Tolerance-conferring defensive symbionts and the evolution of parasite virulence

Cameron A. Smith[1] and Ben Ashby[1,2,3,4]

1. Department of Mathematical Sciences, University of Bath.

2. Milner Centre for Evolution, University of Bath.

3. Department of Mathematics, Simon Fraser University.

4. The Pacific Institute on Pathogens, Pandemics and Society (PIPPS), Simon Fraser University.

Keywords: defensive symbiosis, mutualism, parasitism, biocontrol, coevolution, tolerance

10 ABSTRACT

Defensive symbionts in the host microbiome can confer protection from infection or reduce the

harms of being infected by a parasite. Defensive symbionts are therefore promising agents of

biocontrol that could be used to control or ameliorate the impact of infectious diseases. Previous

theory has shown how symbionts can evolve along the parasitism-mutualism continuum to confer

greater or lesser protection to their hosts, and in turn how hosts may coevolve with their symbionts

to potentially form a mutualistic relationship. However, the consequences of introducing a defensive

symbiont for parasite evolution and how the symbiont may coevolve with the parasite have received

relatively little theoretical attention. Here, we investigate the ecological and evolutionary

implications of introducing a tolerance-conferring defensive symbiont into an established host-

parasite system. We show that while the defensive symbiont may initially have a positive impact on

the host population, parasite and symbiont evolution tend to have a net negative effect on the host

population in the long-term. This is because the introduction of the defensive symbiont always

selects for an increase in parasite virulence and may cause diversification into high- and low-

virulence strains. Even if the symbiont experiences selection for greater host protection, this simply

increases selection for virulence in the parasite, resulting in a net negative effect on the host

population. Our results therefore suggest that tolerance-conferring defensive symbionts may be

poor biocontrol agents for population-level infectious disease control.

LAY SUMMARY

Defensive symbionts – microbes that confer protection to a host against a harmful parasite – are found throughout the natural world and represent promising candidates for biological control to combat infectious diseases. Symbionts can protect their hosts through a variety of mechanisms that may prevent infection (resistance) or increase survival following infection (tolerance), yet our understanding of the ecological and evolutionary impact of defensive symbionts on parasites is limited. Moreover, few theoretical predictions exist for how defensive symbionts are likely to evolve in the presence of parasites, and for the net effect on the host population. Using a mathematical model where defensive symbionts reduce parasite virulence (harm to the host), we investigate the impact of their introduction on the evolution of parasite virulence, how selection increases or decreases host protection, and whether such symbionts are beneficial for the host population. We find that this form of defensive symbiosis always selects for higher parasite virulence and that it can cause the parasite to diversify into high and low virulence strains which specialise on different host subpopulations. Crucially, we show that the introduction of a defensive symbiont will always lead to a long-term reduction in host population size even if they are beneficial in the short-term. Together, our results show that defensive symbionts can have a strong impact on the evolution of virulence and that this form of host protection is not robust, indicating that tolerance-conferring symbionts are likely to be poor candidates for biological control of infectious diseases at the population level.

INTRODUCTION

Defensive symbiosis, where an organism confers protection to its host from a natural enemy such as a parasite or predator is widespread in nature (reviewed in (Ford and King 2016)). For example, ants have long been known to defend acacia trees from herbivores (Belt, Thomas 1874) and various bacteria have been shown to confer protection directly or indirectly against bacterial and fungal parasites across diverse host taxa, including insects (Oliver et al. 2003; Cariveau et al. 2014), plants (Herre et al. 2007; Arnold et al. 2003), invertebrates (Gil-Turnes, Hay, and Fenical 1989; Gil-Turnes and Fenical 1992), and vertebrates (Lauer and Hernandez 2015; Heikkilä and Saris 2003). Protection can be conferred to hosts as resistance (preventing infection) or tolerance (increasing survival following infection) through a variety of mechanisms (Troha and Ayres 2022), including through interactions with the host's immune system (Ford, Drew, and King 2022), interference competition through chemical defences - for example, *Streptococcus pneumoniae* can produce hydrogen peroxide to displace *Staphylococcus aureus* in the nasopharynx (Selva et al. 2009) – and resource competition or priority effects (Hancock, Sinkins, and Godfray 2011; Moreira et al. 2009). Defensive symbionts therefore have potential as agents of biocontrol, especially in the context of infectious diseases for therapeutic use (Bakken et al. 2011) or for population-level control (Utarini et al. 2021). The use of defensive symbionts should be approached with caution, however, as the nature and extent of protection conferred to their hosts is evolvable and they could alter both the ecological and evolutionary dynamics of hosts and parasites, potentially leading to unintended consequences.

Crucially, the protective relationship between a defensive symbiont and its host is not fixed; it may be context dependent, due to changes in the biotic or abiotic environment (González et al. 2021; King et al. 2016; Ashby and King 2017; Rafaluk-Mohr et al. 2018; Rogalski et al. 2021; Chamberlain, Bronstein, and Rudgers 2014; Lin and Koskella 2015), and it is subject to selection (King et al. 2016; Rafaluk-Mohr et al. 2022). For example, the removal of large herbivores can lead to the loss of acacia

tree protection by ants (Palmer et al. 2008) and protective microbes such as *Enterococcus faecalis*

reduce nematode fitness in the absence of *Staphylococcus aureus* but can be experimentally evolved

to rapidly increase protection of their hosts when *S. aureus* is present (King et al. 2016). An organism

may therefore be parasitic to its host in isolation, but may be protective – and may evolve to be

more or less protective – when another parasite is present (Ashby and King 2017; Rafaluk-Mohr et

al. 2018). Understanding evolution along the parasitism-mutualism continuum is therefore a key

challenge for evolutionary biologists, especially in the context of the gut microbiome and infectious

diseases. In particular, understanding the evolutionary robustness of host protection is particularly

important when defensive symbionts are used as biocontrol agents, as their effectiveness will

depend on both on the initial impact on the parasite, as well as the subsequent coevolutionary

dynamics between host protection and parasite virulence.

The evolution of parasite virulence has long been a focus of theoretical studies of host-parasite

systems (S. Levin and Pimentel 1981; R. May and Anderson 1983; Lenski and May 1994). Theoretical

studies of host-associated communities have primarily focused on the effects of within- and

between-host competition on the evolution of virulence (Alizon 2013; Frank 1992; 1996; Brown,

Hochberg, and Grenfell 2002; R. May and Nowak 1995). By comparison, few theoretical studies have

explored microbial evolution in the context of defensive symbiosis. Ashby and King (2017) explore

how host protection evolves in the presence of a non-evolving parasite population, showing that

conferred tolerance and resistance could readily evolve under a wide range of conditions, potentially

leading to symbiont diversification into a highly protective strain and one that conferred no

protection. This model was extended by Rafaluk-Mohr et al. (2018) to explore symbiont coevolution

with the host, showing that the host becomes more mutualistic towards the symbiont at

intermediate levels of protection. Nelson and May (2017) investigate the evolution of symbionts

along the full mutualism-parasitism continuum when there is a shared cost of virulence. They show

that the community of symbionts maintain mutualisms and evolve lower virulence when the shared

costs are sufficiently low, but higher virulence may evolve when shared costs are high. Nelson and

May (2020) extend this model to show that if increased defence is evolved by one symbiont, it may

facilitate the reduction of virulence in both symbionts present, and in some cases cause pathogens

to evolve towards mutualism. Together, these studies highlight the complex context-dependent

nature of coevolution between mutualistic and parasitic symbionts. A key question, yet to be

addressed by previous studies, is how do defensive symbionts drive the evolution of virulence in

obligate parasites? Furthermore, how does the evolution of virulence affect the evolution of host

protection, and what are the consequences for the host population?

Here, we use a mathematical model to address these questions. We explore the (co)evolution of

parasite virulence and host protection – specifically, tolerance – by a defensive symbiont. Although

tolerance can take many form (Rafaluk-Mohr et al. 2022), here we focus on mortality tolerance,

whereby infected hosts that possess the defensive symbiont experience a lower mortality rate than

infected hosts that do not possess the defensive symbiont. We first show how the introduction of a

defensive symbiont always selects for greater parasite virulence, and that the defensive symbiont

can induce the parasite to diversify into high and low virulent strains. We then show how the shape

of life-history trade-offs associated with host protection affect the outcome of symbiont-parasite

coevolution, and that this always results in a reduction in the host population size in the long term.

MATERIALS AND METHODS

MODEL

We consider a well-mixed population of hosts with two co-circulating microbes: an obligate parasite

that increases host mortality and a defensive symbiont that may confer tolerance to infected hosts

by reducing their disease-associated mortality rate. Hosts may exist in one of four states, where they

harbour: no microbes (H), defensive symbionts only (D), parasitic microbes only (P) or both (B).

New hosts are born at rate $v(N) = N(a - qN)$, where $N = H + D + P + B$ is the total number of

hosts, a is the maximum per capita rate of reproduction and q controls the strength of density

dependent competition among hosts. All hosts, regardless of infection status, have a natural

mortality rate b.

We assume that transmission is density-dependent, occurring at a baseline rate of $\hat{\beta}_D$ for the

defensive microbe with a clearance rate of γ_D, and β_P for the parasite with a clearance rate of γ_P.

There is no vertical transmission (all individuals begin life without either microbe), co-transmission

does not occur (i.e., hosts must transition through one of the single-microbe classes to reach class

B), and there is no long-lasting immunity. Both defensive and parasitic microbes increase the

baseline mortality rate of the host, by α_D and $\bar{\alpha}_P$, respectively. We assume that the parasite

experiences a power-law trade-off between transmission and virulence (the additional microbe-

induced mortality) such that $\alpha_P(\beta_P) = \hat{\alpha}_P\left(1 + \beta_P^d\right)$ with $d > 1$ so that there are diminishing returns

for increased virulence. Note that due to this positive correlation between parasite transmission and

virulence, we will interchangeably refer to transmission and virulence evolution throughout.

Defensive microbes may confer protection to parasitised hosts in the form of tolerance, $y \in [0,1]$,

such that the additional mortality rate for hosts with both microbes, $\alpha_B(y, \beta_P)$, satisfies $\alpha_B(y, \beta_P) \leq$

$\alpha_P(\beta_P) + \alpha_D$ (i.e. it is less than or equal to the sum of the additional mortality rates). However, the

defensive microbe incurs a fitness cost when it diverts resources to protect a host, resulting in a

reduction in its transmissibility such that $\beta_D(y) = \hat{\beta}_D\left(1 - c(y)\right)$, where $c(y)$ is an increasing, non-

linear cost function:

$$c(y) = \begin{cases} \dfrac{c_1(1 - e^{c_2 y})}{1 - e^{c_2}}, & c_2 \neq 0 \\ c_1 y, & c_2 = 0 \end{cases}, \tag{1}$$

where $c_1 \in [0,1]$ is the strength of the cost function, denoting the maximum reduction in

transmission when tolerance is maximised at $y = 1$, and c_2 controls the shape of the trade-off:

when $c_2 > 0$, conferring protection is increasingly costly (an accelerating trade-off), and when

$c_2 < 0$ conferring protection is decreasingly costly (a decelerating trade-off). Biologically, an

accelerating (respectively, decelerating) trade-off corresponds to diminishing (increasing) returns,

such that for a given increase in the cost, the amount of additional host protection decreases

(increases) as it gets stronger. Thus, for accelerating trade-offs there is a relatively small initial cost

but the costs become ever more extreme at higher levels of protection, whereas for decelerating

trade-offs there is a relatively large initial cost but the additional costs become less extreme at

higher levels of protection.

The ecological dynamics of monomorphic populations are shown schematically in Fig. 1 and are

governed by the following ordinary differential equations (ODEs)

$$\frac{dH}{dt} = \nu(N) - [b + \beta_D(y)(D + B) + \beta_P(P + B)]H + \gamma_D D + \gamma_P P, \tag{2}$$

$$\frac{dD}{dt} = \beta_D(y)H(D + B) - [b + \gamma_D + \alpha_D + \beta_P(P + B)]D + \gamma_P B, \tag{3}$$

$$\frac{dP}{dt} = \beta_P H(P + B) - [b + \gamma_P + \alpha_P(\beta_P) + \beta_D(y)(D + B)]P + \gamma_D B, \tag{4}$$

$$\frac{dB}{dt} = \beta_D(y)P(D + B) + \beta_P D(P + B) - [b + \alpha_B(y, \beta_P) + \gamma_D + \gamma_P]B. \tag{5}$$

ANALYSIS

We employ evolutionary invasion analysis using a combination of numerical analysis and simulations

to establish how parasite virulence (α_P) evolves following the introduction of the defensive

symbiont, and in turn how the defensive symbiont co-evolves to be more or less protective (y)

following its introduction.

We use the next generation method (Diekmann, Heesterbeek, and Roberts 2010) (see

Supplementary Material) to derive the invasion fitness for a rare defensive symbiont with protection

y^m (denoted w_D), or a rare parasite with transmission rate β_P^m and virulence α_P^m (denoted w_P),

when introduced into a population at equilibrium with resident traits $\boldsymbol{\theta}^r = (y^r, \beta_P^r)$:

$w_D(y^m \mid \boldsymbol{\theta}^r)$ $$= \frac{\beta_D(y^m)\{H^*[b + \gamma_D + \gamma_P + \alpha_B(y^m, \beta_P^r) + \beta_P^r(P^* + B^*)] + P^*[b + \gamma_D + \gamma_P + \alpha_D + \beta_P^r(P^* + B^*)]\}}{\left(b + \gamma_D + \alpha_D + \beta_P^r(P^* + B^*)\right)\left(b + \alpha_B(y^m, \beta_P^r) + \gamma_D + \gamma_P\right) - \gamma_P \beta_P^r(P^* + B^*)},$$	(6)
$w_P(\beta_P^m \mid \boldsymbol{\theta}^r)$ $$= \frac{\beta_P^m\{H^*[b + \gamma_D + \gamma_P + \alpha_B(y^r, \beta_P^m) + \beta_D(y^r)(D^* + B^*)] + D^*[b + \gamma_D + \gamma_P + \alpha_P(\beta_P^m) + \beta_D(y^r)(D^* + B^*)]\}}{\left(b + \gamma_P + \alpha_P(\beta_P^m) + \beta_D(y^r)(D^* + B^*)\right)\left(b + \alpha_B(y^r, \beta_P^m) + \gamma_D + \gamma_P\right) - \gamma_D \beta_D(y^r)(D^* + C^*)},$$	(7)

where each of the steady states (indicated with asterisks) are functions of the resident traits, for

example $H^* \equiv H^*(\boldsymbol{\theta}^r)$. We are unable to obtain an analytical expression for these steady states, so

we approximate them in our numerical analysis by simulating the ODE system for a sufficiently long

period of time so that the population approaches its unique, locally asymptotically stable, endemic

equilibrium. We derive the respective fitness gradients, $\mathcal{F}_D(y) = \left.\frac{\partial w_D}{\partial y^m}\right|_{y^m = y}$ and $\mathcal{F}_P(\beta_P) =$

$\left.\frac{\partial w_P}{\partial \beta_P^m}\right|_{\beta_P^m = \beta_P}$, from equations (6)-(7) (omitted for brevity) and find singular strategies, y^* and β_P^*, by

numerically solving $\mathcal{F}_D(y^*) = 0$ and $\mathcal{F}_P(\beta_P^*) = 0$. Singular strategies are evolutionarily stable if

$\mathcal{E}_D(y^*) = \left.\frac{\partial^2 w_D}{\partial(y^m)^2}\right|_{y^m = y = y^*} < 0$ and $\mathcal{E}_P(\beta_P^*) = \left.\frac{\partial^2 w_P}{\partial(\beta_P^m)^2}\right|_{\beta_P^m = \beta_P = \beta_P^*} < 0$, respectively. For parasite

evolution only, we determine convergence stability by numerically evaluating the derivative

$C_P(\beta_P^*) = \frac{\partial^2 w_P}{\partial \beta_P^m \partial \beta_P}\Big|_{\beta_P^m = \beta_P = \beta_P^*}$ and checking that $\mathcal{E}_P(\beta_P^*) < C_P(\beta_P^*)$. In the case of coevolution, we

assume equal mutation rates for defensive symbionts and parasites, and determine strong

convergence stability using the method presented in (Leimer 2009) (see *Supplementary Material*).

We assume that the defensive symbiont is introduced into a well-established host-parasite system,

with the parasite at its unique continuously stable strategy (CSS) in the absence of the defensive

symbiont (see *Supplementary Material*), $\tilde{\beta}_P^*$, which is given by

$$\tilde{\beta}_P^* = \left(\frac{b + \gamma_P + \hat{\alpha}_P}{(d-1)\hat{\alpha}_P}\right)^{\frac{1}{d}}. \tag{8}$$

In addition to exploring the effects of the defensive symbiont on the (co)evolution of virulence and

host protection, we measure the net effect on the host population size and change in the average

host mortality rate (relative to the initial symbiont-free population). The net effect on the host

population size is measured by comparing the steady state in the presence and absence of the

defensive symbiont, $N^*(y, \beta_P)$ and $\tilde{N}^*$ respectively. Similarly, we calculate the average disease-

associated mortality rate at equilibrium in the presence and absence of the defensive symbiont,

$r^*(y, \beta_P)$ and $\tilde{r}^*$, respectively as:

$$r^*(y, \beta_P) = \alpha_D \frac{D^*(y, \beta_P)}{N^*(y, \beta_P)} + \alpha_P(\beta_P)\frac{P^*(y, \beta_P)}{N^*(y, \beta_P)} + \alpha_C(y, \beta_P)\frac{B^*(y, \beta_P)}{N^*(y, \beta_P)}, \tag{9}$$

$$\tilde{r}^* = \tilde{\alpha}_P \frac{\tilde{P}^*}{\tilde{N}^*}, \tag{10}$$

194   where we have explicitly written the dependence of the trait variables on the steady state values.

195   Note that the values for $\tilde{N}^*$ and $\tilde{r}^*$ are constants calculated at the singular strategy for a parasite

196   circulating in the absence of the defensive symbiont, where we begin all of our evolutionary

197   simulations. We then define the following two measures to determine the net effects on the host

198   population following the introduction of the defensive symbiont:

$$Q_1(y, \beta_P) = 100 \left(\frac{N^*(y, \beta_P)}{\tilde{N}^*} - 1 \right), \tag{11}$$

$$Q_2(y, \beta_P) = 100 \left(1 - \frac{r^*(y, \beta_P)}{\tilde{r}^*} \right). \tag{12}$$

The first measure (equation 11) is the percentage increase in the host population size and the

second measure (equation 12) is the percentage decrease in the disease-associated mortality rate.

SIMULATIONS

The above analysis makes two key assumptions: (1) that there is a separation of the ecological and

evolutionary time scales (i.e. mutations are rare), and (2) that selection is weak, so that mutations

only have a small phenotypic effect (i.e. traits are continuous). We relax these assumptions in our

simulations by allowing new mutants to arise before the ecological dynamics are close to their

ecological attractor and by discretising the trait space so that new mutations have small but finite

effects. Simulations proceed as follows (described for the coevolution case). We initialise a resident

population which has a defensive symbiont protection level of y^r and a parasite transmission of $\tilde{\beta}_P^*$

as defined in equation (8). We simulate the ecological dynamics (1)-(4) for a total (arbitrary) time of

$T_{eco} = 100$. We choose either the defensive symbiont or parasite population with equal probability

and introduce a mutant at low frequency with trait value differing from the resident by a small

amount, ϵ_D or ϵ_P. We then run the ecological dynamics again for another T_{eco} time units, remove

any phenotypes that have dropped below a frequency of $\varepsilon = 10^{-4}$ (this threshold is arbitrary) and

then introduce a new mutant again, by firstly choosing the defensive symbiont or parasite with

equal probability and then choosing a trait to mutate proportional to its frequency. This continues

for a total of T_{evo} evolutionary time-steps.

RESULTS

We begin by exploring how the introduction of a (non-evolving) defensive symbiont affects the

quantitative and qualitative evolution of parasite virulence, before considering coevolution of both

microbes.

DEFENSIVE SYMBIONTS THAT CONFER TOLERANCE ALWAYS SELECT FOR INCREASED

VIRULENCE

The introduction of a non-evolving defensive symbiont, which confers a fixed level of tolerance to

parasitised hosts, always leads to selection for higher parasite virulence (Fig. 2a). This is because the

defensive symbiont not only directly reduces virulence when present with the parasite (hence,

reducing the cost to the parasite of elevated virulence), but also competes with the parasite for

hosts (thus increasing selection for a higher transmission rate, and hence higher virulence) even

when providing little to no protection. The latter effect is more subtle and is typically weaker but is

evident when the defensive symbiont confers no protection to the host ($y = 0$), as the parasite still

evolves increased virulence due to increased competition for hosts. The strength of the first effect

depends on both the level (y) and cost (c_1) of conferred protection, which together determine how

often the parasite shares a host with a symbiont (Fig. 2c). When the cost to the defensive symbiont

of conferring tolerance (c_1) is sufficiently low, greater host protection (y) always selects for higher

parasite virulence because the parasite frequently shares hosts with the defensive symbiont, and so

benefits from decreased realised virulence due to tolerance conferred to the host by the symbiont.

However, when the cost of host protection is relatively high, fewer hosts harbour the defensive

symbiont and so the parasite is less likely to benefit from conferred tolerance, resulting in evolved

virulence peaking at an intermediate level of host protection (Fig. 2a).

As the defensive symbiont confers tolerance to the host, higher evolved virulence does not

necessarily imply that realised virulence will be higher. Yet, following the introduction of the

defensive symbiont, there is always an increase in average realised virulence (i.e. the average level

of virulence experienced by parasitised hosts, with or without the defensive symbiont; Fig. 2b).

Average realised virulence is markedly lower than the increase in evolved virulence (Fig. 2a-b) due to

the presence of the defensive symbiont, but hosts that do not possess the defensive symbiont will

experience the full increase in virulence. Average realised virulence is minimised at an intermediate

level of host protection, where there are relatively more hosts harbouring both microbes (Fig. 2c),

and at high levels of protection there can be a sharp increase in average realised virulence due to a

combination of strong selection for virulence (Fig. 2a) and fewer hosts possessing the defensive

symbiont (Fig. 2c).

DEFENSIVE SYMBIONTS CAN DRIVE PARASITE DIVERSIFICATION

In addition to selecting for higher parasite virulence, the defensive symbiont can also drive

diversification when tolerance is maximised or very close to being maximised ($y \approx 1$), causing the

parasite to branch into two subpopulations (Fig. 3). One of these subpopulations has a high level of

virulence (and transmission), and is primarily found in hosts that also harbour the defensive

symbiont, while the other evolves a much lower level of virulence and is primarily found in hosts

that do not harbour the defensive symbiont (Fig. 3bii). Note that when tolerance is maximised at

$y = 1$, parasite virulence is completely negated in hosts that possess defensive symbionts, but the

two strains are maintained in the population due to their contrasting strategies in isolation (infecting

fully susceptible hosts compared to those that harbour the defensive symbiont) and the frequency

with which they co-occur with the defensive symbiont. Evolutionary branching in parasite virulence

occurs when the strength of the cost to the defensive symbiont is within a relatively narrow range.

When the costs of host protection are below this range, there is only runaway selection for virulence

(Fig. 3ai), and when the costs are above this range, there may be runaway selection for virulence or

a stable level of virulence may evolve (Fig. 3aiii).

SYMBIONT-PARASITE COEVOLUTION CAN BE DETRIMENTAL TO THE HOST POPULATION

We now allow the level of protection conferred by the defensive symbiont to coevolve with parasite

virulence. The parasite, as before, is initialised to its stable level of virulence (equation 8) in the

absence of the defensive symbiont. We then introduce the defensive symbiont at different initial

levels of protection to determine if coevolution results in (i) increased or decreased conferred

protection and (ii) a net cost or benefit to the host population.

We first determine the range of possible evolutionary outcomes for the defensive symbiont as the

cost parameters associated with host protection vary (Fig. 4). It is well-established that trade-off

shapes determine qualitative evolutionary outcomes (Andrew Hoyle et al. 2008) and the range of

outcomes in our model and when they occur is consistent with previous theory (Ashby and King

2017). Under decelerating trade-offs ($c_2 < 0$), the defensive symbiont either maximises or

minimises host protection (potentially depending on the initial level of protection; Fig. 4), as a small

increase from no protection ($y = 0$) is relatively costly, whereas changes at higher levels of

protection are less costly. The defensive symbiont therefore either overcomes the initial cost and

experiences runaway selection for maximal protection, or experiences selection against host

protection. When the costs of host protection accelerate ($c_2 > 0$), the defensive symbiont

maximises protection if the strength of the cost is sufficiently low, and evolves to either an

intermediate level of protection or no protection if the strength of the cost is sufficiently high (Fig.

4).

We now consider how virulence coevolves with host protection to determine the net effect on the

host population following the introduction of the defensive symbiont (Fig. 5). First, we find that

while a defensive symbiont may initially increase the host population size, the host appears

eventually to always suffer a decrease in population size due to parasite-symbiont coevolution,

regardless of the initial strength of protection (indicated by the terminus of each evolutionary

trajectory residing in regions with a negative percentage increase in host population size). This

seems to occur for one or more of the following three reasons: (1) the symbiont may experience

selection against tolerance, resulting in a reduction or even loss of host protection; (2) the defensive

symbiont incurs a small cost to the host; and (3) while the defensive symbiont may confer tolerance

to some hosts, the parasite subsequently experiences selection for higher virulence, and so hosts

without the defensive symbiont experience higher virulence.

Although there is always eventually a net-negative effect on the host population size following

parasite-symbiont coevolution, the same is not necessarily true for realised virulence (i.e. the

average disease-associated mortality rate). In many cases an initially positive effect on average

realised virulence is followed by a long-term negative effect (as observed for the host population

size measure above), but when the costs of protection are sufficiently strong and accelerate, there is

a reduction in average realised virulence (Fig. 5b).

When the costs of protection accelerate, the parasite and symbiont coevolve to co-continuously

stable strategies (Fig. 5a and b), but when the costs of protection decelerate, the outcome may

depend on the initial conditions, with sufficiently low levels of initial protection leading to selection

against any protection and a minor increase in parasite virulence (Fig. 5c and d), and sufficiently high

levels of initial protection leading to selection for maximal protection and high virulence (Fig. 5c).

Somewhat paradoxically, this means that the introduction of a highly protective symbiont can lead

to a much larger negative effect on the host population than the introduction of a symbiont that

confers only a low level of protection.

DISCUSSION

Defensive symbionts are found throughout the natural world and are potentially important agents of

biocontrol, yet the robustness of host protection and their eco-evolutionary impacts on parasite

evolution are poorly understood. In this study, we have theoretically explored the (co)evolutionary

dynamics of parasite virulence and host protection by a defensive symbiont in the form of tolerance.

We have investigated the behaviour of both the parasite and the defensive symbiont, as well as the

net effect on the host population. We have shown that the parasite will always evolve to be more

virulent following the introduction of a tolerance-conferring defensive symbiont, and (for every

parameter combination tested) this always has a negative impact on the host population size even if

the defensive symbiont evolves to confer maximum host protection. Furthermore, our model reveals

that the defensive symbiont can cause diversification in the parasite population for sufficiently high

levels of host protection, leading to the coexistence of low and high virulence phenotypes. Overall,

our results suggest that the introduction of tolerance-conferring defensive symbionts is likely to lead

to higher evolved and realised virulence, resulting in a net negative impact on the host population.

Higher virulence always evolves in our model because the defensive symbiont confers protection to

the host by ameliorating the disease-associated mortality rate, which increases the average

infectious period in coinfected hosts – those harbouring the defensive symbiont and parasite

simultaneously. Although more virulent parasites experience a sub-optimal level of virulence in

hosts that do not harbour the defensive symbiont, this is more than offset by having a higher

transmission rate in coinfections. Thus, the prevalence of the defensive symbiont, and hence the

frequency of coinfections, plays a crucial role in determining the strength of selection for increased

virulence. The fact that tolerance-conferring symbionts always select for higher virulence mirrors the

literature on imperfect vaccination. Gandon et al. (2001) showed theoretically how partially effective

vaccines that prevent or reduce disease (i.e., confer tolerance) but do not prevent transmission

select for higher virulence, a prediction that has since been confirmed for Marek's disease in

chickens (Read et al. 2015). Imperfectly vaccinated individuals are analogous to hosts who harbour

the defensive symbiont in our model; in both cases, the host experiences lower virulence while still

being able to transmit the infectious agent, weakening the evolutionary trade-off between

transmission and virulence and shifting the balance of selection towards higher virulence. Note that

in our model there is no explicit reduction in the pathogen growth rate due to tolerance (although

this need not be the case in general) and there is no direct effect on the transmission rate. Instead,

the defensive symbiont confers protection by reducing the negative effects of infection (akin to the

"anti-toxin" resistance in Gandon et al. (2001)). While we are not aware of any experimental studies

that have explored the evolution of virulence in the presence of a tolerance-conferring symbiont,

the strong parallels with imperfect vaccination suggest that such symbionts should indeed select for

higher parasite virulence.

Although we found that the introduction of a tolerance-conferring defensive symbiont always

selects for higher virulence, this is not necessarily the case for all symbionts. For example, Nelson

and May (2017, 2020) have shown that when the shared costs of virulence are sufficiently low,

communities of symbionts remain mutualistic and evolve lower virulence, and that in certain cases

pathogens can evolve towards mutualism in these communities. There are several key differences

between our model and those studied by Nelson and May (2017, 2020). In particular, Nelson and

May consider shared (additive) costs of virulence, whereas in our model host protection directly (i.e.

multiplicatively) reduces the virulence of the parasite. Furthermore, the mutualistic effects of our

defensive symbiont are context-dependent, as the benefits of protection are only realised in

infected hosts, whereas the mutualistic effects observed by Nelson and May occur regardless of

whether other symbionts are present. Our model therefore highlights the importance of context-

dependent mutualisms and direct interactions between mutualists and parasites for the evolution of

virulence.

Even if evolved virulence is higher in the presence of the defensive symbiont, the realised virulence

experienced by hosts with the symbiont can be lower due to host protection. However, hosts

without the defensive symbiont will experience increased virulence, and so the frequency of

coinfections will determine the variance in the realised level of virulence experienced by parasitised

hosts. While the net effect of the defensive symbiont on the host population size might initially be

positive, we have shown that this is not evolutionary robust, either due to selection for higher

parasite virulence (even if selection also favours higher host protection by the defensive symbiont,

as in Fig. 5a), or due to selection against host protection (Fig. 5c-d). However, if the goal is to reduce

the average virulence experienced by infected hosts rather than to maximise host population size,

then it is possible to achieve modest gains in host survival provided the cost of conferring host

protection accelerates with greater host protection and the overall strength of costs are sufficiently

high (Fig. 5b).

Our results have critical implications for the use of defensive symbionts as biocontrol agents, with

tolerance-conferring symbionts likely to be a poor choice for long-term infectious disease control at

the population level. Moreover, our model demonstrates the need to investigate the possible

evolutionary dynamics of both defensive symbionts and parasites when considering the use of

biocontrols, as short-term ecological dynamics may be a poor predictor of long-term outcomes.

Counter-intuitively, our model reveals that under certain trade-offs (when costs of host protection

have increasing returns, i.e. are decelerating), the introduction of a more protective defensive

symbiont can lead to far worse outcomes for the host population in the long-term compared than if

a less protective symbiont is introduced (Fig. 5c). Decelerating trade-offs often produce evolutionary

repellers (e.g., Ashby and King 2017) as they impose a high initial cost when investment is low (here,

this selects against protection and leads to little change in virulence if the initial level of protection is

low), and a relatively low additional cost when investment is high (here, this selects for higher

protection, and in turn, higher virulence, when the initial level of protection is sufficiently high). Due

to the complex nature of eco-evolutionary feedbacks in these systems and the potential for

unexpected evolutionary trajectories, we therefore urge caution in the use of tolerance-conferring

symbionts.

Our final key result is that the defensive symbiont can drive parasite diversification into high and low

virulence phenotypes. This occurs because the defensive symbiont adds an additional feedback on

the parasite population, which allows the different phenotypes to specialise on hosts that either lack

or possess the defensive symbiont. However, we found that the level of tolerance conferred by the

symbiont must be very high for diversification to occur, which suggests that although this is

theoretically possible, it is unlikely to be common in real populations. Nevertheless, the fact that a

defensive symbiont can facilitate parasite diversification emphasises the importance of considering

community effects on host and parasite diversity, and this finding follows a general pattern in recent

theoretical studies where the addition of a third species induces diversification in the host or

parasite (Best 2018; Kisdi, Geritz, and Boldin 2013; Wood and Ashby 2023; Hoyle, Best, and Bowers

2012). For example, the addition of a predator that differentially feeds on infected hosts has been

shown to lead to diversification in host resistance (Hoyle, Best, and Bowers 2012) and parasite

virulence (Kisdi, Geritz, and Boldin 2013; Best 2018). Similarly, the introduction of a hyperparasite (a

parasite of the parasite) can also cause diversification in parasite virulence (Wood and Ashby 2023),

which potentially presents problems for biocontrol, as with tolerance-conferring symbionts.

To date, few studies have experimentally explored the evolution of parasite virulence in the

presence of defensive symbionts (King et al. 2016; Ford et al. 2017; G. May et al. 2022). May et al.

(2022) have shown that when a plant host (*Zea mays*) is infected by a pathogenic fungus (*Ustilago*

maydis), parasite fitness is maximised at higher levels of virulence in the presence of a defensive

symbiont (*Fusarium verticillioides*), in agreement with our model. However, Ford et al. (2016)

experimentally coevolved pathogenic *Staphylococcus aureus* and protective *Enterococcus faecalis* in

Caenorhabditis elegans hosts, which led to a reduction in pathogen virulence; the same study

system has also been shown to generate fluctuating selection dynamics in pathogen virulence and

host protection (Ford et al. 2017). The contrast with our results is because *E. faecalis* confers

protection through the production of antimicrobial superoxides which directly inhibit *S. aureus*

rather than conferring tolerance to the host. The stark contrast in evolutionary outcomes with our

model emphasises the importance of understanding the mechanism of host protection. Moreover,

most studies of defensive symbionts focus on those that confer protection in terms of a reduced

parasite load (e.g., due to interference competition) (Hoang and King 2022), and tolerance-

conferring symbionts are understudied. Indeed, we are aware of only one study that explicitly shows

defensive symbionts conferring tolerance to the host, with *Bacteroides fragilis* conferring tolerance

by inducing the production of anti-inflammatory proteins against an experimental colitis caused by

the bacterium *Heliobacter hepaticus* (Mazmanian, Round, and Kasper 2008).

Given that many defensive symbionts confer host protection through other mechanisms (including

through upregulation of host immune responses (Ford, Drew, and King 2022), the eco-evolutionary

implications of introducing different types of symbionts should be explored in future theoretical

studies. Furthermore, we have implicitly modelled how the defensive symbiont and parasite interact

at the within-host level, along with trade-offs between transmission and tolerance or virulence. This

simplification makes the model much more tractable, but an important direction for future research

is to explicitly model the within-host dynamics and couple these to between-host transmission.

Coupling within- and between-host modelling has been shown to provide new insights than

population-level modelling on its own cannot provide. Modelling at each level explicitly means that

we can investigate conflicting selection, i.e. where the most successful phenotype at one level is not

necessarily the most successful at the other (Frank 1996; van Baalen and Sabelis 1995; B. R. Levin

and Bull 1994). The overall evolutionary outcomes will heavily depend on this conflicting selection

that cannot be fully captured by simply modelling at the between-host level only. Another important

consequence of explicit within-host modelling is providing insights into trade-offs and recovery

rates. Several nested models have shown that any trade-offs depend heavily on the within-host

dynamics (Ganusov, Bergstrom, and Antia 2002; Gilchrist and Sasaki 2002; André, Ferdy, and Godelle

2003; Alizon and van Baalen 2005; Gilchrist and Coombs 2006), whilst recovery rates, which are

traditionally considered to be constant and independent of other parameters, have been shown to

be important for the evolution of virulence (Ganusov and Antia 2006; André and Gandon 2006). In

the context of defensive symbiont-parasite dynamics, explicit within-host modelling will allow for a

greater understanding of how a range of different mechanisms, such as interference competition,

resource competition, spite, priority effects and interactions with the host immune system impact

on the evolution of virulence and host protection.

Overall, our model reveals how tolerance-conferring defensive symbionts typically have a net

negative impact on the host population over the long-term as they always select for higher parasite

virulence and are therefore poor candidates for biocontrol.

ACKNOWLEDGEMENTS

BA is funded by Natural Environment Research Council grants NE/N014979/1 and NE/V003909/1.

CAS is funded by Natural Environment Research Council grant NE/V003909/1. We acknowledge the

support of the Natural Sciences and Engineering Research Council of Canada (NSERC). Nous

remercions le Conseil de recherches en sciences naturelles et en génie du Canada (CRSNG) de son

soutien. PIPPS receives funding from the BC Ministry of Health.

COMPETING INTERESTS

The authors declare no competing interests

DATA ACCESSIBILITY

The Python code for the implementation of this model can be found here:

https://github.com/CameronSmith50/Defensive-Symbiosis

REFERENCES

Alizon, Samuel. 2013. 'Co-Infection and Super-Infection Models in Evolutionary Epidemiology'.
Interface Focus 3 (6). https://doi.org/10.1098/rsfs.2013.0031.
Alizon, Samuel, and Minus van Baalen. 2005. 'Emergence of a Convex Trade-Off between
Transmission and Virulence'. *The American Naturalist* 165 (6): E155–67.
https://doi.org/10.1086/430053.
André, Jean-Baptiste, Jean-Baptiste Ferdy, and Bernard Godelle. 2003. 'Within-Host Parasite
Dynamics, Emerging Trade-off, and Evolution of Virulence with Immune System'. *Evolution*
57 (7): 1489–97. https://doi.org/10.1111/j.0014-3820.2003.tb00357.x.
André, Jean-Baptiste, and Sylvain Gandon. 2006. 'Vaccination, within-Host Dynamics and Virulence
Evolution'. *Evolution* 60 (1): 13–23. https://doi.org/10.1111/j.0014-3820.2006.tb01077.x.
Arnold, A. Elizabeth, Luis Carlos Mejía, Damond Kyllo, Enith I. Rojas, Zuleyka Maynard, Nancy
Robbins, and Edward Allen Herre. 2003. 'Fungal Endophytes Limit Pathogen Damage in a
Tropical Tree'. *Proceedings of the National Academy of Sciences of the United States of*
America 100 (26): 15649–54. https://doi.org/10.1073/pnas.2533483100.
Ashby, Ben, and Kayla C. King. 2017. 'Friendly Foes: The Evolution of Host Protection by a Parasite'.
Evolution Letters 1 (4): 211–21. https://doi.org/10.1002/evl3.19.
Baalen, Minus van, and Maurice W. Sabelis. 1995. 'The Dynamics of Multiple Infection and the
Evolution of Virulence'. *The American Naturalist* 146 (6): 881–910.
https://doi.org/10.1086/285830.
Bakken, Johan S., Thomas Borody, Lawrence J. Brandt, Joel V. Brill, Daniel C. Demarco, Marc Alaric
Franzos, Colleen Kelly, et al. 2011. 'Treating Clostridium Difficile Infection With Fecal

Microbiota Transplantation'. *Clinical Gastroenterology and Hepatology* 9 (12): 1044–49.
https://doi.org/10.1016/j.cgh.2011.08.014.
Belt, Thomas. 1874. *The Naturalist in Nicaragua*. University Press of the Pacific.
Best, Alex. 2018. 'Host–Pathogen Coevolution in the Presence of Predators: Fluctuating Selection
and Ecological Feedbacks'. *Proceedings of the Royal Society B: Biological Sciences* 285 (1885):
20180928. https://doi.org/10.1098/rspb.2018.0928.
Brown, Sam P., Michael E. Hochberg, and Bryan T. Grenfell. 2002. 'Does Multiple Infection Select for
Raised Virulence?' *Trends in Microbiology* 10 (9): 401–5. https://doi.org/10.1016/S0966-
842X(02)02413-7.
Cariveau, Daniel P., J. Elijah Powell, Hauke Koch, Rachael Winfree, and Nancy A. Moran. 2014.
'Variation in Gut Microbial Communities and Its Association with Pathogen Infection in Wild
Bumble Bees (Bombus)'. *The ISME Journal* 8 (12): 2369–79.
https://doi.org/10.1038/ismej.2014.68.
Chamberlain, Scott A., Judith L. Bronstein, and Jennifer A. Rudgers. 2014. 'How Context Dependent
Are Species Interactions?' Edited by Rampal Etienne. *Ecology Letters* 17 (7): 881–90.
https://doi.org/10.1111/ele.12279.
Diekmann, O., J. A. P. Heesterbeek, and M. G. Roberts. 2010. 'The Construction of Next-Generation
Matrices for Compartmental Epidemic Models'. *Journal of The Royal Society Interface* 7 (47).
https://doi.org/10.1098/rsif.2009.0386.
Ford, Suzanne A., Georgia C. Drew, and Kayla C. King. 2022. 'Immune-Mediated Competition Benefits
Protective Microbes over Pathogens in a Novel Host Species'. *Heredity*, November.
https://doi.org/10.1038/s41437-022-00569-3.
Ford, Suzanne A., Damian Kao, David Williams, and Kayla C. King. 2016. 'Microbe-Mediated Host
Defence Drives the Evolution of Reduced Pathogen Virulence'. *Nature Communications* 7 (1):
13430. https://doi.org/10.1038/ncomms13430.
Ford, Suzanne A., and Kayla C. King. 2016. 'Harnessing the Power of Defensive Microbes:
Evolutionary Implications in Nature and Disease Control'. Edited by Carolyn B Coyne. *PLOS
Pathogens* 12 (4): e1005465. https://doi.org/10.1371/journal.ppat.1005465.
Ford, Suzanne A., David Williams, Steve Paterson, and Kayla C. King. 2017. 'Co-Evolutionary
Dynamics between a Defensive Microbe and a Pathogen Driven by Fluctuating Selection'.
Molecular Ecology 26 (7): 1778–89. https://doi.org/10.1111/mec.13906.
Frank, Steven A. 1992. 'A Kin Selection Model for the Evolution of Virulence'. *Proceedings of the
Royal Society of London. Series B: Biological Sciences* 250 (1329): 195–97.
https://doi.org/10.1098/rspb.1992.0149.
———. 1996. 'Models of Parasite Virulence'. *The Quarterly Review of Biology* 71 (1): 37–78.
https://doi.org/10.1086/419267.
Gandon, Sylvain, Margaret J. Mackinnon, Sean Nee, and Andrew F. Read. 2001. 'Imperfect Vaccines
and the Evolution of Pathogen Virulence'. *Nature* 414 (6865): 751–56.
https://doi.org/10.1038/414751a.
Ganusov, Vitaly V., and Rustom Antia. 2006. 'Imperfect Vaccines and the Evolution of Pathogens
Causing Acute Infections in Vertibrates'. *Evolution* 60 (5): 957–69.
https://doi.org/10.1111/j.0014-3820.2006.tb01174.x.
Ganusov, Vitaly V., Carl T. Bergstrom, and Rustom Antia. 2002. 'Within-Host Population Dynamics
and the Evolution of Microparasites in a Heterogeneous Host Population'. *Evolution* 56 (2):
213–23. https://doi.org/10.1111/j.0014-3820.2002.tb01332.x.
Gilchrist, Michael A., and Daniel Coombs. 2006. 'Evolution of Virulence: Interdependence,
Constraints, and Selection Using Nested Models'. *Theoretical Population Biology* 69 (2).
https://doi.org/10.1016/j.tpb.2005.07.002.
Gilchrist, Michael A, and Akira Sasaki. 2002. 'Modeling Host-Parasite Coevolution: A Nested
Approach Based on Mechanistic Models'. *J. Theor. Biol* 218: 289–308.
https://doi.org/10.1006/yjtbi.3076.

Gil-Turnes, M. S., and W. Fenical. 1992. 'Embryos of *Homarus Americanus* Are Protected by Epibiotic
Bacteria'. *The Biological Bulletin* 182 (1): 105–8. https://doi.org/10.2307/1542184.
Gil-Turnes, M. S., M. E. Hay, and W. Fenical. 1989. 'Symbiotic Marine Bacteria Chemically Defend
Crustacean Embryos from a Pathogenic Fungus'. *Science (New York, N.Y.)* 246 (4926): 116–
18. https://doi.org/10.1126/science.2781297.
González, Rubén, Anamarija Butković, Francisco J. Escaray, Javier Martínez-Latorre, Ízan Melero,
Enric Pérez-Parets, Aurelio Gómez-Cadenas, Pedro Carrasco, and Santiago F. Elena. 2021.
'Plant Virus Evolution under Strong Drought Conditions Results in a Transition from
Parasitism to Mutualism'. *Proceedings of the National Academy of Sciences* 118 (6):
e2020990118. https://doi.org/10.1073/pnas.2020990118.
Hancock, Penelope A., Steven P. Sinkins, and H. Charles J. Godfray. 2011. 'Strategies for Introducing
Wolbachia to Reduce Transmission of Mosquito-Borne Diseases'. Edited by Scott Leslie
O'Neill. *PLoS Neglected Tropical Diseases* 5 (4): e1024.
https://doi.org/10.1371/journal.pntd.0001024.
Heikkilä, M. P., and P. E. J. Saris. 2003. 'Inhibition of Staphylococcus Aureus by the Commensal
Bacteria of Human Milk'. *Journal of Applied Microbiology* 95 (3): 471–78.
https://doi.org/10.1046/j.1365-2672.2003.02002.x.
Herre, Edward Allen, Luis C. Mejía, Damond A. Kyllo, Enith Rojas, Zuleyka Maynard, Andre Butler,
and Sunshine A. Van Bael. 2007. 'Ecological Implications of Anti-Pathogen Effects of Tropical
Fungal Endophytes and Mycorrhizae'. *Ecology* 88 (3): 550–58. https://doi.org/10.1890/05-
1606.
Hoang, Kim L., and Kayla C. King. 2022. 'Symbiont-Mediated Immune Priming in Animals through an
Evolutionary Lens'. *Microbiology* 168 (4). https://doi.org/10.1099/mic.0.001181.
Hoyle, Andrew, Alex Best, and Roger G. Bowers. 2012. 'Evolution of Host Resistance towards
Pathogen Exclusion: The Role of Predators'. *Evolutionary Ecology Research* 14 (2): 125–46.
Hoyle, Andrew, Roger G. Bowers, Andrew White, and Michael Boots. 2008. 'The Influence of Trade-
off Shape on Evolutionary Behaviour in Classical Ecological Scenarios'. *Journal of Theoretical
Biology* 250 (3): 498–511. https://doi.org/10.1016/j.jtbi.2007.10.009.
King, Kayla C, Michael A Brockhurst, Olga Vasieva, Steve Paterson, Alex Betts, Suzanne A Ford,
Crystal L Frost, Malcolm J Horsburgh, Sam Haldenby, and Gregory DD Hurst. 2016. 'Rapid
Evolution of Microbe-Mediated Protection against Pathogens in a Worm Host'. *The ISME
Journal* 10 (8): 1915–24. https://doi.org/10.1038/ismej.2015.259.
Kisdi, Eva, Stefan A.H. Geritz, and Barbara Boldin. 2013. 'Evolution of Pathogen Virulence under
Selective Predation: A Construction Method to Find Eco-Evolutionary Cycles'. *Journal of
Theoretical Biology* 339 (December): 140–50. https://doi.org/10.1016/j.jtbi.2013.05.023.
Lauer, Antje, and Trang Hernandez. 2015. 'Cutaneous Bacterial Species from Lithobates
Catesbeianus Can Inhibit Pathogenic Dermatophytes'. *Mycopathologia* 179 (3–4): 259–68.
https://doi.org/10.1007/s11046-014-9838-1.
Leimer, Olaf. 2009. 'Multidimensional Convergence Stability'. *Evolutionary Ecology Research*.
http://zootis.zoologi.su.se/research/leimar/fulltext/Leimar2009b.pdf.
Lenski, Richard E., and Robert M. May. 1994. 'The Evolution of Virulence in Parasites and Pathogens:
Reconciliation Between Two Competing Hypotheses'. *Journal of Theoretical Biology* 169 (3):
253–65. https://doi.org/10.1006/jtbi.1994.1146.
Levin, Bruce R., and James J. Bull. 1994. 'Short-Sighted Evolution and the Virulence of Pathogenic
Microorganisms'. *Trends in Microbiology* 2 (3): 76–81. https://doi.org/10.1016/0966-
842X(94)90538-X.
Levin, Simon, and David Pimentel. 1981. 'Selection of Intermediate Rates of Increase in Parasite-Host
Systems'. *The American Naturalist* 117 (3): 308–15. https://doi.org/10.1086/283708.
Lin, Derek, and Britt Koskella. 2015. 'Friend and Foe: Factors Influencing the Movement of the
Bacterium *Helicobacter Pylori* along the Parasitism-Mutualism Continuum'. *Evolutionary
Applications* 8 (1): 9–22. https://doi.org/10.1111/eva.12231.

May, Georgiana, Ruth G. Shaw, Charles J. Geyer, and Daniel J. Eck. 2022. 'Do Interactions among Microbial Symbionts Cause Selection for Greater Pathogen Virulence?' *The American Naturalist* 199 (2): 252–65. https://doi.org/10.1086/717679.

May, Robert, and Roy Anderson. 1983. 'Epidemiology and Genetics in the Coevolution of Parasites and Hosts'. *Proceedings of the Royal Society of London. Series B. Biological Sciences* 219 (1216): 281–313. https://doi.org/10.1098/rspb.1983.0075.

May, Robert, and Martin A. Nowak. 1995. 'Coinfection and the Evolution of Parasite Virulence'. *Proceedings of the Royal Society of London. Series B: Biological Sciences* 261 (1361): 209–15. https://doi.org/10.1098/rspb.1995.0138.

Mazmanian, Sarkis K., June L. Round, and Dennis L. Kasper. 2008. 'A Microbial Symbiosis Factor Prevents Intestinal Inflammatory Disease'. *Nature* 453 (7195): 620–25. https://doi.org/10.1038/nature07008.

Moreira, Luciano A., Iñaki Iturbe-Ormaetxe, Jason A. Jeffery, Guangjin Lu, Alyssa T. Pyke, Lauren M. Hedges, Bruno C. Rocha, et al. 2009. 'A Wolbachia Symbiont in Aedes Aegypti Limits Infection with Dengue, Chikungunya, and Plasmodium'. *Cell* 139 (7): 1268–78. https://doi.org/10.1016/j.cell.2009.11.042.

Nelson, Paul G., and Georgiana May. 2017. 'Coevolution between Mutualists and Parasites in Symbiotic Communities May Lead to the Evolution of Lower Virulence'. *The American Naturalist* 190 (6): 803–17. https://doi.org/10.1086/694334.

Nelson, Paul, and Georgiana May. 2020. 'Defensive Symbiosis and the Evolution of Virulence'. *The American Naturalist* 196 (3): 333–43. https://doi.org/10.1086/709962.

Oliver, Kerry M., Jacob A. Russell, Nancy A. Moran, and Martha S. Hunter. 2003. 'Facultative Bacterial Symbionts in Aphids Confer Resistance to Parasitic Wasps'. *Proceedings of the National Academy of Sciences of the United States of America* 100 (4): 1803–7. https://doi.org/10.1073/pnas.0335320100.

Palmer, Todd M., Maureen L. Stanton, Truman P. Young, Jacob R. Goheen, Robert M. Pringle, and Richard Karban. 2008. 'Breakdown of an Ant-Plant Mutualism Follows the Loss of Large Herbivores from an African Savanna'. *Science* 319 (5860): 192–95. https://doi.org/10.1126/science.1151579.

Rafaluk-Mohr, Charlotte, Ben Ashby, Dylan A. Dahan, and Kayla C. King. 2018. 'Mutual Fitness Benefits Arise during Coevolution in a Nematode-Defensive Microbe Model'. *Evolution Letters* 2 (3): 246–56. https://doi.org/10.1002/evl3.58.

Rafaluk-Mohr, Charlotte, Michael Gerth, Jordan E. Sealey, Alice K.E. Ekroth, Aziz A. Aboobaker, Anke Kloock, and Kayla C. King. 2022. 'Microbial Protection Favors Parasite Tolerance and Alters Host-Parasite Coevolutionary Dynamics'. *Current Biology* 32 (7): 1593-1598.e3. https://doi.org/10.1016/j.cub.2022.01.063.

Read, Andrew F., Susan J. Baigent, Claire Powers, Lydia B. Kgosana, Luke Blackwell, Lorraine P. Smith, David A. Kennedy, Stephen W. Walkden-Brown, and Venugopal K. Nair. 2015. 'Imperfect Vaccination Can Enhance the Transmission of Highly Virulent Pathogens'. Edited by Christophe Fraser. *PLOS Biology* 13 (7): e1002198. https://doi.org/10.1371/journal.pbio.1002198.

Rogalski, Mary A., Tara Stewart Merrill, Camden D. Gowler, Carla E. Cáceres, and Meghan A. Duffy. 2021. 'Context-Dependent Host-Symbiont Interactions: Shifts along the Parasitism-Mutualism Continuum'. *The American Naturalist* 198 (5): 563–75. https://doi.org/10.1086/716635.

Selva, Laura, David Viana, Gili Regev-Yochay, Krzysztof Trzcinski, Juan Manuel Corpa, íñigo Lasa, Richard P. Novick, and José R. Penadés. 2009. 'Killing Niche Competitors by Remote-Control Bacteriophage Induction'. *Proceedings of the National Academy of Sciences* 106 (4): 1234–38. https://doi.org/10.1073/pnas.0809600106.

Troha, Katia, and Janelle S Ayres. 2022. 'Cooperative Defenses during Enteropathogenic Infection'.
Current Opinion in Microbiology 65 (February): 123–30.
https://doi.org/10.1016/j.mib.2021.11.003.
Utarini, Adi, Citra Indriani, Riris A. Ahmad, Warsito Tantowijoyo, Eggi Arguni, M. Ridwan Ansari,
Endah Supriyati, et al. 2021. 'Efficacy of Wolbachia-Infected Mosquito Deployments for the
Control of Dengue'. *New England Journal of Medicine* 384 (23): 2177–86.
https://doi.org/10.1056/NEJMoa2030243.
Wood, Jason, and Ben Ashby. 2023. 'Hyperparasitism and the Evolution of Parasite Virulence'.
Preprint. Evolutionary Biology. https://doi.org/10.1101/2023.01.18.524570.
659

Parameter	Description	Default value
a	Maximum per-capita host birth rate	1.0
b	Host natural mortality rate	0.25
c_1	Defensive symbiont cost strength parameter	0.25
c_2	Defensive symbiont cost shape parameter	2
d	Power-law for parasite virulence cost	2
q	Strength of density-dependent competition on host reproduction	0.25
T_{eco}	Duration for ecological time steps	100
T_{evo}	Duration for evolutionary simulations	2000
α_D	Cost of harbouring the defensive symbiont	0.1
$\hat{\alpha}_P$	Virulence of a parasite that cannot transmit	0.1
$\hat{\beta}_D$	Transmission rate of a defensive symbiont with no cost of protection	2
γ_D	Host recovery rate for defensive symbiont	0.05
γ_P	Host recovery rate for parasite	0.05
ε	Extinction threshold	10^{-4}

662   *Table 1: Default parameter values for the model (1)-(4).*

664
665
Fig. 1: Model schematic. Arrows denote transitions into or out of states at the indicated rates:
transmission (green), mortality (red), recovery/clearance (yellow) and birth of new hosts (blue).

Fig. 2: Evolution of parasite virulence following the introduction of the defensive symbiont. (a)

Evolved virulence relative to the initial stable level of virulence in the absence of the defensive

symbiont. (b) The percentage increase in average realised virulence compared with the absence of

the defensive symbiont. (c) The proportion of parasitised hosts that possess defensive symbionts. The

black dashed line corresponds to relatively low costs to the defensive symbiont of conferring

protection   , the red solid line to moderate costs   and the blue dotted line to

relatively high costs   . Values of strength of cost have been chosen to represent strengths of

cost across the entire range   . All other parameters are as in Table 1.

Fig. 3: Parasite diversification driven by a defensive symbiont. Row a: evolutionary

trajectories of parasite transmission with inset pairwise invasion plots (PIPs). Black

regions in the PIPs show where the mutant can invade (where $w_P\left(\beta_P^m \middle| (1,\ \beta_P^r)\right) > 1$), and

white regions are where it cannot. Row b: the proportion of parasitised hosts which

also possess the defensive symbiont. For bii, the red line (right branch) corresponds to

the high virulence strain, and the black line (left branch) corresponds to the low

virulence strain. Costs of host protection: (column i) $c_1 = 0.4$, (column ii) $c_1 = 0.5$ and

(column iii) $c_1 = 0.6$. These values of cost strength represent the narrow range of

values with a qualitative change in evolutionary behaviour. All other parameters as in

Table 1.

Fig. 4: Classification of the coevolutionary outcome for the defensive symbiont as a function of the two cost function parameters; c_1, the strength of cost ranging from 0 (no cost) to 1 (maximal cost), and c_2 the shape of the trade-off with transmission: accelerating $(c_2 > 0)$, linear $(c_2 = 0)$, decelerating $(c_2 < 0)$. The repeller region results in the defensive symbiont either maximising or minimising host protection depending on the initial level of protection. The CSS region corresponds to a continuously stable strategy at an intermediate level of protection. The parasite is also evolving, but its evolutionary outcome is not shown as it always tends to a CSS. Black crosses correspond to the 4 parameter pairs used in Fig. 5 to demonstrate the different qualitative behaviours. All other parameters as in Table 1.

Fig. 5: Heatmaps for the changes in population size and death rate (as given in

equations (11)-(12)) for various mutualist cost functions. We show moderate cost

$(c_1 = 0.4)$ in the left column and strong cost $(c_1 = 0.9)$ in the right, with accelerating

cost $(c_2 = 2)$ and decelerating cost $(c_2 = -2)$ in the top and bottom rows respectively.

Colours and values on the contour plots denote percentage changes for a given trait

space pair (y, β_P). Green dots are the initial value, solid black lines denote an

evolutionary trajectory in trait space, and the red dots are the ends. The black dashed

line is the CSS value for the parasite transmission when it is the only microbe in

circulation. The values of c_1 and c_2 have been chosen to represent the four qualitative

behaviours for the defensive symbiont that we have seen in Fig. 4.