falls to zero (24). At this point, some of the fish population (denoted $x \ll X$) will be found in the vicinity of each FAD (of either type), and the remaining proportion will be distributed evenly across the stock distribution area N. In the absence of any FADs, the time-averaged number of fish that will be found inside the MPA will then be:

$$p(f_F = 0, f_C = 0) = X\frac{M}{N}$$

Equation 1

Equation 1 represents the essential role of an MPA – to protect a proportion of the fish stock distribution from mortality. It is also the basis of the primary critique of blue water MPAs: that their size is small relative to the area of a mobile pelagic stock (i.e., $M \ll N$).

If cFADs and dFADs are present, then the time-averaged proportion of the stock that will be found inside the MPAs (and therefore protected from excess fishing mortality) is:

$$p(f_F, f_C) = \frac{M}{N}\left(X - x(f_F + f_C)\right) + x\left(f_c + f_F\frac{M}{N}\right)$$

Equation 2

The first term on the right-hand side of Equation 2 represents fish that are not associated with any FAD and are protected by the MPA, while the second term represents those fish that are associated with any FAD, and which are currently inside the MPA. We also assume that $x \leq X/(f_F + f_C)$, to ensure non-negative fish populations.

Thus, the presence of f_C conservation cFADs amplify the stock benefits flowing from an MPA by a factor A:

$$A = \frac{p(f_F, f_C)}{p(f_F = 0, f_C = 0)} = 1 + \frac{x}{X}\left(\frac{N}{M} - 1\right) f_C.$$

Equation 3

We parameterised the nonspatial model with area $N = 1$, for an MPA that makes up 1%, 2%, and 5% of the stock distribution, and which contains a normalised stock abundance of $X = 1$. We allow the number of FADs to range between $50 \leq f_F \leq 500$ for fishing FADs, and $0 \leq f_c \leq 10$ for conservation FADs. A proportion $x/X = 10^{-3}$ (i.e., 0.1%) of the total stock abundance are found in the vicinity of each FAD (of either type).

Nonspatial model results

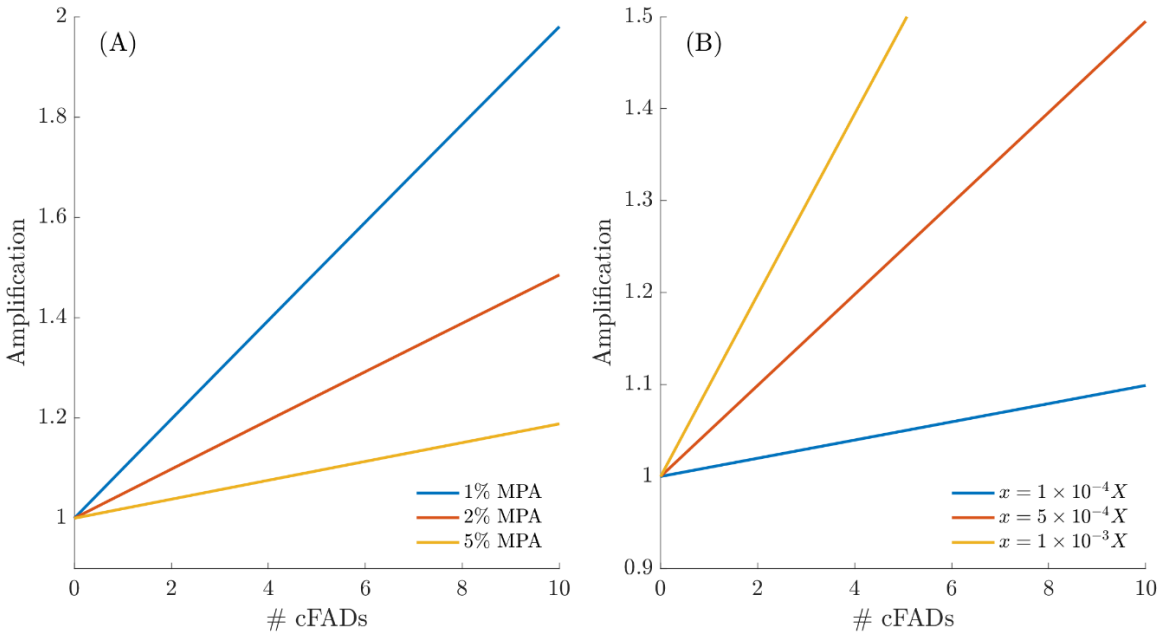

Figure 2: Nonspatial model results. Panel (A) shows the amplification provided to blue water MPAs of three different sizes, as the number of cFADs increases from zero to 10. Panel (B) shows the amplification provided when FADs attract more fish, as the number of cFADs increases from zero to 10. Note the different y-axis limits.

Figure 2 shows that the amplification factor A is an inverse function of MPA size – highest for smaller MPAs (Figure 2A). Small MPAs offer the least protection to fish stocks (Equation 1), and their smaller benefits are therefore easy to amplify. If a particular number of cFADs create an amplification factor of $A = 2$, for example,

then they have effectively doubled the density of targeted fish, halved the exposure of each fish to excess harvest mortality, and doubled the effective size of the MPA, when compared to an MPA without any cFADs present. Figure 3 provides a visual illustration of how cFADs amplify the effective size of a blue water MPA.

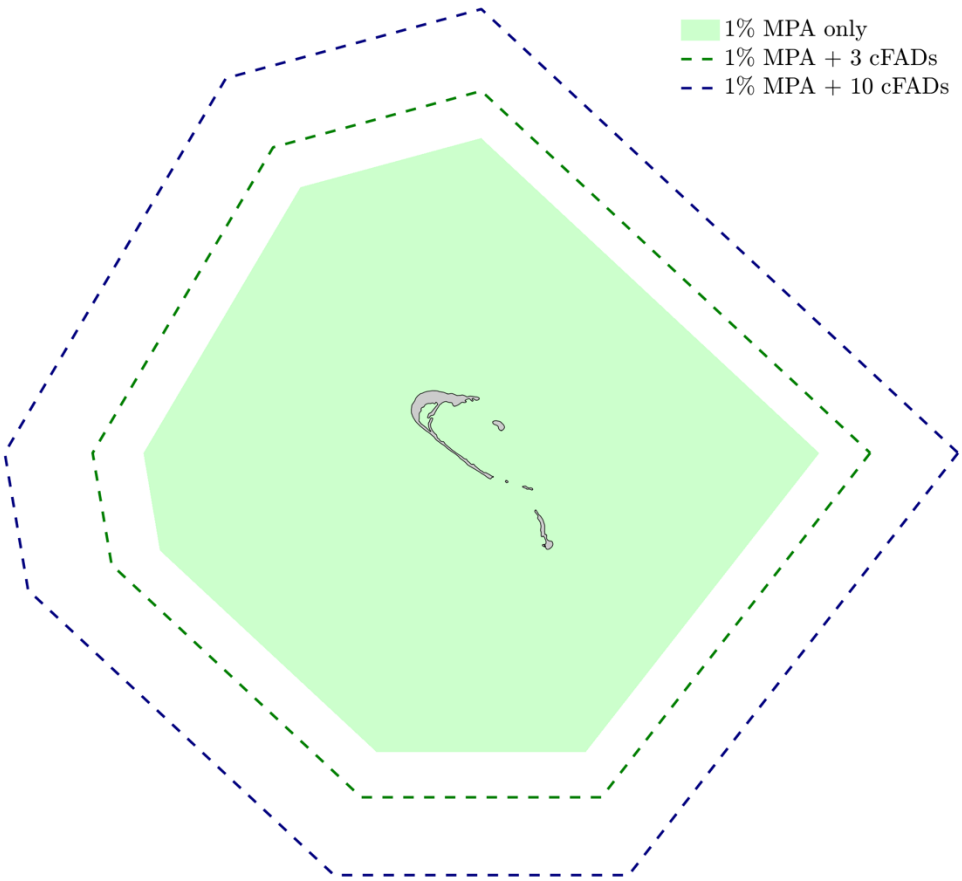

Figure 3: Visual illustration of the benefits provided by cFADs to a hypothetical blue water MPA around a coral atoll. The green region shows a hypothetical MPA surrounding an island (grey outline). With 3 cFADs, inside a 1% MPA, the area will contain as many fish as an MPA that is 30% larger (area enclosed by green dashed line). With 10 cFADs, the MPA will contain as many fish as an MPA that is 100% larger (area enclosed by blue dashed line). The stock distribution area is much larger than domain shown here. Results are based on our nonspatial model in Figure 2A.

The model produces two counter-intuitive results. First, since dFADs are far more numerous, we might therefore expect that more effective FADs would benefit fishers. Instead, we find that amplification is larger when FADs attract more fish (Figure 2B). Second, we might expect that larger numbers of dFADs would dilute the effects of cFADs, reducing the amplification benefits they create. However, the model predicts that amplification is unaffected by the number of dFADs. Both of these results – the positive benefits of more

attractive FADs, and the insensitivity to the number of dFADs – reflect the fact that dFADs spend a proportional amount of time inside MPAs. If dFADs are distributed evenly through space, and if they attract fish from a relatively large surrounding region, then they produce positive conservation effects while they are inside an MPA.

While this model captures the attraction that FADs exert on fish population distributions, it doesn't explicitly model the interaction of FADs with MPA locations (particularly the potential for dFADs to export biomass from no-take zones as they pass through them or along boundaries), nor does it include the negative effects of fishing mortality on stock abundance, which will increase with the number of fishing FADs. To address these factors, we extend the model to include the effects of space, and the negative demographic effects of fishing.

SPATIAL MODEL

Spatial model formulation

Our spatial model is based on a previously-published model designed to measure the effects of aFADs on the movement of skipjack tuna (25). We assume a one-dimensional seascape that is divided into n discrete cells, mapped onto a circle to avoid boundary conditions. A contiguous subset of m cells in the centre of the domain are designated as a blue water no-take MPA. We assume that in f_F of the cells across the domain there is a drifting FAD, and in f_C of the cells inside the MPA there is a stationary conservation FAD.

Within the domain there is a population of fish that is expected to benefit from the cFADs and MPA. The abundance of these fish in cell i at time t is denoted $x_{i,t}$. Individuals in any cell can move left or right, and this may involve entering or exiting the MPA. They can also remain in the same cell. The relative attractiveness of each cell is defined as a_i, and we assume all cells containing FADs have a higher attractiveness, cells without FADs have a uniform, lower attractiveness, and cells containing multiple FADs have the same attractiveness as single-FAD cells. The result is an accumulation of fish in cells occupied by a cFAD or a dFAD, and a minor accumulation of fish in nearby cells.

Individuals face a constant natural mortality probability of μ, and we apply a constant excess fishery harvest mortality rate $e_{i,t}$ to all cells outside the MPA that contain dFADs in timestep t. The mortality rate increases proportionally when multiple dFADs co-occur in space. We assume that the harvest mortality elsewhere is

effectively zero. We investigated the application of dFAD fishing mortality both continuously and stochastically (i.e., according to a probability each time step), and the effects of non-dFAD associated mortality outside the MPA (i.e., additional fishery effort that does not target dFADs). Neither choice affects our results.

The number of new recruits each year is determined by a density-dependent Ricker model with rate parameter k, following harvests, natural mortality, and movement. Recruitment from the larval pool is equally redistributed across the domain, with each cell receiving r_t recruits in timestep t, where:

$$r_t = \frac{1}{n}\left(1 - \exp\left[k \sum_i (1 - e_{i,t} - \mu) x_{i,t}\right]\right)$$

We model reproductive output as a function of local densities rather than global abundance, since density-dependent processes may operate either locally or globally, but this choice does not substantially affect our outcomes. Taken together, these assumptions imply that the abundance of fish in cell i at timestep $t + 1$ is:

$$x_{i,t+1} = \left[\frac{a_i}{a_{i-1}} x_{i-1,t} + \frac{a_i}{a_{i+1}} x_{i+1,t} - x_{i,t}\left(\frac{a_{i+1}}{a_i} + \frac{a_{i-1}}{a_i}\right)\right](1 - e_{i,t} - \mu) + r_t.$$

The system does not reach a spatiotemporal equilibrium because the movement of the dFADs constantly affects the distribution of fishes. We therefore simulate the model until the 100-year time-averaged abundance of the total fish population stabilises.

For our example, we discretise the seascape into $n = 300$ sites, with an MPA that takes up $m = \{1, 3, 6\}$ contiguous sites. We model $f_F = \{25, 250, 500\}$ moving dFADs, and $f_C = \{0, \ldots, 10\}$ stationary cFADs. The dFADs are initialised at random outside the MPA, and the cFADs are placed at random locations within the MPA. At the higher densities, many locations will therefore contain more than one FAD. The relative attractiveness of each location is $a_i = 0.5$ for locations without FADs, and $a_i = 0.6$ for locations with FADs. Natural mortality is assumed to be $\mu = 0.1$, fishery harvest mortality is $e_i = 0.1$, and density-dependent Ricker mortality is governed by parameter $k = 0.1$.

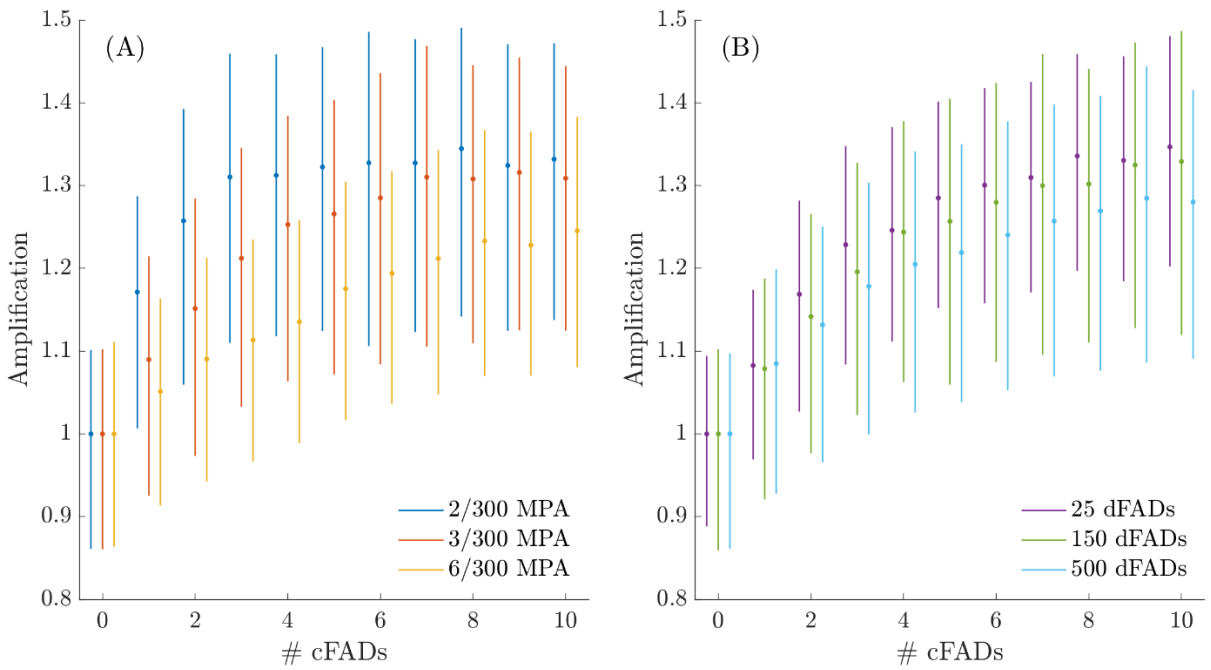

Figure 4: Spatial model results for the number of cFADs varying between 0 and 10. Panel (A) shows that the amplification benefits provided to a blue water MPA decline as the MPA increases in size. There are 100 dFADs in the system. Panel (B) shows that benefits decline as the number of dFADs increases. These results are for an MPA that encompasses 3 cells (i.e., 1% of the domain). In both panels, the bounds illustrate the interquartile range of the observed amplification across the different cells of the MPA, and across many years. That is, how the increase in density inside the MPA varies across space and through time.

Spatial model results

The spatial model results augment the nonspatial results in three important ways. First, unlike the nonspatial model, spatial amplification benefits are concave; that is, increasing the number of cFADs delivers diminishing marginal conservation returns. In the nonspatial model, each cFAD delivers independent benefits, drawing fish into the MPA from across the fished area. In the spatial model by contrast, cFADs can only attract fish from the surrounding seascape. As their numbers increase, these local areas begin to overlap, the cFADs begin to compete for fish density. Such a saturation effect has been observed for skipjack tuna in the Solomon Islands (25).

Second, unlike the nonspatial model, the number of dFADs outside the MPA negatively affects the benefits produced by cFADs inside the MPA (Figure 4B). In the nonspatial model, dFADs offer benefits to MPAs when they are inside their boundaries, and disbenefits when they are outside. By contrast, in the spatial model

these benefits and disbenefits are unequal. dFADs enter the MPA with a lower number of associated fish and leave with a larger number: a net export. This inequality is greatest for large MPAs, since dFADs will spend more time traversing them, accumulating a higher biomass in the process.

Third, the benefits provided by the cFADs vary in space and time, depending on the location of the different FADs and their recent history (e.g., if they have recently exited an MPA). As a consequence, the amplification provided by the cFADs varies in space and time, as captured by the bounds in Figure 4. Although cFADs offer an average amplification, certain locations within the MPA offer higher and lower benefits. For the species and ecosystem functions that benefit from high fish densities (e.g., hunting seabirds), this variation makes it more likely that benefits will be available at some location in the MPA.

DISCUSSION

The world's tuna fishing fleets increasingly rely on FADs to enhance their catches. By producing and maintaining high densities of fish in predictable locations, these fleets use dFADs to increase the efficiency of their harvests. Stationary conservation FADs would operate under similar principles, but for a diametrically opposite motivation. By increasing the residence time of species within MPAs, cFADs could increase the efficiency of blue water MPAs, substantially amplify the benefits that they produce. The addition of cFADs to blue water MPAs could address their greatest limitation – that MPAs in the open ocean are small relative to the distribution of targeted stocks. Our results indicate that the relative benefits of cFADs are likely to be largest when the MPA is small (Figure 2 & 4). The addition of cFADs is likely to be cheaper and less controversial than increasing the area of blue water MPAs.

Conservation FADs do not have to reduce the overall mortality risk faced by pelagic fishes to deliver a benefit to blue water MPAs – the ecosystems within an individual blue water MPA can still benefit from an increased local density of aggregating fishes, even if the total abundance of those fishes is the same. Drifting objects, which FADs mimic, help tuna form and maintain school integrity and size (18), which may increase their feeding efficiency and their visibility to other species that feed in association with them. In the PRIMNM, sooty terns feed obligately with yellowfin tuna schools, and great frigatebirds and red-footed boobies benefit from schooling subsurface predators, including tuna species, for effective foraging (6).

Our results provide a theoretical rationale for deploying cFADs to blue water MPAs. However, the models that we use here are limited in a number of important ways that may affect the potential benefits offered by cFADs. First, cFADs will increase the densities of any species that is attracted to pelagic floating objects, and not all of these will be negatively affected by fishing effort or bycatch mortality. Over 55 bony fish species are frequently found around dFADs used by the tropical tuna purse seine fishery, and many of them will be abundant, fast-growing, and of low conservation concern (26). Second, our models are based on the assumption that the aggregating effects of cFADs and dFADs are equivalent. dFADs move passively with ocean currents, which may place them to current boundary locations that offer better conditions for environmental factors and food. Third, our models are not parameterised for any particular fish species or location; they are instead general and theoretical. Our models cannot accurately predict fish densities around cFADs, nor how these densities would change with the number or location of the cFADs.

The immediate policy question for conservation FADs in blue water MPAs is whether their benefits are large enough to justify their deployment. Moving forward, we believe that the best method to answer this question is experimental deployment of cFADs in existing blue water MPAs, with particular target species and ecosystem functions being identified and measured. The downside risks of such experimental management are low, as the world's oceans are already full of drifting FADs, and even MPAs are continually traversed by dFADs. The enthusiasm and speed with which dFADs have been adopted by pelagic fisheries speaks to the size of the potential opportunity.

ACKNOWLEDGEMENTS

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REFERENCES CITED

1. Wagner D. Big Ocean: A shared research agenda for large-scale marine protected areas. Big Ocean Netw Rep. 2013;

2. O'Leary BC, Ban NC, Fernandez M, Friedlander AM, García-Borboroglu P, Golbuu Y, et al. Addressing criticisms of large-scale marine protected areas. Bioscience. 2018;68(5):359–70.

3. Crespo GO, Dunn DC. A review of the impacts of fisheries on open-ocean ecosystems.

4. Pauly D, Zeller D, Palomares MLD. Sea around us concepts, design and data. Vanc BC. 2015;

5. Green AL, Maypa AP, Almany GR, Rhodes KL, Weeks R, Abesamis RA, et al. Larval dispersal and movement patterns of coral reef fishes, and implications for marine reserve network design. Biol Rev. 2015;90(4):1215–47.

6. Gilman E, Chaloupka M, Fitchett M, Cantrell DL, Merrifield M. Ecological responses to blue water MPAs. PLoS ONE. 2020;15(7).

7. Kaplan D, Chassot E, Gruss A, Fonteneau A. Pelagic MPAs: The devil is in the details. TRENDS Ecol Evol. 2010;25(2):62–3.

8. Gilman E, Kaiser M, Chaloupka M. Do static and dynamic marine protected areas that restrict pelagic fishing achieve ecological objectives? Ecosphere. 2019;10(12):e02968.

9. Koldewey HJ, Curnick D, Harding S, Harrison LR, Gollock M. Potential benefits to fisheries and biodiversity of the Chagos Archipelago/British Indian Ocean Territory as a no-take marine reserve. 2010;

10. Game ET, Grantham HS, Hobday AJ, Pressey RL, Lombard AT, Beckley LE, et al. Pelagic protected areas: the missing dimension in ocean conservation. Trends Ecol Evol. 2009;24(7):360–9.

11. UNEP-WCMC, IUCN. Protected Planet: The World Database on Protected Areas (WDPA). Cambridge, UK;

12. Hanich Q, Davis R, Holmes G, Amidjogbe E-R, Campbell B. Drifting Fish Aggregating Devices (fads): Deploying, Soaking and Setting–When is a fad 'Fishing'? Int J Mar Coast Law. 2019;34(4):731–54.

13. Maufroy A, Kaplan DM, Bez N, De Molina AD, Murua H, Floch L, et al. Massive increase in the use of drifting Fish Aggregating Devices (dFADs) by tropical tuna purse seine fisheries in the Atlantic and Indian oceans. ICES J Mar Sci. 2017;74(1):215–25.

14. Baidai Y, Dagorn L, Amande MJ, Gaertner D, Capello M. Tuna aggregation dynamics at Drifting Fish Aggregating Devices: a view through the eyes of commercial echosounder buoys. ICES J Mar Sci. 2020;77(7–8):2960–70.

15. Wain G, Guéry L, Kaplan DM, Gaertner D. Quantifying the increase in fishing efficiency due to the use of drifting FADs equipped with echosounders in tropical tuna purse seine fisheries. ICES J Mar Sci. 2021;78(1):235–45.

16. Gershman D, Nickson A, O'Toole M. Estimating the use of FAD around the world, an updated analysis of the number of fish aggregating devices deployed in the ocean. Pew Env Gr. 2015;1–24.

17. Clavareau L, Sabarros PS, Escalle L, Bach P, Abascal FJ, Lopez J, et al. Elasmobranch bycatch distributions and mortality: insights from the European tropical tuna purse-seine fishery. Glob Ecol Conserv. 2020;24:e01211.

18. Dagorn L, Holland KN, Restrepo V, Moreno G. Is it good or bad to fish with FAD s? What are the real impacts of the use of drifting FAD s on pelagic marine ecosystems? Fish Fish. 2013;14(3):391–415.

19. Imzilen T, Lett C, Chassot E, Kaplan DM. Spatial management can significantly reduce dFAD beachings in Indian and Atlantic Ocean tropical tuna purse seine fisheries. Biol Conserv. 2021;254:108939.

20. Curnick DJ, Feary DA, Cavalcante GH. Risks to large marine protected areas posed by drifting fish aggregation devices. Conserv Biol. 2021;35(4):1222–32.

21. Jaquemet S, Le Corre M, Marsac F, Potier M, Weimerskirch H. Foraging habitats of the seabird community of Europa Island (Mozambique Channel). Mar Biol. 2005;147(3):573–82.

22. Au DW, Pitman RL. Seabird interactions with dolphins and tuna in the eastern tropical Pacific. The Condor. 1986;88(3):304–17.

23. Visser AW, Fiksen Ø. Optimal foraging in marine ecosystem models: selectivity, profitability and switching. Mar Ecol Prog Ser. 2013;473:91–101.

24. Avgar T, Betini GS, Fryxell JM. Habitat selection patterns are density dependent under the ideal free distribution. J Anim Ecol. 2020;89(12):2777–87.

25. Kleiber P, Hampton J. Modeling effects of FADs and islands on movement of skipjack tuna (Katsuwonus pelamis): estimating parameters from tagging data. Can J Fish Aquat Sci. 1994;51(12):2642–53.

26. Amandé JM, Ariz J, Chassot E, Chavance P, Delgado de Molina A, Gaertner D, et al. By-catch and discards of the European purse seine tuna fishery in the Indian Ocean. Estimation and characteristics for the 2003-2007 period. Indian Ocean Tuna Comm. 2008;

27. Kellner JB, Tetreault I, Gaines SD, Nisbet RM. Fishing the line near marine reserves in single and multispecies fisheries. Ecol Appl. 2007;17(4):1039–54.