Title

Breakdown in seasonal dynamics of ant communities with land-cover change

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Abstract

Concerns about widespread human-induced declines in insect populations are mounting, yet little
is known about how land-use change modifies the dynamics of insect communities, particularly
in understudied biomes. Here we examine how the seasonal patterns of ant activity, key drivers
of ecosystem functioning, vary with human-induced land cover change on a subtropical island
landscape. Using trap captures sampled biweekly from a biodiversity monitoring network
covering Okinawa Island, Japan, we processed 1.2 million individuals and reconstructed activity
patterns within and across habitat types. We determined that communities inside the forest
exhibited more variability than those in more developed areas. Using time-series decomposition
to deconstruct this pattern, we found that ant communities at sites with greater human
development exhibited diminished seasonality, reduced synchrony, and higher stochasticity
compared to those at sites with greater forest cover. We demonstrate that our results cannot be
explained by variation in either regional or *in situ* temperature patterns, or by differences in
species richness or composition among sites. We conclude that the breakdown of natural
seasonal patterns of functionally key insect communities may comprise an important and
underappreciated consequence of global environmental change that must be better understood
across Earth's biomes.

Keywords

ant, insect, monitoring, Japan, subtropics, synchrony, temporal, variability
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Introduction

Insects comprise 95% of described terrestrial animal species on Earth and are key drivers of a multitude of ecosystem functions and services, including pollination, food provisioning, pest control, water filtration, carbon sequestration, and decomposition (Schowalter, 2013). There is also mounting concern about the possibility of long-term global insect declines due to human-induced environmental change, which can disrupt key functions and services (Goulson, 2019; Wagner, 2020). But while such trends measured over broad timescales are informative for monitoring populations (Dornelas et al., 2013), they overlook changes in natural seasonal patterns that can only be detected at fine temporal resolutions. When these seasonal patterns are broken down by human pressures, it can result in the erosion of biodiversity (Tonkin et al., 2017) and degradation of the ecosystem functions and services on which human societies depend (Ross et al., 2021; Stevenson et al., 2015). For this reason, a better understanding is needed of how fine-scale temporal patterns of insect communities are affected in this era of global change.

Land-cover modification is a major driver of biodiversity loss (Cardinale et al., 2012), and many studies have examined its consequences for insect diversity (e.g., Castro et al., 2018; Corro et al., 2019; Knop, 2016; Oliver et al., 2016; Senapathi et al., 2015). However, the corresponding effects on temporal dynamics of insect communities are not well-known. For example, human-altered land cover with higher densities of alien species can lead to boom-and-bust dynamics that cause high variability (Lester & Gruber, 2016; Simberloff & Gibbons, 2004). At the same time, urbanization can lead to biotic homogenization and reduced community variability over time (Hung et al., 2017; McKinney, 2006). Although these documented effects of land-cover change on insect temporal dynamics differ, investigations into how insect communities are affected are crucial for tropical and subtropical biomes undergoing rapid land-cover change (Janzen & Hallwachs, 2021), and also for islands that have restricted land area, frequent introductions of alien species, and exposure to increasingly extreme weather events (Russell & Kueffer, 2019).

Here, we examine the effects of anthropogenic land-cover change on the fine-scale temporal variability of ant community activity. Ants are a dominant insect group in the many environments they inhabit around the world, underpinning a broad spectrum of ecological interactions and playing a major role in regulating ecosystem function (Hölldobler & Wilson, 1990), which makes them ideal bioindicators for ecosystem change (Andersen & Majer, 2004). Community seasonality is linked to the maintenance of key ecosystem functions and services (Stevenson et al., 2015). However, even though ants can exhibit significant seasonality in tropical and subtropical regions (Basu, 1997; Samways, 1990), little is known about how land-cover change affects ant community seasonality in these diverse biomes.

To address this, we used high-resolution (biweekly) monitoring data on ant activity for subtropical Okinawa Island in southern Japan to examine whether, and how, the temporal variability of ant communities varies with land cover change. We then determined which temporal components of variability (i.e., seasonality, trend, or stochasticity) contribute the most to differences across the gradient (Fig. S1). We also tested whether other potential drivers, including differences across sites in climate seasonality, species richness, and community composition, could be responsible for observed differences in temporal community variability, and additionally examined the roles played by invasive species in moderating these effects.

   Materials and Methods

Study sites and land-cover data

Our study focuses on the subtropical main island of Okinawa in the southern Ryukyu archipelago
of Japan. The island has a historical land-cover gradient, spanning from the minimally developed
north to the more urban south (Fig. 1). Okinawa's extensive Yambaru Forest in the north was
recently recognized as part of Japan's newest Natural World Heritage Site for its high endemism
and biodiversity. Although the island is only 0.3% of Japan's total land area, it provides habitat
to more than one-third of the country's ant species (Terayama et al. 2009, Terayama et al. 2014).
The few studies of ant communities in Okinawa include biodiversity surveys (e.g., Ito et al.,
1998) and comparisons between native and alien ant populations in Yambaru (Katayama &
Tsuji, 2010; Suwabe et al., 2009; Yamauchi & Ogata, 1995). This study, the first to sample
island-wide ant diversity and temporal patterns in Okinawa, uses data from a biodiversity
monitoring system covering the land-cover gradient of the island, administered by the Okinawa
Environmental Observation Network (OKEON) Churamori Project (https://okeon.unit.oist.jp/).
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We used a land-cover map for Okinawa (year 2015) to calculate representative land-cover values
for the 24 OKEON monitoring sites. The sites (pairwise distances: minimum = 389 m, mean =
33 km, maximum = 102 km) are located in or near forested areas across multiple land-cover
types varying from broadly contiguous forest to highly agricultural or urban areas (Fig. 1). To
characterize each site, we calculated the proportion of seven main land-cover classes (forest,
agriculture, urban, grass, sand, freshwater, miscellaneous) around circular buffers with radius 1
km (see Ross et al., 2018 for details). Buffers with this size were chosen to characterize
representative land-cover values for areas surrounding sites without including regions that may
have resident unsampled colonies. Sites near the coast included ocean area within the buffer, so
we used relative proportions for all sites that considered only land area. As correlations were
high between the main land-cover classes of interest (that is, forest, agriculture, and urban; Table
S1), we used principal component analysis (R function *prcomp*) after applying an arcsine
transformation to the proportions to improve normality (e.g., Van Buskirk, 2005) and retained
the first and second axes (variance explained: PC1 = 81% and PC2 = 11%) for use as
explanatory variables. PC1 represents the forested (high) to developed (low; urban and/or
agriculture) gradient, while PC2 represents the rural (high; agriculture and/or grass) to urban
(low) gradient (Fig. S2). All analyses were conducted in R (R Core Group, 2021), and spatial
analyses were conducted with R packages *sf* (Pebesma, 2018) for vector data and *raster*
(Hijmans, 2021) for gridded data.
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Ant activity data

We sampled worker ant activity biweekly with Sea, Land, and Air Malaise (SLAM) traps for two
years and identified samples to species level, resulting in 1,378,324 classified individuals.
Specifically, we sampled one SLAM trap (Large BT1005, MegaView Science Co., Ltd.) at each
of three stations per OKEON site (pairwise distances within sites: mean = 90 m, minimum = 19
m, maximum = 195 m) every two weeks from March 2016 to March 2018 (further details in
Supporting Information A). The stations are predominantly deployed under forest canopy,
though several are in nearby open areas. This sampling method mainly targets terrestrial
browsing species, though does occasionally catch arboreal and subterranean individuals. Insect

count data sampled with passive traps are usually interpreted as a measure of activity, or the rate
at which individuals intersect a point in space, rather than direct estimates of abundance, though
changes in abundance are generally reflected in ant activity data (Kaspari et al., 2022a).
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We additionally categorized all species by invasive status as either native, alien, or uncertain
based on expert opinion and biogeographical databases (Guénard et al. 2017, Janicki et al. 2016,
Terayama et al. 2009, https://www.antwiki.org/, https://www.antweb.org/). Native refers to
species with enough data to establish historical occupancy in Okinawa without human-aided
dispersal from other regions. Alien refers to species that definitively have native ranges in
different regions and have dispersed to Okinawa via human activity. Uncertain species lack
sufficient data to make such determinations.
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Some prevalent species had extreme count outliers, such as *Technomyrmex brunneus*
(representing counts greater than 5,000), likely caused by large colony sizes and/or increased
activity following mating flights, heterospecific competition, or other biological phenomena. We
avoided bias in variability estimates by limiting count values to a conservative threshold of 500
individuals for a given species at a station within a two-week period. We also examined changes
in our results for a range of thresholds. For each species during each sampling period, we
summed counts over the three stations per site to determine site-level activity (Fig. 2), and over
all sites per land-cover group for grouped species-level activity. We then calculated total site
density as the sum of all counts per site.
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Sampled abundance and variability can be correlated due to sampling error alone because of
mean-variance scaling (McArdle & Gaston, 1995). For example, even if species dynamics are
identical across two sites, if we sample fewer individuals from site A than site B (e.g., due to
effects of sampling station placement), site B may have a higher temporal variability simply
because its counts are higher. While in some sense this could reflect the actual variability of
observed species at specific locations (that is, at the stations), we wanted to disentangle
differences in community variability from those attributable to sampling error. Thus, we rarefied
the data from all sites so that the total count (sum of all samples across the two years) for each
site equals the total count at the site with the least abundance (Fig. S1, Supporting Information B;
Gaston & McArdle, 1994). Finally, to determine whether species richness differences could
drive any observed differences in temporal variability, we calculated site-specific total species
richness, and the richness of each invasive status group from the observed and rarefied data,
making extrapolations to account for incomplete sampling using Hill numbers ($q = 0$) with the R
package *iNEXT* (Hsieh et al., 2016).
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Temporal variability and time-series decomposition
We calculated two aspects of temporal variability: functional and compositional (Supporting
Information C). Functional variability refers to activity changes at the aggregate scale (e.g.,
Hillebrand & Kunze, 2020) and is calculated as the coefficient of variability (ratio of the
standard deviation to the mean) of total ant counts across all species per site (McArdle & Gaston,
1995). Compositional variability estimates the variation in community composition over time
and is calculated as temporal beta diversity (Legendre & De Cáceres, 2013) using the R package
adespatial (Dray et al., 2022), which additionally records species' individual contributions to

beta diversity. As we use ant count data, compositional changes would thus represent changes in
relative activity rates of different species in the local species pool, which in turn is a function of
the abundance and patterns of behavior (Kaspari et al., 2022a).
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Next, we used temporal decomposition models to determine the relationships of temporal
variability components with observed patterns. Time series can be decomposed into components
describing different underlying temporal patterns in the data: "seasonal" processes that repeat
cyclically over some temporal scale (i.e., phenology), "trend" that describes directional change,
and "remainder" that represents any residual variation not captured by the other components (i.e.,
short-term stochastic fluctuations; Hyndman & Athanasopoulos, 2021). A community with high
temporal variability may have strong cyclical patterns, increasing or decreasing trends, high
stochasticity, or some combination of these. For temporal datasets spanning many annual cycles,
models that can estimate complex seasonal responses (e.g., wavelet analysis; Tonkin et al., 2017)
are often employed, but for studies spanning few annual cycles at a fine temporal resolution,
simpler models with fewer assumptions and problems with overfitting are preferable. We
estimated additive temporal components for the ant activity data by fitting time-series linear
models with temporal predictors using the R package *fable* (O'Hara-Wild et al., 2021). As our
time-series was short, we used simple predictors consisting of a linear trend and a seasonal signal
approximated with an annual Fourier term with the simplest maximum order (K) of 1—this
models seasonality as a sine wave (Hyndman & Athanasopoulos, 2021). We fitted this model to
the rarefied data at both the site-level and species-level for land-cover groups, then decomposed
the count value at each time-step. For each site, we measured the absolute variance of each
temporal component, but also the relative component variance, defined here as the individual
component variance divided by summed variance of all components (Supporting Information D).
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Finally, as spatial autocorrelation between sites did not affect results (Table S3; Supporting
Information E), we used simple linear models to estimate relationships between land cover and
our focal metrics of temporal variability. Specifically, we fitted models for total ant counts
(before rarefaction), functional and compositional temporal variability, and absolute and relative
variances of time-series components. To explore differences between native and alien species,
we also fitted models separately by invasive status for site richness and summed species'
contributions to beta diversity. We performed model selection on linear combinations of the
predictor variables land-cover PC1 and PC2 using the Akaike Information Criterion corrected for
small sample sizes (AICc).
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Assessing effects of differences in temperature and community composition on observed
seasonality patterns
To determine if other differences among sites may be responsible for any observed variation in
ant community activity and relationships with land cover, we additionally examined differences
in 1) regional and site-level temperature and 2) tested whether seasonality differences persisted
after standardizing community composition between land-cover groups. For (1), we downloaded
regional climate data for the collection period from the Japan Meteorological Agency (JMA)
database (http://www.jma.go.jp/en/amedas_h/map65.html, accessed 04/01/2020) and calculated
temperature means and extremes from the six climate stations that collect temperature data
(Miyagijima, Itokazu, Naha, Ashimine, Nago, Oku). We also collected *in situ* site-level data

throughout the collection period at one station per site to characterize local air and soil
temperature patterns (WatchDog 2900ET Weather Station 1.5 m above ground and SMEC
WaterScout 300 Soil Sensor 10 cm below ground, Spectrum Technologies). With the same
workflow as for the ant activity data, we calculated absolute variance of seasonality for regional
and site-level temperature time-series, then estimated relationships with land-cover PC1 using
linear models.
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For (2), we conducted analyses to test whether differences in community composition alone,
without variation in seasonality within species, could be driving the observed patterns in
seasonality, or alternatively if habitat-dependent dynamics of species occurring in both habitat
types were responsible. We used land-cover PC1 to partition the sites into two groups with eight
sites each—forested and developed (comprising sites with more agricultural and/or urban land
cover)—and retained only those species shared between groups and with total counts greater
than 100. We excluded the eight sites with intermediate values of PC1 to better represent the
most characteristic sites for each land-cover group. To make balanced comparisons between
land-cover groups, we rarefied each species' activity data to the minimum count of that species
between groups (Fig. S1, Supporting Information B). To determine whether community
composition affected our results, we fitted time-series linear models to each species in the land-
cover groups and calculated the relative variance of their seasonality (forested versus developed
sites). As differences between the land-cover groups were not normally distributed (Shapiro-
Wilk test; $p = 0.011$), we used the nonparametric Wilcoxon signed-rank test to determine
whether the same species between groups exhibit significant differences in relative seasonality
variance. Additionally, to determine how temporal mismatch changes with invasive status, we
calculated the synchrony of seasonality for each invasive status between land-cover groups and
standardized them to the same scale. We used the R package *codyn* (Hallett et al., 2016) to
calculate synchrony using the method described by Loreau & De Mazancourt (2008). We then
compared seasonal curves to the average seasonality curve for each land-cover group to visualize
the degree of temporal mismatch by invasive status.
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Results

Our sampling for 2016–2018 resulted in the recovery of >1.2 million individuals (before
thresholding) and 16,595 unique count records at the site-level across 91 ant species. Total count
was less than 5 individuals for 21 species and greater than 10,000 individuals for 15 species, with
a minimum of 1 (*Aenictus ceylonicus*, *Crematogaster* c.f. *matsumurai*, *Ectomomyrmex sp.*,
Erromyrma latinodis, *Hypoponera* sp., *Leptogenys confucii*, *Strumigenys hirashimai*,
Strumigenys mazu) and a maximum of 259,180 (*Tetramorium bicarinatum*). The pattern of ant
activity dropping in the winter and rising in the summer qualitatively diminished as human
disturbance increased (Fig. 2). Total ant activity (after thresholding to 500) had a nearly 23-fold
difference between the site with the minimum count (Yona Forest, $n = 3616$) and that with
maximum count (Sueyoshi Forest, $n = 82,781$). The natural log of total density had a strong
negative relationship with land-cover PC1 ($R^2 = 0.71$, $p < 0.001$), indicating that ant activity
overall was higher in developed areas than forested areas (Fig. 3). Rarefaction resampled all sites
to the minimum site count before analysis.
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The linear models suggested strong relationships with the land-cover PC1 for most metrics of
temporal variability (Figs. 3, 4). Based on model selection via AICc, all models included PC1 as
the sole predictor except the relative variances of seasonality and stochasticity, which also used
PC2 (Table S3). Functional and compositional temporal variability both had strong positive
relationships with PC1 (functional: $R^2 = 0.67$, $p < 0.001$; compositional: $R^2 = 0.60$, $p < 0.001$;
Fig. 3), and these two variabilities also had a strong relationship to each other ($R^2 = 0.62$, Fig.
S3). The species' contributions to beta diversity (Fig. S4) that explained individual species'
impacts on compositional variability were also positively correlated with PC1 for native species
($R^2 = 0.40$, $p < 0.001$), but uncertain ($R^2 = 0.28$, $p < 0.01$) species had a weaker, negative
relationship (alien species showed no relationship). Absolute seasonality and stochasticity
variance had positive relationships with PC1, though that for seasonality ($R^2 = 0.57$, $p < 0.001$)
was stronger than that for stochasticity ($R^2 = 0.15$, $p < 0.05$; Fig. 4). On the other hand, relative
seasonality variance ($R^2 = 0.23$, $p < 0.05$) was positively related to PC1, while relative
stochasticity variance ($R^2 = 0.36$, $p < 0.01$) had a negative relationship (both had $p > 0.05$ for
PC2; Fig. 4). All linear regression results are found in Table S3.
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We found no evidence that differences in species richness, temperature at different scales, or
species composition are candidates as drivers of observed differences in ant community temporal
variability. We found no relationship between PC1 and total species richness (either observed or
extrapolated), and a weak correlation with rarefied richness ($R^2 = 0.15$, $p < 0.05$) due to the loss
of rare species during the rarefaction process, though we consider this unlikely to bias our results
(Fig. S5). We did find strong positive relationships with PC1 for native richness values
(observed: $R^2 = 0.47$, $p < 0.001$; extrapolated: $R^2 = 0.35$, $p < 0.005$; rarefied: $R^2 = 0.61$, $p <$
0.001) indicating that more native species can be found in the more forested sites, and weaker
negative relationships with alien richness values (observed: $R^2 = 0.29$, $p < 0.005$; extrapolated:
$R^2 = 0.20$, $p < 0.05$; rarefied: $R^2 = 0.20$, $p < 0.05$) (Table S3, Fig. S5). Moreover, we found no
clear patterns in the absolute variance of regional temperature seasonality from JMA climate
stations spanning the island from north to south, nor did we find relationships between the
absolute seasonality variance of *in situ* air or soil temperature and PC1 (Fig. S6). Only two sites
had exposed sensors that were not located below canopy cover (Oyama Park and OIST Open
[OYA and OIT, respectively, in Fig. S6]), and these had higher absolute variances than the other
sites. Concerning the tests of different threshold sizes, although we observed some differences,
our results remained similar (Supporting Information F, Fig. S7).
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Lastly, individual species showed differing seasonality between land-cover groups (more
forested and more developed). Comparisons revealed that, in general, the same species have a
higher relative seasonality variance at forested sites than at sites with more human development
($p < 0.005$), though the alien species *Tetramorium lanuginosum* was a notable exception (*Tlanu*
in Fig. 5). While seasonality was stronger in the forested group for all species, when separated by
invasive status we observed higher synchrony in the forested group for native ($n = 11$, forested:
0.75, developed: 0.58) and uncertain species ($n = 8$, forested: 0.5, developed: 0.28), but the
opposite pattern for alien species ($n = 4$, forested: 0.73, developed: 0.86) (Fig. 5).
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Discussion

Our study sheds light on how ant community activity dynamics vary across a gradient of human-
induced land-cover change in a subtropical island environment. Our results complement an
extensive number of recent studies focusing on the temperate zone that examine change of insect
populations over time (Hallmann et al., 2017; Kaspari et al., 2022a; Seibold et al., 2019; Uhler et
al., 2021), including those with enough intra-annual sampling to examine changes to seasonality
(Kaspari et al., 2022b). The notable difference is that because climatic seasonality is much
weaker in the subtropics, differences in seasonality of insect populations across environmental
gradients are likely to be more difficult to detect than for populations in the temperate zone that
experience stronger seasonal contrasts (Kishimoto-Yamada & Itioka, 2015).
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We found that temporal variability diminishes for the subtropical ant communities of Okinawa
Island as land cover transitions from forested to more developed, and that this is related to a
breakdown in seasonality. Forested sites were more variable overall, but their resident ant
communities were more seasonal and synchronous, while community variability for developed
sites (i.e., urban and agricultural) was characterized by stochastic variation. Although native ant
richness was higher in forested sites, we determined the observed differences in seasonality
across the land-cover gradient were not due to differences in regional or local temperature
patterns, nor to species composition among sites. Compositional differences between
communities in different land-cover types, such as the prevalence of one or more highly seasonal
species, could drive observed relationships between community seasonality and land cover.
However, we found that populations of the same species occupying forested sites were more
seasonal than those with more human development. Further, when we separated species by
invasive status, we found that the strong seasonal patterns we observed in the forest were more
synchronous for native than alien species, though both synchrony values are reasonably high and
the considerably smaller sample size for alien species should be taken into consideration.
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Our results suggest that increasing agricultural and urban development can dampen seasonal
community variability and thus disrupt phenological patterns for insects. In a number of studies,
loss of seasonality has been explained by biotic homogenization, whereby communities become
more similar and generalist, and this has been linked with urbanization for a variety of taxa,
including bees (Hung et al., 2017), bugs and leafhoppers (Knop, 2016), flowering plants and
butterflies (Uchida et al., 2018), and birds (La Sorte et al., 2014; Leveau, 2018). However, the
majority of studies on the temporal variability of natural communities compare measurements
between years and rarely within them (e.g., Cottingham et al., 2001; de Mazancourt et al., 2013;
Olivier et al., 2020; Tilman et al., 2006), meaning that loss of seasonality due to environmental
or anthropogenic factors may be overlooked. Yearly samples taken for highly seasonal
communities at peak productivity over many annual cycles should show higher stability than for
a more stochastic community, yet more frequent sampling would reveal considerable variation
due to higher synchrony and seasonality. As other studies have demonstrated, temporal scale is a
crucial factor that determines how stability should be interpreted (Clark et al., 2021; McArdle et
al., 1990), and as community variability and its drivers vary across temporal scales (Hatosy et al.,
2013), it is important to measure community responses to the environment and anthropogenic
disturbance across scales to improve future predictions.
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We found a positive relationship between functional and compositional temporal variability,
meaning that those sites with communities that varied more as a whole tended also to have
higher total variation of the individual species in the local pool. The relationship between
functional and compositional variability can range from positive to negative depending on the
system and exposure to disturbance (Hillebrand et al., 2018; Ross et al., 2022; White et al.,
2020). Two characteristics of more forested sites may help explain this positive relationship. The
first is that more forested sites had higher variability in general at the biweekly timescale. The
second is that, at more forested sites, native species had higher richness and the highest
contributions to beta diversity. Sites with high richness that include substantial numbers of alien
species can have community dynamics that differ considerably from similar sites in which native
species dominate (Krushelnycky & Gillespie, 2008; Sanders et al., 2003), thus including alien
species in richness estimates can affect how diversity-stability relationships are interpreted (e.g.,
Moore & Olden, 2017).
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Our results raise the question of what mechanisms drive these patterns. We outline several
hypotheses for why seasonal variation in activity could vary across the land-cover gradient. First,
there is a possibility that ants are simply responding to the temperature they experience on a
physiological level, reflecting temperature and microclimatic variation across habitats. Ants are
well-known to be sensitive to temperature variation, with foraging limited to certain temperature
ranges (Bernstein 1979, Stuble et al., 2013). Regional climatic variation across the island is very
modest, and with no consistent differences in seasonality. Further, we found no relationships
between the seasonal variance of *in situ* air and soil temperature and the main land-cover
gradient PC1. However, differences in canopy openness, vegetation, and urbanization (via heat-
island effects, etc.) may create conditions that are relevant for promoting and depressing ant
foraging, and the prevalence of such conditions might be higher in more degraded habitats.
Comprehensive measurements with small sensors at various habitat strata for representative sites
could help elucidate whether this phenomenon could differ across land-cover types. Another
possibility is that ants are responding to resource availability patterns. In this scenario, it is
possible that resources are more seasonal inside the forest rather than more human-impacted
areas. This could occur, for example, because of anthropogenic food sources that are more static
throughout the year, or because of differences in phenology of vegetation inside or outside the
forest (Penick et al., 2015). A related question is whether these differences in seasonality are
limited to ants or are observed across the arthropod community. Were arthropod seasonality
responding more generally to land cover, this would provide a resource pattern that could affect
ant activity. Related to this, it is unclear why there seems to be little seasonal niche
differentiation for ant species in forested areas, which have activity patterns that follow
temperature quite closely in synchrony. One possibility is that species become most active at
different times during the same warm season to avoid competition—we saw some evidence of
this in the standardized seasonal curves (Fig. 5). This could be due to the inability to achieve
high activity levels during cooler months due to physiological limitations, which may be absent
in more developed environments with anthropogenic temperature refugia. Other potential
explanations relating to differences in the ecology or physical environments across the two
habitats likely exist, but more targeted research is needed to address this question.
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We measured variability over two years, but there may be other types of variability that only
manifest on longer timescales and are also correlated with land cover. For example, introduced

ants are known to have boom-bust dynamics when they reach new localities (Lester & Gruber,
2016). Over longer timescales, communities in more human-dominated areas may prove to be
more dynamic with successive shifts in the community. Indeed, we witnessed such an event in
our two-year period at one of our sites (Sefautaki Forest), where *Pheidole megacephala*, a
notoriously impactful invasive ant, arrived and quickly reached high abundance, raising
variability of this site beyond what was typical of others with similar land-cover characteristics.
Recent research in Yambaru Forest has documented resilience of native ant communities to
disturbance based on surveys of invasions at developed areas after regrowth (Shimoji et al.,
2022), so this site too may eventually recover to its previous structure. Thus, successive regime
shifts in these more disturbed communities could occur over longer timescales, but only
monitoring over more years would reveal such dynamics.
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Conclusions

This study provides novel insight into the seasonal dynamics of subtropical insect communities
across differing levels of human development. Importantly, it is the first study to our knowledge
to measure seasonality of ant activity with high-frequency sampling across a human disturbance
gradient. The results of our study, which link anthropogenic disturbance in the form of
agricultural and urban development to a reduction in intra-annual temporal variability due to
weakening seasonality, support the growing evidence that human development reduces natural
seasonal patterns (Hung et al., 2017; La Sorte et al., 2014; Leveau, 2018; Uchida et al., 2018).
Although this study focuses on correlative relationships and thus cannot fundamentally
demonstrate causal mechanisms, it does provide compelling evidence that continuing habitat loss
and fragmentation can lead to increasing homogenization of insect communities in parts of the
globe with the highest insect diversity, the subtropics and tropics. As loss of seasonal patterns
has been linked to a subsequent reduction in key ecosystem functions and services (Ross et al.,
2021; Stevenson et al., 2015), increasing development could result in disruptions to nutrient
cycling and food security (Bommarco et al., 2013). To better understand the generality of these
patterns, future studies using high-frequency sampling should focus on monitoring a greater
diversity of insect communities across different habitat strata (i.e., arboreal, subterranean; Gotelli
et al., 2011) in a variety of biomes. Greater focus on the impacts of land-cover change on insect
community seasonality should aid ongoing management and conservation efforts to help
preserve the important ecosystem functions and services insects provide.
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Data Availability

Raw species' count data before processing and R code used for analysis will be found on Data
Dryad after article acceptance. Please contact <jamie.m.kass (at) gmail.com> with data requests.

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Figure 1. Major land-cover classes of Okinawa Island with locations of the OKEON
monitoring stations, which cover the full gradient of human development across the
island. Ants were sampled from three stations at each sampling site biweekly from 2016–
2018 (total of 52 sampling periods).

Figure 2. Heat map showing the standardized ant activity (i.e., logs of summed site counts
divided by the log of the grand total count) over two years in Okinawa. Mean temperature
measurements from the Japan Meteorological Agency across the island, ordered from south to
north, are included for reference. Heat map sites are ordered by values of land-cover PC1,
explaining the anthropogenic stress gradient (top: more forested, bottom: more urban). Dotted
lines show the last sampling period before the new year.

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668 Figure 3. Trends in total log activity, functional variability, and compositional variability for ants
669 across a gradient of anthropogenic impact. Relationships with PC1, explaining the anthropogenic
670 stress gradient (low: more developed, high: more forested). Total log ant activity (before
671 rarefaction) correlates negatively with PC1 while indices of temporal variability (functional:
672 coefficient of variation, compositional: beta diversity) correlate positively.

Figure 4. a) Time-series decomposition of rarefied ant activity for the sites with the most forest (Yona Forest) and most human development (Oyama Park) within a 1 km buffer. b) Relationships between absolute and relative variance of seasonality and stochastic temporal components and PC1 across all 24 sites, explaining the anthropogenic stress gradient (low: more developed, high: more forested). Relative variance here is calculated as individual component variance divided by summed variance of all components.

Figure 5. a) Relative seasonality variance calculated per ant species in two representative land-cover groups each containing eight sites: more forested and more developed. Only species shared between groups (abbreviations of those found in Table S2) and with total counts ≥100 were retained, whereupon species counts were rarefied to the minimum between groups. On average, species have a higher relative seasonality variance in the forested group (paired Wilcoxon Test; p = 0.003), and no apparent patterns exist regarding alien status. b) Seasonal component time-series for each species (lines) found in the more forested and more developed groups, standardized by total species activity and separated by invasive species status. The dashed black line shows the mean standardized seasonality across all species per land-cover group, showing much greater temporal mismatch in ant communities at sites with greater human development.

<u>Supporting Information</u>

A. Ant activity data details

As sites across the island were sampled in sequential groups rather than simultaneously for each
biweekly period (differences are on the order of 7 days), to allow for appropriate site
comparisons, we assigned each sample a time step (52 in total) based on the relative sampling
order for that site. To this activity dataset, we filled in absence data when species that were
observed at least once at a site were not observed for other time steps for that site in order to
calculate temporal variability metrics. Values of NA ($n = 8$) were assigned to samples that
experienced trap malfunctions or data loss via extreme weather events and were removed from
analyses.
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B. Rarefaction permutations

Community samples collected over time are typically biased by sampling error, which results in
fluctuations in activity due to site placement or other external factors, but this bias can be
reduced via data rarefaction (Gaston & McArdle, 1994). As natural differences in total
community count across sites were high (from $n = 3,616$ [Yona Forest] to $n = 82,781$ [Sueyoshi
Forest]), we rarefied site- and group-level count while retaining original activity patterns to
reduce the effects of sampling error, allowing us to make more even comparisons of temporal
variability. For site-level activity, we randomly resampled the individual counts of species i at
time t for each site to equal the minimum total site count ($n = 3,616$). For group-level activity,
we first limited the species considered to those with total count greater than 100 for the two-year
study period, then randomly resampled (with replacement) the individual counts of species i at
time t for each group to equal that species' minimum total count between groups. We performed
1000 iterations of each rarefaction and conducted all proceeding analyses on these iterations.
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C. Community and compositional variability calculations

We measured community temporal variability by calculating the coefficient of variation (CV) of
summed ant count by site over the 2-year sampling period. We measured compositional temporal
variability by calculating temporal beta diversity for each site community as the total variability
of the species composition matrix (here, species × time step) after a Hellinger transformation (to
ensure purely relative count data; Legendre & De Cáceres, 2013) using the function *beta.div*
from the R package *adespatial* (Dray et al., 2022). As an additional product of this function, we
derived species contributions to beta diversity per site, which is calculated as species variance
divided by the total community variance (summing to 1). We then summed these contributions
for each invasive status category (native, uncertain, alien) to determine the contribution of each
category. We calculated both indices of temporal variability on each rarefied site activity dataset,
then found the mean values across datasets. These mean rarefied site values were used in
subsequent models.
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D. Formulas for temporal decomposition and associated variance

We used the TSLM() function in the R package *fable* (O'Hara-Wild et al., 2021) to fit time-
series linear models with the following formula:

$$y_t = \beta_0 + \beta_1 t + \beta_2 x_1$$

where y_t is the total count of ants at a site, with t equal to the range 1 to T, the number of time

intervals; and where x_1 is the 1st order Fourier term $\sin(\frac{2\pi t}{m})$, with m equal to the number of

seasonal periods (in this case, $m = 2$ for 2 years).

We calculated the temporal components in the following ways:

$$trend = \beta_0 + \beta_1 t$$

$$season = y_t - trend - resid$$

$$remainder = resid$$

where *resid* is the model residuals.

In addition to calculating absolute variance of temporal components, we calculated relative
variance with the following formula:

$$relative\ var_z = \frac{var(z)}{var(trend) + var(season) + var(remainder)}$$

where z is one of the temporal components: trend, season, or remainder.

E. Assessing effects of spatial autocorrelation on models

We first built models using ordinary least squares (OLS) regression to estimate the relationships
between land cover and the variables of interest. As some OKEON sites are relatively close to
others, we assessed the spatial autocorrelation present in the OLS model residuals by calculating
Moran's I and testing with random permutations using the function *moran.randtest*() from the R
package *adespatial* (999 repetitions; Dray et al., 2022). But as this test was not significant for
any of the models ($p > 0.05$), we fitted OLS models instead of more complex generalized least
squares (GLS) models with spatial structure. We determined goodness-of-fit with the adjusted
coefficient of determination (R-squared) and compared these between different count threshold
choices.

F. Effects of count threshold choice

As we avoided bias in temporal variability estimates by limiting all count values to an assigned
threshold, we examined how robust our results are over a range of different thresholds. We
report results in the main text for the relatively conservative threshold of 500 set at the station-
level, but additionally ran the same analyses with no threshold, and with thresholds 100, 200,
1000, and 2000, applied at both the station- and site-levels. We observed some differences across

the different thresholding schema, but no threshold fundamentally changes our results (which we
report for threshold 500 set at the station-level). The adjusted R-squared values for the metrics
calculated were sensitive in varying degrees to threshold (Fig. S7), with the exception of
compositional temporal variability—this is because our beta diversity calculation includes a
Hellinger transformation that mollifies the effects of outliers in the data. Further, thresholding at
the station- and site-levels showed the same general trends. The highest threshold considered
(2000) for the station-level was most often associated with the lowest R-squared values. The
absence of a threshold had the most dramatic differences. This reduced the R-squared value for
functional temporal variability to 0.11 from a thresholded minimum of 0.40, and seasonality
variance to 0.16 from 0.39. R-squared values for other metrics were similarly reduced, but more
closely followed decreasing trends as thresholds became larger. These reductions in correlations
with PC1 occurred because, without thresholding, sites with extreme count values for one or
several species spike in functional variability (calculated with CV) and lose seasonality, and the
existence of such spikes has little to do with land cover.
789

790   Table S1. Pearson correlations between proportions of land-cover classes within 1 km buffers of
791   sampling sites. "MISC" stands for miscellaneous and includes the classes for unclassified pixels
792   and those classified as bare rock or soil. As pairwise correlations were high, we conducted a
793   principal component analysis (PCA) to derive the main orthogonal land-cover gradients.
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	AGRICULTURE	FOREST	WATER	GRASS	SAND	URBAN	MISC
AGRICULTURE	1						
FOREST	-0.89	1					
WATER	-0.05	0.02	1				
GRASS	0.53	-0.44	0.02	1			
SAND	0.54	-0.48	-0.04	0.23	1		
URBAN	0.56	-0.84	-0.08	0.01	0.20	1	
MISC	0.51	-0.73	0.13	0.18	0.53	0.74	1

797 Table S2. Invasive status for all observed ant species based on existing literature and expert

798 opinion.

species	status
Aenictus ceylonicus	native
Aenictus lifuiae	native
Anoplolepis gracilipes	uncertain
Aphaenogaster concolor	native
Aphaenogaster irrigua	native
Brachyponera chinensis	native
Brachyponera luteipes	native
Camponotus bishamon	native
Camponotus devestivus	native
Camponotus monju	native
Camponotus OK01	native
Camponotus yambaru	native
Cardiocondyla kagutsuchi	uncertain
Cardiocondyla minutior	uncertain
Cardiocondyla obscurior	uncertain
Cardiocondyla wroughtonii	native
Carebara hannya	native
Carebara oni	native
Carebara yamatonis	native
Colobopsis shohki	native
Crematogaster cf. matsumurai	native

Crematogaster nawai	native
Crematogaster vagula	native
Cryptopone tengu	native
Diacamma OK01	native
Discothyrea kamiteta	native
Ectomomyrmex OK01	native
Erromyrma latinodis	native
Euponera pilosior	native
Hypoponera nippona	native
Hypoponera OK01	native
Hypoponera punctatissima	alien
Hypoponera sauteri	native
Leptogenys confucii	native
Lioponera daikoku	native
Monomorium chinense	native
Monomorium floricola	uncertain
Monomorium hiten	native
Monomorium intrudens	native
Monomorium pharaonis	alien
Myrmecina ryukyuensis	native
Nylanderia amia	uncertain
Nylanderia OK02	uncertain
Nylanderia OK03	uncertain
Nylanderia ryukyuensis	native
Nylanderia yambaru	native
Ochetellus glaber	native

Odontomachus kuroiwae	native
Ooceraea biroi	uncertain
Paratrechina longicornis	alien
Pheidole fervens	uncertain
Pheidole megacephala	alien
Pheidole noda	native
Pheidole parva	uncertain
Pheidole pieli	native
Plagiolepis alluaudi	alien
Polyrhachis dives	native
Polyrhachis moesta	native
Ponera takaminei	native
Ponera tamon	native
Pristomyrmex punctatus	native
Proceratium japonicum	native
Protanilla lini	native
Rhopalomastix OK01	native
Solenopsis tipuna	native
Stigmatomma sakaii	native
Stigmatomma silvestrii	native
Strumigenys circothrix	native
Strumigenys emmae	uncertain
Strumigenys exilirhina	native
Strumigenys hexamera	native
Strumigenys hirashimai	native
Strumigenys lewisi	native

Strumigenys mazu	native
Strumigenys membranifera	uncertain
Strumigenys minutula	native
Strumigenys OK01	uncertain
Strumigenys strigatella	native
Tapinoma melanocephalum	uncertain
Tapinoma saohime	native
Technomyrmex brunneus	alien
Temnothorax indra	native
Temnothorax OK01	native
Tetramorium bicarinatum	alien
Tetramorium kraepelini	uncertain
Tetramorium lanuginosum	alien
Tetramorium nipponense	native
Tetramorium simillimum	alien
Tetramorium smithi	alien
Trichomyrmex destructor	uncertain
Vollenhovia OK01	native

801   Table S3. Results from linear regression models built with two land-cover PCA axes: PC1,
802   explaining the forested-developed gradient, and PC2, explaining the rural-urban gradient. All
803   model residuals were tested for spatial autocorrelation by calculating Moran's I and testing with
804   random permutations (*moran.randtest*() from the R package *adespatial*; 999 repetitions), yet
805   none had significant results.

Response variable	Intercept	PC1 Estimate	PC1 Std. Error	PC1 *p*	PC2 Estimate	PC2 Std. Error	PC2 *p*	Adj. R^2	Moran *p*
Total activity (natural log)	10.082	-1.575	0.206	0	NA	NA	NA	0.714	0.269
Total richness (rarefied)	22.742	3.453	1.557	0.037	NA	NA	NA	0.145	0.727
Native richness (rarefied)	13.662	8.351	1.367	0	NA	NA	NA	0.612	0.789
Uncertain richness (rarefied)	5.863	-3.585	1.019	0.002	NA	NA	NA	0.331	0.179
Alien richness (rarefied)	3.217	-1.314	0.509	0.017	NA	NA	NA	0.198	0.59
Total richness (observed)	30.708	NA	NA	NA	NA	NA	NA	0	0.887
Native richness (observed)	18.708	7.483	1.686	0	-7.594	4.611	0.114	0.47	0.8
Uncertain richness (observed)	7.875	-5.188	1.516	0.002	NA	NA	NA	0.318	0.312
Alien richness (observed)	4.125	-2.052	0.643	0.004	NA	NA	NA	0.285	0.271
Total richness (estimated)	38.923	NA	NA	NA	NA	NA	NA	0	0.862
Native richness (estimated)	23.781	13.457	3.646	0.001	NA	NA	NA	0.354	0.767
Uncertain richness (estimated)	8.427	-5.717	1.636	0.002	NA	NA	NA	0.328	0.487
Alien richness (estimated)	4.292	-2.092	0.802	0.016	NA	NA	NA	0.202	0.3

Functional temporal variability	0.67	0.405	0.059	0	NA	NA	NA	0.667	0.269
Compositional temporal variability	0.23	0.141	0.024	0	NA	NA	NA	0.601	0.703
Native SCBD	0.53	0.237	0.059	0.001	NA	NA	NA	0.397	0.776
Uncertain SCBD	0.229	-0.16	0.05	0.004	NA	NA	NA	0.283	0.524
Alien SCBD	0.257	-0.087	0.05	0.094	NA	NA	NA	0.082	0.465
Seasonal variability (absolute)	1209.034	1850.927	328.626	0	NA	NA	NA	0.572	0.918
Seasonal variability (relative)	0.478	0.201	0.081	0.022	-0.373	0.222	0.108	0.232	0.39
Trend variability (absolute)	194.354	311.24	90.185	0.002	NA	NA	NA	0.322	0.567
Trend variability (relative)	0.079	NA	NA	NA	NA	NA	NA	0	0.722
Stochastic variability (absolute)	946.566	660.74	297.568	0.037	NA	NA	NA	0.146	0.591
Stochastic variability (relative)	0.443	-0.223	0.066	0.003	0.345	0.18	0.069	0.363	0.403

Figure S1. Conceptual workflow schematic describing the analysis steps. A) The main analysis
uses ant activity data from all sites. Richness is calculated for observed data and estimated via
Hill numbers, then the dataset is rarefied by randomly resampling individual workers per site to
match the minimum total site activity; rarefied richness is calculated from this rarefied dataset.
Next, the following analyses are conducted: functional temporal variability, compositional
temporal variability (including species contribution to beta diversity [SCBD]), and temporal
decompositions to derive seasonal, trend, and remainder (stochastic) components. B) The group-
level analysis first splits sites into two groups (green and pink points) based on land-cover PC1,
explaining the forested-developed gradient. Both groups share the same species, and those with
mid-level PC1 values are excluded (black points). These grouped data are then rarefied by
randomly resampling individual workers per species within each group to match the minimum
total species activity between groups. The same temporal decomposition is then conducted on
these grouped data.
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826　Figure S2. Biplot of the principal component analysis for the land-cover proportion dataset. PC1
827　explains the main anthropogenic stress gradient (low: more forested, high: more developed),
828　while PC2 explains the rural (high; agriculture and/or grass) to urban (low) gradient.
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831 Figure S3. Plot of compositional variability by functional variability, showing the sampling sites
832 with colors corresponding to the forested-developed gradient (PC1), where higher values (blue)
833 are more forested and lower values (red) have more human development. Site abbreviations
834 correspond to those used in Figure 1. The gray line shows the relationship between two
835 variabilities modeled with linear regression.
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Figure S4. Relationships between PC1 and summed species contributions to beta diversity (SCBD) for native, uncertain, and alien species for the rarefied activity data. PC1 explains the anthropogenic stress gradient (low: more developed, high: more forested).

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Figure S5. Relationships between PC1 and total richness, as well as richness of native, uncertain, and alien species for the rarefied activity data. PC1 explains the anthropogenic stress gradient (low: more developed, high: more forested).

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Figure S6. Absolute seasonality variance of temperature at the a) regional and b) site-level scales
over the two-year study period (2016 – 2018). Regional temperature measurements were taken
from Japanese Meteorological Association weather stations, and station names correspond to
those on the map. Site-level temperature measurements were taken for air and soil at one station
per sample site. Oyama Park (OYA) and OIST Open (OIT) were the only sites with sensors not
located below forest canopy, and thus have higher variance for soil temperature seasonality.
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Figure S7. The adjusted R-squared value for the metrics measured in this study at different count
thresholds and spatial levels for thresholding (station, site). The threshold value "none" is used as
a reference and is symbolized by the red point and orange line. For reference, the analysis
described in this paper is a threshold value of 500 at the station level.

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