Functional genomic mechanisms of opioid action and opioid use disorder: a systematic review of animal models and human studies

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Abstract

In the past two decades, over-prescription of opioids for pain management has driven a steep increase in opioid use disorder (OUD) and death by overdose, exerting a dramatic toll on western countries. OUD is a chronic relapsing disease associated with a lifetime struggle to control drug consumption, suggesting that opioids trigger long-lasting brain adaptations, notably through functional genomic and epigenomic mechanisms. Current understanding of these processes, however, remain scarce, and have not been previously reviewed systematically. To do so, the goal of the present work was to synthesize current knowledge on genome-wide transcriptomic and epigenetic mechanisms of opioid action, in primate and rodent species. Using a prospectively registered methodology, comprehensive literature searches were completed in PubMed, Embase, and Web of Science. Of the 2705 articles identified, 69 met our inclusion criteria and were considered for qualitative analysis. Focusing on the 5 most studied nervous system structures (nucleus accumbens, frontal cortex, whole striatum, dorsal striatum, spinal cord; 44 articles), we also conducted a quantitative analysis of differentially expressed genes, in an effort to identify a putative core transcriptional signature of opioids. Only one gene, Cdkn1a, was consistently identified in eleven studies, and globally, our results unveil surprisingly low consistency across published work, even when considering most recent single-cell approaches. Analysis of putative sources of variability detected significant contributions from species, brain structure, duration of opioid exposure, strain, time-point of analysis, and batch effects, but not type of opioid. To go beyond those limitations, we leveraged threshold-free methods to illustrate how genome-wide comparisons may generate new findings and hypotheses. Finally, we discuss current methodological development in the field, and their implication for future research and, ultimately, better care.

Keywords

Opioid, substance use disorder, genomics, epigenomics, next generation sequencing

Introduction

Opioids have been used for millennia for their analgesic and euphoric properties[1]. While they remain reference pain treatments, their chronic use also associates with tolerance, physical dependence and, in some individuals, the emergence of an opioid use disorder (OUD). OUD is defined as a problematic use leading to significant impairment. This severe and chronic disorder associates with a life expectancy decreased by more than 10 years due to somatic and psychiatric comorbidities. Over the last 2 decades, western countries have faced an increase in death by opioid overdose, due to more frequent prescription for pain management, increasing use of illegal compounds, or misuse of substitution therapies. OUD imposes a major socio-economic burden, with an estimated annual cost in the trillion dollar range in the US[2], and increasing use and harm in Europe[3]. This results, overall, in a worsening public health crisis.

OUD results from a variety of interacting biological, psychological and socio-economic factors[4]. At biological level, it originates from pharmacological effects of opioid drugs, which trigger chemical and molecular brain adaptations, under modulation by genetic vulnerability and epigenetic regulation. In turn, these effects mediate behavioral and cognitive dysfunctions, including the inability to control drug use despite harmful consequences, or persistent and intense desire for the drug, even after years of abstinence.

Since the 1990s, high-throughput approaches have been harnessed to characterize the molecular underpinnings of opioid effects in brain tissue, throughout the full genome[5], across various experimental models and species. In this context, the goal of the present systematic review was to synthesize current knowledge on functional genomic mechanisms of opioid action and OUD, defined as changes in gene expression or epigenetic regulation. To do so, we used a preregistered methodology, and performed an unbiased survey of bibliographic repositories, focusing on high-throughput studies, most notably microarrays and next-generation sequencing. While we acknowledge the important contribution of candidate gene studies (already reviewed in the past, see[6, 7]), this focus on genome-wide analyses

reflects the conviction that the understanding of heterogeneous phenotypes such as OUD may

benefit from analyzing the full genome.

Based on studies identified through this screening, a qualitative synthesis was first

performed to summarize experimental designs and findings. Second, a focused quantitative

analysis of bulk tissue transcriptomic studies was conducted, with the goal of defining a core

signature of opioids. To our knowledge, while numerous reviews are regularly published in the

opioid field[8–12], such a systematic synthesis across brain structures and technologies had

never been performed. Our results indicate that, although more than 40 genome-wide studies

have been published, available evidence for convergent findings is surprisingly limited. Third,

we reviewed most recent work that leveraged cell-type specific and single-cell approaches, or

that interrogated epigenetic regulatory mechanisms contributing to opioid plasticity. Finally,

we discuss current challenges in the field, as well as avenues and recommendations for future

work.

Methods

Full methodology and code are available as Supplementary Material and File.

Results

The identification and selection of eligible studies is presented in Fig.1. For qualitative

synthesis, 69 papers were selected, covering both transcriptomic and epigenomic

approaches. Among these, the 44 articles that performed transcriptomic analyses in the 5

most frequently investigated regions were selected for more detailed quantitative synthesis:

of these, 34 investigated a single brain region, 9 covered 2 regions, and only 1 characterized

5 regions[13]. This resulted in 52 differential expression analyses: 18, 11, 11, 6 and 6 in the

nucleus accumbens (NAc), frontal cortex, whole striatum, dorsal striatum and spinal cord,

respectively.

1. Qualitative analysis (n=69 studies)

Species and sex ratio. Most studies were conducted in rodents (Fig.2A): 30 (43%) in rats,

28 (41%) in mice, and 1 in both species. Only a small number investigated humans (n=8, 12%)

and macaques (n=2, 3%), reflecting well-known practical limitations. As consequence, current

knowledge on conserved or species-specific opioid effects is limited.

Regarding sex, epidemiological surveys indicate that OUD prevalence is higher in

males than in females[3, 14] (although smaller differences have been reported with DSM-5

criteria[15]), with a number of sex-specific risk factors (e.g., ethnicity, income). This clinical

situation has long been modeled in rodents, as sex differences in behavioral effects of

opioids[16] have been described for analgesia[17], tolerance[18], withdrawal symptoms[19],

reward[20] or motivation[21]. In comparison, molecular mechanisms underlying these

differences are poorly characterized. The majority of OUD studies reviewed here investigated

males only (n=48, 70%; Fig.2B-D), with few conducted in females (n=5, 7%), and only 14

(20%) that included both sexes (7 in rodents, 7 in humans). These numbers are consistent

with the current landscape of neuroscience research, in which the ratio of articles reporting on

males only, as opposed to females only, is around 5 to 1[22]. We nevertheless note an

encouraging trend, as the large majority of studies published in 2021 (8/10) investigated both

sexes. Several factors likely underlie this evolution: policies from funding agencies (such as

the NIH; https://grants.nih.gov/grants/guide/notice-files/not-od-15-102.html); increased

recognition that females do not display more experimental variability than males[23]; and

evidence that pathophysiological mechanisms may differ across sexes, as shown for

depression[24], schizophrenia[25] or alcohol use disorder[26].

Specifically in rodents (Fig.2C), 80% of studies investigated males only (n=47). Seven

articles investigated both sexes (n=7, 12%), using surprisingly low sample sizes (n=1-4 in

male/female, and opioid-treated/control, groups; Fig.2D). Among these, 2 studies treated sex

as a covariate[27, 28], and 3 pooled both sexes for the analysis of opioid effects[29–31],

therefore providing no direct description of male/female differences. This is concerning,

considering that the 2 remaining studies found little overlap, from 5 to 35% (see[32] and[33]),

among morphine-induced changes occurring in each sex. Therefore, available evidence

points toward substantial sex-specificity in opioid-induced transcriptomic plasticity, which will have to be confirmed in future work. In human, even less is known (Fig.2E). Among 8 articles, 7 investigated males and females, while only 1 investigated males only. However, these 7 studies showed a strong bias towards examination of men (Fig.2F), with only one that used equal sample sizes in both sexes[34]. In the latter, sex was treated as a covariate, with no specific description of its impact. Altogether, this significant gap in the literature calls for additional work, with the hope that better understanding of sex-specific pathophysiological routes may unravel more efficient therapeutic strategies.

Route and duration of opioid administration. Among studies included in qualitative analysis (n=69), the majority used chronic administration (n=55, 80%; see Fig.3A), mostly via self-administration (SA, n=19, 34%, including 8 human studies) and intraperitoneal injection (ip; n=18, 33%), followed by subcutaneous injection (n=8, 15%). Other procedures were less frequently used: continuous administration (with pellets or mini-pumps; n=5, 9%), intrathecal administration (n=3, 5%), or gavage (n=2, 4%).

There is limited information regarding differences among administration procedures. Interestingly, Lefevre et al[35] recently compared 3 conditions: continuous and systemic morphine infusion using osmotic mini-pumps, with ("Interrupted") or without ("Continuous") the induction, twice daily, of a precipitated withdrawal episode (using naloxone, a non-specific opioid antagonist), compared to Controls (saline mini-pumps). Results indicated 687 and 407 differentially expressed genes (DEG) when comparing Interrupted and Control groups in the NAc and dorsal striatum, respectively, with only 1 DEG between Continuous and Control groups. Therefore, rather than opioid exposure per se, it is the repetition of intermittent withdrawal episodes that may drive the intensity of transcriptomic adaptations affecting the mesolimbic system (whether precipitated, as in this study, or spontaneous, in-between bolus injections). This is consistent with the clinical notion that the alternation of intoxication and withdrawal phases is critical for the development of OUD.

Opioid effects have also been investigated using acute administration (10 studies,

14%), corresponding mostly to ip injection (9 articles), with only 1 using subcutaneous

injection. While these studies were mainly interested in the use of opioids as analgesics or

anesthetics[36, 37], or sought to compare acute effects of various psychoactive drugs[38],

their results may be relevant for the understanding of OUD. Indeed, comparisons of inbred

mouse strains found striking variability in the intensity of opioid physical dependence, with the

same strains showing severe or mild signs of withdrawal when precipitated after acute or

chronic opioid injections[39, 40]. In other words, there may be shared genetic determinants,

within each strain, for brain adaptations triggered by acute or chronic exposure. A recent

single-cell RNA-sequencing study (scRNA-Seq; see *3. High cellular resolution approaches*

and [41]) provides an interesting illustration of how such strain-specific genetic traits may

translate into at least partially similar transcriptional plasticity during acute or chronic morphine

exposure.

Another important consideration is that most studies (51/69, 73%) used passive

administration, which do not model volitional or compulsive drug-taking. This reflects the

technical difficulty of intravenous SA experiments (IVSA), particularly in the mouse. Few

studies have conducted direct comparison of passive and active administrations. Among

these, Tapocik et al[42] compared "yoker" mice (IVSA group) with groups of "yoked-morphine"

and "yoked-saline" animals which, each time the yoker self-administered morphine, received

passive IV morphine or saline injections. Results significantly differed depending on the mouse

strain used. In C57BL/6J, a much higher number of DEG were identified between the yoker

and yoked-saline groups (n=1176) than between yoked-morphine and yoked-saline mice

(n=244), with limited overlap between the 2 (n=103). In DBA/2J mice that did not self-

administer morphine (no behavioral differences among the 3 same groups), transcriptional

effects associated with operant responding were milder than in C57BL/6J (n=107 DEG

between yoker and yoked-saline groups), as expected, while again a modest number of DEG

between yoked-morphine and yoked-saline groups (n=262, likely corresponding to

pharmacological morphine response). This suggests that distinct transcriptional adaptations,

partly dependent on genetic background, may underlie voluntary drug-taking or reflect non-

contingent passive pharmacological effects, and together contribute to OUD.

New paradigms have been developed to facilitate such studies, which notably include

oral SA[43–45]), or devices for opioid vapor SA[46]. In addition, optogenetic tools now enable

light-induced self-stimulation (optogenetic intracranial self-stimulation, oICSS)[47]. The latter

models are based on selective manipulation, using opsins, of the activity of anatomically- or

genetically-defined neuronal populations. While these approaches do not recapitulate the

systemic effects of opioid IVSA, they are nevertheless able to recapitulate similar behavioral

deficits, including uncontrolled consumption, resistance to punishment, or excessive

motivation[48, 49]. It is therefore possible that they may allow dissecting the neuronal

pathways that are necessary or sufficient for each of these behavioral dimensions, as well as

underlying molecular mechanisms. Another avenue for improvement relates to the mode of

action of genetic tools used for oICSS. Upon light-stimulation, opsins open ion channels that

modulate neuronal excitability. Their high temporal resolution does not necessarily represent

the best model for the longer time-frame of pharmacological action of drugs of abuse. To more

faithfully mimic such kinetics and associated intracellular signaling, chemogenetics or chimeric

opsins (for light-induced metabotropic signaling, with a proof-of-concept available for

MOR[50]) might represent better tools.

Nervous system structures. Among articles eligible for qualitative analysis, 18 regions of the

nervous system were explored (Fig.2B). Not surprisingly, a large majority focused on the

mesocorticolimbic dopaminergic pathway: the NAc (21 articles[13, 27, 30, 33–35, 37, 51–64]),

frontal cortex (13 ; [13, 29, 32, 34, 62, 65–72]), whole/unspecified striatal complex (11 ; [31,

38, 73–81]), dorsal striatum (7 ; [13, 35, 59–61, 82, 83]) and ventral tegmental area (VTA, 5 ;

[33, 62, 84–86]). Other regions included the spinal cord (7 ; [73, 87–92]); hippocampus (5 ;

[13, 74, 93–95]); amygdala (3 ; [13, 96, 97]); locus coeruleus[85, 98], ventral midbrain[42, 99],

hypothalamus[100, 101] (2 each), whole brain with[102] or without[103] cerebellum,

periaqueductal gray matter[104], pituitary gland[101], arcuate nucleus[105], nucleus of the

tractus solitarius[36], brainstem[28], cerebellum[95] or dorsal root ganglia[106] (DRG, 1 each).

Below, we briefly describe the rationale for studying such diverse structures.

The course of OUD involves 3 stages: intoxication, withdrawal/negative affect, and

anticipation of next intoxication. These stages reflect gradual adaptation of the brain to drug

exposure, and rely on distinct mechanisms. The intoxication stage, mostly driven by acute

reward, notably associates with dopamine release, in the striatum and frontal cortex, by

neurons located in the VTA. The second stage corresponds to negative reinforcement,

whereby drug consumption alleviates physical and negative affective states of withdrawal,

involving the locus coeruleus[107], NAc[108], and amygdala[109], among others. Finally,

anticipation results from memories of drug-associated cues and contexts (hippocampus), as

well as impaired goal-directed behaviors, implicating higher-order structures (e.g. prefrontal

and orbital cortices[110]). These relationships are not exclusive, with some structures

implicated at multiple stages (e.g., the striatum, also involved in habit formation). Other regions

(spinal cord, DRG, brainstem, periaqueductal gray matter, PAG) were investigated in relation

to opioid analgesia, tolerance and hyperalgesia. Finally, few articles were interested in how

opioids modulate neuroendocrine systems (hypothalamus, arcuate nucleus, pituitary gland).

In addition to these associations, it is also important to consider the distribution of the

endogenous opioid system. This system is composed of 3 opioid receptors (mu, delta and

kappa; MOR, DOR and KOR), among which the MOR is necessary for opioid-induced

analgesia, reward, and physical dependence[111]. Neurons expressing this receptor therefore

represent pharmacological entry points for brain adaptations to opioids. Comparison of studies

reviewed here with MOR brain distribution (assessed using ligand autoradiography[112–117];

Fig.3C) indicates that: i) MOR is highly expressed among structures most frequently studied,

as expected; ii) surprisingly, other areas with similarly high expression have received little to

no attention (NTS, locus coeruleus, dorsal raphe nucleus, medial habenula, thalamus), calling

for more work.

Techniques. Thirty-seven and 29 studies used microarrays and RNA-seq, respectively, to analyze the transcriptome (Fig.3D). While the first RNA-seq study was published in 2014[37], this technique has since progressively replaced microarrays, reflecting well-recognized advantages (higher sensitivity and throughput, gene discovery). For epigenomic profiling, 3 studies recently performed bisulfite sequencing[62, 70, 72], and 1 used ATACseq (Assay for Transposase-Accessible Chromatin with sequencing)[83]. Considering rapid progress in sequencing methodologies (see below), these trends are expected to strengthen.

2. Quantitative analysis of transcriptomics studies

To identify reproducible transcriptomic opioid signatures, we ran quantitative analyses of bulk-tissue results available for the 5 most studied structures (44 articles; Fig.4A). To do so, gene-level results were retrieved when available. This yielded 24 lists of DEG comparing opioid-treated animals and control groups, originating from 17 articles. Among these, 6 articles did not report any p-values, while the remaining 11 used very diverse thresholds on p-values and fold changes (FC; Supplementary Table 1). Only a small minority (3/44, 6.8%[31, 33, 34]) provided p-values and FC for all genes. In addition, raw data were available in the public repository GEO in less than half of studies reporting DEG (8/17, 47%). Overall, this underscores strong heterogeneity in data analysis and reporting, which limits re-use[118]. To counter this, efforts have been made to harmonize practices in the field[119, 120], with guidelines enforced by funding agencies[121]. For the present review, a genome-wide meta-analysis combining, for every gene, effect and sample sizes across studies was not possible. We nevertheless: i) Systematically assessed concordance among available lists of DEG; ii) Conducted a threshold-free comparison of the 2 NAc studies for which genome-wide results were available.

2.1 Nucleus accumbens

Among 18 articles, only 8 provided lists of DEGs: 4 in mice (males only[35, 37, 51, 54]), 2 in rats (1 in males[56], 1 in both sexes[33]), 2 in humans (both sexes[34, 58]). The 8 articles

provided a total of 3502 DEG (Fig.4B-E). Among these, the vast majority were identified by 1 study only (n=3017/3502, 86%), while 424, 57 and 4 DEG were identified across 2, 3 or 4 studies, respectively. No single genes were common to 5 or more articles. In addition, the best overlap was found among the 2 DEG lists reported by Imperio et al[56] (Fig.3E). Overall, this indicates low concordance among NAc studies.

To define factors contributing to this heterogeneity, we intersected DEG lists in each species separately (Fig.3C-E). This, however, did not improve concordance: in humans, only 54 genes were common to both studies (Jaccard Index, JI=2.4%, Fig.4C, Supplementary Table 2 for the matrix of pair-wise JI across all studies); in mice, 7 were identified by at least 2 studies (JI=1%, Fig. 4D); in rats, the JI increased to 15.5% (Fig.4E), mostly driven by high overlap among the 2 aforementioned DEG lists from Imperio et al. In the latter work, the authors identified large- (LS) and small-suppressors (SS): in LS rats, greater avoidance of a natural reward (saccharine) that predicted heroin availability was associated, compared to SS rats, with worse outcomes during later heroin IVSA (increased consumption, higher motivation and relapse). Gene expression changes occurring in the LS group (compared to control rats given access to saccharine but not heroin; n=597 genes), and in the SS group (same controls, n=205) showed a JI of 25.5% (with striking concordance in up- and down-regulation). This was much higher than overlap between Imperio et al and the other rat study, by Townsend et al (Imperio-SS/Townsend, JI=1.35%; Imperio-LS/Townsend, 3.43%), suggesting that differences across laboratories and animal facilities may significantly contribute to poor concordance. Importantly, while the relatively small number of studies in the NAc precluded definitive conclusions, a more powerful analysis of sources of variability was possible across our 5 regions of interest (see section *2.6*).

2.2 Dorsal striatum

Only 2 dorsal striatum studies provided DEG (Fig.3F). One study used chronic morphine administration interrupted by naloxone injections in male mice[35], as described above, while the other used acute morphine injection in male rats[82]. Reflecting these differences, a very

small overlap was found between the 2 (0.62%). Therefore, while the dorsal striatum is thought

to gradually gain control over drug-taking in OUD[122], underlying transcriptomic plasticity is

poorly characterized.

2.3 Whole striatum

Six of the 11 whole striatum studies provided DEG[31, 38, 75–78] (Fig.5B): 2 investigated

chronic heroin[78] or morphine[31], while 2 compared acute effects of distinct opioids

(morphine and heroin)[38, 76], and 2 compared acute and chronic morphine[75, 77]. Most

DEG were identified by 1 study only (3308/3736, 88.5%), with only 112 genes common to 3

or more studies. Consistent with NAc results, strongest overlaps were observed for lists of

DEG coming from the same article, with for example 54 (JI=17%, Korostynski 2013[38]) and

16 (21%, Piechota 2010[76]) DEG common to acute morphine and heroin injections,

respectively, and 139 DEG (31%, Korostynski 2007 [77]; Fig.5A) common to acute and chronic

morphine administration. In sharp contrast, comparisons across, rather than within, studies

yielded poor overlaps, even when considering those that used similar drug administration:

among chronic morphine studies (Fig.S1A), the JI varied from 0.1 to 2.6% (Korostynski

2007/Skupio 2017), and, among acute morphine studies, from 2.7% to 7.8% (Fig.S1B), with

only one comparison standing out among acute heroin studies (30 common DEG, JI=19%,

Piechota 2010/Korostinsksy 2013). Importantly, 5 of these 6 studies in the whole striatum were

conducted by the same group, suggesting that significant variability may persist even among

studies conducted in the same facility, but at different periods (see also section 2.6).

2.4 Frontal cortex

Seven studies provided DEG (Fig.5B): 2 in humans[34, 69], 5 in rats[29, 32, 66–68]. Data from

Odegaard et al[29] were discarded, as they corresponded to transgenerational effects in

animals not directly exposed to opioids. For 2 additional studies [66, 68], we could not map

microarray probes to genes. Among the 4 remaining articles, little overlaps were again found

(Kuntz-Melcavage/Mendez, JI=0.5%; Kuntz-Melcavage/Seney, 1.1%; Liu/Mendez, 1.4%;

Liu/Seney, 1.6%), with no improvement within species (Liu/Kuntz-Melcavage in rats, JI=0%;

Seney/Mendez in humans, 4.1%).

2.5 Spinal cord

Among the 6 spinal cord studies, 5 did not provide DEG[87–91], while the 6th one used a

homemade microarray[73], for which correspondence between probe IDs and genes was only

partially provided. As such, no comparison was possible.

2.6 Sources of variability

Considering that pharmacological recruitment and intra-cellular signaling of MOR may trigger

a set of common transcriptional adaptations across the 5 regions of interest, we next pooled

all lists of DEGs (Fig.5D). Again, the vast majority of DEG (70.1%, 4940/7047) were unique to

a single study, while only 21.4% common to 2 studies (1509), 5.9% to 3 (418), and 1.7% to 4

or more (117; Supplementary Table 3). Interestingly, Cdkn1a (Cyclin Dependent Kinase

Inhibitor 1A) was identified by 11 studies. This gene encodes for p21, a protein involved in the

regulation of the cell-cycle and oligodendrocytes[123]. Interestingly, Avey et al[51] found

Cdkn1a to be one of the most significantly upregulated genes in response to acute morphine.

This upregulation occurred in oligodendrocytes, and was blocked by the opioid antagonist

naltrexone, indicating a MOR-dependent mechanism. Of note, cocaine, another major drug of

abuse, similarly recruits p21, as cocaine-induced behavioral responses were modified in p21

knockout mice[124]. Using this knock-out line, or virally-mediated approaches for p21

manipulation, represent appealing perspectives to investigate this gene in OUD.

We also performed Gene Ontology (GO) analysis using DEG common to at least 3, 4

or 5 studies (Supplementary Table 4). No results were significant for the small list of 63 DEG

common to 5 or more studies. When considering the 180 DEGs common to at least 4 studies,

only 3 GO terms were enriched: cell periphery, plasma membrane and hormone activity.

Results more directly related to nervous system function emerged for the 598 DEG common

to at least 3 studies, with 46 significant GO terms, mostly related to synaptic signaling and

cell-cell communication, likely reflecting opioid-induced electrophysiological and synaptic adaptations across brain structures.

Because weak DEG overlaps were identified within individual regions (even when grouping studies based on species, Fig.4C-E, or treatment chronicity, Fig.S1), we sought to more systematically assess, across the 5 regions, the impact of following factors: species, brain structure, treatment chronicity, drug, strain, time-point of assessment following last exposure, and article (Fig.5D). For each factor (e.g., species), pair-wise JI were classified in 2 categories depending on whether they belonged to the same sub-group within that factor (2 studies in humans; "intra"), or not (1 human study compared to another in rats; "inter"). As expected, grouping list of DEG by species (p=2.5E-3), brain structures (3.4E-8), treatment chronicity (p=1.4E-3), mouse genetic background (3.9E-5) or the time-point of analysis after opioid exposure (6.7E-10) all increased concordance, with higher JI. This is consistent with the notion that, although they remain poorly characterized, these 5 variables contribute to opioid-induced transcriptional plasticity. In contrast, the type of opioid used had none (p=0.059) when considering the 13 and 5 rodent DEG lists related to morphine or heroin, respectively (human studies were discarded, as the types of opioids consumed are diverse and/or poorly characterized). This is surprising, considering that these 2 MOR agonists, known to exhibit strong pharmacokinetic differences (heroin being a lipophilic prodrug that more rapidly crosses the blood brain barrier to be metabolized in 6-monoacetylmorphine and morphine, which activate MOR), may be expected to generate distinct transcriptomic adaptations. Time-course and dose-response experiments may therefore be necessary to detect such potential differences. Importantly, concordance increased most when DEG lists came from the same publication (p=2.5E-5), despite the fact that only 7 comparisons fell under this category. As already mentioned, this suggests that differences across laboratories may explain a substantial part of the lack of concordance across studies.

Finally, we refined our analysis by considering the directionality (up/down) of DEG, focusing on genes identified by at least 5 studies (n=63). Among results from 17 out of 24 studies that reported on directionality (Fig.S2), most genes (37/63, 59%) showed discordant

results (while, for 8 others, directionality was reported by only 1 or no study). Only 18 genes

showed 100% concordance across studies (29%; 17 up-regulated, 1 down), among which 11

were identified by at least 3 studies. The latter included the aforementioned Cdkn1a, as well

as Slc2a1, Fkbp5, Sult1a1, Arrdc3, Ccdc117, Plin4, Wscd1, Arid5b, Pla2g3, and Tsc22d3.

Slc2a1 encodes for the glutamate transporter GLT1, which plays a major role in extracellular

glutamate uptake and the regulation of excitatory transmission. A recent study showed that

during prolonged morphine withdrawal, activation of KOR in the amygdala leads to increased

GLT1 expression and excitatory drive in the NAc, thereby mediating withdrawal-induced

depressive-like behaviors. Another interesting hit is Fkbp5, a chaperone and intra-cellular

negative regulator of glucocorticoid signaling, that has been largely investigated for its role in

stress-related disorders[125], including OUD[126]. The 8 remaining genes, although

comparatively less studied in relation to opioids (Fig.S2), represent additional candidates for

future work.

2.7 Threshold-free genome-wide comparison

Next, we compared the 2 NAc studies (Seney et al[34], Townsend et al[33]) for which full

genome-wide results were available. While both included males and females, only Townsend

reported sex-specific results. Therefore, we reprocessed Seney's raw data to identify DEG in

each sex separately (*Supplementary Material*). This slightly increased concordance: while JI

were equal to 7.5 and 4% when comparing initial Seney data (male/females pooled) with

Townsend data in females or males, respectively (Fig.S3A-B), they increased to 9.5% and

4.5% when considering Seney's sex-specific results (Fig.6A-B). This argues for the

importance of accounting for sex.

We then used RRHO2 (Rank–Rank Hypergeometric Overlap[127]) to compare the 2

studies. RRHO2 iteratively performs hypergeometric testing for all combinations of thresholds

applied to each dataset, generating a "threshold-free" analysis. Because this approach takes

directionality into account, we also computed, for comparison, overlaps among either up- or

down-regulated DEG reported by each study (white crosses, Fig.6C-D). Compared to those,

much more significant overlaps were identified when considering less stringent p-value

thresholds with RRHO2 (best hypergeometric results shown as black crosses). By definition,

these increased overlaps, came at the expense of the identification of large gene lists which,

although commonly dysregulated in similar direction, individually exhibited milder FC.

Interestingly, these overlaps were detected despite the fact that Seney investigated human

post mortem tissue from individuals with OUD, while Townsend focused on rat tissue collected

after 24 hours of withdrawal from fentanyl IVSA. Also, different patterns of convergence were

observed in each sex: in females, overlaps were highly significant among both up- and down-

regulated genes while, in males, a lower concordance predominantly affected down-regulated

genes. This suggests that the rat dataset may differentially recapitulate part of the sex-specific

signatures of human OUD. Overall, these analyses illustrate the utility of reporting full genome-

wide results which, in combination with threshold-free approaches, generate deeper

comparisons (see *Discussion*).

3. High cellular resolution approaches

Studies reviewed above indicate that opioids induce genome-wide transcriptomic changes,

with striking variability across laboratories, experimental designs, and brain regions. This likely

reflects a combination of: i) direct pharmacological opioid effects, corresponding to the

recruitment of the MOR intracellular signaling, and resulting in decreased neuronal

excitability[128]; ii) indirect effects: depending on the chemical identity and connectivity pattern

of MOR-expressing cells, their inhibition leads to secondary changes affecting other

neurotransmitters[113]. This raises 2 questions: how is each cell type, or even each brain cell,

affected by these changes? What are underlying molecular regulatory principles?

To address the first question, several groups recently used cell-type specific and

single-cell methodologies. Coffey et al[31] focused on microglial cells of the whole striatum,

using translational ribosome affinity purification (TRAP, to capture ribosomes and the

"translating transcriptome"[129, 130]). Chronic morphine effects were analyzed with, or

without, the induction of naloxone-precipitated withdrawal. Results showed that both chronic

morphine and withdrawal triggered numerous changes in gene expression, which mostly

affected GO terms related to cAMP signaling, regulation of synapses, and the cytoskeleton. A

striking negative correlation was observed between the 2 sets of results, providing a correlate,

at the level of the microglial transcriptome, of the pharmacological antagonism between

morphine and naloxone. Considering the inter-strain variability in the sensitivity to both acute

and chronic opioid physical dependence described earlier (see section *1.Qualitative analysis-*

Route and duration of opioid administration and [39, 40]), an interesting perspective would be

to determine whether naloxone similarly antagonizes transcriptomic effects of acute opioid

exposure, and how this is modulated by underlying genetic variation. Of note, surprisingly,

such cell-type specific approaches have not been used to interrogate opioid effects in neuronal

cells, let alone opioid-responsive MOR-expressing neurons which, arguably, may be

particularly sensitive. This is feasible, as knock-in lines in which the Cre recombinase is

selectively active in MOR-expressing cells are available[131–133].

Achieving higher resolution, Avey and colleagues applied single-cell RNA-seq

(scRNA-Seq, using Drop-seq[134]) to analyze transcriptomic changes induced by acute

morphine in more than 23,000 cells of the mouse NAc [51]. As is typical in scRNA-Seq,

sequencing depth yielded data for ~1,600 genes, corresponding to a reduced genomic

representation. Changes were widespread and, interestingly, most frequent in

oligodendrocytes, followed by neurons, astrocytes, and microglia, therefore uncovering a

cellular hierarchy in transcriptional sensitivity to opioids. More recently, Reiner et al[41] used

single-nucleus RNA-Seq (snRNA-seq, 10x Genomics 3') to analyze a larger number of nuclei

(190,030) and genes (>2000) in the rat NAc, also following acute morphine passive injection.

Compared to the former study, DEG mostly affected neurons and oligodendrocytes, followed

by astrocytes and microglia. Ten days of morphine IVSA was also investigated, and compared

to acute effects. Among DEG shared across the 2 conditions (n=431), the vast majority were

dysregulated in a similar direction (up/down), suggesting that they may reflect morphine

exposure rather than volitional drug-taking (a "yoked" group chronically exposed to morphine

would help substantiate this distinction). Importantly, a larger number of DEG were specifically

dysregulated in only one condition (n=868, with more DEG associated with IVSA), with distinct

distributions among NAc cell-types (e.g., Dopamine receptor D1- or D2-expressing neurons).

By providing the first single-cell description of morphine IVSA, these results reinforce the

notion that voluntary drug-intake may recruit partly distinct molecular mechanisms from those

of non-contingent administration.

We next compared results on acute morphine from these 2 studies. Both used

Seurat[135] and dimensionality reduction (t-SNE[51] or UMAP[41]) to identify clusters, which

were then assigned to cell-types using external data. We first compared marker genes

reported to define those clusters, when available (Avey: 11/29 clusters; Reiner: 11/19

clusters). Low agreement was found (Fig.S4A), with JI ranging from 0 up to 7.1% (for Avey's

activated medium spiny neurons, MSNs, and Reiner's D1R MSN-1), possibly because both

studies reported low numbers of marker genes (Avey: 39-51; Reiner: 4-12). We then

confronted morphine-induced DEG for each cell-type (Fig.6E). Low JI were found, with

maximal overlap (6.4%) obtained when comparing DEG affecting Avey's "myelin-forming and

mature oligodendrocytes" (MO_all) with Reiner's "Oligodendrocyte-3". Surprisingly, while

some clusters assigned to similar cell-types in the 2 studies showed some similarity in DEG

(green squares, Fig.6E), strong concordance was also observed across clusters with different

identities (e.g. Reiner's "Astrocyte-2" and Avey's MO_all; yellow squares). Overlaps among

DEG (Fig.6E) did not increase as a function of cell-type similarity across studies (Fig.S4B), as

would be expected. While particular emphasis has been put in sc/snRNA-Seq studies on the

number of individual cells to be sequenced, these results suggest that in order to maximize

their utility: i) marker genes used to assign cellular identities to clusters should be reported

comprehensively; and ii) rather than short lists of DEG, differential expression results should

be provided for all genes analyzed in each cluster. In addition, classical issues related to

sample size, power or statistical models for group comparisons remain pregnant. The Avey

and Reiner studies used similar sample sizes (4 and 5 replicates per opioid or control group,

respectively), in the upper range of bulk tissue studies (Fig.2D), but applied distinct tools to

identify DEG (the SCDE and MAST packages, respectively), which may partly account for

discrepancies. With the rapid evolution of the sc/snRNA-Seq field, and emerging controversies

(e.g. on dimensionality reduction[136]), additional efforts will be necessary to reach consensus

on designs and analyses[137, 138].

4. Non-coding and epigenomic mechanisms

In parallel to efforts to deepen cellular resolution, others have sought to decipher underlying

non-coding RNA and epigenomic regulations[139]. Although neurons are post-mitotic cells,

they exhibit peculiar plasticity features that may contribute to the encoding of life events,

including exposure to psychoactive drugs[140]. Several non-coding RNA species have been

investigated. The first report used custom-made arrays to characterize micro-RNA (miR) and

identify bidirectional expression changes affecting 7 of them, in rat primary neuronal cultures

or the mouse hippocampus, following morphine or heroin exposure[95]. Among these, miR-

339-3p was shown to target the 3'-UTR and down-regulate MOR in response to opioid

treatment[141]. A second study used more recent arrays to interrogate all mouse miR in the

NAc, following chronic heroin injections[63]. 111 miR exhibited >25% expression changes (no

significance reported). The study focused on miR-218, and showed that its over-expression

inhibited heroin-induced reinforcement (in conditioned place preference and IVSA), possibly

by targeting MeCP2 (a protein that binds methylated DNA), suggesting a potential link

between opioids, miR and DNA methylation. Other studies explored long non-coding (lncRNA)

or circular RNA (circRNA). One group used 2 arrays interrogating 9,000 lncRNA[89] and

15,000 circRNA[92] to characterize chronic morphine effects in the rat spinal cord: 136 lncRNA

(84 up, 52 down; nominal p-value<0.05, |log2FC|>1.5) and 2038 circRNA (896 up, 1142 down;

nominal p-value<0.05, FC>2) were affected. Another report used RNA-Sequencing to focus

on circRNA in the NAc[64], and found differences for 112 of them when comparing mice

receiving chronic morphine injections with or without electroacupuncture (51 up, 61 down;

nominal p-value<0.05, |log2FC|>1.5). Results from these studies remain descriptive, and

numerous mechanisms have been proposed to explain how non-coding RNA may exert

biological functions (including the sequestering of miR by circRNA, or complementarity

interactions among lncRNA and mRNA). Understanding how these mechanisms are recruited

by opioids will require further work.

Regarding the epigenome, 3 recent studies (1 in rats[62], 2 in humans[142, 143])

investigated DNA methylation[144]. The rat study used Whole Genome Bisulfite Sequencing

(WGBS) to analyze DNA methylation changes after heroin IVSA, in a standard or enriched

environment, in 3 brain regions (VTA, NAc and medial prefrontal cortex)[62]. Because of low

sequencing depth, DNA methylation levels were averaged across the whole genome for

selected features (promoters, CG islands). As expected, neither IVSA nor environmental

enrichment had any impact on these metrics, suggesting that the methylome is not massively

reprogrammed. Opioids may still, however, trigger more discrete DNA methylation changes,

a hypothesis supported by: i) the intuition that the large transcriptomic changes associated

with opioid exposure or OUD reviewed above are unlikely to occur in the absence of any

chromatin plasticity; ii) candidate studies which, although outside our scope, provide evidence

for differential DNA methylation at specific loci (the MOR gene in human blood[145–147]; few

loci in the rat brain[148]); and iii) results from the 2 human studies. Focusing on Brodmann

area 9 (BA9), the first applied EPIC arrays to a post-mortem cohort of OUD patients (n=19)

and healthy controls (n=11)[143]. Among >800,000 CG sites (where DNA methylation mostly

occurs in the mammalian brain), 11,917 located at transcription start sites (TSS) showed

nominally significant differences (p-value<0.05), with bidirectional changes (unfortunately,

genomic localization of these probes, and full genome-wide results, were not provided).

Such human studies face the difficult task of accounting for confounding factors.

Variation in genotype, age, sex, or socio-demographic factors, for example, have all been

associated with DNA methylation changes. Also, because epigenetic landscapes determine

cellular identity, group differences observed in bulk tissue may reflect variations in cellular

composition (possibly contributing to pathogenesis), rather than transcriptional regulation. The

second human study started to address these questions[142]. Using EPIC arrays to analyze

bulk tissue, the authors adjusted for socio-demographic characteristics, ancestry and cellular

composition. They investigated a large cohort (n=153), focusing on death from acute opioid

overdose. Although no individual CG site survived multiple testing correction, 13 passed a

relaxed significance threshold (p<1.0E-05), one of which was located in Netrin-1, a gene

implicated in KOR activity. At genome-wide level, opioid use associated with accelerated

epigenetic aging, along with enriched differential methylation in the KEGG pathway related to

dopaminergic signaling. Overall, these 2 human studies support the notion that the brain DNA

methylome may be modulated by opioids. More work will be necessary to better understand

these effects, as well as potential roles for non-canonical, brain-specific forms of DNA

methylation (methylation in non-CG context[149], hydroxymethylation at CG sites[150].

Beyond DNA modifications, epigenomic plasticity also relies on histone modifications

and changes in chromatin structure. Surprisingly, no data characterizing histone changes in

relation to opioid exposure were identified during our systematic screening. However, a recent

study compared chromatin accessibility in a cohort of 10 heroin users who died from heroin

overdose, and 10 healthy individuals[83], in the NAc and putamen. ATAC-Seq was used to

identify genomic regions where chromatin was "open", and DNA accessible. The technique

was applied to neuronal and non-neuronal cells, isolated using NeuN staining and

Fluorescence-Activated Nuclear Sorting (FANS). Heroin use associated with an enrichment

of ATAC-Seq peaks in specific gene features (CG islands, promoters, 5' untranslated regions)

and annotated regulatory regions (the histone mark H3K4me1, binding sites for the EZH2

transcription factor), as well as more frequent changes in neurons than in non-neuronal cells

(affecting GO terms related to the synapse, among others). At behavioral level, the authors

then showed that FYN (a tyrosine kinase with changes in chromatin accessibility in human)

significantly contributed to heroin IVSA in the rat. Overall, this work illustrates how epigenetic

analyses may uncover chromatin regulation contributing to OUD.

Discussion

Here we systematically assessed current knowledge on functional genomic mechanisms of

opioid action and OUD, and uncovered a surprisingly low convergence across studies. As a

result, a lot remains to be done to move the field forward. Below, we propose

recommendations for study design and data sharing, as well as perspectives to refine

behavioral modelling, improve cellular resolution, and advance multiomic data integration.

Recommendations for study design and data sharing. The low sample sizes observed in

the present review (Fig.2D,F) were surprising, considering that this factor is arguably the most

important one influencing statistical power[151–153]. Accordingly, a more rigorous analysis of

power, and larger sample sizes, will likely lead in future work to better reproducibility, and

should be enforced, especially when studying rodent outbred strains, or humans, who display

higher inter-individual variability[154, 155]. In humans, similar to what has been observed for

Genome-Wide Association Studies (GWAS[156]), functional genomic studies of OUD are

expected to evolve in the next decade towards collaborative efforts involving multiple labs (e.g.

the SCORCH consortium[157]), and the progressive aggregation of ever larger cohorts.

Another major recommendation relates to data sharing. While public repositories have been

specifically implemented for collecting and disseminating omic data (GEO[158], the IHEC data

portal[159]) or bioinformatic pipelines (Zenodo, GitHub), their systematic use is not yet current

practice in the OUD field, as exemplified in the present work (with similar criticisms raised in

other research domains[160]). This hampers re-analysis of published data, comparisons of

results across laboratories, and the use of threshold-free methods. We believe this last point

is particularly meaningful. Most opioid studies report subsets of DEG identified using arbitrary

significance cut-offs, which can mask subtle changes holding relevant biological information

(Fig.6[127]). We therefore strongly advocate for generalized use of threshold-free algorithms

for enrichment analysis of single datasets (GSEA), or comparison across pairs of differential

analysis results (RRHO2). Within this line, newer packages are expected to scale-up the

applicability of such tools to epigenomic data, which frequently involve larger number of

observations than gene-level counts[161].

Refine behavioral modeling. The face validity of SA or oICSS models of OUD has

significantly increased over last 2 decades [162], and we now face the task of more

systematically combining them with most recent molecular approaches. Another important

step will be to decipher the molecular mechanisms that may differentially account for distinct

behavioral dimensions, among impulsive or persistent drug-seeking, resistance to

punishment, or excessive motivation (including those modelled using passive administration,

such as tolerance, sensitization, or withdrawal). While the vast majority of studies reviewed

here used a categorical comparison of animals exposed or not to opioids, another design,

based on a dimensional approach within the exposed group, could help address the inter-

individual variability in susceptibility or resilience to OUD. The latter design has been used by

only one article in the present review[56] (section *2.1*), which contrasts with the amount of data

available for models of other psychiatric disorders, in particular depression (e.g. chronic social

defeat[163–165]. To better understand such inter-individual susceptibility, epigenetic

mechanisms are obvious candidates[166], as illustrated by 2 recent studies on DNA

methylation and oral SA of alcohol in macaques[167], and miRNA in a rat model of cocaine

SA[168]. While similar mechanisms are likely at play in OUD, they have not been addressed

by open-ended approaches yet, pointing toward an important avenue for future work.

Improve cellular resolution. This is another major avenue for future work, considering the

extreme heterogeneity of brain tissue. In addition to cell-type specific (TRAP) and sc/snRNA-

Seq (already applied to OUD, see above), a newer generation of tools now enables similar

molecular profiling while taking neuroanatomical context into consideration, with 2 possible

factors: i) connectivity patterns, probed using antero- or retrograde viruses, in combination

with TRAP (transcriptomics[169]) or FANS (epigenomics[170]); or ii) spatial context, with

methodologies based on sequencing or microscopy to detect RNA "in histological space";

while spatial capture followed by transcriptome sequencing (e.g. Slide-seq[171]) enables

genome-wide analysis with 10µm resolution, single-molecule fluorescence in situ hybridization

(smFISH) allows for better spatial resolution (1µm), but requires probe design and target gene

selection[146,147] (covering a few hundred to a few thousand genes, lower than sc/snRNA-

Seq[174]). Future work using these techniques, by characterizing molecular effects of opioids

in a particular neuronal pathway, or with exquisite spatial resolution, should deepen the

understanding of OUD.

Advance multiomic data integration. The epigenomic and transcriptomic mechanisms

reviewed above are thought to contribute to neurophysiological changes that are ultimately

mediated by changes at protein level. Proteomic studies were initially not included in the

present systematic bibliography, considering the low number of studies available and the fact

that, compared to RNA- or DNA-sequencing methodologies, they are limited to the analysis of

a reduced genome (a few hundred proteins; see [175–178]). Nevertheless, to go further in the

understanding of OUD, analyses at proteomic level will be necessary, ideally in combination

with the characterization of aforementioned molecular layers. Unfortunately, up to now,

numerous epigenetic mechanisms have not yet been investigated at genome-wide scale in

relation to opioids, including chromatin accessibility (ATAC-Seq) and conformation (4C), or

histone modifications (ChIP-Seq, Cut&Tag). This contrasts with studies conducted on

cocaine[150, 179]. Because the latter uncovered widespread histone post-translational

modifications, it can be hypothesized that opioids, notably through convergent dopaminergic

signaling, may recruit similar types of plasticity, whose reciprocal interaction with e.g. opioid-

induced changes in DNA methylation or gene expression remain unknown. The next

challenge, therefore, will be to integrate those numerous layers of analysis into multiomic

models of pathophysiology, which are likely to recruit genomic locus- and cell type-specific

mechanisms. Illustrating these concepts, we recently conducted a human study suggesting

that early-life adversity and depression associate with 2 forms of DNA methylation changes in

the canonical CG, and the brain-enriched CAC, contexts[149]. By combining WGBS with

ChIP-Seq analysis of 6 histone marks, the 2 forms of plasticity were found to affect distinct

genomic sites and features, and to be defined by different histones modifications and

chromatin states. Similar multiomic investigations of opioid effects should be conducted and,

importantly, will require innovative bioinformatic frameworks, which are only emerging in

psychiatry[180]. Only 2 studies reviewed here followed that path: Mendez et al performed

transcriptomic and proteomic analyses, while Liu et al analyzed the transcriptome and methylome (see section *4.Non-coding and epigenomic mechanisms of opioid plasticity*). In both studies, multiomic integration consisted: first, in the identification of features (single mRNA, DNA methylation site, or protein) passing a significance threshold, and their overlap; second, in the construction of co-expression or co-methylation networks, and the prioritization of groups of features, defined as modules, most significantly associated with OUD, and their biological interpretation (enrichment in functional GO terms, significantly affected features, GWAS results, etc). While these systems-biology approaches will undoubtedly contribute to the understanding of OUD, they remain based on step-by-step aggregation of results generated individually for each layer. In the coming years, it is expected that advanced multiomic integration approaches (e.g. similarity network fusion, or multiblock methods[181–183]), may more globally define, throughout the genome, how multiple regulatory layers interact, and the chain of events linking genetic, developmental, and environmental factors to abnormal behavior and compulsive drug use[184].

In conclusion, although numerous functional genomic studies of opioid action and OUD have been conducted across animal models and patient cohorts, replicable findings appear limited. In the future, combining refined behavioral models with multiomic integration, at the single-cell scale, will hopefully pave the way towards the identification of innovative therapeutic targets and, ultimately, contribute to better care.

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Conflict of Interest

The authors declare no conflicts of interest.

Code availability

All code used in the present review is available as Supplementary File 1.

Figure legends

Figure 1. PRISMA diagram. The review protocol was registered prospectively in Prospero (record ID#CRD42022270113), and followed the Preferred Reporting Items for Systematic Review and Meta-Analysis protocol (PRISMA[185]). *Identification of studies*. Three databases were used for systematic screening: MEDLINE, Embase and Web of Science. The literature search was performed using the following keywords: "microarray" OR "RNA sequencing" OR "bisulfite sequencing" OR "chromatin immunopurification sequencing" OR "single cell RNA sequencing" AND each of the following terms: "opiates", "opioid", "morphine", "fentanyl", "oxycodone", "heroin", "methadone" or "buprenorphine". The initial search provided 2705 potentially eligible studies. *Screening*. Articles were screened based on their title and abstract, and included if they used a high-throughput, genome-wide methodology to assess modifications of gene expression or epigenetic mechanisms as a function of opioid exposure, in the nervous system of primates or rodents (rat, mouse), and were published in English in peer-reviewed journals. Exclusion criteria included tissue other than the nervous system and candidate genes approaches. Once duplicates were removed, a total of 1225 articles were screened, among which 1067 were excluded because their title or abstract didn't meet eligibility criteria. *Eligibility*. After a more thorough examination of articles' full-text, an additional 89 articles were excluded. As a result, 69 papers were selected for qualitative synthesis. Among these, 44 articles reporting on transcriptomic analyses in the 5 most frequently investigated regions were selected for a more detailed quantitative synthesis. Of these, 37 articles investigated a single brain region, 6 covered 2 regions, and only 1 characterized 5 regions [13]. Overall, this resulted in 52 differential expression analyses: 18, 11, 11, 6 and 6 in the nucleus accumbens (NAc), frontal cortex, whole striatum, dorsal striatum and spinal cord, respectively.

Figure 2. Characteristics of studies included: Species & Sex. (A) Species repartition in studies identified during systematic search. **(B)** Distribution of sex and year of publication across studies. **(C-D)** Sex distribution specifically in rodent and human studies. **(E-F)** Sample

sizes in rodent and human studies comparing opioid-exposed animals to controls (rodent,

panel E), or individuals with a diagnosis of opioid use disorder (OUD) to healthy controls.

Figure 3. Characteristics of studies included: Injection parameters, Brain structure &

Techniques. Among the 69 studies considered during qualitative analysis, a large panel of

injection parameters, nervous system structures and techniques were represented. **(A)**

Injection parameters used for opioid administration. NB: "mixed" refers to a single study that

used a combination of chronic oral administration (through bottles) and acute ip injections of

morphine[75]. **(B)** Number of articles investigating each nervous system structure. **(C)** Side-

by-side comparison of the number of articles investigating each structure with the protein

expression profile of the molecular target of opioids, the mu opioid receptor (MOR; as

assessed using [^{3}H]-DAMGO autoradiography, data in fmol/mg of tissue, adapted from[111,

114–116]). **(D)** Distribution of high-throughput methodologies and year of publication across

studies. *Abbreviations:* FCx, Frontal cortex; NAc, Nucleus accumbens; dStr, Dorsal Striatum;

Hc, Hippocapus; HT, Hypothalamus; Th, Thalamus; Amy, Amygdala; VTA, Ventral Tegmental

Area; MHb, Medial Habenula; LC, Locus Coeruleus; DRN, Dorsal Raphe Nucleus.

Figure 4. Data availability for eligible studies and quantitative analysis in the nucleus

accumbens. (A) The graph depicts the number of: i) differential expression analysis results

considered for quantitative analysis (with the corresponding number of articles between

brackets); ii) cases when lists of differentially expressed genes (DEG) were available; iii) cases

when full genome-wide results were available (with both fold changes, FC, and p-values). **(B)**

Number of DEG identified in studies that investigated the nucleus accumbens. **(C-E)** Venn

diagram of DEG identified in each species (human, mice and rat). **(F)** Venn diagram of DEG

in the 2 studies that investigated the dorsal striatum.

Figure 5. Quantitative analyses in the dorsal striatum, whole striatum and frontal cortex.

(A) Number of differentially expressed genes (DEG) identified in studies that investigated the

striatum (see Fig.S1 for separate analyses of studies that used acute or chronic opioid

administration). **(B)** Number of DEG in studies of the frontal cortex. **(C)** Number of genes

identified as differentially expressed (DE) across different studies. **(D)** Across the 5 brain

regions considered during quantitative analysis, we assessed the impact of the following 7

factors on consistency among studies, using pairwise Jaccard indexes: species, brain

structure, treatment chronicity (acute/chronic), opioid drug (morphine/heroin), strain

(C57BL6J/others), time of analysis following last exposure (5 lists of DEG from tissue collected

1 hour after opioid exposure, 5 after 2 hours, 10 after 4 hours, 4 after 8 hours, 1 after 14 hours,

and 3 after 24 hours) and article. For each factor (e.g., species), pair-wise JI were classified

in 2 categories describing whether they belonged to the same sub-group (e.g., 2 studies

conducted in humans; "intra"), or not (e.g., 1 study in humans compared to another in rats;

"inter"), within that factor. For the drug factor, studies in human were not considered as they

cannot be assigned to specific opioids; a single study investigated fentanyl, and was

excluded[33]. For the strain factor, among the 14 mouse studies available, JI significantly

increased among the 12 that used C57BL6J mice, while a similar analysis was not possible

for 129P3/J, DBA/2J, SWR/J or Kunming strains, as only 1 list of DEG was available for each.

One study reporting on the combined analysis of C57BL6J and DBA2J mice was excluded[54].

No similar analysis was possible in rats, as all available studies used Sprague-Dawley (except

one that did not state the strain used[67]). Note that the numbers of JI values considered in

each comparison are indicated at the bottom of each bar.

Figure 6. (A-D) Threshold-free comparison of 2 bulk tissue RNA-sequencing studies

(Seney et al and Townsend et al), and cell-type specific comparison of 2 single-cell

(Avey et al) and single-nucleus (Reiner et al) RNA-sequencing studies. (A-B) Venn

diagram of differentially expressed genes (DEG) identified across the 2 bulk tissue RNA-

Sequencing studies by Seney et al and Townsend et al (DEG defined by arbitrary significance

thresholds in each study: p-value < 0.05 for both) for women/female rats (A) and men/male

rats (B). **(C-D)** Genome-wide, threshold-free comparison of the same 2 studies using Rank-

Rank Hypergeometric Overlap (RRHO2) for women/female (C) rats and men/male rats (D). In

RRHO2, briefly, genes are ranked using p-values (signed according to directionality), from

upregulated (Seney: left, Townsend: bottom) to down-regulated genes (Seney: right, fentan:

top). Overlaps along the 2 distributions are then iteratively assessed using hypergeometric

testing. High p-values in the bottom-left and top-right quadrants indicates concordance,

whereas high p-values in the other 2 quadrants (bottom-right and top-left) indicate discordance

between the 2 datasets. The white crosses indicate the location of the overlap corresponding

to Venn diagrams depicted in panels A-B (split in subgroups of up- or down-regulated genes),

while black crosses correspond to most significant overlaps identified using RRHO2. **(E)**

Heatmap of pairwise Jaccard indexes for DEG identified across each of the 26 and 27 cell-

types defined by Avey et al and Reiner et al, respectively.

References

1. Rosenblum A, Marsch LA, Joseph H, Portenoy RK. Opioids and the treatment of chronic pain: controversies, current status, and future directions. Exp Clin Psychopharmacol. 2008;16:405–416.

2. Florence C, Luo F, Rice K. The economic burden of opioid use disorder and fatal opioid overdose in the United States, 2017. Drug Alcohol Depend. 2021;218:108350.

3. European Monitoring Centre for Drugs and Drug Addiction. European drug report 2017 :trends and developments. LU: Publications Office; 2017.

4. Heilig M, MacKillop J, Martinez D, Rehm J, Leggio L, Vanderschuren LJMJ. Addiction as a brain disease revised: why it still matters, and the need for consilience. Neuropsychopharmacology. 2021;46:1715–1723.

5. Maze I, Shen L, Zhang B, Garcia BA, Shao N, Mitchell A, et al. Analytical tools and current challenges in the modern era of neuroepigenomics. Nat Neurosci. 2014;17:1476–1490.

6. Przewlocki R. Opioid abuse and brain gene expression. Eur J Pharmacol. 2004;500:331–349.

7. McClung CA. The molecular mechanisms of morphine addiction. Rev Neurosci. 2006;17:393–402.

8. Koob GF, Volkow ND. Neurobiology of addiction: a neurocircuitry analysis. Lancet Psychiatry. 2016;3:760–773.

9. Reed B, Kreek MJ. Genetic Vulnerability to Opioid Addiction. Cold Spring Harb Perspect Med. 2021;11:a039735.

10. Bodnar RJ. Endogenous opiates and behavior: 2020. Peptides. 2022;151:170752.

11. Przewlocki R. Opioid abuse and brain gene expression. European Journal of Pharmacology. 2004;500:331–349.

12. McClung CA. The molecular mechanisms of morphine addiction. Reviews in the Neurosciences. 2006;17:393–402.

13. Yamada K, Nagai T, Nabeshima T. Drug dependence, synaptic plasticity, and tissue plasminogen activator. J Pharmacol Sci. 2005;97:157–161.

14. UNODC, World Drug Report 2022 (United Nations publication, 2022).

15. Kerridge BT, Saha TD, Chou SP, Zhang H, Jung J, Ruan WJ, et al. Gender and nonmedical prescription opioid use and DSM-5 nonmedical prescription opioid use disorder: Results from the National Epidemiologic Survey on Alcohol and Related Conditions - III. Drug Alcohol Depend. 2015;156:47–56.

16. Bravo IM, Luster BR, Flanigan ME, Perez PJ, Cogan ES, Schmidt KT, et al. Divergent behavioral responses in protracted opioid withdrawal in male and female C57BL/6J mice. Eur J Neurosci. 2020;51:742–754.

17. Craft RM. Sex differences in opioid analgesia: 'from mouse to man'. Clin J Pain.
2003;19:175–186.

18. Craft RM, Stratmann JA, Bartok RE, Walpole TI, King SJ. Sex differences in development
of morphine tolerance and dependence in the rat. Psychopharmacology (Berl).
1999;143:1–7.

19. Cicero TJ, Nock B, Meyer ER. Gender-linked differences in the expression of physical
dependence in the rat. Pharmacol Biochem Behav. 2002;72:691–697.

20. Karami M, Zarrindast MR. Morphine sex-dependently induced place conditioning in adult
Wistar rats. Eur J Pharmacol. 2008;582:78–87.

21. Cicero TJ, Aylward SC, Meyer ER. Gender differences in the intravenous self-
administration of mu opiate agonists. Pharmacol Biochem Behav. 2003;74:541–549.

22. Beery AK, Zucker I. Sex bias in neuroscience and biomedical research. Neurosci
Biobehav Rev. 2011;35:565–572.

23. Prendergast BJ, Onishi KG, Zucker I. Female mice liberated for inclusion in neuroscience
and biomedical research. Neurosci Biobehav Rev. 2014;40:1–5.

24. Labonté B, Engmann O, Purushothaman I, Menard C, Wang J, Tan C, et al. Sex-specific
transcriptional signatures in human depression. Nat Med. 2017;23:1102–1111.

25. Hoffman GE, Ma Y, Montgomery KS, Bendl J, Jaiswal MK, Kozlenkov A, et al. Sex
Differences in the Human Brain Transcriptome of Cases With Schizophrenia. Biol
Psychiatry. 2022;91:92–101.

26. Hitzemann R, Bergeson SE, Berman AE, Bubier JA, Chesler EJ, Finn DA, et al. Sex
Differences in the Brain Transcriptome Related to Alcohol Effects and Alcohol Use
Disorder. Biol Psychiatry. 2022;91:43–52.

27. Vassoler FM, Oliver DJ, Wyse C, Blau A, Shtutman M, Turner JR, et al.
Transgenerational attenuation of opioid self-administration as a consequence of
adolescent morphine exposure. Neuropharmacology. 2017;113:271–280.

28. Borrelli KN, Yao EJ, Yen WW, Phadke RA, Ruan QT, Chen MM, et al. Sex Differences
in Behavioral and Brainstem Transcriptomic Neuroadaptations following Neonatal Opioid
Exposure in Outbred Mice. ENeuro. 2021;8:ENEURO.0143-21.2021.

29. Odegaard KE, Schaal VL, Clark AR, Koul S, Sankarasubramanian J, Xia Z, et al. A
Holistic Systems Approach to Characterize the Impact of Pre- and Post-natal Oxycodone
Exposure on Neurodevelopment and Behavior. Front Cell Dev Biol. 2020;8:619199.

30. Odegaard KE, Schaal VL, Clark AR, Koul S, Gowen A, Sankarasubramani J, et al.
Characterization of the intergenerational impact of in utero and postnatal oxycodone
exposure. Transl Psychiatry. 2020;10:329.

31. Coffey KR, Lesiak AJ, Marx RG, Vo EK, Garden GA, Neumaier JF. A cAMP-Related
Gene Network in Microglia Is Inversely Regulated by Morphine Tolerance and
Withdrawal. Biol Psychiatry Glob Open Sci. 2022;2:180–189.

32. Liu SX, Gades MS, Swain Y, Ramakrishnan A, Harris AC, Tran PV, et al. Repeated
morphine exposure activates synaptogenesis and other neuroplasticity-related gene

networks in the dorsomedial prefrontal cortex of male and female rats. Drug Alcohol
Depend. 2021;221:108598.

33. Townsend EA, Kim RK, Robinson HL, Marsh SA, Banks ML, Hamilton PJ. Opioid
withdrawal produces sex-specific effects on fentanyl-vs.-food choice and mesolimbic
transcription. Biol Psychiatry Glob Open Sci. 2021;1:112–122.

34. Seney ML, Kim S-M, Glausier JR, Hildebrand MA, Xue X, Zong W, et al. Transcriptional
Alterations in Dorsolateral Prefrontal Cortex and Nucleus Accumbens Implicate
Neuroinflammation and Synaptic Remodeling in Opioid Use Disorder. Biol Psychiatry.
2021;90:550–562.

35. Lefevre EM, Pisansky MT, Toddes C, Baruffaldi F, Pravetoni M, Tian L, et al. Interruption
of continuous opioid exposure exacerbates drug-evoked adaptations in the mesolimbic
dopamine system. Neuropsychopharmacology: Official Publication of the American
College of Neuropsychopharmacology. 2020;45:1781–1792.

36. Sugino S, Konno D, Abe J, Imamura-Kawasawa Y, Kido K, Suzuki J, et al. Crucial
involvement of catecholamine neurotransmission in postoperative nausea and vomiting:
Whole-transcriptome profiling in the rat nucleus of the solitary tract. Genes Brain Behav.
2021:e12759.

37. Ye J, Yang Z, Li C, Cai M, Zhou D, Zhang Q, et al. NF-κB signaling and vesicle transport
are correlated with the reactivation of the memory trace of morphine dependence. Diagn
Pathol. 2014;9:142.

38. Korostynski M, Piechota M, Dzbek J, Mlynarski W, Szklarczyk K, Ziolkowska B, et al.
Novel drug-regulated transcriptional networks in brain reveal pharmacological properties
of psychotropic drugs. BMC Genomics. 2013;14:606.

39. Klein G, Juni A, Waxman AR, Arout CA, Inturrisi CE, Kest B. A survey of acute and
chronic heroin dependence in ten inbred mouse strains: evidence of genetic correlation
with morphine dependence. Pharmacol Biochem Behav. 2008;90:447–452.

40. Kest B, Palmese CA, Hopkins E, Adler M, Juni A, Mogil JS. Naloxone-precipitated
withdrawal jumping in 11 inbred mouse strains: evidence for common genetic
mechanisms in acute and chronic morphine physical dependence. Neuroscience.
2002;115:463–469.

41. Reiner BC, Zhang Y, Stein LM, Perea ED, Arauco-Shapiro G, Ben Nathan J, et al. Single
nucleus transcriptomic analysis of rat nucleus accumbens reveals cell type-specific
patterns of gene expression associated with volitional morphine intake. Transl
Psychiatry. 2022;12:374.

42. Tapocik JD, Luu TV, Mayo CL, Wang B-D, Doyle E, Lee AD, et al. Neuroplasticity, axonal
guidance and micro-RNA genes are associated with morphine self-administration
behavior. Addiction Biology. 2013;18:480–495.

43. Berger AC, Whistler JL. Morphine-induced mu opioid receptor trafficking enhances
reward yet prevents compulsive drug use. EMBO Mol Med. 2011;3:385–397.

44. Slivicki RA, Earnest T, Chang Y-H, Pareta R, Casey E, Li J-N, et al. Oral oxycodone self-
administration leads to features of opioid misuse in male and female mice. Addict Biol.
2023;28:e13253.

45.   Phillips AG, McGovern DJ, Lee S, Ro K, Huynh DT, Elvig SK, et al. Oral prescription
opioid-seeking behavior in male and female mice. Addict Biol. 2020;25:e12828.

46.   Moussawi K, Ortiz MM, Gantz SC, Tunstall BJ, Marchette RCN, Bonci A, et al. Fentanyl
vapor self-administration model in mice to study opioid addiction. Sci Adv.
2020;6:eabc0413.

47.   Corre J, van Zessen R, Loureiro M, Patriarchi T, Tian L, Pascoli V, et al. Dopamine
neurons projecting to medial shell of the nucleus accumbens drive heroin reinforcement.
Elife. 2018;7:e39945.

48.   Corre J, van Zessen R, Loureiro M, Patriarchi T, Tian L, Pascoli V, et al. Dopamine
neurons projecting to medial shell of the nucleus accumbens drive heroin reinforcement.
ELife. 2018;7:e39945.

49.   Pascoli V, Terrier J, Hiver A, Lüscher C. Sufficiency of Mesolimbic Dopamine Neuron
Stimulation for the Progression to Addiction. Neuron. 2015;88:1054–1066.

50.   Siuda ER, Copits BA, Schmidt MJ, Baird MA, Al-Hasani R, Planer WJ, et al.
Spatiotemporal control of opioid signaling and behavior. Neuron. 2015;86:923–935.

51.   Avey D, Sankararaman S, Yim AKY, Barve R, Milbrandt J, Mitra RD. Single-Cell RNA-
Seq Uncovers a Robust Transcriptional Response to Morphine by Glia. Cell Rep.
2018;24:3619-3629.e4.

52.   Liang J, Chen J-H, Chen X-H, Peng Y-H, Zheng X-G. Gene expression of conditioned
locomotion and context-specific locomotor sensitization controlled by morphine-
associated environment. Behav Brain Res. 2011;216:321–331.

53.   Tapocik JD, Luu TV, Mayo CL, Wang B-D, Doyle E, Lee AD, et al. Neuroplasticity, axonal
guidance and micro-RNA genes are associated with morphine self-administration
behavior. Addict Biol. 2013;18:480–495.

54.   Grice DE, Reenilä I, Männistö PT, Brooks AI, Smith GG, Golden GT, et al. Transcriptional
profiling of C57 and DBA strains of mice in the absence and presence of morphine. BMC
Genomics. 2007;8:76.

55.   Ikeda H, Miyatake M, Koshikawa N, Ochiai K, Yamada K, Kiss A, et al. Morphine
modulation of thrombospondin levels in astrocytes and its implications for neurite
outgrowth and synapse formation. J Biol Chem. 2010;285:38415–38427.

56.   Imperio CG, McFalls AJ, Colechio EM, Masser DR, Vrana KE, Grigson PS, et al.
Assessment of individual differences in the rat nucleus accumbens transcriptome
following taste-heroin extended access. Brain Res Bull. 2016;123:71–80.

57.   Sillivan SE, Whittard JD, Jacobs MM, Ren Y, Mazloom AR, Caputi FF, et al. ELK1
transcription factor linked to dysregulated striatal mu opioid receptor signaling network
and OPRM1 polymorphism in human heroin abusers. Biol Psychiatry. 2013;74:511–519.

58.   Albertson DN, Schmidt CJ, Kapatos G, Bannon MJ. Distinctive profiles of gene
expression in the human nucleus accumbens associated with cocaine and heroin abuse.
Neuropsychopharmacology. 2006;31:2304–2312.

59.   Zhang Y, Liang Y, Randesi M, Yuferov V, Zhao C, Kreek MJ. Chronic Oxycodone Self-
administration Altered Reward-related Genes in the Ventral and Dorsal Striatum of
C57BL/6J Mice: An RNA-seq Analysis. Neuroscience. 2018;393:333–349.

60.   Zhang Y, Liang Y, Levran O, Randesi M, Yuferov V, Zhao C, et al. Alterations of
expression of inflammation/immune-related genes in the dorsal and ventral striatum of
adult C57BL/6J mice following chronic oxycodone self-administration: a RNA sequencing
study. Psychopharmacology (Berl). 2017;234:2259–2275.

61.   Yuferov V, Zhang Y, Liang Y, Zhao C, Randesi M, Kreek MJ. Oxycodone Self-
Administration Induces Alterations in Expression of Integrin, Semaphorin and Ephrin
Genes in the Mouse Striatum. Front Psychiatry. 2018;9:257.

62.   Imperio CG, McFalls AJ, Hadad N, Blanco-Berdugo L, Masser DR, Colechio EM, et al.
Exposure to environmental enrichment attenuates addiction-like behavior and alters
molecular effects of heroin self-administration in rats. Neuropharmacology.
2018;139:26–40.

63.   Yan B, Hu Z, Yao W, Le Q, Xu B, Liu X, et al. MiR-218 targets MeCP2 and inhibits heroin
seeking behavior. Sci Rep. 2017;7:40413.

64.   Zhang H, Wang Q, Wang Q, Liu A, Qin F, Sun Q, et al. Circular RNA expression profiling
in the nucleus accumbens: Effects of electroacupuncture treatment on morphine-induced
conditioned place preference. Addict Biol. 2020;25:e12794.

65.   Martin JA, Caccamise A, Werner CT, Viswanathan R, Polanco JJ, Stewart AF, et al. A
Novel Role for Oligodendrocyte Precursor Cells (OPCs) and Sox10 in Mediating Cellular
and Behavioral Responses to Heroin. Neuropsychopharmacology. 2018;43:1385–1394.

66.   Ammon-Treiber S, Tischmeyer H, Riechert U, Höllt V. Gene expression of transcription
factors in the rat brain after morphine withdrawal. Neurochem Res. 2004;29:1267–1273.

67.   Kuntz-Melcavage KL, Brucklacher RM, Grigson PS, Freeman WM, Vrana KE. Gene
expression changes following extinction testing in a heroin behavioral incubation model.
BMC Neurosci. 2009;10:95.

68.   Ammon S, Mayer P, Riechert U, Tischmeyer H, Höllt V. Microarray analysis of genes
expressed in the frontal cortex of rats chronically treated with morphine and after
naloxone precipitated withdrawal. Brain Res Mol Brain Res. 2003;112:113–125.

69.   Mendez EF, Wei H, Hu R, Stertz L, Fries GR, Wu X, et al. Angiogenic gene networks are
dysregulated in opioid use disorder: evidence from multi-omics and imaging of
postmortem human brain. Mol Psychiatry. 2021;26:7803–7812.

70.   Liu A, Dai Y, Mendez EF, Hu R, Fries GR, Najera KE, et al. Genome-Wide Correlation
of DNA Methylation and Gene Expression in Postmortem Brain Tissues of Opioid Use
Disorder Patients. Int J Neuropsychopharmacol. 2021;24:879–891.

71.   Jiang C, Wang X, Le Q, Liu P, Liu C, Wang Z, et al. Morphine coordinates SST and PV
interneurons in the prelimbic cortex to disinhibit pyramidal neurons and enhance reward.
Mol Psychiatry. 2021;26:1178–1193.

72.   Shu C, Sosnowski DW, Tao R, Deep-Soboslay A, Kleinman JE, Hyde TM, et al.
Epigenome-wide study of brain DNA methylation following acute opioid intoxication. Drug
Alcohol Depend. 2021;221:108658.

73. Loguinov AV, Anderson LM, Crosby GJ, Yukhananov RY. Gene expression following
acute morphine administration. Physiol Genomics. 2001;6:169–181.

74. Choi MR, Jin Y-B, Bang SH, Im C-N, Lee Y, Kim H-N, et al. Age-related Effects of Heroin
on Gene Expression in the Hippocampus and Striatum of Cynomolgus Monkeys. Clin
Psychopharmacol Neurosci. 2020;18:93–108.

75. Skupio U, Sikora M, Korostynski M, Wawrzczak-Bargiela A, Piechota M, Ficek J, et al.
Behavioral and transcriptional patterns of protracted opioid self-administration in mice.
Addict Biol. 2017;22:1802–1816.

76. Piechota M, Korostynski M, Solecki W, Gieryk A, Slezak M, Bilecki W, et al. The
dissection of transcriptional modules regulated by various drugs of abuse in the mouse
striatum. Genome Biol. 2010;11:R48.

77. Korostynski M, Piechota M, Kaminska D, Solecki W, Przewlocki R. Morphine effects on
striatal transcriptome in mice. Genome Biol. 2007;8:R128.

78. Piechota M, Korostynski M, Sikora M, Golda S, Dzbek J, Przewlocki R. Common
transcriptional effects in the mouse striatum following chronic treatment with heroin and
methamphetamine. Genes Brain Behav. 2012;11:404–414.

79. Korostynski M, Piechota M, Golda S, Przewlocki R. High-throughput gene expression
profiling of opioid-induced alterations in discrete brain areas. Methods Mol Biol.
2015;1230:65–76.

80. Choi MR, Jin Y-B, Kim H-N, Chai YG, Im C-N, Lee S-R, et al. Gene expression in the
striatum of cynomolgus monkeys after chronic administration of cocaine and heroin.
Basic Clin Pharmacol Toxicol. 2021;128:686–698.

81. Cha HJ, Choi M-J, Ahn J-I, Jeon S-H, Kang H, Kim EJ, et al. Comparison of gene
expression profiles in drug-withdrawn rats. Mol Cell Toxicol. 2016;12:197–207.

82. Valderrama-Carvajal A, Irizar H, Gago B, Jiménez-Urbieta H, Fuxe K, Rodríguez-Oroz
MC, et al. Transcriptomic integration of D4R and MOR signaling in the rat caudate
putamen. Sci Rep. 2018;8:7337.

83. Egervari G, Akpoyibo D, Rahman T, Fullard JF, Callens JE, Landry JA, et al. Chromatin
accessibility mapping of the striatum identifies tyrosine kinase FYN as a therapeutic
target for heroin use disorder. Nat Commun. 2020;11:4634.

84. Heller EA, Kaska S, Fallon B, Ferguson D, Kennedy PJ, Neve RL, et al. Morphine and
cocaine increase serum- and glucocorticoid-inducible kinase 1 activity in the ventral
tegmental area. J Neurochem. 2015;132:243–253.

85. McClung CA, Nestler EJ, Zachariou V. Regulation of gene expression by chronic
morphine and morphine withdrawal in the locus ceruleus and ventral tegmental area. J
Neurosci. 2005;25:6005–6015.

86. Zhang H, Wang Q, Sun Q, Qin F, Nie D, Li Q, et al. Effects of Compound 511 on BDNF-
TrkB Signaling in the Mice Ventral Tegmental Area in Morphine-Induced Conditioned
Place Preference. Cell Mol Neurobiol. 2021;41:961–975.

87. Jokinen V, Sidorova Y, Viisanen H, Suleymanova I, Tiilikainen H, Li Z, et al. Differential Spinal and Supraspinal Activation of Glia in a Rat Model of Morphine Tolerance. Neuroscience. 2018;375:10–24.

88. Cheng Y-C, Tsai R-Y, Sung Y-T, Chen I-J, Tu T-Y, Mao Y-Y, et al. Melatonin regulation of transcription in the reversal of morphine tolerance: Microarray analysis of differential gene expression. Int J Mol Med. 2019;43:791–806.

89. Shao J, Wang J, Huang J, Liu C, Pan Y, Guo Q, et al. Identification of lncRNA expression profiles and ceRNA analysis in the spinal cord of morphine-tolerant rats. Mol Brain. 2018;11:21.

90. Peng Y, Guo G, Shu B, Liu D, Su P, Zhang X, et al. Spinal CX3CL1/CX3CR1 May Not Directly Participate in the Development of Morphine Tolerance in Rats. Neurochem Res. 2017;42:3254–3267.

91. Santiago JM, Rosas O, Torrado AI, González MM, Kalyan-Masih PO, Miranda JD. Molecular, anatomical, physiological, and behavioral studies of rats treated with buprenorphine after spinal cord injury. J Neurotrauma. 2009;26:1783–1793.

92. Weng Y, Wu J, Li L, Shao J, Li Z, Deng M, et al. Circular RNA expression profile in the spinal cord of morphine tolerated rats and screen of putative key circRNAs. Mol Brain. 2019;12:79.

93. Juul SE, Beyer RP, Bammler TK, Farin FM, Gleason CA. Effects of neonatal stress and morphine on murine hippocampal gene expression. Pediatr Res. 2011;69:285–292.

94. Marie-Claire C, Courtin C, Robert A, Gidrol X, Roques BP, Noble F. Sensitization to the conditioned rewarding effects of morphine modulates gene expression in rat hippocampus. Neuropharmacology. 2007;52:430–435.

95. Zheng H, Zeng Y, Zhang X, Chu J, Loh HH, Law P-Y. mu-Opioid receptor agonists differentially regulate the expression of miR-190 and NeuroD. Mol Pharmacol. 2010;77:102–109.

96. Befort K, Filliol D, Ghate A, Darcq E, Matifas A, Muller J, et al. Mu-opioid receptor activation induces transcriptional plasticity in the central extended amygdala. Eur J Neurosci. 2008;27:2973–2984.

97. Rodriguez Parkitna JM, Bilecki W, Mierzejewski P, Stefanski R, Ligeza A, Bargiela A, et al. Effects of morphine on gene expression in the rat amygdala. J Neurochem. 2004;91:38–48.

98. Zachariou V, Liu R, LaPlant Q, Xiao G, Renthal W, Chan GC, et al. Distinct roles of adenylyl cyclases 1 and 8 in opiate dependence: behavioral, electrophysiological, and molecular studies. Biol Psychiatry. 2008;63:1013–1021.

99. Saad MH, Rumschlag M, Guerra MH, Savonen CL, Jaster AM, Olson PD, et al. Differentially expressed gene networks, biomarkers, long noncoding RNAs, and shared responses with cocaine identified in the midbrains of human opioid abusers. Scientific Reports. 2019;9:1534.

100. Befort K, Filliol D, Darcq E, Ghate A, Matifas A, Lardenois A, et al. Gene expression is altered in the lateral hypothalamus upon activation of the mu opioid receptor. Ann N Y Acad Sci. 2008;1129:175–184.

101. Anghel A, Jamieson C a. M, Ren X, Young J, Porche R, Ozigbo E, et al. Gene expression
profiling following short-term and long-term morphine exposure in mice uncovers genes
involved in food intake. Neuroscience. 2010;167:554–566.

102. Hayase T, Yamamoto Y, Yamamoto K, Muso E, Shiota K. Microarray profile analysis of
toxic cocaine-induced alterations in the expression of mouse brain gene sequences: a
possible 'protective' effect of buprenorphine. J Appl Toxicol. 2004;24:15–20.

103. Hassan HE, Myers AL, Lee IJ, Chen H, Coop A, Eddington ND. Regulation of gene
expression in brain tissues of rats repeatedly treated by the highly abused opioid agonist,
oxycodone: microarray profiling and gene mapping analysis. Drug Metab Dispos.
2010;38:157–167.

104. Gaspari S, Purushothaman I, Cogliani V, Sakloth F, Neve RL, Howland D, et al.
Suppression of RGSz1 function optimizes the actions of opioid analgesics by
mechanisms that involve the Wnt/β-catenin pathway. Proc Natl Acad Sci U S A.
2018;115:E2085–E2094.

105. Toorie AM, Vassoler FM, Qu F, Schonhoff CM, Bradburn S, Murgatroyd CA, et al. A
history of opioid exposure in females increases the risk of metabolic disorders in their
future male offspring. Addict Biol. 2021;26:e12856.

106. Thibault K, Calvino B, Rivals I, Marchand F, Dubacq S, McMahon SB, et al. Molecular
mechanisms underlying the enhanced analgesic effect of oxycodone compared to
morphine in chemotherapy-induced neuropathic pain. PLoS One. 2014;9:e91297.

107. Delfs JM, Zhu Y, Druhan JP, Aston-Jones G. Noradrenaline in the ventral forebrain is
critical for opiate withdrawal-induced aversion. Nature. 2000;403:430–434.

108. Zhu Y, Wienecke CF, Nachtrab G, Chen X. A thalamic input to the nucleus accumbens
mediates opiate dependence. Nature. 2016;530:219–222.

109. Koob GF. The dark side of emotion: the addiction perspective. Eur J Pharmacol.
2015;753:73–87.

110. Hearing M. Prefrontal-accumbens opioid plasticity: Implications for relapse and
dependence. Pharmacol Res. 2019;139:158–165.

111. Lutz P-E, Kieffer BL. The multiple facets of opioid receptor function: implications for
addiction. Curr Opin Neurobiol. 2013;23:473–479.

112. Lutz P-E, Almeida D, Filliol D, Jollant F, Kieffer BL, Turecki G. Increased functional
coupling of the mu opioid receptor in the anterior insula of depressed individuals.
Neuropsychopharmacology. 2021;46:920–927.

113. Lutz P-E, Kieffer BL. Opioid receptors: distinct roles in mood disorders. Trends Neurosci.
2013;36:195–206.

114. Kitchen I, Slowe SJ, Matthes HW, Kieffer B. Quantitative autoradiographic mapping of
mu-, delta- and kappa-opioid receptors in knockout mice lacking the mu-opioid receptor
gene. Brain Res. 1997;778:73–88.

115. Goody RJ, Oakley SM, Filliol D, Kieffer BL, Kitchen I. Quantitative autoradiographic
mapping of opioid receptors in the brain of delta-opioid receptor gene knockout mice.
Brain Res. 2002;945:9–19.

116. Slowe SJ, Simonin F, Kieffer B, Kitchen I. Quantitative autoradiography of mu-,delta- and kappa1 opioid receptors in kappa-opioid receptor knockout mice. Brain Res. 1999;818:335–345.

117. Xia Y, Haddad GG. Ontogeny and distribution of opioid receptors in the rat brainstem. Brain Res. 1991;549:181–193.

118. Miyakawa T. No raw data, no science: another possible source of the reproducibility crisis. Mol Brain. 2020;13:24.

119. Field D, Sansone S-A, Collis A, Booth T, Dukes P, Gregurick SK, et al. Megascience. 'Omics data sharing. Science. 2009;326:234–236.

120. FAIR Principles. GO FAIR. https://www.go-fair.org/fair-principles/. Accessed 14 February 2023.

121. NIH Genomic Data Sharing.

122. Murray JE, Belin-Rauscent A, Simon M, Giuliano C, Benoit-Marand M, Everitt BJ, et al. Basolateral and central amygdala differentially recruit and maintain dorsolateral striatum-dependent cocaine-seeking habits. Nat Commun. 2015;6:10088.

123. Zezula J, Casaccia-Bonnefil P, Ezhevsky SA, Osterhout DJ, Levine JM, Dowdy SF, et al. p21cip1 is required for the differentiation of oligodendrocytes independently of cell cycle withdrawal. EMBO Rep. 2001;2:27–34.

124. Scholpa NE, Briggs SB, Wagner JJ, Cummings BS. Cyclin-Dependent Kinase Inhibitor 1a (p21) Modulates Response to Cocaine and Motivated Behaviors. J Pharmacol Exp Ther. 2016;357:56–65.

125. Häusl AS, Brix LM, Hartmann J, Pöhlmann ML, Lopez J-P, Menegaz D, et al. The co-chaperone Fkbp5 shapes the acute stress response in the paraventricular nucleus of the hypothalamus of male mice. Mol Psychiatry. 2021;26:3060–3076.

126. Levran O, Peles E, Randesi M, Li Y, Rotrosen J, Ott J, et al. Stress-related genes and heroin addiction: a role for a functional FKBP5 haplotype. Psychoneuroendocrinology. 2014;45:67–76.

127. Cahill KM, Huo Z, Tseng GC, Logan RW, Seney ML. Improved identification of concordant and discordant gene expression signatures using an updated rank-rank hypergeometric overlap approach. Scientific Reports. 2018;8:9588.

128. Al-Hasani R, Bruchas MR. Molecular mechanisms of opioid receptor-dependent signaling and behavior. Anesthesiology. 2011;115:1363–1381.

129. Nectow AR, Moya MV, Ekstrand MI, Mousa A, McGuire KL, Sferrazza CE, et al. Rapid Molecular Profiling of Defined Cell Types Using Viral TRAP. Cell Rep. 2017;19:655–667.

130. Heiman M, Schaefer A, Gong S, Peterson JD, Day M, Ramsey KE, et al. A translational profiling approach for the molecular characterization of CNS cell types. Cell. 2008;135:738–748.

131. Bailly J, Del Rossi N, Runtz L, Li J-J, Park D, Scherrer G, et al. Targeting morphine-responsive neurons: generation of a knock-in mouse line expressing Cre recombinase

from the mu opioid receptor gene locus. ENeuro. 2020. 7 May 2020. https://doi.org/10.1523/ENEURO.0433-19.2020.

132. Okunomiya T, Hioki H, Nishimura C, Yawata S, Imayoshi I, Kageyama R, et al. Generation of a MOR-CreER knock-in mouse line to study cells and neural circuits involved in mu opioid receptor signaling. Genesis. 2020;58:e23341.

133. Märtin A, Calvigioni D, Tzortzi O, Fuzik J, Wärnberg E, Meletis K. A Spatiomolecular Map of the Striatum. Cell Rep. 2019;29:4320-4333.e5.

134. Macosko EZ, Basu A, Satija R, Nemesh J, Shekhar K, Goldman M, et al. Highly Parallel Genome-wide Expression Profiling of Individual Cells Using Nanoliter Droplets. Cell. 2015;161:1202–1214.

135. Stuart T, Butler A, Hoffman P, Hafemeister C, Papalexi E, Mauck WM, et al. Comprehensive Integration of Single-Cell Data. Cell. 2019;177:1888-1902.e21.

136. Chari T, Pachter L. The Specious Art of Single-Cell Genomics. Genomics; 2021.

137. Gibson G. Perspectives on rigor and reproducibility in single cell genomics. PLoS Genet. 2022;18:e1010210.

138. Schmid KT, Höllbacher B, Cruceanu C, Böttcher A, Lickert H, Binder EB, et al. scPower accelerates and optimizes the design of multi-sample single cell transcriptomic studies. Nat Commun. 2021;12:6625.

139. Browne CJ, Godino A, Salery M, Nestler EJ. Epigenetic Mechanisms of Opioid Addiction. Biol Psychiatry. 2020;87:22–33.

140. Belzeaux R, Lalanne L, Kieffer BL, Lutz PE. Focusing on the Opioid System for Addiction Biomarker Discovery. Trends Mol Med. 2018;24:206–220.

141. Wu Q, Hwang CK, Zheng H, Wagley Y, Lin H-Y, Kim DK, et al. MicroRNA 339 down-regulates μ-opioid receptor at the post-transcriptional level in response to opioid treatment. FASEB J. 2013;27:522–535.

142. Shu C, Sosnowski DW, Tao R, Deep-Soboslay A, Kleinman JE, Hyde TM, et al. Epigenome-wide study of brain DNA methylation following acute opioid intoxication. Drug Alcohol Depend. 2021;221:108658.

143. Liu A, Dai Y, Mendez EF, Hu R, Fries GR, Najera KE, et al. Genome-Wide Correlation of DNA Methylation and Gene Expression in Postmortem Brain Tissues of Opioid Use Disorder Patients. Int J Neuropsychopharmacol. 2021;24:879–891.

144. Lutz P-E, Mechawar N, Turecki G. Neuropathology of suicide: recent findings and future directions. Mol Psychiatry. 2017;22:1395–1412.

145. Ebrahimi G, Asadikaram G, Akbari H, Nematollahi MH, Abolhassani M, Shahabinejad G, et al. Elevated levels of DNA methylation at the OPRM1 promoter region in men with opioid use disorder. Am J Drug Alcohol Abuse. 2017:1–7.

146. Nielsen DA, Yuferov V, Hamon S, Jackson C, Ho A, Ott J, et al. Increased OPRM1 DNA methylation in lymphocytes of methadone-maintained former heroin addicts. Neuropsychopharmacology. 2009;34:867–873.

147. Doehring A, Oertel BG, Sittl R, Lotsch J. Chronic opioid use is associated with increased
DNA methylation correlating with increased clinical pain. Pain. 2013;154:15–23.

148. Barrow TM, Byun HM, Li X, Smart C, Wang YX, Zhang Y, et al. The effect of morphine
upon DNA methylation in ten regions of the rat brain. Epigenetics. 2017;12:1038–1047.

149. Lutz P-E, Chay M-A, Pacis A, Chen GG, Aouabed Z, Maffioletti E, et al. Non-CG
methylation and multiple histone profiles associate child abuse with immune and small
GTPase dysregulation. Nat Commun. 2021;12:1132.

150. Feng J, Shao N, Szulwach KE, Vialou V, Huynh J, Zhong C, et al. Role of Tet1 and 5-
hydroxymethylcytosine in cocaine action. Nat Neurosci. 2015;18:536–544.

151. Ching T, Huang S, Garmire LX. Power analysis and sample size estimation for RNA-Seq
differential expression. RNA. 2014;20:1684–1696.

152. Baccarella A, Williams CR, Parrish JZ, Kim CC. Empirical assessment of the impact of
sample number and read depth on RNA-Seq analysis workflow performance. BMC
Bioinformatics. 2018;19:423.

153. Hoffman GE, Jaffe AE, Gandal MJ, Collado-Torres L, Sieberts SK, Devlin B, et al.
Comment on: What genes are differentially expressed in individuals with schizophrenia?
A systematic review. Mol Psychiatry. 2023;28:523–525.

154. Hekman JP, Johnson JL, Kukekova AV. Transcriptome Analysis in Domesticated
Species: Challenges and Strategies. Bioinform Biol Insights. 2015;9:21–31.

155. Williams AG, Thomas S, Wyman SK, Holloway AK. RNA-seq Data: Challenges in and
Recommendations for Experimental Design and Analysis. Curr Protoc Hum Genet.
2014;83:11.13.1-20.

156. Crist RC, Reiner BC, Berrettini WH. A review of opioid addiction genetics. Curr Opin
Psychol. 2019;27:31–35.

157. scorch - Home. https://scorch.igs.umaryland.edu/. Accessed 19 May 2023.

158. Barrett T, Wilhite SE, Ledoux P, Evangelista C, Kim IF, Tomashevsky M, et al. NCBI
GEO: archive for functional genomics data sets--update. Nucleic Acids Res.
2013;41:D991-995.

159. Data Portal | IHEC. https://epigenomesportal.ca/ihec/. Accessed 19 May 2023.

160. Savage CJ, Vickers AJ. Empirical study of data sharing by authors publishing in PLoS
journals. PLoS One. 2009;4:e7078.

161. Piron A, Szymczak F, Alvelos MI, Defrance M, Lenaerts T, Eizirik DL, et al. RedRibbon:
A new rank-rank hypergeometric overlap pipeline to compare gene and transcript
expression signatures. Bioinformatics; 2022.

162. Deroche-Gamonet V, Belin D, Piazza PV. Evidence for addiction-like behavior in the rat.
Science. 2004;305:1014–1017.

163. Domingo-Rodriguez L, Ruiz de Azua I, Dominguez E, Senabre E, Serra I, Kummer S, et
al. A specific prelimbic-nucleus accumbens pathway controls resilience versus
vulnerability to food addiction. Nat Commun. 2020;11:782.

164. Laine MA, Trontti K, Misiewicz Z, Sokolowska E, Kulesskaya N, Heikkinen A, et al.
Genetic Control of Myelin Plasticity after Chronic Psychosocial Stress. ENeuro.
2018;5:ENEURO.0166-18.2018.

165. Bagot RC, Cates HM, Purushothaman I, Lorsch ZS, Walker DM, Wang J, et al. Circuit-
wide Transcriptional Profiling Reveals Brain Region-Specific Gene Networks Regulating
Depression Susceptibility. Neuron. 2016;90:969–983.

166. Roy B, Wang Q, Dwivedi Y. Long Noncoding RNA-Associated Transcriptomic Changes
in Resiliency or Susceptibility to Depression and Response to Antidepressant Treatment.
Int J Neuropsychopharmacol. 2018;21:461–472.

167. Cervera-Juanes R, Wilhelm LJ, Park B, Grant KA, Ferguson B. Alcohol-dose-dependent
DNA methylation and expression in the nucleus accumbens identifies coordinated
regulation of synaptic genes. Transl Psychiatry. 2017;7:e994.

168. Domingo-Rodriguez L, Cabana-Domínguez J, Fernàndez-Castillo N, Cormand B,
Martín-García E, Maldonado R. Differential expression of miR-1249-3p and miR-34b-5p
between vulnerable and resilient phenotypes of cocaine addiction. Addict Biol.
2022;27:e13201.

169. Ekstrand MI, Nectow AR, Knight ZA, Latcha KN, Pomeranz LE, Friedman JM. Molecular
profiling of neurons based on connectivity. Cell. 2014;157:1230–1242.

170. Zhang Z, Zhou J, Tan P, Pang Y, Rivkin AC, Kirchgessner MA, et al. Epigenomic diversity
of cortical projection neurons in the mouse brain. Nature. 2021;598:167–173.

171. Rodriques SG, Stickels RR, Goeva A, Martin CA, Murray E, Vanderburg CR, et al. Slide-
seq: A scalable technology for measuring genome-wide expression at high spatial
resolution. Science. 2019;363:1463–1467.

172. Eng C-HL, Lawson M, Zhu Q, Dries R, Koulena N, Takei Y, et al. Transcriptome-scale
super-resolved imaging in tissues by RNA seqFISH. Nature. 2019;568:235–239.

173. Borm LE, Mossi Albiach A, Mannens CCA, Janusauskas J, Özgün C, Fernández-García
D, et al. Scalable in situ single-cell profiling by electrophoretic capture of mRNA using
EEL FISH. Nat Biotechnol. 2022. 22 September 2022. https://doi.org/10.1038/s41587-
022-01455-3.

174. Armand EJ, Li J, Xie F, Luo C, Mukamel EA. Single-Cell Sequencing of Brain Cell
Transcriptomes and Epigenomes. Neuron. 2021;109:11–26.

175. Mendez EF, Wei H, Hu R, Stertz L, Fries GR, Wu X, et al. Angiogenic gene networks are
dysregulated in opioid use disorder: evidence from multi-omics and imaging of
postmortem human brain. Molecular Psychiatry. 2021;26:7803–7812.

176. Li KW, Jimenez CR, van der Schors RC, Hornshaw MP, Schoffelmeer ANM, Smit AB.
Intermittent administration of morphine alters protein expression in rat nucleus
accumbens. Proteomics. 2006;6:2003–2008.

177. Van den Oever MC, Lubbers BR, Goriounova NA, Li KW, Van der Schors RC, Loos M,
et al. Extracellular matrix plasticity and GABAergic inhibition of prefrontal cortex
pyramidal cells facilitates relapse to heroin seeking. Neuropsychopharmacology.
2010;35:2120–2133.

178. Bu Q, Yang Y, Yan G, Hu Z, Hu C, Duan J, et al. Proteomic analysis of the nucleus
accumbens in rhesus monkeys of morphine dependence and withdrawal intervention. J
Proteomics. 2012;75:1330–1342.

179. Feng J, Wilkinson M, Liu X, Purushothaman I, Ferguson D, Vialou V, et al. Chronic
cocaine-regulated epigenomic changes in mouse nucleus accumbens. Genome Biol.
2014;15:R65.

180. Mokhtari A, Porte B, Belzeaux R, Etain B, Ibrahim EC, Marie-Claire C, et al. The
molecular pathophysiology of mood disorders: From the analysis of single molecular
layers to multi-omic integration. Prog Neuropsychopharmacol Biol Psychiatry.
2022;116:110520.

181. Li C-X, Wheelock CE, Sköld CM, Wheelock ÅM. Integration of multi-omics datasets
enables molecular classification of COPD. Eur Respir J. 2018;51:1701930.

182. Yang M, Matan-Lithwick S, Wang Y, De Jager PL, Bennett DA, Felsky D. Multi-omic
integration via similarity network fusion to detect molecular subtypes of ageing. Brain
Commun. 2023;5:fcad110.

183. Revilla L, Mayorgas A, Corraliza AM, Masamunt MC, Metwaly A, Haller D, et al. Multi-
omic modelling of inflammatory bowel disease with regularized canonical correlation
analysis. PLoS One. 2021;16:e0246367.

184. Hasin Y, Seldin M, Lusis A. Multi-omics approaches to disease. Genome Biol.
2017;18:83.

185. Liberati A, Altman DG, Tetzlaff J, Mulrow C, Gotzsche PC, Ioannidis JPA, et al. The
PRISMA statement for reporting systematic reviews and meta-analyses of studies that
evaluate healthcare interventions: explanation and elaboration. BMJ. 2009;339:b2700–
b2700.

1305

Figure 1

Figure 2

Figure 3

Figure 4

A

55 (44)

24 (17)

4 (from 3 articles)

Articles included in quantitative analysis

Lists of DEG with variable criteria

Complete results (with FC and p-val)

B

Nucleus accumbens

Intersection size

967 720 460 292 270 142 129 114 44 41 35 30 26 25 25

Grice
Ye
Imperio_SS
Lefèvre
Imperio_LS
Townsend
Albertson
Seney

Gene set size

C Human

Albertson 970 54 Seney 1252

D Mouse

Lefevre 362

4 2

Ye 160 1 Grice 148

E Rat

Imperio LS 400

154 9

Townsend 653

Imperio SS 39

F

Dorsal striatum

Lefevre 462

Valderrama

Figure 5

A

Striatum
Acute + chronic
studies

Intersection size

2352, 348, 213, 175, 132, 79, 54, 38, 37, 32, 28, 23, 21, 20, 19, 11, 10, 9, 8, 7

Gene set size	Study																				
18	Piechota_chronic_heroin_2012																				●
34	Piechota_acute_morphine_2010																	●		
58	Piechota_acute_heroin_2010															●				
124	Korostynski_acute_heroin_2013						●							●			●		
195	Korostynski_acute_2007						●			●			●			●			
221	Skupio_chronic_morphine			●								●						●	
247	Korostynski_acute_morphine_2013					●				●				●				
390	Korostynski_chronic_2007			●			●		●				●			●		
552	Skupio_acute_morphine		●				●									●		●
2579	Coffey_acute_morphine	●					●	●	●			●			●		

B

Frontal cortex

357, 350, 280, 60, 28, 11, 8, 3, 1, 1

Gene set size	Study										
49	Kuntz-Melcavage (rats)			●				●	●		
333	Mendez (human)		●				●		●	●	
340	Liu (rats)	●			●			●			●
1148	Seney (human)		●		●	●				●	●

C

Number of genes identified as
DE across 24 studies in 5 main
brain regions

4940, 1509, 418, 117, 34, 18, 5, 3, 2, 0, 1

D Sources of variability (24 studies in 5 brain regions)

Species — ** — inter (164), intra (112)
Structures — *** — inter (196), intra (80)
Treatment chronicity — ** — inter (128), intra (148)
Type of opioid — ns — inter (78), intra (93)
Strain — *** — inter (25), intra (66)
Timepoint after exposure — *** — inter (331), intra (75)
Article — *** — inter (269), intra (7)

Figure 6