CiliaMiner: an integrated database for Ciliopathy Genes and Ciliopathies

Merve Gül Turan[1,2][†], Mehmet Emin Orhan[2][†], Sebiha Cevik[1], Oktay I. Kaplan[1*]

[1] Rare Disease Laboratory, School of Life and Natural Sciences, Abdullah Gul University, Kayseri,

Turkey

[2] Department of Bioengineering, School of Life and Natural Sciences, Abdullah Gul University,

Kayseri, Turkey

† These authors equally contributed to this work

*co-corresponding authors: oktay.kaplan@agu.edu.tr

Abstract

Cilia are found in eukaryotic species ranging from single-celled organisms, such as *Chlamydomonas reinhardtii*, to humans, but not in plants. The ability to respond to repellents and/or attractants, regulate cell proliferation and differentiation, and provide cellular mobility are just a few examples of how crucial cilia are to cells and organisms. Over 30 distinct rare disorders generally known as ciliopathy are caused by abnormalities or functional impairments in cilia and cilia-related compartments. Because of the complexity of ciliopathies and the rising number of ciliopathies and ciliopathy genes, a ciliopathy-oriented and up-to-date database is required. In addition, disorders not yet known as ciliopathy but have genes that produce cilia localizing proteins have yet to be classified. Here we present CiliaMiner, a manually curated ciliopathy database that includes ciliopathy lists collected from articles and databases. Analysis reveals that there are 55 distinct disorders likely related to ciliopathy, with over 4000 clinical manifestations. Based on comparative symptom analysis and subcellular localization data, diseases are classified as primary, secondary, or atypical ciliopathies. CiliaMiner provides easy access to all of these diseases and disease genes, as well as clinical features and gene-specific clinical features, as well as subcellular localization of each protein. Additionally, the orthologs of disease genes are also provided for mice, zebrafish, Xenopus, Drosophila, and *C. elegans*. CiliaMiner (https://kaplanlab.shinyapps.io/ciliaminer) aims to serve the cilia community with its comprehensive content, and highly enriched interactive heatmaps, and will be continually updated.

Introduction

Cilia are cellular organelles that extend out of the cell in unicellular and multi-organismal organisms. Within the cilia structure, nine pairs of peripheral microtubules are radially positioned and encircled by the cilia membrane, and this microtubule-based core structure is called the axoneme. Despite cilia being tiny cellular organelles, cilia have several subcompartments, including the transition zone (TZ), the basal body (a modified centriole), the axoneme, and the distal segment (1). Importantly, the ciliary structures and subcompartments have been well-preserved throughout evolution. Furthermore, depending on their structural differences and functional distinctions, cilia are categorized as either motile (9+2 axonemal structures) or non-motile (9+0 axonemal structure; primary cilium) **(Figure 1)** (2). In humans, motile cilia exist as multiple cilia on cell surfaces, requiring cell motility and fluid movement, whereas a single non-motile cilium emerges from a cell surface and is primarily involved in sensation (chemosensation and photosensation), developmental and signaling pathway regulation (Wnt, Hedgehog) (3,4). Any structural or functional anomalies in primary or motile cilia can cause a variety of rare disorders, and the term "ciliopathy" can be used to describe the complete spectrum of diseases that are related to primary or motile cilia (5).

The number of ciliopathies and ciliopathy-related genes has expanded dramatically over the last 20 years, owing to advances in medical technology, the ease of disseminating clinical knowledge, and the ability to sequence people (6). Apart from the growing number of ciliopathies and disease-associated genes, compiling, categorizing, and displaying the numerous ciliopathies and their associated genes has become more difficult due to symptomatic differences between ciliopathies, changes in symptom frequency, and the same gene causing multiple types of ciliopathies. Even though all the relevant details and information about diseases and disease genes are available, they are scattered among different biological and medical databases, necessitating users to visit these databases like MalaCards (7) and OMIM (8). It is critical to keep all of this massive data up to date and in one place, allowing the ciliopathy community to access all of the most recent data.

Here we present a ciliopathy-specific database, called CiliaMiner, that compiles and presents the updated list of ciliopathies, ciliopathy-associated genes, and the disease symptoms for each

condition. Furthermore, CiliaMiner provides classifications of ciliopathies and associated
disorders based on the subcellular localization of disease-associated genes in conjunction with
disease symptoms. CiliaMiner offers easy access to all ciliopathies, disease symptoms, ciliopathy
genes, ciliopathy candidate genes, and orthologs of ciliopathy genes and ciliopathy candidate
genes, seeking to serve as a major place in the ciliopathy database. Each piece of information is
manually acquired and confirmed before being uploaded to the appropriate part of the CiliaMiner
database, and users can also easily submit their data to CiliaMiner.

Materials and Methods

Data collection and curation
Data presented in the CiliaMiner were manually collected from six separate data sources, including
Online Mendelian Inheritance in Man (OMIM) (8), PubMed, Congruent clinical Variation
Visualization Tool (ConVarT) (9), OrthoList 2 (OL2) (10), Wormbase (11), and Protein Atlas (12)
databases. We specifically utilized the phrase "ciliopathy" in PubMed to find a comprehensive list
of ciliopathy disorders, followed by confirmation of ciliopathy disease using an OMIM search.
The manual search, together with OMIM validation, yielded 55 possible ciliopathy diseases. Once
the list of ciliopathy diseases is finalized, we collected the clinical symptoms and features of each
disease using OMIM and PubMed web pages. The disease-associated genes and relevant
references were collected from OMIM and PubMed. Additionally, we looked for potential
ciliopathy genes in PubMed using the phrase "cilia" between 2018-2022, and if the protein product
of the gene localizes to cilia, we next searched for disease relevance and collected symptoms for
that disease. Finally, we visited PubMed for each gene to determine whether the protein encoded
by a ciliopathy gene is detected in cilia or cilia-related compartments such as the basal body and
transition zone. Finally, we obtained the localization from ProteinAtlas (12), an excellent source
for protein localization, for those for which we were unable to find it.

Orthologs of Disease-Associated Genes
The ciliopathy disease list was created using human genes. Following that, the ortholog genes of
other organisms, *Mus musculus, Danio rerio, Xenopus laevis, Drosophila*
melanogaster, and *Caenorhabditis elegans*, are created using ConVarT (9) and OrthoList 2 (10)

(just for *C. elegans*). Wormbase used to be certain of the common gene name, sequence number,

and WormBase ID of *C. elegans*. We created a dedicated webpage for orthology search, called

"Ciliopathy Genes and Orthologs".

Collection of Detailed Clinical Symptoms of Diseases

Following the collection of the list of ciliary diseases, 4092 distinct clinical features are discovered

for disorders connected to ciliopathy (OMIM and the articles). A meticulous collection of clinical

symptoms suggests that there are 2354 and 1784 unique symptoms for primary ciliopathy and

secondary disease groups, respectively. Because the list of clinical features is extensive, we choose

the limited numbers of clinical symptoms to generate representative heatmaps on the "Ciliopathy

Names" page in the CiliaMiner. However, all ciliopathy clinical features are available on the

"Symptoms and Diseases" page. These symptoms are assigned to organ symbols and presented on

the same disease symptom summary panels on the "Ciliopathy Names" page.

Classification of diseases

All disease searches combined with symptoms and subcellular localization data yield a list of 55

different diseases and over 4000 clinical symptoms. The ciliopathy disorders exhibit a variety of

diverse symptoms that vary between ciliopathies as well as among the same ciliopathies (**Table

1**). For example, many clinical features, including finger abnormalities like polydactyly,

brachydactyly, and syndactyly, situs inversus, cerebral and skeletal anomalies are manifested in

the most common ciliopathies, including Bardet-Biedl Syndrome (BBS) (13), Meckel-Gruber

Syndrome (MKS) (14), Nephronophthisis (NPHP) (15), and Joubert Syndrome (JBTS) (16) while

we noticed that there are disease-specific clinical features, such as the molar tooth sign for JBTS.

We gathered symptoms from all these ciliopathies, collectively labelled as "core ciliopathies", as

a starting point for developing a list of ciliopathy symptoms. As a result, we categorized the most

prevalent ciliopathy symptoms as "core ciliopathy symptoms". It is worth noting that the core

ciliopathy symptoms are only obtained from the primary cilia-related ciliopathies; however, the

list of motile ciliopathies and motile ciliopathy-associated symptoms are gathered and listed in a

separate subgroup on the web page called "Motile Ciliopathy".

Furthermore, we noticed that some diseases, such as Birt-Hogg-Dubé syndrome and retinitis
pigmentosa, do not display many core ciliopathy symptoms, but the many protein products
encoded by the disease-causing genes localize to the cilia and cilia-related compartments, whereas
others, such as cone-rod dystrophy, STAR, and SOFT syndromes, have several symptoms that
overlap with core ciliopathies but lack genes encoding proteins that localize to cilia. Categorizing
diseases that do not have core ciliopathy symptoms but have cilia localizing genes poses a bigger
issue in determining whether they should be included in the ciliopathy list. The symptoms of these
diseases might also arise due to the non-ciliary functions of these cilia localizing proteins. The real
question is what criteria should be utilized to categorize ciliopathy diseases. Reiter et al. offered
the following forms of disease classification for ciliopathies: first-order and second-order
ciliopathies (17). Second-order ciliopathies are caused by genes encoding non-cilia localizing
proteins, but displaying cilia-related functions, including building cilia or regulating cilia function.
First-order ciliopathies are diseases caused by genes encoding proteins localizing to cilia and cilia-
related compartments. Even though this review offers an in-depth analysis of both motile and non-
motile ciliopathies, the clinical characteristics of disorders were not taken into consideration.
However, the disease classification should also consider the clinical features of diseases for several
reasons. 1) It is now well known that cilia-localizing proteins have non-ciliary functions, and
various clinical symptoms unrelated to ciliopathy may develop as a result of these genes' non-
ciliary functions. 2) Several disorders contain proteins that localize to cilia, including those that
cause conditions like Cornelia de Lange syndrome (CDLS), whose genes *SMC1A* and *SMC3* do
so (25) To correctly classify them, the symptoms of ciliopathy and CDLS should be compared.
For this reason, the CiliaMiner provides clinical features, subcellular protein localization, and
disease-specific clinical features, and uses heatmaps and interactive tables to display all of this
data. Similar to Reiter et al., we classified ciliopathies using subcellular localization, but disease
classification also takes into account clinical characteristics.
181
Based on our classification, the diseases are divided into primary, secondary, and atypical
ciliopathies. The following criteria were used to determine whether a disease is a primary
ciliopathy: 1) comparing disease clinical symptoms to core ciliopathy symptoms; and 2) the
localization of disease-causing gene products in cilia and cilia-related compartments (basal body,
transition zone, and centrosome). Furthermore, if a disease had previously been proposed as a

ciliopathy, we included it in the primary ciliopathy disease without considering subcellular
localization evidence. Secondary diseases are documented by independently checking clinical
features and subcellular localization data of condition-associated genes. The disease is listed as a
secondary disease if it presents similarity with core clinical symptoms but lacks a protein localizing
to the cilia and cilia-related compartments, and vice versa. In brief, well-known ciliopathy diseases
and reported ciliopathy disease to fall into the primary ciliopathy while the rest of the diseases are
regarded as secondary diseases if it presents similarity with core clinical symptoms but lacks a
protein localizing to the cilia and cilia-related compartments, and vice versa. Many single gene
disorders have not been classified into any BBS; NPHP or other types of ciliopathies but have been
reported to be atypical ciliopathies, and these atypical ciliopathies are collected directly from
research articles.
198
199   Primary ciliopathy consists of 34 diseases (**Supplementary Table 1**), while secondary disease
200   includes 18 diseases (**Supplementary Table 2**). Whole diseases are listed in the tables with
201   disease-associated genes and some clinical features. Bold lettering is used in tables to indicate
202   ciliary proteins which are localized in the cilia and cilia-related compartments as well as all clinical
203   symptoms that are common to all diseases when compared to core ciliopathies.
204
Atypical ciliopathies were created using the "ciliopathy" keyword search in PubMed between
2018-2022. This subtab shows unclassified ciliopathy genes with their general ciliopathy groups.
The original papers in this subtab report the atypical ciliopathy genes as potential ciliopathy genes.
We downloaded the CiliaCarta (18) gene list and supported it with either ciliary localization or
cilia-related process papers to provide candidate ciliary genes for ciliopathy diseases. This list also
includes additional genes from the "ciliopathy" search from PubMed. We also received a gene list
from this search that localized cilia and related sections, but they are unrelated to ciliopathies. Both
genes are considered potential ciliopathy genes.
213

214   **Database implementation**
215

CiliaMiner is a novel, user-friendly database that provides an up-to-date list of ciliopathy diseases
and disease genes, as well as detailed clinical symptoms and other disease-related information. R

Shiny (v1.7.1) (19) was used to generate the CiliaMiner website. In addition to the Shiny package,
other main libraries like DT (v0.20) (20), ggplot2 (v3.3.5) (21), heatmaply (v1.3.0) (22), and plotly
(v4.10.0) (23) are used for generating visual representations. The DT package is employed for
ordering, searching, and creating data tables in the user interface. ggplot2 is used for creating
graphical representations of the number of ciliopathy genes and their localization presentation on
the home page. Plotly and heatmaply are used for creating interactive figures comprising
interactive heatmaps and data tables.

Our GitHub repository contains a complete list of the manually curated data along with detailed
excel files for each sub-panel (https://github.com/thekaplanlab/CiliaMiner).

Results

Database overview and statistics

The last two decades have seen a dramatic expansion of the number of ciliopathies and
ciliopathies-associated genes. Even though all relevant data can be found in multiple databases
and supplementary files, there is an urgent need for a single and comprehensive database for
ciliopathies and ciliopathies-associated genes, so users can access the following: 1. A detailed list
of ciliopathies and potential ciliopathies, including symptoms and gene names 2. The updated list
of the ciliopathy genes and ciliopathy candidate genes. We, therefore, introduce a new ciliopathy
database, called CiliaMiner, which presents a total of 507 genes for different types of ciliopathies
based on our disease classification approach. Users may access the CiliaMiner
at https://kaplanlab.shinyapps.io/ciliaminer.

Primary ciliopathy has a total of 230 genes associated with 34 different ciliopathy diseases. To
assist users, the database includes disease names (also alternative disease names), the names of
disease-associated genes as well as the symptoms of each condition. Furthermore, the subcellular
localization of proteins encoded by disease-causing genes is supplied for each gene, along with
appropriate references, allowing users to view where the products of disease-causing genes are
located in cells. The data obtained demonstrated that the protein products of 72, 52, and 28

ciliopathy-related genes, respectively, are found in cilia, the basal body, and the transition zone.

78 ciliopathy encoding proteins do not seem to localize to the cilia-related compartments, but the

subcellular localization of data for these genes are collected from the ProteinAtlas, and these

proteins localize in the different subcellular departments. Some of the proteins are localized in the

plasma membrane (CC2D1A), mitochondria (PAM16, GPX4, XPNPEP3), endoplasmic reticulum

(GANAB, DNAJB11, ALG9, LRAT), Golgi apparatus (LRAT, TXNDC15, PI4KB), and

centrosome (KIAA0753, LRRCC1) **(Figure 2A)** (24). Additionally, some diseases do not have

published genes; just, only the disease condition is known.

The secondary disease group has 202 genes belonging to 18 secondary ciliopathy diseases. The

subcellular localization of proteins encoded by disease-causing genes revealed that 12 proteins are

found in the transition zone, 24 in the basal body, and 63 in the cilia; the localization of the rest of

the secondary disease-associated genes are localized in the other subcellular departments like

plasma membrane (PITPNM3, GUCA1A, MERTK, KLHL7, RGR, CNGA1, DHDDS, SLC7A14,

GPC3, SPTBN2, eEF2, TDP1, CC2D1A), mitochondria (IDH3B, HK1, IDH3A, AFG3L2,

COA7), endoplasmic reticulum (PNPLA6, CLCC1, MERTK, REEP6, CACNA1A, ELOVL5,

PLD3), Golgi apparatus (DRAM2) **(Figure 2B)** (24).

Motile ciliopathies have three diseases: primary ciliary dyskinesia, Birt-Hogg-Dubé syndrome,

and Juvenile myoclonic epilepsy. This group presents 75 established disease-associated genes for

all. Forty-eight are localized in the cilia, eight are localized in the basal body, and one is found in

the transition zone. Other genes are located in the out of the cilia like plasma membrane

(GABRA1), Golgi apparatus (GOLGA3, TP73, GABRD), cytosol (DNAAF2, SPAG1, CFAP54,

DNAH7, CLCN2). The number of all genes is given in **Figure 2C**.

In addition to primary and secondary diseases, all atypical and potential ciliopathy lists are

gathered and presented in the databases. There are 42 genes in the "Atypical Ciliopathy" sub-tab,

of which we present unclassified ciliopathy-associated genes **(Figure 2D)**. 274 ciliary proteins

encoding genes were carefully gathered, and a list of these hitherto unconnected to ciliopathy

genes, as well as data on subcellular localization, is presented in CiliaMiner. We consider each of

them as a possible ciliopathy candidate gene. In addition, the list of CilaCarta 934 genes has been

included in CiliaMiner for users to examine under the "Potential Ciliary Genes" sub-tab. It is

noteworthy that of these genes, 505 of them localize to cilia and cilia-related compartments.

Comparing the genes that cause primary, secondary, and atypical ciliopathies demonstrates that

numerous genes are responsible for multiple different diseases. A Venn diagram of the common

and group-specific genes of primary, secondary, and atypical ciliopathies is shown in **Figure 3**.

All ciliopathy subgroups share the gene TTC21B, and both primary and secondary ciliopathies

share the genes IFT140, BBS1, BBS2, ARL6, TTC8, CEP290, IFT172, C8orf37, IFT43, GLI3,

INPP5E, OFD1, SUFU, ARL3, CC2D1A, GUCY2D, RPE65, RPGRIP1, CRX, CRB1, IMPDH1,

RDH12, TULP1, PRPH2, CEP55, CEP83, DCDC2, and DYNC2I2. CFAP45 and PROM1 are

found in both atypical and secondary ciliopathies, whereas TTC26, SCLT1, DNAJB11,

DYNC2LI1, ALMS1, IFT80, and IFT74 are shared between primary and atypical

ciliopathies **(Figure 3)**.

For primary ciliopathy, our collection reveals a total of 2354 distinct clinical features, compared

to 1784 for secondary ciliopathy. Additionally, motile ciliopathy presents 341 clinical symptoms.

We feel that a supplementary table will be outdated if we provide one after some time, so we direct

users to the CiliaMiner website for the regularly updated list. All lists can be downloaded from the

website. The website was created in a way that users may either search for a symptom name to list

diseases where it is present or list all symptoms of a condition. Even for the same condition, distinct

symptoms have been recorded for different genes; as a result, the database includes a screening of

gene-specific symptoms, enabling users to look for gene-specific symptoms.

User interface and reactivity

The CiliaMiner homepage provides two search options. Users can do searches using either a

disease name or a gene name. Gene Search queries can include human gene names, gene IDs, and

Ensembl IDs. On the menu item "Symptoms and Diseases," there is also an option to search by

symptom names. Human gene names, gene IDs, and Ensembl IDs can all be used in Gene Search

queries. Gene names, gene IDs, OMIM numbers, ciliopathy names, and localization references can

all be utilized to do specialized searches across all pages and sub-tabs.

CiliaMiner has different visualization tools for the comparative analysis of ciliopathies and clinical
symptoms. Relative heat maps have been integrated into the Ciliopathy Names page. Specifically,
primary ciliopathies and secondary ciliopathies tabs can compare different ciliopathies regarding
clinical symptoms. In addition, heatmaps can be regenerated by user-selected ciliopathies and a
graphical representation of user inputs that can be used for comparing cilia-related clinical
features.

The representative figures of symptoms are also another visualization way for summarizing
ciliopathies. This visualization of symptoms was created by using symptom supergroups to
understand ciliopathies' effect on organs and systems in the human body. These 16 supergroups;
aural, neural, ophthalmic, skeletal, respiratory, hormonal, reproductive, facial, cerebral, renal,
coronary and vascular, nasal, liver, cognitive, digestive, and organ anomalies were created for a
straightforward understandable clinical representation by using ciliopathy based clinical
symptoms.

Strengths of CiliaMiner

The continuing increases in the number of ciliopathies and ciliopathy genes present enormous
challenges for clinical and basic scientists. They need to visit multiple databases and look up
specific information. In this regard, CiliaMiner, a novel manually-curated database for ciliopathy,
will be helpful since it makes all ciliopathies, ciliopathy genes, disease- and gene-specific
symptoms, and prospective ciliopathy candidate genes easily accessible. Furthermore, while
providing a thorough list of well-known ciliopathies, CiliaMiner also lists a potential ciliopathy
candidate, such as Cornelia de Lange syndrome. Cornelia de Lange syndrome (CDLS) has seven
disease-associated genes; 2 of them localize (*SMC1A* and *SMC3*) to cilia (25), and 2 of them
(*ANKRD11* and *HDAC8*) are implicated in cilia (26, 27). Although the precise relationship
between CDLS and cilia is not yet understood, localization and functional data suggest that several
symptoms, including hearing loss, abnormal hands and limbs, and cardiac issues, may be brought
on by the cilia-related functions of these genes. Additionally, the network analysis indicates CDLS
disease genes interact with many ciliary genes **(Figure 4)**. To establish a connection between cilia
and these genes, more work is required.

Researchers that work with model species are crucial to the cilia field, and CiliaMiners makes it simple to find the orthologs of genes that cause disorders. The database enables searches for mouse, zebrafish, clawed frog, fruit fly, and worm orthologs of human genes linked to diseases.

In conclusion, CiliaMiner is an intuitive database for the exploration of cilia fields, offering a detailed list of ciliopathies, ciliopathy genes, clinical characteristics of each illness, and prospective ciliopathy candidate genes. The content will be constantly updated and users will be able to add and/or correct the relevant information on the website.

Figure 1: The primary cilium is depicted, with a 9 + 0 microtubular axoneme surrounded by a ciliary membrane. Two ciliary subcompartments, including the basal body (BB) and transition zone (Y-shaped linkers) are shown. The intraflagellar transport (IFT), including motor proteins (kinesin and dynein), IFT-A, IFT-B, and Bardet-Biedl syndrome proteins (BBSome) move in both directions (anterograde and retrograde). Ciliary localization of proteins encoded by primary and secondary ciliopathy-causing genes is shown in a representative cilia structure.

Figure 2: The subcellular localization of ciliopathy disease genes within cells is displayed. Numbers in the bar graph demonstrate how the proteins encoded by the primary, secondary, and atypical ciliopathies-causing genes are localized throughout the cell.

Figure 3: The Venn diagram shows shared genes among ciliopathy-related genes of primary, secondary, and atypical ciliopathies.

Figure 4: Genes that cause Cornelia de Lange syndrome (CDLS) were analysed using a network.

Red represents the ciliary genes while blue points CDLS associated genes.

Ciliopathy Names	Clinical Symptoms	Disease Associated Genes	PubMed ID for Symptoms
Bardet–Biedl Syndrome	Anosmia/hyposmia, Ataxia, Behavioral Problems, Brachydactyly, Cardiac Anomalies/Arrhythmias/Anomalies, Central Nervous System Anomalies, Diabetes Mellitus, Facial Dysmorphism, Growth Deficiency, Hearing Loss, Hormonal Anomalies, Hypertension, Hypogenitalism, Impaired Psychomotor Speed and Motor Function, Infertility, Liver Anomalies, Mental Deficiency/Retardation, Obesity, Polydactyly, Renal Anomalies, Reproductive Anomalies, Retinal Degeneration, Short Stature, Syndactyly, Widely Spaced or Irregular Teeth	**BBS1, BBS2, ARL6, BBS4, BBS5, MKKS, BBS7, TTC8, BBS9, BBS10,** TRIM32, **BBS12, MKS1, CEP290, WDPCP, SDCCAG8, LZTFL1, BBIP1, IFT27, IFT172, C8orf37, IFT74, TTC21B,** SCLT1	26762677, 19252258
Joubert Syndrome	Ataxia, Coloboma, Cystic Kidney Dysplasia, Eye Anomalies, Growth Deficiency, Hepatic Developmental Abnormalities, Liver Anomalies, Nephronophthisis, Polydactyly, Respiratory Insufficiency, Widely Spaced or Irregular Teeth	**INPP5E, TMEM216, AHI1, NPHP1, CEP290, TMEM67, RPGRIP1L, ARL13B, CC2D2A, OFD1, TTC21B, KIF7, TCTN1, TMEM237, CEP41, TMEM138, CPLANE1, TCTN3,** ZNF423, **TMEM231, CSPP1, PDE6D, KIAA0586, TCTN2, CEP104, KATNIP, B9D1, MKS1, TMEM107, ARMC9, CEP120, SUFU,** PIBF1, **B9D2, ARL3,** FAM149B1, **TOGARAM1,** KIAA0753, **TMEM218, IFT74, CBY1, LRRCC1, ADAMTS9,** LAMA1	5816874, 26092869, 9438658, 16541367
Meckel–Gruber Syndrome	Anencephaly, Congenital Heart Defects, Cranial Anomalies, Cystic Kidney Dysplasia, Genital Anomalies, Hepatic Developmental Abnormalities, Hypoplasia, Liver Anomalies, Microcephaly, Nephronophthisis, Occipital Encephalocele, Polydactyly, Renal Anomalies, Situs Inversus	**MKS1, TMEM216, TMEM67, CEP290, RPGRIP1L, CC2D2A, NPHP3, TCTN2, B9D1, B9D2, TMEM231, KIF14, TMEM107, TMEM218, CEP55, TCTN3,** TXNDC15	29479449, 29209597, 7246621, 6859092, 4997715, 6486167, 3130875, 6654326
Nephronophthisis	Bronchiectasis, Cardiac Anomalies/Arrhythmias/Anomalies, Cerebral Anomalies, Coloboma, Cone-rod Dystrophy, Hypopituitarism, Intellectual Disability, Liver Anomalies, Occipital Encephalocele, Polydactyly, Retinitis Pigmentosa, Situs Inversus, Skeletal Abnormalities	**NPHP1, INVS, NPHP3, NPHP4, GLIS2, NEK8, TMEM67, TTC21B, WDR19,** ZNF423, **CEP164, ANKS6, CEP83, DCDC2,** MAPKBP1, XPNPEP3, **CC2D2A, ADAMTS9, SDCCAG8, CEP290**	17513324, 29717526

Table 1: The clinical features and disease-associated genes of selected ciliopathies (non-motile

ciliopathies) are shown in the table. The genes in bold are those that have been reported to localize

to cilia, while the other genes are not.

Supplementary Table 1: The clinical features and published genes of all primary ciliopathies.

Only the genes shown in bold localize to cilia; all other genes do not. Clinical characteristics

related with ciliopathies are shown in bold.

Supplementary Table 2: The clinical features and established genes of all secondary diseases. The only genes found to localize to cilia are those in bold. Clinical features associated with ciliopathies are highlighted (bold).

Acknowledgments

We thank Associate Professor Duygu Sacar for allowing Mehmet Emin Orhan to help this project. We appreciate Mustafa Pir's assistance with Figure 4. His coding and CilioGenics work enable us to produce Figure 4. We thank Dr. Abdullah Sezer for critical reading of the manuscript.

References:

1. Greenan, G.A., Vale, R.D. and Agard, D.A. (2020) Electron cryotomography of intact motile cilia defines the basal body to axoneme transition. J. Cell Biol., 219, e201907060.

2. Ishikawa, T. (2017) Axoneme Structure from Motile Cilia. Cold Spring Harb. Perspect. Biol., 9, a028076.

3. Lancaster, M.A., Schroth, J. and Gleeson, J.G. (2011) Subcellular spatial regulation of canonical Wnt signalling at the primary cilium. Nat. Cell Biol., 13, 700–707.

4. Huangfu, D. and Anderson, K.V. (2005) Cilia and Hedgehog responsiveness in the mouse. Proc. Natl. Acad. Sci. U. S. A., 102, 11325–11330.

5. Badano, J.L., Mitsuma, N., Beales, P.L., et al. (2006) The ciliopathies: an emerging class of human genetic disorders. Annu. Rev. Genomics Hum. Genet., 7, 125–148.

6. Wheway, G., Mitchison, H.M., and Genomics England Research Consortium (2019) Opportunities and Challenges for Molecular Understanding of Ciliopathies-The 100,000 Genomes Project. Front. Genet., 10, 127.

7. Rappaport, N., Twik, M., Plaschkes, I., et al. (2017) MalaCards: an amalgamated human disease compendium with diverse clinical and genetic annotation and structured search. Nucleic Acids Res., 45, D877–D887.

8. McKusick, V.A. (2007) Mendelian Inheritance in Man and Its Online Version, OMIM. Am. J. Hum. Genet., 80, 588–604.

9. Pir, M.S., Bilgin, H.I., Sayici, A., et al. (2022) ConVarT: a search engine for matching human

genetic variants with variants from non-human species. Nucleic Acids Res., 50, D1172–D1178.

10. Kim, W., Underwood, R.S., Greenwald, I., et al. (2018) OrthoList 2: A New Comparative

Genomic Analysis of Human and Caenorhabditis elegans Genes. Genetics, 210, 445–461.

11. Davis, P., Zarowiecki, M., Arnaboldi, V., et al. (2022) WormBase in 2022—data, processes,

and tools for analyzing Caenorhabditis elegans. Genetics, 220, iyac003.

12. Thul, P.J., Åkesson, L., Wiking, M., et al. (2017) A subcellular map of the human proteome.

Science, 356, eaal3321.

13. Valente, E.M., Rosti, R.O., Gibbs, E., et al. (2014) Primary cilia in neurodevelopmental

disorders. Nat. Rev. Neurol., 10, 27–36.

14. Hartill, V., Szymanska, K., Sharif, S.M., et al. (2017) Meckel-Gruber Syndrome: An Update

on Diagnosis, Clinical Management, and Research Advances. Front. Pediatr., 5, 244.

15. Luo, F. and Tao, Y.-H. (2018) Nephronophthisis: A review of genotype-phenotype correlation.

Nephrol. Carlton Vic, 23, 904–911.

16. Parisi, M.A. (2009) Clinical and molecular features of Joubert syndrome and related disorders.

Am. J. Med. Genet. C Semin. Med. Genet., 151C, 326–340.

17. Reiter, J.F. and Leroux, M.R. (2017) Genes and molecular pathways underpinning ciliopathies.

Nat. Rev. Mol. Cell Biol., 18, 533–547.

18. van Dam, T.J.P., Kennedy, J., van der Lee, R., et al. (2019) CiliaCarta: An integrated and

validated compendium of ciliary genes. PloS One, 14, e0216705.

19. Chang, W., Cheng, J., Allaire, J.J., et al. (2022) shiny: Web Application Framework for R.

shiny: Web Application Framework for R (2022).

20. Xie, Y., Cheng, J., Tan, X., et al. (2022) DT: A Wrapper of the JavaScript Library

"DataTables." DT: A Wrapper of the JavaScript Library "DataTables" (2022).

21. Wickham, H. (2016) ggplot2: Elegant Graphics for Data Analysis. ggplot2: Elegant Graphics

for Data Analysis; Springer-Verlag New York, (2016) .

22. Galili, T., O'Callaghan, A., Sidi, J., et al. (2018) heatmaply: an R package for creating

interactive cluster heatmaps for online publishing. Bioinformatics, 34, 1600–1602.

23. Inc, P.T. (2015) Collaborative data science. Collaborative data science https://plot.ly.

24. Uhlén, M., Fagerberg, L., Hallström, B.M., et al. (2015) Tissue-based map of the human

proteome. Science, 347, 1260419.

25. Khanna, H., Hurd, T.W., Lillo, C., et al. (2005) RPGR-ORF15, Which Is Mutated in Retinitis

Pigmentosa, Associates with SMC1, SMC3, and Microtubule Transport Proteins. J. Biol. Chem.,

280, 33580–33587.

26. Deardorff, M.A., Bando, M., Nakato, R., et al. (2012) HDAC8 mutations in Cornelia de Lange

syndrome affect the cohesin acetylation cycle. Nature, 489, 313–317.

27. Parenti, I., Gervasini, C., Pozojevic, J., et al. (2016) Broadening of cohesinopathies: exome

sequencing identifies mutations in ANKRD11 in two patients with Cornelia de Lange-overlapping

phenotype. Clin. Genet., 89, 74–81.