Evolution of mate harm resistance in females from *Drosophila melanogaster* populations selected

for faster development and early reproduction

Tanya Verma[1], email id: tanya17@iiserbpr.ac.in

Susnato Das[2], email id:daskpa201@gmail.com

Saunri Dhodi-Lobo[3], email id: dhodi.lobo@students.iiserpune.ac.in

Ashish Mishra[4], email id: ashish.akm1999@gmail.com

Soumi Bhattacharyya[5] email id: soumisayantan2000@gmail.com

Bodhisatta Nandy[1]*

[1]Indian Institute of Science Education and Research Berhampur, Odisha, India

[2]Institute of Animal Welfare and Animal Husbandry, Friedrich-Loeffler-Institut, Celle, Germany

[3]Indian Institute of Science Education and Research Pune, Maharashtra, India

[4]National Institute of Science Education and Research, Odisha, India

[5]Indian Institute of Science Education and Research Kolkata, West Bengal, India

*Corresponding author, email address: nandy@iiserbpr.ac.in, ORCID iD: 0000-0002-9588-0316

Authors Contribution:

BN and TV conceptualized the study, designed the experiments, analysed and interpreted the results, and

prepared the manuscript. SD helped with some parts of manuscript writing at earlier stage. TV, SD, SDL,

AM, SB executed the experiments, including data collection.

Acknowledgements:

The study was financially supported by a research grant from Department of Science and Technology,

Govt. of India (INSPIRE Faculty award, Grant no. DST/INSPIRE/04/2013/000520). We thank Subhasish

Halder and Purbasha Dasgupta for help in the experiments and Rabisankar Pal for help in data analysis

and plotting. We thank Sadanjeet Kumar Kar for help in experimental observations. TV thanks Indian

Institute of Science Education and Research, Berhampur for financial support in the form of Junior and

Senior Research Fellowship. SD thanks Scholarship for Higher Education, Govt. of India for financial

support in form of INSPIRE-SHE fellowship. SD-L thanks IASc, INSA and NASI, Government of India

for financial support in the form of IASc-INSA-NASI Summer Research fellowship. AM thanks

Department of Atomic Energy, Govt. of India for financial support in the form of DISHA scholarship.

Abstract

Interlocus sexual conflict is predicted to result in sexually antagonistic coevolution between male

competitive traits, which are also female-detrimental, and mate harm resistance (MHR) in females.

Though such antagonistic coevolution has been experimentally shown, little is known about its

connection with life-history evolution. Here, we investigated the evolution of MHR in a set of

experimentally evolved populations selected for faster development and early reproduction. Previously

we showed the reduction of harming ability of males in these populations. Here, we measured mortality

and fecundity of females in these populations and those of their matched controls under different male

exposure conditions. As predicted by the coevolution theory, we observed that the evolved females were

more susceptible to mate harm - suffering from significantly higher mortality under continuous exposure

to control males. We used fecundity data to show that this higher mortality in evolved females is unlikely

due to cost of reproduction *per se*.

Keywords: Interlocus sexual conflict, sexually antagonistic coevolution, life history evolution, cost of

reproduction, post-mating response in females

Introduction

Evolutionary interests of sexes are often not aligned leading to evolutionary conflict over traits with sexually antagonistic fitness effects (1). In one form of such conflict, commonly referred to as interlocus sexual conflict, expression of male-benefitting traits (for example, courtship and mating behavioural traits) reduces female fitness as an incidental side effect (2–4). Effect of such antagonistic male effect, often referred to as mate harm, constitutes a significant portion of female cost of reproduction (5). Theories predict evolution of female counter measures that allow females mitigate such cost of mate harm – potentially resulting in sexually antagonistic coevolution (6). While interlocus conflict has been reported in a wide diversity of animals in the form of mate harm (7,8) male-female coevolution has also been shown in a number of studies (9,2,10–17). Though sexual conflict has emerged as one of the most exciting areas of investigation, with an ever increasing body of empirical evidence and a solid theoretical framework, it is now very important to integrate it in the broader framework of life history evolution as conflict related traits including MHR can only evolve within the life-history constrains such as, cost of reproduction and reproductive lifespan (18).

In *Drosophila melanogaster*, a classical model for interlocus conflict, exposure to males increases female mortality due to persistent courtship (19), and effects of the seminal fluid proteins transferred during a copulation (5,20). Resistance traits, collectively hereafter referred to as mate-harm resistance (MHR), can include behavioural and physiological traits such as, mating rejection (21), avoidance of male encounter by finding refuges (22), alteration in production of proteins and peptides that respond to male seminal fluid proteins (23). There is now ample evidence for evolution of female resistance (11,24). If there is a substantial cost of expressing MHR, life-history theories would predict a trade-off between these and other traits such as somatic maintenance. However, if expression of MHR leads to reduction in reproductive cost in general, detecting cost of MHR *per se* might be difficult. Alternatively, cost of MHR can be indirectly inferred from the evolutionary reduction in MHR when selection on such traits is removed (11,24).

Mital et al. (2021) reported a reduction in MHR in females in populations of *D. melanogaster*

experimentally evolved for faster development and early reproduction, possibly due to a change in

breeding system to near-monogamy (25,26). In a similar study, we showed reduction in mate mating

behaviour, including mate harming ability (27). Our results seem to suggest roles of body size evolution

as well as that of the changes in the breeding ecology, most notably extreme shrinkage of effective adult

reproductive life (27). Here we extend our investigation to incorporate the evolution of MHR in females

of the faster developing and early reproducing populations, and their controls.

If maintaining MHR is costly, populations should divest in MHR if it does not have any fitness benefit.

Therefore, since selection for early reproduction resulted in reduced interlocus conflict (27), MHR in

females from these populations should evolve reduced MHR. Such females should thus be more

susceptible to mate harm. To test this theory, we measured mortality rates under virgin (i.e., non-

reproducing), single mating (i.e., limited male exposure, but reproducing), and continuous male exposure

(i.e., reproducing under constant male presence) of evolved and control females. To differentiate between

the cost of male exposure *per se* and cost of reproduction, we also measured female reproductive output.

The above-mentioned traits were measured using a standard control population male to equalise the male

effects.

Materials and methods

We used a set of experimentally evolved *D. melanogaster* populations, ACOs and their paired controls,

COs to conduct the experiments. Detailed information on these populations can be found in the

supplementary information. Briefly, ACO populations are subjected to selection for faster pre-adult

development and early reproduction, and have an extremely short effective adult life of 24-36 hours.

ACOs were derived from COs in 1989 by Chippindale et al. (1997). While COs have a discrete

generation cycle of 28 days, ACOs are maintained on a 9-day discrete generation cycles. There are five

replicate ACOs, derived from five replicates of COs. By the time the following experiments were

conducted, ACO populations had evolved for >1200 generations. Driven by strong experimental

selection, ACO populations have evolved markedly faster pre-adult development, smaller body size, and

shorter adult lifespan (28,29). For the purpose of this investigation, three replicates of the population sets

were randomly selected. All assays described below were conducted with ACO_1, ACO_2, ACO_3, ACO_4

and their paired control CO populations. All experimental flies were generated from a subset of the stock

populations, after one generation of common garden rearing to equalise non-genetic parental effects.

<u>Mate harm resistance assay setup:</u>

In *D. melanogaster*, MHR can be measured by comparing female mortality under limited and extended

exposure to males (11,24,30). Females with lower MHR are expected to show sharper increase in

mortality under extended male exposure compared to those with higher MHR.

Assay vials were set up with 1-2 day old virgins (see supplementary information). The 45 vials, each

having ten virgin females from a population, were randomly assigned to three assay conditions - virgin,

single exposure, and continuous exposure such that each assay condition consisted of an initial count of

15 vials. The experimental vials were set up by introducing flies in fresh food vials. For the virgin assay

condition, females were held without any male exposure for the entire assay duration. Single exposure

and continuous exposure vials were set up by introducing 10 virgin control (i.e., CO) males along with

the ten experimental females in a fresh food vial. For the single exposure vials, matings were manually

observed and after a single round of mating, sexes were separated under CO_2-anaesthesia to discard the

males. The females were then returned back in the same vials. For the continuous exposure treatment,

males and the females were kept together in the same vials till the end of the assay. To ensure similar

handling of flies across all treatments, flies under virgin and continuous exposure treatments were also

exposed to anaesthesia. Throughout the experiment, except sorting of sexes, all other fly handling were

done without anaesthesia. All vials were maintained for twenty days and the flies in each vial were

flipped to fresh food vials every alternate day. For all vials regardless of assay condition, mortality in

females was recorded daily until day 20. Our previous observation suggests that the difference in effects

of mate harm on female mortality can be detected in the first twenty days of adult life (27). In addition,

this period represents early-to-mid life in this system, most relevant to both control (CO) and

experimental (ACO) population ecology. Further, the difference in age-dependent mortality rate between

the two selection regimes has minimal impact on mortality difference within this duration (data not

shown). Dead flies were aspirated out during vial-to-vial flips. In the continuous exposure assay

condition, in case a female fly was found dead in a vial, along with the dead female, a male was also

removed from the same vial to maintain a 1:1 sex ratio.

Female fecundity was recorded twice a week starting from the onset of the assay until day 20 (i.e., day 1,

3, 6, 9, 12, 15, 18, and 20). On each of these days, flies were flipped to a fresh food vial (hereafter

referred to as a fecundity vial) and were left undisturbed for 24 hours. Following this, the flies were

transferred to a fresh food vial, while the fecundity vial was frozen immediately to prevent further

development of the already deposited eggs. The number of eggs laid in a fecundity vial was counted

under microscope. Fecundity count was carried out for single exposure and continuous exposure

treatments. Per capita fecundity, calculated as total number of eggs in a vial divided by the number of

females alive in that vial on that given day, from individual vials was taken as the unit of analysis. A few

vials were removed from the assay for a variety of reasons, including accidental escape, a few females

failing to mate, etc. The final sample size throughout the entire experiment was 13-15 vials per

population.

<u>Data analysis:</u>

Female survivorship was analysed using Cox's Proportional hazards model. Selection regime (levels:

ACO and CO) and assay condition (levels: virgin, single exposure and continuous exposure) were

modelled as fixed factor and block as random factor using R package Coxme (31). Cox partial likelihood

(log-likelihood) estimates across selection regimes were compared.

Per capita fecundity was analysed in two ways. Cumulative fecundity i.e., per capita fecundity pooled

across all eight age classes was analysed to compare to total early-to-mid life reproductive output of the

females. In addition, age-specific per capita fecundity was analysed to compare the age related pattern of

reproduction. The latter was done only for the continuous exposure set to minimise model complication.

Both cumulative female fecundity and age-specific fecundity data were square root transformed before

analysis. A linear mixed effect model was fitted to the transformed data. `lme4` package (32) and

`lmerTest` (33) in R version 4.2.1 (R Core Team, 2022). In the cumulative fecundity model, selection

regime (levels: ACO and CO), assay condition (levels: single exposure and continuous exposure) and

their two-way interactions as fixed factors, block as a random factor. In the analysis of age-specific per

capita fecundity, selection regime and age (levels: 1, 3, 6, 9, 12, 15, 18, 20) were the fixed factors, and

block and all interaction terms involving block were modelled as random factors. All models are

mentioned in the supplementary information. Post-hoc pairwise comparisons using Tukey's HSD method

were performed with the package `Emmenas` (34). The ANOVA table was obtained following

Satterthwaite's method using type III sum of squares.

Results

Cox partial likelihood estimates suggested that the effects of selection regime, assay condition, and

selection regime $\times$ assay condition interaction on female mortality were significant (Figure 1, Table 1).

Pairwise comparisons indicated a significant difference in survivorship of ACO and CO females only

under continuous exposure, with ACO females more than 9.5 times likely to succumb compared to CO

females (estimated hazard ratio: 9.83).

The effects of selection regime and assay condition on cumulative fecundity were significant (Table 2).

While females under continuous exposure had significantly higher fecundity regardless of the selection

regime, cumulative fecundity of ACO females was 27% less than that of the control CO females (Figure

2a). Age-specific fecundity analysis indicated significant effects of selection regime, and age (Figure 2b).

However, we found a two-way and a three-way interaction term involving random block to be significant

(Table S2). Hence, we analysed each block separately (see supplementary information, Table S1).

Though across blocks the age-specific pattern seemed to vary, CO females generally showed higher per-

capita fecundity in most age points (Figure S1). Fecundity on day 1 was of particular interest as ACO

maintenance regime selects for fecundity at this age. Hence, we analysed day 1 fecundity separately,

using a linear mixed model similar to that used to analyse cumulative fecundity. The results indicated

significant effects of selection regime and assay condition, with CO females consistently showing higher

fecundity (Table 2, Figure S2).

Discussion

Our results suggest that selection to faster development and early reproduction has led to the evolution of

sexually antagonistic traits in both sexes. The evolved ACO population males were previously shown to

be significantly less harming to their mates (27). Results of our MHR assay reported here suggest that

ACO females are significantly more susceptible to continued male interaction. When held with males,

ACO females showed close to ten times higher mortality rate compared to that of the control (CO)

females in the same condition. Such higher mortality is unlikely due to the difference in baseline rate of

ageing as we did not find any difference in mortality rate of non-reproducing, and singly mated

reproducing females of the two selection regimes. Further, ACO females were found to be consistently

less fecund, regardless of the length of male exposure, and age. Hence, higher mortality rate of ACO

females mentioned above is unlikely due to an increased cost of reproduction *per se* but a result of

reduction of MHR.

If MHR is costly to express, it is expected to be constrained by the resource availability (35). Females in

resource deprived condition should therefore be limited in terms of their ability to resist mate harm. Such

condition dependence of MHR has been recently demonstrated (17,36). In addition, for reproducing

females, the cost of producing progeny can further constrain resources available for other physiological

processes - potentially making them vulnerable to stresses including mate harm. The evolved ACO

females in our study are small in size (see supplementary information), and can thus be expected to be

resource limited (37). However, they have a lower reproductive rate - hence, lower absolute investment in

reproduction. Though it is difficult to assess relative reproductive investment, as evident from our data

from the single mating treatment, there appears to be a baseline reduction in reproductive rate of ACO

females. However, evidently this baseline difference in reproduction did not result in mortality rate

difference, which is only evident under extended male exposure. In addition, there was no evidence that

this difference in reproductive investment between the evolved ACO and control CO females was higher

under continuous male presence. Hence, it is very unlikely that observed differences in susceptibility is a

mere reflection of the size difference of the experimental females. Our conclusions are also in line with

those of Mital et al. (2021) who used phenocopied females to demonstrate the size independent reduction

in MHR.

Several experimental evolution studies have shown the evolution of MHR (2,10–14,16,17,24,38). Of

these, only two have directly connected evolution of conflict related traits to life history traits such as,

condition, adult lifespan, development time, and size (17,39). Though evolution of MHR is important for

a population's survival (40), continuation of sexual selection (41), and maintenance of genetic variation

(42), it cannot evolve in the vacuum of sexually antagonistic traits only. Our results is an important

addition to the growing list of evidences suggesting that sexual conflict is subjected to a typical eco-

evolutionary feedback process. Herein, breeding ecology sets the stage of sexual conflict and drives

antagonistic coevolution between sexes, but life history affects such evolution by (a) setting physiological

and genetic constraints, and (b) constraining breeding ecology. Hence, selection for life history traits such

as, lifespan, reproductive schedule etc. should be important drivers of sexually antagonistic coevolution as

such selection can impact breeding ecology and offset the fitness premium on sexually antagonistic traits.

References

1. Hosken DJ, Archer CR, Mank JE. Sexual conflict. Curr Biol. 2019;29(11):R451–5.

2. Holland B, Rice WR. Experimental removal of sexual selection reverses intersexual antagonistic coevolution and removes a reproductive load. Proc Natl Acad Sci. 1999;96(9):5083–8.

3. Pitnick S, García–González F. Harm to females increases with male body size in Drosophila melanogaster. Proc R Soc Lond B Biol Sci. 2002;269(1502):1821–8.

4. Arnqvist G, Rowe L. Sexual conflict. Vol. 31. Princeton university press; 2005.

5. Chapman T, Liddle LF, Kalb JM, Wolfner MF, Partridge L. Cost of mating in Drosophila melanogaster females is mediated by male accessory gland products. Nature. 1995;373(6511):241–4.

6. Rice WR, Holland B. The enemies within: intergenomic conflict, interlocus contest evolution (ICE), and the intraspecific Red Queen. Behav Ecol Sociobiol. 1997;41(1):1–10.

7. Koene JM. Sexual conflict in nonhuman animals. 2012;

8. Pischedda A, Stewart AD. Sexual conflict. 2016;

9. Rowe L, Arnqvist G, Sih A, Krupa JJ. Sexual conflict and the evolutionary ecology of mating patterns: water striders as a model system. Trends Ecol Evol. 1994;9(8):289–93.

10. Martin OY, Hosken DJ. Costs and benefits of evolving under experimentally enforced polyandry or monogamy. Evolution. 2003;57(12):2765–72.

11. Wigby S, Chapman T. Female resistance to male harm evolves in response to manipulation of sexual conflict. Evolution. 2004;58(5):1028–37.

12. Crudgington HS, Beckerman AP, Brüstle L, Green K, Snook RR. Experimental removal and elevation of sexual selection: does sexual selection generate manipulative males and resistant females? Am Nat. 2005;165(S5):S72–87.

13. Maklakov AA, Fricke C, Arnqvist G. Sexual selection affects lifespan and aging in the seed beetle. Aging Cell. 2007;6(6):739–44.

14. Michalczyk \Lukasz, Millard AL, Martin OY, Lumley AJ, Emerson BC, Gage MJ. Experimental

evolution exposes female and male responses to sexual selection and conflict in Tribolium

castaneum. Evol Int J Org Evol. 2011;65(3):713–24.

15. Nandy B, Gupta V, Sen S, Udaykumar N, Samant MA, Ali SZ, et al. Evolution of mate-harm,

longevity and behaviour in male fruit flies subjected to different levels of interlocus conflict. BMC

Evol Biol. 2013;13(1):1–16.

16. Hollis B, Koppik M, Wensing KU, Ruhmann H, Genzoni E, Erkosar B, et al. Sexual conflict drives

male manipulation of female postmating responses in Drosophila melanogaster. Proc Natl Acad Sci.

2019;116(17):8437–44.

17. Rostant WG, Fowler EK, Chapman T. Sexual Conflict Theory: Concepts and Empirical Tests. In:

The Sage Handbook of Evolutionary Psychology: Foundations of Evolutionary Psychology. SAGE

Publications; 2020. p. 241–59.

18. Bonduriansky R, Maklakov A, Zajitschek F, Brooks R. Sexual selection, sexual conflict and the

evolution of ageing and life span. Funct Ecol. 2008;443–53.

19. von Schilcher F, Dow M. Courtship behaviour in Drosophila: sexual isolation or sexual selection? Z

Für Tierpsychol. 1977;43(3):304–10.

20. Wolfner MF. Tokens of love: functions and regulation of Drosophila male accessory gland products.

Insect Biochem Mol Biol. 1997;27(3):179–92.

21. Cook R, Connolly K. Rejection responses by female Drosophila melanogaster: their ontogeny,

causality and effects upon the behaviour of the courting male. Behaviour. 1973;44(1–2):142–65.

22. Byrne PG, Rice GR, Rice WR. Effect of a refuge from persistent male courtship in the Drosophila

laboratory environment. Am Zool. 2008;48(2):e1–e1.

23. Yapici N, Kim YJ, Ribeiro C, Dickson BJ. A receptor that mediates the post-mating switch in

Drosophila reproductive behaviour. Nature. 2008;451(7174):33–7.

24. Nandy B, Gupta V, Udaykumar N, Samant MA, Sen S, Prasad NG. Experimental evolution of female

traits under different levels of intersexual conflict in Drosophila melanogaster. Evolution.

2014;68(2):412–25.

25. Mital A, Sarangi M, Nandy B, Pandey N, Joshi A. Shorter effective lifespan in laboratory populations

of D. melanogaster might reduce sexual selection. Behav Ecol Sociobiol. 2022;76(4):1–12.

26. Mital A, Sarangi M, Dey S, Joshi A. Evolution of lower levels of inter-locus sexual conflict in D.

melanogaster populations under strong selection for rapid development. BioRxiv. 2021;

27. Verma T, Mohapatra A, Senapati HK, Muni RK, Dasgupta P, Nandy B. Evolution of reduced mate

harming tendency of males in Drosophila melanogaster populations selected for faster life history.

Behav Ecol Sociobiol. 2022;76(6):1–15.

28. Chippindale AK, Alipaz JA, Chen HW, Rose MR. Experimental evolution of accelerated

development in Drosophila. 1. Developmental speed and larval survival. Evolution.

1997;51(5):1536–51.

29. Rose MR. Passananti HB, Matos M. Methuselah Flies. Singapore: World Scientific Publishing; 2004.

30. Jiang PP, Bedhomme S, Prasad NG, Chippindale A. Sperm competition and mate harm unresponsive

to male-limited selection in Drosophila: an evolving genetic architecture under domestication. Evol

Int J Org Evol. 2011;65(9):2448–60.

31. Therneau T. Coxme: mixed effects Cox models. R package. Version; 2012.

32. Bates D, Kliegl R, Vasishth S, Baayen H. Parsimonious mixed models. ArXiv Prepr

ArXiv150604967. 2015;

33. Kuznetsova A, Brockhoff PB, Christensen RH. lmerTest package: tests in linear mixed effects

models. J Stat Softw. 2017;82:1–26.

34. Lenth RV. Least-squares means: the R package lsmeans. J Stat Softw. 2016;69:1–33.

35. Adler MI, Bonduriansky R. Sexual conflict, life span, and aging. Cold Spring Harb Perspect Biol.

2014;6(8):a017566.

36. Iglesias-Carrasco M, Jennions MD, Zajitschek SR, Head ML. Are females in good condition better

able to cope with costly males? Behav Ecol. 2018;29(4):876-884.

https://doi.org/10.1093/beheco/ary059.

37. Chippindale AK, Leroi AM, Kim SB, Rose MR. Phenotypic plasticity and selection in Drosophila

life-history evolution. I. Nutrition and the cost of reproduction. J Evol Biol. 1993;6(2):171–93.

38. Mital A, Sarangi M, Dey S, Joshi A. Evolution of lower levels of inter-locus sexual conflict in D.

melanogaster populations under strong selection for rapid development. BioRxiv. 2021;

39. Mital A, Sarangi M, Nandy B, Pandey N, Joshi A. Shorter effective lifespan in laboratory populations

of D. melanogaster might reduce sexual selection. bioRxiv. 2022;2021–04.

40. Rankin DJ, Dieckmann U, Kokko H. Sexual conflict and the tragedy of the commons. Am Nat.

2011;177(6):780–91.

41. Snow SS, Alonzo SH, Servedio MR, Prum RO. Female resistance to sexual coercion can evolve to

preserve the indirect benefits of mate choice. J Evol Biol. 2019;32(6):545–58.

42. Härdling R, Karlsson K. The dynamics of sexually antagonistic coevolution and the complex

influences of mating system and genetic correlation. J Theor Biol. 2009;260(2):276–82.

326 Figure1: Survivorship curves obtained from Cox proportional hazard analysis on the mortality of ACO (red line) and

327 CO (dark cyan line) regime females held under virgin (V), single exposure (SE) and continuous exposure (CE)

328 condition for 20 days during the assay. The differences between survivorship ACO and CO females were found to

329 be nonsignificant under virgin and SE conditions. Under CE condition, ACO females showed significantly higher

330 mortality rate.

Figure 2: Results from the fecundity measurements. The panel shows (a) cumulative fecundity per capita,

and (b) age-specific per capita fecundity across ACO and CO selection regime females held with control

(CO) males. Age specific fecundity was analysed only for continuous exposure assay condition to

minimise model complication. Filled circles and error bars represent means, and standard error

respectively. Standard errors are calculated using block means (i.e., population means). Effects of

selection regime, and assay condition on cumulative per capita fecundity were found to be significant.

Effects of selection regime and age were found to be significant on age specific per capita fecundity.

Fixed Coefficients	Hazard Ratios	Lower CI	Upper CI	z	p
Selection Regime ACO	1.111	-0.737	0.947	0.24	**< 0.001**
Assay condition SE	0.501	-1.891	0.509	-1.13	**< 0.001**
Assay condition CE	1.831	-0.264	1.473	1.36	**< 0.001**
Selection Regime ACO: Assay condition SE	1.975	-0.686	2.047	0.98	**< 0.001**
Selection Regime ACO: Assay condition CE	8.702	1.171	3.156	4.27	**< 0.001**
Random effects	**Variance**				
Block	0.0843				

Table 1: Output of mixed effect Cox proportional hazard model for analysis of female survivorship ACO and CO

regime females held under virgin (V), single exposure (SE) and continuous exposure (CE) condition with ancestral

CO males. Hazard ratios are relative to the default level for each factor which is set to 1. The default level for

selection regime was 'CO', and the default level for assay condition was 'Virgin'. Lower CI and Upper CI indicate

lower and upper bounds of 95% confidence intervals. Level of significance was considered to be $\alpha = 0.05$, and

significant p-values are mentioned in bold font style

Trait	Effect	SS	DF	MS	Den	F	p

					DF		
Cumulative fecundity	Selection Regime (SR)	4054.8	1	4054.8	3.008	25.826	**0.015**
	Assay condition	5476.4	1	5476.4	2.987	34.881	**0.010**
	Selection Regime × Assay condition	583.0	1	583.0	3.016	3.713	0.149
Day 1 per capita fecundity	Selection Regime (SR)	301.22	1	301.22	5.649	11.673	**0.016**
	Assay condition	296.34	1	296.34	4.470	11.484	**0.023**
	Selection Regime × Assay condition	129.31	1	129.31	2.077	5.011	0.150
Age-specific per capita fecundity	Selection regime	14.201	1	14.2008	40.027	42.210	**<0.001**
	Age	48.574	7	6.9391	21.019	20.626	**<0.001**
	Selection regime × Age	5.194	7	0.7420	23.470	2.205	0.071
Body size	Selection Regime	1.1065		1.1065	235	1446.6	**<0.001**

Table 2: Summary of the results of linear mixed model (LMM) analysis of cumulative fecundity, day 1 per capita fecundity, age-specific per capita fecundity and body size using `lmerTest` function in R. Selection regime and assay condition in cumulative and day 1 per capita fecundity and regime and Selection regime in body size were modelled as fixed factors and block as a random factor. All tests were done considering α = 0.05 and significant p-values are mentioned in bold font style.