Pan-genome study underling the extent of genomic variation of invasive *Streptococcus pneumoniae* in Malawi

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Impact Statement

We applied pan-genomics principles to characterize diversity in the pneumococcus genome. The main

objective of the research was to highlight pneumococcal virulence genes, a crucial requirement for

vaccine design or drug development. We identified Serotypes 1 and 5 as the most abundant and

invasive pneumococcal strains in Malawi with a short period of nasopharyngeal colonization that could

imply their quickness in infecting the sterile sites in the human body, such as blood and the central

nervous system. They genetically diverged from other serotypes. Genes from an operon known as RD8a

that perform aerobic respiration and ATP synthesis did not exist in serotypes 1 and 5. In contrast, these

serotypes significantly harbored transporters that do not require ATP to function. The pattern of gene

presence-absence in serotypes 1 and 5 could, to some extent, explain why they are invasive with a short

colonization period in the upper respiratory tract.

Abstract

Streptococcus pneumoniae is a common cause of acute bacterial infections in Malawi. Understanding

the underlying molecular mechanisms of the pathogen's invasive behavior is crucial for designing new

therapeutic strategies. We conducted a pan-genome analysis to identify potential virulence genes in *S.*

pneumoniae by comparing the gene pool of the isolates from carriers' nasopharyngeal secretions to

isolates from the blood and cerebrospinal fluid of patients. We identified 56 serotypes in the cohort.

Most serotypes showed a similar prevalence in carriage and disease groups. However, serotypes 1, 5,

and 12F were disproportionately more commonly detected in specimens from patients compared to the

carriage group, as expected, given the known invasiveness of these serotypes. These serotypes were

genetically distinct from other isolates of other serotypes. The lack of genes from a genomic island

known as RD8a was the most pronounced difference between serotypes 1, 5, and 12F compared to

significantly prevalent serotypes in the nasopharynx. Genes in RD8a are involved in binding to epithelial

cells and performing aerobic respiration to synthesize ATP through oxidative phosphorylation. The

absence of RD8a from serotypes 1, 5, and 12F may result in a short duration in the nasopharynx,

theoretically because of decreased capacity to bind to epithelial cells and access free oxygen molecules

for aerobic respiration (needed to maintain the carriage state). Available ATP may be lower in isolates

without RD8a, like serotypes 1, 5, and 12F, causing them to alternatively use phosphotransferase

systems to transport carbohydrates since these transporters use phosphoenolpyruvate as the energy

source instead of ATP. In conclusion, serotypes 1, 5, and 12F, the most prevalent invasive pneumococcal

strains in Malawi, showed a considerable genetic distinction from other strains that may explain their

invasiveness.

Author summary

Despite introducing the pneumococcal conjugate vaccine in 2011, *Streptococcus pneumoniae* remains a

major cause of bacterial infection in Malawi. Whilst some pneumococcal strains harmlessly colonize the

nasopharynx, others find their way into normally sterile sites, such as lungs, blood, and nervous system,

resulting in serious illness. We found specific pneumococcal strains the most invasive in Malawi, they

had a short colonization period and were genetically distinct from other strains. Their remarkable

genetic divergence included the absence of several genes involved in aerobic respiration and the

presence of genes that contribute to ATP-independent carbohydrate transport. The presence or absence

of these genes may be one of the underlying causes of their high invasiveness with a short colonization

period. This hypothesis could make these genes potential candidate targets for future therapeutic

strategies. We hypothesized that the specific gene gain and/or loss in invasive vs. other serotypes are

possibly associated with invasive pneumococcal disease development.

Introduction

Streptococcus pneumoniae, also known as *pneumococcus*, is a Gram-positive, facultatively anaerobic

bacteria and is one of the leading causes of mortality worldwide. Despite reductions in the incidence of

pneumococcal disease in countries that introduced pneumococcal conjugate vaccines (PCV), the

pneumococcal mortality rate is still high. Pneumococci are estimated to be responsible for 317,300

deaths in children aged 1 to 59 months worldwide in 2015 [1]. In the post-PCV13 era, a high disease

burden has still been reported in low-income countries in Africa, such as Malawi [2].

Although pneumococcal nasopharyngeal colonization is asymptomatic, it is a prerequisite for

transmission and disease development [3][4]. Symptoms appear when isolates from the nasopharynx

spread to normally sterile sites such as the lung, blood, and central nervous system. Depending on the

infected organ, *S. pneumoniae* can cause two types of infection: (i) non-invasive (mucosal)

pneumococcal diseases such as otitis media and sinusitis, and (ii) invasive pneumococcal diseases (IPD)

such as bacteremia and meningitis. IPD incidence is highest among infants, the elderly, and

immunosuppressed people, most likely due to their less efficient immune systems [5].

Pneumococci possess several virulence factors, including the polysaccharide capsule, surface proteins,

and enzymes [6][7]. The polysaccharide capsule is the most important virulence factor as it aids the

pneumococcus in evading the immune response during colonization and invasion [8]. Its biosynthesis is

regulated by a cluster of genes in the capsular polysaccharide (*cps*) locus [8][9]. Pneumococcal serotypes

are defined by the type and order of monosaccharides that compose the capsule structure. To date, one

hundred pneumococcal serotypes have been identified [10]. Each strain has a set of capsular synthesis

genes in the *cps* locus that determine its serotype. Immunogenic properties of the capsular

polysaccharide were utilized to develop all pneumococcal vaccines in use, including PCV7, PCV10, and

PCV13 that cover 7, 10, and 13 serotypes, respectively. Although the introduction of PCVs has

significantly reduced the burden of disease caused by vaccine types (VTs), serotype replacement has

increased the non-vaccine types (NVTs) carriage rate and IPD incidence [11][12].

In November 2011, PCV13 was introduced in Malawi, which markedly decreased the health system

burden and rates of severe childhood pneumonia [13]. PCV13 includes serotypes 1, 3, 4, 5, 6A, 6B, 7F,

9V, 14, 18C, 19A, 19F, and 23F. A case-control study in Malawi showed vaccine effectiveness against VT-

IPD of 80·7% [14]. A nasopharyngeal carriage survey conducted in the Karonga district showed that

although the vaccine reduced the VT colonization rate, a moderate level of serotype replacement was

observed among carriers [15]. Moreover, the emergence of antibiotic-resistant pneumococci due to the

overuse of antibiotics is a global concern in the 21st century [16]. To develop new, more effective

vaccines against the vaccine-escape clones and design effective drugs against antibiotic-resistant strains,

it is critical to understand the functions of genes involved in pneumococcal colonization and

pathogenesis. During the past decade, the evolution of high-throughput sequencing technologies has

generated enormous amounts of genomic data that have enabled researchers to perform large-scale

genomic analysis. A well-known example is pan-genome studies. The pan-genome is the entire gene set

in a collection of closely related strains within a specie [17]. Determining the genetic drivers is an active

and promising area of research that can provide insights into pneumococcal disease prevention and

treatment to reduce mortality rates. The pan-genome is useful for analyzing recombinogenic pathogens

such as *pneumococcus*[18]. The recombination level is high in thirteen pneumococcal genomic loci

known as regions of diversity (RDs) numbered from RD1 to RD13, some of which are involved in

virulence [19][20].

In this study, we used whole-genome sequencing (WGS) of 1477 pneumococcal samples recovered from

people residing in Blantyre, Karonga, and Lilongwe in Malawi to address three complementary issues: (i)

identifying the serotype distribution, (ii) understanding the structure of the pneumococcal population,

and (iii) identifying potential invasive driver genes and their biological functions.

Materials and methods

Study design and sample collection

The study was conducted using the samples archived by Malawi-Liverpool Wellcome Trust Clinical

Research Programme (MLW). The study cohort included isolates obtained from asymptomatic carriers

and symptomatic patients that reside in three regions of Malawi including Blantyre in the south,

Karonga in the north, and Lilongwe in the center.

Carriage samples were collected from the nasopharynx of healthy individuals living in the area covered

by the Karonga Health and Demographic Surveillance System (HDSS) [16] and those presenting at Queen

Elizabeth Central Hospital (QECH) in Blantyre [21]. Invasive pneumococcal samples were collected from

the blood and cerebrospinal fluid (CSF) of symptomatic patients visiting QECH and Kamuzu Central

Hospital in Lilongwe[22]. In this study, the term "sterile sites" refers to blood and CSF, and "invasive

samples" are those obtained from the sterile sites.

There were no paired samples. From each person, only one sample was collected, either a

nasopharyngeal swab from a healthy individual or a blood or CSF from symptomatic patients.

Pneumococcal isolates were identified using the procedure previously described [16]. Briefly, isolates

were identified on Blood Agar Gentamicin, based on colony morphology, alpha-hemolysis, and optochin

susceptibility as per standard pneumococcal isolates protocols.

Whole-genome sequencing and quality control

To account for the true diversity of carriages, only a single isolated colony was sequenced and

serotyped, therefore no carriage samples included in this study had multiple serotypes. Archived

samples were sequenced under the Global Pneumococcal Sequencing project and Pneumococcal African

Genomic Consortium at Wellcome Trust Sanger Institute in the United Kingdom. The bacterial DNA was

extracted using QIAamp DNA mini kit and QIAgen Biorobot manufactured by QIAGEN. The whole

148   genome sequencing was accomplished using the Illumina Genome Analyzer II and HiSeq platforms,

149   generating 125 nucleotides paired-end reads. The quality of the reads was evaluated by Fastqc [23].

In-silico serotyping, sequence typing, and quantification of serotype invasiveness

SeroBA version 1.23.4 was used to infer the serotype of samples [24]. SeroBA applies a k-mer method to

determine serotypes directly from the paired-end reads in FASTQ format. Any serotype with a relative

frequency greater than 5% was considered as an abundant serotype. Fisher's exact test was applied to

identify serotypes with a significant presence in the nasopharynx and sterile sites. P-values were

adjusted by the Benjamini-Hochberg method, and serotypes with an adjusted p-value less than 0.01

were considered significant. The odds ratio (OR) was calculated as follows OR = $(ad)/(bc)$, where for

serotype k, a is the number of invasive serotype k, b is the number of carriage serotype k, c is the

number of invasive non-serotype k, and d is the number of carriage non-serotype k. For OR calculation,

zero values were replaced by 0.5. Any abundant serotype with a significant presence in the nasopharynx

was considered to have low invasiveness, and high invasiveness was assigned to abundant serotypes

with a significant presence in the sterile sites. Fisher's exact test was also applied to identify serotypes

whose frequencies changed significantly after introducing PCV13 in 2011 (adjusted p-value < 0.01).

Genome assembly and annotation

164   Genomes were assembled using Velvet Optimiser version 2.2.5 [25]. The assembler was set to produce

165   contigs longer than 500 base pairs. The starting hash value was 61, and the end hash value was 119. The

166   quality of the assembled genomes was evaluated by Quast version 5.2[26]. The assembled genomes

167   were annotated using Prokka version 13.1 [27].

Pan-genome construction

169   The pan-genome for samples was generated using Roary version 3.12.0 [28]. Roary was run to perform

170   the core gene alignments with MAFFT version 7.313 [29]. Genes in the pan-genome were divided into

171 three groups based on their abundance among samples. Genes that existed in 100% of samples were

172 considered as core genes, those that were not core but present in more than 95% of samples were

173 regarded as a soft-core genes, and the rest were considered as accessory genes.

Analysis of the population structure

Since core genes are conserved between all samples, diversity in the core genome exists in the form of

small-scale variations such as single nucleotide polymorphisms (SNPs) and short indels. SNPs and indels

distribution in the core-genome was analyzed to gain insight into the population structure. The genetic

separation between samples was illustrated by the phylogenetic tree built from SNPs and indels in the

alignment of core genes as the phylogeny markers. The core genome alignment was used as input into

the IQ-TREE version 2 [30] to draw a phylogenetic tree visualized by iTol version 3 [31].

Diversity in the accessory genome exists in the form of large-scale gene presence-absence variations.

The R package Nonnegative Matrix Factorization [32] visualized the pan-genome matrix as a gene

presence-absence heatmap. Furthermore, to determine which factors, including isolation sites

(nasopharynx, blood, and CSF), serotypes, geographical locations, and vaccination eras, have the

greatest impact on the gene presence-absence profile of samples, a PCA of gene distribution was

accomplished by the R package MixOmics version 6.20.0 [33].

Gene presence-absence statistical analysis

The phenotypic trait of samples was assigned based on the population structure and isolates'

invasiveness. The gene presence-absence statistical test was performed using Scoary version 1.6.1643

[34]. This tool scores the components of the pan-genome for associations with observed phenotypic

traits while accounting for population stratification. The test was conducted between samples from

different sources and serotypes to find putative virulence factors. Every gene with a Bonferroni

corrected p-value less than 0.05 was reported as a significant gene.

Functional and gene ontology (GO) enrichment analysis

The list of significant genes was submitted to STRING webtool version 11.5 [35] for the functional

enrichment analysis. The STRING database is a network that integrates information from all publicly

available protein-protein interaction databases. The STRING's computational pipeline can predict direct

(physical) and indirect (functional) interactions between proteins. Interactions in STRING are derived

from 5 sources, including (i) genomic context predictions, (ii) high-throughput lab experiments, (iii) co-

expression, (iv) automated text mining, and (v) previous knowledge in databases. Functional enrichment

analysis in STRING uses information from classification systems such as the Kyoto Encyclopedia of Genes

and Genomes (KEGG) [36] and the Protein families database (Pfam) [37]. The tool entitled "Multiple

Sequences" was selected, and the *S. pneumoniae TIGR4* was chosen as the reference organism. STRING

reports the associated enriched pathways with a false discovery rate of less than 0.05.

Results

206   In total, 825 isolates from the nasopharynx of healthy carriers, 368 isolates from the blood of

207   bacteremia patients, and 284 isolates from the CSF of meningitis patients were sequenced. The

208   demographics of the samples are shown in Table 1.

Table 1. Demographics of 1477 pneumococcal isolates collected in Malawi.

Characteristics	Categories	Nasopharynx	Blood	CSF
	< 5	538	165	141
Age (in years)	5-19	109	42	60
	20-40	60	67	50
	> 40	7	24	7
	Unknown	111	70	26
	Female	401	131	111
Sex	Male	313	122	122

	Unknown	111	115	51
	Blantyre	169	357	259
City	Karonga	656	0	0
	Lilongwe	0	0	23
	Unknown	0	11	2
Sampling period		2009-2014	1997-2015	2000-2015

Serotypes 1, 5, and 12F had the highest invasiveness, likely with a short period of

nasopharyngeal colonization

In total, isolates were assigned to 56 serotypes. Regardless of the isolation sites, serotypes 1 (8.7%), 5

(7.8%), 6B (6.6%), 23F (6.3%), and 19F (5.5%) were abundant, with a relative frequency greater than 5%

of the entire cohort. 66% of samples were obtained before the PCV13 rollout in 2011, and 27% were

from the post-PCV13 era (S1 Fig).

Amongst the carriage isolates, serotypes 19F (7.88%), 6B (7.27%), 16F (6.79%), and 23F (5.21%) were

abundant. Serotype 5 (20.38%) followed by 1 (16.58%), 23F (8.42%), and 6B (5.98%) dominated the

blood samples, while serotypes 1 (21.13%), 5 (8.1%), 12F (7.04%), 23F (6.69%), 6A (5.63%), and 6B

(5.28%) were predominant in the CSF. When aggregating blood and CSF groups, the most common

serotypes were 1 (18.56%), 5 (15.03%), 23F (7.67%), 6B (5.67%), and 6A (5%).

As illustrated in Fig 1 and S1 Table, of the abundant serotypes in the blood or CSF, serotypes 1 (p =

1.96E-34), 5 (p = 3.97E-19), and 12F (p=5.29E-06) had a significant presence among patients with a low

frequency in the nasopharynx including serotype 1 (1%), serotype 5 (2%) and serotype 12F (0.6%). Given

that the colonization phase is a pre-requisite for infection, serotypes 1, 5, and 12F may have a short

period of nasopharyngeal colonization before infecting the sterile sites. Since serotypes 1 and 5 were

also the most frequent in the entire cohort, they could be considered as the most prevalent serotypes

with the highest invasiveness. In this study, the terms "significant invasive serotypes" or "hyper-invasive

serotypes" were assigned to serotypes 1, 5, and 12F. Comparatively, abundant serotypes 16F and 19F in

the nasopharynx had a significant presence among carriers suggesting that they have a lower potential

to cause invasive disease. Other frequently detected serotypes, such as 6A, 6B, and 23F, were abundant

and equally distributed among carriers and patients. They may need to colonize the nasopharynx longer

than the hyper-invasive serotypes before infecting the sterile sites. Serotypes 6A, 6B, and 23F were

previously reported as abundant and common serotypes between non-invasive carriers and invasive

infection [38][39].

Fig 1. The distribution of the 56 pneumococcal serotypes assigned to 1477 samples from Malawi. (a) The relative frequency of each serotype in the nasopharynx of carriers, the blood of bacteremia patients, and the CSF of meningitis patients is shown in blue, red, and yellow, respectively (UT: Un-Typeable). (b) The log-transformed odds ratio of the significantly over- and under-abundant serotypes in the sterile sites (blood and CSF). Fisher's exact test was applied to identify serotypes with a significant differential abundance among carriers and patients (nasopharynx and sterile sites) at the significance level of the Benjamini-Hochberg adjusted p-value < 0.01 (BH: Benjamini-Hochberg).

The temporal distribution of the hyper-invasive serotypes with respect to the vaccine rollout time was of

note. The relative frequency of serotype 1 significantly increased (pre-PVC13=8.6% and post-

PCV13=11.3%), serotype 5 significantly decreased (pre-PVC13=11% and post-PCV13=1.5%), and serotype

12F significantly increased (pre-PVC13=1.1% and post-PCV13=5.7%) after the vaccination program in

Malawi. Vaccination might be effective against serotype 5 but did not reduce the burden of IPDs caused

by serotype 1. Serotype 12F, not included in PCV13, may be an invasive strain emerging in the post-

PCV13 era. However, a more extensive dataset with more recent samples is needed to characterize the

long-term PCV13 effects.

High diversity in the pneumococcal pan-genome

For the genome assembly, the average optimized assembly hash value and the average N50 were 96 and

113,986, respectively. The mean assembled genome size was estimated at 2,116,779 nucleotides with a

standard deviation of 106,481. This length is in the range of previously reported *S. pneumoniae* genome

size [40]. The length of the pan-genome was 5,178,167 base pairs, composed of 6,803 genes, including

729 core-(10.7%), 820 soft-core- (12.1%), and 5,254 accessory genes (77.2%). The pan-genome was open

as it did not reach sampling saturation as the number of genes increased as the sample size grew (S2

Fig). In Fig 2, the pan-genome is demonstrated as a gene presence-absence heatmap that shows

serotypes as the strongest determinant of the gene distribution in the pan-genome. The distinction was

evident for the hyper-invasive serotypes 1, 5, and 12F since they clustered in unique clades. The sets of

genes specifically present in different serotypes appear as small blue blocks in the heatmap.

Gene presence-absence heatmap

Specimen sources
- Blood
- CSF
- Nasopharynx

Serotypes
- 1
- 11A
- 12F
- 15B
- 16F
- 19F
- 21
- 23F
- 5
- 6A
- 6B
- others

PCV13 era
- Post-PCV13
- Pre-PCV13
- Unknown

Fig 2. The pan-genome matrix of 1477 pneumococcal isolates from Malawi. The pan-genome is visualized as a gene presence-absence heatmap representing the hierarchical unsupervised clustering of samples based on the distribution of genes in the pan-genome. Each row is a sample, and each column is a gene. A blue dot denotes the presence of each gene. On the right side of the heatmap, the large blue block shows core genes present in all samples. The left side of the heatmap represents the accessory genome along with the clustering bands. In addition to the significant serotypes 1, 5, 12F, 16F, and 19F, other abundant serotypes, including 6A, 6B, and 23F, as well as serotypes with source-based p-value < 0.05, including 21, 11A, and 15B, are also highlighted on the heatmap.

The significant invasive serotypes showed the highest distinction in the core- and accessory-

genome

The maximum-likelihood tree characterized by the distribution of the small-scale variants (SNPs and

indels) in the core-genome identified the hyper-invasive serotypes 1, 5, and 12F as monophyletic

clusters (Fig 3). The distinction of hyper-invasive serotypes that were separated as unique clusters was

higher than other abundant serotypes, such as 6B, 19F, and 23F, which appeared as multiple clusters on

the phylogenetic tree (Fig 3).

Fig 3. The phylogenetic population structure of 1477 pneumococcal samples from Malawi. The phylogenetic tree was built based on the multiple sequence alignment of the core genome using the maximum likelihood method. Colors on the loops show the serotypes, specimen sources (isolation sites), and PCV13 eras. In addition to the significant serotypes 1, 5, 12F, 16F, and 19F, other abundant serotypes, including 6A, 6B, and 23F, as well as serotypes with source-based p-value < 0.05, including 21, 11A, and 15B, are also highlighted on the tree.

265   The PCA of large-scale variants distribution in the accessory-genome (gene presence/absence) showed

266   serotypes 1 and 5 clustered distantly from other strains (Fig 4). Meanwhile, a moderate level of

267   separation was observed for serotypes 12F, 19F, and 23F. This highlighted the distinction of serotypes 1

268   and 5 in their gene content as observed in their SNPs profiles demonstrated by the phylogenetic tree.

Fig 4. The PCA of the gene distribution in the pan-genome of pneumococcal isolates from 1477 Malawians. The PCA of variants (gene presence-absence) in the accessory-genome indicates the influence of (a) specimen sources (isolation sites), (b) serotypes, (c) PCV13 (vaccination) era, and (d) geographical locations on the gene presence-absence profile of pneumococcal isolates in Malawi. Serotypes 1 and 5 were clearly separated from other samples.

Fig 3 and Fig 4 show that factors such as time, locations, and isolation sites (specimen sources) could not

explain the small- and large-scale genetic variants in the pan-genome. Indeed, the serotype of samples

was the best driver of the population structure. Of importance was that the hyper-invasive serotypes

had the maximum core and accessory distances from other strains, which means they are genetically

different from other serotypes. Since these serotypes were associated with disease, their high genetic

distinction may be associated with their invasive potential. Nonetheless, the analysis must rigorously

account for population structure and carefully curate any hits to show statistical association with the

disease across the population. The first thing to point out is that the genetic distinction of serotype 1

and 5 is not observed in 12F. This may be due to the lower number of 12F (n=35) compared to 1 (n=129)

and 5 (n=116). Moreover, the near-perfect separation between serotypes 1 and 5 and the other

serotypes may be just because these strains were hardly found in the carriage group. Therefore, these

patterns may primarily reflect biases in sampling rather than genuine genetic phenomena. To address

these issues, ten samples of each hyper-invasive serotype and the vaccine types involved in PCV13 were

randomly selected from the nasopharynx, blood, and CSF. The PCA of the gene distribution was

repeated for the downsampled dataset and identified the stratification so that each of serotypes 1, 5,

and 12F formed a distinct group clustered distantly from other strains (S3 Fig).

Typically, serotypes are separated from each other since they have different serotype-defining capsule

genes. The issue that must be addressed is why hyper-invasive serotypes cluster more distantly in

contrast with other serotypes. The hypothesis we explored is that serotypes 1, 5, and 12F gained or lost

a list of genes that caused their quick invasiveness. There are other abundant serotypes in the blood and

CSF (such as 6B and 23F), however, they have a similar prevalence in the nasopharynx, which means

they may stay in the nasopharynx longer than the hyper-invasive serotypes.

The gene presence-absence statistical analysis

The following issues could skew the gene presence-absence analysis:

a) The batch effect introduced by geographical locations: 85% of carriage samples were from

295   Karonga, whereas 95% of disease samples were from Blantyre (Table 1). A comparison between

296   carriage and disease groups may only identify the difference between pneumococcal genomes

297   from two geographical locations rather than between the non-invasive and invasive groups.

298   b) Study limitation: The likely presence of invasive serotypes in the carriage group made it unclear

299   which nasopharyngeal samples progressed to disease after collection. Indeed, the carriage

population likely contained invasive serotypes that could bias the test between carriage and

disease samples to identify potential virulence genes.

c) Population structure: The significant abundance of the hyper-invasive serotypes in the patient

group and their highest genetic distinction would skew the test between the carriage and

patient groups. The difference between carriage and disease groups would actually be the

difference between the carriage and hyper-invasive serotypes (and not all strains in the blood

and CSF).

To measure the geographical batch effect, the carriage isolates from Karonga were compared to

carriage isolates from Blantyre. The test did not detect any significant gene in either Blantyre or Karonga

groups, implying that the gene content of the carriage samples from Blantyre and Karonga was similar.

The serotype distributions in the carriage group from Karonga and Blantyre were also similar. Therefore,

the effect of the geographical locations on the pneumococcal genomes was not remarkable. This was

expected as the distance between Karonga and Blantyre is about 830 kilometers, and the race of people

in these cities is similar.

Despite the possible presence of the invasive serotypes in the carriage group (e.g., 6A, 6B, and 23F that

were prevalent among the carriers and patients), comparing the whole carriage and patient groups

could still make sense since an invasive serotype in the nasopharynx may change its genome before

entering the sterile sites. Indeed, the potential invasive serotypes in the nasopharynx were isolated

during their colonization phase when they could exchange genes with other pneumococci or bacterial

species through recombination and horizontal transfer of genes. Thus, the genome of an invasive

serotype in the nasopharynx is not necessarily identical to its genome in the blood and CSF. The

association test was applied to compare the whole carriage and disease group, while the hyper-invasive

serotypes were excluded (location-based analysis). The exclusion of the hyper-invasive serotypes

prevented them from skewing the difference between the carriage and patient groups. The location-

based analysis identified 27 significant genes including 11 genes significantly present in the blood and

CSF and 16 genes in the nasopharynx (Table2, Table 3, and S2 Table).

Table 2. Significant genes (p-value < 0.05) in the blood and CSF (hyper-invasive serotypes were excluded).

ID	Annotation	P-value	Odds ratio
SP_0360	Capsular polysaccharide biosynthesis protein (from RD3)	8.17E-05	4.897059
SP_0358	Capsular polysaccharide biosynthesis protein (from RD3)	8.17E-05	4.897059
SP_0351	Capsular polysaccharide biosynthesis protein (from RD3)	8.17E-05	4.897059
SP_0359	Capsular polysaccharide biosynthesis protein (from RD3)	8.17E-05	4.897059
SP_0357	Capsular polysaccharide biosynthesis protein	0.000175	4.758421
SP_1037	Type II restriction endonuclease BcgI	0.001074	2.878307
SP_1953	Bacteriocin/lantibiotic secretion ABC transporter permease protein	0.001864	4.631892
SP_0535	Putative immunity protein	0.004674	1.988523
SP_1056	Relaxase/Mobilisation nuclease domain (From RD6)	0.012604	2.932153
SP_1656	Hypothetical protein	0.014050	1.963212
SP_0347	Capsular polysaccharide biosynthesis protein (from RD3)	0.020560	1.808997

Table 3. Significant genes (p-value < 0.05) in the nasopharynx (hyper-invasive serotypes were excluded).

ID	Annotation	P-value	Odds ratio
SP_1770	Glycosyl transferase, glyB (from RD10)	0.000132	0.502513
SP_1771	Glycosyl transferase, family 2/family 8 (from RD10)	0.012770	0.560236
SP_1763	Preprotein translocase secY family protein (from RD10)	0.017254	0.563256
SP_1765	Glycosyl transferase, glyF (from RD10)	0.018150	0.564542
SP_0939	Hypothetical protein	0.020740	0.483266
SP_1766	Glycosyl transferase, glyE (from RD10)	0.020740	0.483266
SP_1767	Glycosyl transferase, glyD (from RD10)	0.023196	0.568879
SP_1762	Accessory secretory protein asp1 (from RD10)	0.023196	0.568879
SP_1761	Accessory secretory protein asp2 (from RD10)	0.023196	0.568879
SP_1760	Accessory secretory protein asp3 (from RD10)	0.023196	0.568879
SP_1755	Hypothetical protein	0.031054	0.558554
SP_1757	Glycosyl transferase, glyB (from RD10)	0.031054	0.558554
SP_1764	Glycosyl transferase, glyG (from RD10)	0.023196	0.568879
SP_1768	Conserved hypothetical protein (from RD10)	0.031054	0.558553
SP_1758	Poly(glycerol-phosphate) alpha-glucosyltransferase, tagE (from RD10)	0.031122	0.571941
SP_1759	Preprotein translocase, secA subunit (from RD10)	0.031321	0.574557

The most significant genes in the blood and CSF were from *cps* locus (RD3) may be due to a higher level

of encapsulation during the disease. Other significant genes, such as SP_1953, SP_0535, SP_1037, and

SP_1056, contribute to toxic secretion and recombination. SP_1056 is from a region known as

pneumococcal pathogenicity island 1 (PPI1), located in RD6. This gene encodes a mobilization protein

required for the horizontal transfer of genes and plasmids through bacterial conjugation. SP_1056 forms

the relaxation complex or relaxosome with the help of other enzymes [41].

Significant genes in the nasopharynx (absent from samples in the blood and CSF) were from RD10, which

is known as SecY2A2 island and is responsible for the secretion of pneumococcal serine-rich repeat

protein or PsrP (S4 Fig)[42]. RD10 contains several glycosyltransferases and secretory components that

were significantly absent from the genome of samples collected from blood and CSF.

To explore the divergence of the hyper-invasive serotypes, they were compared to serotypes 16F and

19F, which were significantly present in the nasopharynx (serotype-based analysis). Serotypes 16F and

19F could represent non-invasive strains better than the whole nasopharyngeal population that

potentially contained some invasive serotypes. Indeed, it was a test between serotypes with the highest

and lowest invasiveness to characterize the genomes of serotypes 1, 5, and 12F. The gene gain and loss

profiles in the hyper-invasive serotypes may include components contributing to their virulence and

short colonization period. The serotype-based analysis identified 184, 157, and 186 significant genes in

serotypes 1, 5, and 12F, respectively (S3 Table, S4 Table, and S5 Table) that were much larger than the

number of significant genes identified by the location-based analysis. The functional enrichment analysis

identified the phosphotransferase system (PTS, KEGG ID: spn02060) as over-represented and oxidative

phosphorylation (KEGG ID: spn00190) as under-represented pathways in the hyper-invasive serotypes

(p-value < 0.05).

In total, there were 18 significant genes jointly present in the hyper-invasive serotypes (Fig 5.a),

including elements of the PTSs that transport sucrose and lactose across the membrane (SP_0302,

SP_0303, SP_0304, SP_0305, SP_0306, SP_0308, SP_0309, and SP_0310), bacteriocins (SP_0544 and

SP_1051) and a permease protein (SP_1527). The over-represented pathway (spn02060) was associated

with significant genes that code for PTS transporters involved in carbohydrate metabolism. Seven genes

were unannotated. The PTS transporters genes were also present in a high proportion of abundant

serotypes in sterile sites such as 6B (67%) and 23F (86%).

There were 57 significantly absent genes from serotypes 1, 5, and 12F (Fig 5.a). The most significant

ones were from RD8a, a region composed of two operons, RD8a1 (SP_1315-1324) and RD8a2 (SP_1325-

SP_1331) (S5 Fig). Genes in RD8a1 include eight *ntp* genes that code for the multiunit *V-type*

proton/sodium ATP synthase complex that produces ATP through oxidative phosphorylation in the

presence of a Na$^+$ gradient across the membrane [43]. RD8a2 includes *neuraminidase, N-*

acetylneuraminate lyase (nanA), and *N-acetylmannosamine-6-phosphate epimerase (nanE).* These genes

cleave carbohydrates from the glycoproteins on the surface of epithelial cells. Other genes in RD8a2

encode the Sodium/solute symporter subunits that use Na$^+$ gradient to import the carbohydrates [44].

Symporter means the channel transports the solute (carbohydrates) and co-solute (Na$^+$) in the same

direction using the energy stored in an inwardly directed sodium gradient. In summary, genes in RD8a

operons work together to produce ATP, cleave carbohydrates from the host epithelial cells, and import

them into the bacterial cell. RD8a was not present in any samples assigned to serotypes 1, 5, and 12F,

whereas it was present in other abundant serotypes such as 6A, 6B, 16F, and 19F. The under-

represented pathway (sp00190) in the hyper-invasive serotypes was linked to genes in RD8a. The lack of

RD8a in the hyper-invasive serotypes may relate to their speed at entering the blood and CSF, where

they do not have access to free oxygen molecules that are required for oxidative phosphorylation [45].

Other significant genes absent from the hyper-invasive serotypes were from RD4 and RD7. RD4 is

composed of a group of sortase enzymes that catalyze the assembly of pilins into pili and the anchoring

of pili and other surface proteins to the cell wall [46][47]. The pilus is a hair-like structure associated

with bacterial adhesion and colonization [48]. Due to the possible short colonization period of the

hyper-invasive serotypes, they may not benefit from genes in RD4 that assemble the pilus. Genes in RD7

have remained uncharacterized to date.

There were similarities between hyper-invasive serotypes and other strains in the sterile sites. As

observed for samples in the blood and CSF, capsule genes were significantly present in the hyper-

invasive serotypes, and RD10 was absent from serotypes 1 and 12F, however, this region was fully

conserved in serotype 5. RD10 was also conserved in 100% of serotypes 16F and 19F. The summary of

the gene present-absent analysis is illustrated in Fig 5.b.

Fig 5. The summary of the gene presence-absence analysis. (a) The number of significant genes present-absent in the hyper-invasive serotypes. The gene presence-absence analysis was applied using Scoary to compare the gene pools of the hyper-invasive serotypes and serotypes 16F and 19F. P-values were corrected by the Bonferroni method, and significant genes had an adjusted p-value of less than 0.05. (b) The significant presence and absence of RDs in samples from blood and CSF is shown as a presence-absence heatmap.

As mentioned, serotypes 6B and 23F were abundant in both carrier and patient groups, the test

between serotype 6B from the nasopharynx (n=60) and sterile sites (n=37) and serotype 23F from the

nasopharynx (n=43) and sterile sites (n=50) did not identify any significant gene. The gene content of

these two serotypes in the nasopharynx and sterile sites was similar.

To highlight genes that might assist pneumococci in crossing the blood-brain barrier, a test was

conducted between the samples from blood and CSF. The analysis between whole blood (n=368) and

CSF (n=284), serotype 1 samples from blood (n=60) and CSF (n=61), and serotype 5 samples from blood

(n=75) and CSF (n=23) did not identify any significant genes.

Discussion

The *S. pneumoniae* genome is highly diverse, and only a small fraction of genes is conserved between all

strains. In this species, the pan-genome is open and provides an unlimited gene repertoire for the highly

recombinogenic pneumococci. Changing the *S. pneumoniae* habitat could promote using different

combinations of genes by the organisms to diversify their genome and efficiently respond to

environmental stresses. In this study, the genetic diversity was identified to be very high, with only

10.7% of genes being core. These genes have been conserved in all samples over a long time, at least

from 1997 to 2015, and their presence may be vital for cell survival, making them potential targets for

drug design and vaccine development. In particular, core genes with no SNP in their structure would of

interest. Examples of the conserved core genes identified in this study included SP_1961 (*rpoB*, DNA-

dependent RNA polymerase), SP_0251 (formate acetyltransferase), SP_1891 (*amiA*, Oligopeptide

binding protein), and SP_0855 (*parC*, Topoisomerase IV). The conserved core genes are involved in DNA

transcription and translation.

Serotypes 1, 5, and 12F were significantly prevalent among patients and scarce in the carrier group. This

characteristic was interpreted as their high invasiveness likely with a short period of nasopharyngeal

colonization. On the other hand, serotypes 16F and 19F were significantly found among carriers,

implying that they dominate the nasopharynx with low invasiveness. Most of the other serotypes were

common between carriers and patients. Knowing that pneumococcal virulence strongly depends on the

serotype of isolates, we sought to address why several serotypes were shared across both nasopharynx

and sterile sites. Here, we discuss two possible scenarios that could justify the ubiquitous presence of

some serotypes in both nasopharynx and sterile sites.

The first scenario is related to the colonization of *S. pneumoniae*, which is known as a prerequisite for

virulence[4]. Several samples from the carriage group may be actually the invasive serotypes collected

during their colonization phase. Abundant serotypes such as 6B and 23F that had a similar frequency

amongst carriers and patients need to colonize the upper respiratory tract longer than the hyper-

invasive serotypes before entering the sterile organs. In contrast, the hyper-invasive serotypes 1, 5, and

12F colonize the nasopharynx for a short period and quickly enter the sterile sites.

The second possible scenario relates to the differential gene expression pattern of shared genes in the

ubiquitous serotypes. Although the type-specific *cps* genes were identified in the isolates of both

nasopharynx and sterile sites, the expression pattern of these genes could vary within each strain, which

would contribute to the invasiveness of some strains. Several studies described a cycle of encapsulation

and un-encapsulation amongst pneumococcal strains. Isolates benefit from mutations in the *cps* locus to

either cease or re-start the capsule expression [49]. The lack of a capsule at the epithelial surface

enables the bacterium to expose its surface proteins on the cell wall underneath the capsule and

promote adherence to the host epithelial cells. It has been estimated that 15% of isolates in the upper

respiratory tract are unencapsulated and adhere to the respiratory epithelial cells more efficiently than

encapsulated isolates [50][51]. Lack of capsule also facilitates acquiring virulence and resistance genes

from other isolates. The thick capsule prevents immunoglobulins from interacting with the pathogen

surface proteins during disease. Meanwhile, the negatively charged CPS interferes with the function of

the host phagocytes [52][53]. Taken together, the presence of the *cps* locus in the genome of isolates

assigned to the same serotype does not necessarily reflect the encapsulation of all cells.

Serotypes 1 and 5 can infect all age groups and cause severe IPDs [54]. Serotype 1 is genetically distinct

between different geographical regions [55] and is known as the leading cause of pneumococcal

meningitis in Africa [56][57]. Our findings support previous research showing that serotypes 1, 5, and

12F are the major cause of IPDs in Malawi [58]. Serotype 1 was persistently dominant in pre- and post-

PCV13 eras, serotype 5 was only predominant in pre-PCV3, and serotype 12F emerged after vaccination.

The study also characterized the high genetic distinction of the hyper-invasive serotypes in Malawi by

identifying significant genes that are present or absent in their genome structure compared to other

serotypes. Many of the significant genes present in the nasopharynx and sterile sites were homologous

and had the same function, and there were significant genes with an unknown function (hypothetical

proteins). However, of greatest interest, we did find several significant genes with specific functions that

could explain the difference between the biology of the hyper-invasive serotypes and nasopharyngeal

samples.

Genes from RD8a were not present in the genome of serotypes 1, 5, and 12F. The presence of this

region has been reported to be associated with the virulence of serotypes 6B and 14 in the United States

[59]. We found this region conserved in serotypes 13, 14, 16F, and 19F in Malawi. One may hypothesize

that RD8a may be required by serotypes that colonize the nasopharynx longer than serotypes 1, 5, and

12F. The functions of genes in RD8a strengthen the assumption to some extent that RD8a may be

essential for long nasopharyngeal colonization. RD8a harbors genes such as *neuraminidase*, *nanA,* and

nanE that cleave terminal sialic acid residues from mucoglycans and epithelial glycoconjugates and assist

the pathogen in penetrating the mucus layer and adhering to the epithelial cells in the nasopharynx. In

the meantime, because free carbohydrates are scarce in the upper respiratory tract [60], cleaved sialic

acid can be used as the carbon source for metabolism. During colonization, secretion of the

pneumococcal toxins elevates the level of sodium ions (Na^+) in the nasopharynx [61], the sodium-solute

symporter in RD8a imports a wide variety of substrates with the sodium ions into the cell [62].

Most importantly, the *ntp* gene cluster in RD8a encodes the *V-type sodium ATP synthase* that pumps the

extra sodium ions out of the cell [43] and uses the sodium-motive force for oxidative phosphorylation

and ATP synthesis [63]. Oxidative phosphorylation is the final step of aerobic respiration that requires

free oxygen molecules for ATP synthesis. Pneumococci are facultative anaerobes that can either

perform aerobic or anaerobic respiration with or without oxygen. In the upper respiratory tract, they

access atmospheric oxygen molecules that can be used by *ntp* genes to perform oxidative

phosphorylation. However, genes in RD8a may not be beneficial for hyper-invasive serotypes that

supposedly do not stay in the nasopharynx for long. Thus, the level of aerobic ATP synthesis is

presumably higher in serotypes 13, 14, 16F, and 19F in contrast with serotypes 1, 5, and 12F, which lack

RD8a.

Pneumococci can ferment up to 30 types of carbohydrates, imported mainly by two types of membrane

transporters, including ATP-binding cassette (ABC) transporters and PTS transporters [64]. The major

differences between ABC and PTS transporters are: (i) ABC transporters use energy from ATP, but PTS

transporters use energy from phosphoenolpyruvate, and (ii) ABC transporters do not modify the

imported substrate, but PTS transporters phosphorylate the incoming sugar upon transport. Generally,

ABC transporters require more energy than PTS transporters, albeit they can transport longer and more

complicated carbohydrates [65]. Unlike isolates in the nasopharynx, serotypes 1, 5, and 12F have access

to more simple and free host dietary carbohydrates in the blood and the central nervous system. Due to

a potential lower ATP synthesis level in serotypes 1, 5, and 12F (due to the lack of RD8a), they may

prefer to use PTS transporters to uptake sugars such as fructose and lactose, and that is why genes that

encode PTS transporters are significantly more present in the hyper-invasive serotypes. In addition to

the sugar uptake, PTS transporters regulate several pathways in bacteria, such as gene expression and

communication between cells. Thus, the phenotypic effects of the PTS transporters should not be

limited just to their ability to import carbohydrates [66].

Genes from RD10 were absent from serotypes 1 and 12F. However, they were conserved in serotypes 5,

16F, and 19F. Moreover, the location-based gene presence-absence analysis showed that RD10 was

significantly present in the nasopharyngeal samples compared to samples from sterile sites. These genes

have homology to the general secretion pathway protein B sceA2/Y2 system components in

Streptococcus gordonii that contribute to the secretion of the general secretion pathway protein B

involved in infective endocarditis [67]. In the *S. pneumoniae* genome, the homolog of general secretion

pathway protein B is the PsrP that is transported to the bacterial cell surface by the SecA2/Y2 system

encoded by genes in RD10. Studies conducted on *Streptococcus gordonii* showed that the presence of

SecA2/Y2 promotes the adhesion to epithelial cells in the nasopharynx and erythrocytes in the blood

[68][69], which is why SecA/Y2 is significantly present in nasopharyngeal samples and serotype 5

(abundant in the blood). The presence of the secA2/Y2-like component should also facilitate the export

of pneumolysin that enhances adhesion to the host cell and survival in the blood [70][71].

In conclusion, some of the genes specifically present (absent) in the hyper-invasive serotypes (1, 5, and

12F) may contribute to the invasiveness and low colonization rate. However, experimental verification is

required to confirm the outcome of computational work in this study. The serotype is the best driver of

the pneumococcal population structure, however, the study characterized the genetic divergence of

serotypes 1, 5, and 12F to be much greater than other serotypes. Their significant presence in the blood

and CSF accounted for the most pronounced genomic and functional differences observed between the

nasopharynx and sterile sites. The low frequency of serotypes 1, 5, and 12F among carriers may be due

to their short-term colonization before entering the sterile sites. These invasive serotypes harbor the

elements of PTS transporters while they lack genes from RD8a. In serotype 5, RD10 is highly conserved,

but this region was absent from serotypes 1 and 12F. The study demonstrated that isolation sites do not

significantly influence the genomic structure of pneumococcal isolates. Although a few genes were

associated with the virulence of commonly present serotypes in both nasopharynx and sterile sites, we

speculated that the differences between them could be captured by other high throughput techniques

such as gene expression analysis. In summary, the results shed light on the pneumococcal population

structure and serotypes in Malawi. The unique cluster of significant genes in the hyper-invasive

serotypes and highly conserved core genes could be potential therapeutic targets.

519 Author contributions

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All authors have given consent to participate in the study.

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Conflicts of interest

The author(s) declare that there are no conflicts of interest.

Ethical approval

Not required as the research delas with bacterial samples.

References

[1]   T. D. Swarthout *et al.*, 'High residual carriage of vaccine-serotype Streptococcus pneumoniae

after introduction of pneumococcal conjugate vaccine in Malawi', *Nature Communications 2020*

11:1, vol. 11, no. 1, pp. 1–12, May 2020, doi: 10.1038/s41467-020-15786-9.

[2]   P. Kamthunzi, 'Impact of PCV13 vaccination in Blantyre, Malawi', *Lancet Glob Health*, vol. 9, no. 7,

pp. e893–e894, Jul. 2021, doi: 10.1016/S2214-109X(21)00258-8.

[3]   L. Paixão *et al.*, 'Host glycan sugar-specific pathways in streptococcus pneumonia: Galactose as a

key sugar in colonisation and infection', *PLoS One*, 2015, doi: 10.1371/journal.pone.0121042.

[4]   D. Bogaert, R. de Groot, and P. W. M. Hermans, 'Streptococcus pneumoniae colonisation: The key

to pneumococcal disease', *Lancet Infectious Diseases*. 2004. doi: 10.1016/S1473-3099(04)00938-

7.

[5]   E. Backhaus *et al.*, 'Epidemiology of invasive pneumococcal infections: Manifestations, incidence

and case fatality rate correlated to age, gender and risk factors', *BMC Infect Dis*, 2016, doi:

10.1186/s12879-016-1648-2.

[6]   A. Kadioglu, J. N. Weiser, J. C. Paton, and P. W. Andrew, 'The role of Streptococcus pneumoniae

561   virulence factors in host respiratory colonization and disease', *Nature Reviews Microbiology*.

562   2008. doi: 10.1038/nrmicro1871.

563   [7]   J. N. Weiser, D. M. Ferreira, and J. C. Paton, 'Streptococcus pneumoniae: Transmission,

564   colonization and invasion', *Nature Reviews Microbiology*. 2018. doi: 10.1038/s41579-018-0001-8.

565   [8]   K. A. Geno *et al.*, 'Pneumococcal capsules and their types: Past, present, and future', *Clin*

566   *Microbiol Rev*, 2015, doi: 10.1128/CMR.00024-15.

567   [9]   A. L. Nelson, A. M. Roche, J. M. Gould, K. Chim, A. J. Ratner, and J. N. Weiser, 'Capsule enhances

568   pneumococcal colonization by limiting mucus-mediated clearance', *Infect Immun*, 2007, doi:

569   10.1128/IAI.01475-06.

570   [10]   J. Brown, S. Hammerschmidt, and C. Orihuela, *Streptococcus Pneumoniae: Molecular*

571   *Mechanisms of Host-Pathogen Interactions*. 2015. doi: 10.1016/C2012-0-00722-3.

572   [11]   F. Ganaie *et al.*, 'A new pneumococcal capsule type, 10D, is the 100th serotype and has a large

573   cps fragment from an oral streptococcus', *mBio*, 2020, doi: 10.1128/mBio.00937-20.

574   [12]   D. M. Weinberger, R. Malley, and M. Lipsitch, 'Serotype replacement in disease after

575   pneumococcal vaccination', *The Lancet*. 2011. doi: 10.5455/apd.239006.

576   [13]   D. R. Feikin *et al.*, 'Serotype-Specific Changes in Invasive Pneumococcal Disease after

577   Pneumococcal Conjugate Vaccine Introduction: A Pooled Analysis of Multiple Surveillance Sites',

578   *PLoS Med*, 2013, doi: 10.1371/journal.pmed.1001517.

579   [14]   E. D. McCollum *et al.*, 'Impact of the 13-valent pneumococcal conjugate vaccine on clinical and

580   hypoxemic childhood pneumonia over three years in central Malawi: An observational study',

581   *PLoS One*, 2017, doi: 10.1371/journal.pone.0168209.

582   [15]   N. Bar-Zeev *et al.*, 'Impact and Effectiveness of 13-Valent Pneumococcal Conjugate Vaccine on

583   Population Incidence of Vaccine and Non-Vaccine Serotype Invasive Pneumococcal Disease in

584   Blantyre, Malawi, 2006-2018: Prospective Observational Time-Series and Case-Control Studies',

585   *SSRN Electronic Journal*, Dec. 2020, doi: 10.2139/SSRN.3745169.

586   [16]   E. Heinsbroek *et al.*, 'Pneumococcal carriage in households in Karonga District, Malawi, before

587   and after introduction of 13-valent pneumococcal conjugate vaccination', *Vaccine*, 2018, doi:

588   10.1016/j.vaccine.2018.10.021.

589   [17]   C. Cillóniz, C. Garcia-Vidal, A. Ceccato, and A. Torres, 'Antimicrobial Resistance Among

590   Streptococcus pneumoniae', in *Antimicrobial Resistance in the 21st Century*, Springer, 2018, pp.

591   13–38.

592   [18]   A. Iranzadeh and N. J. Mulder, 'Bacterial Pan-Genomics', in *Microbial Genomics in Sustainable*

593   *Agroecosystems*, Springer, 2019, pp. 21–38.

594   [19]   A. Embry, E. Hinojosa, and C. J. Orihuela, 'Regions of Diversity 8, 9 and 13 contribute to

595   Streptococcus pneumoniae virulence', *BMC Microbiol*, 2007, doi: 10.1186/1471-2180-7-80.

596   [20]   R. Brückner, M. Nuhn, P. Reichmann, B. Weber, and R. Hakenbeck, 'Mosaic genes and mosaic

597   chromosomes-genomic variation in Streptococcus pneumoniae', *International Journal of Medical*

598   *Microbiology*. 2004. doi: 10.1016/j.ijmm.2004.06.019.

599   [21]   A. W. Kamng'ona *et al.*, 'High multiple carriage and emergence of Streptococcus pneumoniae

600   vaccine serotype variants in Malawian children', *BMC Infect Dis*, vol. 15, no. 1, pp. 1–11, Jun.

601   2015, doi: 10.1186/S12879-015-0980-2/FIGURES/7.

[22] C. Chaguza *et al.*, 'Population genetic structure, antibiotic resistance, capsule switching and

evolution of invasive pneumococci before conjugate vaccination in Malawi', *Vaccine*, vol. 35, no.

35 Pt B, pp. 4594–4602, Aug. 2017, doi: 10.1016/J.VACCINE.2017.07.009.

[23] S. Andrews, 'FastQC: A quality control tool for high throughput sequence data.', *Babraham*

Bioinformatics, p. http://www.bioinformatics.babraham.ac.uk/projects/, 2010, doi: citeulike-

article-id:11583827.

[24] L. Epping, A. J. van Tonder, R. A. Gladstone, S. D. Bentley, A. J. Page, and J. A. Keane, 'SeroBA:

rapid high-throughput serotyping of Streptococcus pneumoniae from whole genome sequence

data', *Microb Genom*, vol. 4, no. 7, Jul. 2018, doi: 10.1099/MGEN.0.000186.

[25] S. Gladman and T. Seemann, 'VelvetOptimiser', *Free Software Foundation*. 2008. doi:

10.1016/S0925-8574(99)00040-3.

[26] A. Gurevich, V. Saveliev, N. Vyahhi, and G. Tesler, 'QUAST: Quality assessment tool for genome

assemblies', *Bioinformatics*, 2013, doi: 10.1093/bioinformatics/btt086.

[27] T. Seemann, 'Prokka: Rapid prokaryotic genome annotation', *Bioinformatics*, vol. 30, no. 14, pp.

2068–2069, 2014, doi: 10.1093/bioinformatics/btu153.

[28] A. J. Page *et al.*, 'Roary: Rapid large-scale prokaryote pan genome analysis', *Bioinformatics*, vol.

31, no. 22, pp. 3691–3693, May 2015, doi: 10.1093/bioinformatics/btv421.

[29] K. Katoh and D. M. Standley, 'MAFFT multiple sequence alignment software version 7:

Improvements in performance and usability', *Mol Biol Evol*, 2013, doi: 10.1093/molbev/mst010.

[30] B. Q. Minh *et al.*, 'IQ-TREE 2: New Models and Efficient Methods for Phylogenetic Inference in the

Genomic Era', *Mol Biol Evol*, vol. 37, no. 5, pp. 1530–1534, May 2020, doi:

10.1093/MOLBEV/MSAA015.

[31] I. Letunic and P. Bork, 'Interactive tree of life (iTOL) v3: an online tool for the display and

annotation of phylogenetic and other trees', *Nucleic Acids Res*, 2016, doi: 10.1093/nar/gkw290.

[32] R. Gaujoux, 'An introduction to NMF package', *BMC Bioinformatics*, 2010, doi: 10.1186/1471-

2105-11-367.

[33] F. Rohart, B. Gautier, A. Singh, and K. A. Lê Cao, 'mixOmics: An R package for 'omics feature

selection and multiple data integration', *PLoS Comput Biol*, 2017, doi:

10.1371/journal.pcbi.1005752.

[34] O. Brynildsrud, J. Bohlin, L. Scheffer, and V. Eldholm, 'Rapid scoring of genes in microbial pan-

genome-wide association studies with Scoary', *Genome Biol*, vol. 17, no. 1, p. 238, 2016, doi:

10.1186/s13059-016-1108-8.

[35] D. Szklarczyk *et al.*, 'STRING v11: Protein-protein association networks with increased coverage,

supporting functional discovery in genome-wide experimental datasets', *Nucleic Acids Res*, 2019,

doi: 10.1093/nar/gky1131.

[36] H. Ogata, S. Goto, K. Sato, W. Fujibuchi, H. Bono, and M. Kanehisa, 'KEGG: Kyoto encyclopedia of

genes and genomes', *Nucleic Acids Research*, vol. 27, no. 1. pp. 29–34, 1999. doi:

10.1093/nar/27.1.29.

[37] A. Bateman *et al.*, 'The Pfam protein families database', *Nucleic Acids Res*, vol. 32, no. suppl_1,

pp. D138–D141, Jan. 2004, doi: 10.1093/NAR/GKH121.

[38] Q. Wang *et al.*, 'Serotype distribution of Streptococcus pneumoniae isolated from children

hospitalized in Beijing children's hospital (2013–2019)', *Vaccine*, vol. 38, no. 49, pp. 7858–7864,

2020, doi: 10.1016/j.vaccine.2020.10.005.

[39]   M. Alizadeh Chamkhaleh *et al.*, 'Serotype distribution of Streptococcus pneumoniae among

646   healthy carriers and clinical patients: a systematic review from Iran', *European Journal of Clinical*

647   *Microbiology and Infectious Diseases*. 2020. doi: 10.1007/s10096-020-03963-z.

648   [40]   N. L. Hiller *et al.*, 'Comparative genomic analyses of seventeen Streptococcus pneumoniae

649   strains: Insights into the pneumococcal supragenome', *J Bacteriol*, vol. 189, no. 22, pp. 8186–

650   8195, Nov. 2007, doi: 10.1128/JB.00690-07/SUPPL_FILE/SUPPLEMENTARY_TABLE_3.ZIP.

651   [41]   S. Zhang and R. Meyer, 'The relaxosome protein MobC promotes conjugal plasmid mobilization

652   by extending DNA strand separation to the nick site at the origin of transfer', *Mol Microbiol*,

653   1997, doi: 10.1046/j.1365-2958.1997.4861849.x.

654   [42]   C. Obert *et al.*, 'Identification of a candidate Streptococcus pneumoniae core genome and

655   regions of diversity correlated with invasive pneumococcal disease', *Infect Immun*, vol. 74, no. 8,

656   pp. 4766–4777, 2006, doi: 10.1128/IAI.00316-06.

657   [43]   K. Yokoyama and H. Imamura, 'Rotation, structure, and classification of prokaryotic V-ATPase',

658   *Journal of Bioenergetics and Biomembranes*. 2005. doi: 10.1007/s10863-005-9480-1.

659   [44]   J. Reizer, A. Reizer, and M. H. Saier, 'A functional superfamily of sodium/solute symporters',

660   *Biochim Biophys Acta*, vol. 1197, no. 2, pp. 133–166, Jun. 1994, doi: 10.1016/0304-

661   4157(94)90003-5.

662   [45]   P. Gaudu, 'Aerobic respiration metabolism in lactic acid bacteria and uses in biotechnology', doi:

663   10.1146/annurev-food-022811-101255.

664   [46]   C. Manzano *et al.*, 'Sortase-Mediated Pilus Fiber Biogenesis in Streptococcus pneumoniae',

665   *Structure*, 2008, doi: 10.1016/j.str.2008.10.007.

666 [47] J. Lemieux, S. Woody, and A. Camilli, 'Roles of the sortases of Streptococcus pneumoniae in

667    assembly of the RlrA pilus', *J Bacteriol*, 2008, doi: 10.1128/JB.00379-08.

668 [48] L. Williams, F. Stapleton, and N. Carnt, 'Microbiology, lens care and maintenance', *Contact*

669    *Lenses*, pp. 65–96, Jan. 2019, doi: 10.1016/B978-0-7020-7168-3.00004-0.

670 [49] J. D. Langereis and M. I. de Jonge, ' Non-encapsulated Streptococcus pneumoniae , vaccination as

671    a measure to interfere with horizontal gene transfer ', *Virulence*, 2017, doi:

672    10.1080/21505594.2017.1309492.

673 [50] D. van der Windt, H. J. Bootsma, P. Burghout, C. E. van der Gaast-de Jongh, P. W. M. Hermans,

674    and M. van der Flier, 'Nonencapsulated Streptococcus pneumoniae resists extracellular human

675    neutrophil elastase- and cathepsin G-mediated killing', *FEMS Immunol Med Microbiol*, 2012, doi:

676    10.1111/j.1574-695X.2012.01028.x.

677 [51] U. M. Talbot, A. W. Paton, and J. C. Paton, 'Uptake of Streptococcus pneumoniae by respiratory

678    epithelial cells', *Infect Immun*, 1996.

679 [52] C. J. Lee, S. D. Banks, and J. P. Li, 'Virulence, immunity, and vaccine related to streptococcus

680    pneumoniae', *Crit Rev Microbiol*, 1991, doi: 10.3109/10408419109113510.

681 [53] L. E. Keller, D. A. Robinson, and L. S. McDaniel, ' Nonencapsulated Streptococcus pneumoniae :

682    Emergence and Pathogenesis ', *mBio*, 2016, doi: 10.1128/mbio.01792-15.

683 [54] W. P. Hausdorff, 'The roles of pneumococcal serotypes 1 and 5 in paediatric invasive disease',

684    *Vaccine*, 2007, doi: 10.1016/j.vaccine.2006.09.009.

685 [55] J. E. Cornick *et al.*, 'Region-specific diversification of the highly virulent serotype 1 Streptococcus

686    pneumoniae', *Microb Genom*, vol. 1, no. 2, 2015.

687   [56]   J. Leimkugel *et al.*, 'An outbreak of serotype 1 Streptococcus pneumoniae meningitis in northern

688   Ghana with features that are characteristic of Neisseria meningitidis meningitis epidemics', *J*

689   *Infect Dis*, vol. 192, no. 2, pp. 192–199, 2005.

690   [57]   B. D. Gessner, J. E. Mueller, and S. Yaro, 'African meningitis belt pneumococcal disease

691   epidemiology indicates a need for an effective serotype 1 containing vaccine, including for older

692   children and adults', *BMC Infect Dis*, 2010, doi: 10.1186/1471-2334-10-22.

693   [58]   J. E. Cornick *et al.*, 'Invasive streptococcus pneumoniae in children, Malawi, 2004-2006', *Emerg*

694   *Infect Dis*, 2011, doi: 10.3201/eid1706.101404.

695   [59]   C. Obert *et al.*, 'Identification of a Candidate Streptococcus pneumoniae core genome and

696   regions of diversity correlated with invasive pneumococcal disease', *Infect Immun*, vol. 74, no. 8,

697   pp. 4766–4777, Aug. 2006, doi: 10.1128/IAI.00316-06.

698   [60]   D. M. Walters, V. L. Stirewalt, and S. B. Melville, 'Cloning, sequence, and transcriptional

699   regulation of the operon encoding a putative N-acetylmannosamine-6-phosphate epimerase

700   (nanE) and sialic acid lyase (nanA) in Clostridium perfringens', *J Bacteriol*, 1999.

701   [61]   L. E. Bakeeva, K. M. Chumakov, A. L. Drachev, A. L. Metlina, and V. P. Skulachev, 'The sodium

702   cycle. III. Vibrio alginolyticus resembles Vibrio cholerae and some other vibriones by flagellar

703   motor and ribosomal 5S-RNA structures', *BBA - Bioenergetics*, 1986, doi: 10.1016/0005-

704   2728(86)90115-5.

705   [62]   M. H. Saier, 'Families of transmembrane sugar transport proteins', *Molecular Microbiology*. 2000.

706   doi: 10.1046/j.1365-2958.2000.01759.x.

707   [63]   P. D. Boyer, 'THE ATP SYNTHASE—A SPLENDID MOLECULAR MACHINE', *Annu Rev Biochem*, 2002,

708   doi: 10.1146/annurev.biochem.66.1.717.

709   [64]   A. Bidossi *et al.*, 'A functional genomics approach to establish the complement of carbohydrate

710   transporters in Streptococcus pneumoniae', *PLoS One*, 2012, doi: 10.1371/journal.pone.0033320.

711   [65]   C. M. Buckwalter and S. J. King, 'Pneumococcal carbohydrate transport: Food for thought', *Trends*

712   *in Microbiology*. 2012. doi: 10.1016/j.tim.2012.08.008.

713   [66]   M. H. Saier, 'The Bacterial Phosphotransferase System: New Frontiers 50 Years after Its

714   Discovery', *Journal of Molecular Microbiology and Biotechnology*. 2015. doi: 10.1159/000381215.

715   [67]   B. A. Bensing, B. W. Gibson, and P. M. Sullam, 'The Streptococcus gordonii Platelet Binding

716   Protein GspB Undergoes Glycosylation Independently of Export', *J Bacteriol*, 2004, doi:

717   10.1128/JB.186.3.638-645.2004.

718   [68]   B. A. Bensing and P. M. Sullam, 'Transport of preproteins by the accessory Sec system requires a

719   specific domain adjacent to the signal peptide', *J Bacteriol*, 2010, doi: 10.1128/JB.00373-10.

720   [69]   M. Yamaguchi *et al.*, 'Streptococcus pneumoniae Invades Erythrocytes and Utilizes Them to

721   Evade Human Innate Immunity', *PLoS One*, 2013, doi: 10.1371/journal.pone.0077282.

722   [70]   M. Bandara *et al.*, 'The accessory Sec system (SecY2A2) in Streptococcus pneumoniae is involved

723   in export of pneumolysin toxin, adhesion and biofilm formation', *Microbes Infect*, 2017, doi:

724   10.1016/j.micinf.2017.04.003.

725   [71]   R. Wu and H. Wu, 'A molecular chaperone mediates a two-protein enzyme complex and

726   glycosylation of serine-rich streptococcal adhesins', *Journal of Biological Chemistry*, 2011, doi:

727   10.1074/jbc.M111.239350.

Supporting information

S1 Fig. Characteristics of the 1477 pneumococcal isolates used in the study. (a) The relative frequency of serotypes in the entire cohort, samples were assigned to 56 serotypes. For each sample, the in-silico serotyping was accomplished by SeroBA. (b) Frequency of isolates in the pre- and post-PCV13 eras in Malawi. (c) Frequency of isolates obtained from each specimen source.

S2 Fig. The pan-genome of 1477 pneumococcal samples isolated in Malawi was obtained from 1997 to 2015. The pan-genome is an open pan-genome, which means the number of total genes increases unlimitedly when the sample size grows. The dashed line represents the number of total genes, and the solid line represents the number of conserved genes in the pan-genome.

S3 Fig. The three-dimensional PCA of the gene distribution in the vaccine types. For each serotype and for downsampling, 10 samples were randomly selected from the nasopharynx, blood, and CSF. The PCA was conducted using the R package MixOmics. Hyper-invasive serotypes 1, 5, and 12F clustered separately from other strains.

S4 Fig. Genes in RD10 are absent from serotypes 1 and 12F but conserved in serotype 5, 16F, and 19F. Genes from RD10 encode the components of the secretory system SecA2/Y2 that transports glycoproteins to the bacterial cell surface, which are required for binding to the human proteins on the surface of epithelial cells and erythrocytes.

S5 Fig. RD8a consists of two operons RD8a1 (SP_1315-1324) and RD8a2 (SP_1325-1331). This region is not detected in the significant invasive serotypes 1, 5, and 12F, but it is present in more than 80% of serotype 16F and 19F that significantly dominates the nasopharynx. The important biological processes carried out by these genes are the transport of ions across the membrane and the synthesis of ATP molecules.

S1 Table. Statistical analysis of serotypes' prevalence across specimen sources.

S2 Table. Gene presence-absence analysis (Invasive vs Nasopharyngeal, serotypes 1, 5, and 12F were excluded).

S3 Table. Gene presence-absence analysis (Serotype 1 vs 16F & 19F).

S4 Table. Gene presence-absence analysis (Serotype 5 vs 16F & 19F).

S5 Table. Gene presence-absence analysis (Serotype 12F vs 16F & 19F).

Figure captions

Fig 1. The distribution of the 56 pneumococcal serotypes assigned to 1477 samples from Malawi. (a)

The relative frequency of each serotype in the nasopharynx of carriers, the blood of bacteremia

patients, and the CSF of meningitis patients is shown in blue, red, and yellow, respectively (UT: Un-

Typeable). (b) The log-transformed odds ratio of the significantly over- and under-abundant serotypes in

the sterile sites (blood and CSF). Fisher's exact test was applied to identify serotypes with a significant

differential abundance among carriers and patients (nasopharynx and sterile sites) at the significance

level of the Benjamini-Hochberg adjusted p-value < 0.01 (BH: Benjamini-Hochberg).

Fig 2. The pan-genome matrix of 1477 pneumococcal isolates from Malawi. The pan-genome is

visualized as a gene presence-absence heatmap that represents the hierarchical unsupervised clustering

of samples based on the distribution of genes in the pan-genome. Each row is a sample, and each

column is a gene. A blue dot denotes the presence of each gene. On the right side of the heatmap, the

large blue block shows core genes present in all samples. The left side of the heatmap represents the

accessory genome along with the clustering bands. In addition to the significant serotypes 1, 5, 12F, 16F,

and 19F, other abundant serotypes, including 6A, 6B, and 23F, as well as serotypes with source-based p-

value < 0.05, including 21, 11A, and 15B, are also highlighted on the heatmap.

Fig 3. The phylogenetic population structure of 1477 pneumococcal samples from Malawi. The

phylogenetic tree was built based on the multiple sequence alignment of the core genome using the

maximum likelihood method. Colors on the loops show the serotypes, specimen sources (isolation sites),

and PCV13 eras. In addition to the significant serotypes 1, 5, 12F, 16F, and 19F, other abundant

serotypes, including 6A, 6B, and 23F, as well as serotypes with source-based p-value < 0.05, including 21,

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Fig 4. The PCA of the gene distribution in the pan-genome of pneumococcal isolates from 1477

Malawians. The PCA of variants (gene presence-absence) in the accessory-genome indicates the

influence of (a) specimen sources (isolation sites), (b) serotypes, (c) PCV13 (vaccination) era, and (d)

geographical locations on the gene presence-absence profile of pneumococcal isolates in Malawi.

Serotypes 1 and 5 were clearly separated from other samples.

Fig 5. The summary of the gene presence-absence analysis. (a) The number of significant genes

present-absent in the hyper-invasive serotypes. The gene presence-absence analysis was applied using

Scoary to compare the gene pools of the hyper-invasive serotypes and serotypes 16F and 19F. P-values

were corrected by the Bonferroni method, and significant genes had an adjusted p-value of less than

0.05. (b) The significant presence and absence of RDs in samples from blood and CSF is shown as a

presence-absence heatmap.

Data summary

S6 Table. Samples IDs on European Nucleotide Archive (ENA)

S1 Fig. Characteristics of the 1477 pneumococcal isolates used in the study. (a) The relative frequency of serotypes in the entire cohort, samples were assigned to 56 serotypes. For each sample, the in-silico serotyping was accomplished by SeroBA. (b) Frequency of isolates in the pre- and post-PCV13 eras in Malawi. (c) Frequency of isolates obtained from each specimen source.

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Ensemble Bacteria Streptococcus pneumonia TIGR4 Version 97.1 (ASM688v1) Chromosome: 1,238,537-1,256,136

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