Signal detectability and boldness are not the same: the function of defensive colouration in
nudibranchs is distance-dependent

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Abstract

Aposematic signals advertise underlying defences in many species. They should be detectable

(highly contrasting against the background) and bold (high internal pattern contrast) to enhance

predator recognition, learning and memorisation. However, the signalling function of aposematic

colour patterns may be distance-dependent: signals may be undetectable from a distance to reduce

costs of increased attacks from naïve predators but bold when viewed up close. To test this hypothesis,

we quantified the chromatic and achromatic detectability and boldness of colour patterns in 13

nudibranch species that varied in the strength of their chemical defences, in terms of unpalatability

and toxicity, using Quantitative Colour Pattern Analysis (QCPA) and data on the visual perception of a

triggerfish (*Rhinecanthus aculeatus*). When viewed from larger distances, there were no differences

in detectability and boldness between well-defended and undefended species. However, when

viewed at close distances, well-defended species were more detectable and bolder than undefended

species. The detectability of defended species decreased more significantly with increased viewing

distances compared to boldness but remained relatively consistent over viewing distances for

undefended species. We provide evidence for distance-dependent signalling in aposematic

nudibranchs and highlight the importance of distinguishing between signal detectability and boldness

in studies of aposematism.

Keywords

Aposematism, crypsis, visual modelling, secondary defences, predator psychology, signalling
honesty.

Introduction

Conspicuous colour patterns displayed by aposematic species are used to educate predators

about underlying defences during prey encounters [1,2], or via eavesdropping (e.g. Hämäläinen et al.,

2021). Aposematic colour patterns should be readily detectable against their visual background, and

detectability is considered crucial to the initial evolution of aposematism [4,5]. Most aposematic

signals are also bold, defined as exhibiting high internal colour and luminance contrast [6,7], and

striking colour pattern geometry [1,2,8,9], which is thought to enhance the formation and

maintenance of predator avoidance behaviour [10]. Theoretically, aposematic colour patterns should

be most efficient when they are both easy to detect and are bold [4].

However, it is important to differentiate between detectability and boldness to understand

the appearance of a visual signal in the context of its background and the pattern itself. Evidence for

or against the relative contribution of each to signal quality and efficacy is mixed [11,12]. For example,

Sillén-Tullberg [13] demonstrated that chicks (*Gallus gallus domesticus*) learned to avoid coloured

artificial prey independent of background contrast. In contrast, Aronsson and Gamberale-Stille [14]

failed to find an effect of internal patterning on the strength of avoidance behaviour in chicks. A key

benefit of increased detectability in aposematic species may be to reduce predator recognition errors

by allowing more time for accurate decision-making [15]. However, Gamberale-Stille *et al.* [16]

demonstrated that increased detectability in chickens (*Gallus gallus domesticus*) did not improve the

survival of aposematic prey. In fact, if detectable aposematic species are attacked more frequently by

naive predators, or by those that are capable and willing to tolerate secondary defences [e.g. 17,18],

detectability may be a potential handicap of bold aposematic signals, rather than a fitness benefit

[4,19–23].

To adapt to optimal levels of detectability and signal boldness, aposematic species may

benefit from distance-dependent signalling [21,24], which could reduce the detectability of bold

colour patterns from a distance while enabling predator deterrence up close. This principle is now

supported by multiple theoretical and empirical studies [e.g. 25–31]. For example, Barnett and Cuthill

[22] demonstrated that, when searched for by human observers, camouflaged stimuli with boldly

contrasting markings were detected at the same distance compared to camouflaged stimuli without

bold markings. This was later confirmed by using artificial stimuli and avian predators [32–34].

The apparent paradox of distance-dependent signalling can be investigated by considering the

visual perception of a signal in its ecological context, including the spatial acuity of a visual system and

the distance at which it is observed [21,35,36]. In the predator sequence, prey detection precedes

prey identification and subsequent decisions by predators to further engage with prey [37,38].

Therefore, context-specific signal processing is used by predators as they proceed from detection to

discrimination and subsequent attack [12,39,40]. It is likely that the relative importance of primary

and secondary defences shifts along such a predation sequence [see 41 for review]. For example,

visual defences relying on the avoidance of detection precede bold deimatic displays by threatened

prey warning a predator of underlying secondary defences [see 41,42 for reviews]. However, it is

unknown how detectability and/or boldness change along an escalating predations sequence in the

context of permanently displayed warning signals.

Few studies have differentiated between the detectability and boldness of aposematic

animals [but see 43]. Fewer studies have done so at different viewing distances while considering the

physiological limitations of ecologically relevant observers [but see 26]. This is partly due to the

challenge of capturing spatiochromatic properties of complex visual backgrounds according to

observer-specific physiological limitations, such as chromatic and achromatic contrast perception and

spatial acuity.

In this study, we investigated the detectability and boldness of defensive animal colouration

in 13 species of nudibranch molluscs with differing chemical defences (Fig. 1). Nudibranch molluscs

display a stunning diversity of defensive colouration and secondary defences, and are valuable in the

study of the ecology and evolution of aposematism [43–46]. We used Quantitative Colour Pattern

Analysis (QCPA) [47] and considered the distance at which nudibranchs would be perceived by an

ecologically relevant observer, a triggerfish (*Rhinecanthus aculeatus*). We hypothesized that when

viewed from larger distances, the detectability of nudibranchs against their backgrounds should be

low for all species, irrespective of their bold colour patterns or strength of chemical defences.

However, when viewed up close, we hypothesized that species with strong chemical defences would

show significantly stronger visual contrasts against their backgrounds than those of undefended

species. Lastly, we expected an unequal change in the relationship of boldness and detectability

between chemically defended and undefended species across different viewing distances.

2. Material and Methods

(a) Image collection

We took calibrated digital images of 13 Dorid nudibranch species (n = 226 individuals):

Aphelodoris varia (n = 24), *Chromodoris elisabethina* (n = 21), *Chromodoris kuiteri* (n = 17), *Discodoris*

sp. (n = 15), *Doriprismatica atromarginata* (n = 27), *Glossodoris vespa* (n = 15), *Goniobranchus*

collingwoodi (n = 15), *Goniobranchus splendidus* (n = 25), *Hypselodoris bennetti* (n = 10)*, Phyllidia*

elegans (n = 8)*, Phyllidia ocellata* (n = 23), *Phyllidia varicosa* (n = 8) and *Phyllidiella pustulosa* (n = 18).

Species were visually identified using various taxonomic books [48–50]. We grouped individuals that

visually resembled *Sebadoris fragilis* and were found in the same locations as *Discodoris* sp.; however,

these individuals could be a mixture of *Sebadoris fragilis, Thayuva lilacina, Jorunna pantheris* and

perhaps other undescribed species that cannot be identified without molecular sequencing.

Fieldwork was conducted on SCUBA between October 2017 and July 2021 at dive sites along

the east coast of Australia: Sunshine Coast (SE Queensland), Gold Coast (SE Queensland) and Nelson

Bay (New South Wales). Individual nudibranchs were located and photographed underwater at depths

of 2-18m against their natural habitat using a calibrated digital Olympus EPL-5 with a 60mm macro

lens in an Olympus PT-EP10 underwater housing. We used white LED illumination from a combination

of VK6r and PV62 Scubalamp video lights for illumination [47]. All images were taken at roughly a 90°

angle (top-down) relative to the animal and its background with the animals in a naturally stretched-

out, straight, forward-moving position (Fig. 1).

Figure 1: Representative images of species used in this study grouped by their chemical defences
scaled approximately according to size: A) *Discodoris sp.; B) Aphelodoris varia; C) Phyllidiella pustulosa;*
D) Phyllidia elegans; E) Goniobranchus splendidus; F) Phyllidia ocellata; G) Goniobranchus
collingwoodi; H) Glossodoris vespa; I) Chromodoris kuiteri; J) Doriprismatica atromarginata; K)
Phyllidia varicosa; L) Chromodoris elisabethina; M) Hypselodoris bennetti. The level of chemical
defence is provided as values below each image determined from toxicity and unpalatability assays in
Winters et al. 2021. The first value indicates class of chemical defence: 'NR' indicates nudibranch
species that have limited or no chemical defences; class 'II' indicate species that are highly unpalatable
but weakly toxic and class 'I & II' indicate species are highly unpalatable and highly toxic. The second
value is data from unpalatability feeding assays with Palaemon shrimp (presented here as 1-ED_{50}
values). ED_{50} refers to the median effective dose.

(b) Aposematism and chemical defences

With the exception of *Discodoris sp.* and *Aphelodoris varia*, we consider these species to be

aposematic as they harbour potent chemical defences and display bold colour signals [46,51] Fig. 1).

Aphelodoris varia has no known chemical defences [46] and likely relies on crypsis via background

matching for their primary defence. *Sebadoris fragilis* does not have any known chemical defences

[46] and we, therefore, treat *Discodoris sp.* as non-toxic and palatable.

We quantified chemical defences in two ways in our study. First, we used the classification

from [46] to delineate between species belonging to the following three classes: NR - nudibranch

species that have limited or no chemical defences, class 'II' – species that are highly unpalatable but

only weakly toxic, class 'I & II' – highly unpalatable and highly toxic. *Glossodoris vespa* was classified

as class II from assay data reported in [51]. Second, we ranked the species from 0 to 1, according to

the average ED_{50} response in *Palaemon* shrimp feeding assays reported in [46,51], calculated as 1-ED_{50}

so that 1 represented the most unpalatable species (Fig 1).

(b) Quantification of detectability and boldness

Image analysis was conducted with visual modelling parameter choices as per [52] at the

following viewing distances: 2cm, 5cm, 10cm and 30cm. At distances beyond 30cm, the majority of

small nudibranchs and much of the internal patterning in larger individuals are unlikely to be visible

to a triggerfish (*R. aculeatus*) due to its spatial acuity of about 3 cycles per degree (cpd) [53].

To quantify detectability (i.e. a measure of background matching), we calculated the absolute

difference of the abundance weighted coefficient of variation of achromatic (Lum.CoV) and chromatic

(Col.CoV) local edge contrast (LEIA, measured in ΔS) for each animal relative to that of its respective

visual background. Thus, by considering the difference in the overall appearance of an animal by itself

relative to its immediate background, we specifically quantify background matching. The coefficient

of variation of chromatic and achromatic boundary contrast between colour pattern elements has

previously been shown to indicate mate choices in guppies (*Poecilia reticulata*) [54,55]. Instead of

using the boundary strength analysis [BSA, 56], we make use of the non-parametric approach of LEIA

and its ability to consider colour pattern contrast at roughly the scale of an edge-detecting receptive

field of a triggerfish [57]. Unlike BSA, LEIA does not rely on image segmentation. After applying

Gaussian acuity modelling, we use receptor noise limited (RNL) ranked filtering [47] as per [52].

The RNL model [58] allows us to estimate the perceived colour and luminance contrast of non-

human observers and, in addition to RNL ranked filtering, is used to describe the contrast of edges

within each animal and its corresponding natural background. This contrast measure is expressed as

ΔS and reflects the distance between two contrasting points in the RNL colour space [59]. To describe

the colour and luminance contrast at each location in the image, we used the average LEIA contrast

across the vertical, horizontal and diagonal filter axis. For a detailed description of LEIA parameters,

see [47].

To quantify boldness, we used Lum.Cov and Col.CoV of each animal without considering their

respective visual backgrounds, as we define the boldness of an aposematic signal as the strength of

the signal once prey detection has taken place. High values for each of these pattern statistics indicate

highly contrasting colour pattern elements.

(c) Statistical analysis

All colour pattern statistics were normalised to range from 0-1 using the 'range' argument of

the PreProcess function (caret package, [60], v6.0-90) using R Software ([61], v4.1.2), with infinite and

NA values treated as 0.

When we considered differences between species without considering the class of their

chemical defences, detectability and boldness data did not meet the assumptions of normality due to

large variation in the number of individuals and individual trait data, so we used a Kruskal-Wallis test

[62]. This analysis was performed using the *stats* package and post hoc analyses comparing individuals

was done using Dunn tests [63] with a Bonferroni correction [64] in the *FSA* package ([65],v0.9.3).

When considering differences between species in relation to their class of chemical defence

at different viewing distances, we applied a linear mixed effect (lme) model using the lme4 [66], v1.1-

28) and lmertest [67], v3.1-3) packages and type III ANOVA tables using Saterwaite's method in the

stats package after applying a square-root transform to the left-skewed data. The meeting of

assumptions was assessed for each model fit. We further ran independent lme models for chemical

defence and viewing distance. Species was treated as a random effect for all lme models.

To determine the relationship between unpalatability data and colour pattern statistics, we

used a Pearson product-moment correlation (R) in the stats package [61], v4.1.2). Detectability and

boldness were measured as the species median value of Lum.Cov and Col.Cov at each viewing

distance. For this analysis, we applied an ordered quantile normalisation to the raw data using the

bestNormalize package [68], v.1.8.2) to ensure normality throughout the dataset. The distance-

dependent change in the correlation between unpalatability and visual defences was then assessed

by fitting linear regressions to the obtained Pearson product-moment correlations.

3. Results

(a) Detectability

We first considered differences in detectability between species (i.e. the difference between

each animal and its background) at different distances without considering chemical defences. We

found significant differences in achromatic detectability (Lum.CoV) between species at all viewing

distances (Kruskal-Wallis 2cm: $\chi^2_{(12)}$ = 126.35, p < 0.001; 5cm: $\chi^2_{(12)}$ = 111.44, p < 0.001; 10cm: $\chi^2_{(12)}$ =

75.389, p < 0.001; 30cm: $\chi^2_{(12)}$ = 24.94, p = 0.015). However, only one species remained significantly

more detectable than others at 30cm, with *H. bennetti* being more detectable than *G. vespa* (Dunn-
test: p_{adj} = 0.021, Table S1). Similarly, there were significant differences in chromatic detectability
between species at close viewing distances (Kruskal-Wallis 2cm: $\chi^2_{(12)}$ = 95.23, *p* < 0.001; 5cm: $\chi^2_{(12)}$ =
97.69, *p* < 0.001; 10cm: $\chi^2_{(12)}$ = 76.81, *p* < 0.001) but not at 30cm (Kruskal-Wallis: $\chi^2_{(12)}$ = 16.62, *p* =
0.164, Table S2)

When considering the class of chemical defence, species without chemical defences (NR) had
higher achromatic detectability (Lum.Cov) at larger viewing distances ($F_{4,189}$ = 6.27, *p* < 0.001). In
contrast, chemically defended species were less detectable when viewed from a distance, with the
most defended species (class I and II) displaying the largest differences between viewing distances
(weakly toxic / highly unpalatable (class II): $F_{4,510}$ = 12.31, *p* < 0.001; highly toxic / highly unpalatable
(class I & II): $F_{4,406}$ = 31.38, *p* < 0.001) (Fig. 2a).

For species without chemical defences (NR), there was no difference in chromatic
detectability (Col.Cov) between viewing distances ($F_{4,190}$ = 0.50, *p* = 0.734). In contrast, species with
chemical defences (classes II / I & II) showed a strong reduction in chromatic detectability with
increasing viewing distances (weakly toxic / highly unpalatable (class II): $F_{4,510}$ = 12.24, *p* < 0.001; highly
toxic / highly unpalatable (class I & II): $F_{4,509.98}$ = 16.13, *p* < 0.001). Again, this relationship was stronger
for species with both toxic and unpalatable chemical defences (class I & II) (Fig. 2b).

There was no significant differences in either achromatic (Lum.CoV - 5cm: $F_{2,9.94}$ = 2.07, *p* =
0.178, 10cm: $F_{2,10.17}$ = 0.27, *p* = 0.771, 30cm: $F_{2,8.58}$ = 0.07, *p* = 0.937) or chromatic detectability (Col.CoV
- 2cm: $F_{2,9.82}$ = 1.09, *p* = 0.374, 5cm: $F_{2,9.88}$ = 1.03, *p* = 0.391, 10cm: $F_{2,9.95}$ = 0.75, *p* = 0.498, 30cm: $F_{2,8.68}$
= 0.22, *p* = 0.807) between chemical defence groups at either viewing distance. However, at 2cm
achromatic detectability species with both toxic and unpalatable chemical defences (class I & II) were
more detectable than either species without chemical defences (class NR) or unpalatable ones (class
II) ($F_{2,9.8}$ = 4.26, *p* = 0.047).

Unpalatability data was weakly positively correlated with both achromatic and chromatic
detectability at close viewing distances, whereas no or weakly negative correlations were observed at
larger viewing distances (Fig. 3a). Both, achromatic (r^2 = 0.98, $F_{1,2}$ = 117.7, *p* = 0.008) and chromatic (r^2 =
0.98, $F_{1,2}$ = 110, *p* = 0.009) detectability showed a significant reduction in their correlation with
unpalatability over increasing viewing distances (Fig. 3b).

(b) Boldness

When we considered differences in boldness (i.e. analysing each animal without its
background) without considering chemical defences, we found significant differences in achromatic

boldness between species at all viewing distances (Kruskal-Wallis 2cm: $\chi^2_{(12)}$ = 159.94, p < 0.001; 5cm:

$\chi^2_{(12)}$ = 161.11, p < 0.001; 10cm: $\chi^2_{(12)}$ = 148.26, p < 0.001; 30cm: $\chi^2_{(12)}$ = 74.27, p < 0.001). At 30cm,

significant differences between multiple species remained (Fig. 2c, Table S3). Similarly, there were

significant differences in chromatic boldness between species across all viewing distances (Kruskal-

Wallis 2cm: $\chi^2_{(12)}$ = 165.46, p < 0.001; 5cm: $\chi^2_{(12)}$ = 163.55, p < 0.001; 10cm: $\chi^2_{(12)}$ = 147.8, p < 0.001,

30cm: $\chi^2_{(12)}$ = 88.89, p < 0.001) (Table S4).

When considering the class of chemical defence, species with and without chemical defences

showed decreased achromatic boldness at larger viewing distances, with the most defended species

displaying the largest differences between distances (Undefended (class NR): $F_{4,189}$ = 38.34, p < 0.001;

weakly toxic / highly unpalatable (class II): $F_{4,510.05}$ = 244.74, p < 0.001; highly toxic / highly unpalatable

(class I & II): $F_{4,406}$ = 324.83, p < 0.001) (Fig. 2).

Similarly, species with and without chemical defences showed decreased chromatic boldness

at larger viewing distances with the most defended species displaying the largest differences between

distances (Undefended (NR): $F_{4,189}$ = 6.27, p < 0.001; Weakly toxic / highly unpalatable (II): $F_{4,510}$ = 12.31,

p < 0.001; Highly toxic / highly unpalatable (I & II): $F_{4,406}$ = 31.38, p < 0.001) (Fig. 2d).

There were no significant differences in either achromatic (Lum.CoV - 2cm: $F_{2,9.95}$ = 2.43, p =

0.139, 5cm: $F_{2,9.92}$ = 1.33, p = 0.309, 10cm: $F_{2,9.87}$ = 0.34, p = 0.721, 30cm: $F_{2,9.48}$ = 0.19, p = 0.828) or

chromatic boldness (Col.CoV - 2cm: $F_{2,9.95}$ = 1.08, p = 0.375, 5cm: $F_{2,9.97}$ = 1.28, p = 0.320, 10cm: $F_{2,9.80}$

= 0.70, p = 0.519, 30cm: $F_{2,9.77}$ = 0.05, p = 0.947) between chemical defence groups at either viewing

distance.

When considering chemical defences on a continuous scale using unpalatability data,

achromatic boldness showed a moderate correlation with unpalatability at close viewing distances

whereas chromatic boldness showed a weak correlation at close viewing distances (Fig. 3a). However,

neither achromatic (r^2 = 0.84, $F_{1,2}$ = 10.38, p = 0.084) nor chromatic (r^2 = 0.19, $F_{1,2}$ = 0.48, p = 0.560)

detectability showed a significant reduction in their correlation with unpalatability over increasing

viewing distances (Fig. 3b).

Figure 2 Relative detectability and boldness of 13 Dorid nudibranch species. The strength of chemical defences is indicated by colours relative to the class of chemical defence. A) Species median detectability quantified as the abundance weighted coefficient of variation of achromatic local edge contrast (Lum.CoV) measured relative to the natural background of every individual. B) Species median detectability quantified as the abundance weighted coefficient of variation of chromatic local edge contrast (Col.CoV) measured relative to the natural background of every individual. C) Species median achromatic pattern boldness measured as the abundance weighted coefficient of variation of achromatic local edge contrast (Lum.CoV) of each animal. D) Species median chromatic pattern boldness measured as the as the abundance weighted coefficient of variation of chromatic local edge contrast (Col.CoV) of each animal.

Figure 3 Summary of the correlation of the median chromatic (Col.CoV) and achromatic (Lum.CoV) boldness and detectability
of each species with the strength of chemical defences (1-ED$_{50}$) across different viewing distances. A) Individual Pearson
product-moment correlations (R) for each viewing distance and colour pattern statistic. Grey shading shows 95% confidence

intervals; B) Linear regressions fitted to the Pearson product-moment correlations (R) show variable distance-dependent
decreases in the correlation between colour pattern statistics.

4. Discussion

Our study demonstrates that the appearance of nudibranch colouration in the context of

natural backgrounds (detectability) or by itself (boldness) depends on the distance at which it is

viewed by an animal observer. For highly defended nudibranchs, both achromatic and chromatic

detectability and boldness significantly reduced with increasing viewing distance. In contrast, the

detectability and boldness of undefended species remained relatively consistent over viewing

distances. Our results support predictions [21,24] and empirical findings [e.g. 22] suggesting that

colour patterns with bold markings can be as camouflaged at a distance as colour patterns that are

camouflaged using background matching.

While chromatic and achromatic boldness decreased equally with increasing viewing

distances, various significant differences in achromatic and chromatic boldness remained between

species when viewed at 30 cm (Fig. 2). This could indicate that some species remain distinct in

appearance at viewing distances where the likelihood of detection is already significantly reduced.

Therefore, some well-defended species might still profit from a bold appearance in the unlikely event

of being detected at larger distances. These findings align with the suggestion from Barnett *et al.* [34]

that detectability and signalling function of defensive colouration can be differentially influenced by

viewing distance and that a reduction in detectability does not necessarily mean an equal reduction

in signal quality. Our study is the first to support this hypothesis using animals observed in their natural

habitat.

The loss of perceived higher spatial frequencies with increasing viewing distance affects both

apparent features and detail in visual backgrounds, as well as within an animal. As viewing distances

increase, smaller-scale colour pattern elements are expected to blur together, leading to the additive

blending of colour pattern elements [69] and subsequent change in the visual appearance of colour

patterns and visual backgrounds. How, and if, such blending happens, is influenced by the size, shape

and colour of individual colour pattern elements. However, in many animals chromatic detail is

perceived at lower spatial acuity than achromatic contrast [e.g. 70]. Thus, how chromatic and

achromatic contrast is arranged in colour patterns can allow for independently variable detectability

or boldness to a given predator at a given distance. Furthermore, object detection and discrimination

rely on vastly different processing pathways with object detection not necessarily requiring object

recognition [71]. As a result, individual or multiple colour pattern design elements can be subject to

individual or multiple selective pressures [39,72]. Therefore, despite being influenced by the loss of

spatial detail as viewing distances increase, colour pattern boldness and detectability in aposematic

animals may not be subject to equal selective pressures at varying viewing distances [e.g. 34].

311   Regulating the relative importance of detectability and boldness has been proposed to be

crucial in aposematic species to balance the costs and benefits of increased detectability induced by

bold colouration [34]. Therefore, detectability and boldness are thought to coincide with aposematism

at close viewing distances whereas aposematic and cryptic species are likely to profit from camouflage

at a distance [21,24]. Our findings support this assumption, as animals with both toxic and unpalatable

chemical defences were more detectable at close viewing distances than animals lacking chemical

defences or those with only unpalatable chemical defences (Fig. 3). We also did not find significant

differences in detectability between species with chemical defences and no chemical defences at

larger viewing distances (Fig. 3). However, while there were limited differences in detectability

between species at 30cm viewing distance, substantial variation in terms of chromatic and achromatic

boldness remained between species (Fig. 2, Table S3 & S4). Our findings suggest that the boldness and

detectability of colour patterns in aposematic nudibranch molluscs show distinct distance-dependent

correlations with increasing viewing distances. In line with predictions from Barnett et al. (2017), our

results show that detectability in chemically defended nudibranchs decreases faster than boldness as

viewing distances increase (Fig. 3). Specifically, both chromatic and achromatic detectability show a

significant decrease in their correlation with unpalatability, whereas boldness does not. This highlights

the cumulative importance of distinguishing between both, signal detectability and boldness as well

as ecologically relevant viewing distances when investigating signalling honesty in aposematic species.

However, it is important to note that the relative importance of detectability and colour pattern

boldness is context specific, as a bold animal is not necessarily easy to detect in a visual background

containing similar levels of colour pattern contrast [21,73,74], such is the case, for example, if the

animal's colour pattern represents a random sample of its complex and highly contrasty visual

background [21].

334   Interestingly, the intersection of the distance-dependent decreases of the correlation

between detectability and boldness is situated at around 10cm (Fig. 3). At this distance, many of the

more intricate colour pattern details are no longer visible to triggerfish. This likely hinders the

identification of and discrimination between species by potential predators, further emphasising the

importance of distinguishing between cognitive processes involved in prey detection and

identification. Such distance-dependent effects on the perception of colour patterns could be crucial

in the ecology and evolution of mimicry [75]. For example, the bold, outline-enhancing yellow rims

displayed by some of the species in this study are likely to be more detectable at a distance than the

finer internal patterning. This could explain the presence and constancy of such outline enhancing
yellow rims [9] among many aposematic nudibranch species and their putative mimics [72].

We show that the distance-dependent correlation between boldness and detectability
depends on the definition and strength of both underlying chemical and visual defences (Fig. 2 & 3).
For example, when correlating unpalatability on a continuous scale with each species' median colour
pattern statistic, we found that the correlation between detectability and the strength of chemical
defences decreases significantly with increasing viewing distances, whereas this does not happen for
boldness (Fig. 3). Instead, the correlation between boldness and unpalatability stays constant as
viewing distance increases. However, when looking at chemical defences defined as categories
considering both unpalatability and toxicity, we find significant decreases in boldness for chemically
defended species (Fig. 2). Furthermore, the effect size for the decrease of achromatic boldness is much
larger than that for the decrease of chromatic boldness. Therefore, in addition to highlighting the
importance of considering achromatic and chromatic aspects of aposematic signals separately, our
results highlight the importance of considering the type of analysis applied to such data.

As with many aposematic animals [see 76 for review], colour patterns within species of Dorid
nudibranchs are known to vary substantially [e.g. 72,77]. Furthermore, species of Dorid nudibranchs
display substantial degrees of seasonal and geographic variation in their chemical defences [46,72]
and sample sizes (n = 1-3 extractions of pooled individuals per species) in [46] are unlikely to capture
this variability. However, larger sample sizes for each species for both, chemical defences and colour
patterns, will be of great interest in future studies. Despite such limitations, our results show that
considering individual variability as opposed to using species averages and defining chemical defences
in terms of categories as opposed to a continuous scale shapes the conclusions on the distance-
dependent signalling function of aposematic animals. This is especially true considering that, despite
considering many of the most common species, this study only captures 13 out of hundreds of species
of sea slugs present in the study area [78]

Perceptual mechanisms underlying prey detection, identification and assessment are complex
and remain insufficiently quantified in studies of visual ecology [41,47,52,79]. Furthermore, rather
than considering the entire colour pattern space available in QCPA we quantify detectability and
boldness using a pre-determined selection of image statistics based on existing literature. These
statistics are generated using a specific set of modelling parameters guided by the best available
knowledge. However, a significant amount of work remains to be done to identify relevant visual
features and modelling parameters in each behavioural context [52,79]. Therefore, revisiting distance
dependent signalling in nudibranch molluscs using a larger dataset with a more informed approach on

modelling and statistical analysis will remain of great interest for future research. And yet, our study

and represents the currently most common approach towards the quantification of visual signals in

our field, even when using analyses such as QCPA [e.g. 80,81].

In summary, our results suggest that many aposematic Dorid nudibranchs are equally as

cryptic by means of background matching as undefended species at greater viewing distances. Our

results further suggest a multifunctional, multicomponent design of colour patterns in aposematic

Dorid nudibranchs. Moreover, our findings emphasize the need for a cautious and well-defined use of

the term conspicuousness when discussing aposematism. It is important to distinguish between the

detectability and boldness of aposematic signals when investigating colour pattern functionality,

particularly when investigating signal honesty in aposematic species [82,83]. However, differential

selection on detectability and colour pattern boldness likely also applies to other forms of visual

signalling in prey animals, such as sexual or territorial signalling, providing a great area of investigation

for future studies.

Data accessibility

The data will be uploaded to UQ's e-space upon acceptance.

Author's contributions

CPvdB conceived the original study with KLC & JAE providing critical input; CPvdB collected and

analysed the data; CPvdB and KLC led the writing of the manuscript. All authors contributed critically

to the drafts and gave final approval for publication.

Competing interests

We declare we have no competing interests.

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References

1.   Cott HB. 1940 Animal coloration and natural selection. *Nature* **146**, 144–145.

2.   Poulton EB. 1890 *The colours of animals, their meaning and use, especially considered in the case of insects*. 2nd editio. London, UK: Kegan Paul, Trench, Trubner.

3.   Hämäläinen L, Hoppitt W, Rowland HM, Mappes J, Fulford AJ, Sosa S, Thorogood R. 2021 Social transmission in the wild can reduce predation pressure on novel prey signals. *Nat. Commun.* **12**, 1–11. (doi:10.1038/s41467-021-24154-0)

4.   Endler JA, Mappes J. 2004 Predator Mixes and the Conspicuousness of Aposematic Signals. *Am. Nat.* **163**, 532–547. (doi:10.1086/382662)

5.   Gittleman JL, Harvey PH. 1980 Why are distasteful prey not cryptic? *Nature*. **286**, 149–150. (doi:10.1038/286149a0)

6.   Cook SE, Roper TJ. 1989 Responses of chicks to brightly coloured insect prey. *Behaviour* **110**, 276–293. (doi:10.1163/156853989X00510)

7.   Prudic KL, Skemp AK, Papaj DR. 2007 Aposematic coloration, luminance contrast, and the benefits of conspicuousness. *Behav. Ecol.* **18**, 41–46. (doi:10.1093/beheco/arl046)

8.   Kenward B, Wachtmeister CA, Ghirlanda S, Enquist M. 2004 Spots and stripes: The evolution of repetition in visual signal form. *J. Theor. Biol.* **230**, 407–419. (doi:10.1016/j.jtbi.2004.06.008)

9.   Green NF, Urquhart HH, van den Berg CP, Marshall NJ, Cheney KL. 2018 Pattern edges improve predator learning of aposematic signals. *Behav. Ecol.* **29**, 1481–1486. (doi:10.1093/beheco/ary089)

10.   Speed M. 2000 Warning signals, receiver psychology and predator memory. *Anim. Behav.* **60**, 269–278. (doi:10.1006/anbe.2000.1430)

11.   Rowe C. 2013 Receiver psychology: A receiver's perspective. *Anim. Behav.* **85**, 517–523. (doi:10.1016/j.anbehav.2013.01.004)

12.   Guilford T, Dawkins MS. 1993 Receiver psychology and the design of animal signals. *Trends Neurosci.* **16**, 430–436. (doi:10.1016/S0003-3472(05)80600-1)

13.   Sillén-Tullberg B. 1985 The significance of coloration per Se, independent of background, for predator avoidance of aposematic prey. *Anim. Behav.* **33**, 1382–1384. (doi:10.1016/S0003-3472(85)80211-6)

14.   Aronsson M, Gamberale-Stille G. 2009 Importance of internal pattern contrast and contrast against the background in aposematic signals. *Behav. Ecol.* **20**, 1356–1362.

444   (doi:10.1093/beheco/arp141)

445   15.   Guilford T. 1986 How do 'warning colours' work? Conspicuousness may reduce recognition
446   errors in experienced predators. *Anim. Behav.* **34**, 286–288. (doi:10.1016/0003-
447   3472(86)90034-5)

448   16.   Gamberale-Stille G, Bragée C, Tullberg BS. 2009 Higher survival of aposematic prey in close
449   encounters with predators: an experimental study of detection distance. *Anim. Behav.* **78**,
450   111–116. (doi:10.1016/j.anbehav.2009.04.015)

451   17.   Sherratt TN, Speed MP, Ruxton GD. 2004 Natural selection on unpalatable species imposed
452   by state-dependent foraging behaviour. *J. Theor. Biol.* **228**, 217–226.
453   (doi:10.1016/j.jtbi.2003.12.009)

454   18.   Aubier TG, Sherratt TN. 2020 State-dependent decision-making by predators and its
455   consequences for mimicry. *Am. Nat.* **196**. (doi:10.1086/710568)

456   19.   Sherratt TN. 2002 The coevolution of warning signals. *Proc. R. Soc. B* **269**, 741–746.
457   (doi:10.1098/rspb.2001.1944)

458   20.   Barnett CA, Skelhorn J, Bateson M, Rowe C. 2012 Educated predators make strategic
459   decisions to eat defended prey according to their toxin content. *Behav. Ecol.* **23**, 418–424.
460   (doi:10.1093/beheco/arr206)

461   21.   Endler JA. 1978 A predator's view of animal color patterns. *Evol. Biol.* **11**, 320–364.

462   22.   Barnett JB, Cuthill IC. 2015 Distance-dependent defensive coloration. *Curr. Biol.* **24**, R1157–
463   R1158. (doi:10.1016/j.cub.2014.11.015)

464   23.   Speed MP, Ruxton GD. 2007 How bright and how nasty: Explaining diversity in warning signal
465   strength. *Evolution (N. Y).* **61**, 623–635. (doi:10.1111/j.1558-5646.2007.00054.x)

466   24.   Rothschild M. 1964 A note on the evolution of defensive and repellant odours of insects.
467   *Entomol.* **97**, 276–280.

468   25.   Barnett JB, Cuthill IC, Scott-Samuel NE. 2018 Distance-dependent aposematism and
469   camouflage in the cinnabar moth caterpillar (*Tyria jacobaeae* , Erebidae). *R. Soc. Open Sci.* **5**,
470   171396. (doi:10.1098/rsos.171396)

471   26.   Barnett JB, Michalis C, Scott-Samuel NE, Cuthill IC. 2018 Distance-dependent defensive
472   coloration in the poison frog *Dendrobates tinctorius* , Dendrobatidae. *Proc. Natl. Acad. Sci.* ,
473   201800826. (doi:10.1073/pnas.1800826115)

474   27.   Honma A, Mappes J, Valkonen JK. 2015 Warning coloration can be disruptive: Aposematic
475   marginal wing patterning in the wood tiger moth. *Ecol. Evol.* **5**, 4863–4874.
476   (doi:10.1002/ece3.1736)

477   28.   Marples NM, Kelly DJ, Thomas RJ. 2005 The evolution of warning coloration is not
478   paradoxical. *Evolution (N. Y).* **59**, 933–940. (doi:10.1111/j.0014-3820.2005.tb01032.x)

479   29.   Sherratt TN, Beatty CD. 2003 The evolution of warning signals as reliable indicators of prey
480   defense. *Am. Nat.* **162**, 377–389. (doi:10.1086/378047)

481   30.   Caro T, Stankowich T, Kiffner C, Hunter J. 2013 Are spotted skunks conspicuous or cryptic?
482   *Ethol. Ecol. Evol.* **25**, 144–160. (doi:10.1080/03949370.2012.744359)

483   31.   Tullberg BS, Merilaita S, Wiklund C. 2005 Aposematism and crypsis combined as a result of
484   distance dependence: functional versatility of the colour pattern in the swallowtail butterfly

485   larva. *Proc. R. Soc. B* **272**, 1315–1321. (doi:10.1098/rspb.2005.3079)

486   32.   Barnett JB, Scott-Samuel NE, Cuthill IC. 2016 Aposematism: balancing salience and
487   camouflage. *Biol. Lett.* **12**, 20160335. (doi:10.1098/rsbl.2016.0335)

488   33.   Barnett JB *et al.* 2016 Stripes for warning and stripes for hiding: spatial frequency and
489   detection distance. *Behav. Ecol.* **00**, 1–9. (doi:10.1093/gbe/evw245)

490   34.   Barnett JB, Cuthill IC, Scott-Samuel NE. 2017 Distance-dependent pattern blending can
491   camouflage salient aposematic signals. *Proc. R. Soc. B* **284**, 20170128.
492   (doi:10.1098/rspb.2017.0128)

493   35.   Campbell FW, Green DG. 1965 Optical and retinal factors affecting visual resolution. *J.*
494   *Physiol.* **181**, 576–593. (doi:10.1113/jphysiol.1965.sp007784)

495   36.   Mottram JC. 1915 Some Observations on Pattern-Blending with reference to Obliterative
496   Shading and Concealment of Outline. *Proc. Zool. Soc. London* **85**, 679–692.
497   (doi:10.1111/j.1469-7998.1915.00679.x)

498   37.   Endler JA. 1991 Interactions between predators and prey. In *Behavioural Ecology* (eds JR
499   Krebs, NB Davies), pp. 169–196. Oxford: Blackwell Scientific.

500   38.   Mappes J, Marples N, Endler JA. 2005 The complex business of survival by aposematism.
501   *Trends Ecol. Evol.* **20**, 598–603. (doi:10.1016/j.tree.2005.07.011)

502   39.   Hebets EA, Papaj DR. 2005 Complex signal function: Developing a framework of testable
503   hypotheses. *Behav. Ecol. Sociobiol.* **57**, 197–214. (doi:10.1007/s00265-004-0865-7)

504   40.   Rowe C. 1999 Receiver psychology and the evolution of multicomponent signals. *Anim.*
505   *Behav.* **58**, 921–931. (doi:10.1006/anbe.1999.1242)

506   41.   Ruxton GD, Allen WL, Sherratt TN, Speed MP. 2018 *Avoiding Attack*. New York: Oxford
507   University Press. (doi:10.1093/oso/9780199688678.001.0001)

508   42.   Drinkwater E *et al.* 2022 A synthesis of deimatic behaviour. *Biol. Rev.* **3**.
509   (doi:10.1111/brv.12891)

510   43.   Cortesi F, Cheney KL. 2010 Conspicuousness is correlated with toxicity in marine
511   opisthobranchs. *J. Evol. Biol.* **23**, 1509–18. (doi:10.1111/j.1420-9101.2010.02018.x)

512   44.   Winters AE, Wilson NG, van den Berg CP, How MJ, Endler JA, Marshall NJ, White AM, Garson
513   MJ, Cheney KL. 2018 Toxicity and taste: unequal chemical defences in a mimicry ring. *Proc. R.*
514   *Soc. B Biol. Sci.* **285**, 20180457. (doi:10.1098/rspb.2018.0457)

515   45.   Rudman WB. 1991 Purpose in pattern - the evolution of color in chromodorid Nudibranchs. *J.*
516   *Molluskan Stud.* **57**, 5–21.

517   46.   Winters AE, Chan W, White AM, Berg CP, Garson MJ, Cheney KL. 2022 Weapons or
518   deterrents? Nudibranch molluscs use distinct ecological modes of chemical defence against
519   predators. *J. Anim. Ecol.* **91**, 831–844. (doi:10.1111/1365-2656.13643)

520   47.   van den Berg CP, Troscianko J, Endler JA, Marshall NJ, Cheney KL. 2020 Quantitative Colour
521   Pattern Analysis (QCPA): A comprehensive framework for the analysis of colour patterns in
522   nature. *Methods Ecol. Evol.* **11**, 316–332. (doi:10.1111/2041-210X.13328)

523   48.   Coleman N, Cobb G, Mullins D. 2015 *Nudibranchs Encyclopedia*. Oakland, CA: Thomas Slone,
524   Masalai Press and Tim Hochgrebe.

525   49.   Debelius H, Kuiter RH. 2007 *Nudibranchs of the world*. Frankfurt, Germany: IKAN-

526   Unterwasserarchiv.

527   50.   Gosliner TM, Valdes A, Behrens DW. 2018 *Nudibranch & Sea Slug Identification - Indo-Pacific*.
528   2nd edn. Jacksonville, Fla: New World Publications.

529   51.   Winters AE, White AM, Dewi AS, Mudianta IW, Wilson NG, Forster LC, Garson MJ, Cheney KL.
530   2018 Distribution of Defensive Metabolites in Nudibranch Molluscs. *J. Chem. Ecol.* **44**, 384–
531   396. (doi:10.1007/s10886-018-0941-5)

532   52.   van den Berg CP, Endler JA, Papinczak DEJ, Cheney KL. 2022 Using colour pattern edge
533   contrast statistics to predict detection speed and success in triggerfish (Rhinecanthus
534   aculeatus) . *J. Exp. Biol.* (doi:10.1242/jeb.244677)

535   53.   Champ CM, Wallis G, Vorobyev M, Siebeck U, Marshall J. 2014 Visual acuity in a species of
536   coral reef fish: *Rhinecanthus aculeatus*. *Brain. Behav. Evol.* **83**, 31–42.
537   (doi:10.1159/000356977)

538   54.   Sibeaux A, Cole GL, Endler JA. 2019 The relative importance of local and global visual contrast
539   in mate choice. *Anim. Behav.* **154**, 143–159. (doi:10.1016/j.anbehav.2019.06.020)

540   55.   Sibeaux A, Camduras T, Endler JA. 2021 The role of boundary length and adjacent patch
541   contrast in guppy mate choice. *Behav. Ecol.* **32**, 30–40. (doi:10.1093/beheco/araa097)

542   56.   Endler JA, Cole GL, Kranz AM. 2018 Boundary strength analysis: Combining colour pattern
543   geometry and coloured patch visual properties for use in predicting behaviour and fitness.
544   *Methods Ecol. Evol.* **9**, 2334–2348. (doi:10.1111/2041-210X.13073)

545   57.   Hubel DH, Wiesel TN. 1962 Receptive fields, binocular interaction and functional architecture
546   in the cat's visual cortex. *J. Physiol.* **160**, 106–154. (doi:10.1113/jphysiol.1962.sp006837)

547   58.   Vorobyev M, Osorio D. 1998 Receptor noise as a determinant of colour thresholds. *Philos.*
548   *Trans. R. Soc. B Biol. Sci.* **265**, 351–358. (doi:10.1098/rspb.1998.0302)

549   59.   Hempel de Ibarra N, Giurfa M, Vorobyev M. 2001 Detection of coloured patterns by
550   honeybees through chromatic and achromatic cues. *J. Comp. Physiol. A Sensory, Neural,*
551   *Behav. Physiol.* **187**, 215–224. (doi:10.1007/s003590100192)

552   60.   Kuhn M. 2021 caret: Classification and Regression Training. R package version 6.0-90.
553   https://CRAN.R-project.org/package=caret.

554   61.   R Core Team. 2021 R: A Language and Environment for Statistical Computing.

555   62.   Hollander M, Wolfe DA. 1973 *Nonparametric Statistical Methods*. New York: John wiley &
556   Sons Ltd.

557   63.   Dunn OJ. 1964 Multiple Comparisons Using Rank Sums. *Technometrics* **6**, 241.
558   (doi:10.2307/1266041)

559   64.   Bland j. M, Altman DG. 1995 Statistics notes: Multiple significance tests: the Bonferroni
560   method. *BMJ* **310**, 170–170. (doi:10.1136/bmj.310.6973.170)

561   65.   Ogle H. D, Doll J, Wheeler P, Dino A. 2022 FSA: Fisheries Stock Analysis.

562   66.   Bates D, Maechler M, Bolker B, Walker S. 2015 Fitting Linear Mixed-Effects Models Using
563   lme4. *J. Stat. Softw.* **67**, 1–48. (doi:10.18637)

564   67.   Kuznetsova A, Brockhoff PB, Christensen RHB. 2017 lmerTest Package: Tests in Linear Mixed
565   Effects Models. *J. Stat. Softw.* **82**, 1–26. (doi:10.18637/JSS.V082.I13)

68. Peterson RA, Cavanaugh JE. 2020 Ordered quantile normalization: a semiparametric
transformation built for the cross-validation era. *J. Appl. Stat.* **47**, 2312–2327.
(doi:10.1080/02664763.2019.1630372)

69. Marshall NJ. 2000 Communication and camouflage with the same 'bright' colours in reef
fishes. *Philos. Trans. R. Soc. London. Ser. B Biol. Sci.* **355**, 1243–1248.
(doi:10.1098/rstb.2000.0676)

70. Cheney KL, Hudson J, de Busserolles F, Luehrmann M, Shaughnessy A, van den Berg C, Green
NF, Marshall NJ, Cortesi F. 2022 Seeing Picasso: an investigation into the visual system of the
triggerfish *Rhinecanthus aculeatus*. *J. Exp. Biol.* **225**, jeb243907. (doi:10.1242/jeb.243907)

71. Revonsuo A, Newman J. 1999 Binding and consciousness. *Conscious. Cogn.* **8**, 123–127.
(doi:10.1006/ccog.1999.0393)

72. Winters AE, Green NF, Wilson NG, How MJ, Garson MJ, Marshall NJ, Cheney KL. 2017
Stabilizing selection on individual pattern elements of aposematic signals. *Proc. R. Soc. B* **284**.

73. Dimitrova M, Merilaita S. 2010 Prey concealment: Visual background complexity and prey
contrast distribution. *Behav. Ecol.* **21**, 176–181. (doi:10.1093/beheco/arp174)

74. Dimitrova M, Merilaita S. 2012 Prey pattern regularity and background complexity affect
detectability of background-matching prey. *Behav. Ecol.* **23**, 384–390.
(doi:10.1093/beheco/arr201)

75. Stevens M. 2007 Predator perception and the interrelation between different forms of
protective coloration. *Proc. R. Soc. B* **274**, 1457–64. (doi:10.1098/rspb.2007.0220)

76. Briolat ES *et al.* 2019 Diversity in warning coloration: selective paradox or the norm? *Biol.*
Rev. **94**, 388–414. (doi:10.1111/brv.12460)

77. Layton KKS, Gosliner TM, Wilson NG. 2018 Flexible colour patterns obscure identification and
mimicry in Indo-Pacific Chromodoris nudibranchs (Gastropoda: Chromodorididae). *Mol.*
Phylogenet. Evol. **124**, 27–36. (doi:10.1016/j.ympev.2018.02.008)

78. Smith SDA, Davis TR. 2019 Slugging it out for science: volunteers provide valuable data on the
diversity and distribution of heterobranch sea slugs. *Molluscan Res.* **39**, 214–223.
(doi:10.1080/13235818.2019.1594600)

79. Troscianko J, Skelhorn J, Stevens M. 2017 Quantifying camouflage: how to predict
detectability from appearance. *BMC Evol. Biol.* **17**, 7. (doi:10.1186/s12862-016-0854-2)

80. Nokelainen O, Scott-Samuel NE, Nie Y, Wei F, Caro T. 2021 The giant panda is cryptic. *Sci. Rep.*
11, 1–10. (doi:10.1038/s41598-021-00742-4)

81. Rohr VA, Volkmer T, Metzler D, Küpper C. 2021 Neoptile feathers contribute to outline
concealment of precocial chicks. *Sci. Rep.* **11**, 5483. (doi:10.1038/s41598-021-84227-4)

82. Summers K, Speed MP, Blount JD, Stuckert AMM. 2015 Are aposematic signals honest? A
review. *J. Evol. Biol.* **28**, 1583–1599. (doi:10.1111/jeb.12676)

83. White TE, Umbers KDL. 2021 Meta-analytic evidence for quantitative honesty in aposematic
signals. *Proc. R. Soc. B Biol. Sci.* **288**. (doi:10.1098/rspb.2021.0679)

605