Epigenetic and Signaling Determinants of Blood-Brain Barrier-Regulatory Genetic Networks

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Abstract

The blood-brain barrier (BBB) controls the movement of molecules into and out of the central nervous system (CNS). Since a functional BBB forms by mouse embryonic day E15.5, we reasoned that gene cohorts expressed in CNS endothelial cells (EC) at E13.5 contribute to BBB formation, whereas adult gene signatures reflect BBB maintenance mechanisms. Supporting this hypothesis, transcriptomic analysis revealed distinct cohorts of EC genes during BBB formation and maintenance. Here we demonstrate that epigenetic regulator's histone deacetylase 2 (HDAC2) and polycomb repressive complex 2 (PRC2) control EC gene expression for BBB development and prevented Wnt/β-catenin (Wnt) target genes from being expressed in adult CNS ECs. Low Wnt activity during development modifies BBB genes epigenetically for the formation of functional BBB. As a Class-I HDAC inhibitor induces adult CNS ECs to regain Wnt activity and BBB genetic signatures that support BBB formation, our results inform strategies to promote BBB repair.

Keywords: Blood-brain barrier, Epigenetic, HDAC2, PRC2, Wnt signaling, β-catenin

Introduction

Central nervous system vessels possess a blood-brain barrier (BBB) that prevents toxins and pathogens from entering the brain. A leaky BBB can lead to deleterious consequences for CNS disorders including stroke, traumatic brain injury, and brain tumors[1]. Currently, no treatment options are available to sustain and/or regenerate BBB integrity. The identification and targeting of the mechanism that forms and maintains the BBB is an attractive strategy to regain BBB integrity.

By comparing the EC transcriptome of peripheral and brain vessels, the genomic profile that contributes to BBB was described[2,3]. Further, extensive gene expression changes in CNS ECs during development were also reported [4,5]. However, molecular mechanisms governing BBB gene transcription and how such mechanisms contribute to BBB establishment and maintenance are unresolved.

Epigenetic modifications of histones and chromatin-modifying enzyme activities are critical determinants of gene expression[6]. Histone deacetylases (HDACs) associate with specific transcription factors and participate in gene repression[7]. HDAC inhibitors important experimental tools for elucidating HDAC functions[8], and four HDAC inhibitors are FDA-approved drugs for cancer treatment[9]. More than thirty HDAC inhibitors are being investigated in clinical trials[10]. Similarly, polycomb repressive complex 2 (PRC2), a complex of polycomb-group proteins (such as EZH2, EED, and SUZ12), represses transcription via its methyltransferase activity that catalyzes tri-methylation of H3K27[11,12]. An EZH2 inhibitor is FDA-approved[13] for Follicular lymphoma.

Wnt/β-catenin (Wnt) signaling is essential for establishing the BBB [4,5,14-16], but the activity of this pathway declines gradually thereafter and is reported to be minimal in adults[15,17,18]. Evidence linking Wnt to the regulation of BBB genes includes an EC-specific knockout (KO) of β-catenin that affects BBB gene expression and integrity[5,19-21]. In CNS ECs, Wnt signaling requires the binding of Wnt ligands Wnt7a/7b to Frizzled receptors. This interaction

stabilizes the intracellular signaling molecule β-catenin, by suppressing a cytoplasmic destruction complex, which would otherwise degrade β-catenin. Stabilized β-catenin translocates to the nucleus and regulates Wnt target gene transcription by interacting with DNA binding transcription factors TCF/LEF (T-cell factor/lymphoid enhancing factor). The mechanisms underlying the reduced Wnt signaling and how Wnt regulates the BBB gene transcription are not established.

We describe the discovery of epigenetic mechanisms responsible for regulating BBB gene expression during development, how Wnt signaling is reduced, and the significance of these mechanisms for BBB development. Furthermore, we demonstrated that HDAC inhibitors activate BBB gene cohorts expressed during BBB formation, suggesting a potential therapeutic intervention.

Results

Transcriptional downregulation of key BBB genes in adult cortical ECs: evidence that distinct EC gene cohorts regulate BBB establishment versus maintenance

An intact, non-leaky BBB forms at E15.5[22]. We hypothesized that the EC gene cohorts involved in BBB formation are expressed on E13.5, and 3-4 month-old adult CNS EC will express gene cohorts required for BBB maintenance. To test this, we defined transcriptomes of primary cortical ECs isolated from E13.5 and 3-4-month-old adults. mRNA-seq analysis revealed strong expression of EC genes including *PECAM1*, *CDH5*, and *CLDN5* in comparison to perivascular cell types such as pericytes, neuronal and glial genes (S.Fig-1A). To define the gene cohorts responsible for BBB formation vs. BBB maintenance, we identified differentially expressed genes (DEGs) expressed in natural log (fold change) in E-13.5 relative to adult mice (Fig-1A,B). DEGs with a P-value < 0.05) were considered significant. In comparison to E13.5, 50% of genes were upregulated, 35% were downregulated in adults and 15% of genes were unchanged (Fig-1C).

GO enrichment analysis of the DEGs identified five functional categories: angiogenesis, cell-to-cell junction, transporters, extracellular matrix, and DNA binding transcription factors

(Fig-1D). Except for transporters, the other categories were characterized by a greater number of downregulated vs. upregulated genes. The DEGs included important BBB genes (Fig-1E). For example, tight junction (TJ) gene *CLDN1*, *CLDN5*, BBB transporters *MFSD2A*, *CAV1*, and BBB-related transcription factors *ZIC3*, *FOXF2*, and *SOX17* were differentially expressed (Fig-1E). The complete dataset is presented in supplementary table 1. Overall the transcriptomic analysis identified EC gene cohorts that expressed during the formation or maintenance of the BBB.

We validated the differential expression of important BBB and related genes (*e.g.*, *CLDN1*, *CLDN5*, *MFSD2A*, *ZIC3*, and *SOX17*) at E-13.5, E-17.5, P0, P7, and in the adult. This analysis revealed that *CLDN1*, *MFSD2A*, and *ZIC3* expression were significantly downregulated by E-17.5 and subsequent developmental stages (Fig-1F). Thus, the expression of these genes might be required for the establishment, but not maintenance, of the BBB. By contrast, adult expression of *CLDN5* was significantly upregulated compared to other developmental stages and *SOX17* was significant to other developmental stages except for P7(Fig-1F). Additionally, we validated the mRNA expression of CLDN11 and FOXF2 as well as the protein expression of *CLDN1*, *CLDN5* and *ZIC3* (S.Fig-B&C).

HDAC2 and PRC2 mediated transcriptional regulation of BBB genes.

Histone deacetylases (HDACs) have critical roles in development and tissue homeostasis, and HDAC inhibitors are instructive experimental tools [23,24]. To assess whether the transcriptional downregulation of BBB genes involves HDAC-dependent epigenetic repression, we treated adult ECs with a pan HDAC inhibitor trichostatin A (TSA) for 48 hours. TSA increased *CLDN1* (Fig-2A) and *ZIC3* (S.Fig-2A) expression relative to the control. Since there are four major HDAC classes, class-I (HDAC 1, 2, 3, and 8), class II (HDAC 4, 5, 6, 7, 9, 10), class III (SIRTs 1–7), and class IV (HDAC 11), we tested whether specific HDACs mediate the repression. The class-I HDAC inhibitor MS-275 significantly upregulated *CLDN1*, MFSD2A and ZIC3 while downregulated *CLDN5* (Fig-2A, S.Fig-2A). No significant difference in CLDN1 mRNA expression was observed with class-II HDAC inhibitor (S.Fig-2A). In adult ECs, HDAC2 exhibited greater expression than other class I HDACs (Fig-2B). To analyze the HDAC2 function, we utilized siRNAs to downregulate HDAC1, HDAC2, or HDAC3 in adult ECs. Knockdown (KD) of HDAC2 significantly

upregulated the repressed *CLDN1*, *MFSD2A*, and *ZIC3* genes and reduced *CLDN5* expression (Fig-2C). By contrast, HDAC1 and HDAC3 downregulation did not affect these genes. BBB genes analyzed above were selected based on their expression patterns and to represent important functional attributes of BBBs, such as tight junctions (*CLDN1* and *CLDN5*), transporters (*MFSD2A*), and transcription factors (*ZIC3*). We used ChIP-qPCR to test whether HDAC2 directly regulates BBB gene expression in E13.5 and adult cortical EC contexts. Three primers were designed ((-)500, TSS & (+)500) spanning the 1 kb region on each side of the promoter. HDAC2 occupancy was detected in the (-)and(+) 500 regions of *CLDN1* in adults with no significant enrichment in E13.5. *ZIC3* showed occupancy at all three regions (1kb) in the adult stage with E13.5 showed enrichment in TSS only (Fig-2D & S.Fig2B). Conversely, HDAC2 occupied *MFSD2A* only at E13.5((-)500 & TSS), and in *CLDN5* occupancy was detected at both E13.5 (1kb) and in adults ((-) & (+) 500). These results link HDAC2 to the developmental control of BBB genes (Fig-2D & S.Fig2B).

In our initial screening, we found that the PRC2 inhibitor DZNEP significantly increased *CLDN1* expression in adult ECs (S.Fig-2A). To analyze the PRC2 function in this context, we downregulated the PRC2 subunit EZH2 from adult ECs. Compared to the control, EZH2 downregulation significantly increased *CLDN1*, *ZIC3*, and *MFSD2A* expression and decreased *CLDN5* expression (Fig 2E). ChiP-qPCR of PRC2 subunit EED revealed EED occupancy at various regions of *CLDN1*, *MFSD2A,* and *ZIC3* at E13.5 and in the adult (Fig2F,G & S.Fig-2). EED occupied *CLDN5* at E13.5, but not in adult ECs. Together our data support a model in which HDAC2 and PRC2 are important determinants of EC BBB transcriptomes during BBB development.

Distinct histone modifications delineate the transcription program of BBB genes.

Since HDAC2 and PRC2 are regulating multiple BBB genes we sought whether they share similar post-translational histone modifications. To examine this, we performed ChIP-qPCR of potentially involved histone marks, including the repressive marks H3K27me3, and H3K9me3 as well as the activating marks H3K4me3 and H3K9ac. We have scanned approximately 1kb genomic region surrounding the TSS of *CLDN1*, *CLDN5*, *MFSD2A*, *ZIC3,* and *SOX17* in E13.5 and adults. H3K9me3 ChIP-qPCR on our selected BBB genes didn't show significant binding in either

stage (S.Fig-3A). We detected abundant enrichment of repressive histone mark H3K27me3 on *CLDN1* (Fig-3A,C), *MFSD2A*, and *ZIC3* (Fig-3C & S.Fig-3B,C) in adult ECs compared to E13.5. *CLDN1, MFSD2A,* and *ZIC*3 showed significant enrichment of H3K27me3 compared to IgG in E13.5 (Fig-3A & S.Fig-3B,C). While *CLDN5* showed significant enrichment of H3K27me3 on 1kb region in both E13.5 and adults (Fig-3B). However, E13.5 showed abundant enrichment of H3K27me3 in the -500 region compared to adults (Fig-3B).

Active histone mark, H3K4me3 showed significantly increased enrichment on *CLDN1* (TSS), *MFSD2A* (1KB), and *ZIC3* (-500 and +500) at E13.5 compared to adult (Fig-3A and S.Fig-3B, C). *CLDN1* (-)500 region didn't show any significant binding for H3K4me3 in both stages, while the (+)500 region showed significant binding compared to IgG with no difference between E-13.5 and adult. *ZIC3* TSS region showed significant enrichment of H3K4me3 in the adult compared to E-13.5(S.Fig-3). Supporting its abundant expression in adults, *CLDN5* (1KB) showed significant enrichment of H3K4me3 in adults compared to E13.5. Another active histone mark H3K9ac showed significant enrichment on *CLDN1* (TSS and +500) at E13.5 compared to the adult with no significant binding in the -500 region in both stages (Fig-3A & S.Fig-3). *ZIC3* (TSS and +500) showed a significant binding for H3K9ac in E13.5 compared to the adult while the -500 region didn't show any enrichment in both stages. *MFSD2A* showed significant enrichment of H3K9ac in the -500 and +500 regions at E-13.5 compared to adults. While the TSS region showed significant enrichment in adults compared to E-13.5. Histone modifications on *SOX17* is shown in S.Fig-3D. These results indicate that BBB genes acquire a unique epigenetic signature during development.

HDAC2 activity is critical for the maturation of BBB, while PRC2 is dispensable.

To examine the role of HDAC2 and PRC2 in BBB maturation, we knock out (KO) *HDAC2* and PRC2 subunit *EZH2* from ECs during embryonic development. To conditionally KO *HDAC2* or *EZH2* we used tamoxifen-inducible Cdh5(PAC)-CreERT2 mice. Tamoxifen was injected into the mother starting at E-12.5 and on alternate days until E-16.5 (Fig-4A & 4E). This allowed the KO of *HDAC2* or *EZH2* before the maturation of the BBB.

It was observed that *HDAC2 and EZH2* KO embryos grew normally and were alive on the day of sacrifice E17.5. Significantly decreased mRNA expression of *HDAC2* in the whole brain analysis confirmed the loss of HDAC2 (S.Fig-4). Compared to WT, *HDAC2* ECKO pial vessels were dilated and showed increased angiogenesis (Fig-4B). BBB permeability was assessed in *HDAC2* ECKO mice using a 70KD FITC-conjugated dextran tracer. The tracer remained confined inside the vessels of E17.5 WT embryos, supporting previous findings that the BBB matures by E-15.5 (Fig-4C). Conversely in HDAC2 ECKO, dextran leaked into the cortical parenchyma (Fig-4C). Green fluorescent intensity measurements confirmed the immature BBB in the *HDAC2 ECKO* (Fig-4D). A vascular analysis using angiotool determined that HDAC2 ECKO had a significantly higher vascularized brain area than W.T, indicating that HDAC2 plays a key role in angiogenesis (Fig-4D). While the pharmacological inhibition of class I HDAC using MS-275 in timed pregnant WT mice at E-13.5 showed significant leakage of tracer into the brain parenchyma at E-15.5 compared to vehicle-treated control (S.Fig-4A). The MS-275 treated embryos also showed a thin cortex compared to the control possibly due to the leakage of the drug into the brain which affects brain development (S.Fig-4A). These findings should be taken into account when considering the clinical use of MS-275 in a developing brain. In the whole brain mRNA analysis between the HDAC2 ECKO and WT showed significantly upregulated *CLDN1* in HDAC2 ECKO (S.Fig-4B). No significant difference was observed in *CLDN5*, *MFSD2A,* and *ZIC3* genes.

Compared to WT, at E-17.5 *EZH2 ECKO* showed dilated vessels with no evident increase in the pial vessel angiogenesis (Fig-4F). Confirming the loss of *EZH2*, mRNA analysis on the whole brain showed a significant reduction. BBB permeability assay showed a subtle leakage of 70KD fluorescent tracer into the brain parenchyma (Fig-4G). However, the fluorescence intensity analysis showed no significant difference (Fig-4H). Further, the mRNA analysis on the whole brain showed a significant decrease in *EZH2* expression with no difference in the expression of *CLDN1*, *MFSD2A*, and *CLDN5* (S.Fig-4C). Together, *HDAC2* and *EZH2* ECKO data suggest that HDAC2 is critical for BBB maturation, whereas PRC2 is dispensable or is a support mechanism that is required during later BBB development.

Despite Wnt pathway activity in the adult, Wnt target genes are epigenetically repressed.

Wnt pathway has been shown to influence BBB gene expression [5]. However, this pathway activity is reported to be minimum in adults[15,17]. In our transcriptomic analysis, 67% of Wnt signaling-related genes were downregulated in adults, while 33% were upregulated (Fig-5A). Interestingly, downstream Wnt targets genes including *AXIN2*, *LEF1*, and *VEGFA* have downregulated in adult ECs, while the upstream Wnt pathway components like *FZD4/6*, *LRP5*, and *CTNNB1* were upregulated (Fig-5a). Since Wnt-regulated BBB genes such as *CLDN1*, and *ZIC3* expression was also minimum in adults, we hypothesize that the Wnt pathway is still active in adult CNS ECs while Wnt target genes are epigenetically repressed.

To test this, we activated the Wnt pathway in E13.5 and adult ECs using identical concentrations of Wnt3a ligand or GSK3B inhibitor CHIR99021. mRNA analysis revealed that Wnt target genes *AXIN2* and *LEF1* can be significantly activated in E13.5 ECs when treated with Wnt3a or CHIR99021 while no significant activation was observed in adults (Fig-5B). Further, validating this finding, transcriptome analysis on adult ECs after Wnt3a treatment showed only activation of one Wnt target gene CD44 (Fig-5C). Intriguingly, the Wnt3a treatment downregulated 16 Wnt-related genes (Fig-5C). Next using, immunohistochemistry of β-catenin in adult control and CHIR 99021 treated ECs we demonstrated that Wnt pathway activation could translocate or stabilize the Wnt transducer β-catenin into the nucleus (Fig-5D). These results indicate that the Wnt pathway is active in adult CNS ECs.

Next, we investigated whether Wnt target genes are epigenetically repressed. Wnt target genes *AXIN2* and *LEF1* were significantly upregulated in adult CNS ECs when treated with HDAC2 or EZH2 siRNA, MS-275, and in HDAC2 ECKO mutants (Fig-5E & S.F-5A,B). Furthermore, the *AXIN2* promoter in adults showed significantly increased occupancy of HDAC2, repressive histone mark H3K27me3 in adults while active histone marks H3K4me3 and H3K9ac were significantly reduced compared to E-13.5 as analyzed by ChIP-qPCR (Fig 5F). *LEF1* also showed a similar repressive histone modification (Fig-5C). Further MS-275 and LiCl (Wnt agonist) treatment increase the AXIN2 protein expression in adult cortical vessels but not by just LiCl (S.Fig-5D). Combined, our data explain the mechanism behind the low Wnt pathway in adult CNS ECs.

Low Wnt signaling epigenetically modifies the BBB genes to achieve BBB maturation.

We investigated the relevance of the low Wnt pathway to BBB development. To this we treated primary E13.5 cortical ECs with LF3 (inhibits the interaction of β-catenin and TCF4) for 48 hours. LF3 activity was confirmed by reduced mRNA expression of the Wnt target gene *AXIN2* (Fig-6A). LF3 treatment induces adult/BBB maintenance type gene expression patterns in E13.5 with a significant decrease in *CLDN1*, *ZIC3*, and *MFSD2A* expression and an increase in *CLDN5* and *SOX17* expression (Fig-6A). Another Wnt inhibitor IWR-1-endo also showed similar results (not shown) while β-catenin siRNA KD showed a similar gene expression pattern for AXIN2, CLDN1, ZIC3, and MFSD2A and no difference in *CLDN5* (S.Fig-6A).

To determine whether Wnt pathway inhibition induces epigenetic modifications on its target genes and BBB genes, we performed Chip-qPCR analysis of HDAC2, EED, and histone mark H3K27me3, H3K4me3, and H3K9ac on the promoter of Wnt target genes *AXIN2*, and *LEF1*, BBB genes *CLDN1*, *CLDN5*, *MFSD2A,* and *ZIC3*. An increased HDAC2 occupancy was observed on the promoter of *AXIN2, LEF1*, *CLDN1*, and *MFSD2A* when E13.5 ECs were treated with LF3 (Fig-6B & S.Fig-6B). *ZIC3* and *CLDN5* showed no significant difference (not shown). EED and histone mark H3K27me3 showed an increased enrichment on the *AXIN2 while LEF1* showed increased enrichment for H3K27me3 in the promoter after LF3 treatment (Fig-6B & S.Fig-6B) with no difference in other genes analyzed (not shown). *CLDN1* and *MFSD2A* showed a significant decrease in the enrichment of active histone mark H3K4me3 after treatment with LF3, whereas *CLDN5* showed a significant increase (Fig-6B). Another active histone mark H3K9ac showed significantly increased enrichment on *CLDN5* (Fig-6B) with no difference in the promoter of other genes analyzed (not shown).

We then investigate the significance of the physiological reduction of Wnt signaling on BBB maturation. For this we use Ctnnb1lox(ex3)$^{+/+}$; Cdh5-CreERT2 mice which are widely used to attain the inducible EC specific β-catenin gain of function (GOF) (Fig-6C). Upon tamoxifen treatment exon 3 of *CTNNB1* (encoding β-catenin) will be deleted leading to the expression of a stabilized form of β-catenin protein and thereby constitutive activation of canonical Wnt signaling (Fig6C). Tamoxifen was injected into the pregnant mother as explained in section 2.4.

At E-17.5 β-catenin GOF embryos showed significantly increased pial angiogenesis (Fig-6D) compared to WT. BBB permeability assay using 70KD FITC dextran revealed immature BBB in β-catenin GOF (Fig-6E & F). Confirming this result pharmacological activation of Wnt signaling by Wnt agonist LiCl also showed a significant BBB leakage compared to the control (S.Fig-6C). Together, our data suggest that a low Wnt pathway supports BBB maturations by epigenetically modifying BBB genes.

Class-I HDAC inhibitor treatment of adult CNS ECs re-establishes gene cohorts required for BBB formation.

To obtain a detailed picture of CNS EC genes regulated after MS-275 treatment we performed an mRNA-seq analysis of MS-275-treated adult ECs and compared this with the control. MS-275 treatment upregulated 48% of genes, downregulated 34% of genes, and did not alter the expression of 18% of genes. Differentially expressed genes were grouped into six relevant categories and all the categories showed a significant no.of differentially regulated genes (Fig-7A). Fig-7B illustrates the potential of MS-275 in regaining BBB, angiogenesis, Wnt, and transcription factors that are differentially regulated during development.

We next assessed whether the gene expression changes acquired by adult ECs following MS-275 treatment were reversible. To this end, we treated adult ECs with MS-275 for 48 hours and collected ECs at 48 hours of treatment and 7 days after treatment. As previously shown, *CLDN1* was significantly upregulated and *CLDN5* was significantly downregulated after 48 hours (Fig-7C). While the expression of *CLDN1* and *CLDN5* in adult ECs returned to normal in ECs collected 7 days after the treatment (Fig-7C).

Finally, we examined if human vessels show similar reactivation when treated with MS-275. To this end, the human brain vessels collected from epilepsy surgery irrespective of age and gender. We tested three genes with MS-275 treatment: *CLDN1*, *CLDN5*, and *AXIN2*. We found that MS-275 treatment significantly activates the expression of *CLDN1* and *AXIN2* compared to the control, with no difference in *CLDN5* expression (Fig-7D).

Discussion

The mechanisms that create and maintain the BBB are vitally important, but they are poorly understood. We identified EC gene cohorts that differentially support BBB formation and maintenance, describe the genetic and signaling mechanisms that establish the BBB, and present an attractive strategy to activate gene cohorts involved in BBB formation that may promote BBB repair.

MFSD2A is required for BBB formation[22], and the transcription factors *ZIC3* and *FOXF2* can induce BBB markers even in peripheral ECs[5]. Our transcriptomic analysis revealed that *CLDN1, MFSD2A*, *ZIC3*, and *FOXF2* were significantly expressed in E13.5 compared to adults, validating their requirement in BBB formation. Increased levels of *CLDN5*, *PECAM*, *ABCG1*, and *SOX17* in adult CNS ECs point to its significance in BBB maintenance. The results of our study agree with those of prior gene expression studies, but provide new concepts regarding downstream mechanisms governing BBB gene expression. Although a high level of purity is achieved in primary EC culture, we cannot exclude the fact that transcripts from other cell populations may be identified.

It is not known how BBB gene transcription is regulated. In CNS ECs, HDAC2 and PRC2 directly regulate the transcription of important BBB genes including *CLDN1*, *CLDN5*, *MFSD2A*, and *ZIC3*. While HDAC2 and PRC2 commonly occupy repressed genes, we detected them at BBB genes in both active and repressive states. This result is consistent with the established dual transcriptional role of these epigenetic regulators[25-32]. Furthermore, we present evidence that HDAC2 is required for the formation of a functional BBB and induction of anti-angiogenic signals. Even though the loss of vascular integrity and lethality at E-13.5 was reported in non-inducible conditional EZH2 KO using Tie2 Cre-mouse [33], loss of *EZH2* from E-13.5 did not significantly affect BBB permeability. These results indicate that, during the differentiation phase, HDAC2 initiates the transcriptional control, and PRC2 functions in a supporting mechanism. However, the KD of these regulators from adult CNS ECs induces a similar gene expression pattern indicating the possibility that PRC2 facilitates the epigenetic modification during later BBB development and maintenance.

Since we did not detect H3K9me3, the repression of genes involved in BBB formation is mainly mediated through H3K27me3. On varying abundance repressive histone mark H3K27me3 and active histone mark, H3K4me3 were detected in our selected BBB genes, suggesting that the promoter is modified by repressive and activating histone methylation marks. Bivalent histone states can correlate with genes transcribed at low levels, suggesting that these genes are poised for activation[34]. In addition, we detected H3K9ac at the active and repressed BBB genes. Bivalent promoters can harbor H3K9ac. Thus, among the five BBB genes analyzed, each gene exhibits a different epigenetic signature, defined by histone acetylation and methylation. These results systematically demonstrate the complexity of epigenetic regulation in BBB genes and the evidence of unique epigenetic signatures. The data presented here is not completely representing all the histone modifications on BBB genes, as other important histone modifications such as H3K27ac and H3K14ac have not been examined.

It is still unknown what mechanism confers low Wnt signaling in adult CNS ECs. Our results demonstrate that Wnt pathway components are still active in adult CNS ECs, yet the Wnt target genes are epigenetically inactive. Thus, in the adult CNS ECs, Wnt pathway activation permits stabilized β-catenin to enter the nucleus. Whether this nuclear β-catenin binds to Wnt target genes is unknown. Nevertheless, we demonstrated that HDAC2 and PRC2 repress Wnt target genes. Other studies have revealed that the basal or minimal Wnt pathway maintains adult BBB integrity, but also its activation prevents stroke-induced BBB damage. This also suggests the possibility of switching Wnt-regulated genes during development. Our transcriptomic analysis revealed the Wnt-regulated adult CNS EC genes associated with BBB (Table S2).

It is unknown how Wnt regulates BBB genes. We demonstrated that inhibiting the active Wnt pathway at E13.5 can induce the BBB gene expression pattern associated with BBB maintenance. We demonstrate that the low Wnt pathway in E13.5 causes epigenetic modifications on BBB genes mediated by HDAC2. However, the link between Wnt and epigenetic mechanisms is unclear. Since inhibiting β-catenin induce the aforementioned results, in principle, it may be mediated through β-catenin. In support of this model, β-catenin GOF during BBB development prevents the maturation of BBB and inhibits the differentiation of

angiogenic vessels which results in increased brain vascularization. Similar morphological characteristics between HDAC2 ECKO and β-catenin GOF point to its close association.

Wnt signaling in CNS ECs is believed to require Wnt7a and Wnt7b produced by neural progenitors[35-38]. Our transcriptomic data revealed significant expression of Wnt7a in E13.5 ECs with null expression in adults. Interestingly, adult CNS ECs showed significant activation of Wnt7a mRNA with MS-275 treatment (S.Fig7A). Moreover, β-catenin KD and GOF affect the expression of Wnt7a, and β -catenin staining after MS-275 treatment showed localization of β-catenin to the nucleus. These data indicate an innate mechanism of ECs to regulate the Wnt pathway.

A natural consequence of targeting epigenetic regulators or the Wnt pathway is that these mechanisms regulate numerous genes. Our mRNA sequencing results revealed that class-1 HDAC inhibitor treatment regulates EC genes associated with key functions including angiogenesis, barriergenesis, and Wnt signaling. Epidrugs are attractive therapeutics since epigenetic changes are reversible, with the potential to reestablish function after treatment. Our results support this, as the expression of *CLDN1* and *CLDN5* returns to the normal level after 7 days of treatment. Furthermore, our results illustrate the potential of MS-275 to reinstate the developmental characteristics in mice and partially in human adult CNS ECs.

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Author Contributions

JS and ST performed the experiments and analyzed the data under the supervision of PKT. IEM performed western blots and analyzed RNA-seq data. PKT conceived and supervised the project, designed, and performed experiments, interpreted the data, and wrote the manuscript. HZ primarily contributed to the idea of Wnt target gene repression in adult cortical

ECs. EHB provided critical comments on the manuscript and ChIP data. NT provide the human cortical vessels. AH and HL analyzed the RNA-seq data. All the authors critically commented on the manuscript.

Figure legends

Figure-1 Distinct cohort of EC genes regulates the formation vs maintenance of BBB. A) Workflow for transcriptomic analysis. Primary ECs were isolated from E13.5 and adult (2-3 months old) cortex followed by RNA isolation and mRNA sequencing and transcriptomic analysis. B) Comparative transcriptome analysis of ECs from E13.5 and adult cortex. Volcano plot depicting downregulated and upregulated genes in adult primary cortical ECs compared to E-13.5. Genes marked in red are significant ($P<0.05$) N=3. C) Diagram depicting the percentage of genes downregulated, and upregulated, and shows no differential expression in adult cortical primary ECs compared to E-13.5. D) Downregulated and upregulated genes were categorized with 5 important EC functions. E) Enrichment analysis revealed important BBB-related genes that were differentially regulated during development. F) Relative mRNA expression of *CLDN1*, *MFSD2A, ZIC3*, *SOX17,* and *CLDN5*. in primary cortical ECs isolated from E13.5, E-17.5, P0, P7 and adult. Significant differences are observed between E-13.5 and all consecutive stages for *CLDN1*(*** $p<0.001$, N=3/group), *MFSD2A* (*** $p<0.001$, N=3-4/group) and ZIC3(*** $p<0.001$N=3-4/group). CLDN1 showed significant differences between E-17.5 vs P7 and adult and P0 vs P7 and adult (*** $p<0.001$). ZIC3 showed a significant difference between E-17.5 vs P7 and adult es (** $p<0.01$). Significant differences are observed between E-13.5 vs P7 and adults for *CLDN5* and *SOX17* (* $p<0.001$ N=3-4/group).

Epigenetic regulators HDAC2 and PRC2 regulate the transcription of BBB genes. A) qPCR of adult primary cortical ECs treated with TSA (200nm) and MS-275(10um) 48hrs showed significantly increased mRNA expression of *CLDN1* compared to DMSO treated control while

CLDN5 was significantly decreased with MS-275 treatment when compared to control (* $p<0.001$ vs Control #$p<0.001$ vs TSA treatment N=3/group). B) mRNA expression level of class-I HDAC family members in adult primary cortical ECs. Expression was normalized to housekeeping genes GAPDH and HDAC1. Significant mRNA expression of HDAC2 was observed in adult primary cortical ECs compared to other Class-I HDACs (* $p<0.05$ vs HDAC1,3 and 8 N=3/group). HDAC1 showed significantly higher expression compared to HDAC3 and HDAC8 (* $p<0.05$) and HDAC8 showed significantly higher expression compared to HDAC3 (* $p<0.05$). C) Effect of HDAC2 siRNA on BBB gene expression in adult cortical ECs. Using lipofectamine adult cortical ECs were transfected with HDAC2 siRNA (500µg). qPCR analysis revealed that compared to control siRNA treated group HDAC2 siRNA treated ECs showed significantly increased expression of *CLDN1*(* $p<0.001$), *MFSD2A* (* $p<0.05$) and *ZIC3* (* $p<0.05$) while *CLDN5* (* $p<0.05$) showed significantly decreased expression. N=3/group D) HDAC2 occupancy of the indicated chromatin regions in primary cortical ECs from E-13.5 and adult. Occupancy was measured by ChIP followed by quantitative PCR (ChIP-qPCR). Chromatin regions are named by the adjacent gene and the distance to the TSS. (* $p<0.05$ vs IgG N=3-4/group) E) qPCR analysis of *CLDN1*, *CLDN5*, *MFSD2A,* and ZIC3 in EZH2 and control siRNA-treated adult primary cortical ECs. Compared to control siRNA treated group EZH2 siRNA treated ECs showed significantly increased expression of *CLDN1* (* $p<0.001$), *MFSD2A* (* $p<0.001$) and *ZIC3* (* $p<0.05$) while *CLDN5* (* $p<0.05$) showed significantly decreased expression. N=3/group. ChIP-qPCR analysis of PRC2 subunit EED on indicated chromatin regions and genes in E13.5 and adult primary cortical ECs. * $p<0.05$ vs IgG, N=3-4/ group. Schematic representation of HDAC2 and PRC2 binding on indicated genes in cortical ECs at E-13.5 and adult.

Figure-3 BBB genes exhibit diverse post-translational histone modifications. A) ChIP-qPCR analysis of histone marks H3K27me3, H3K4me3, and H3K9ac on 1KB region *CLDN1* gene in E13.5 and adult cortical ECs. *CLDN1* gene was downregulated in ECs during development. The ChIP signals were normalized to IgG. B) ChIP-qPCR showing the H3K27me3, H3K4me3, and H3K9ac density at the CLDN5 gene in E13.5 and adult cortical ECs. CLDN5 gene was upregulated during development. Data are shown as mean ± SD. ***$p < 0.001$, **$p < 0.01$, *$p < 0.05$ vs IgG & ###$p < 0.001$, ##$p < 0.01$, #$p < 0.05$ vs respective E-13.5 histone mark. N = 3-4/group. C)

Schematic representation of H3K27me3, H3K4me3, and H3K9ac binding density on *CLDN1*, *CLDN5*, *MFSD2A*, and *ZIC3* in E13.5 and adult primary cortical ECs. Shape size indicates the binding density in the indicated chromatin regions.

Figure-4. HDAC2 activity is required for the formation of a functional BBB while PRC2 is dispensable. A) Effect of deletion of HDAC2 from the ECs during embryonic development. Schematic representation of breeding scheme and generation of EC-specific KO of HDAC2. Tamoxifen was delivered to the pregnant mother at E-12.5 and brains were harvested at E-17.5. B) Representative phase microscopy images of the dorsal surface and ventral surface of the brain at E-17.5. HDAC2 ECKO shows a significant increase in pia vessels and as pointed out by black arrows dorsal surface. Black arrows in the ventral surface showed dilated vessels. C) BBB permeability assay using 70KD tracer and isolectin B4 (IB4) staining to image the vessels. In HDAC2 ECKO Green fluorescent tracer was leaked out of the vessels as indicated by white arrows. 10x images are acquired and merged using tile scanning. Scale bar, 500µm D) Vessel percentage area of HDAC2 ECKO was significantly higher than WT (* $p<0.001$ vs control N=10). A significant increase in fluorescent intensity was quantified in HDAC2 ECKO compared to WT (* $p<0.0001$ vs control N=3 for W.T and N=5 for HDAC2 ECKO). E) Schematic representation of breeding scheme and generation of EZH2 ECKO. Tamoxifen was injected as same as for HDAC2 ECKO. F) Representative phase microscopic image of WT and EZH2 ECKO. As shown by black arrows EZH2 ECKO brain shows dilated vessels with no visible increase in pial angiogenesis. G) BBB permeability assay using 70KD-FITC Dextran shows subtle leakage of tracer out of the vessels in EZH2 ECKO, represented using white arrows. IB4 staining reveals the vessels in the brain. 10x images are acquired and merged using tile scanning. Scale bar, 500 µm H) Quantification of vessel percentage area (W.T N=9 EZH2 ECKO N=10) and fluorescent intensity didn't show any significant difference between WT and EZH2 ECKO (W.T N=5 EZH2 ECKO N=5).

Figure-5 Wnt pathway is active in adult CNS ECs but Wnt target genes are epigenetically repressed. A) Diagram depicting the percentage of Wnt-related genes downregulated, and upregulated adult cortical primary ECs compared to E-13.5. Selected important Wnt-related genes are shown. B) Ligand-independent transcriptional repression of Wnt target genes. In primary cortical ECs from E-13.5, activating the Wnt pathway with Wnt3a (200ng/mL) or CHIR-

99021 (5uM) for 48 hrs. caused increased mRNA expression of Wnt target genes *AXIN2* and *LEF1* (measured via qRT-PCR). However, activation of the Wnt pathway in primary adult mouse brain ECs does not increase Wnt target gene expressions. *p<0.001 vs E-13.5 control, ns-no significant difference n=3/group. C) mRNA sequencing was performed in control and Wnt3a (200ng/mL) treated adult primary cortical ECs. Differentially expressed genes were categorized into six selected categories that are important to CNS endothelial cells. D) Immunofluorescence staining of β-catenin (green) in control and Wnt agonist Chir-99021 treated endothelial cells. White arrows indicate the nuclear localization of β-catenin to the nucleus. 20x images are acquired and cropped and enlarged. Scale bar, 1 µm. E) Adult primary cortical ECs transfected with control, HDAC2 & EZH2 siRNA showed significant upregulation of Wnt target genes AXIN2. ***p<0.001 vs control siRNA & #p<0.05 vs control siRNA. N=3/group F) HDAC2, histone marks H3K27me3, H3K4me3, and H3K9ac occupancy on the AXIN2 TSS regions in primary cortical ECs from E-13.5 and adult. Occupancy was measured by ChIP-qPCR. N=3/group ***p < 0.001, **p < 0.01, *p < 0.05 vs IgG & #p < 0.05 vs H3K4me3 E-13.5.

Figure-6. Low Wnt signaling epigenetically modifies the BBB genes to achieve BBB maturation. A) Effect of Wnt pathway inhibition on BBB genes in E13.5 primary cortical ECs. E-13.5 ECs were treated with LF3(50um) for 48hrs to inhibit the Wnt pathway. Significantly decreased mRNA expression of *AXIN2* confirmed the reduced Wnt pathway. mRNA expression of *CLDN1*, *MFSD2A* and *ZIC3* was significantly decreased *CLDN5* and *SOX17* expression was significantly increased after LF3 treatment. Data are shown as mean ± SD. ***p < 0.001, **p < 0.01, *p < 0.05 vs E13.5 control N=3/group. B) Wnt pathway inhibition *via* LF3 induce epigenetic modifications in target gene *AXIN2* and BBB genes *CLDN1*, *MFSD2A*, and CLDN5. First row- *AXIN2* showed significant enrichment of HDAC2 and EED in LF3 treated E13.5 ECs when compared to control (*p < 0.05 vs IgG). Histone mark H3K27me3 showed significant enrichment in both conditions compared to IgG however LF3 treated ECs showed significantly increased enrichment compared to the control. **p < 0.01 vs IgG, #p < 0.05 vs control N=3-4/group. Second row- LF3 treated E13.5 ECs showed significant enrichment of HDAC2 in *CLDN1* TSS (*p < 0.05 vs IgG N=3-4/group). *MFSD2A* showed significant enrichment in both conditions while LF3 treatment showed a significantly increased enrichment compared to the control (*p < 0.05 vs

IgG & ##$p < 0.01$ vs control N=3/group). *CLDN5* didn't show any significant difference in HDAC2 binding (not shown) while active histone marks H3K9ac showed significant enrichment in both conditions with an increased enrichment with LF3 treatment (***$p < 0.001$, *$p < 0.05$ vs IgG & #$p < 0.05$ vs control N=3/group). Third row- H3K4me3 ChIP-qpcr on the TSS region of *CDLN1*, *MFSD2A* and *CLDN5* showed a significant enrichment in both conditions with a decreased enrichment with LF3 treatment on *CDLN1* and *MFSD2A* and an increased enrichment with LF3 treatment on *CLDN5*. ***$p < 0.001$, *$p < 0.05$ vs IgG & ##$p < 0.01$, #$p < 0.05$ vs control N=3-4/group. C) Schematic representation of breeding scheme and generation of EC-specific gain of function (GOF) of β-catenin. Tamoxifen was delivered to the pregnant mother at E-12.5 and brains were harvested at E-17.5. D) Representative Phase microscopy image of the dorsal brain from W.T and β-catenin-GOF. Images in the square box were enlarged to show the increase in pial vessel angiogenesis. E) BBB permeability assay using the 70KD FITC-Dextran tracer. Cortical vessels are stained using IB4.10X tile scanning images were acquired and merged. FITC dextran was leaked out of the vessels in the brain of β-catenin GOF compared to W.T. 10x images are acquired and merged using tile scanning. Scale bar, 500 µm F) Quantification of brain vessels percentage area (***$p < 0.001$ vs WT N=4-5/group) and green fluorescent intensity showed a significant increase in β-catenin GOF compared to W.T. ***$p < 0.001$, *$p < 0.05$ vs WT N=5-6/group.

Figure-7 In adult ECs, treatment with MS-275 induces partial reactivation of BBB and angiogenesis-supporting gene cohorts. A) Diagram depicting the percentage of genes downregulated, upregulated and shows no differential expression in MS-275 treated adult cortical ECs compared to Control. Downregulated and upregulated genes were categorized with 6 important EC functions. B) Heat map of expression values (Z score) for differentially expressed genes (*p adj < 0.05) in E13.5, adult control, and adult treated with MS-275. Five different gene categories which show a significant difference between E13.5 vs adult control and Adult Control vs Adult MS-275 treatment are presented. N=3 for E13.5 and adult control, N=2 for MS-275. C) Adult primary cortical ECs treated with MS-275 for 48 hrs. showed significant activation of *CLDN1* and downregulation of *Cldn5*. mRNA analysis on ECs after 7 days of withdrawing MS-275 showed a significant reversal of expression back to normal. *p<0.001 compared to control 48

hrs. # p<0.001 compared to MS-275 48 hrs. D) Representative phase contrast image of human temporal lobe vessels in culture collected from epilepsy patients who underwent surgery. MS-275 treated human vessels showed significantly increased mRNA expression of *CLDN1*, and *AXIN2* when compared to control. ***p*<0.01 and **p*<0.05 N=5/group. No significant difference was observed in *CLDN5* expression. E) Schematic diagram illustrating the mechanisms underlying BBB formation and maintenance led by epigenetic regulators HDAC2 and PRC2. EC gene cohorts that support BBB formation are epigenetically repressed by HDAC2 and PRC2 during development. Active Wnt signaling supports the expression of gene cohorts required for the BBB formation, while a reduction in Wnt signaling recruits HDAC2 to these gene cohorts to support the formation of a functional or intact BBB. Inhibiting HDAC2 in adult ECs induce the reacquisition of gene cohorts that support BBB formation and thus represents a potential therapeutic opportunity to repair damaged BBB.

References

1 Profaci, C. P., Munji, R. N., Pulido, R. S. & Daneman, R. The blood-brain barrier in health and disease: Important unanswered questions. *J Exp Med* **217** (2020). https://doi.org:10.1084/jem.20190062

2 Sabbagh, M. F. *et al.* Transcriptional and epigenomic landscapes of CNS and non-CNS vascular endothelial cells. *Elife* **7** (2018). https://doi.org:10.7554/eLife.36187

3 Daneman, R. *et al.* The mouse blood-brain barrier transcriptome: a new resource for understanding the development and function of brain endothelial cells. *PLoS One* **5**, e13741 (2010). https://doi.org:10.1371/journal.pone.0013741

4 Corada, M. *et al.* Fine-Tuning of Sox17 and Canonical Wnt Coordinates the Permeability Properties of the Blood-Brain Barrier. *Circ Res* **124**, 511-525 (2019). https://doi.org:10.1161/CIRCRESAHA.118.313316

5 Hupe, M. *et al.* Gene expression profiles of brain endothelial cells during embryonic development at bulk and single-cell levels. *Sci Signal* **10** (2017). https://doi.org:10.1126/scisignal.aag2476

6 Allis, C. D. & Jenuwein, T. The molecular hallmarks of epigenetic control. *Nat Rev Genet* **17**, 487-500 (2016). https://doi.org:10.1038/nrg.2016.59

7 Grunstein, M. Histone acetylation in chromatin structure and transcription. *Nature* **389**, 349-352 (1997). https://doi.org:10.1038/38664

8 Li, Y. & Seto, E. HDACs and HDAC Inhibitors in Cancer Development and Therapy. *Cold Spring Harb Perspect Med* **6** (2016). https://doi.org:10.1101/cshperspect.a026831

9 Yoon, S. & Eom, G. H. HDAC and HDAC Inhibitor: From Cancer to Cardiovascular Diseases. *Chonnam Med J* **52**, 1-11 (2016). https://doi.org:10.4068/cmj.2016.52.1.1

10 He, X. *et al.* Medicinal chemistry updates of novel HDACs inhibitors (2020 to present). *Eur J Med Chem* **227**, 113946 (2022). https://doi.org:10.1016/j.ejmech.2021.113946

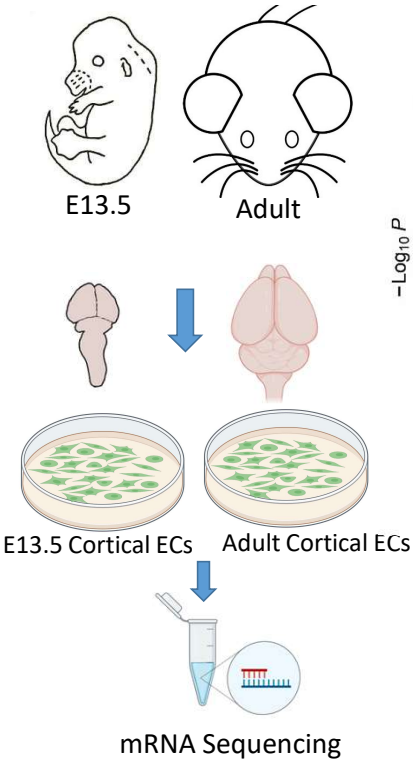
11 Kuzmichev, A., Nishioka, K., Erdjument-Bromage, H., Tempst, P. & Reinberg, D. Histone methyltransferase activity associated with a human multiprotein complex containing the Enhancer of Zeste protein. *Genes Dev* **16**, 2893-2905 (2002). https://doi.org:10.1101/gad.1035902
12 Cao, R. *et al.* Role of histone H3 lysine 27 methylation in Polycomb-group silencing. *Science* **298**, 1039-1043 (2002). https://doi.org:10.1126/science.1076997
13 Straining, R. & Eighmy, W. Tazemetostat: EZH2 Inhibitor. *J Adv Pract Oncol* **13**, 158-163 (2022). https://doi.org:10.6004/jadpro.2022.13.2.7
14 Daneman, R. *et al.* Wnt/beta-catenin signaling is required for CNS, but not non-CNS, angiogenesis. *Proc Natl Acad Sci U S A* **106**, 641-646 (2009). https://doi.org:10.1073/pnas.0805165106
15 Liebner, S. *et al.* Wnt/beta-catenin signaling controls development of the blood-brain barrier. *J Cell Biol* **183**, 409-417 (2008). https://doi.org:10.1083/jcb.200806024
16 Stenman, J. M. *et al.* Canonical Wnt signaling regulates organ-specific assembly and differentiation of CNS vasculature. *Science* **322**, 1247-1250 (2008). https://doi.org:10.1126/science.1164594
17 Ma, S., Kwon, H. J., Johng, H., Zang, K. & Huang, Z. Radial glial neural progenitors regulate nascent brain vascular network stabilization via inhibition of Wnt signaling. *PLoS Biol* **11**, e1001469 (2013). https://doi.org:10.1371/journal.pbio.1001469
18 Reis, M. *et al.* Endothelial Wnt/beta-catenin signaling inhibits glioma angiogenesis and normalizes tumor blood vessels by inducing PDGF-B expression. *J Exp Med* **209**, 1611-1627 (2012). https://doi.org:10.1084/jem.20111580
19 Wang, Y. *et al.* Beta-catenin signaling regulates barrier-specific gene expression in circumventricular organ and ocular vasculatures. *Elife* **8** (2019). https://doi.org:10.7554/eLife.43257
20 Gastfriend, B. D. *et al.* Wnt signaling mediates acquisition of blood-brain barrier properties in naive endothelium derived from human pluripotent stem cells. *Elife* **10** (2021). https://doi.org:10.7554/eLife.70992
21 Tran, K. A. *et al.* Endothelial beta-Catenin Signaling Is Required for Maintaining Adult Blood-Brain Barrier Integrity and Central Nervous System Homeostasis. *Circulation* **133**, 177-186 (2016). https://doi.org:10.1161/CIRCULATIONAHA.115.015982
22 Ben-Zvi, A. *et al.* Mfsd2a is critical for the formation and function of the blood-brain barrier. *Nature* **509**, 507-511 (2014). https://doi.org:10.1038/nature13324
23 Delcuve, G. P., Khan, D. H. & Davie, J. R. Roles of histone deacetylases in epigenetic regulation: emerging paradigms from studies with inhibitors. *Clin Epigenetics* **4**, 5 (2012). https://doi.org:10.1186/1868-7083-4-5
24 Marks, P. A. Histone deacetylase inhibitors: a chemical genetics approach to understanding cellular functions. *Biochim Biophys Acta* **1799**, 717-725 (2010). https://doi.org:10.1016/j.bbagrm.2010.05.008
25 Ferrari, K. J. *et al.* Polycomb-dependent H3K27me1 and H3K27me2 regulate active transcription and enhancer fidelity. *Mol Cell* **53**, 49-62 (2014). https://doi.org:10.1016/j.molcel.2013.10.030
26 Wang, X. *et al.* Molecular analysis of PRC2 recruitment to DNA in chromatin and its inhibition by RNA. *Nat Struct Mol Biol* **24**, 1028-1038 (2017). https://doi.org:10.1038/nsmb.3487
27 Greer, C. B. *et al.* Histone Deacetylases Positively Regulate Transcription through the Elongation Machinery. *Cell Rep* **13**, 1444-1455 (2015). https://doi.org:10.1016/j.celrep.2015.10.013
28 Jahan, S., Sun, J. M., He, S. & Davie, J. R. Transcription-dependent association of HDAC2 with active chromatin. *J Cell Physiol* **233**, 1650-1657 (2018). https://doi.org:10.1002/jcp.26078

29 Somanath, P., Herndon Klein, R. & Knoepfler, P. S. CRISPR-mediated HDAC2 disruption identifies two distinct classes of target genes in human cells. *PLoS One* **12**, e0185627 (2017). https://doi.org:10.1371/journal.pone.0185627
30 Wang, Z. *et al.* Genome-wide mapping of HATs and HDACs reveals distinct functions in active and inactive genes. *Cell* **138**, 1019-1031 (2009). https://doi.org:10.1016/j.cell.2009.06.049
31 Zupkovitz, G. *et al.* Negative and positive regulation of gene expression by mouse histone deacetylase 1. *Mol Cell Biol* **26**, 7913-7928 (2006). https://doi.org:10.1128/MCB.01220-06
32 Kaneko, S., Son, J., Shen, S. S., Reinberg, D. & Bonasio, R. PRC2 binds active promoters and contacts nascent RNAs in embryonic stem cells. *Nat Struct Mol Biol* **20**, 1258-1264 (2013). https://doi.org:10.1038/nsmb.2700
33 Delgado-Olguin, P. *et al.* Ezh2-mediated repression of a transcriptional pathway upstream of Mmp9 maintains integrity of the developing vasculature. *Development* **141**, 4610-4617 (2014). https://doi.org:10.1242/dev.112607
34 Bernstein, B. E. *et al.* A bivalent chromatin structure marks key developmental genes in embryonic stem cells. *Cell* **125**, 315-326 (2006). https://doi.org:10.1016/j.cell.2006.02.041
35 Vallon, M. *et al.* A RECK-WNT7 Receptor-Ligand Interaction Enables Isoform-Specific Regulation of Wnt Bioavailability. *Cell Rep* **25**, 339-349 e339 (2018). https://doi.org:10.1016/j.celrep.2018.09.045
36 Eubelen, M. *et al.* A molecular mechanism for Wnt ligand-specific signaling. *Science* **361** (2018). https://doi.org:10.1126/science.aat1178
37 Kuhnert, F. *et al.* Essential regulation of CNS angiogenesis by the orphan G protein-coupled receptor GPR124. *Science* **330**, 985-989 (2010). https://doi.org:10.1126/science.1196554
38 Cho, C., Smallwood, P. M. & Nathans, J. Reck and Gpr124 Are Essential Receptor Cofactors for Wnt7a/Wnt7b-Specific Signaling in Mammalian CNS Angiogenesis and Blood-Brain Barrier Regulation. *Neuron* **95**, 1056-1073 e1055 (2017). https://doi.org:10.1016/j.neuron.2017.07.031

Figure-1

Comparative transcriptome analysis of cortical ECs from E13.5 and adult

A



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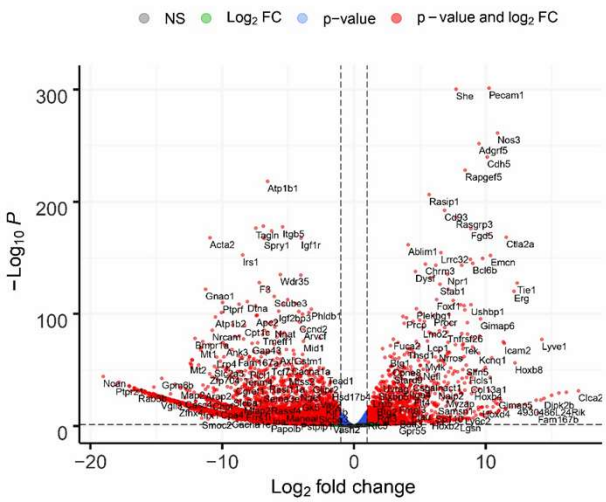

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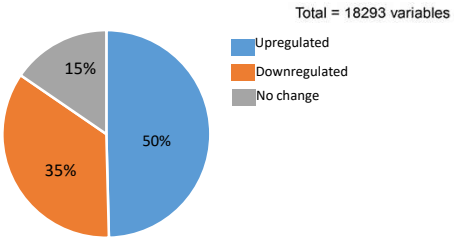

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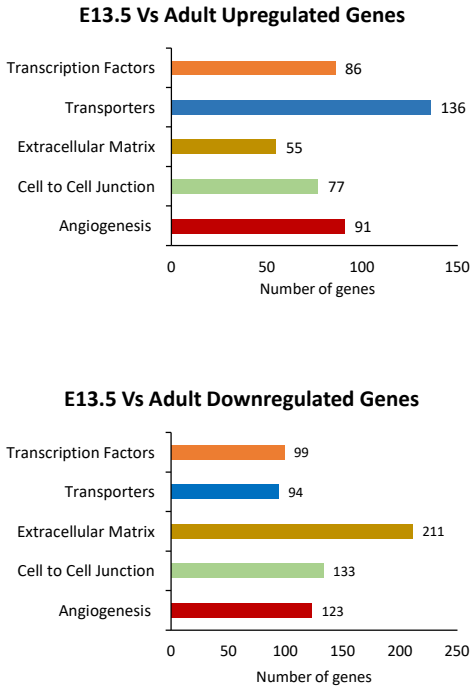


E

Selected differentially regulated BBB genes during development

Adherens/Tight Junctions

Downregulated	Upregulated
CLDN1	*CLDN5*
CLDN11	*JAM2*
CLDN12	*PECAM1*
MARVELD3	*ESAM*

BBB Transporters

Downregulated	Upregulated
MFSD2A	*CAV1*
AQP4	*ABCG1*
ABCG2	*SLC7A7*

Transcription Factors

Downregulated	Upregulated
ZIC3	*SOX17*
FOXF2	*ERG*
LEF1	*SOX18*

F

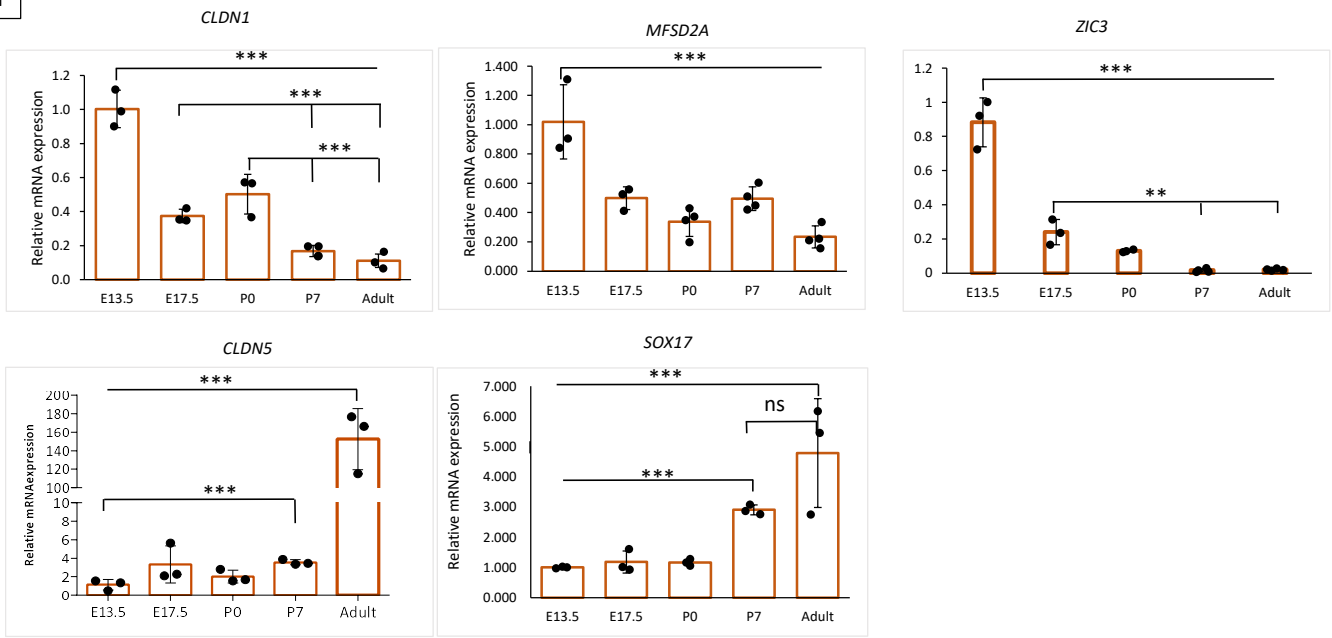
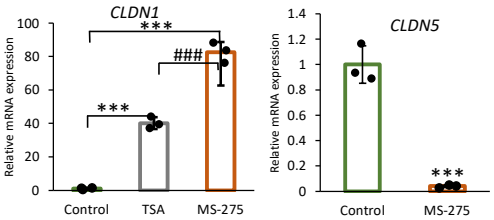


Figure-2

HDAC2 mediated transcriptional repression of BBB genes

A

Class-I HDAC regulate BBB gene expression



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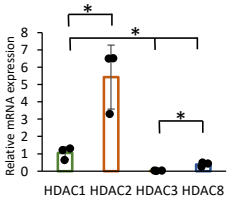
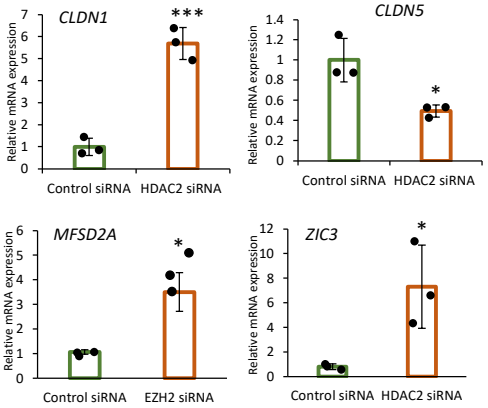
HDAC2 shows significant expression in adult ECs



C

HDAC2 Knockdown in adult cortical ECs establish gene expression pattern similar to E13.5



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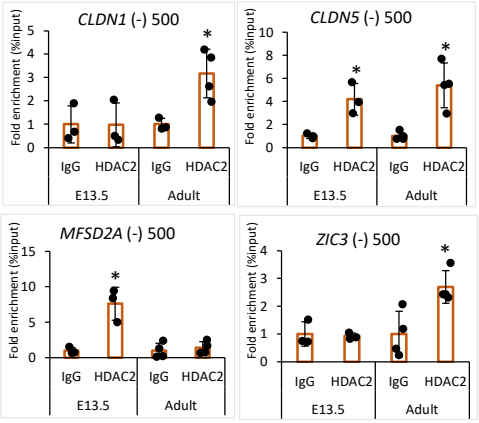
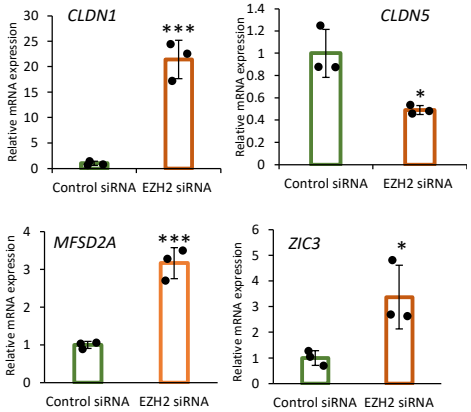
HDAC2 regulate repression by binding to BBB genes



PRC2 mediated repression of BBB genes

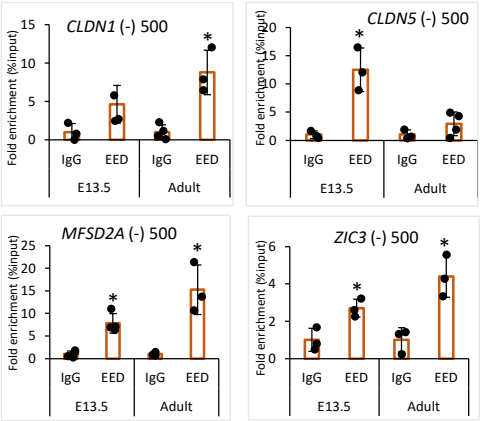
E

EZH2 knockdown also establish a E13.5 like gene expression pattern



F

PRC2 subunit EED binds to BBB genes to regulated its expression



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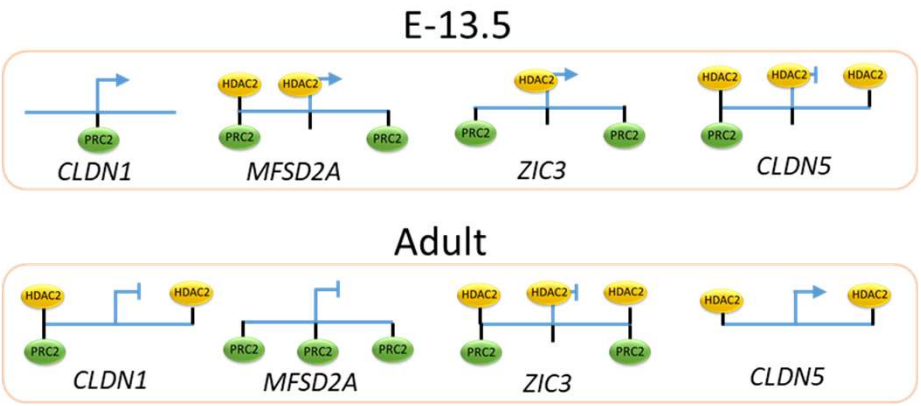
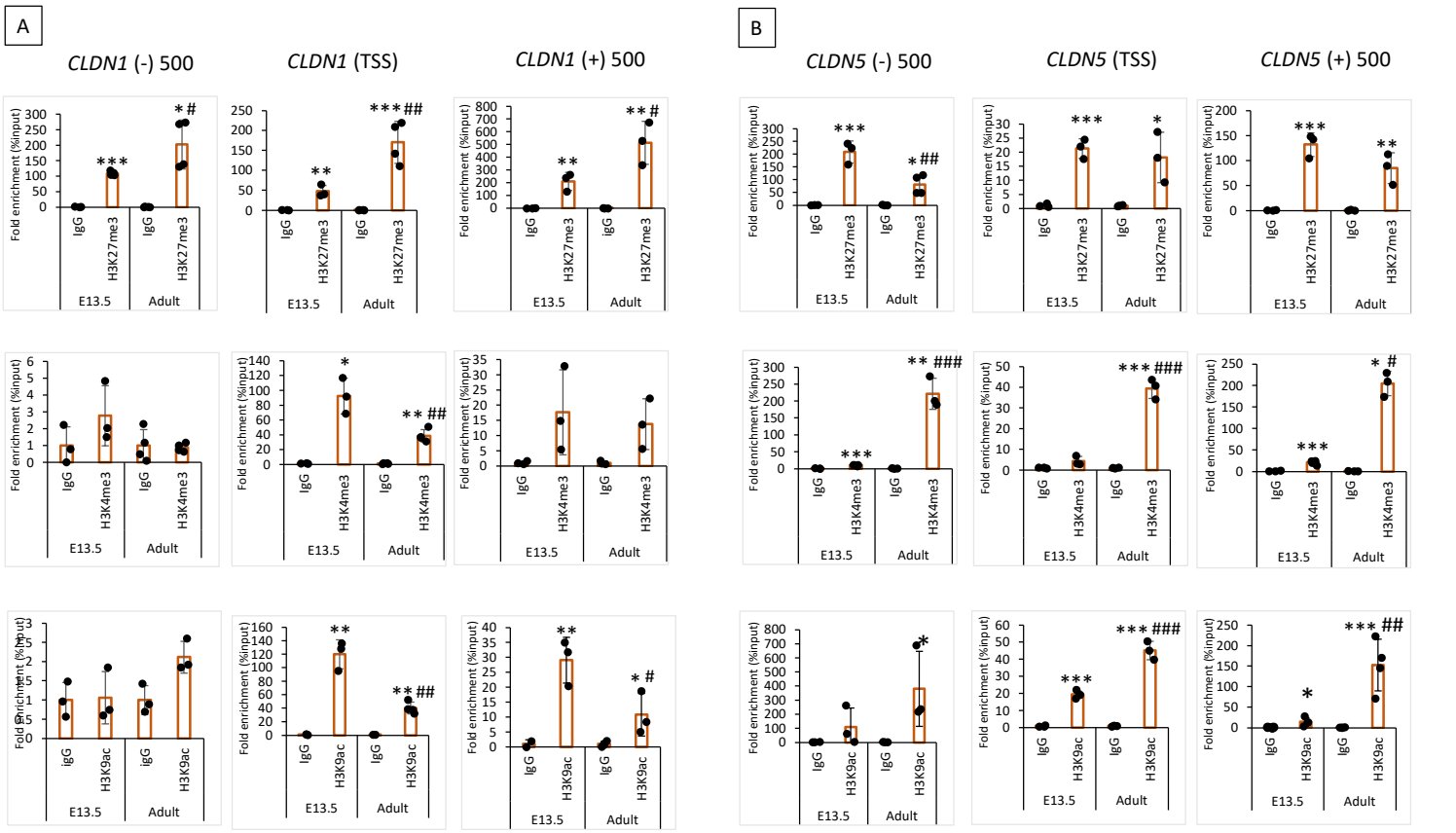


Figure-3


A
CLDN1 (-) 500
CLDN1 (TSS)
CLDN1 (+) 500
B
CLDN5 (-) 500
CLDN5 (TSS)
CLDN5 (+) 500
Fold enrichment (%input)
IgG
H3K27me3
H3K4me3
H3K9ac
E13.5
Adult

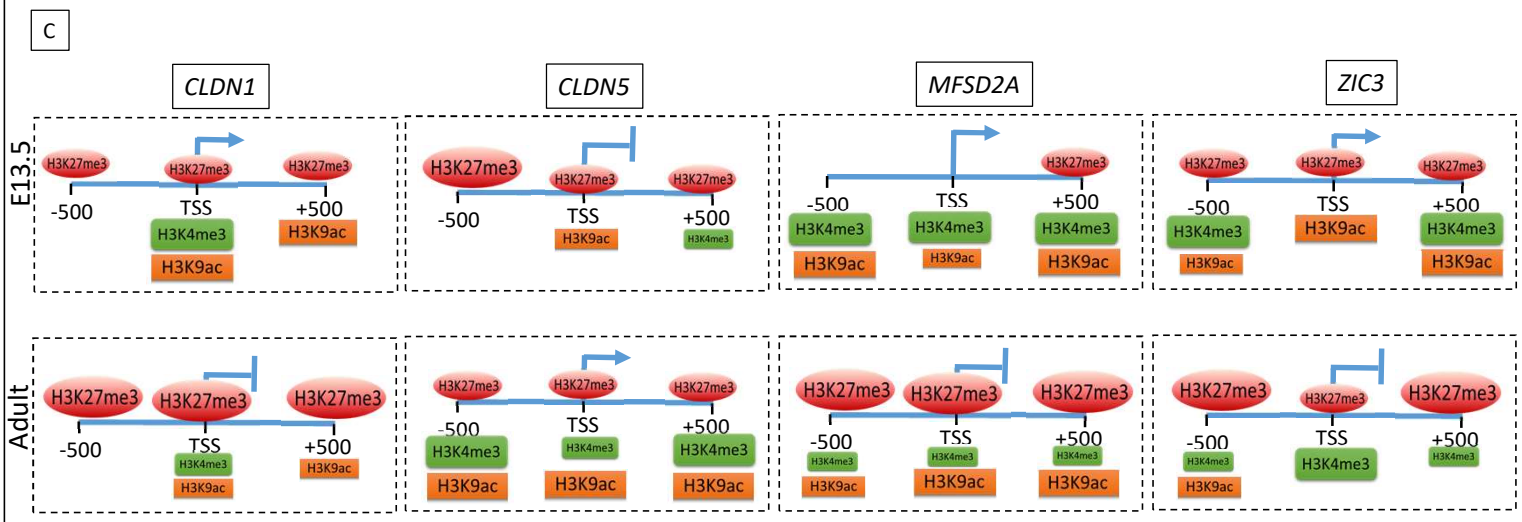

C
CLDN1
CLDN5
MFSD2A
ZIC3
E13.5
Adult
H3K27me3
H3K4me3
H3K9ac
-500
TSS
+500

Figure-4

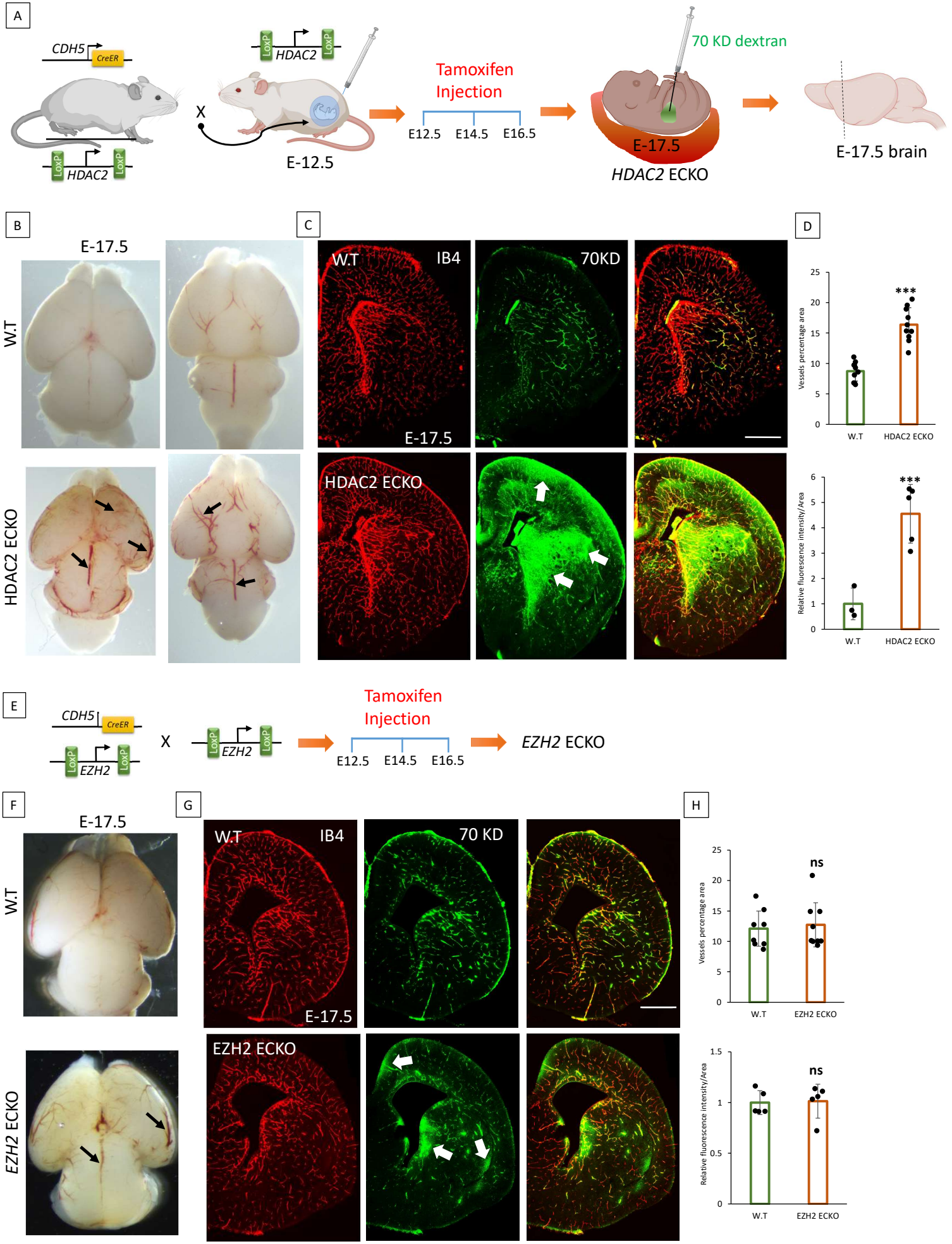
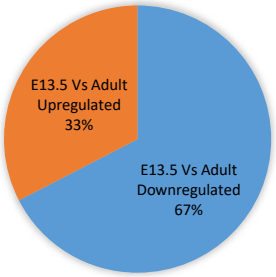

A
CDH5
CreER
LoxP
HDAC2
LoxP
X
LoxP
HDAC2
LoxP
E-12.5
Tamoxifen
Injection
E12.5
E14.5
E16.5
70 KD dextran
E-17.5
HDAC2 ECKO
E-17.5 brain
B
E-17.5
W.T
HDAC2 ECKO
C
W.T
IB4
70KD
E-17.5
HDAC2 ECKO
D
Vessels percentage area
***
W.T
HDAC2 ECKO
Relative fluorescence intensity/Area
***
W.T
HDAC2 ECKO
E
CDH5
CreER
LoxP
EZH2
LoxP
X
LoxP
EZH2
LoxP
Tamoxifen
Injection
E12.5
E14.5
E16.5
EZH2 ECKO
F
E-17.5
W.T
EZH2 ECKO
G
W.T
IB4
70 KD
E-17.5
EZH2 ECKO
H
Vessels percentage area
ns
W.T
EZH2 ECKO
Relative fluorescence intensity/Area
ns
W.T
EZH2 ECKO

Figure-5

A

E13.5 Vs Adult differentially regulated Wnt genes



Downregulated

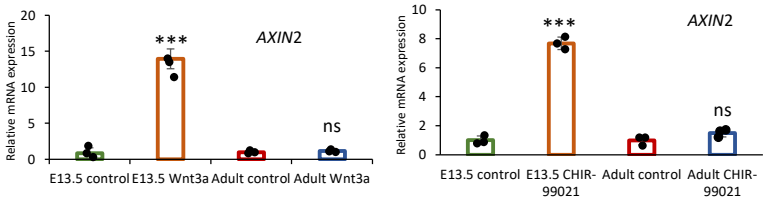
- *Axin2*
- *Lef1*
- *Wnt7a/7b*
- *Tcf7*

Upregulated

- *Fzd4/6*
- *LRP5*
- *Ctnnb1*
- *Sox17*

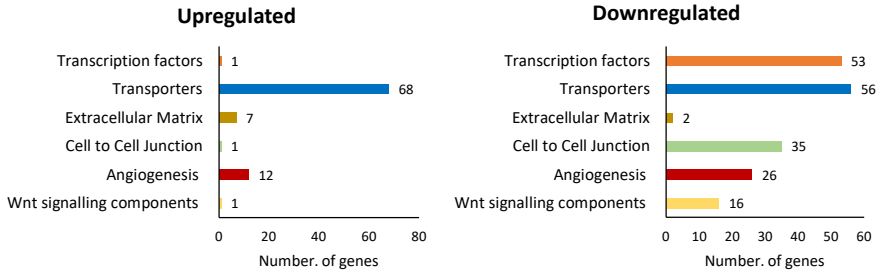
B

Independent of ligand Wnt target genes are not active



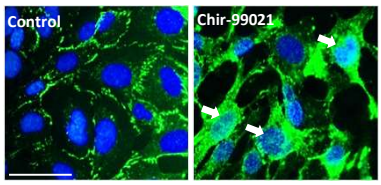
C

Effect of Wnt3a treatment in adult cortical ECs



D

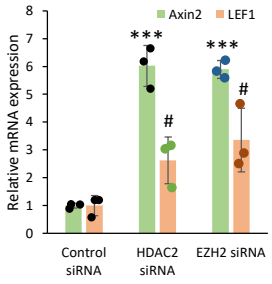
Nuclear localization of β-catenin in adult ECs



β-catenin/DAPI

Epigenetic repression of Wnt target genes in the adult CNS ECs

E



F

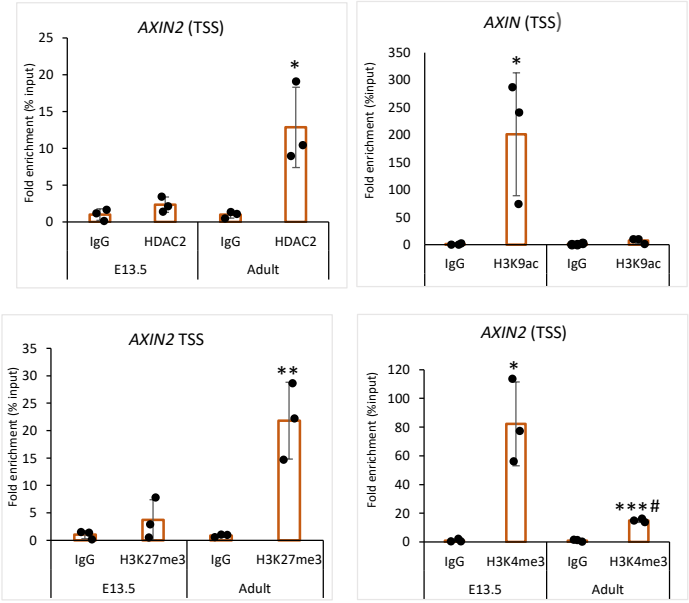
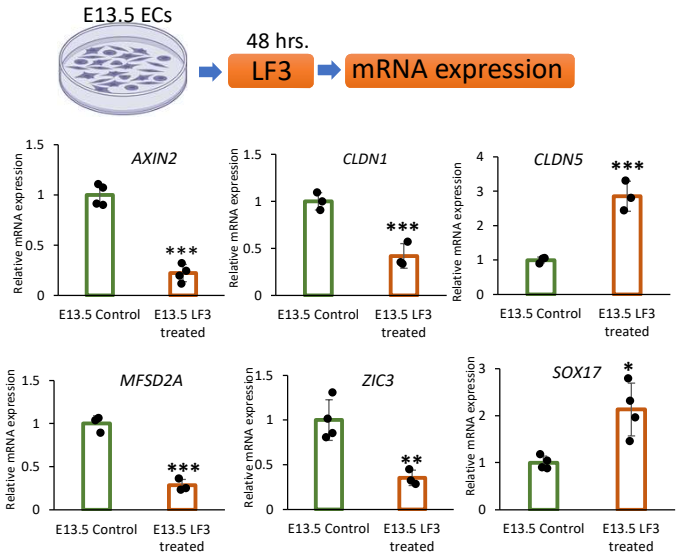
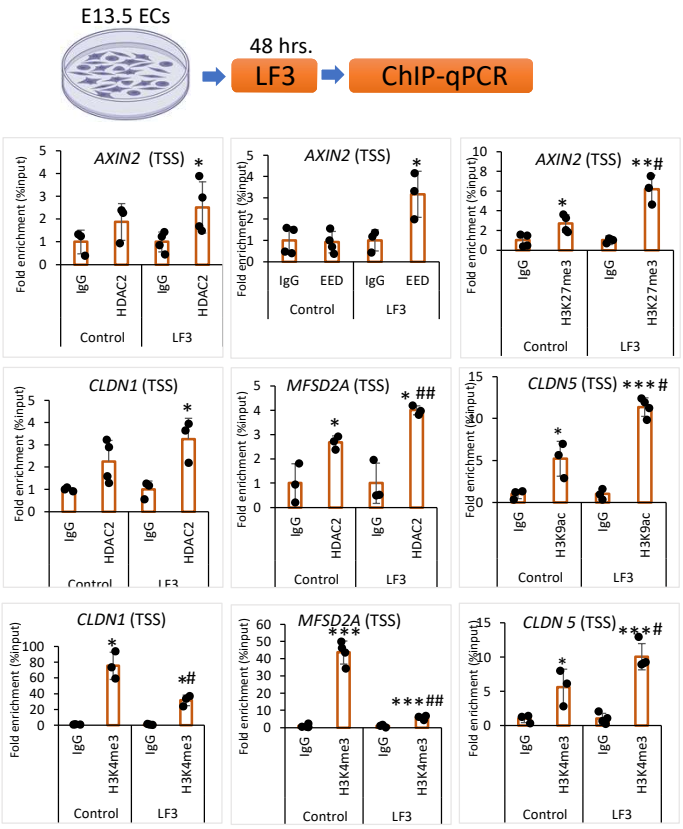


Figure-6

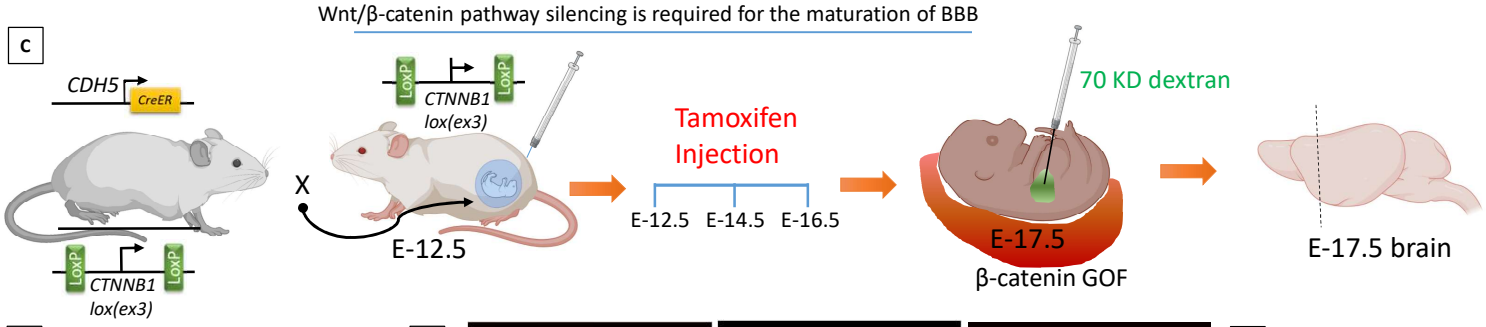
A Inhibiting Wnt/β-catenin pathway in E-13.5 ECs induces adult like BBB gene expression pattern.



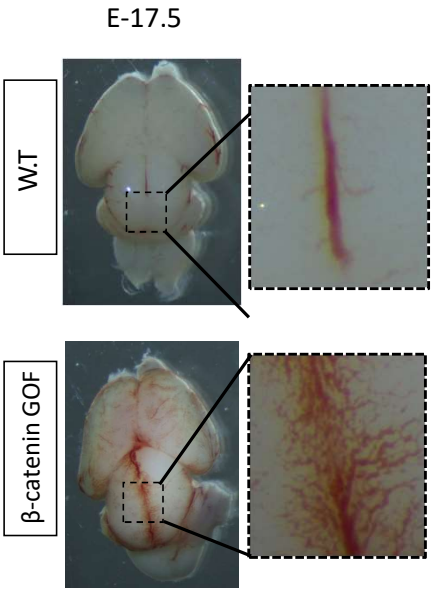
B Wnt/β-catenin pathway inhibition initiate epigenetic modifications in BBB and Wnt target genes.



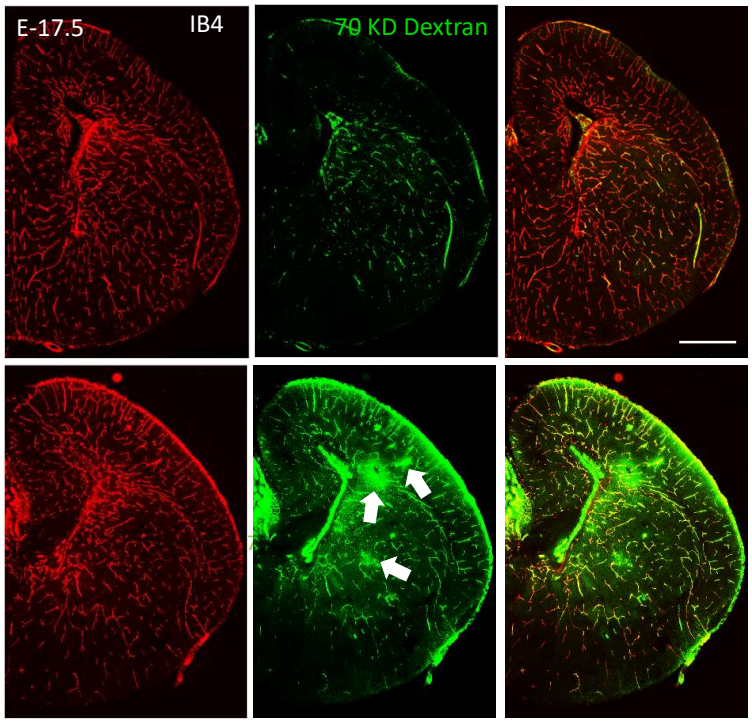
C Wnt/β-catenin pathway silencing is required for the maturation of BBB



D



E



F

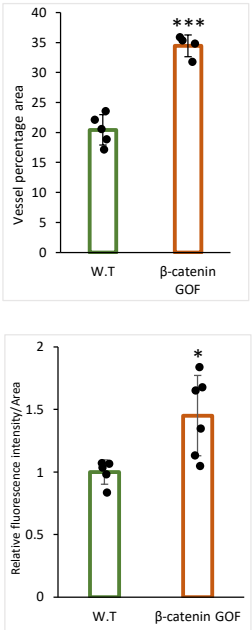
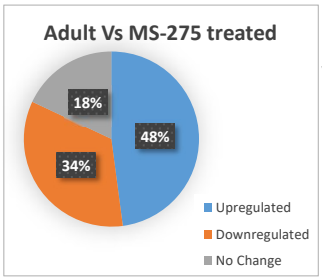
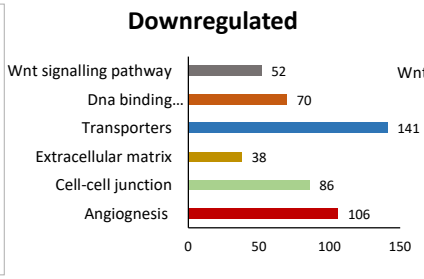


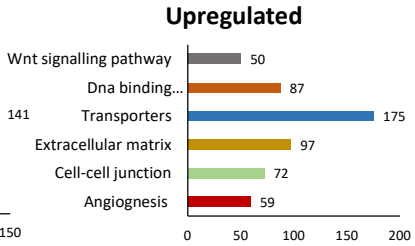
Figure-7

A

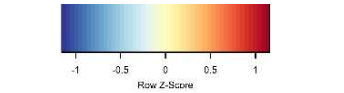
Important differentially regulated EC genes in adult with MS-275 treatment

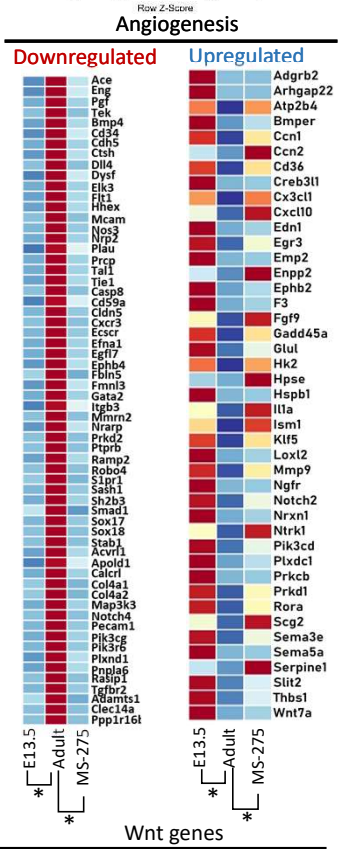


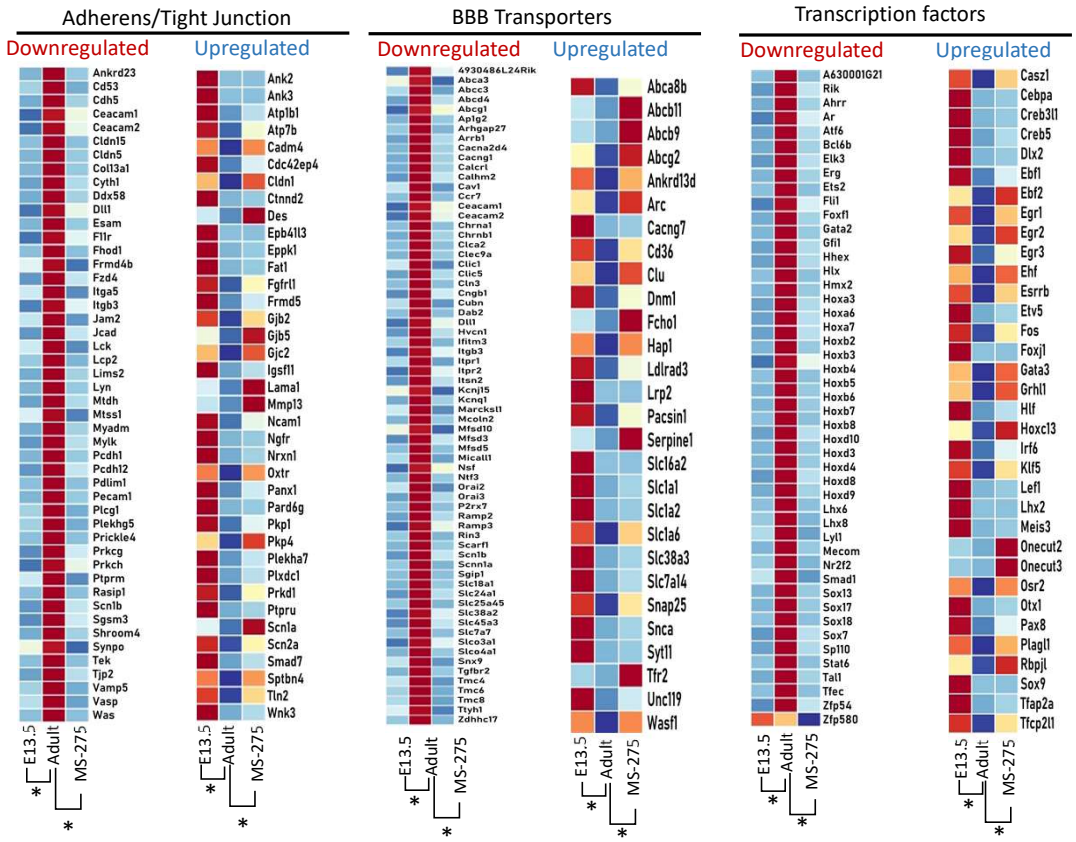


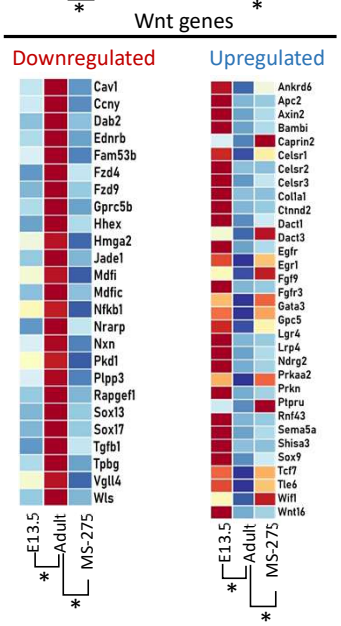


B



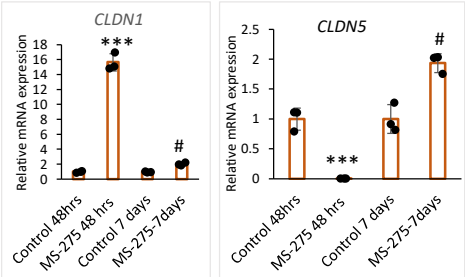






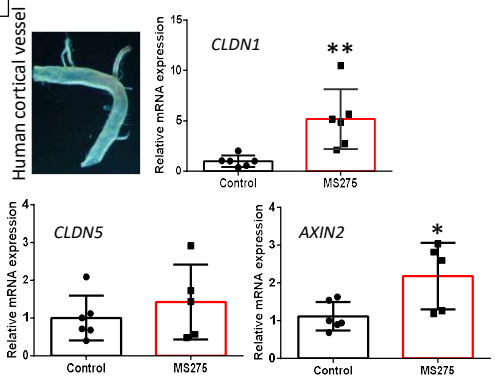
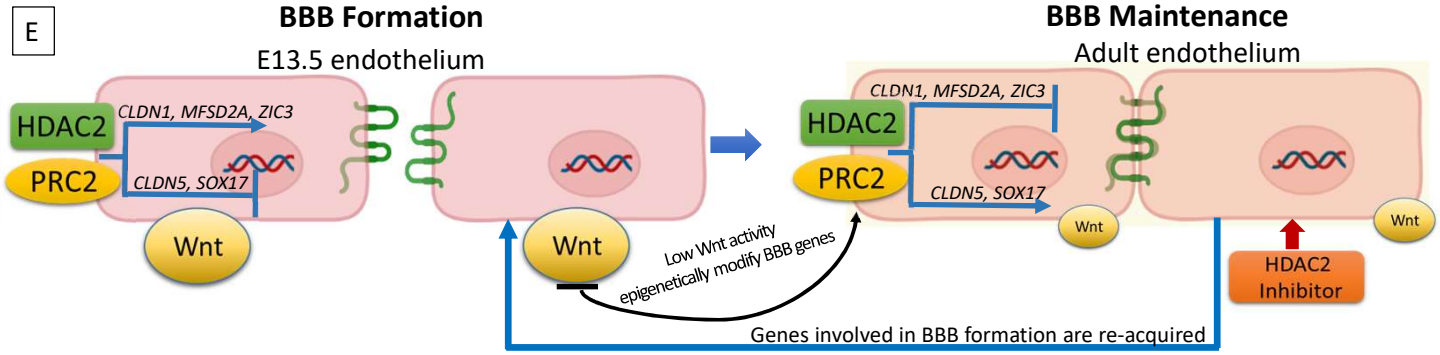
C

Epigenetic modifications induced by MS-275 in BBB genes is reversible



D

MS-275 treatment in human cortical vessels



E