Dysfunctional Hippocampal-Prefrontal Connectivity without Neuronal Loss Underlies Psychotic-Like Behaviors Following Early-Life Seizure

Running head: HPC-PFC dysfunction and psychosis in epilepsy

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Abstract

Cognitive impairments and psychiatric symptoms affect up to half of patients with temporal lobe epilepsy and are often more detrimental to their quality of life than the seizures themselves. Evidence indicates that the neurobiology of epileptogenesis shares common pathophysiological mechanisms with psychiatric comorbidities. However, these mechanisms and how they relate to specific behavioral alterations are unclear. We hypothesized that a dysfunctional communication between the hippocampus and the prefrontal cortex, as a consequence of epileptogenesis, would be linked to behavioral and cognitive symptoms observed in the comorbidities of temporal lobe epilepsy. Here, we performed a multilevel study to investigate behavioral, electrophysiological, histopathological, and neurochemical long-term consequences of early-life *Status Epilepticus* in male rats. We found that adult animals submitted to early-life seizure (ELS) presented behavioral alterations typically found in animal models of psychosis, such as working memory deficits, hyperlocomotion, reduction in sensorimotor gating, and sensitivity to psychostimulants. Surprisingly, cognitive deficits were linked to an aberrant increase in hippocampus-prefrontal cortex (HPC-PFC) long-term potentiation (LTP) while sensorimotor alterations were associated with increased neuroinflammation, as

verified by glial fibrillary acidic protein (GFAP) expression, and altered dopamine neurotransmission. Noteworthy, ELS rats did not exhibit neuronal loss. Furthermore, ELS rats displayed impaired HPC-PFC theta-gamma coordination and an abnormal brain state during active behavior oddly similar to rapid eye movement (REM) sleep oscillatory dynamics. Our results point to impaired HPC-PFC functional connectivity as a possible pathophysiological mechanism by which an epileptogenic insult can cause cognitive deficits and psychosis-related manifestations even without structural damage, bearing translational implications for understanding and monitoring psychiatric comorbidities in epilepsy.

Introduction

Psychiatric disorders affect up to half of the patients with epilepsy and significantly impact their quality of life (Salpekar and Mula, 2019). Understanding the neurobiological mechanisms of these comorbidities is crucial to develop better treatments for patients with epilepsy. It is currently speculated that the neurobiology of epileptogenesis shares common pathophysiological mechanisms with non-ictal symptoms, especially cognitive and psychiatric symptoms (Jensen, 2011). Temporal lobe epilepsy (TLE) is the most prevalent type of acquired epilepsy and presents a higher incidence of psychiatric comorbidities than the general population and other extratemporal epilepsies (Dalmagro et al., 2012). Patients with TLE frequently have a history of *Status Epilepticus* (SE) during infancy, which is seen as an initial injury that triggers epileptogenic processes (Mathern et al., 2002).

Experimental studies indicate that early-life seizures (ELS) induce long-term cognitive and behavioral alterations with translational validity for the psychiatric manifestations observed in clinical epilepsies (Holmes, 2016). However, unlike clinical or experimental TLE, ELS models are not associated with severe neuronal loss (Baram et al., 2002). One possible mechanism is that ELS could have functional and molecular effects at the brain circuit level, producing relevant cognitive impairments (Kandratavicius et al., 2012b, 2012a). The hippocampus (HPC) is a fundamental brain region affected by ELS and TLE. HPC interaction with the prefrontal cortex (PFC) underlies cognitive processes and is impaired in neuropsychiatric disorders such as schizophrenia (Ruggiero et al., 2021). Impairments of HPC-PFC oscillatory dynamics are observed in genetic,

developmental, and pharmacological models of schizophrenia and are thought to be essential to produce psychotic-like behavior (Sigurdsson and Duvarci, 2016).

Although it is well recognized that PFC and HPC are important for neuropsychiatric disorders, little is known about how their neuronal dynamics are affected by ELS. Here, we hypothesized that ELS would produce long-term alterations in HPC-PFC neuronal dynamics related to cognitive deficits and behavioral abnormalities (Ruggiero et al., 2012). To test this hypothesis, we induced SE as a form of prolonged ELS and performed behavioral tests to assess cognitive processes and psychotic-like behaviors. Then, we performed neurophysiological, immunohistochemical, and neurochemical experiments to study potential changes in the HPC-PFC circuits that could underlie such behavioral alterations. We found that adult ELS rats present a myriad of behavioral impairments that are typically observed in animal models of psychosis and schizophrenia: working memory deficits, hyperlocomotion, sensorimotor gating impairment, and behavioral sensitivity to psychostimulants. Remarkably, we found an enhanced long-term potentiation (LTP) in the HPC-PFC pathway of ELS animals, which was strongly associated with cognitive impairment. Our results also describe neuroinflammation and alterations in dopaminergic neurotransmission as major contributors to psychotic-relevant behavior. Freely-moving experiments allowed us to observe HPC-PFC dynamics during different brain states. ELS rats displayed an increased HPC theta and impaired coordination of PFC gamma activity during active behavior. During rapid eye movement (REM) sleep, ELS rats showed a reduction in HPC-PFC connectivity. Notably, ELS rats presented an aberrant brain state during active behavior characterized by oscillatory dynamics similar to REM sleep. Finally, we implemented a linear discriminative model, which demonstrated that collective patterns of oscillatory alterations accurately distinguished ELS from control animals. Therefore, we identified a convergence of dysfunctional patterns at multiple levels linking epileptogenesis and psychotic-like behaviors. These convergent patterns indicate that epileptogenic processes triggered during early life, such as neuroinflammation, could lead to altered neurotransmission and consequently aberrant dynamics in the HPC-PFC coordination, potentially contributing to the psychiatric comorbidities of epilepsy.

Materials and Methods

Animals

Wistar rats were housed in a colony room with controlled temperature (22 ± 2 °C) and a 12-h light/dark cycle with free access to food and water. Litters contained up to 8 animals with a maximum of 5 males. Rats were weaned at 21 days postnatally (P21). All procedures were performed according to the National Council for the Control of Animal Experimentation guidelines for animal research and approved by the local Committee on Ethics in the Use of Animals (Ribeirão Preto Medical School, University of São Paulo; protocol: 159/2014).

Experimental Designs

We used four cohorts (a total of 17 litters) to compare the effects of early-life *Status Epilepticus* (ELS) *versus* control (CTRL) animals. The first cohort was used for acute electrophysiology during the induction of ELS (n=5, ELS only). The second was used for behavioral tests (CTRL n=11, and ELS n=14), synaptic plasticity experiments (CTRL n=7, and ELS n=7), and immunohistochemistry (CTRL n=9, and ELS n=11). The third was used to investigate HPC-PFC dynamics in freely moving rats (CTRL n=6, and ELS n=9). Finally, in the fourth cohort, we assessed behavioral sensitization to psychostimulants (CTRL n=15, and ELS n=14) and neurochemical quantification (CTRL n=8, and ELS n=9).

Early-Life Status Epilepticus

At P11-P12 we induced a 2-hour SE using lithium chloride (LiCl, 127 mg/kg, i.p., Sigma-Aldrich, USA) 18-20 prior to injection of methylscopolamine (1 mg/kg, i.p., Sigma-Aldrich) and pilocarpine (80 mg/kg, s.c., Sigma-Aldrich, USA) (Kubová and Mareš, 2013). Behavioral seizures were monitored following the Racine scale (Racine, 1971). SE was interrupted by diazepam (5 mg/kg, i.p., Teuto, Brazil) at the 2-hour mark. During the recovery period (1 week post-ELS) we provided food supplementation and subcutaneous rehydration with saline and monitored body temperature and weight.

Behavior

The radial-arm maze working memory task was based on Floresco et al. (1997). Briefly, rats were kept at 85-90% of their initial body weight throughout

the 21 days of the experiment. Training consisted of single daily sessions, each one divided into the training phase and test phase, 30 min apart. During the training session, four doors were kept open providing access to food pellets. In the test session, all doors remained open, but with pellets available only in the four arms that were closed in the previous training session. The sessions were terminated upon full pellet consumption or after 5 min. Two types of errors were quantified: Reference error: reentering arms previously visited during the training session; Working memory error: reentering an arm already visited during the test session. The sum of both categories denoted the total errors for each animal.

Three days after returning to *ad libitum* diet, locomotor activity was evaluated for 30 min in the open field test (Wolf et al., 2016).

Immediately after the open field test, rats were examined for prepulse inhibition (PPI) of the acoustic startle. The PPI test consisted of a habituation period (5 min) followed by 72 trials (intertrial interval: 15 ± 8 s) with prepulse alone (PP), startling pulse alone (P), prepulse followed by a startling pulse (PP + P), and no stimulus, all of which accompanied by background noise (65 dB white noise). Startle-eliciting stimuli were presented at 120 dB for 40 ms (P) and prepulse stimuli were presented for 20 ms (PP, 1000 Hz; 71, 77, and 83 dB; 20 ms). Trials were presented in a random order, with each trial type being presented six times. We used the equation: %PPI = 100 − 100 × (mean startle amplitude at PP + P)/(mean startle amplitude at P) (Wolf et al., 2016).

In order to measure the effects of ELS on sensitization to psychostimulants, we performed the apomorphine-induced locomotion test. At P80-90, rats were placed in the open field apparatus and were allowed to roam for 60 min. Then, we injected apomorphine (APO, 1.5 mg/kg, i.p., Sigma-Aldrich, USA) and measured locomotion for another 60 min. The distance across blocks was normalized by Z-score to the mean of baseline blocks.

Synaptic Plasticity

HPC-PFC synaptic plasticity studies were performed as described in

previous works from our group (Bueno-Junior et al., 2018; Ruggiero et al., 2018).

Briefly, rats were anesthetized with urethane (1.2 mg/Kg in NaCl 0.15 M, i.p.) for

electrode implantation. A recording electrode (Teflon-coated tungsten wires 60

μm, AM-Systems) was inserted in the medial PFC (prelimbic region (PL), anterior-

posterior: 3.0 mm; medial-lateral, ML: 0.5 mm; dorsal-ventral, DV: 3.2 mm), and

a bipolar stimulation electrode (twisted wires; ~500 μm inter-pole distance) was

inserted in the ipsilateral HPC (intermediate CA1 AP: −5.6 mm; ML: 4.5 mm; DV:

2.5 mm). An additional burr hole was drilled over the right parietal cortex for a

reference screw. Temperature was kept constant at 37 ± 0.5°C during the

experiment.

HPC was stimulated with single monophasic pulses (200 μs, 0.05 Hz,

150–300 μA, 80-ms inter-pulse interval) while recording PFC field postsynaptic

potentials (fPSPs). Stimulation intensity (60–500 μA) was calibrated to evoke 50–

60% of the maximal fPSP amplitude. Field recordings were conditioned through

a preamplifier (100× gain, 0.03–3 kHz bandpass; Grass) and digitized at 10 kHz

(ADInstruments). For LTP induction, we used a high-frequency stimulation (HFS)

protocol consisting of 2 series (10 min apart) of 10 trains (0.1 Hz), each train with

50 pulses at 250 Hz (Ruggiero et al., 2018). fPSP amplitudes and slopes were

computed as the difference and slope between the positive and negative peaks,

respectively. Short-term synaptic plasticity was estimated by paired-pulse

facilitation (PPF), calculated as the ratio of the fPSP2 and fPSP1 (Lopes-Aguiar

et al., 2020). Amplitudes and slopes were normalized as ratios of the baseline

mean and averaged into 10-min bins.

Brain Tissue Processing

Immunohistochemistry

We employed a standardized immunohistochemistry protocol (Peixoto-
Santos et al., 2015) to estimate NeuN+ neuronal density and to evaluate
parvalbumin-expressing (PV) interneurons, mGluR5 expression, and glial
fibrillary acid protein (GFAP) reaction. Briefly, endogenous peroxidases were
blocked and the antigens were exposed by microwave-induced retrieval. We
used the primary antibodies against NeuN (Chemicon, USA; 1:1000), PV
(Calbiochem, USA; 1:500), mGluR5 (Millipore, USA; 1:200), and GFAP (Dako,
Denmark; 1:500) proteins, and biotinylated secondary antibodies (Dako,
Denmark; 1:200). Revelation was performed with Elite ABC Kit (Vector) and
diaminobenzidine solution (Pierce).

Images from the regions of interest were captured at 200× magnification
in the Axio Scope.A1 microscope system (Carl Zeiss). The regions of interest
included the hippocampus (CA1 and GL, granule layer of dentate gyrus),
entorhinal cortex (EC), infralimbic (IL) and prelimbic cortex (PL), and thalamic
reticular nucleus (TRN). Images were then processed in Image J software (NIH,
USA; 1.48v). Data are shown as the density of NeuN positive cells and
immunopositive area for PV, mGluR5, and GFAP.

High-Performance Liquid Chromatography (HPLC)

Samples were taken bilaterally from the nucleus accumbens (NAc), PFC,
and HPC. The samples were homogenized with ultrasound in a 0.1 mol/L $HClO_4$
0.02% $Na_2S_2O_2$ solution containing dihydroxybenzylamine (DHBA, 146.5 ng/mL),
and homoserine (HSER, 10 µg/mL) as internal standards and at a dilution of 15
µL per sample milligram. We employed Shimadzu LC-10AD isocratic systems
with a fluorescence detector for glutamate (Glu) and gamma-aminobutyric acid
(GABA), and an electrochemical detector for dopamine (DA) and its metabolites
3,4-dihydroxyphenylacetic acid (DOPAC) as previously published (Castro-Neto
et al., 2013). The neurotransmitter levels were considered to reflect
neurotransmitter stock in synaptic vesicles, their metabolites reflected the
release, and metabolite/neurotransmitter ratios were taken as an index of
neurotransmitter turnover. Standards containing all amino acids and
monoamines/metabolites of interest were employed to define their respective
peaks in the samples.

 Freely Moving Electrophysiology

Electrophysiological recordings

Chronic head caps consisted of two eight-channel connectors (Omnetics),

one for the PFC (PL, 7 wires) and another for the HPC (intermediate CA1, 7 wires)

(Bueno-Junior et al., 2017; Marques et al., 2022). We additionally implanted a

recording wire into the trapezius muscle to serve as an electromyogram (EMG).

The EMG wire was soldered to the HPC connector. Microscrews were fastened

into the skull, including a ground reference in the intraparietal area. Animals were

allowed to recover for 8-9 days before recordings.

To record electrophysiological signals through sleep-wake states, rats

were placed into a shuttle box apparatus inside a soundproof box with food and

water *ad libitum*. Recordings varied between 30 to 48 hours, divided later into

periods of 6 hours for analysis. Local field potentials (LFP) were recorded through

a multichannel acquisition processor (Plexon) (1000× gain, 0.7-500 Hz bandpass

filter, 1-kHz sampling rate).

Classification of sleep-wake states

Classification of the sleep-wake cycle states was carried out through a

semi-supervised machine learning pipeline. The HPC theta (4.5-12 Hz), PFC

delta (0.7-4 Hz), and EMG (100-200 Hz) power were computed in 5-second

epochs without overlaps (4 tapers; time-bandwidth product: 2.5). A support vector

machine (SVM) (radial basis function) was trained to discriminate the epochs into:

(1) awake or sleep epochs using EMG power; (2) REM or NREM epochs based

on theta/delta ratio; (3) quiet wake (QWK) or active (ACT) epochs based on

theta/delta ratio. Later, we classified our recordings based on 30-second epochs

by grouping sets of six 5-second epochs and considering the most representative

state.

Spike wave-discharge detection (SWD)

For spike wave-discharge detection, we used the PFC recordings. First,

the short-time Fourier transform (STFT) was computed within 3-70 Hz throughout

250-ms windows with 50% overlap. A detection threshold was defined as the

mean + 1.5 standard deviation of the absolute value of the STFT of the frequency

bands 4.4-16.4, 17.6-32.8, and 35.1-55 Hz throughout the entire recording. SWD

events were detected when the average STFT of the aforementioned frequency

bands at one epoch exceeded the threshold. Consecutive epochs above the

threshold were grouped as the same event. The beginning and the end of an

event were defined as the closest epochs when the average STFT was lower

than the mean + 1 standard deviation of STFT of the frequency bands of interest.

Lastly, the detected putative events were manually sorted by an expert.

Spectral estimates

We divided 30-s epochs into 3-s epochs for spectral analyses. Power

spectrum densities (PSD) and magnitude-squared coherence were obtained

using Welch's method with 1-s segments, 50% overlap, and Hamming

windowing. Relative power was obtained by dividing values by the sum within

0.5-55 Hz for each epoch. We defined the frequency bands: delta (1-4 Hz), theta

(4.5-11 Hz), low gamma (30-55 Hz), and high-gamma (65-100 Hz) to compare

estimates and obtained the spectral peak of each epoch using MATLAB function

findpeaks. We used the low and high-gamma peaks values to estimate a

probability density function using kernel density estimate method (*kde* function in

MATLAB) and compare ACT and REM state distributions.

To quantify theta oscillation power we used the FOOOF algorithm

(Donoghue et al., 2020), which parametrizes the PSD separating the periodic and

aperiodic components of the signal. We used frequencies within 3-40 Hz, the

"fixed" aperiodic mode, a minimum peak height of 0.05, and a maximum number

of peaks of 4 (https://fooof-tools.github.io/fooof/). EMG data were low-cut filtered

(>30 Hz) and the sum of power within 30-200 Hz was obtained.

As a synchrony measure, we also computed cross-correlations between

each pair of LFPs (low-pass filter of 50 Hz) recorded from the HPC and PFC using

a maximum lag of 1 s. Coefficients were normalized so that the autocorrelations

at zero lag were one. We observed that the highest coefficients occurred in the

range of -80 to -20 ms (HPC leading the PFC), so we obtained the maximum

coefficient within this range for each rat average for comparisons. Cross-

spectrum power was calculated using Welch's method for the normalized

correlation coefficients from -1 to 1 s.

For cross-frequency amplitude spectrum correlations, we computed the

absolute STFT for each 3-s epoch throughout 100-ms segments with 50%

overlap and Hamming windowing across 0.5-Hz frequency bins within 0.5-200 Hz. We calculated the correlation matrix for each frequency pair using Pearson's R.

State maps

After state classification and spectral analysis, we computed two-dimensional state maps to investigate the natural distinction between sleep-wake states. The state maps were computed according to Gervasoni et al. (2004) and Dzirasa et al. (2006). First, we obtained the power *ratio 1* (4.5-9/4.5-50 Hz) and *ratio 2* (2-20/2-50 Hz) for each region and the first principal component, estimated by singular value decomposition algorithm considering both regions, for each ratio. Finally, we applied a Hann window of 10 epochs on scores throughout time for smoothing.

To identify the intermediate states (IS), we first estimated the trajectory *speed* of epochs throughout state map dimensions as the Euclidean distance between each epoch and its predecessor. Then, we defined a threshold of 0.1 and labeled the epochs with greater speed as IS. This threshold was determined heuristically, corresponding closely to the expected value plus a standard deviation from a Poisson distribution across rats.

From visual inspection and the literature (Gervasoni et al., 2004; Dzirasa et al., 2006), the state maps usually separate three well-defined clusters heuristically corresponding to awake, REM, and NREM sleep. To assure the correspondence between state map clusters and the classification outcomes, we performed Gaussian mixture models clustering of the state maps for 3 clusters and 80% posterior probabilities and computed Chi-squared tests with the classified NREM, REM, and awake (QWK+ACT) labels. Three animals were excluded for having outlier lower Chi-values. Although QWK and ACT states are often evenly distributed inside the awake cluster, QWK epochs may overlap with intermediate states closer to NREM, so the ACT epochs were considered to represent more precisely the localization of neural dynamics associated with behavior in the state map. The dissimilarity between ACT and REM states was estimated by the average pairwise state map Euclidean distance of all epochs between the two states.

We used machine learning algorithms to decode sleep-wake states from the state map. We used the ACT and REM state map *ratios 1* and *2* values. We applied a linear and radial basis function SVM *fictcsvm* function in MATLAB ('KernelFunction', 'linear' or 'rbf') as well as a Random Forest algorithm using the *fitcensemble* function in MATLAB ('Method', 'bag', 'NumLearningCycles', 50, for Random Forest) to decode states. We used the ACT and REM epochs from all the rats creating one single model for the CTRL or ELS condition. Using the same approach, we also constructed a random forest decoder model with non-reduced variables using 1 to 55 Hz (0.5 Hz bins) for the PFC and HPC power and coherence. Discrimination was evaluated using the area under the receiver operating characteristic curve (AUC-ROC). The point-wise confidence interval was calculated using a 1000-repetition bootstrap. Average AUC-ROC values were calculated using the prediction of the trained model on 20% of the epochs as test data and repeating the process 100 times.

Statistical Analysis

Data were analyzed using GraphPad Prism Software version 9.00 (GraphPad Software, LLC), and R studio version 2021. We used paired Student *t*-tests for within-group comparisons and two-sample *t*-tests for between-group comparisons. Two-way ANOVA with repeated measures (RM) was performed for comparisons across time bins, using Sidak's multiple comparison test as a *post-hoc*. To examine linear model assumptions, we analyzed the distribution of the residuals using a Q-Q plot, the Kolmogorov-Smirnov test, homoscedasticity, and the residual plot. We used the Mann-Whitney test to compare two groups and the Wilcoxon signed-rank test to paired comparisons as non-parametric alternatives. We calculated Pearson's linear correlation and Spearman's rank correlation as a nonparametric equivalent. To compare correlation and spectral coherence estimates we applied Fisher's Z transform to the data. For multiple comparisons in the neurochemical quantification, we performed Holm's procedure for multiple comparisons correction across neurotransmitters and metabolites per region.

Data are expressed as the mean ± SEM. The significance level was set to 0.05 unless stated otherwise. For some multiple comparisons, statistical values are expressed as the range. Significances are expressed as [†]$p < 0.1$, *$p < 0.05$, **$p < 0.01$, and ***$p < 0.001$.

Linear mixed-effects models

We also constructed statistical inferential models across time controlling for individual variability and litter using linear mixed-effects models. We compared power and coherence estimates throughout time by dividing the electrophysiological recording into 5 periods (6 hours). Group and time (periods) were assigned as fixed effects and subject, litter, and time were assigned as random effects. We used this approach to construct a maximal random effects structure model as preconized in the literature as more generalizable (Barr et al., 2013). The model was estimated by restricted maximum likelihood using the *lme* function of the *nlme* package (Pinheiro et al., 2023) with an unstructured covariance matrix.

Multivariate analysis

Each variable was Z-scored before multivariate analysis. We constructed correlograms to assess the relationship within behavioral and neurobiological measures. Variables were sorted by agglomerative hierarchical clustering using Ward's method and correlation as the metric. We also performed agglomerative hierarchical clustering of subjects using Ward's method and Euclidean distance as the metric to assess how unsupervised clustering by our multivariate data might match the experimental groups. An optimal number of clusters was estimated using the silhouette method and final definition was based on parsimonious interpretability of the results.

Principal component analysis (PCA), using singular-value decomposition, was used for dimensionality reduction to find collective patterns of variance within behavioral variables (behavior factors) and neurobiological variables (neurobiology factors), and to reduce each immunohistochemical measure across all brain regions into single scores. Principal components (PCs) were projected onto data and the mean score was compared between conditions.

Canonical correlation analysis (CCA) was performed to describe multivariate relationships between behavioral and neurobiological measures. Missing data were imputed using the multiple imputation by chained equations algorithm. For CCA, we used the first PC (PC1) and second PC (PC2) of behavioral variables as inputs for the behavioral factors and the PC1 of each

immunohistochemical variable (NeuN, PV, mGluR5, and GFAP,) across all brain regions and the Z-scored LTP data as inputs for the neurobiological factors. Variables were narrowed down from 30 to 7, which is statistically appropriate for our sample size. We used the CCA package in R to obtain the canonical dimensions and performed the Hotelling-Lawley trace statistic to assess the significance of each dimension.

We used multivariate generalized linear models (GLM) with a Gaussian family and an identity link function. Quasi-Poisson regression (quasi-Poisson family and logarithmic link function) were used for count data with overdispersion (Errors in the radial maze) using a quadratic covariate (LTP). We constructed two types of models: 1) a full model considering all the covariates; and 2) a restricted model using backward stepwise model selection based on the Akaike information criterion.

To assess the effectiveness of neurophysiological features in discriminating CTRL from ELS rats, we fitted a regularized linear discriminant classifier model on the PCA scores of selected electrophysiological variables. We used the MATLAB function *fitcdiscr* without hyperparameter optimization. The linear classifier using two predictors allowed the graphical representation and avoided the overfitting of a high number of variables. Accuracy was determined as the probability of correct category assignments using leave-one-out cross-validation. We also determined the contribution of each original neurophysiological variable by assessing their respective AUCs using a univariate logistic regression model.

Results

ELS induces a psychotic-like behavioral phenotype

SE induced by lithium-pilocarpine in early life (Fig. 1A) produced an unequivocal and stereotyped seizure, consisting mainly of forelimb myoclonus and orofacial automatism (Racine 3), with some animals showing further development into bilateral forelimb myoclonus, rearing and falling (Racine 4 and 5)(Racine, 1971). Seizures were confirmed electrophysiologically in freely moving recordings of the frontal cortex and HPC, with paroxysms starting 10 min after pilocarpine injection and persisting for at least 120 min (Fig. 1B).

Next, we investigated the long-term behavioral consequences of ELS (Fig. 2A). First, we evaluated possible weight differences as a confounder of behavioral measures. ELS rats displayed weight loss on the day after SE (Two-way RM ANOVA, treatment effect: $F_{(9,207)}$ = 11.7, $p < 0.0001$, Sidak's *post-hoc: p < 0.0001*; Fig. 2B). However, ELS animals recovered the pace of normal weight gain in the subsequent days (*post-hoc: p > 0.05*) and into adulthood (Fig. 2C), showing no distinction to CTRL before (*t*-test, $t_{(23)}$ = 1.440, $p = 0.1634$) or during food restriction (*t*-test, $t_{(23)}$ = 0.0535, $p = 0.9577$), indicating that behavioral effects were unrelated to weight differences or alterations in food-seeking.

Importantly, when examined for spatial memory in the radial-arm maze, ELS rats presented a specific increase in working memory errors (Mann-Whitney test, U = 36.5, $n(CTRL, ELS)$ = 11, 14, $p = 0.0254$; Fig. 2D). We found no differences in the total reference errors (*t*-test, $t_{(23)}$ = 1.719, $p = 0.0991$; Fig 2D) and in the learning curve (Two-way RM ANOVA, treatment effect: $F_{(1,23)}$ = 0.8032, $p = 0.3794$; Fig 2E). ELS rats presented significantly higher locomotion in the open field test (Mann-Whitney test, U = 27, $n(CTRL, ELS)$ = 11/14, $p = 0.0051$; Fig. 2F). ELS rats also presented a higher startle response (*t*-test, $t_{(23)}$ = 2.757, $p = 0.011$; Fig. 2G) and a deficit in sensorimotor gating represented by a reduction in PPI of the acoustic startle (Two-way RM ANOVA, treatment effect: $F_{(2,46)}$ = 19.6, $p < 0.0001$, Sidak's *post-hoc:* PPI 71 dB: $p = 0.0017$; PPI 77 dB: $p = 0.1040$; PPI 83 dB: $p = 0.030$; Fig. 2H).

Interestingly, all these behavioral alterations are typical of animal models of psychosis (Sigurdsson, 2016). We investigated if collective patterns of behavioral alterations induced by ELS could reflect latent factors representing more comprising behavioral dimensions associated with psychosis. Working memory errors in the radial-arm maze showed little correlation with the other behavioral variables, while psychotic-like behaviors presented a strong correlation with each other. PPI exhibited a positive correlation with every intensity and a negative correlation with startle response and locomotion (Fig. 2I). We also performed PCA to distinguish patterns of variance between multivariate behavioral data (Fig. 2J-M). Our analysis indicated that dimensionality reduction by PCA using the first two components explained 71.32% of the variability of the data (Fig. 2J). By analyzing the loadings of the PC1, the variables that contribute more strongly (>0.5) to this dimension were: high total distance, startle reflex, and

low PPI scores, indicating this component represents a latent factor of sensorimotor dysfunction related to psychotic-like behaviors. In turn, the PC2 represents the cognitive dimension of the behavioral variables, as it presents a high loading value (>0.5) for working memory errors (Fig. 2K). Finally, we found that these PCs clearly distinguished CTRL from ELS animals, which presented higher values of PC1 (PC1: t-test, $t_{(23)}$ = 3.9713, p = 0.0006; PC2: t-test, $t_{(23)}$ = 1.6466, p = 0.1132; Fig. 2L-M). These results indicate that the covariance across behavioral tests can be summarized by a latent *psychotic factor* (PC1) and a *cognitive factor* (PC2), representing two distinct dimensions of the behavioral phenotype and suggesting possibly different neurobiological bases.

Aberrant HPC-PFC synaptic plasticity in animals submitted to ELS

Next, we tested the hypothesis that behavioral abnormalities in ELS rats could be associated with synaptic plasticity impairments of the HPC-PFC pathway (Fig. 3A-C). First, we looked for ELS-related alterations in basal synaptic efficacy before inducing LTP. Synaptic efficacy was assessed in terms of basal fPSP amplitude and slope (amplitude: t-test, $t_{(12)}$ = 1.278, p = 0.2255; slope: t-test, $t_{(12)}$ = 0.5470, p = 0.5944; Fig. 3D), input-output curve (fPSP amplitude as a function of stimulus intensity) (Two-way RM ANOVA, treatment effect: $F_{(1,12)}$ = 0.6596, p = 0.4325; Fig. 3E) and PPF curve (PPF as a function of the inter-pulse interval) (Two-way RM ANOVA, treatment effect: $F_{(1,12)}$ = 0.3345, p = 0.5737; Fig. 3F). None of these measures indicated alterations in ELS when compared to CTRL (Fig. 3D-F).

Then, we proceeded to the actual synaptic plasticity experiments and found that ELS rats presented an abnormally augmented LTP compared to CTRL (Fig. 3G; Two-way RM ANOVA, treatment effect: $F_{(1,12)}$ = 7.54, p = 0.0177; interaction effect: $F_{(26,312)}$ = 7.393, p < 0.0001; Sidak's *post-hoc: p* < 0.05 for 10-60 min; t-test, $t_{(12)}$ = 2.747, p = 0.0177; Fig. 3G inset), despite the lack of alterations in basal synaptic efficacy described above (Fig. 3D-F). Aberrant LTP in the ELS rats was also observed when analyzing the fPSP slope (Two-way RM ANOVA, treatment effect: $F_{(1,12)}$ = 4.757, p = 0.0498; interaction effect: $F_{(26,312)}$ = 4.120, p < 0.0001; Sidak's *post-hoc: p* < 0.05 for 10-40 min; Fig. 3H; t-test, $t_{(23)}$ = 2.794, p = 0.0162; Fig. 3H inset). However, short-term plasticity measured by PPF decreased during the initial 30 min of recording (Two-way RM ANOVA,

interaction effect: $F_{(26,312)} = 3.457$, $p < 0.0001$; Sidak's *post-hoc: p < 0.05* for 20-30 min; Fig. 3I), possibly reflecting a ceiling effect on fPSP amplitude after LTP induction. With these results, we demonstrate that, although ELS does not affect basal synaptic efficiency, it significantly increases the ability of HPC-PFC circuits to undergo LTP.

ELS induces long-term neural inflammation, but not neuronal loss

We sought to investigate histopathological alterations in cortico-limbic circuits that could underlie the abnormalities in behavior and HPC-PFC synaptic plasticity of ELS rats (Fig. 4A). Noteworthy, ELS rats did not present neuronal loss when adults (Mixed-effects model, $F_{(1,12)} = 0.7683$, $p = 0.3980$; uncorrected Mann-Whitney test, $p = 0.1666$-0.9372; Fig. 4B). Since neuronal loss is a conspicuous finding related to cognitive deficits and behavioral alterations in clinical and experimental TLE, our data show that cognitive impairments and psychotic-like phenotypes can occur even without significant neuronal loss in the HPC or PFC. Also, ELS did not produce long-term effects in the expression of PV (Mixed-effects model, $F_{(1,13)} = 0.008$, $p = 0.9297$; uncorrected Mann-Whitney test, $p = 0.3659$-0.9307; Fig. 4C) or mGluR5 receptors (Mixed-effects model, $F_{(1,11)} = 0.0011$, $p = 0.9735$; uncorrected Mann-Whitney test, $p = 0.2467$-0.8181; Fig. 4D). Finally, we found a strong increase in neuroinflammation, verified by the marked expression of GFAP in ELS rats in all regions investigated except the infralimbic cortex (Mixed-effects model, $F_{(1,18)} = 5.368$, $p = 0.0325$; uncorrected Mann-Whitney test, $p = 0.0067$-0.0419 for GL, CA1, EC, PL, and TRN, and $p = 0.2298$ for IL; Fig. 4E). These results show that ELS produces brain inflammation later in life but without an evident neuronal loss.

Cognitive impairment and psychotic-like behavior are distinctly correlated with abnormal LTP and neuroinflammation

By understanding the association between behavioral and neurobiological variables, we would be able to infer putative mechanisms underlying the behavioral comorbidities observed in ELS animals. To investigate multivariate relationships between the behavioral (cognitive and psychotic factors) and neurobiological (synaptic plasticity and immunohistochemical measures) dimensions of ELS effects, we conducted CCA (Fig. 5A). CCA produced a

significant high-correlation model ($r = 0.94$, Hotteling-Lawley trace test, *F-*
approximation$_{(10,12)}$ = 3.2311, $p = 0.0164$; Fig. 5B) that clearly separated CTRL
from ELS (behavior canonical dimension, *t*-test, $t_{(11)}$ = 3.693, $p = 0.0035$;
neurobiology canonical dimension, *t*-test, $t_{(11)}$ = 3.917, $p = 0.0024$; Fig. 5C). By
analyzing the coefficients, we observed that the behavior canonical dimension
showed higher values for the psychotic factor, but also included the cognitive
factor, although at a lower degree (Fig. 5D). The neurobiology canonical
dimension showed a larger value for the GFAP, intermediate values for LTP and
PV and the lowest coefficients for NeuN and mGluR5 (Fig. 5D). A similar pattern
was observed when analyzing the loadings for each dimension (Fig 5E). Notably,
the neurobiological variables have only weak correlations between them (Fig.
5F). To further dissect specific multivariate relationships between behavioral and
neurobiological measures, we constructed generalized linear models (GLM). The
GLM indicates GFAP, PV, and LTP as the most significant variables that explain
the psychotic factor variance (coefficients = 2.3775, 0.8457, and 0.5187; $p =$
0.1164, 0.00548, 0.08474 for GFAP, PV, and LTP, respectively, (Fig. 5G).
Accordingly, GLM multivariate regression indicates a strong association between
the psychotic factor and all significant GLM coefficients (GFAP + PV + LTP: $r^2 =$
0.7974, $p = 0.0116$; Fig. 5H), as well as psychotic factor and univariate
neurobiological measures (GFAP: $r = 0.6840$, $p = 0.0099$; Fig. 5I, PV: $r = 0.6548$,
$p = 0.0151$; Fig. 5J). Strikingly, we found a quadratic relationship between
synaptic plasticity effects and cognitive impairment. Poor performance on the
working memory test was observed in rats that presented too low or too high
values of HPC-PFC LTP, suggesting a U-shaped relationship between LTP and
cognitive performance (Fig. 5K-L). We observed this quadratic relationship using
both the cognitive factor (gaussian GLM, LTP coefficient = 1.22, $p = 0.0162$; Fig.
5 K) and raw error values from the radial maze test using a quasi-Poisson GLM
($p = 0.00843$; Fig. 5L).

Taken together, both neuroinflammation and synaptic plasticity alterations
may underlie the behavioral effects of ELS, with neuroinflammation being more
associated with the psychotic-like dimension and LTP with the cognitive
dimension.

ELS animals present epileptiform-like activity when adults

Given the results above, we decided to further investigate the relationships

between circuit dynamics and behavioral dysfunctions during different brain

states using chronic electrophysiology recordings. We classified LFP epochs into

active wake (ACT), non-REM (NREM) sleep, rapid eye movement sleep (REM),

and quiet wake (QWK; Fig. 6A-D). No CTRL *vs.* ELS differences were found in

sleep architecture, according to the mean consecutive duration of each state,

percentage of time awake, number of persistent episodes in each state (Two-way

RM ANOVA, $F_{(1,13)}$ = 0.6500, p > 0.4346; Fig. 6E-G), and percentage of time in

each state (Two-way RM ANOVA, $F_{(1,13)}$ = 0.5425, p > 0.4745; Fig. 6H).

We also examined if ELS animals presented ictal activity during

recordings. We found that 44.45% of the ELS animals presented electrographic

discharges similar to spike-wave discharges (SWD) or rhythmical spiking (Fig.

6H-I). SWDs occurred mainly during NREM sleep and less often in QWK and

ACT but rarely occurred during REM epochs (Fig 6J-L). The SWDs were not

found in the CTRL rats, indicating that ELS induces an epileptogenic process.

Increased HPC theta power during behavioral activity in ELS animals

We initially concentrated our investigation on the ACT behavioral state to

associate oscillatory patterns with the behavioral abnormalities observed. We

found that ELS animals presented a higher theta power during ACT states in the

HPC (*t*-test, $t_{(13)}$ = 2.259, p = 0.0417; Fig. 7A-D), but not in the PFC (*t*-test, $t_{(13)}$ =

0.2636, p = 0.7961; Fig. 7B). This increased theta activity is more prominent

during the first periods of recording and exploration of the new environment

(linear mixed-effects model, t = 2.10, p = 0.055; Uncorrected *t*-tests for period

comparison: $t_{(13)}$ = 2.145, 2.367, and 2.319, p = 0.051, 0.034, and 0.037 for

periods 1, 2, and 3 respectively; Fig. 7E). To further explore whether theta power

increase reflects a true oscillation and is not due to alterations in the aperiodic

components of the PSD, we also performed a PSD parametrization to separate

the theta oscillation (Fig. 7F). We found an increase in the theta oscillatory activity

in ELS animals during ACT states, indicating an actual oscillatory effect (Mann-

Whitney, U = 7, n = (6,9), p = 0.0176; Fig. 7G-H). Hippocampal theta is well

known to occur during periods of locomotor activity, e.g., postural changes, head

movements, and rearing (Buzsáki and Moser, 2013) representing a possible

confounder. To control for this, we co-examined HPC theta power and EMG distributions during active states in CTRL *vs.* ELS rats (Fig. 7I). Indeed, theta oscillatory increase in ELS animals did not appear to be related to more movement, as we did not observe a significant association between EMG and theta power. In contrast, we found higher theta power in ELS animals even at low levels of EMG activity (Fig. 7I).

ELS promotes abnormal gamma coordination in the PFC during behavioral activity

Given the well-documented PFC gamma alterations in animal models of psychosis (Sigurdsson, 2016), we investigated the possible effects of ELS on PFC gamma activity during active states. ELS animals showed a reduced theta-gamma amplitude correlation during ACT states (*t*-test, $t_{(13)}$ = 2.851, p = 0.0136; Fig. 8A-C), indicating that high-gamma activity is less coordinated, despite the increase in theta power. Interestingly, there were no increases in both low or high-gamma power (low-gamma: Mann-Whitney, U = 16, n = (6,9), p = 0.2238; high-gamma: Mann-Whitney, U = 24, n = (6,9), p = 0.7756; Fig. 8D). Amplitude correlation is an indicator of large-scale cortical interactions that govern cognition (Siegel et al., 2012). Thus, the lower theta-high gamma correlations found here may be related to the cognitive disruptions caused by ELS.

Next, we found a reduction in the PFC spectral exponent in ELS animals accessed by aperiodic PFC activity during ACT state (Fig. 8E), distribution of the aperiodic component (exponent and offset, Fig 8.F), and comparison in exponent parameter between groups (*t*-test, $t_{(13)}$ = 2.010, p = 0.0656; Fig. 8G). Reductions of the spectral exponent have been discussed to indicate increased excitation/inhibition imbalance (Gao et al., 2017). This abnormal gamma dynamics in the ELS indicates a less coordinated oscillatory activity, which may reflect impaired limbic-cortical or cortico-cortical interactions that underlie cognitive dysfunctions.

ELS impairs HPC-PFC coherence during REM sleep

HPC-PFC connectivity during REM sleep is essential for memory consolidation (Lesting et al., 2013). Because we found cognitive impairments in ELS animals, we investigated HPC-PFC interactions during REM. First, we

observed that the theta power in the PFC and HPC during REM is not different between groups (PFC: *t*-test, $t_{(13)}$ = 1.320, *p* = 0.2096; HPC: *t*-test, $t_{(13)}$ = 0.3557, *p* = 0.7278; Fig. 9A-B). However, we found that ELS rats show reduced HPC-PFC connectivity in the theta band, measured by spectral coherence during REM sleep (*t*-test, $t_{(13)}$ = 2.183, *p* = 0.0480; Fig. 9C-D). This difference is present throughout the entire recording, as observed by the comparisons for all 6-hour periods (linear mixed-effects model, *t* = -2.27, *p* = 0.0408; Fig. 9D, right). We further confirmed this difference by analyzing the cross-correlation and cross-spectrum power of the HPC-PFC LFP during REM (Fig. 9E-G). Cross-correlation analysis reveals a clear directionality between signals from the HPC to the PFC in both groups that happens markedly at a specific lag (-80 to -20 ms; Fig. 9E), in which ELS rats present the most profound decrease in the correlation peak (*t*-test, $t_{(13)}$ = 2.533, *p* = 0.0250; Fig. 9F-G).

We noticed from inspecting the theta coherence changes between ACT and REM – the two states with the strongest theta coherence – that all CTRL animals exhibited greater theta coherence during REM (paired *t*-test, $t_{(5)}$ = 8.023, *p* = 0.0005; Fig. 9H), while in the ELS group, part of the animals did not present such an increase (paired *t*-test, $t_{(8)}$ = 1.924, *p* = 0.09; Fig. 9H). This dynamic is further illustrated by observing the coherence changes across REM, QWK, and ACT states. While most CTRL rats present an inclined decrease from REM to QWK and a slight increase from QWK to ACT, ELS rats display a flatter curve (Fig. 9I). By quantifying the homogeneity of the distributions of average theta coherence values across states with entropy, we found a significantly higher entropy (more homogeneous) in ELS than CTRL (paired *t*-test, $t_{(13)}$ = 2.695, *p* = 0.0184; Fig. 9JI). These results indicate that ELS promotes impairment in HPC-PFC connectivity during REM sleep and in the regulation of state-dependent dynamics.

ELS rats display REM-like oscillatory dynamics during active behavior

After we noticed that ELS animals show less distinction of some electrophysiological activities between states, particularly REM and ACT, we decided to further characterize the long-term effects of ELS on HPC-PFC state-dependent oscillatory dynamics across the sleep-wake cycle. For that, we constructed a state map based on two spectral power ratios (Gervasoni et al.,

2004; Dzirasa et al., 2006). This state map framework allows consistent characterization of all major brain states, revealing a dynamic global structure produced by the collective activity of forebrain structures (Gervasoni et al., 2004). First, we verified that the state maps reflected other brain and muscle activities that were not directly used for the map construction, such as the concentrated distribution of the PFC delta power during NREM, of the HPC theta/delta ratios in REM, of the EMG power in awake states, and of HPC-PFC theta coherence in both REM and awake states (Fig. 10A). Thus, as expected, our state maps presented three distinct clusters that precisely located REM, NREM, and ACT epochs (Fig. 10B).

Using this approach, a previous study reported an overlap between awake and REM states in a mouse model of schizophrenia (Dzirasa et al., 2006). Because we found ELS-related functional changes specifically during ACT and REM, we sought to investigate the convergence between these two states. Remarkably, we also found that ELS rats showed a significant similarity between the REM and ACT states (Fig. 10B). While in CTRL rats the distributions of Euclidean distances were larger, homogeneous, and symmetrical (Fig. 10C), ELS rats presented highly variable patterns across animals, most of which with skewed distributions concentrated at shorter distances (Fig. 10C). Indeed, the Euclidean distance was significantly reduced in ELS (*t*-test, $t_{(13)}$ = 2.326, *p* = 0.0369; Fig. 10D), suggesting that the spectral features of REM and ACT are more similar in ELS animals.

Interestingly, we observed that this difference in states similarity was stronger in the initial 12 hours of recording, which corresponds to the periods of habituation to the novel environment (*t*-test, for periods 1-2: $t_{(13)}$ = 2.45, *p* = 0.029; for periods 3-4: $t_{(13)}$ = 0.9409, *p* = 0.3638; Fig. 10E).

To further demonstrate the higher similarity between REM and ACT states in ELS animals, we used machine learning algorithms in an attempt to separate these brain states through a *brute-force* approach. We observed that the two algorithms tested had the worst discriminative performance in the ELS rats, as verified by the smaller AUC-ROC (*t*-test, for SVM: $t_{(198)}$ = 99.59, *p* < 0.0001; for random forest: $t_{(198)}$ = 107.6, *p* < 0.0001; Fig. 10F). This worse discriminative performance also arises when using random forest algorithm (which presents the best discriminative performance) on the original whole-spectrum high-

733 dimensional variables (*t*-test, for PFC power: $t_{(198)}$ = 21.34, *p* < 0.0001; for HPC

734 power: $t_{(198)}$ = 58.78, *p* < 0.0001; for coherence: $t_{(198)}$ = 80.82, *p* < 0.0001; for all

735 the spectral estimates combined: $t_{(198)}$ = 39.23, *p* < 0.0001; Fig. 10G). This lack

736 of discriminative performance once again corroborates spectral similarities

737 between REM and ACT states in ELS rats.

739 ELS rats present behavioral and neurochemical sensitivity to dopaminergic

740 activation

REM-like neural oscillations during the awake state were described in

pharmacological and genetic animal models of hyperdopaminergia (Dzirasa et

al., 2006). In this context, we conducted a final experiment to investigate the

susceptibility of ELS rats to dopaminergic activation using apomorphine

(unspecific dopaminergic agonist). We examined behavioral sensitivity by

comparing the locomotion produced by apomorphine in the open field (Fig. 11A).

Our data demonstrate that apomorphine produces a higher increase in

locomotion in ELS rats when compared to CTRL across time blocks (Two-way

RM ANOVA, treatment effect: $F_{(1,27)}$ = 4.260, *p* = 0.0487, Sidak's *post-hoc:* 5

min *p* = 0.0376; Fig. 11B) or between average normalized distances (Mann-

Whitney, *n(CTRL, ELS)* = 14/15, *U* = 32, *p* =0.0009; Fig. 11B inset).

Immediately after the behavioral test, we collected samples of NAc, PFC,

and HPC to quantify neurotransmitters. The NAc was included in this analysis

because it is highly implicated in dopamine sensitivity and exploratory behavior

(Gomes et al., 2020). Our data demonstrate that apomorphine reduces dopamine

in the NAc of ELS rats, with no effects on dopamine turnover, as measured by

DOPAC/DA ratio (Sidak's multiple correction procedure, $t_{(12)}$ = 3.656*, p* = 0.0033;

Fig. 11C). Still, a GLM model implicated NAc dopamine turnover as a significant

predictor of apomorphine-induced hyperlocomotion (*coefficient* = -0.7108*, p* =

0.00297*;* Fig. 11D). Consistently, apomorphine is known to reduce endogenous

DA release and DA metabolism in rodents and humans (Ozaki et al., 1989; de La

Fuente-Fernández et al., 2001). We also observed a reduction in dopamine

turnover in the HPC of ELS rats (Sidak's multiple correction procedure, *U* = 9*, p* =

0.0164; Fig. 11E) but no significant effects in the PFC.

Interestingly, we found that the HPC of ELS rats shows a higher Glu

content and higher Glu/GABA ratio than CTRL animals following apomorphine

injection (Sidak's multiple correction procedure, Glu: $t_{(15)}$ = 2.566, p = 0.0215;

Glu/GABA: U = 9, p = 0.0079; Fig. 12F) indicating possible disturbances in the

excitation/inhibition balance.

HPC-PFC oscillatory dynamics discriminate ELS individuals

After identifying the relevance of specific electrophysiological variables

throughout the study, we sought to investigate if collective patterns of the

electrophysiological covariates across individuals could assemble a

multidimensional oscillatory signature that distinguishes the ELS subjects. Based

on relevance, particularity, and state specificity, for this investigation, we chose:

(1) HPC theta power (Fig. 7H), (2) theta coherence entropy (Fig. 9J), (3) PFC

exponent (Fig. 8G), (4) PFC theta-gamma correlation (Fig. 8C), (5) theta

coherence (Fig. 9D), and (6) state map distance (Fig. 10D). By performing PCA,

we observed that the first two principal components completely discriminated

ELS from CTRL animals (Fig. 12A). This discrimination was further confirmed by

fitting a linear discriminant classifier (traces on Fig. 12A) that achieved a 100%

accuracy using the entire dataset and 80% accuracy on leave-one-out cross-

validation.

Furthermore, unsupervised clustering almost completely discriminated

ELS from CTRL rats dividing all the animals into two major clusters: a

predominantly ELS cluster (81.82% of ELS animals) and a complete CTRL

cluster (100% of CTRL animals; Fig. 12B-C). This data demonstrates that the

electrophysiological variables used here are jointly affected by ELS at the

functional level. In addition, the hierarchical clustering approach evidenced a

progression of ELS-related oscillatory alterations, with a subcluster presenting an

overlap between ELS and CTRL but the other with a more pronounced abnormal

spectral fingerprint (Fig. 12B). Finally, we show that all the single

electrophysiological variables have a high discriminative value (AUC > 0.7) (Fig.

12D), yet, their multivariate linear combination represented by PC1 scores (Fig.

12A) has the best classification performance with an AUC of 1.

Discussion

799 Here we showed that ELS leads to long-term behavioral abnormalities that

800 are well-established translational features of psychosis. Remarkably, these

behaviors occurred independently of neuronal loss. Instead, they correlated with neuronal inflammation, hypersensitive dopaminergic neurotransmission, and aberrant HPC-PFC synaptic plasticity. We also found that ELS rats exhibit brain state-dependent alterations – particularly during active behavior and REM sleep – characterized by increased theta power and impaired coordination of local and long-range activity. In addition, ELS presents an abnormal oscillatory state during active behavior oddly similar to REM sleep. Finally, the collective pattern of oscillatory alterations accurately distinguishes ELS from control animals.

Common pathophysiological mechanisms underlying epileptogenesis, psychosis, and cognitive impairments

Studies usually describe increased locomotion (Kubová and Mareš, 2013) and working memory deficits (Tsai et al., 2012) as consequences of ELS. In addition to confirming these findings, we also found reduced PPI, which is less studied, but also a well-established correlate of positive symptoms in clinical psychosis (Turetsky et al., 2007; Ziermans et al., 2011). Here, we presented a multivariate *behavioral factor* comprising phenotypes of psychosis frequently reported in pharmacological, genetic, and developmental models of schizophrenia (Ruggiero et al., 2017; Sigurdsson, 2016). Furthermore, we presented a *cognitive factor* uncorrelated with the latter but also impaired after ELS. This lack of association between psychotic-like behaviors and cognitive impairments suggests two neurobiological substrates, which are confirmed by our multivariate analysis, indicating the association of LTP to cognitive deficits and neuroinflammation to sensorimotor alterations.

Surprisingly, we found that cognitive impairment was associated with an abnormally increased susceptibility of HPC-PFC circuits to undergo LTP. This appears to be in contrast with previous studies, which reported reduced hippocampal LTP and cognitive deficits after ELS (Chang et al., 2003; O'Leary et al., 2016). However, no hyperexcitability or spontaneous seizures were described in these studies (O'Leary et al., 2016). In turn, Notenboom et al. (2010) used an animal model of febrile ELS and demonstrated increased LTP in HPC slices related to the reduction of seizure threshold, hyperexcitability, and spontaneous seizures later in life (i.e., epileptogenesis) (Dubé et al., 2006; Dubé et al., 2000). Indeed, paroxysmal activity had been shown to acutely increase synaptic efficacy

and hippocampal LTP, thereby reducing the threshold for seizure induction (Ben-Ari and Gho, 1988; Minamiura A' et al., 1996). In this context, the enhanced LTP observed in our data indicates a relationship with epileptogenesis, which concurs with the epileptiform discharges we observed in ELS rats and the increases we observed in the hippocampal Glu/GABA ratio.

We describe an inverted-U relationship between LTP and working memory, suggesting an optimal level of LTP susceptibility is required for adequate working memory performance. Aberrant synaptic plasticity could be a common mechanism between cognitive deficits and epileptogenesis. Hyperexcitable HPC-PFC circuits could facilitate the spread of epileptiform activity and promote alterations in synaptic saturation and excitatory/inhibitory balance, ultimately leading to cognitive deficits. In this sense, the abnormally increased LTP could impair sensory processes during working memory and explain the poor performance we observed in ELS rats. In a neural network with aberrantly enhanced LTP, synaptic plasticity could be biased toward potentiation, resulting in a lower ability to retain information or adapt to the environment (Navakkode et al., 2022). This rationale is supported by several genetic studies, including on risk genes for neuropsychiatric disorders, that described relationships between enhanced LTP and impaired learning (Garcia-Alvarez et al., 2015; Meng et al., 2002; Nuytens et al., 2013; Yokoi et al., 2015). Additionally, enhanced HPC-PFC LTP is found in the methylazoxymethanol acetate (MAM) neurodevelopmental model of schizophrenia, which also presents impaired spatial working memory, reduced PPI, hypersensitivity to psychostimulants, and increased DA neurotransmission (Goto and Grace, 2006). Modulation of the dopaminergic system attenuated the MAM-enhanced LTP, indicating that altered DA neurotransmission could underlie the LTP abnormalities in ELS rats.

We also report an increase in GFAP expression in ELS animals. Inflammatory processes initiated by a brain insult could lead to functional impairments and ultimately psychiatric symptoms (Vezzani et al., 2013). In ELS models, long-term glial activation has been related to behavioral impairments and epileptogenesis (de Oliveira et al., 2008), while treatment with inflammatory inhibitors resulted in reductions of GFAP and cognitive impairments (Abraham et al., 2012; Somera-Molina et al., 2007). High levels of pro-inflammatory substances and neuroinflammation have been documented in both patients with

schizophrenia and animal models (Vezzani et al., 2013). Noteworthy, the HPC of patients with TLE and interictal psychosis present greater GFAP and microglia activation compared to TLE patients with or without depression (Kandratavicius et al., 2015). These findings are in accordance with our analysis describing enhanced GFAP as significantly contributing to psychotic-like behavioral alterations in ELS animals.

In adult SE studies, behavioral abnormalities and cognitive deficits have been positively correlated to neuronal loss and spontaneous seizure frequency (Wolf et al., 2016). It is suggested that neuropsychiatric mechanisms and epileptic excitability spread progressively over time. Indeed, the behavioral impairments in ELS models tend to be more subtle than in animal models of TLE or psychosis. It is possible that the observed neuroinflammation, neuronal dynamics, and neurochemical alterations represent initial epileptogenic pathological mechanisms that could progress to spontaneous seizures, progressive neuronal loss, and more intense behavioral manifestations (Dubé et al., 2006; Kubová and Mareš, 2013).

Dysfunctional patterns of HPC-PFC oscillatory dynamics

Our results demonstrate a locomotion-unrelated increase in HPC theta power during active behavior in ELS animals. Theta oscillations underlie various cognitive processes and are believed to serve as a temporal organizer for multiple functions, such as sensorimotor integration (Buzsáki and Moser, 2013). However, increases in HPC theta power are also observed in altered psychotic-like states, such as following ketamine administration (Caixeta et al., 2013). Increases in theta power are reported in most studies in patients with schizophrenia (Newson and Thiagarajan, 2019) and have been correlated with poor cognitive performance (Curic et al., 2021). Thus, stronger HPC theta power does not necessarily reflect an optimal cognitive state. In fact, here we observed lower theta-gamma correlations in the PFC of ELS animals, suggesting a disconnection between theta power and local coordination of PFC activity. Similarly, the ketamine model of psychosis also presents disrupted gamma coordination in the PFC (Jodo et al., 2005; Lopes-Aguiar et al., 2020). In addition, we also observed a flatter PFC power spectrum in ELS animals compared to the normal tilt of CTRL, which has been discussed to indicate excitation/inhibition imbalance (Gao et al.,

2017). These findings occurring during active behavior could explain the
cognitive-behavioral alterations we observed, at least to some extent.

Desynchronization between limbic-frontal circuits is also reflected by
reduced theta coherence during REM. Genetic and neurodevelopmental models
of schizophrenia exhibit a decrease in HPC-PFC theta synchrony correlated with
poor spatial working memory performance (Sigurdsson, 2016; Sigurdsson and
Duvarci, 2016). We observed impaired theta coherence only during REM sleep,
which is the brain state with the expected higher coherence values, indicating
that ELS animals could not reach or maintain this connectivity at the same
maximum capacity as CTRL animals. The results from oscillatory dynamics in the
HPC-PFC circuit indicate abnormalities in the temporal organization and
interaction of local circuits that can contribute to behavioral alterations and
cognitive deficits.

Our results also demonstrated that during ACT behavior, frontal-limbic
oscillatory dynamics of ELS rats are unusually similar to REM sleep. Importantly,
this finding could not be explained by altered locomotion or sleep architecture.
This aberrant active state was more pronounced in the initial 12 h of recording,
corresponding to the habituation to the novel environment. Dzirasa et al. (2006)
found similar results when recording hyperdopaminergic mice. The authors
described a REM-like awake state in DAT-KO mice and wild-type mice injected
with amphetamine. Similar to our results, the hyperdopaminergic wake state was
marked by a significant increase in hippocampal theta oscillations. Additionally,
treatment with antipsychotics reduced these REM-like hippocampal oscillations
(Dzirasa et al., 2006). Our similar findings on a developmental model resulting in
a psychotic-like phenotype support the relevance of intraindividual REM-wake
similarity as a neural mechanism of psychosis. These results also advance
previous studies, suggesting a shared involvement of dopamine in the impaired
regulation of state-dependent global network dynamics in animal models of
psychosis and epilepsy. Also interestingly, it is speculated that REM-wake
similarity may represent a neural correlate of oneiric intrusion onto awake
consciousness (Dzirasa et al. 2006), which, in the clinical context, could underlie
potential phenomenological manifestations in epilepsy-comorbid psychosis.

Apomorphine sensitivity has been associated with reduced PPI,
hyperlocomotion, and increased NAc dopamine release (Ellenbroek and Cools,

2002). Corroborating our data, neonatal SE promotes amphetamine sensitization and affects both dopaminergic and glutamatergic levels in prefrontal-striatal circuits (Lin et al., 2009). These results point to enhanced dopaminergic neurotransmission as a potential mechanism underlying the functional abnormalities observed in ELS rats.

In conclusion, we describe a comprehensive set of behavioral and neurobiological alterations following ELS that represent common patterns of dysfunctional circuit dynamics between epilepsy and psychosis. A prominent theory about the pathogenesis of schizophrenia links stressful insults during development to increased HPC excitability, which would culminate in the hyperactivation of the mesolimbic system, resulting in behavioral dysfunctions (Gomes et al., 2020). Our results add depth to this theory by finding relationships among (1) epileptogenesis, (2) hyperexcitable HPC, (3) increased DA drive, (4) disrupted information flow in limbic-frontal circuits, and (5) psychosis-relevant behaviors. Given the common pathophysiological and behavioral traits linking epileptogenesis and behavioral-cognitive comorbidities, we emphasize the model's validity to investigate the neurobiological mechanisms of how an injury during development can lead to psychiatric symptoms. Our study also demonstrates the potential of a multilevel strategy combining electrophysiology, behavior, immunohistochemistry, and neurochemistry in revealing integrated multidimensional mechanisms linking epilepsy, psychosis, and cognition.

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Figures and Legends

Figure 1. ELS induction. *A,* Experimental design representing early-life SE induction and monitoring. ***B,*** Representative traces of LFP during baseline, motor seizure induced by pilocarpine, and ELS interruption with diazepam injection (top). Representative LFP from the frontal cortex and HPC throughout the experiment (middle). LFPs are aligned with their respective spectrograms and energy curves (bottom).

Figure 2. ELS induces a psychotic-like behavioral phenotype. *A,* Experimental design representing early-life SE induction, age of the animals during the experiments, and behavioral tests performed. ***B,*** Weight gain after ELS induction. ELS rats showed weight loss only on the ELS induction day. ***C,*** No weight differences between ELS and CTRL during experiments in adulthood, regardless if before food restriction (left) or during food restriction (right). ***D,*** Working memory task on the radial-arm maze. ELS rats showed an increase in working memory errors across sessions (left) without differences in the number of reference errors (right). E, ELS rats showed a similar learning curve when compared to CTRL. ***F,*** Hyperlocomotion of ELS rats in the open field test. ***G,*** Higher acoustic startle of ELS rats. ***H,*** Impaired sensorimotor gating in ELS rats as measured in the PPI test with three stimulus intensities. ***I,*** Correlation matrix of all behavioral variables. Note a cluster of positive correlations formed by the PPI measures and another cluster of negative correlations formed by the PPI measures with distance traveled on the open field and the acoustic startle. Working memory errors were not correlated with the other behavioral features. ***J,*** Explained variance from principal components analysis (PCA). Note that the two first PC explain 71.32% of the data. ***K,*** Loadings of the behavioral variables on each principal component (PC) reveal a *psychotic factor* (PC1) and an uncorrelated *cognitive factor* (PC2). ***L,*** PCA of the behavioral features showing each animal projected onto the new factors. ***M,*** The psychotic factor distinguishes ELS from CTRL animals, as we compare the PC scores from each group. † p < 0.1, *p < 0.05, **p < 0.01, ***p < 0.001. Error bars represent the mean ± SEM.

Figure 3. Aberrant HPC-PFC synaptic plasticity in animals submitted to ELS. *A,* Experimental design including high-frequency stimulation (HFS) to LTP induction. *B,* Electrode placement and representative electrolytic lesions in Nissl-stained coronal sections. *C,* Representative evoked fPSPs in the PFC. No difference between ELS and CTRL rats in terms of basal neurotransmission and synaptic efficacy, as shown by: *(D)*, fPSP amplitude (left) and slope (right); *(E),* input-output curve; and *(F),* paired-pulse facilitation (PPF) curve. *G,* Despite no changes in basal synaptic efficacy, ELS rats showed aberrantly increased LTP, represented by higher fPSP amplitude ratios from baseline (BL) post-HFS, both across time blocks (main curve graph) and their averages (bar graph, inset). *H,* As in *G,* but from fPSP slopes. *I,* ELS rats showed reduced PPF in the initial 30 min (curve graph). Averaged data resulted in no differences between the groups (bar graph, inset). † p < 0.1, *p < 0.05, **p < 0.01, ***p < 0.001. Error bars represent the mean ± SEM.

Figure 4. ELS induces long-term neural inflammation, but not neuronal loss. *A* 3-D brain diagram and coronal sections representing the brain regions investigated. *B-E*, Representative photomicrographs from immunohistochemistry each pair of columns represents a group comparison (left CTRL and right ELS) and each line is related to CA1 and PL, respectively. *B,* NeuN immunohistochemistry showing that ELS rats did not present neuronal loss. *C,* Parvalbumin (PV) immunohistochemistry. *D,* mGluR5: *E*, GFAP, showing increased astrogliosis induced by ELS. GL: Granule layer of dentate gyrus; CA1: *Cornu Ammonis* 1; EC: Entorhinal cortex; IL: Infralimbic cortex; PL: Prelimbic cortex; TRN: Thalamic Reticular Nucleus. *p < 0.05 Error bars represent the mean ± SEM.

Figure 5. Cognitive impairment and psychotic-like behavior are distinctly correlated with abnormal LTP and neuroinflammation. *A,* Steps performed in data processing and analysis for canonical correlation analysis (CCA). *B,* CCA analysis showing CTRL and ELS animals projected onto the first canonical correlation. We found a strong correlation between the neurobiology canonical dimension and the behavior canonical dimension. *C,* Canonical scores showing the clear CTRL *vs.* ELS difference that the CCA was able to capture. *D,* Coefficients for behavior and neurobiology canonical dimensions, showing greater contributions of the psychotic factor, GFAP, PV, and LTP for the correlation. *E,* Loadings of the behavioral and neurobiological variables on each canonical correlation dimension indicating an association between GFAP and PV with the psychotic behavioral factor. *F,* Correlation matrix showing only weak correlations between the neurobiological features. *G,* GLM regression between neurobiological features and the psychotic factor. *H*, GLM model between neurobiological features and the psychotic factor show a strong association with GFAP, PV, and LTP. *I,* GLM regression between LTP and the cognitive factor shows a quadratic association (left). This quadratic association was further confirmed performing a quasi-Poisson regression between LTP and raw values of working memory errors (right). **p < 0.01. Error bars represent the mean ± SEM.

Figure 6. Classification of sleep-wake states and electrographic discharges in ELS rats. *A,*
Electrode placement and representative electrolytic lesions in Nissl-stained coronal sections. ***B,***
Representative spectrogram, electromyogram (EMG) root mean squared (RMS), and HPC
theta/delta ratio used to score the sleep-wake cycle and to generate the hypnogram (bottom). ***C,***
Representative LFPs (top) recorded from PFC and HPC during ACT, NREM, REM, and QWK. ***D,***
Average power spectral density for each state in the PFC and HPC of a representative CTRL
animal. Note the increase in HPC theta power during ACT and REM. ELS rats did not present
alterations in the sleep-wake cycle. ***E-G***, There were no differences between ELS and CTRL in
the ***(E)*** Mean duration of consecutive epochs for each state, ***(F)*** percentage of time awake across
6-h blocks, and in the ***(G)*** number of persistent episodes (episodes lasting longer than 3 s, top)
or the percentage of time spent in each state (bottom). ***H,*** Representative LFP from PFC and
HPC showing a representative spike-wave discharge (SWD, top) and a poly-spike discharge
(bottom). ***I,*** Representative LFPs with clear electrographic discharges of two ELS rats (top and
bottom). Electrographic discharges were found in 44.5% of ELS animals and were absent in CTRL
animals. ***J,*** Percentage of events occurring in each sleep-wake stage. ***K,*** Average total number
of SWD events in CTRL and ELS animals. ***L,*** Frequency of SWDs occurring per state. SWDs
events rarely occurred during REM. Error bars represent the mean ± SEM.

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Figure 7. Increased HPC theta power during behavioral activity in ELS animals. *A,*
Representative raw LFPs (transparent black lines) and theta-filtered signal of CTRL (black) and
ELS (orange) from the PFC (left) and HPC (right) during ACT. *B,* Relative PSD of the PFC during
ACT. *C,* Relative PSD of the HPC during ACT. *D,* ELS rats present an enhanced HPC theta power
during ACT. *E,* Theta relative power during each of the 6-hour periods of recording, showing a
general increase in the ELS animals that is more pronounced in the initial 18 h of recording. *F,*
Parametric model used to distinguish oscillatory from aperiodic component. *G,* Oscillatory
components extracted from ACT PSDs used to calculate theta power. *H,* Theta power increase
in ELS rats is due to a true oscillatory component and not from the aperiodic activity in ACT states.
I, Theta power increase in ELS rats is independent of locomotor activity. Note from the distribution
of relative theta power and Z-scored EMG (trapezius) values for all rats that EMG power is not
related to the regions of greater theta power in the ELS animals. *p < 0.05. Error bars represent
the mean ± SEM.

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Figure 8. ELS promotes abnormal gamma coordination in the PFC during behavioral activity. ELS rats present an abnormal gamma activity in the PFC. *A,* Grand-average amplitude correlogram for CTRL (left) and ELS (right) group. Note the absence of amplitude correlation between theta and gamma activity in the ELS group. *B,* High-gamma (65-100 Hz) correlation with low-frequency activity. Note the peak around theta oscillation in the CTRL group and the absence in ELS. *C,* ELS present an impaired theta-gamma amplitude correlation. *D,* There is no difference between the low- (top) and high-gamma (bottom) average power. *E,* Aperiodic activity extracted from the PFC of each rat during ACT state. *F,* Distribution of the parameters of the aperiodic component (exponent and offset) for each rat. *G,* ELS rats show a tendency to reduction (*p* = 0.0656) in the exponent parameter of the aperiodic component. † p < 0.1, *p < 0.05. Error bars represent the mean ± SEM.

Figure 9. ELS impairs HPC-PFC coherence during REM sleep. *A,* PSD in the PFC during REM. ***B,*** PSD in the HPC during REM. No significant alterations in theta power during REM are observed in ELS rats. ***C,*** Theta coherence is diminished in ELS rats. ***D,*** Significant effect on the HPC-PFC theta coherence by averaging the entire recording (left) and across all periods (right). ***E,*** Time domain cross-correlation between PFC and HPC LFPs during REM epochs highlighting CTRL (left) and ELS (right) group averages. ***F,*** Reduction in the peak of correlation between -80 to -20 ms in ELS rats*. **G,*** Average cross-spectrum power density showing a reduced theta peak in the ELS rats. ***H,*** Theta HPC-PFC coherence increases from ACT to REM states. We can observe that all the CTRL rats present an increase from ACT to REM, while in ELS rats this is not observed. Paired *t*-test *p < 0.05. ***I,*** Theta coherence dynamics across the sleep-wake cycle. CTRL rats display a homogeneous dynamics with theta coherence being maximal at REM, minimal in QWK, and intermediate in ACT states. This dynamics is disrupted in ELS rats that display a higher variance between animals, with some animals displaying similar coherence in REM and ACT. ***J,*** Entropy of the theta coherence dynamics is higher in ELS rats, representing the more homogeneous values across states.

Figure 10. ELS rats display REM-like oscillatory dynamics during active behavior. *A,* State map construction reflects the sleep-wake cycle and spectral dynamics for one representative CTRL (top) and ELS (bottom) rat. PFC delta power is evident during NREM, theta/delta ratio is maximum in REM, EMG power describes ACT states, and HPC-PFC theta coherence is stronger close to REM. Note that in the ELS rat, the coherence is also broadly high during the ACT state. ***B,*** Representative state maps from three CTRL (top) and three ELS (bottom) rats. CTRL rats displayed a clear separation between REM (red), ACT (light green), and NREM (blue) clusters. ELS displayed an overlap between REM and ACT states. ***C,*** Distributions of the Euclidean distances between each REM and ACT epoch for CTRL (top) and ELS (bottom) rats. ELS distributions were heterogeneous, some of which were skewed. ***D,*** Average Euclidean distance showing that ELS rats have REM-like oscillatory dynamics during active behavior. ***E,*** REM-like oscillations during ACT were more pronounced in ELS during the initial 12 h of recording. ***F,*** Machine learning approach to decode REM *versus* ACT states. We used radial basis function

support vector machine (SVM) and random forest (RF) algorithm in order to classify REM and ACT epochs in CTRL (black) or ELS (orange) rats. Using a bootstrapped confidence interval, we observed that the discriminative performance was worse in ELS animals, even when using "brute-force" algorithms. *G,* A machine learning algorithm (random forest) fitted on whole-spectrum data also shows a significantly worse performance in ELS for PFC power, HPC power, coherence, and all estimates combined. † p < 0.1, *p < 0.05, **p < 0.01, ***p < 0.001. Error bars represent the mean ± SEM.

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Figure 11. ELS rats present behavioral and neurochemical sensitivity to dopaminergic

activation. *A,* Experimental design representing quantification of locomotion in the open field,

apomorphine (APO) injection, and neurochemical analysis. ***B,*** Normalized locomotion across time

blocks after APO injection, demonstrating increased locomotion of ELS rats induced by APO.

Average normalized locomotion after APO injection (inset). ***C,*** Concentration of dopamine and

metabolites in the NAc. ***D,*** GLM for the total distance after APO indicating DOPAC/DA ratio in the

NAc as a strong significant variable associated with behavioral sensitivity to APO. ***E,***

Concentration of dopamine and metabolites (left) in the PFC (upper panel) and HPC (lower

panel). Concentration of glutamate and GABA (right) in the PFC (upper panel) and HPC (lower

panel). †Uncorrected $p < 0.05$, * multiple comparison corrected $p < 0.05$. Error bars represent the

mean ± SEM.

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Figure 12. HPC and PFC oscillatory dynamics discriminate ELS rats. *A,* A linear discriminant

classifier (dotted orange line) using the first two principal components (PCs) of relevant

electrophysiological variables completely distinguishes ELS from CTRL rats (100% accuracy;

93.34% leave-one-out cross-validation). Note that the PC1 alone completely discriminates ELS

rats. ***B,*** Unsupervised hierarchical clustering using the electrophysiological variables (bottom

dendrogram) almost completely dissociates ELS from CTRL rats (left dendrogram). Inside the

ELS cluster, we can observe one complete ELS branch, but the other overlapping with some

CTRL animals, suggesting a graduality in the pathophysiological effects of ELS. ***C,*** Cluster

centroids of the ELS and CTRL clusters showing distinct patterns of activity for each cluster. ***D,***

All the electrophysiological variables are good discriminators of CTRL and ELS rats (AUC-ROC

> 0.7). † p < 0.1, *p < 0.05, **p < 0.01, ***p < 0.001. Error bars represent the mean ± SEM.

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