The lncRNA landscape of cardiac resident macrophages and identification of *SchlafenInc* as a regulator of macrophage migratory function

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ABSTRACT

Cardiac resident macrophages (crMPs) were recently shown to exert pivotal functions in cardiac homeostasis and disease, but the underlying molecular mechanisms are largely unclear. Long non-coding RNAs (lncRNAs) are increasingly recognized as important regulatory molecules in a number of cell types, but neither the identity nor the molecular mechanisms of lncRNAs in crMPs are known. Here, we have employed deep RNA-seq and single cell RNA sequencing to resolve the crMP lncRNA landscape from healthy and diseased murine myocardium. CrMPs express previously unknown and highly cell type-specific lncRNAs, among which one lncRNA, termed *Schlafenlnc*, was particularly abundant and enriched in crMPs. We found *Schlafenlnc* to be necessary for migration-associated gene expression in macrophages *in vitro* and *in vivo* and essential for their adhesion and migration. Collectively, our data provide a basis to the systematic characterization of lncRNAs in crMPs and establish *Schlafenlnc* as a critical regulator of macrophage migratory functions.

INTRODUCTION

Long non-coding RNAs (lncRNAs) are the largest group of non-coding RNA molecules, with 14,764 lncRNA genes currently annotated in mouse and 19,933 in humans (Gencode in October 2022). Although only a small fraction of these has been functionally characterized, there is strong evidence that lncRNAs, through a variety of mechanisms, adopt critical roles in disease, for example in cancer[1], metabolic, neurologic[2] and cardiovascular disease[3]. Macrophage lncRNAs in cardiovascular disease have been primarily characterized in the vasculature and within the context of atherosclerosis, e.g., formation of foam cells or polarization (reviewed in [4]). With regard to cardiac disease, the analysis of lncRNAs and their functions has largely focused on cardiomyocytes[5], endothelial cells[6] and fibroblasts[7] (reviewed in [3]). By contrast, very little is known about lncRNAs in cardiac resident macrophages (crMPs), although this cell type constitutes the largest population of immune cells in the myocardium during homeostasis and also determines cardiac disease processes (reviewed in [8]). CrMPs have in recent years been characterized with respect to their developmental origin[9,10], their composition[10,11] and their putative functions such as roles in cardiac conduction[12,13], hypertrophy[14] or lymph vessel growth[15]. Although it can be readily assumed that many of these functions are subject to lncRNA-mediated regulation, a comprehensive description of the lncRNome from crMPs or functional data on a crMP-specific lncRNA is missing thus far.

In this study, we established the first comprehensive analysis of the lncRNA portfolios of the major cardiac cell types in health and disease, with particular emphasis on macrophages. In mouse myocardium, we identified lncRNAs that are specific for, and define, this cell type. One crMP lncRNA that has previously not been analyzed and which we termed *Schlafenlnc*, stands out for its strong enrichment and high abundance in crMPs. Upon functional characterization of a *Schlafenlnc* knockout in a mouse model for left ventricular pressure overload, we found that this lncRNA regulates macrophage abundance in the heart and the ability of this cell type to exert chemotactic migration. We therefore propose that *Schlafenlnc* critically regulates the function of cardiac macrophages in disease.

RESULTS

lncRNAs are important determinants for the definition of specific cell types

To determine the long non-coding RNA (lncRNA) repertoire in the major cardiac cell types and its changes under disease conditions, we subjected mice to chronic left ventricular pressure overload (induced by transverse aortic constriction, TAC), an established model of cardiac inflammation that leads to hypertrophic and fibrotic remodeling of the heart. At day 6 after surgery, we sacrificed the mice, isolated primary cells by Langendorff digestion[16], and performed droplet-based single cell RNA sequencing (scRNA-seq, Chromium platform) (Fig. 1A). This experimental approach analyzes all cardiac non-myocytes, while the number of cardiac myocytes is decreased due to size constraints of the method. Unsupervised clustering of both samples based on the expression of both coding and non-coding RNAs with the Seurat analysis tool (Seurat V3.2[17], Gencode vM22[18]) yielded a total of 20,010 cells after quality control. These comprised all major non-myocyte cardiac cell types (Seurat clustering resolution 0.3, Fig. 1B), including subpopulations of immune cells, fibroblasts, endothelial cells and mural cells (Fig. 1B). Differential gene expression analysis from these cell types revealed distinct sets of protein-coding genes that define each major cardiac cell type as individual clusters (Extended Data Fig. S1), in line with previous reports[19,20].

Despite the generally lower expression levels of lncRNAs in comparison to protein-coding genes[21–24], we were able to readily detect over 440 lncRNAs. Several of the most strongly expressed lncRNAs exhibited a very high level of cell type specificity (Fig. 1C and 1D), suggesting that cardiac cell types can be defined based on their lncRNA transcriptomes. As Fig. 1D shows, our data not only confirmed the previously reported expression of *Meg3* in fibroblasts[25], *H19* in endothelial cells[26] and *Mhrt* in cardiac myocytes[27], but also revealed lncRNAs that had hitherto not been reported to be cell type-specific in the heart. These lncRNAs may reasonably be presumed to potentially exert important roles in their specific cell type, prompting for functional characterization.

To validate the contribution of lncRNAs to cell type identity, we performed differential clustering of the scRNA-seq data sets with or without lncRNA genes, and tested for clustering precision. Independent of whether the top 3,000 variable genes defined by the Seurat pipeline, all annotated genes (n=55,487) or expressed genes (n=19,719, expression value >0) were used to perform clustering, the inclusion of lncRNA marker genes significantly improved clustering compared to protein coding genes alone (Fig. 1E-G). Compared to clustering based on protein-coding genes alone, this resulted in an overall increased sensitivity (indicated by an increased adjusted Rand Index value) and yielded additional subclusters of cell types. Specifically, the detection of clusters representing venous capillary endothelial cells, pericytes, *Fabp4*$^{+}$ macrophages and *Notch2*$^{+}$ fibroblasts was dependent on the inclusion of lncRNAs (resolution 0.65) (Fig. 1H, Extended Data Fig. S1B).

In conclusion, lncRNAs were shown to be a valuable determinant of defining clusters and fine-tuning cell type assignations.

As a means to approach lncRNAs of potential disease relevance, we next thought to assess the expression of lncRNAs during cardiac disease by comparing the datasets obtained from sham-treated vs. TAC-treated mice. Upon increasing the clustering resolution from 0.3 to 0.7, a more detailed and granular cell assignment of lncRNA genes to cellular subtypes was achieved (Extended Data Fig. S2A

110 and S2B). With this setup, we performed differential gene expression analysis for the sub-populations
111 of the major cell types in the heart, and visualized the top de-regulated lncRNAs (Fig. 2A-C, Extended
112 Data Fig. S2C). Some lncRNAs were regulated after TAC throughout all subpopulations of a cell type,
113 e.g., Gm29686 in fibroblasts (Fig. 2A), whereas others were only significantly regulated within a defined
114 subpopulation, e.g., Gm49785 in the fibroblast subpopulation (FB6) (Fig. 2A). This subpopulation of
115 fibroblasts represents activated myofibroblasts (high expression of periostin), a cell type involved in
116 extracellular matrix secretion and immune cell recruitment[28]. Similarly, several lncRNAs displayed
117 dysregulation within cardiac endothelial cells (Fig. 2B) and cardiac macrophages (Fig. 2C), resolved to
118 the level of cellular subtypes.
119 Of particular interest are lncRNAs in macrophages of the myocardium, since no lncRNA had thus far
120 been identified or functionally characterized in this cardiac cell type. As a first approach to the potential
121 function of the lncRNAs we had identified as deregulated in crMPs in the TAC model, we correlated
122 these lncRNAs with biological processes of their overlapping neighboring genes (Extended Data Fig.
123 S2D). Since lncRNAs are often co-expressed and co-regulated with proximal genes in *cis*[29], we
124 speculated that the latter may give insight into the biological context within which these lncRNAs
125 function. For example, genes antisense to downregulated lncRNAs in the Mac1 cluster (*Lyve1*
126 macrophages) after TAC are associated with programmed cell death processes and genes antisense
127 to downregulated lncRNAs in the Mac2 cluster (*Cd74*+ macrophages) after TAC were predominantly
128 linked to adhesion pathways (Extended Data Fig. S2D), suggesting that the reported de-regulated
129 lncRNAs might also be involved in the regulation of these processes.
130 Taken together, we report a comprehensive inventory of regulated lncRNA genes in the major non-
131 myocyte cell types of the heart during pressure overload. These lncRNA genes will be interesting
132 candidates to be studied in more detail.

133 **Determination and annotation of the lncRNA landscape of cardiac resident macrophages**

134 As the sequencing depth of current single cell sequencing methodology limits detection sensitivity, we
135 therefore complemented our scRNA-seq data by performing deep RNA-Seq on basal total murine
136 myocardium as well as crMPs that were isolated using well-established cell markers (CD45, CD64,
137 CD11b and F4/80[9,10]). These surface proteins collectively labelled essentially all crMPs in our scRNA-
138 seq data (Extended Data Fig. S3A), corroborating our purification strategy. Using a pipeline to identify
139 and quantify novel lncRNAs (Extended Data Fig. S3B), we identified 250 genes that had previously not
140 been annotated by Gencode vM22[18]. The identified novel lncRNAs showed the same characteristics as
141 the known lncRNAs with respect to having no coding probability (true non-coding) and transcript length
142 (Extended Data Fig. S3C upper panel), but were more highly expressed in our data set than the known
143 lncRNA population (Extended Data Fig. S3C lower panel). RNA-seq from fractionation experiments
144 showed for known lncRNAs a distribution of 80% in the nucleus (and 20% to the cytoplasm), the novel
145 lncRNAs are slightly less nuclear in localization (70% nuclear, 30% cytoplasmic; Extended Data Fig.
146 S3D).
147 As a further approach to the function of the newly identified lncRNAs, we built on the observation that
148 the products of lncRNA genes and protein-coding genes which are in proximity of 5kb or less and in
149 antisense overlapping orientation are prone to interact in the same biological processes[30]. The

150 distribution of the interaction categories (e.g. “antisense overlapping”) was comparable between novel
151 and known lncRNAs (Extended Data Fig. S3E). Notably, 15 novel lncRNA genes were found without
152 any interaction, categorizing them as novel long intergenic RNAs (lincRNA). For both known and novel
153 lncRNAs we found many well-expressed genes (> 5 transcripts per million reads, TPM) that are specific
154 in cardiac macrophages compared to total murine myocardium (containing myocytes, endothelial cells,
155 fibroblasts, and macrophages) (Fig. 3A).

156 **Identification of lncRNA *SchlafenInc* as an abundant and enriched macrophage lncRNA**

157 In order to select lncRNA candidates for experimental validation, we further filtered the datasets from
158 scRNA-Seq data and RNA-Seq based on the following stringent criteria: low coding probability
159 according to CPAT score (true non-coding RNA < 44% coding probability), absolute expression > 25
160 TPM in macrophages, enrichment of log2FC >3 and FDR <0.05, and de-regulation in TAC (scRNA-seq
161 data) FC >0.25). This yielded 13 lncRNA candidates that are highly abundant, enriched and regulated
162 in cardiac macrophages (Fig. 3B).

163 We found one of these candidates, lncRNA *AI662270*, particularly interesting due to its genomic
164 association with a cluster of eight genes on chromosome 11 which encode members of the Schlafen
165 family of proteins (Fig. 3C). Schlafen proteins are found in human and mouse and known to regulate
166 important processes and functions such as immune cell development, invasion and migration[31].
167 *AI662270*, however, is a hitherto uncharacterized gene within this locus. The genomic location of
168 lncRNA *AI662270* in the Schlafen cluster led us to name the transcript *SchlafenInc.*

169 The Schlafen locus of protein-coding genes is very similar between human and mouse in general,
170 yet the internal order of the proteins (and lncRNAs) is slightly different. A study by Ulitsky and colleagues
171 developed a pipeline to identify syntenic genes across multiple organisms, where lncRNAs are tested
172 on the relative location and orientation to orthologous protein-coding genes[32]. By comparing Gencode
173 annotations[18] of human (V30) and mouse (vM21) with their pipeline, the gene *ENSG00000267074* was
174 designated as the putative human *SchlafenInc* homolog (Fig. 3C). It is expressed in various human cell
175 types of leukocyte origin (using the RNA-Get module Encode data sets[33], Extended Data Fig. S4A),
176 making it an attractive molecule for studying the observed phenotypes in human cells. Besides the
177 analysis of expression, we tested if the two lncRNAs share common potential binding partners using a
178 pretrained deep neural network (DNN)[34] on eCLIP-seq data[35]. Indeed, the mouse *SchlafenInc* and
179 human *ENSG00000267074* contain three major binding modules of the same or similar proteins
180 (Extended Data Fig. S4B and S4C), supporting the hypothesis of conservation.

181 Murine *SchlafenInc* was up-regulated after TAC in both macrophage populations, but even more
182 pronounced in Mac2 (*MHCII*-high, *Cd74*+) macrophages (Fig. 3D). Its strong expression made it readily
183 detectable by Northern blotting (Fig. 3E). Interrogation of the RNA-seq datasets from the nucleus and
184 the cytosol, as well as RNA *in situ* hybridization, showed that *SchlafenInc* localizes to both compartments
185 (Fig. 3F and G, Extended Data Fig. S4D).

186 ***SchlafenInc* determines myocardial macrophage infiltration and polarization in response to**
187 **pressure overload**

188 To assess the phenotype of a macrophage-specific deficiency of *SchlafenInc in vivo*, we generated a
189 *SchlafenInc*-deficient mouse line (*SchlafenInc*$^{-/-}$) by CRISPR/Cas9 and examined these mice under
190 basal and disease conditions. The dominant transcript was targeted with specific sgRNAs, including the
191 promoter region upstream of *SchlafenInc* (determined by H3K4me3 signals in murine bone marrow-
192 derived macrophages, ENCODE data set ENCSR000CFF[36]) to abrogate all transcription from that locus
193 (Extended Data Fig. 5 A). Deletion was verified by PCR from genomic DNA (Extended Data Fig. 5 B).
194 The early compensation stage of the TAC model is characterized by sterile inflammation and both an
195 influx of monocytes as well as proliferation of resident macrophages. We therefore performed sham and
196 TAC surgery (Fig. 4 A) and analyzed the heart phenotype and monocyte/macrophage numbers after 6
197 days (Fig. 4 B). The heart weight of *SchlafenInc*-deficient mice was significantly reduced (Fig. 4 C) and
198 heart function (comparing ejection fraction of sham and TAC mice) was significantly improved (Fig. 4
199 D). Furthermore, *SchlafenInc*$^{-/-}$ hearts showed a substantial decrease in pressure overload-induced
200 interstitial fibrosis (Fig. 4 E) and interestingly a significantly reduced monocyte/macrophage infiltration
201 after TAC, as determined by immunostaining of left ventricular myocardial sections against the
202 monocyte/macrophage marker CD68 (Fig. 4 F).

203 To assess the composition of immune cells in more detail we next performed scRNA-seq of the CD45+
204 cell fraction isolated from the myocardium and blood (Fig. 5 A). As expected, macrophages are the
205 dominant cell population within the heart whereas the blood contains more monocytes (Fig. 5 B).
206 Activated macrophages and monocytes release signals in an auto- and paracrine fashion[19].
207 *SchlafenInc*$^{-/-}$ mice showed decreased signaling between almost all macrophage and monocyte
208 populations after 6 days of TAC (Fig. 5 C), with a significant decrease in inflammatory signaling
209 pathways such as Osteopontin (*Spp1*) and Macrophage Inhibitory Factor (*Mif*) signaling (Fig. 5 D), both
210 are important factors in recruiting monocytes and other inflammatory cells from the blood after cardiac
211 injury[37,38]. Performing RNA velocity analysis of cardiac leukocytes after TAC (Fig. 5 E) showed gene
212 dynamics of *SchlafenInc* WT *Ly6C*-high monocytes towards non-classical monocytes as well as a
213 moderate dynamic from *MHCII*-high macrophages to *Lyve1* macrophages. In
214 *SchlafenInc*$^{-/-}$, the dynamics between different monocyte states was largely abolished while transition
215 dynamics from monocyte-derived macrophages (*MHCII*-high) to resident-like macrophages (*Lyve1*) was
216 strongly increased (Fig. 5 E). Taken together, the analysis of *SchlafenInc*$^{-/-}$ myocardium indicated
217 stronger polarization of cardiac macrophages towards the *Lyve1* macrophage population, which is
218 generally believed to play a more anti-inflammatory role in disease.

219 Next, we asked which genes were changing globally in *Lyve1* macrophages after TAC and upon
220 *SchlafenInc* deletion. Among the genes that were de-regulated during TAC (WT TAC vs. sham), 64
221 genes were additionally strongly de-regulated in *SchlafenInc*$^{-/-}$ vs. *SchlafenInc* WT *Lyve1* macrophages
222 (|Log2FC|>0.5, FDR<0.05). 17 of these genes are classified as coding for secreted proteins (Fig. 5 F,
223 categorization according to Human Protein Atlas[39]), for example, *Lgals3* (Galectin-3) and *Spp1*
224 (Osteopontin), which were de-regulated in all macrophage populations (Extended Data Fig. 5 C). An

225 analysis of the associated biological processes of de-regulated genes in *Lyve1* macrophages (Fig.5 F)
226 showed leukocyte chemotaxis and migration as the strongest impacted pathways (Fig. 5 G).

227 Taken together, *SchlafenInc* deletion in immune cells of the heart altered the cardiac macrophage
228 response to early pressure overload towards an anti-inflammatory phenotype and disease-associated
229 phenotypes such as fibrosis and heart weight were improved.

230 ***SchlafenInc* is required for macrophage adhesion and migration**

231 To elucidate the function of *SchlafenInc* in more detail and explore its potential role in migration
232 processes, we generated *SchlafenInc*-deficient macrophages from Hoxb8-FL macrophage
233 progenitors[40,41] (Extended Data Fig. S6A). Gene deletion was validated by RNA-Seq (Extended Data
234 Fig. S6A, middle track). As a genomic deletion might have an impact on nearby genes due to a
235 mechanism in *cis* by the deleted lncRNA or due to the deletion of a regulatory element within the DNA,
236 the surrounding region of *SchlafenInc* (0.5 Mbp in each direction) was analyzed for de-regulation
237 (Extended Data Fig. S6B). 9 of 64 genes were found to be moderately dysregulated (with |log2FC| > 1,
238 FDR < 0.05) in the vicinity of *SchlafenInc* revealing that there was not a general activation or de-
239 activation of that locus by *SchlafenInc*. To identify major perturbations on gene expression level,
240 *SchlafenInc*-deficient monoclonal cell lines (2 clones, each measured in n=4) were subjected to RNA-
241 Seq. These samples were compared to control monoclonal cell lines (two clones with n=3 each) that
242 were generated using an unrelated sgRNA (sequenced as wildtype at the *SchlafenInc* locus). We
243 observed 2,660 genes significantly de-regulated (|log2FC| > 1, FDR < 0.05) (Fig. 6 A), among those
244 *SchlafenInc* that dropped from 67 TPM to 0.33 TPM. Among the downregulated genes, we observed an
245 enrichment of genes involved in migration and adhesion similarly to our *in vivo* study (Fig. 6 B, e.g.,
246 *Mmp14* and *Lgals3*). Lower amounts of *Lgals3* could not only be observed in our RNA-Seq data but
247 were validated on protein level (Galectin-3 is the gene product of *Lgals3*) by measurement of cell culture
248 supernatant using an enzyme-linked immunosorbent assay (ELISA) (Fig. 6 C). More generally, gene
249 ontology analysis of all significantly de-regulated genes implicated pathways such as regulation of
250 cytokine production, regulation of cell adhesion and chemotaxis that are promoted by *SchlafenInc* (Fig.
251 6 D). To validate these results, we performed an adhesion assay by rigorous washing (Fig. 6 E) as well
252 as tested the chemotactic migration ability of *SchlafenInc*-deficient macrophages with a gradient of the
253 chemokine C5a (Fig. 6 F). Adhesion was diminished by approximately 23% (Fig. 6 E) and chemotaxis
254 was about 50% reduced (Fig. 6 F) in *SchlafenInc* KO cells compared to wild type controls.
255 Together these data obtained on *SchlafenInc*-deficient macrophages corroborate a cell autonomous
256 role for this lncRNA in macrophages, in line with its specific expression in this cell type. *SchlafenInc* is
257 required for the activation of macrophage gene programs underlying cell adherence and migration
258 providing a mechanistic basis for the phenotype observed *in vivo* in *SchlafenInc*-deficient mice.

DISCUSSION

This study provides the first comprehensive lncRNome of cardiac resident macrophages. One of the most highly enriched and abundant lncRNAs in cardiac resident macrophages – lncRNA *AI662270*, which we termed *Schlafenlnc* – was shown to be involved in adhesion and migration. As for *Schlafenlnc*, a particular characteristic of lncRNAs in general is their high degree of cell type-specificity[21–23] which makes them important for processes such as development, differentiation of cell types, cellular fate and function (reviewed in [3]). This characteristic could therefore make lncRNAs the natural choice when describing cell population expression signatures, especially in single cell RNA sequencing (scRNA-seq). scRNA-seq allows for an unbiased approach to identify main- and sub-populations of cells, yet little focus has been set to expressly include lncRNA marker genes. Initially, methodical problems such as library preparation, sequencing depth or analysis software hampered the detection of lncRNAs in such data sets. LncRNAs are still not as readily measured as mRNAs due to their on average lower expression levels, but together with improved scRNA-seq depth and improved annotations, our data suggest that lncRNAs are underestimated markers of cell type definition. Emerging platforms measuring RNA transcripts in tissue such as spatial transcriptomics or *in situ* sequencing (reviewed in [42]) might be able to leverage that potential further.

Focusing on lncRNAs and their potential regulation in ventricular myocardium during disease, we could identify an extensive set of regulated lncRNAs for all major cell populations. In general, we find more up-regulated than down-regulated genes upon induction of disease, which we believe to be due to the detection limit of scRNA-seq concerning very lowly expressed genes[43]. LncRNAs more specific to and regulated in a subset of cell types might offer new possibilities for therapeutic intervention and to gain insight into regulatory mechanisms specific for defined cell types. As we have shown for the three macrophage clusters (Mac1 (*Lyve1*+), Mac2 (*Cd74*+) and transitional macrophages), the different sets of regulated lncRNAs are implicated in distinct gene ontology pathways, which points to distinct functions of these macrophage populations.

Cardiac resident macrophages have been shown to be key players in diseases such as heart failure and myocardial infarction (reviewed in [44–47]). In the mouse model of left ventricular pressure overload, crMPs are proliferating during the compensation phase (from 0 days to about 7-14 days). An infiltration of monocyte-derived MPs starts early in disease and increases in the late, decompensated phase[48]. The infiltration of monocyte-derived MPs is typically associated with a long-term detrimental cardiac function[48–55]. This recruitment of monocytes is at least to some degree dependent on crMPs[56], and suppression of this communication axis might be beneficial. Galectin-3, for example, is known to be increased in heart failure patients[57] and is associated with murine cardiac remodeling during pressure-overload[58]. It is well known to be involved in the monocyte as well as macrophage migration and infiltration in cardiovascular diseases[59–61]. Our combination of unbiased scRNA-seq measurements and in-depth coverage by RNA-Seq allowed us to identify the hitherto uncharacterized lncRNA *Schlafenlnc* as a regulator of macrophage adhesion and migration. *Schlafenlnc* is up-regulated after 6 days of TAC as well as other genes involved in adhesion and migration such as Galectin-3 and its deletion impacts Galectin-3 expression both *in vivo* and *in vitro*. Our quantitative assessment of macrophage/monocyte numbers (CD68 staining) and the analysis in relative immune cell composition by scRNA-seq after 6 days of TAC point to a mechanism of defective signaling by *Schlafenlnc*-deficient crMPs to initiate

300 recruitment of inflammatory cells such as monocytes. Given its selective expression in crMPs, we
301 believe that targeting of *Schlafenlnc* warrants further exploration as a potential anti-inflammatory
302 therapeutic strategy.

303 Similar to its orthologue in mice, human *Schlafenlnc* is specific for immune cell types (Extended Data
304 Fig. S4A). In support of conserved function in immune cells, our protein interactor analysis of both
305 murine and human *Schlafenlnc* using deep learning showed similar binding modules for RNA binding
306 proteins (Extended Data Fig. S4B and S4C). Further studies will be required to dissect molecular
307 interactors for *Schlafenlnc* in mouse and human.

308 Taken together, our study demonstrates that lncRNAs act as critical contributors to the
309 transcriptome-based analysis of cell clusters in line with their crucial role in development and cell fate
310 determination[3], leading to the definition of cell identities. The combination of single cell sequencing and
311 RNA-Seq approaches led us to the identification of multiple novel candidate lncRNAs that warrant
312 further analysis in cardiac homeostasis and disease. One of the most abundant and macrophage-
313 enriched lncRNAs, *Schlafenlnc*, acts as a crucial regulator of macrophage adhesion and migration.

314 **METHODS**

315 Experimental animals, surgery conditions

316 Surgeries on mice were performed according to the guidelines and regulations of the responsible
317 authorities. Consent was given by the institutional review board of the Regierung of Oberbayern (ROB-
318 55.2-2532.Vet_02-19-194).
319 Wild type male mice of C57BL/6N background were purchased from Jackson Laboratories and
320 submitted to transverse aortic constriction (TAC) or sham procedure as reported previously[16] at an age
321 of 8-10 weeks.

322 To produce *Schlafenlnc* -knockout mice, two sgRNAs (5' cut site: 5'- GAGGTTCCATCAGTCTGACA-
323 3', 3' cut site: 5'-ACATGTCTTTGGTAGGAGCA-3') targeting the flanking region of the *Schlafenlnc* gene
324 in the mouse genome were designed and synthesized by IDT (Coralville, IA, USA). A mixture of Cas9
325 protein (IDT) and sgRNAs was electroporated into C57BL/6 mouse zygotes. Genomic DNA of newborn
326 mice was amplified using the primers Slfnlnc1 (5'-TTCCTTCAGAGGTCCATCAG-3') and Slfnlnc4 (5'-
327 AGCCAGTCTAAAGACTCTGG-3'). The appearance of a 360 bp band identified founder mice with a
328 complete loss of the *Schlafenlnc* gene. The animals were bred to homozygosity and used for the
329 experiments.

330 Echocardiography was performed as reported before[16].

331 Primary cell isolation from adult murine heart

332 Hearts of C57BL/6N mice were harvested at an age of 8-10 weeks and subjected to digestion by
333 perfusion via the coronary arteries using Collagenase II (Worthington, Lakewood, NJ) as described
334 previously[62]. Cell suspension was processed according to subsequent protocol/experiment.

335 Single cell cloning and sequencing

336 Primary cells from a sham[16] and an adult mouse heart 6d after TAC were washed three times with PBS
337 to remove cell debris and passed successively through 100µm, 70µm and 30µm filters. The single cell
338 library was generated using the Chromium Single Cell 3' GEM, Library & Gel Bead Kit v2 (10X
339 Genomics) with a target of 10,000 cells per sample. The single cell library was cloned according to the
340 manufacturer's instructions. Sequencing was performed on a HiSeq4000 (Illumina).

341 Primary cells from the hearts from *Schlafenlnc*$^{-/-}$and littermate wildtype mice in basal condition as well
342 as after 6d TAC were passed through a 100µm filter and centrifuged for 1min at 100*xg* to pellet
343 cardiomyocytes. The supernatant (non-myocytes) was passed through a 40µm filter and washed twice
344 in magnetic-activated cell sorting (MACS) buffer (PBS + 0.5% BSA + 2mM EDTA) to result in 200µl cell
345 suspension. For the isolation of PBMCs, blood was diluted with 5 ml of MACS buffer and layered onto
346 5ml of Ficoll-Paque (17144002, Cytiva), followed by centrifugation at 400xg and 4°C for 30min in a
347 swinging rotor without a brake. The white layer containing the cells was aspirated, transferred to a 50ml
348 tube and mixed with 30ml of MACS buffer. After centrifugation for 5 min at 300xg and 4°C, the pellet
349 was washed once more with 30ml MACS buffer (200xg and 4°C for 5min) and resuspended in 200µl

350 MACS buffer. Cardiac and blood leukocytes (basal condition) were further purified using MACS, labelled
351 with cell hashing antibodies and isolated by FACS (all steps at 4°C): Cell suspensions were incubated
352 with Fc-block (1:50, BD Pharmingen 553142) for 10min and subsequently incubated with mouse CD45
353 MicroBeads (Miltenyi Biotec) for 15min. Cells were washed with MACS buffer and MACS was performed
354 using an AutoMACS (Miltenyi Biotec) running the program *possel_s*. After elution, pelleted cells were
355 resuspended in MACS buffer containing anti-CD45-FITC (1:1000, clone 30-F11, Biolegend, 103108),
356 PI (1:100) and cell hashing antibodies (1:1000, #1 for heart, #2 for blood, Biolegend) and incubated for
357 30min protected from light. For TAC samples, PBMCs were purified as above, cardiac non-myocytes
358 were directly used for cell sorting. Both cell fractions were stained with cell hashing antibodies (1:1000,
359 #2 for heart, #3 for blood, Biolegend) and Propidium iodide (1:100, Thermo) and incubated for 30min.
360 After washing, live cells of all samples were sorted on a Sony cell sorter SH800S, heart and blood cells
361 of each genotype and condition were pooled and run on a Chromium controller using the Chromium
362 Next GEM Single Cell 3' v3.1 Library & Gel Bead Kit (10X Genomics) with a target of 10,000 cells per
363 pool. Libraries were cloned according to the 10x Genomics user guide with feature barcode technology
364 for cell surface protein (RevD) and Biolegend's protocol for amplification of the cell hashing libraries.
365 Libraries were pooled according to the recommendations of 10x genomics protocol and sequenced on
366 a NovaSeq6000 (Illumina).

367 Purification of primary cells and cardiac macrophages by F4/80 MACS and cell sorting for RNA-seq

368 For a total myocardial cell sample, 100µl of the raw cell suspension of primary cells from adult wildtype
369 mouse hearts were washed twice with PBS and lysed in Trizol (ThermoFisher) for subsequent RNA
370 preparation. For the purification of cardiac macrophages, the cell suspension was filtered through cell
371 sieves of 70µm and 40µm of size and centrifuged at 100 *x g* for 3min at 4°C to completely remove
372 cardiac myocytes. All following steps were performed on ice/at 4°C. The supernatant (non-myocytes)
373 was washed twice in magnetic-activated cell sorting (MACS) buffer (PBS + 0.5% biotin-free BSA + 2mM
374 EDTA). MACS was performed in a two-step process using anti-F4/80-biotin coupled antibody (clone
375 BM8, Biolegend, 123106) and Streptavidin microbeads (Miltenyi Biotec) on LS columns (Miltenyi
376 Biotec). The eluted macrophage fraction was resuspended in a final volume of 200µl MACS buffer and
377 incubated 10 min with Fc block (1:50, BD Pharmingen). 200µl of staining solution containing flow
378 cytometry antibodies (anti-CD45-PECy7 (1:100, clone 30-F11, Invitrogen, 25-0451-82), anti-CD64-
379 PE/Dazzle594 (1:50, clone X54-5/7.1, Biolegend, 139320), anti-CD11b-PE (1:100, clone M1/70, BD
380 Pharmingen, 557397)) was added and incubated for 30min in the dark. Cells were washed twice with
381 MACS buffer and sorted on a Biorad S3 cell sorter. CD45+/CD64+/CD11+ macrophages were collected
382 (between 100k-170K cells), washed with PBS and lysed in Trizol (Invitrogen) for RNA preparation.

383 Fibrosis staining

384 Paraffin sections (8 µm) of ventricular myocardium were stained with Sirius red and Fast green. Images
385 of the whole heart were taken with a 10x objective using a AxioObserver.Z1 (Zeiss) motorized scanning-
386 stage microscopy (130 x 85; Märzhäuser). Fibrosis of the left ventricular myocardium was quantified as
387 the ratio of signals from Sirius red to Fast green in each section using the Metamorph software
388 (Molecular Devices).

389 CD68 staining

390 Cryosections (5 µm) of ventricular myocardium were fixed with methanol for 10 min at -20°C, washed
391 thoroughly with tap water and treated with acetone for 1 min at -20°C. Sections were air dried and
392 subsequently blocked with 5% goat serum in PBS for 1 h. Sections were stained with anti-mouse CD68
393 (1:100, clone FA-11, AbD Serotec, MCA 1957) at 4°C overnight. The sections were washed three times
394 with PBS + 0.05 % Tween 20 and stained with anti-rat AF564 (1:100, Invitrogen, A11007) and SYTOX™
395 Green (1:1000, Life technologies, S7020). Images were acquired by confocal microscopy using 63x
396 glycerol objective (Leica SP5) and analyzed using Metamorph (Molecular Devices).

397 Generation of knockout macrophages

398 Conditionally immortalized hematopoietic progenitor cells were generated from a R26-Cas9-eGFP
399 transgenic mouse line[41] as previously described[40]. For inducing CRISPR-Cas9 mediated knockout in
400 Hoxb8-FL cells, cells were electroporated with an sgRNA pair targeting the *SchlafenInc* locus (IDT: Alt-
401 R® CRISPR-Cas9 sgRNA, 5' cut site 5'-GAGGTTCCATCAGTCTGACA-3', 3' cut site 5'-
402 GGCTTGGAAGTAGAAAACCG-3'), as control, a single sgRNA was used (5'-
403 ACCCCTTCAGCTTCACCTCC-3'). Transfected cells were single-cell cloned by limiting dilution assay
404 into 96-well plates and screened for knockout clones. CRISPR-directed deletions on *SchlafenInc* were
405 identified by a PCR-based strategy to amplify the genomic DNA of the clones. Single-cell clones carrying
406 the induced deletions were expanded and frozen for further use. To do this, extraction of genomic DNA
407 was performed with QuickExtract™ DNA Extraction Solution (Lucigen) according to the manufacturer's
408 instruction. Briefly, approximately 100,000 cells were lysed in 50µl of QuickExtract by incubation as
409 follows: 65°C – 15min, 68°C – 15min, 98°C – 10min. 50ng of genomic DNA was then subjected to PCR
410 amplification with two primer pairs to distinguish successful *SchlafenInc* knockout clones from wildtype
411 or heterozygous knockout. *SchlafenInc*_KO_forward 5'-CAGCCCTGAGTCAGAACATAC-3' and
412 *SchlafenInc*_KO_reverse 5'-TCCTTAGGGTGCACATCTACT-3' were used for detection of a
413 successfully cut site with a resulting amplicon length of 480bp, *SchlafenInc*_WT_forward 5'-
414 TCTGTCTCTACTCCCTCAACTC-3' and *SchlafenInc*_WT_reverse 5'-AAGGCATGTTCTGCTCCTAC-
415 3' were used for detection of the wildtype allele/uncut gene locus with a resulting amplicon length of
416 320bp.

417 Hoxb8-FL *SchlafenInc* KO cell culture and differentiation

418 Hoxb8 progenitor cells were cultured in RPMI 1640 supplemented with 10% FCS, 1% Pen/Strep, 50
419 ng/mL FLT3L, 50nM b-mercaptoethanol and 1nM b-estradiol. FLT3L supernatant was obtained from a
420 FLT3L-producing B16 melanoma cell line.
421 For differentiation of HoxB8 cells into macrophages, Hoxb8 progenitor cells were cultured in
422 differentiation medium consisting of the culturing medium described above without b-estradiol and
423 supplemented with 20% MCSF supernatant, which was obtained from a MCSF-producing L929 cell line.
424 Differentiation was accomplished after 6 days.

425 Generation of bone marrow-derived macrophages

426 Femurs and tibias from 6-12 week old C57BL/6N mice were prepared and bone marrow was isolated
427 by flushing the bones with R10 medium (RPMI medium, 10% FCS, 1% antibiotics) with a syringe. To
428 generate bone marrow derived macrophages, 3-4 mio. isolated bone marrow cells were seeded in a
429 10cm petri dish containing R10 medium supplemented with 20% MCSF supernatant. Medium was
430 added on day 3 and exchanged on day 6 after seeding, macrophages were fully differentiated on day 8
431 and used for experiments.

432 Cytoplasmic/nuclear fractionation

433 Cytoplasmic/nuclear fractionation was performed as described before[63]. Briefly, bone marrow-derived
434 macrophages (BMDMs) were washed two times with cold PBS and harvested. 10% of the cells was
435 transferred into a new tube and used as a sample for the total gene expression. Cells were centrifuged
436 at 200 *x g* for 10 minutes and the total expression sample was directly used for RNA isolation. The
437 fractionation sample was resuspended in 200µl lysis buffer A (10 mM Tris pH=8, 140 mM NaCl, 1.5 mM
438 MgCl2, 0.5% Nonidet P-40) and incubated for 5 minutes on ice. The lysate was centrifuged at 1000 *x g*
439 for 3 minutes at 4°C. The supernatant containing the cytoplasmic fraction was directly used for RNA
440 isolation through lysis with Trizol. The sediment containing the nuclear fraction was washed two times
441 with lysis buffer A and one time with lysis buffer B (Buffer A supplemented with 1% Tween-40, 0.5%
442 deoxycholic acid) and afterwards resuspended in Trizol for RNA isolation.

443 RNA cloning and sequencing

444 Libraries from primary cardiac macrophages and total myocardium as well as *Schlafenlnc*-deficient and
445 control cell lines were generated using the TruSeq Stranded RNA Kit (Illumina) according to the
446 manufacturer's protocol, except for one step. Instead of using the SuperScriptII (ThermoFisher) as the
447 reverse transcriptase, SuperScriptIII (ThermoFisher) was used at a reaction temperature of 50°C.
448 Sequencing was performed on a HiSeq4000 platform (Illumina).
449 Libraries from fractionation samples were generated using the Stranded mRNA Prep, Ligation Kit
450 (Illumina) according to the manufacturer's instructions. Sequencing was performed on a NovaSeq6000
451 machine (Illumina).

452 RNA *in situ* hybridization (RNAscope)

453 Bone marrow derived macrophages were seeded on a removable 8-well chambers slide (Ibidi) and RNA
454 *in situ* hybridization was performed using the RNAscope Fluorescent Multiplex Reagent Kit (Advanced
455 Cell Diagnostics (ACD)) according to manufacturer's instructions. A custom designed RNA probe by
456 ACD was used for detection of *Schlafenlnc* (ENSMUST00000143673.2). Following the RNA *in situ*
457 hybridization, the slides were incubated with Phalloidin-AF647 (Abcam) for 60 minutes at room
458 temperature. Subsequently, slides were washed three times with PBS and mounted using ProLong™
459 Gold Antifade Mountant (Invitrogen). Images were acquired using the confocal microscope Leica TCS
460 SP II and the Leica Application Suite.

461 Galectin-3 ELISA

462 Cell culture supernatant of differentiated Hoxb8 macrophages was collected and the amount of soluble
463 Galectin-3 was quantitatively assessed using the Galectin-3 Mouse SimpleStep enzyme-linked immuno-
464 sorbent assay (ELISA) kit (Abcam) according to the manufacturer's instructions. The cell culture
465 supernatant was diluted 1:10 in sample diluent before use.

466 Chemotaxis

467 An IncuCyte® ClearView 96-Well Chemotaxis Plate (Sartorius) was coated on both sides of the
468 membrane using 50 µg/mL Matrigel (Corning) at 37°C for 30 minutes. After coating, plates were
469 incubated at room temperature for 30 minutes and afterwards washed with D-PBS. Differentiated HoxB8
470 macrophages were harvested and 2,000 cells were seeded in each well in differentiation medium
471 containing 0.5% FCS. Cells were allowed to settle at the bottom of the upper well for 30 minutes.
472 Subsequently, 100nM recombinant murine C5a (Pepro Tech) was added to the lower well of the
473 ClearView migration plate. Cells were imaged every 2 h for 72 hours using an Incucyte ZOOM
474 instrument and a 10x objective with chemotaxis plate settings. Chemotaxis was analyzed using the
475 IncuCyte ZOOM software.

476 Adhesion Assay

477 Ibidi black 96 well µ-Plate was coated with 20µg/mL Fibronectin (Sigma Aldrich) for 1 h at 37°C. 10,000
478 differentiated Hoxb8 macrophages were seeded in each well in differentiation medium and were
479 incubated for 2 hours at 37°C in a 5% CO2 atmosphere. After the incubation, cells were stained using
480 NucBlue Live ReadyProbes reagent (Invitrogen) and following the manufacturers' instructions. Images
481 of the Nuclei were taken before and after removing the non-adherent cells by washing three times with
482 pre-warmed medium using an EVOS FL Auto 2 cell imaging system (Invitrogen). Cell nuclei were
483 counted automatically using the MetaMorph analysis software (Molecular Devices).

484 Northern blotting

485 Northern blotting was performed as reported previously[64]. Briefly, 10µg total RNA from fibroblasts
486 (NIH/3T3) and *in vitro* differentiated macrophages were run on a 1.2% formaldehyde agarose gel,
487 transferred to a Hybond-N+ membrane (Ge Healthcare) and UV-crosslinked. Probes were end-labelled
488 using [gamma-^{32}P]-ATP and purified using an Illustra ProbeQuant G-50 micro column (GE Healthcare).
489 A mixture of two different probes was used per target. Probes were incubated over night with a pre-
490 hybridized membrane, signal was accumulated using a phophoimager screen and detected using a
491 Cyclone Plus (PerkinElmer). mGAPDH probe1 5'-
492 CATGGACTGTGGTCATGAGCCCTTCCACAATGCCAAAGTTG-3', mGAPDH probe2 5'-
493 GAATTTGCCGTGAGTGGAGTCATACTGGAACATGTAGACCATGTAGTTGAGG-3', Slfnlnc probe1
494 5'-CAGCTCTGCGTCTCTGCTCCTGTTCAGGTCCATCTTTCATCTCCG-3', Slfnlnc probe2 5'-
495 CAGTCGGGGTTCTGCCTCTTGATGTGGTTGTGCTAACCGGTGGGAC-3'.

496 Quantitative real-time PCR

497 1 µg of total RNA was reverse transcribed with Protoscript II reverse transcriptase (NEB), both according
498 to the manufacturer's instructions using random hexamer primers. Quantitative real-time PCR (qPCR)
499 was carried out with primers listed below, using the FastStart universal SYBR Green master mix
500 (Roche). The sample volume of 12.5 µl contained 1x SYBR Green master mix, 5 pmol of each primer
501 and 10 ng of cDNA. PCR was performed using StepOnePlus Real-Time PCR systems (Applied
502 Biosystems). Slfnlnc_qPCR fwd 5'- ATGACTGGTCCTGGTTTTCG -3', Slfnlnc_qPCR fwd 5'-
503 TCTGCCTCTTGATGTGGTTG -3', mRPL32 fwd (housekeeping gene) 5'-
504 ACATCGGTTATGGGAGCAAC -3', mRPL32 rev (housekeeping gene) 5'-
505 GGGATTGGTGACTCTGATGG -3'.

506 Data analysis

507 *Single cell sequencing data analysis*
508 Raw reads generated by single cell RNA sequencing data from sham[16] and 6d after TAC (this study)
509 was aligned against the mouse genome (mm10, Gencode annotation vM22[18]) using CellRanger 5.0.0
510 (10x Genomics). Cell that were called by CellRanger were subsequently analyzed using the R package
511 Seurat v3.2[65]. After quality control (retaining cells with >500 or <3,600 detected genes, <10%
512 mitochondrial cell content), 7,913 cells of the sham condition and 13,803 of the TAC condition were
513 used for further downstream analysis. SCTransform[66] was used to normalize and scale the data as well
514 as remove unwanted variation (regressing out mitochondrial gene content and cell cycle score).
515 SCTransform was then applied to identify variable feature which were used to integrate both data sets
516 ("IntegrateData" function). By using the ElbowPlot function on Principal Component Analysis results, the
517 number of relevant components was determined (set to 40) and used for embedding and visualization
518 by Uniform Manifold Approximation and Projection (UMAP). Seurat's "FindAllMarkers" function was
519 used for determination of cell population marker genes.
520 For the comparison of clusterings with adjusted Rand index score, gene sets were defined as follows:
521 all genes = all genes listed in the vM22 annotation of Gencode; protein coding genes = all genes with
522 gene biotype "protein coding" in vM22; lncRNAs = all genes in vM22 lncRNA annotation of Gencode
523 (gene biotype "lncRNA"+"TEC"); "expressed" gene sets = same definitions as above with filter for
524 average expression >0 in at least one cell cluster; all variable genes = gene list generated by Seurat's
525 "SelectIntegrationFeatures", default is 3,000 genes; subset of variable genes based on gene lists
526 defined above. Next, clusterings with resolutions from 0.01 to 1.00 with 0.01 increments were iteratively
527 calculated ("FindClusters" function) and cluster ID per cell was saved in a data frame. The adjusted
528 Rand Index is a measure for the degree of similarity between two clusterings, for example. The R
529 package mclust[67] with inbuilt function "adjustedRandIndex" was used to compare all gene sets to the
530 respective complete gene set (e.g. the subset of expressed lncRNAs to all expressed genes).
531 For calculating de-regulated genes after TAC, first Adaptively thresholded Low-Rank Approximation
532 (ALRA)[68] was performed in order to impute drop out gene expression values followed by using Seurat's
533 "FindMarkers" function to compare each cell population before and after TAC. For the analysis of
534 scRNA-seq data of *Schlafenlnc*-deficient and wildtype animals, raw reads from basal and 6d after TAC
535 from WT and KO were aligned against the mouse genome (mm10, Gencode annotation vM25[18]) using

536 CellRanger 6.0.1 (10x Genomics) and analyzed using the R package Seurat v3.2[65]. Quality control was
537 applied to basal (>200 or <5,000 detected genes, <10% mitochondrial cell content) and TAC (>200 or
538 <4,000 detected genes, <10% mitochondrial cell content) samples. SCTransform and FindAllMarkers
539 was applied as above to normalize, scale, integrate data and identify marker genes for cell populations.
540 The integrated object was subset on all cell clusters expressing *Cd45* (Leukocytes). Deregulated genes
541 were calculated using Seurat's "FindMarkers" function to compare each cell population between
542 *Schlafenlnc*-deficient and wildtype cells. GO Term analysis of the deregulated genes was performed
543 using "DAVID 2021"[69].

544 *Cell-cell communication analysis*

545 A subset of cells of interest from blood and heart of *Schlafenlnc*-decient and wildtype *Cd45*+ (*Lyve1*
546 macrophages, *MHCII*-high macrophages, *Isg* macrophages, intermediate macrophages, classical
547 monocytes and non-classical monocytes) was used for analysis of secreted signaling pathways within
548 the CellChat database. The *Schlafenlnc*-deficient and wildtype objects were analyzed separately using
549 CellChat[70] and merged with "mergeCellChat". The differential number of interactions between the cell
550 clusters in *Schlafenlnc*-deficient vs. wildtype was analyzed and displayed using the CellChat function
551 "netVisual_diffInteraction" and the differential signaling pathways were assessed using "rankNet".

552 *Estimating RNA velocity*

553 To calculate the RNA velocity of single cells, the abundance of the spliced and unspliced RNA was
554 quantified by running Velocyto CLI (v0.17.5) on CellRanger output BAM files and the corresponding
555 gene annotation (GENCODE vM22). The resulting loom files were incorporated into the Seurat objects
556 using SeuratWrapper R package (v0.3.0), and after integration and clustering, the objects were
557 converted to AnnData format (h5ad) to be compatible for downstream analysis with the Python package
558 scVelo (v0.2.5). Using scVelo, cell-to-cell transition probabilities were estimated and RNA-velocity was
559 projected as streamlines on the UMAP embedding.

560 *Identification of novel lncRNAs*

561 Most steps of this pipeline were performed using the Galaxy platform[71]. Reads were mapped using RNA
562 STAR aligner (Galaxy version 2.6.0b-2)[72] using a built-in index based on Gencode vM22 annotation and
563 otherwise default parameters. StringTie (Galaxy Version 1.3.3.2)[73] was used to predict novel transcripts
564 (strand information RF, reference file Gencode vM22, average read length 90). StringTie merge (Galaxy
565 version 1.3.6) with default parameters was used to merge all new annotations together, including
566 Gencode annotation vM22 as reference annotation. Novel genes were extracted by Bedtools Intersect
567 tool (Galaxy version v2.29.0)[74], intersecting the merged annotation with the Gencode vM22 annotation.
568 Transcripts were extracted with the "Extract features" tool (Galaxy 1.0.0) and analyzed for coding
569 potential with FEELnc (Galaxy version 0.1.1.1)[30]. FEELnc produces a gene transfer format (gtf)
570 annotation file containing only transcripts classified as long non-coding RNAs. "cat" command was used
571 to merge this lncRNA annotation to Gencode vM22 reference annotation file to create reference
572 annotation file with additional novel lncRNA genes. All RNA-Seq libraries were measured again against
573 this new transcriptome using StringTie's "count" option. For visualizing coding probability, the python
574 version of the Coding-Potential Assessment Tool (CPAT, v2.0.0)[75] was run on the fasta sequences of

575 the novel lncRNAs ($ cpat.py -g novel_lncRNAs_gffread.fasta -d Mouse_logitModel_CPAT.RData -x
576 Mouse_Hexamer_CPAT.tsv -o output_novel_lncRNAs.txt). GraphPad Prism 8 was used to calculate
577 the frequency distribution in percentages. For plotting transcript lengths, the length determined in the
578 CPAT output was used and again transformed to frequency distribution in Prism. For the categorization
579 of interactions between lncRNAs and other genes, the classifier output of FEELnc was used.
580 Interactions with the same gene were discarded, for known lncRNAs only genes expressed
581 predominantly in cardiac macrophages were considered (log_2 fold change >1). To compute differential
582 gene expression analysis, DESeq2 package was used (Galaxy version 2.11.40.2)[76], outputting also a
583 normalized counts file. Normalized counts served as input for clustering and heatmap visualization by
584 ClustVis R package (v. 0.0.0.9)[77].

585 *RNA-Seq analysis of Hoxb8 SchlafenInc-deficient macrophages*

586 Fastq reads were mapped using RNA STAR aligner (Galaxy version 2.6.0b-2)[72] using a built-in index
587 based on Gencode vM22 annotation and otherwise default parameters. StringTie (Galaxy Version
588 1.3.3.2)[73] count option was used to quantify gene expression using Gencode vM22 as reference
589 annotation. DESeq2 was used to calculate differential gene expression (Galaxy version 2.11.40.2)[76].
590 The normalized counts file was used for heatmap visualization with ln(x) transformation of the top 20
591 down-regulated genes (ClustVis R package[77]).

592 *Neighboring genes analysis and Gene Ontology Analysis*

593 Up- or downregulated lncRNAs during pressure overload (p-value<0.05) in either macrophage,
594 fibroblast or endothelial cell populations were analyzed for neighboring genes using FEELnc (Galaxy
595 Version 0.2[30]) and filtered for antisense protein coding genes. Gene ontology analysis for these gene
596 sets for each cell population was performed using the web-based tool "DAVID 2021"[69]. Neighboring
597 genes of known and novel lncRNAs were also identified using FEELnc (Galaxy Version 0.2[30]).
598 Biological processes associated with the differentially regulated genes in *SchlafenInc*-deficient
599 macrophages (log2FC > 1 or < -1, FDR < 0.05) were determined using ClueGO application (v2.5.8)[78]
600 ran within Cytoscape (v3.8.2)[79] network analysis tool (p-value cutoff = 0.01, Min GO level = 4, Max GO
601 Level = 10, Number of Genes = 2, Min. Percentage = 4.0).

602 *Kipoi RBP-eclip analysis (Protein interactors analysis)*

603 Model training data: To access the binding probability of known RBPs on *SchlafenInc*, a pretrained deep
604 neural network (DNN)[34] on eCLIP-seq data[35] was downloaded from the Kipoi repository[80]. In order to
605 test the binding probability of the 101 proteins on *SchlafenInc*, a bed file containing the coordinates of
606 *SchlafenInc*'s 4 exons (ENSMUST00000143673.2 on Ensembl) was obtained. Then for each exon, we
607 took sliding windows of 101nt length resulting on a new bed6 file where each row is of 101nt length.
608 Kipoi builds on environments, making the sharing and loading of models easy. We started our analysis
609 by generating an environment with all the needed dependencies. Afterwards for each protein of interest,
610 we programmatically obtained the dataloader, load the fasta sequence for GRCm38.p6, load the
611 GENCODE annotation for mouse version vM22[18], and a bed file with the pre-extracted 101nt length
612 sliding windows and ran the predict function with default arguments (considering all the genomic
613 landmarks also for non-protein coding genes). Finally, for each protein of interest a csv file was
614 generated, containing the following info for each 101nt sequence considered: the chromosome, the

615 starting position, the end position, a sequence ID, a score as in the bed6 file, the strand and the binding
616 score [0 (no binding probability), 1 (high binding probability)] of the protein of interest on that window.

617 Software list

618 Leica Application Suite Advanced Fluorescence, version 2.7.9723
619 Invitrogen EVOS FL Auto 2 Software
620 CellRanger, version 5.0.0
621 Seurat, version 3.2
622 RNA STAR aligner, version 2.6.0b-2
623 StringTie and StringTie merge, version 1.3.3.2 and 1.3.6
624 Bedtools Intersect tool, version v2.29.0
625 "Extract features" tool, version 1.0.0
626 FEELnc, version 0.1.1.1 (Galaxy) and version 0.2
627 Coding-Potential Assessment Tool, CPAT, version 2.0.0
628 DESeq2, version 2.11.40.2
629 ClustVis R package, version 0.0.0.9
630 Cytoscape, version 3.8.2
631 ClueGO, version 2.5.8
632 MetaMorph Offline, version 7.10.1
633 Prism 8 for Mac OS, version 8.4.1
634 Adobe Illustrator for Mac OS, version 24.2.1
635 R studio, version 3.6.3
636 Microsoft Office for Mac

637 Statistical Analysis

638 Statistical analysis was performed using GraphPad Prism 8. To test for outliers Grubbs test (alpha=0.05)
639 was used. Data distribution and the difference between variances were tested using the Shapiro-Wilk
640 normality test. For statistical analysis of two groups, unpaired t-test was performed for normal distributed
641 samples, otherwise Mann-Whitney test was used. For more than two groups, two-way ANOVA was
642 performed. For chemotaxis experiments, area under the curve (AUC) was calculated and statistical
643 significance was tested using unpaired t-test. Significance is depicted as $*p < 0.05$, $**p < 0.01$,
644 $***p < 0.001$, $****p < 0.0001$.

645 **ACCESSION NUMBERS**

646 All sequencing data sets can be accessed at GEO database under GSE210732.

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661 **Author contributions**

662 A.D. and S.E. designed the study and wrote the manuscript. A.D., L.A. D.E., S.B., E.T., K.H., N.P. and
663 A.U. performed the experiments or analyzed the resulting data. M.B. provided the *SchlafenInc* knockout
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667 **CONFLICT OF INTEREST**

668 None declared.

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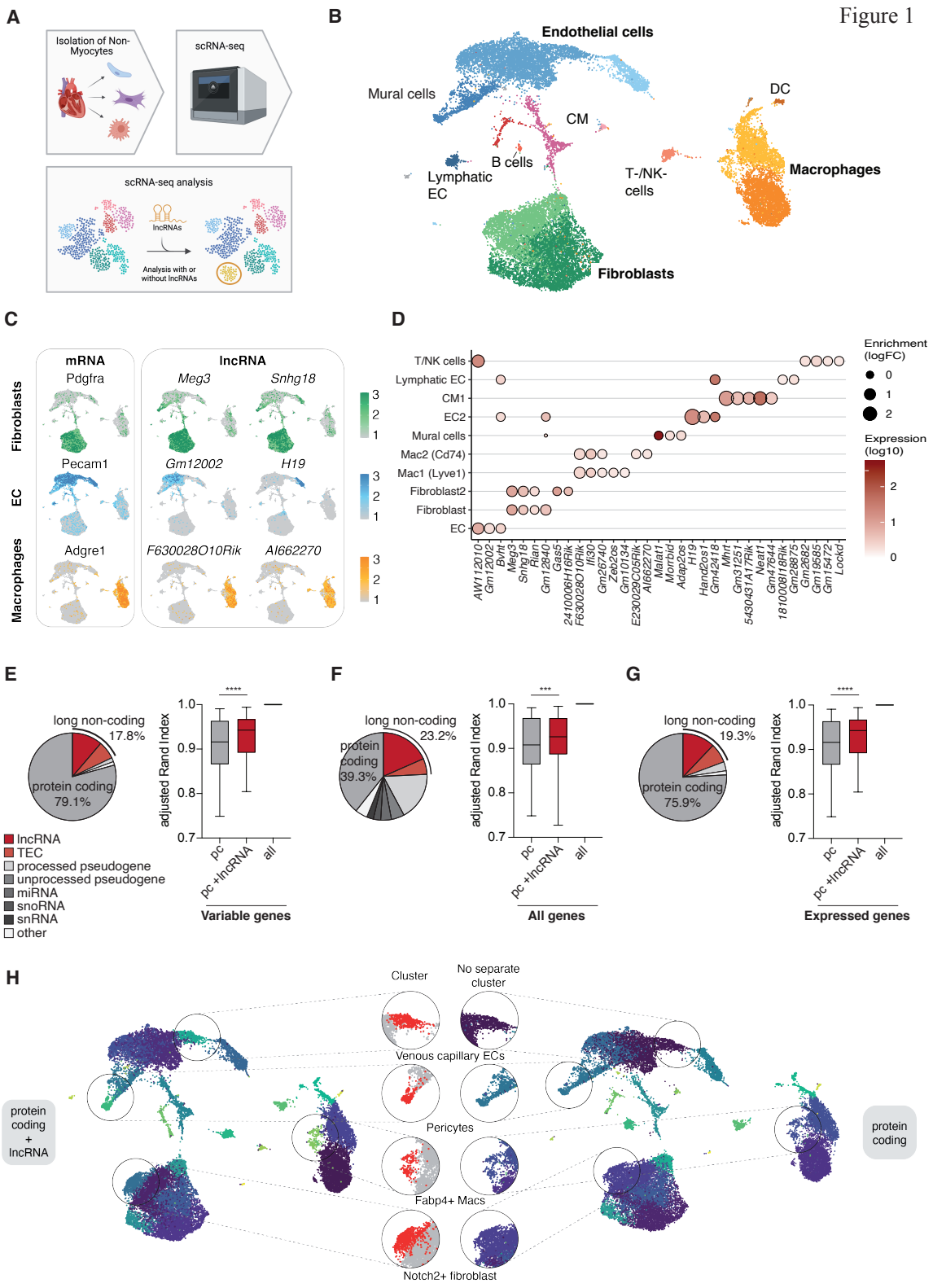


Figure 1: LncRNA are important determinants for definition of specific cell types.

(A) Illustration for the measurement of lncRNA in murine myocardium and re-clustering analysis. (B) UMAP projection of murine myocardium as measured by single-cell sequencing (scRNA-seq). Cell populations are colored by type as indicated by the label. DC: dendritic cells, EC: endothelial cells, CM:

839 cardiac myocytes. (C) Bubble plot showing lncRNA marker genes conserved across conditions. Bubble
840 size corresponds to enrichment (logFC) and fill color to expression value (log10). Only values/bubbles
841 with a significant enrichment (p-val < 0.05) are shown. (D) Feature plots showing mRNA and lncRNA
842 markers for fibroblasts, endothelial cells (EC) and macrophages. (E), (F), (G) Adjusted Rand Index (ARI)
843 calculation. In order to compare the clusterings with and without lncRNAs, the respective complete gene
844 set was considered to yield the optimal clustering. We iterated the clustering in 0.01 steps with
845 resolutions ranging from 0.10 to 1.00, testing for the similarity to optimal clustering using adjusted Rand
846 index scoring. Here, the more similar a clustering would be to the "optimal" one the closer it will be to 1.
847 Displayed as boxplot with median and quartiles. (E) ARI calculation for the top variable genes (3,000
848 genes). (F) ARI calculation for all annotated genes (Gencode vM22[18]). (G) ARI calculation for all
849 expressed genes (expression value >0). pc: protein coding (H) Umap projection of murine myocardium
850 with cell population clustering based on protein coding genes plus lncRNA genes (left panel) or only
851 protein coding genes (right panel). The middle panel shows differentially assigned cell population
852 clusters.

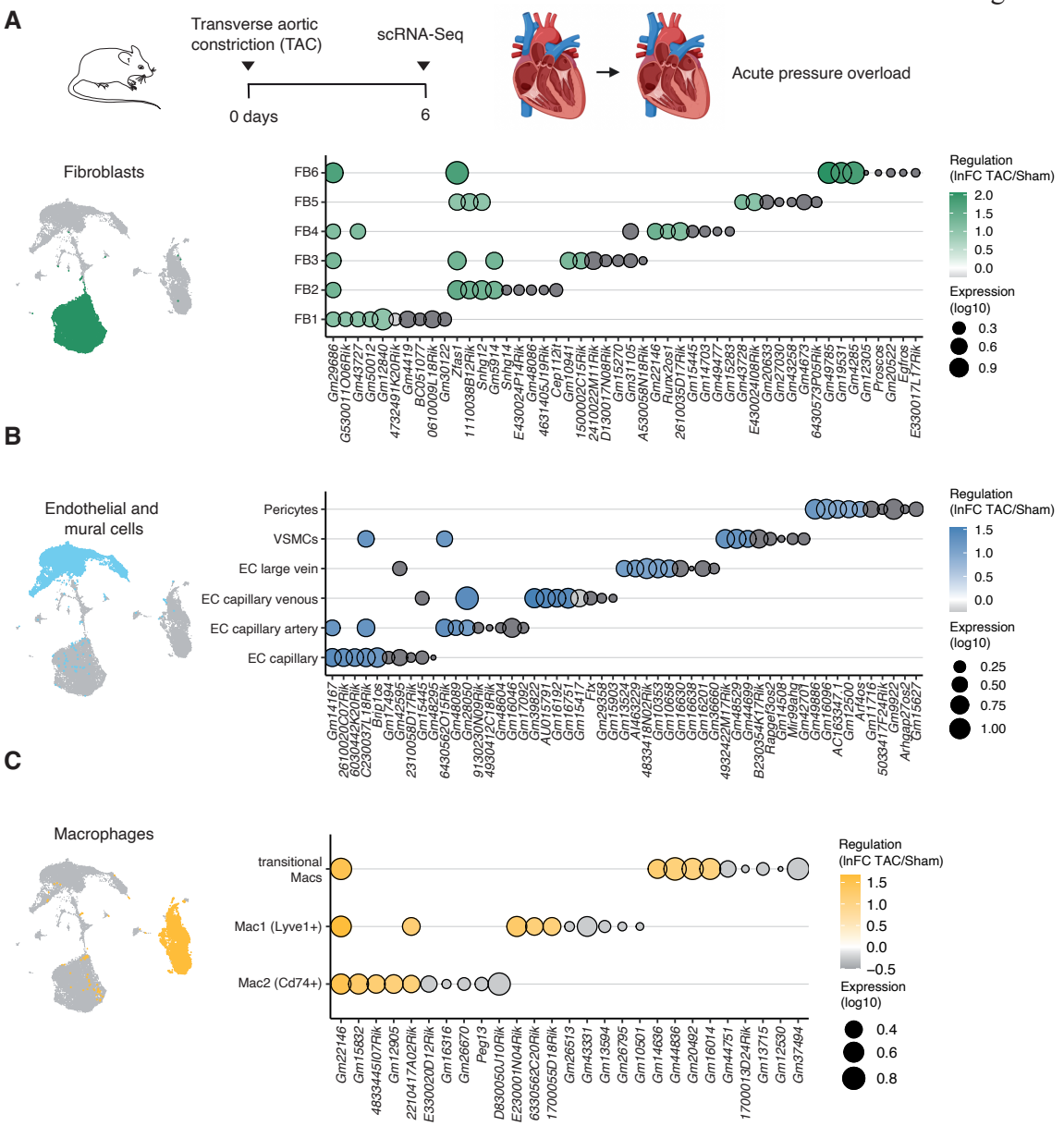


Figure 2: Dynamic changes of lncRNA expression in crMPs during pressure overload.

Bubble plot showing the top 5 up-regulated and top 5 down-regulated lncRNA genes after TAC for the sub-populations of the major cell types in the heart: (A) Fibroblasts (B) Endothelial cells and (C) Macrophages. Fill color corresponds to regulation in TAC vs. sham (lnFC TAC/sham) and bubble size corresponds to expression value (log10). Only values with a significant enrichment (p-val < 0.05) are shown. Some genes are among the top regulated genes in more than one cell type.

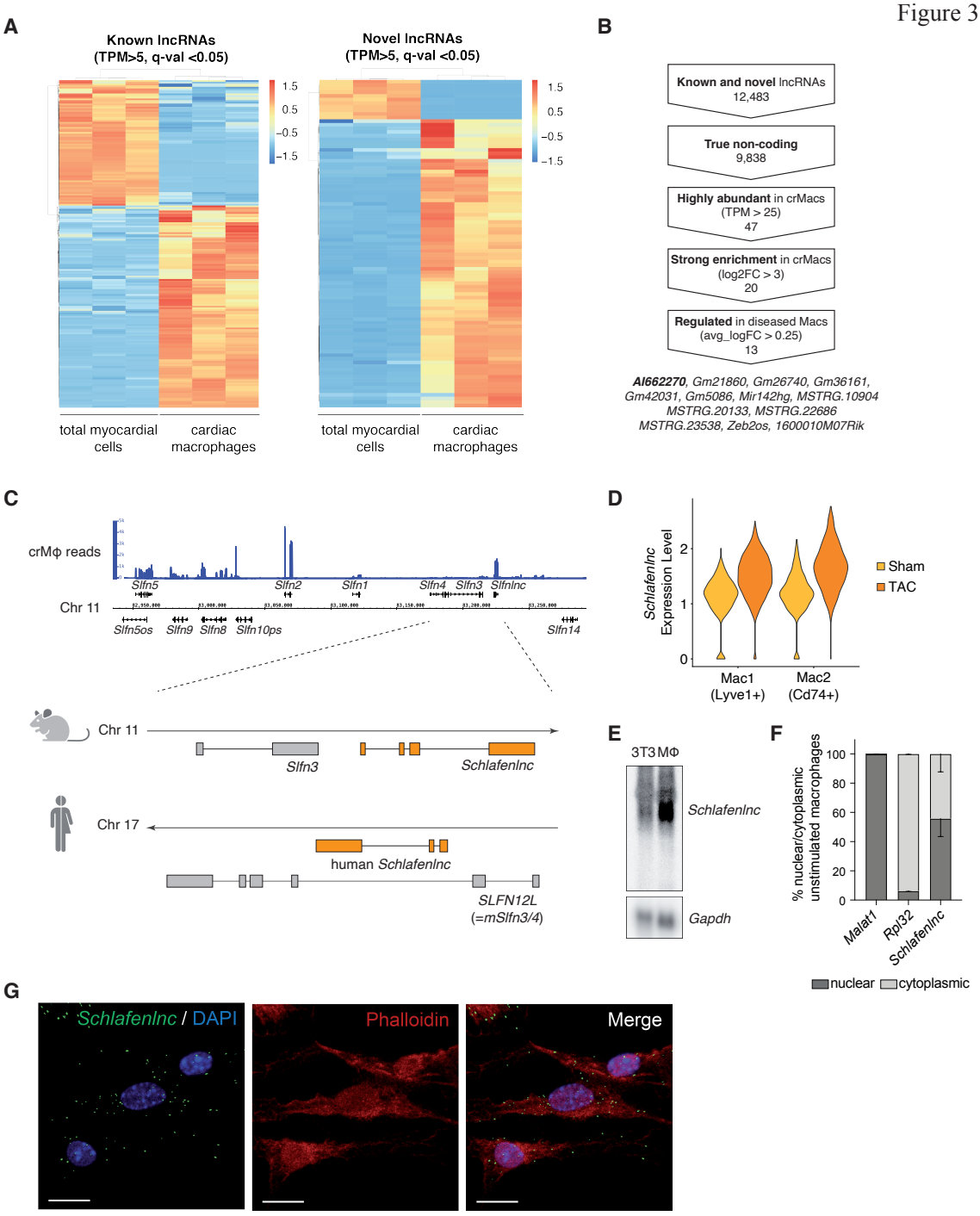


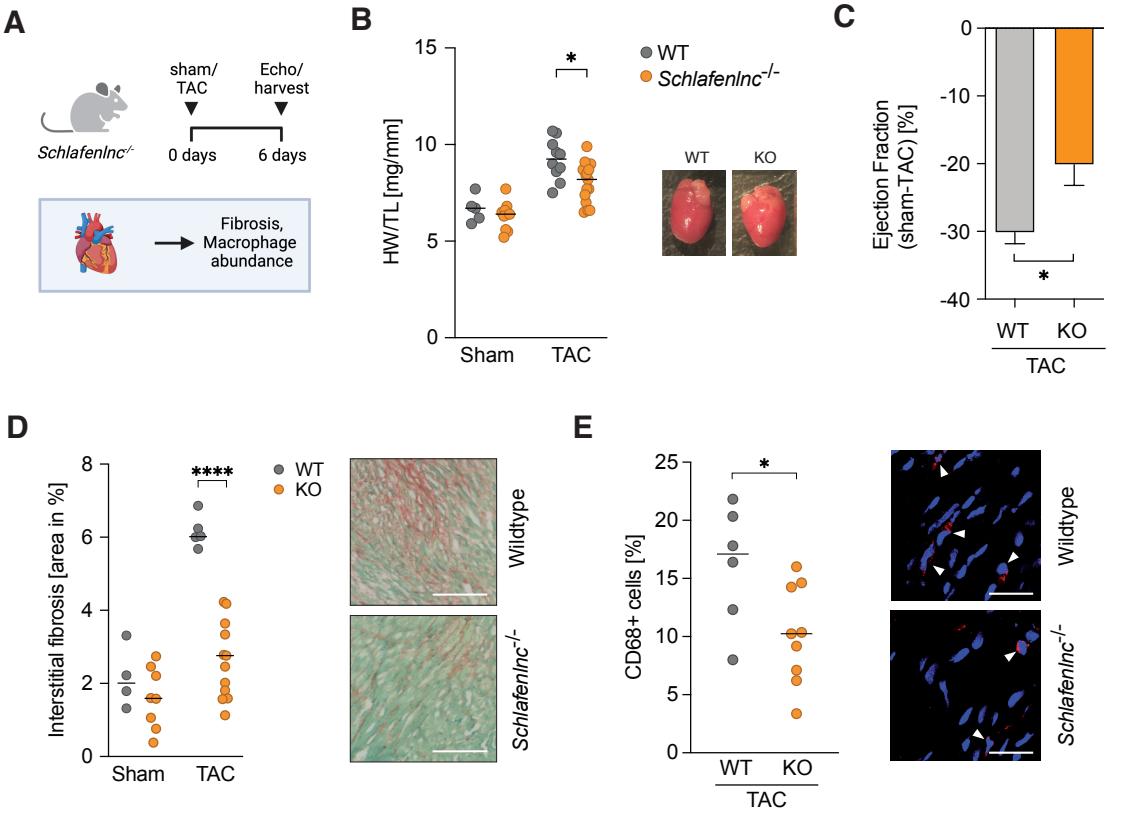
Figure 3: Determination and annotation of the lncRNA landscape of cardiac resident macrophages.

(A) Unsupervised clustering of known and novel lncRNAs using normalized counts values, rows were averaged across samples for clustering. Genes shown are expressed with a TPM value >5 and differentially regulated (enriched) with a q-val of <0.05. (B) Flow scheme for the identification of lncRNA candidates using the indicated filters. (C) The Schlafen family locus showing the expression in cardiac resident macrophages (accumulated plus and minus strand). Lower panel: Zoom in of the *Schlafenlnc* locus in mouse as well as human for comparison. (D) Expression of the lncRNA *Schlafenlnc* in Mac1 (*Lyve1*+ macrophages) and Mac2 (*MHCII* high, *Cd74*+ macrophages) after sham or TAC in the scRNA-

870 seq data. (E) Northern Blot showing the expression of *SchlafenInc* transcripts in bone marrow-derived
871 macrophages in comparison to fibroblasts (NIH3T3 cell line). (F) Relative nuclear and cytoplasmic
872 localization of *Malat1* (nuclear lncRNA), *Rpl32* (mRNA) and *SchlafenInc* as determined by fractionation
873 of macrophages followed by quantification through RNA-Seq. Data are normalized counts displayed as
874 mean with SEM. (G) Detection of *SchlafenInc* (green) in macrophages by RNA *in situ* hybridization
875 (RNAScope). Nuclei were stained with DAPI (blue), Phalloidin (red) was used for outlining the cellular
876 body (Scale bars = 15µm).

877

Figure 4



878

879 **Figure 4: *SchlafenInc*-deficient mice show improved cardiac function and reduced inflammatory**
880 **response after transverse aortic constriction (TAC).**

881 (A) Experimental setup of cardiac phenotyping of *SchlafenInc*$^{-/-}$ and wildtype mice. (B) Quantification of
882 normalized heart weight after TAC. HW= heart weight, TL= tibia length. Black line = median. C) Ejection
883 fraction changes after 6 days TAC. Data displayed as mean with SEM. (C) Left: Quantification of
884 interstitial fibrosis by Sirius red/Fast green staining. Black line = median. Right: Representative images
885 of interstitial fibrosis, scale bar = 200µm. (D) Representative images and quantification of
886 macrophage/monocyte content by CD68 staining in WT and *SchlafenInc*$^{-/-}$ hearts after 6d after TAC.
887 Black line = median. White arrows label CD68-positive cells, scale bar = 25µm.

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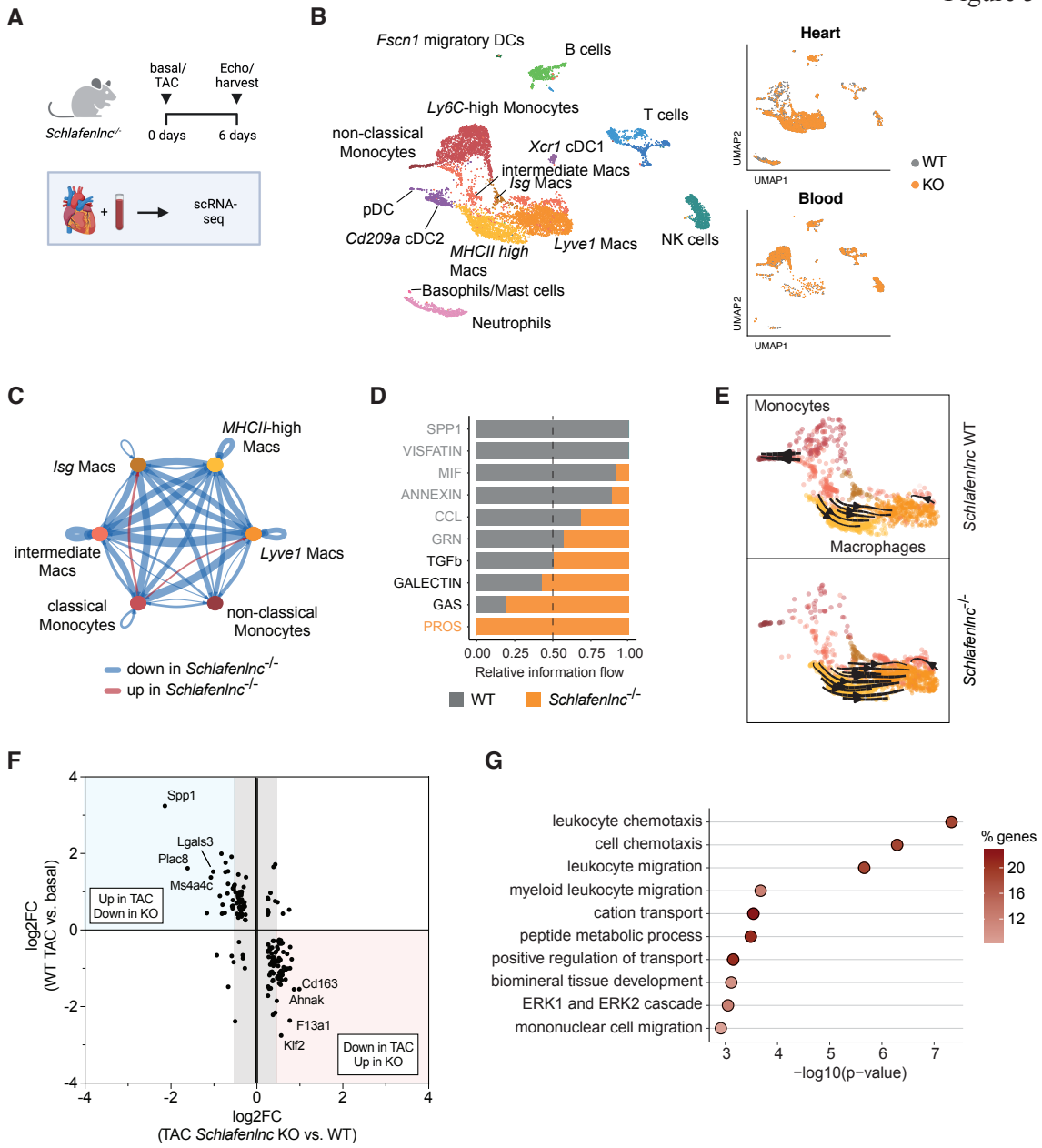

Figure 5: scRNA-seq of *Schlafenlnc*$^{-/-}$ mice reveals macrophage polarization towards an anti-inflammatory phenotype after TAC.

(A) Experimental setup of scRNA-seq of *Schlafenlnc*$^{-/-}$ and wildtype mice. (B) Dimensional reduction (Umap) representation of combined measured cells with assigned cell types. (C) Cell interaction analysis of *Schlafenlnc*$^{-/-}$ and wildtype mice (*Cd45*+ cells from blood and heart) after TAC using CellChat[70]. (D) Differential signaling in *Schlafenlnc*$^{-/-}$ and WT macrophage and monocyte populations after TAC. Light grey and orange highlighted pathways are significantly enriched in wildtype or *Schlafenlnc*$^{-/-}$, respectively (p-val<0.05). (E) RNA Velocity of *Schlafenlnc*$^{-/-}$ and WT cells from heart after 6 days of TAC. Shown are macrophage and monocyte cell clusters. (F) Comparison of de-regulated genes (q-val<0.05) in TAC vs. basal condition in WT compared to de-regulated genes after 6d TAC KO

900 vs. WT in *Lyve1*$^{+}$ macrophages (G) Gene ontology terms of genes de-regulated in *SchlafenInc*-deficient
901 mice and after 6 days TAC.

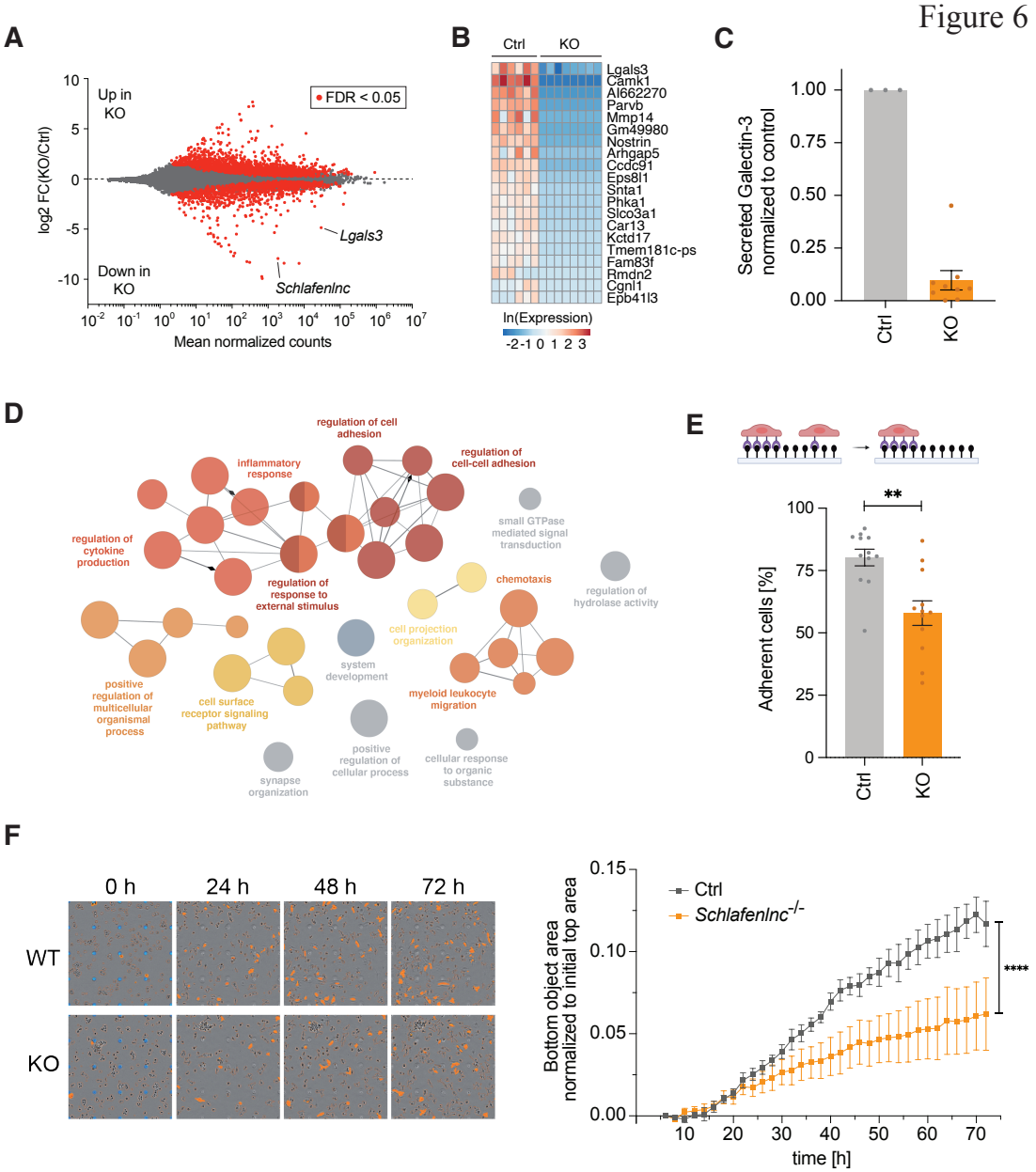

Figure 6: *SchlafenInc* is required for the activation of macrophage gene programs for cell adhesion and migration.

(A) MA plot showing differentially expressed genes in *SchlafenInc*$^{-/-}$ vs. Ctrl macrophages. Significantly deregulated genes (FDR < 0.05) are shown in red. (B) Heatmap of an unsupervised clustering showing the top 20 downregulated genes in *SchlafenInc*$^{-/-}$ macrophages. (C) Measurement of Galectin-3 protein secretion by control and *SchlafenInc*$^{-/-}$ macrophages as determined by ELISA. The supernatant of one control cell line and three *SchlafenInc*$^{-/-}$ clones were used in n=3 independent experiments. Data points are displayed as mean ±SEM. (D) Functionally grouped gene ontology term analysis of biological processes associated with significantly downregulated genes (FDR < 0.05, log2FC < -1) in *SchlafenInc*$^{-/-}$ macrophages. (E) Adhesion assay with control and *SchlafenInc*$^{-/-}$ macrophages (Ctrl clones n=2, KO clones n=3; 4 independent experiments). Data points are displayed as mean ±SEM. (F) Chemotaxis assay against a gradient of the chemokine C5a with control and *SchlafenInc*$^{-/-}$ macrophages

915 stimulated with 10ng/ml LPS overnight (Ctrl clones n=1, KO clones n=3; 3 independent experiments).
916 Data points are displayed as mean ±SEM.