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Meta-analysis of public RNA sequencing data of queens and workers in social Hymenoptera and termites

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Simple Summary: Social insects form complex, sophisticated societies in which members work cooperatively. Ants, bees, wasps, and termites are examples of social insects. The key characteristic of their structured societies is the reproductive division of labor. The queens reproduce within the nests, whereas workers engage in other tasks for colony development. Although gene expression profiles associated with the reproductive division of labor have been revealed for each social species, the existence of core regulatory genes commonly shared by all or multiple social insects is largely unknown. To address this issue, we conducted a meta-analysis by collecting and reanalyzing massive amounts of data from social bees, social wasps, ants, and termites. We identified 20 genes with queen- and worker-biased expression. Twelve of these genes have not been previously associated with reproductive division of labor. The expression of these genes is likely necessary for social insects to exhibit reproductive division of labor.

Abstract: Eusociality in insects has evolved independently many times. One of the most notable characteristics of eusociality is the reproductive division of labor. In social insects, the reproductive division of labor is accomplished by queens and workers. Transcriptome analyses of queens and workers have been conducted for various eusocial species. However, the genes that regulate the reproductive division of labor across all or multiple eusocial species have not yet been fully elucidated. Therefore, we conducted a meta-analysis using publicly available RNA-sequencing data from four major groups of social insects. In this meta-analysis, we collected 258 pairs (queen vs. worker) of RNA-sequencing data from 34 eusocial species. The meta-analysis identified a total of 20 genes that were differentially expressed in queens or workers. Out of these, 12 genes have not previously been reported to be involved in the reproductive division of labor. Functional annotation of these 20 genes in other organisms revealed that they could be regulators of behaviors and physiological states related to the reproductive division of labor. These 20 genes, revealed using massive datasets of numerous eusocial insects, may be key regulators of the reproductive division of labor.

Keywords: Division of labor; eusociality; social insect; meta-analysis; RNA sequencing

1. Introduction

Organisms have undergone several evolutionary transitions from cellular to multicellular organisms and their organized societies [1]. Sociogenomics aims to comprehensively understand social life from a molecular perspective [2]. Eusociality is the most advanced social form, characterized by a high level of cooperation among members of society. An essential characteristic of eusociality is the reproductive division of labor, which is accomplished by reproductive and non-reproductive castes. Reproductive castes are physiologically and behaviorally specialized in reproduction, whereas non-reproductive castes engage in brood care or foraging. The division of labor is believed to increase the

overall productivity of eusocial colonies [3].

Eusociality, which is based on the reproductive division of labor, has evolved independently in several insect lineages [4,5]. Eusocial Hymenoptera (bees, ants, and wasps) and termites are extensively studied insects that have complex and sophisticated societies. The reproductive and non-reproductive castes of these social insects are referred to as queen/king and workers/soldiers, respectively[6]. The investigation of genomic changes underlying the convergent evolution of eusociality among several lineages (bees and ants) has identified the genes that molecular evolution has occurred with increased social complexity [7–10]. Transcriptome analysis using the pharaoh ant *Monomorium pharaonis* and the honeybee *Apis mellifera* revealed that convergent eusocial evolution is based on the expression of highly conserved reproductive and lineage-specific genes [11]. In addition, many transcriptome analyses of queens and workers have been conducted for each species of bee, ant, wasp, and termite [12–33]. However, no comprehensive transcriptome analysis has been performed to understand the regulatory mechanisms of reproductive division of labor across a wide range of eusocial species.

Physiological differences between queens and workers are mainly produced by quantitative differences in juvenile hormones (JH), neurotransmitters, and neuropeptides. JH generally acts as a gonadotropic hormone in eusocial and solitary insects [34–38]. Vitellogenin (Vg) is precursor protein of the egg yolk, and is taken up by oocytes via receptor (Vg receptor)-mediated endocytosis [39,40]. Vg synthesis and uptake into ovary are promoted by JH [Soller et al 1999]. Vg and its receptors are highly expressed in the reproductive castes of various social insects [11,30,41–46]. The regulation of the secretion of neuropeptide and neurotransmitter plays a central role in controlling the reproductive division of labor. The expression of insulin-like peptide (ILP) is upregulated in the queens of several ants and termite *Macrotermes natalensis* [47,48]. In *A. mellifera*, however, *ILP* expression is lower in the old queen than in the old workers [49]. Neuropeptides other than insulin also play an important role in the reproductive division of labor. Corazonin is a neuropeptide that is highly expressed in the workers of three ants (*Harpegnathos saltator*, *Camponotus floridanus* and *Monomorium pharaonis*) and one wasp (*Poistes canadensis*) [16]. Biogenic amines are the primary neuroactive substances that control the ovary development of reproductive caste and worker behavior [50].

Meta-analysis is common in clinical research, but less general in basic biology including entomology[51]. Recently, however, a meta-analysis has provided interesting insights into several fields of entomology. A meta-analysis of the number of insect species provided a different view of spatiotemporal patterns of insect abundance than previously suggested. In the field of genomics, a search for horizontally transmitted genes within many number of insect genomes led to the discovery of genes involved in courtship behavior in lepidopteran insects. A meta-analysis that combines transcriptomic data obtained from multiple studies is effective identifying novel gene expression associated with specific biological processes [52–54]. Additionally, it is possible to identify master regulatory genes by combining transcriptome data from a wide range of species. We developed meta-analysis methods for publicly available transcriptome data from humans, plants, and insects [52,55–58]. In this study, we used a meta-analysis method to identify regulatory genes related to the reproductive division of labor in ants, bees, wasps, and termites. We collected RNA sequencing data for 258 pairs (queens vs. workers) from 34 eusocial species. Meta-analysis detected 212 genes that were conserved in all species examined in this study, 20 of these 212 genes were significantly differentially expressed between queens and workers.

2. Materials and Methods

2.1. Curation of public RNA sequencing data

Accession IDs (Project ID) related to eusociality were searched from the public databases All Of gene Expression (AOE) [59] and DBCLS SRA (https://sra.dbcls.jp/

(accessed on 11 May 2022)) using the keywords caste, eusocial, eusociality, and sociality. Metadata was obtained using NCBI SRA Run Selector (https://www.ncbi.nlm.nih.gov/Traces/study/ (accessed on 12 May 2022)). The acquisition of RNA sequencing data was limited to species for which the genome or reference transcriptome was available. RNA sequencing data from each species were obtained in pairs (queen and worker) to investigate gene expression related to the reproductive division of labor.

2.2. RNA-seq data retrieval, processing, and quantification

SRA retrieval and conversion to FASTQ files were performed using the prefetch and fasterq-dump programs in the SRA Toolkit (v3.0.0) in the Docker environment (v3.3.1) [60]. To decrease the disk space usage, the obtained FASTQ files were immediately compressed using pigz (v. 2.4). For trimming and quality control, Trim Galore! (v.0.6.6) [61] and Cutadapt (v.3.4) [62] software were used. Trim Galore! was run with the parameter fastqc --trim1--paired (if data were paired-end). Trim Galore! included Cutadapt (v.3.4) to trim low-quality base calls. Because all the data passed through this quality check, low-quality data were not included in this study. Biological replicates were treated as one experiment, and technical replicates were combined into one expression dataset.

The files of the reference genome and transcripts used for quantifying gene expression levels are listed in Table S1 (https://doi.org/10.6084/m9.figshare.21524286.v3). If available, these data were obtained from the NCBI Reference Sequence Database (RefSeq) or open data repositories used in the cited literature (accessed on 5 Aug 2022). Transcript expression was quantified using Salmon (v.1.5.0) [63] with the parameters -i index -l IU. Gene expression levels were quantified using transcripts per million (TPM). For species with only genome sequences, RNA sequencing reads were mapped to the reference genome using HISAT2 (v.2.2.1) [64] with the parameters -q --dta -x, and then quantification of the mapped data was performed using StringTie (2.1.5) [65]. TPM was used as a quantitative value of the gene expression level.

2.3. The detection of genes related to the reproductive division of labor

A calculation method developed for the meta-analysis of RNA sequencing data was employed [52,55–58]as described below. Pairs of the queen and worker RNA sequencing data (Table S2: https://doi.org/10.6084/m9.figshare.21524286.v3) were used to calculate the queen/worker ratio of TPM for each transcript (referred to as the QW ratio). The QW ratio for each transcript was calculated using Equation (1).

$$\text{QW ratio} = \log_2\left(\text{TPM}_{queen} + 0.01\right) - \log_2(\text{TPM}_{worker} + 0.01), \quad (1)$$

This pairwise calculation was performed for all the pairs described in Table S2 (https://doi.org/10.6084/m9.figshare.21524286.v3). The value "0.01" was added to TPM to convert zero into a logarithm. QW ratios were classified as "upregulated," "downregulated," or "unchanged" according to the thresholds. Transcripts with a QW ratio greater than 0.58496250072116 (1.5-fold expression change) were treated as "upregulated." Transcripts with QW ratios less than –0.58496250072116 (1/1.5-fold expression changes) were treated as "downregulated." Others were treated as "unchanged." The QW score was calculated to identify the genes involved in the reproductive division of labor. The QW score was calculated by subtracting the number of pairs with "downregulated" from those with "upregulated, " as shown in Equation (2):

$$\text{QW score} = \text{count number}_{\text{upregulated}} - \text{count number}_{\text{downregulated}}, \quad (2)$$

Python codes for calculating QW-ratios and QW-scores were obtained from a previous study[55](https://github.com/no85j/hypoxia_code/blob/master/CodingGene/HN-score.ipynb (accessed on 13 Aug 2022)).

2.4. Annotation for transcripts and enrichment analysis

To determine homologous relationships, the protein BLAST (BLASTP) program in the NCBI BLAST software package (v2.6.0) was used with parameters -evalue 1e-10 -outfmt 6 -max_target_seqs 1. The transcript files used for protein translation are listed in Table S1 (https://doi.org/10.6084/m9.figshare.21524286.v3). For species with only genome sequences, GFFread (v0.12.1) [66] was used to extract transcript sequences from the genome sequence. TransDecoder (v5.5.0) (https://transdecoder.github.io/ (accessed on 22 Jun 2021)) was used to identify and translate the coding regions into amino acid sequences. Open reading frames were extracted using TransDecoder.Longorfs with parameters -m 100, and coding regions were predicted using TransDecoder.Predict with the default parameters. As a result of the above processes, the transcript IDs of all species were converted to the protein IDs of *A. mellifera*. The protein IDs of *A. mellifera* were then converted to those of other species using the annotation table for transcripts of *A. mellifera* [67]. Transcripts and amino acid sequences created in this study can be downloaded from figshare (Supplemental File S1). Enrichment analysis was performed using the Metascape software with default settings (GO Biological Processes, Canonical Pathways, KEGG Pathway and WikiPathways with p-value < 0.01, a minimum count of 3 and an enrichment factor > 1.5 (the ratio between the observed counts and the counts expected by chance)) [68]. Gene symbols for *D. melanogaster* and *H. sapiens* were used for Metascape.

3. Results

3.1. Data collection of transcriptomes related to queens and workers

RNA sequencing data of queens and workers have been collected from many social insects to identify the genes underlying the reproductive division of labor. Consequently, 258 pairs (queen vs. worker) of RNA sequencing data were collected from 34 eusocial insect species (Figure 1A, Table S2; https://doi.org/10.6084/m9.figshare.21524286.v3). Ants (*Harpegnathos saltator* and *Monomorium pahraonis*), termites (*Cryptotermes secundus*), honeybees (*A. mellifera*), and wasps (*Polistes dominula*) were among the higher ranked species, indicating that RNA sequencing data from a wide range of eusocial species were included in the dataset. When compared among the tissues, brain-derived RNA sequencing data were the most abundant (Figure 1B).

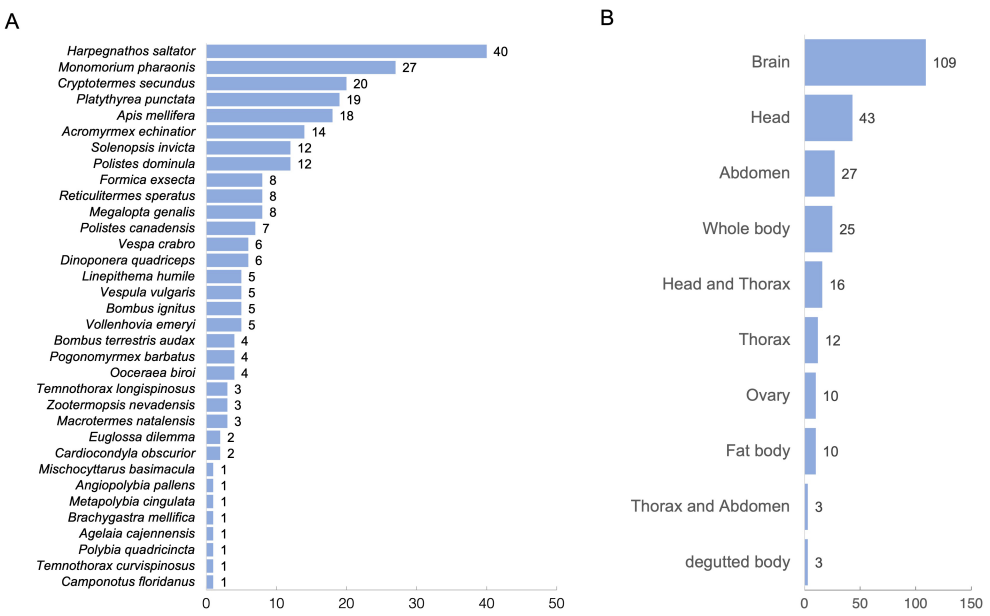

Figure 1. Number of RNA sequencing data compared across species (A) and tissues (B). The number of RNA-seq data used is indicated by bar graphs and numerical values.

3.2. Meta-analysis of RNA sequencing data in social insects

The expression ratio (queen vs. worker) (QW-ratio) was calculated for each species. At this point, the transcript IDs in the QW ratio table differ from species to species, even for homologs. Therefore, we converted the transcript IDs of each species to the protein IDs of *A. mellifera*, and then merged the tables in which QW-ratio of each species was described. Among the social insects used in this study, 212 genes were conserved. In addition, for each of 212 genes, 258 pairs of QW-ratio values were obtained (Table S3: https://doi.org/10.6084/m9.figshare.21524286.v3). The QW score for each gene was then calculated based on the QW ratio table. The QW score for each gene was calculated by subtracting the number of pairs with downregulated (less than 1/1.5-fold change) gene expression from the number of pairs with upregulated (more than 1.5-fold change) gene expression. This method provides the overall trends of gene expression changes in 258 pairs of RNA sequencing data. High scores indicate highly expressed genes in the queen, while low scores indicate highly expressed genes in the workers. The 212 identified genes were ranked based on their QW-score values (Table S4: https://doi.org/10.6084/m9.figshare.21524286.v3). These genes were annotated by the corresponding homologs of other species (*D. melanogaster*, *H. sapiens*, *Mus musculus*, *Bombyx mori*, *Bombus terrestis*, and *Nasonia vitripennis*) using the annotation table created by Yokoi et al 2022 [67] (Table S4: https://doi.org/10.6084/m9.figshare.21524286.v3). We focused on the top 10 and bottom 10 genes in Table S4 because they showed abrupt score changes (Figure 2 and Table 1). Abrupt score changes indicated that their expression was remarkably variable in the dataset used in this study. QW-scores for each species are available in Table S5 (https://doi.org/10.6084/m9.figshare.21524286.v3). The QW-score of *H. saltor* is not extremely high or low compared to other species, despite this species have many replicates (Figure 1). Therefore, it was unlikely that the QW-score was biased by the number of replicates of *H. saltor*.

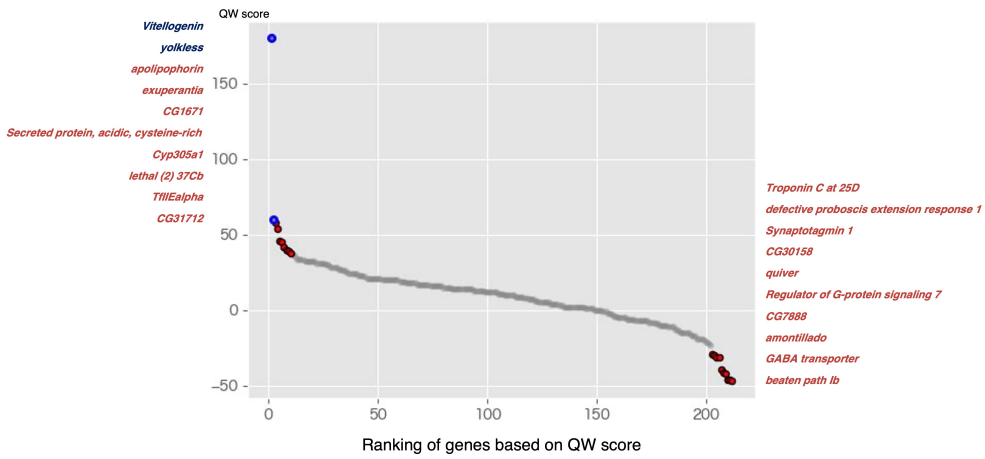

Figure 2. Scatter plots of QW-score of all genes (212 genes) identified in this study. Gene ranking based on QW score values. The gene names follow those of *D. melanogaster*. Red and blue dots indicate the top 10 best-ranked and worst-ranked genes, respectively. The blue dots indicate *Vitellogenin* and *yolkless* (Vitellogenin receptor), which are significantly expressed in queens of many eusocial species. Gene names indicate the top 10 and bottom 10 genes.

Table 1. Top 10 best-ranked and worst-ranked genes based on QW-score values.

RANK	QW score	Gene ID of *A. mellifera*	Gene symbol of *D.melanogaster*	Gene symbol *of H. sapiens*	Tissue-expression profile in female adult *D. melanogaster*	Caste with high expression that were reported in previous study	Biological/molecular function
1	180	NP_001011578.1 (Vitellogenin)	-	-	-	Queen in many species [53-57]	Oogenesis [51, 52]

2	60	XP_026295652.1	yl	LRP2	ovary	Queen in *Diacamma sp.*; *A. mellifera*; *M. pharaonis*; *T. longispinosus* [11, 58, 59]	Oogenesis [51, 52]
3	58	XP_026298285.1	apolpp	GTPBP2	fat body, spermatheca, head, eye	Queen in *R. speratus* [31]	JH binding [66]
4	54	XP_623977.2	exu	-	ovary	Queen in *A. mellifera*; *T. longispinosus* [59, 70]	Oogenesis [69]
5	46	XP_001120493.3	CG1671	TBL3	relatively high in ovary	-	Ribosome biogenesis [71]
6	45	XP_623079.2	SPARC	SPARC	heart, eye, head thoracicoabdominal ganglion	-	Insulin secretion [74]
7	42	NP_001314895.1	Cyp305a1	CYP2A13	spermatheca, head	-	JH synthesis [65]
8	40	XP_026300350.1	l(2)37Cb	DHX16	relatively high in heart	-	Splicesome [78]
9	39	XP_395253.2	TfIIEalpha	GTF2E1	relatively high in ovary	-	Ribosome biogenesis [72]
10	38	XP_395437.4	CG31712	GKAP1	brain, thoracicoabdominal ganglion,head	-	Splicesome [79]
203	-29	XP_026299328.1	TpnC25D	CALM3	head, eye, gut	-	A calcium signal transduction pathway (Uniprot)
204	-30	XP_026300722.1	dpr1	CNTN4	brain, thoracicoabdominal ganglion, head	-	Immunoglobulin superfamily [80,81]
205	-31	XP_026295297.1	Syt1	SYT1	thoracicoabdominal ganglion, brain, head	worker (forager) in *A. mellifera*; *P. metricus*; *S. invicta* [83]	Neurotransmitter release at synapses [83]
206	-31	XP_026298569.1	CG30158	RASD1	brain, head, thoracicoabdominal ganglion	-	Circadian entrainment [91, 92]
207	-39	XP_026302000.1	qvr	LYPD6	thoracicoabdominal ganglion, head, eye	worker (forager) in *B. terrestris* [87]	Sleep [87]
208	-41	XP_393403.3	RSG7	RGS7	brain, thoracicoabdominal ganglion, head	-	Insulin secretion [74]

209	-42	XP_393138.2	CG7888	SLC36A1	hindgut, thoracico-abdominal ganglion, brain	-	Amino acid transport (Uniprot)
210	-46	XP_392366.2	amon	PCSK2	brain, thoracicoab-dominal ganglion, head	-	The production of bi-oactive neuropeptide hormones [86]
211	-46	XP_006571520.1	Gat	SLC6A1	thoracicoabdominal ganglion, brain, head	Worker in A. mellifera [70]	regulation of circa-dian sleep [89]
212	-47	XP_026298737.1	beat	PSG4	brain, thoracicoab-dominal ganglion, head	-	Immunoglobulin su-perfamily [80, 82]

To estimate the function of these genes, the information of tissue-expression profile in *D. melanogaster* were added (Table 1). Nine out of ten highly expressed genes in workers were found primarily in the brain and thoracoabdominal ganglia in *D. melanogaster*, while highly expressed genes in queens were found in various tissues. If high expression has already been reported in queens or workers, caste and species names are noted. In addition, we added the biological/molecular functions predicted by the cited literature.

3.3. Enrichment analysis

We selected the top 30 best-ranked and worst-ranked genes based on the QW score, and then performed enrichment analysis using the corresponding IDs of *D. melanogaster* and *H. sapiens* (Figure 3). The molecular functions of many genes are highly conserved across species. In addition, since there is abundant information on human gene function, we can use this to infer the gene function of social insects. The number 30 was set arbitrarily. This setting was determined with reference to the scatterplot curve shown in Figure 2. No GO terms with markedly low p-values (-log10(P) values greater than 4) were detected when *D. melanogaster* IDs were used (Figure 3AB). However, when *H. sapiens* IDs were used, rRNA processing was significantly enriched in the top 30 genes (Figure 3C), whereas the positive regulation of ion transport and modulation of chemical synaptic transmission were enriched in the last 30 genes (Figure 3D).

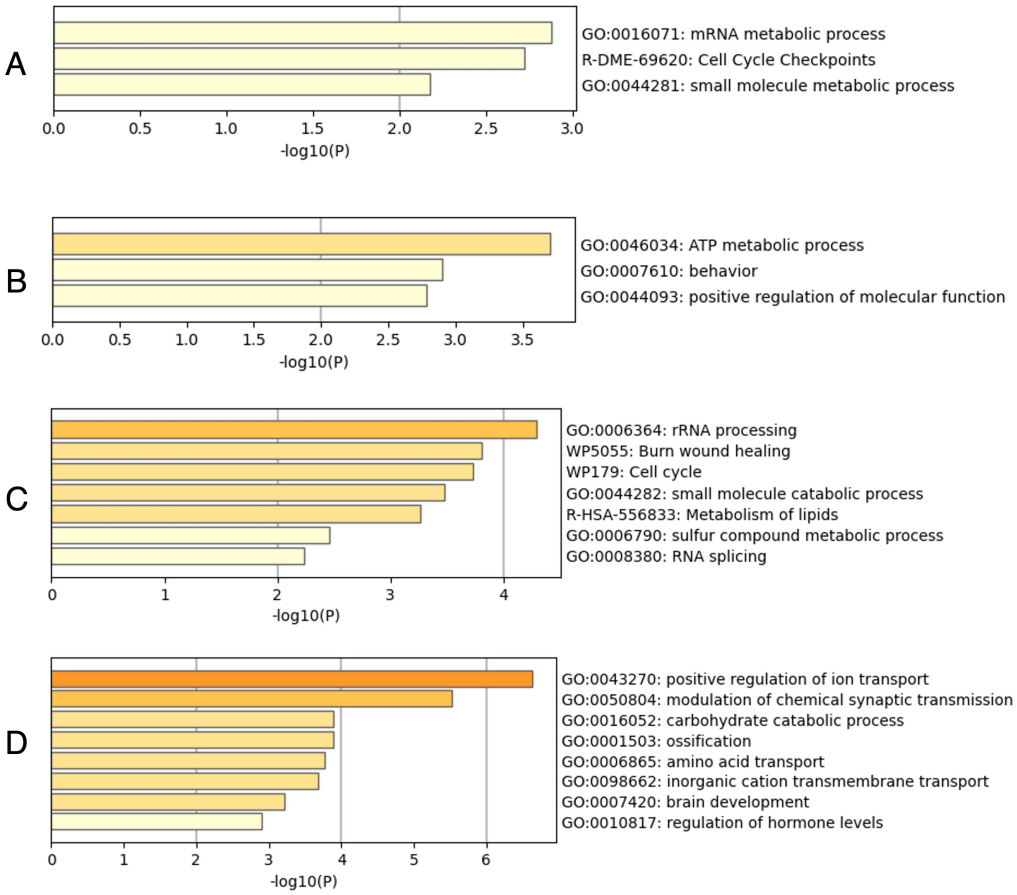

Figure 3. Results of enrichment analysis using *D. melanogaster* IDs (A, B) and *H. sapiens* IDs (B, C). Bars are colored differently according to the value of -log10(P).In this study, we focused on GO terms with -log10(P) values greater than 4.

4. Discussion

In this study, we aimed to identify genes underlying the reproductive division of labor across all or multiple eusocial insects by performing a meta-analysis of RNA-seq data. Meta-analysis revealed 20 genes with differential expression between queens and workers. Among these genes, vitellogenin and vitellogenin receptors, which are highly expressed in queens across many social insects [11,30,41–46], showed the highest QW score (Figure 1). This suggests that the current meta-analysis method is suitable for identifying differences in gene expression between queens and workers.

Among the genes identified in this study, we focused on 20 genes with the highest or lowest QW scores (Table 1). As the results of this study represent a vast amount of data from a wide variety of eusocial species, it is likely that these genes are the key factors underlying the reproductive division of labor in many social insects. In addition, several of the 20 genes were not previously implicated in the reproductive division of labor, providing novel insights into the underlying mechanisms of the reproductive division of labor. In the following sections, we summarize the biological and molecular functions of the corresponding homologs of these 20 genes in other organisms and discuss their role in social insects. Topics other than those listed below were available in the supplemental materials (Supplemental Discussion DOI: https://doi.org/10.6084/m9.figshare.21524286.v3).

4.1 Upregulated genes in queen

It has been known that the reproductive performance of queen is mainly regulated by JH (+ vitelogenin) and insulin in several social insects. We found several genes involved in these processes, as discussed below.

4.1.1 Oogenesis, Juvenile hormone (JH) binding, and synthesis (Vitellogenin, yolkless, Cyp305a1, apolipophorins, and exuperantia)

In queens, expression of JH-related genes (Vitellogenin, yolkless, Cyp305a1 and apolipophorins) and oogenesis gene (*Exuperantia*) were upregulated as expected. *Cyp305a1*, which encodes JH epoxidase, is required for JH biosynthesis in *D. melanogaster* [69]. *Apolpp* is a JH-binding protein in the cockroach *Blattella germanica* [70], and is upregulated in the termite queens, *Reticuliterms speratus* [31]. *Exuperantia* is a maternal factor that is involved in oocyte formation[71]. These genes may contribute to queen ovarian development via an upregulated JH titer. Although these genes, except for vitellogenin and vitellogenin receptor, are known to be expressed in the small number of species (Table 1), this study suggest that these genes are involved in queen ovary development of many social insects. However, in the honeybees *A. mellifera* and the ant *H. saltator,* the gonadotropic function of JH was lost [72,73]. The gonadotropic effects of these genes may depend on species.

4.1.2. The regulation of insulin secretion (SPARC (secreted protein acidic and cysteine rich), RSG7 (regulator of G protein signaling 7)

The expression of insulin-like peptide (ILP) is upregulated in the queens of several ants and in the termite *Macrotermes natalensis* [47,48]. To date, however, the regulatory mechanisms of insulin secretion in social insects are largely unknown. In this study, *SPARC* was highly expressed in queens, and *RSG7* was highly expressed in workers. In mice, *SPARC* promotes insulin secretion via downregulation of *regulator of G protein 4* (*RGS4*) expression in pancreatic β cells [79]. It also may regulate insulin secretion in eusocial insects via downregulation of *RSG7* expression. The discovery of SPARC and RSG7 is a major achievement of the meta-analysis, because these genes have not been reported as regulators of insulin secretion in social insects. In *A. mellifera*, however, *ILP* expression is lower in the old queen than in the old workers [49]. Therefore, the relationship between *SPARC* and *RGS* may possibly not apply to all eusocial species.

4.2. Upregulated genes in workers

Tissue expression profiles in *D. melanogaster* were referred for 20 listed genes using FlyAtras2 [80] (accessed on 14 Oct 2022). Unlike the genes significantly expressed in queens, many of the gene sets upregulated in workers were expressed in the nervous system of adult *D. melanogaster* (Table 1). Enrichment analysis also suggests the importance of genes expressed in the nervous system. Genes involved in positive regulation of ion transport (GO:0043270) were enriched in the results of enrichment analysis. It has been reported that genes related to ion transport are also expressed in the brain of *Polistes canadensis* [81], suggesting the contribution to increased cell activity in the brain. In addition, enrichment analysis showed that genes involved in the modulation of chemical synaptic transmission (GO:0050804) were enriched in the gene set that is highly expressed in workers. These results suggest active gene expression in the nervous system of workers, leading to worker-specific behaviors. The following sections, we discussed the genes that may be associated with worker-specific behaviors.

4.2.1. The regulation of secretion of neuropeptide and neurotransmitter (*amon* (*amontillado*) and *Syt1*(*Synaptotagmin 1*))

amon may be involved in secretion of neuropeptide related to determining worker behaviors. *amon,* which encodes a member of the prohormone convertase family, is required for propeptide processing to form bioactive neuropeptides [82]. In *D. melanogaster*,

amon protein is an important enzyme that catalyses many neuropeptides in the larval corpora cardiaca and perineuronal organs of the thorax and abdomen. Corazonin is a neuropeptide whose production is dependent on catalysis of amon protein [82]. Actually, corazonin plays a central role in determining the worker identity in *Harpegnathos saltator*, and is also highly expressed in the workers of three ants (*Harpegnathos saltator*, *Camponotus floridanus* and *Monomorium pharaonis*) and one wasp (*Poistes canadensis*) [16]. In addition to corazonin, *amon* may regulate the production of neuropeptides that regulate worker behavior. This gene has not been reported to control the worker behavior in social insects.

Serotonin secretion regulates worker responsiveness to trail pheromones in the ant *Pheidole dentata*, leading to cooperative foraging behavior [83]. *Syt1* may control the secretion of neurotransmitters such as serotonin involved in the regulation of social behavior. *Syt1,* which encodes a synaptic vesicle calcium-binding protein, is required for synaptic vesicles to interact with the plasma membrane [84]. $Ca2^{+}$ ions act on molecular complexes, including *Syt1*, triggering rapid exocytosis at the synapse. *Syt1* is highly expressed in ants (*S. invicta*), honeybees (*A. mellifera*), and wasps (*P. metricus*) [85]. *Syt1* might regulate the secretion of neurotransmitters that cause foraging and brood care.

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4.2.2. Regulation of circadian sleep (Gat (GABA transporter), quiver (qvr), and CG30158)

In honeybees, *A. mellifera*, sleep deprivation negatively affects the accuracy of the waggle dance, which informs colony members of the location of food resources [91]. This suggests that sleep control is essential for organized social behavior. *qvr,* which encodes a glycosylphosphatidylinositol (GPI)-anchored membrane protein, is a sleep-promoting factor in *D. melanogaster* [92]. *Gat* (*GABA transporter*) encodes a protein belonging to the solute carrier transporter family. Mutations in *Gat* cause excessive sleep because of the reduced uptake of GABA (a sleep inhibitor) from synapses to astrocytes in *D. melanogaster* [93]. It is possible that the amount of sleep of workers was optimized by *Gat* and *quiver*. Although these two genes are known to be expressed in workers of two bee species [74,94], and the results of this study suggest that they may be involved in worker sleep regulation in many social insect species.. It is unknown what effect excessive sleep has on the task performance of workers, but it is believed that the longer workers sleep, the less time they spend foraging and providing brood care for the colony.

A cooperative division of labor within the colony should require unifying the timing of when workers engage in foraging and feeding. *CG30158* is homologous to mammalian *Dexras1*, which encodes a Ras-like G protein. *Dexras1* is required for entrainment of circadian rhythms in the environmental light cycle in mice [95,96]. In worker foragers of the honeybee, *A. cerana japonica,* the circadian rhythm of locomotion activity is synchronized with light and temperature [97]. The *A. mellifera* foragers are often inactive at night [98]. In addition, the sleep rhythms of *A. mellifera* foragers are also known to be synchronized with the timing of the food available [99]. In social insects, the homolog of *CG30158* may play an important role in synchronizing foraging timing with environmental cycles.

5. Conclusions

This study is the first meta-analysis conducted on massive datasets of numerous eusocial species in two phylogenetically distant lineages, social Hymenoptera and termites, and provides two novel findings. First, the meta-analysis found 12 genes that had not been reported to be associated with the reproductive division of labor. This is because meta-analysis removes the data bias that occurs in the results of individual studies. Second, the 20 genes retrieved from massive datasets of a large number of eusocial species may be key regulators of the reproductive division of labor. Functional analyses of the genes

identified in this study are expected in the future. As there is increase in the publicly available RNA sequencing data , meta-analyses should prove useful in providing new insights into targeted biological phenomena, despite the fact that meta-analysis of RNA sequencing data is hardly widespread in entomology. The present study demonstrated the effectiveness of a meta-analysis of RNA sequencing data.

Supplementary Materials: Supporting information can be downloaded from figshare (https://doi.org/10.6084/m9.figshare.21524286.v3). Table S1: Reference genome and transcriptome used for quantifying gene expression levels (DOI: https://doi.org/10.6084/m9.figshare.21524286.v3). Table S2: Data sets for meta-analysis of public RNA sequencing data in eusocial insects (DOI: https://doi.org/10.6084/m9.figshare.21524286.v3). Table S3: Gene expression ratios when comparing queens and workers (QW ratio) (DOI: https://doi.org/10.6084/m9.figshare.21524286.v3). Table S4: QW score calculated based on the QW ratio. Table S5: QW-score for each species (DOI: https://doi.org/10.6084/m9.figshare.21524286.v3). The protein ID of *Apis mellifera* was functionally annotated using homologs of other species (DOI: https://doi.org/10.6084/m9.figshare.21524286.v3). Information from Unigene and Pham has also been added to this table. Supplemental File S1: Amino acid sequence newly created for the present study (peptide.tar.gz). The files of transcript sequence newly created for the present study (transcript.tar.gz) (DOI: https://doi.org/10.6084/m9.figshare.21524286.v3). Supplemental Discussion: Additional discussion of the genes identified in this study was described.

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Data Availability Statement: The data presented in this study are openly available in figshare [DOI: https://doi.org/10.6084/m9.figshare.21524286.v3 (accessed 19 Dec 2022)].

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