#### 1 Exploring the genetic diversity of the Japanese Population: Insights

### 2 from a Large-Scale Whole Genome Sequencing Analysis

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# 61 Abstract

62 The Japanese archipelago is a terminal location for human migration, and the contemporary 63 Japanese people represent a unique population whose genomic diversity has been shaped by 64 multiple migrations from Eurasia. Through high-coverage whole-genome sequencing 65 (WGS) analysis of 9,850 samples from the National Center Biobank Network, we analyzed 66 the genomic characteristics that define the genetic makeup of the modern Japanese 67 population from a population genetics perspective. The dataset comprised populations from 68 the Ryukyu Islands and other parts of the Japanese archipelago (Hondo). Low frequency 69 detrimental or pathogenic variants were found in these populations. The Hondo population 70 underwent two episodes of population decline during the Jomon period, corresponding to 71 the Late Neolithic, and the Edo period, corresponding to the Early Modern era, while the 72 Ryukyu population experienced a population decline during the shell midden period of the 73 Late Neolithic in this region. Genes related to alcohol and lipid metabolism were affected 74 by positive natural selection. Two genes related to alcohol metabolism were found to be 75 12,500 years out of phase with the time when they began to be affected by positive natural 76 selection; this finding indicates that the genomic diversity of Japanese people has been 77 shaped by events closely related to agriculture and food production.

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# **83** Author summary

84 The human population in the Japanese archipelago exhibits significant genetic diversity, 85 with the Ryukyu Islands and other parts of the archipelago (Hondo) having undergone 86 distinct evolutionary paths that have contributed to the genetic divergence of the 87 populations in each region. In this study, whole genome sequencing of healthy individuals 88 from national research hospital biobanks was utilized to investigate the genetic diversity of 89 the Japanese population. Haplotypes were inferred from the genomic data, and a thorough 90 population genetic analysis was conducted. The results indicated not only genetic 91 differentiation between Hondo and the Ryukyu Islands, but also marked differences in past 92 population size. In addition, gene genealogies were inferred from the haplotypes, and the 93 patterns were scrutinized for evidence of natural selection. This analysis revealed unique 94 traces of natural selection in East Asian populations, many of which were believed to be 95 linked to dietary changes brought about by agriculture and food production.

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## 99 Introduction

100 The Japanese archipelago is located in the eastern part of the Eurasian continent and is one 101 of the final destinations of the human migration out of Africa. While the identity of the first 102 human groups to reach the Japanese archipelago is uncertain, the Jomon people, who were 103 hunter-gatherers known for their pottery, lived in the region after 16,000 years ago. The 104 genetic diversity of the peoples of the Japanese archipelago underwent a dramatic 105 transformation following the Yayoi Period, which began around 3,000 years ago with the 106 migration of agriculturalists from Eurasia. Genome analysis of ancient and modern humans 107 has shown that they admixed with the originally inhabited Jomon people, resulting in the 108 genetic diversity of the modern Japanese population from the Yayoi people. This process is 109 thought to have started on Kyushu Island and then gradually spread throughout the 110 archipelago. Although the Ryukyu Islands are separated from Kyushu Island by a 111 significant distance, the agricultural culture known as the Gusuku period began 800 years 112 ago, and it is believed that this agriculture was introduced by migrants from the mainland. 113 These long histories of migration and admixture have shaped the genetic diversity of the 114 peoples of the Japanese archipelago. Previous genome analyses have attempted to reveal 115 this genetic diversity, but most have only sampled specific regions and therefore have been 116 insufficient to fully examine the genetic diversity of the Japanese archipelago as a whole. In 117 this study, we used whole-genome sequencing data from subjects from a wide range of 118 regions in the Japanese archipelago to more fully understand the genetic diversity of the 119 peoples of the archipelago.

120	There are six national research hospitals in Japan that specialize in advanced medical care
121	and research, and each of them maintains its own biobank that collects and stores biological
122	samples from patients. National Center Biobank Network (NCBN) is a federation of these
123	centers that collaborate to provide samples, genomic and clinical information, and public
124	relations. In this study, we performed WGS of 9,850 individual DNA specimens stored in
125	the biobanks of NCBN. These biobanks are located in three distinct regions of Japan:
126	NCGM, NCCHD, and NCNP in the Tokyo area; NCGG in Aichi Prefecture in the central
127	area in the Honshu Island; and NCVC in Osaka Prefecture in the western area in Honshu
128	Island. Therefore, the genomic information obtained in the present study is expected to
129	reflect the regional diversity of Japan to some extent. Here, we characterized the data
130	obtained from this analysis and described the genetic diversity in the Japanese population.

# 131 **Results**

### 132 Whole genome sequencing analysis

133 DNA samples from 9,850 individuals from five National Center Biobanks were analyzed

using WGS, and the data in FASTQ format were received from the outsourced laboratory.

135 The received data were processed through the primary data analysis pipeline to obtain

136 mapping results and variant call results. Quality control (QC) metrices were calculated to

137 confirm that stable quality analysis was obtained (Fig 1). The autosomes had an average

read depth of  $34.0 \pm 2.4$ , and the average insert length of the reads was  $703 \pm -30$  bp. The

139 mapping rate per sample was  $99.99\% \pm 0.39\%$ . These statistics did not vary significantly

- 140 between samples, and there was no clear bias between biobanks except for saliva samples.
- 141 The saliva samples showed lower mapping rates than the blood samples, probably due to
- 142 the foreign DNA in the saliva.

#### 143 Summary and accuracy of SNP and short insertion and

#### 144 deletions

145 Variants were characterized by joint calling to integrate individual variant information. In
146 this analysis, we performed joint calling of the gVCFs of 9,287 samples from NCBN,
147 analyzed at the time of writing, together with 2,504 samples from the International 1000

- 148 Genomes project (S1 Table). The VCF obtained after the joint call contained a total of
- 149 208,785,859 records, of which 88.5% (184,864,563) passed the filtering using Variant
- 150 Quality Score Recalibration (VQSR). We found 122,459,307 variants after focusing only
- 151 on the variants in the NCBN samples, and 86.3% (105,729,588) of them passed the filter of
- 152 VQSR. Of the variants that passed the filter, 87,246,166 were single nucleotide variants
- 153 (SNVs) and 18,483,422 were short insertion and deletions (INDELs); 47% (41,046,547) of
- the SNVs and 39.8% (7,361,318) of the INDELs that passed the filter were novel variants
- 155 not registered in dbSNP151. Most of the novel variants were very rare. For example,
- 156 34.56% of the known SNVs were singletons found as the heterozygous genotypes of one
- sample out of 9,287 individuals, and 86.73% were observed at a very low frequency of less
- than 0.5%. Conversely, 67.46% of the novel variants not registered in dbSNP were
- singletons, and the percentage of SNVs with a frequency less than 0.5% was more than

99.99%. This is consistent with previous reports that most novel variants are foundprivately [3–6].

162 We evaluated the accuracy of the variants using two approaches. First, we performed the 163 genotyping using SNP array to estimate the degree of genotype concordance with WGS 164 results. For this purpose, genome-wide genotyping using the SNP array on the DNA 165 samples of 448 individuals who had undergone WGS analysis was conducted. The 639,508 166 autosomal SNPs remaining after variant QC in the SNP array were compared with the 167 results obtained after WGS analysis. The number of mismatches ranged from 66 to 7,205 168 per sample, with an average of 408.7. As a result, the average discordance rate between the 169 two sets of variants was 0.063%. This estimate appears to be a conservative estimate of the 170 error, as it is primarily concentrated in a region that is easy to analyze and for which probes 171 are designed on SNP arrays. Then, we compared the genotypes of the trio samples to 172 estimate the frequency with which the offspring of a trio had heterozygous or non-reference 173 homozygous variants whose parents' genotypes did not follow the pattern expected from 174 Mendelian low. The sample analyzed in this study contained 148 trios of parents and 175 offspring. We observed the inheritance pattern of genotypes from an average of 4,284,264 176 variants per trio. Of these, 6,448.4 (0.15%) had an abnormal inheritance. This percentage 177 became more pronounced when stratified by the novelty of the variants, e.g., the known 178 SNVs and INDELs had error rates of 0.09% and 0.42%, respectively, with errors in the 179 inheritance pattern, whereas the novel SNVs and INDELs had error rates of 2.26% and 180 10.9%, respectively. The Mendelian heritability errors can include the sequencing or

181 genotyping error, *de novo* mutations, and gene conversions in the parents' gametes.

182 However, our estimates approximate the error rate of genotyping in this study.

#### **183** Ancestry inference and allele frequency distribution

184 We conducted the principal component analysis (PCA) to identify the ancestry of NCBN 185 samples. After removing 20 samples with a call rate below 95%, Identical-by-Descent 186 (IBD) was used to detect related samples, resulting in 8,972 and 2,493 unrelated samples 187 from NCBN and the International 1000 Genomes project, respectively. PCA using these 188 samples detected 21 NCBN samples not belonging to the East Asian populations (Fig 2A). 189 Furthermore, when PCA was performed only on East Asians, the samples were divided into 190 two clusters: one consisting of continental populations (Han Chinese in Beijing; CHB, Han 191 Chinese South; CHS, Kinh Vietnamese; KHV, Chinese Dai in Xishuangbanna, China; 192 CDX) and the other including Japanese in Tokyo (JPT) from 1000 Genomes and NCBN 193 samples (Fig 2B). In addition, the latter cluster was divided into large and small clusters 194 consistent with the previous studies [7-9] in which the larger one was called the Hondo 195 population and the smaller one was called the Ryukyu population [7]. In this study, we 196 followed this convention (S1 Fig). The Hondo cluster consisted of 8,524 people, whereas 197 the Ryukyu cluster consisted of 182 people. We compared the allele frequencies of the 198 Japanese population (GEM Japan Whole-genome Aggregation) estimated based on the 199 WGS analysis in previous studies with those of the Hondo sample and found significant 200 frequency agreement (Fig 3A). While the allele frequencies between the Hondo and 201 Ryukyu populations also showed high agreement, the breadth of the distribution was wider

than the comparison between Hondo and GEM Japan (Fig 3B). This could be due to thedifference in the mainland and Ryukyu populations and the subsequent genetic drift.

#### 204 Functional landscape of variants

205 The variants identified by WGS analysis were annotated for their biological functions. The 206 impact of the variants was classified based on the criteria of the annotation software and the 207 database as described in the Methods section. Deleterious mutations are more likely to be 208 kept at low frequencies in the population, as such mutations are less likely to spread in the 209 population because of negative selection. In fact, variants with a high impact on annotation 210 showed a clear tendency to have a low frequency in the population. The LOFTEE plugin of 211 Variant Effect Predictor was used to detect loss-of-function (LoF) variants in the Hondo 212 and Ryukyu populations. For comparison, we also detected LoF variants in 26 populations 213 in the 1000 Genomes Project phase 3 dataset [10]. 14,145 SNVs and 16,823 INDELs were 214 detected as high confident LoF specific to the Hondo population. For the Ryukyu 215 population, 211 SNVs and 288 INDELs were detected. The vast majority of LoF SNVs 216 exhibited a very low frequency in the Hondo population (Fig 4A). In fact, 76.0% of these 217 SNVs exhibited allele frequencies below 0.01%. We compared the number of LoF alleles 218 and the number of homozygous of LoF alleles per individual for Hondo, Ryukyu, and 219 populations in the 1000 Genomes Project (Fig 4 B and C, S2 Fig). Since homozygous LoFs 220 result in a complete loss of gene function, the number of homozygous LoFs in an 221 individual's genome can be used to measure the individual's genetic burden. Both indices 222 were highest in Africa, lowest in West Eurasia, and moderate in Hondo and Ryukyu. The

number of homozygous LoF alleles per individual by allele frequency was generally higher
in Africa across all allele frequencies (Fig 4D), which is consistent with the trend observed
in a previous study [11].

226 We compared the variants of NCBN samples with ClinVar registered variants [12]. A total 227 of 103,833 variants found in the Hondo population are registered in ClinVar. Of these, 228 2,427 were classified as "pathogenic" or "likely pathogenic" variants. Seven variants were found in the four-star category, the most reliable classification based on the ClinVar review 229 230 status. Only one of them was "pathogenic" and a singleton variant (i.e., heterozygous in a 231 person) of the CTFR gene. The remaining six were polymorphic variants related to drug 232 responsiveness of CYP2C19. There were 1,130 variants in the 3-star category reviewed by 233 the expert panel. Of these, 56 were "pathogenic," and 13 were "likely pathogenic." The 234 frequencies of these variants were the highest, at 1.0%, and most of them were extremely 235 rare; only a few were observed in the population. Most of the less well-reviewed variants 236 with <3 stars had frequencies of less than 1%, but 34 variants had a frequency of 1% or 237 more.

### 238 Allele frequency estimation of HLA loci

239 Three-field HLA calling results from the WGS dataset in the present study were compared

240 with HLA allele frequencies HLA Foundation Laboratory (Kyoto, Japan) (S3 Fig). All

common HLA alleles (frequencies >1%) were concordant between the two datasets with

observed differences of less than 1%. To further validate our HLA calling results, a subset

243 of 94 samples was subjected to high-resolution HLA genotyping. Three-field HLA class I

#### 244 (HLA-A, -C, and -B) accuracies were 96.3%, 97.9%, and 96.8%, respectively, and 3-field

HLA class II (HLA-DRB1, -DQA1, -DPA1, and -DPB1) accuracies were 98.9%, 100.0%,

246 98.9%, 100.0%, and 96.8%, respectively. The accuracy of 2-field HLA class I (HLA-A, -C,

247 and -B) increased to 97.9%, 98.4%, and 97.3%, respectively.

### 248 Evolutionary perspective of genomic diversity

249 The recent decrease in population size was detected in Hondo and Ryukyu populations.

250 Figure 5A shows the population histories of Hondo and Ryukyu populations inferred using

IBDNe [13], which estimated the changes in population size in recent (~200 generations

ago) past based on IBD sharing among individuals. In Hondo, the population size decreased

from about 75 to 50 generations ago, and from 17 to 11 generations ago. In the Ryukyu

population, a reduction in population size was observed from about 100 to 25 generations

ago. The distributions of IBD length were multimodal in both populations, indicating

256 fluctuations in population size (Fig 5B and 5C). We also estimated the long-term changes

in the effective population size from the genome-wide genealogy using Relate software

[14]. We estimated genome-wide genealogy based on the whole-genome data of 1,000

randomly selected samples from Hondo, 182 samples from the Ryukyu, and the CHB

260 population from the 1000 Genomes Project. The Ryukyu population showed a bottleneck

that peaked around 2,700 years ago (S4 Fig). Hondo/CHB population and Ryukyu

262 population diverged around 3,700 years ago, consistent with previous estimations of the

263 divergence time using SNP arrays [15,16].

264 We detected positive natural selection based on genome-wide genealogy of 1,000 Hondo 265 samples and found SNPs with p-values below the genome-wide significance level (p < 5.0266  $\times$  10<sup>-8</sup>) (S5 Fig, S2 Table). As the QQ plot suggested inflation of the test statistics (S6 Fig), 267 it is possible that the results contain false positives. However, the genes reported in 268 previous studies, which may have undergone positive natural selection, were correctly 269 included in the results. It is therefore important to consider this when interpreting the results. For example, ALDH2 rs671 G/A (p-value =  $2.0 \times 10^{-17}$ ) and ADH1B rs1229984 270 T/C (p-value =  $6.8 \times 10^{-10}$ ), which are associated with alcohol metabolism, showed positive 271 272 natural selection signals [17,18]. The genealogies of the genes showed that the derived 273 alleles were spreading rapidly through the population (S7 Fig). The second example is 274 signals of positive natural selection on the non-synonymous rs76930569 C/T (p-value = 1.1 $\times$  10<sup>-12</sup>) variant in the OCA2 gene. This variant is in complete linkage equilibrium with 275 276 rs1800414 T/C, involved in melanin biosynthesis, and has been shown to be associated 277 with light skin color and tanning ability in Asian populations [19–21]. The third example of 278 the positive selection is the FADS gene family. Multiple SNPs (rs174599, rs174600, 279 rs174601, rs97384, rs57535397, rs76996928) showed the signatures of positive selection. 280 FADS1 and FADS2 encode catalytic proteins, which synthesize long-chain fatty acids from 281 short-chain fatty acids [22], and have been subjected to natural selection related to diet in 282 several human populations [22–26]. We further analyzed change in allele frequency with 283 time for these genes under positive natural selection. We used CLUES software [27] to 284 estimate the allele frequency trajectory of SNPs in ALDH2, ADH1B, OCA2, and the 285 FADS gene family. The frequency of the derived alleles in ADH1B rs1229984 increased

286	about 20,000 years ago (Fig 6A). In contrast, the frequency of ALDH2 rs671 increased
287	from about 7,500 years ago (Fig 6B). The allele frequency trajectory of OCA2 rs1800414
288	(Fig 6C) showed that the frequency of derived allele of OCA2 rs1800414 began to increase
289	due to natural selection around 25,000 years ago. The frequency of derived allele of
290	rs174599 began increasing around 25,000 years ago, slowed down 15,000 years ago, and
291	started increasing again 10,000 years ago (Fig 6D).

# 292 **Discussion**

293 In the present study, we conducted a WGS analysis of samples from five biobanks in Japan. 294 Although the data obtained in this study are intended to be provided as control data for 295 genomic studies of various diseases, the analysis in this study focused on data quality and 296 population genetics properties. A uniform quality of data was obtained through the use of a 297 single procedure that encompassed both sequencing and data analysis. Population-based 298 studies using WGS analysis have been conducted in various populations [5,28,29]. Studies 299 on Japanese populations have already been reported [28], and the allele frequency 300 distributions in previous studies are consistent with the results of the present study (Fig. 301 3A). The samples analyzed in the present study were provided by biobanks in three regions 302 of Japan: NCGM, NCCHD, and NCNP in the Tokyo area; NCGG in Aichi Prefecture in the 303 central area in the Honshu Island; and NCVC in Osaka Prefecture in the western area in 304 Honshu Island. Therefore, the genomic information obtained in the present study is 305 expected to reflect the regional diversity of Japan to some extent. For instance, the

306 population genetic analysis identified two clusters representing the ancestry of Ryukyu 307 Islands, comprising Okinawa Prefecture and the islands of Kagoshima Prefecture and the 308 Hondo region (mainland). This supports the idea that the Hondo and Ryukyu populations 309 are genetically differentiated, as suggested by anthropological studies [7–9]. We further 310 found that past population sizes differed between Hondo and Ryukyu. There was a 311 reduction in the Hondo population from 17 to 11 generations ago (Fig 5A). The 312 corresponding period was 476 and 308 years ago, and the assumption is that each 313 generation spanned 28 years; most of this duration overlaps with the Edo period in Japan. 314 This is consistent with findings from historical demography studies, which suggest that the 315 population not only increased but also remained stagnant due to limited economic growth, 316 population concentration in cities, and famine caused by cold weather-related damage 317 during this period. In contrast, the Ryukyu populations showed population reduction from 318 100 generations ago to 25 generations ago but then increased until the present (Fig 5A). 319 This population growth about 700 years ago was close to the beginning of farming in the 320 Ryukyu Islands (12th century). Assuming that agriculture was brought to the Ryukyu 321 Islands by migrants from the mainland of Japan, the population decline observed in the 322 Ryukyu population can be considered a bottleneck associated with the migration. The 323 population size estimated from the modern genome reflects the past population of the 324 migrants and should be influenced negligibly by the genetic diversity of the original 325 inhabitants of the Ryukyu Islands. Indeed, although the several human skeletal remains 326 have been discovered from Pleistocene sites in Ryukyu Islands [30,31], the previous 327 population genetic analysis based on genome-wide SNPs suggested minor genetic

328 contribution of the Pleistocene Ryukyu Island population to the modern Ryukyu population
329 [15,16]. The estimated time of divergence between Hondo/CHB and Ryukyu was 3,700

330 years ago (S4 Fig), suggesting that migration to the Ryukyu Islands occurred recently.

331 Studies of rare genetic diseases require data on the frequency of variants in the population.

332 Most of the variants we found in this study were rare, and many of them were newly

333 discovered in this study, as expected from population genetics theory. However, the lower

the frequency of the variants, the more difficult it becomes to distinguish them from errors.

335 In this study, we evaluated the accuracy of genotype detection, estimating a discordance

rate of 0.063% compared to genotype detection using WGS and SNP arrays. However, this

is an overestimation of the error rate due to the combined error of both technologies. We

also used the data obtained from the WGS analysis of the trio for validation. We estimated

the Mendelian error rate, which is the proportion of genotypes detected in the offspring of a

trio that is inconsistent with Mendel's laws of heredity. This method has the advantage of

being able to examine the entire genome compared to the use of SNP arrays. We found that

342 the Mendelian error rate is much higher for novel variants, i.e., previously reported

343 variants. The Mendelian error rate for novel SNVs was 2.26%, much higher than that of the

known SNVs (0.09%). This has important implications for the identification of causative

345 mutations in rare genetic diseases, as many causative mutations for these conditions are

newly discovered rare variants. This means that the discovery of such pathological variants

in patient sequencing is subject to a non-negligible degree of error.

348	We conducted the functional annotation of the variants discovered in this study. Consistent
349	with previous studies [5,28,29], variants that were expected to have a high biological
350	impact were less common in the population, confirming that negative natural selection
351	shapes the diversity of variants. Most of the LoF mutations were extremely rare, and most
352	of them were heterozygous (Fig 4A). The number of LoF mutations in the genome was
353	comparable to that in other Eurasian populations (Fig 4B). Although the Ryukyu population
354	has experienced population decline (Fig 5A), the frequency of LoF variants was
355	comparable to that in the Hondo population, and no evidence of differences in the profile of
356	rare functional variants due to the bottleneck effect was noted. The number of LoF sites and
357	homozygous LoF per individual in this study were higher than those detected in a previous
358	study [32]. Among these, the number of stop gained SNVs was consistent with that
359	recorded in the previous study [32], whereas the number of splice site SNVs and frameshift
360	INDELs was higher than that in the previous study [32] (S2 Fig). The number of LoF sites
361	was generally consistent with the number of LoF sites before manual curation in a previous
362	study [33]; thus, it may be possible to remove false-positive homozygous LoFs through
363	manual filtering, as in the previous study [33].
364	We also examined pathogenic variants that have been reported in the past. Pathogenic

364 We also examined pathogenic variants that have been reported in the past. Pathogenic

365 variants assessed by an expert panel (4-star status) on ClinVar were found only in one to a

366 few individuals in the population. On the other hand, some variants that were less reviewed

- 367 were polymorphic with high frequency. These results reinforce the importance of utilizing
- 368 the frequency of the variants in the population to evaluate their pathogenicity.

369 Genes that have undergone positive natural selection in the East Asian populations are 370 related to the metabolism. This study supported that the dietary changes in the ancestors 371 seem to have shaped gene frequencies. Candidate regions undergoing positive natural 372 selection were found on a genome-wide scale using genealogy analysis (S5 Fig). ADH1B is 373 involved in metabolizing alcohol to acetaldehyde, and ALDH2 is involved in metabolizing 374 acetaldehyde. Both the non-synonymous A allele of ALDH2 rs671 and the C allele of 375 ADH1B rs1229984 affect the retention of acetaldehyde in the body and cause alcohol flush 376 in Asians [17,18]. These alleles have been suggested to be associated with Japanese dietary habits and diseases, such as esophageal cancer [34,35]. Previous studies have hypothesized 377 378 that positive selection may have acted to maintain acetaldehyde in the blood against 379 parasite infection, which correlates with large-scale rice cultivation [36–39]. We also 380 observed that the increase in the frequency of ADH1B occurred earlier than that of 381 ALDH2, indicating that positive selection began to act at different times for these two 382 genes (Fig 6A and 6B). Based on the geographic distribution of haplotype structures around 383 ADH1B and ALDH2, according to Koganebuchi et al., positive selection on ADH1B 384 rs1229984 started before the beginning of the Jomon period, while positive natural 385 selection on ALDH2 began around 8,000 years ago, in association with the beginning of 386 rice cultivation in China [39]. Our dating by genome-wide genealogy of the Japanese 387 population genome is consistent with the above consideration. Using HapMap data, OCA2 388 rs1800414 has been shown in previous studies to be the effect of positive natural selection 389 on East Asians [19]. For the OCA2 gene, positive natural selection signals were found in 390 the European population for skin color-related SNPs other than those detected in this study

391 [19]. As natural selection works for light skin color, a previous study mentioned that it 392 enhances vitamin D synthesis capacity in regions with low sunlight [20]. For East Asians as 393 well, positive natural selection may have operated in relation to vitamin D synthesis in 394 regions with low sunlight. However, since the derived allele of rs1800414 is not necessarily 395 more frequent at the high latitudes of East Asia, other possibilities, such as sexual selection, 396 cannot be ruled out at this time [19]. The derived allele of rs1800414 has been shown to be 397 associated with light skin color and tanning ability in Chinese and Japanese populations [20,21] and is widely observed in modern East Asians [19], suggesting that the derived 398 399 allele of rs1800414 originated in the common ancestor of East Asians and spread 400 throughout East Asia at very early stages of the East Asian population history. We 401 estimated that the derived allele of OCA2 rs1800414 began to increase in frequency around 402 25,000 years ago (Fig 6C). Future analyses of older East Asian lineages, such as the ancient 403 genome of the Jomon people, may reveal the original variant of this allele that led to 404 positive natural selection. FADS1 and FADS2 participate in fatty acid metabolism. For 405 example, in the Inuit population, which relies heavily on a marine animal diet, there are 406 positive natural selection signals on SNPs of FADS2 genes, which are responsible for the 407 increase in the concentrations of short-chain fatty acids [40]. Signals of positive natural 408 selection on alleles that promote long-chain fatty acid synthesis have also been identified in 409 African [22], European [25,26,41], and South Asian populations [24]. In particular, studies in European populations have shown that the derived alleles of rs174594 and rs1714546 are 410 411 associated with increased total cholesterol and LDL cholesterol levels, increased expression 412 of FADS2, and decreased expression of FADS1 [25]. In European populations, increased

413 reliance on plant diets seemed to have resulted in positive natural selection on alleles that 414 promote long-chain fatty acid synthesis pathways of the FADS gene family [23,25,26]. The 415 SNPs in the FADS gene family detected in this study were associated with total cholesterol 416 and LDL cholesterol levels, increased expression of FADS2, and decreased expression of 417 FADS1 (S3 Table), like the SNPs subjected to natural selection in the European population 418 (S3 Table). These results suggest that in Hondo populations, as in Europeans, the dietary 419 change was accompanied by positive natural selection for alleles that promote the long-420 chain fatty acid synthesis. The frequency of the derived allele of rs174599 in FADS2 began 421 to increase around 25,000 years ago, but the increase was not continuous, and there was a 422 period of stagnation from 15,000 years ago for 5,000 years (Fig 6D). Interestingly, the 423 frequency of this allele varies widely among East Asian populations. The derived allele was 424 major in CHB (64%) and Japanese (63%), whereas it was minor in Dai (Chinese Dai in 425 Xishuangbanna, China) (22%), Han Chinese in South (42%), and Kinh Vietnamese (20%); 426 these data suggest that the positive natural selection of the FADS gene family in East 427 Asians may reflect the association with agriculture and the complex dietary differences 428 among regional populations. Notably, Mathieson and Mathieson (2018) disproved the 429 simple idea that these derived alleles underwent positive natural selection in relation to the 430 introduction of agriculture and speculated that there were complex underlying factors, such 431 as unknown dietary changes [26].

432 In this study, we demonstrated that the data presented here can be used as a foundation for

433 analysis of human genetics. While this study focused on population genetic characterization

434 of the Japanese population, the data can be used in disease studies, as a resource for

genotype imputation in studies of common diseases, and as a control in studies on rarediseases.

# 437 Materials and Methods

### 438 Sample preparation

439 DNA samples stored in the biobanks of five national centers (National Cerebral and 440 Cardiovascular Center; NCVC, National Center for Geriatrics and Gerontology; NCGG, 441 National Center for Global Health and Medicine; NCGM, National Center of Neurology 442 and Psychiatry; NCNP and National Center for Child Health and Development; NCCHD) 443 were submitted for WGS analysis. Samples derived from healthy individuals or patients 444 with some common diseases were selected as control groups for future disease studies. This 445 study was conducted with approval from the ethics review committee of the NCGM. 446 Informed consent for the analysis of these samples was received from all subjects in each 447 biobank. Approximately 50  $\mu$ l of DNA at a concentration of 80 ng/ $\mu$ l per sample was 448 aliquoted into 96-well plates and shipped to an outsourced laboratory (TakaraBio, Shiga, 449 Japan) for WGS analysis.

### 450 WGS

451 To avoid quality fluctuations and batch effects, all samples were analyzed by a single

452 outsourced laboratory. WGS analysis was performed using NovaSeq6000 (Illumina, San

453 Diego, CA, US), and sample preparation was performed using the procedures and reagents

454	recommended by the manufacturer. DNA molecules were sonicated with a protocol
455	targeting an average size of 550 bp. DNA libraries were prepared using the TruSeq DNA
456	PCR-Free HT Library Prep Kit, and index sequences were added for multiplex analysis.
457	The insert size was confirmed by electrophoresis in the range of 400–750 bp before
458	sequencing runs. WGS was performed at 150 bp paired-end and repeated in multiplex until
459	an output of $>90$ Gbases without duplicated reads was obtained.

#### 460 Data analysis

461 We received the quality controlled FASTQ data from the outsourced laboratory and

462 performed mapping and variant calling in an in-house data analysis pipeline. The mapping

463 and variant calls were performed using the Parabricks v3.1.0 (Nvidia, Santa Clara, CA,

464 US), which provides the capability to perform the analysis recommended by GATK at high

speed using a GPU [42]. The GRCh38 was used as the reference sequence. The pipeline

466 used in this study implements algorithms equivalent to those of bwa (v0.7.15) [43] and

467 GATK (v4.1.0). We flagged duplicates from mapped reads but did not perform realignment

and base quality score recalibration to reduce the computational time. The mapped data

469 were outputted in BAM format and converted into CRAM format using samtools [44] to

470 reduce the file size. Variant calls were output in gVCF format for joint calling. QC metrics

471 were obtained to evaluate the quality of the analyzed data. The depth and map rate were

472 calculated using GATK's CollectWgsMetrics tool. These QC metrics were continuously

473 monitored throughout the analysis. The sex chromosomes were analyzed assuming both

474 male and female genders. Variant calls were performed for chromosome X in the diploid (-

475 ploidy 2) model for females and the monoploid (-ploidy 1) model for males.
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- 476 of the pseudoautosomal region were performed in the diploid model. Variants of
- 477 chromosome Y were called in the monoploid mode regardless of the sample's gender.
- 478 Finally, the appropriate gVCF file for each sex was used during joint calling. Data from the
- 479 high-coverage WGS analysis of 2,504 individuals of the International 1000 Genomes
- 480 Project phase 3 [10] were used as population references for this study. The CRAM files
- 481 reanalyzed using high depth WGS were downloaded from a public database, and variant
- 482 calls were performed with the protocol described in this study.

#### 483 **Integrated analyses**

484 To properly estimate the frequencies of variants found after WGS in the population, we 485 integrated the gVCF files. The joint calling was conducted by combining samples from the 486 biobank of NCBN and samples from the International 1000 Genomes Project phase 3. For 487 the joint calling, we used the gVCFtyper program of the Sentieon package [45]. This 488 program produces results equivalent to those of GeomicsDBImport followed by 489 GenotypeGVCFs programs for the joint calling of GATK. To perform efficient 490 computation in a cluster computation environment, we divided the autosomes into 29 491 regions evenly. Each variant was scored using VQSR to filter the integrated VCF. The 492 VarCal and ApplyVarCal programs of the Sentieon package corresponding to GATK's 493 VariantRecalibrator and ApplyVQSR, respectively, were used for this process. The 494 HapMap and International 1000 Genomes Omni2.5 sites, the high-confidence SNPs of the 495 International 1000 Genomes Project, and the dbSNP151 sites were used as true, training,

and known datasets, respectively. Variants identified as PASS, which correspond to
filtering with 99.9% sensitivity, were used in subsequent analyses unless otherwise noted.
The INDELs in the present variant set were normalized by performing left align, and
multiallelic variants were split into multiple variant records using the norm subcommand in
beftools [44].

#### 501 Variant annotation

502 Variants were annotated with the Variant Effect Predictor v102 [46]. We ran the loftee

503 plugin to evaluate the effects of the LoF variants. For the other evaluation of the functional

504 effects, dbNSFP4.1 [47] was used to assign precomputed evaluation values to the variants.

505 The metrics used for the assignment included LRT, SIFT, MutationTaster, and Polyphen2.

#### 506 Genotyping by SNP array

507 Using JaponicaArray [48], genome-wide genotyping was performed on a subset of samples 508 for comparison with WGS results. Ninety-four samples each from five biobanks were 509 analyzed using the residual DNA after WGS analysis. The analysis was performed by an 510 outsourced laboratory (Toshiba, Tokyo, Japan), and the raw data in CEL format was 511 received. Four samples were dropped from the genotyping due to a low call rate (<97%) in 512 the first step of genotyping. Clustering for SNP genotyping of variants was performed on 513 the data of 466 individuals using the Analysis Power Tools (ver. 2.10.2.2, Thermo Fisher 514 Scientific, MA, USA). The clustering results for each probes' intensity were classified 515 using the SNPolisher program bundled with the Analysis Power Tools, and the 639,508

516	SNPs classified as "Recommended" in autosomes were used for subsequent analyses. The
517	genotype concordance with WGS was estimated using the hap.py software. To compare the
518	positions for which probes were designed in the SNP array, SNVs with the same position as
519	the SNP array were extracted from the results of WGS analysis. The SNP array results were

- 520 used as true data and the WGS results as query data. The genotype discordance rate
- 521 between the SNP array and WGS was calculated by dividing the number of false positives
- 522 by 639,508, which is the total number of SNPs compared.

### 523 Allele frequency estimation

524 To calculate the accurate allele frequencies, the ancestry of the samples was estimated

525 using PCA. Variants were filtered under more stringent criteria for this purpose.

526 Individual's genotypes were considered no calls if they had a genotype quality (GQ) of less

527 than 20, a depth outside the range of 11 to 64, or if less than 25% of the reads supported the

528 minor allele for heterozygous calls. Then, sites with SNPs that had a VQSR filter of PASS,

a minor allele frequency of >1%, and a call rate of >95% were retained. The KING

530 program [49] selected samples of unrelated individuals in the third-degree kinship or more.

531 For this dataset, independent SNPs were extracted using PLINK1.9 [50] with "-indep-

pairwise 500 50 0.1", and PCA was performed to calculate the principal component values

533 for each sample using PLINK1.9 [50]. Clusters were identified visually on the scatter plot

of the first and second principal components.

535 Allele and genotype frequencies were estimated for each ancestry group and biobank. The

536 fill-tags plugin of beftools was used for these calculations. To compare the allele

537 frequencies in the Japanese population, we downloaded the GEM Japan frequency panel

information from TogoVar. Since the GEM Japan panel only provides information in hg19

539 coordinates, we converted it to GRCh38 coordinates. We used GATK's LiftoverVcf

540 program for the conversion.

#### 541 HLA analysis

542 Three-field HLA alleles calling was performed using HLA-HD v1.3.0 [51] based on IPD-

543 IMGT/HLA v3.43.0 [52]; a score based on the weighted read counts considering variations

in and outside of the domain for antigen presentation was calculated to select the most

545 suitable pair of alleles amongst the candidate HLA alleles. To validate the accuracies of

546 HLA calling from WGS, HLA allele frequency distribution was compared with the HLA

547 frequency dataset from HLA Foundation Laboratory (Kyoto, Japan). To evaluate the

548 accuracy of HLA calling from the WGS dataset, a subset of the samples (n = 94) was

549 subjected to high-resolution experimental HLA genotyping for eight HLA genes (HLA-A, -

550 C, -B, -DRB1, -DQA1, -DQB1, -DPA1, and -DPB1) using next-generation sequencing and

551 AllType assay (One Lambda, West Hills, CA, US). Experimental HLA genotyping was

552 carried out following the vendor instructions, which consist of HLA gene amplification,

553 HLA library preparation, HLA template preparation, and HLA library loading onto an ion

554 530v1 chip (Thermo Fisher Scientific) in the Ion Chef (Thermo Fisher Scientific), followed

555 by final sequencing on the Ion S5 machine (Thermo Fisher Scientific). HLA genotype

assignments were carried out using HLATypeStream Visual (TSV v2.0; One Lambda,

557 West Hills, CA, US) and NGSengine® (v2.18.0.17625, GenDX, Utrecht, the Netherlands).

### 558 Haplotype phasing

559	Variant phasing was performed using shapeit v4.2 [53] in a haplotype-based analysis. SNPs
560	of unrelated samples identified using the ancestry inference were extracted for phasing. The
561	variant phasing was performed by dividing the autosomes into regions containing overlaps
562	for efficient computation. Each region was about 10 Mb in length, with a 500 kb overlap
563	margin at both ends. After phasing, VCFs were concatenated using the concat subcommand
564	in beftools.

### 565 Estimation of recent population size change

566 We estimated the effective population size change of the Japanese population from IBD

sharing, which can estimate the population size change in the recent past (~200 generations

ago) using WGS data [13]. Population size change was estimated for each population based

on the whole-genome data of Hondo (8,524 individuals) and Ryukyu (182 individuals).

570 First, the hapibd software [54] was used to detect the IBD segments shared by each

571 individual. For the genetic distance, we referred to the HapMap genetic map data

572 distributed with hapibd. We then estimated the population size change of the Hondo and

573 Ryukyu populations using IBDNe (ibdne.23Apr20.ae9.jar). The shortest threshold of the

574 IBD segment length was set at 2 cM.

#### 575 Estimation of genome-wide genealogy, estimation of population

### 576 size change, and detection of positive natural selection

577 We conducted the analysis of gene genealogy using the Relate software [14] to detect long-

term population size change and positive natural selection in Hondo and Ryukyu

579 populations. Relate is a software that can estimate genealogy on a genome-wide scale for

580 over 10,000 samples [55]. In this study, we used 1,000 randomly selected individual

genomes of Hondo, 182 Ryukyu samples, and 103 CHB samples of 1000 Genomes Project

582 [10]. First, input files (.haps, .samples) were created from vcf files using the

583 PrepareInputFiles.sh script in Relate software. We retrieved the Homo sapiens ancestral

sequences (GRCh38) of Ensembl 103 for the ancestral sequence and StrictMask of 1000

585 Genomes Project for genomic mask. Next, genome-wide genealogy was estimated using

the "Relate" command of Relate software packages. The mutation rate was set to  $1.25 \times 10^{-10}$ 

<sup>8</sup> per base per generation and the effective population size was set to 30,000. We assumed

588 28 years as the generation time in humans. The estimated genome-wide genealogy

589 (.anc, .mut) was used as input for population size estimation of Hondo and Ryukyu

590 populations using the EstimatePopulationSize.sh script. This script simultaneously

591 conducts estimation of population sizes, re-estimation of branch lengths using the estimated

592 population sizes, and estimation an average mutation rate. Finally, based on genome-wide

593 genealogy, we detected the target SNPs of positive natural selection acting on Hondo and

594 Ryukyu populations. Relate calculates a p-value of each SNP for positive selection that

595 quantifies how quickly a mutation has spread in the population. The p-values were

calculated for each SNP using the DetectSelection.sh script using the output genealogies of
population size estimation (.anc, .mut). We evaluated the quality of each SNP by
"RelateSelection –mode Quality," and SNPs inferred to be inaccurate tree estimation were
excluded.

### 600 Estimating the allele frequency trajectory

601 Changes in the allele frequency through the time were estimated using CLUES to infer

allele frequency trajectories [27]. CLUES uses the genome-wide genealogy inferred by

603 Relate. First, the sampleBranchLengths.sh script implemented in Relate was used to

604 MCMC sample the gene trees for the focal SNPs. Then, using the sampled tree file (.timeb)

as input, we estimated the allele frequency trajectory using CLUES's inference.py

606 command. The coalescence rate estimated by Relate (.coal file) can be used as an input to

607 modify the population size change using the -coal option. In this study, we estimated the

allele frequency trajectory by focusing on ALDH2 rs671, ADH1B rs1229984, OCA2

rs1800414, and FADS2 rs174600 among the SNPs that showed signals of natural selection

610 in Relate.

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- 639 Y.K., N.E., M.M., and T.K. designed the study. Y.O., R.Mi., H.G., K.Hata., K.Hatt., A.I.,
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- 641 Y.G., Y.Mar., Y.Mat., and S.N. contributed to the whole-genome sequencing. Y.O., R.Mi.,
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- 644 S.N., and K.T. contributed to the management of biobank. Y.K., W.Y., Y.O., and T.K.
- 645 wrote the manuscript with input from all authors.

# 646 Supporting information

647 Supporting information includes seven figures and three tables.

# 648 Competing interests

649 The authors declare no competing interests.

### 650 Web resources

- 651 IGSR: The International Genome Sample Resource, https://www.internationalgenome.org
- 652 Hap.py, https://github.com/Illumina/hap.py
- 653 GEM Japan, https://www.amed.go.jp/en/aboutus/collaboration/ga4gh\_gem\_japan.html
- 654 TogoVar, https://togovar.biosciencedbc.jp/
- 655 GATK, https://gatk.broadinstitute.org/hc/en-us
- 656 HLA frequency dataset from HLA Foundation Laboratory (Kyoto, Japan),
- 657 http://hla.or.jp/index.html
- 658 Homo sapiens ancestral sequences (GRCh38),
- 659 ftp://ftp.ensembl.org/pub/current\_fasta/ancestral\_alleles/homo\_sapiens\_ancestor\_GRCh38.t
- 660 ar.gz
- 661 dbSNP, https://www.ncbi.nlm.nih.gov/snp/

# 662 Data and code availability

- 663 The allele and genotype frequency data are available in the NBDC human database;
- Accession: hum0331. The raw genomic data are available upon request to corresponding
- authors and will soon be shared on a computational infrastructure currently under
- 666 construction by the Japan Agency for Medical Research and Development.

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# 668 **References**

669 670 671	1.	Turnbull C, Scott RH, Thomas E, Jones L, Murugaesu N, Pretty FB, et al. The 100 000 Genomes Project: bringing whole genome sequencing to the NHS. BMJ. 2018;361: k1687. doi: 10.1136/bmj.k1687.
672 673 674	2.	Taliun D, Harris DN, Kessler MD, Carlson J, Szpiech ZA, Torres R, et al. Sequencing of 53,831 diverse genomes from the NHLBI TOPMed Program. Nature. 2021;590: 290–299. doi: 10.1038/s41586-021-03205-y.
675 676	3.	The 1000 Genomes Project Consortium. An integrated map of genetic variation from 1,092 human genomes. Nature. 2012;491: 56–65. doi: 10.1038/nature11632.
677 678 679	4.	Fu W, O'Connor TD, Jun G, Kang HM, Abecasis G, Leal SM, et al. Analysis of 6,515 exomes reveals the recent origin of most human protein-coding variants. Nature. 2013;493: 216–220. doi: 10.1038/nature11690.
680 681 682	5.	The Genome of the Netherlands Consortium. Whole-genome sequence variation, population structure and demographic history of the Dutch population. Nat Genet. 2014;46: 818–825. doi: 10.1038/ng.3021.
683 684 685	6.	Nagasaki M, Yasuda J, Katsuoka F, Nariai N, Kojima K, Kawai Y, et al. Rare variant discovery by deep whole-genome sequencing of 1,070 Japanese individuals. Nat Commun. 2015;6: 8018. doi: 10.1038/ncomms9018.
686 687	7.	Yamaguchi-Kabata Y, Nakazono K, Takahashi A, Saito S, Hosono N, Kubo M, et al. Japanese population structure, based on SNP genotypes from 7003 individuals

688 compared to other ethnic groups: Effects on population-based association studies.
689 Am J Hum Genet. 2008;83: 445–456. doi: 10.1016/j.ajhg.2008.08.019.

- 8. Jinam T, Nishida N, Hirai M, Kawamura S, Oota H, Umetsu K, et al. The history of
  human populations in the Japanese Archipelago inferred from genome-wide SNP
  data with a special reference to the Ainu and the Ryukyuan populations. J Hum
  Genet. 2012;57: 787–95. doi: 10.1038/jhg.2012.114.
- Watanabe Y, Isshiki M, Ohashi J. Prefecture-level population structure of the
  Japanese based on SNP genotypes of 11,069 individuals. J Hum Genet. 2021;66:
  431–437. doi: 10.1038/s10038-020-00847-0.
- Byrska-Bishop M, Evani US, Zhao X, Basile AO, Abel HJ, Regier AA, et al. High
  coverage whole genome sequencing of the expanded 1000 Genomes Project cohort
  including 602 trios. bioRxiv. 2021; 2021.02.06.430068. doi:
  10.1101/2021.02.06.430068.
- 11. Lek M, Karczewski KJ, Minikel E V, Samocha KE, Banks E, Fennell T, et al.
  Analysis of protein-coding genetic variation in 60,706 humans. Nature. 2016;536:
  285–291. doi: 10.1038/nature19057.
- Landrum MJ, Lee JM, Benson M, Brown G, Chao C, Chitipiralla S, et al. ClinVar:
  public archive of interpretations of clinically relevant variants. Nucleic Acids Res.
  2016;44: D862–D868. doi: 10.1093/nar/gkv1222.
- Browning SR, Browning BL. Accurate non-parametric estimation of recent effective population size from segments of identity by descent. Am J Hum Genet. 2015;97:
  404–418. doi: 10.1016/j.ajhg.2015.07.012.
- 710 14. Speidel L, Forest M, Shi S, Myers SR. A method for genome-wide genealogy
  711 estimation for thousands of samples. Nat Genet. 2019;51: 1321–1329. doi:
  712 10.1038/s41588-019-0484-x.
- Sato T, Nakagome S, Watanabe C, Yamaguchi K, Kawaguchi A, Koganebuchi K, et
  al. Genome-wide SNP analysis reveals population structure and demographic history
  of the Ryukyu islanders in the southern part of the Japanese Archipelago. Mol Biol
  Evol. 2014;31: 2929–2940. doi: 10.1093/molbev/msu230.
- Matsunami M, Koganebuchi K, Imamura M, Ishida H, Kimura R, Maeda S. Finescale genetic structure and demographic history in the Miyako Islands of the Ryukyu
  Archipelago. Mol Biol Evol. 2021;38: 2045–2056. doi: 10.1093/molbev/msab005.
- Harada S, Agarwal DP, Goedde HW. Aldehyde dehydrogenase deficiency as cause
  of facial flushing reaction to alcohol in Japanese. Lancet. 1981;2: 982. doi:
  10.1016/s0140-6736(81)91172-7.

18. Edenberg HJ, McClintick JN. Alcohol dehydrogenases, aldehyde dehydrogenases,
and alcohol use disorders: A critical review. Alcohol Clin Exp Res. 2018;42: 2281–
2297. doi: 10.1111/acer.13904.

- Donnelly MP, Paschou P, Grigorenko E, Gurwitz D, Barta C, Lu R-B, et al. A global view of the OCA2-HERC2 region and pigmentation. Hum Genet. 2012;131: 683–
  696. doi: 10.1007/s00439-011-1110-x.
- Yang Z, Zhong H, Chen J, Zhang X, Zhang H, Luo X, et al. A genetic mechanism
  for convergent skin lightening during recent human evolution. Mol Biol Evol.
  2016;33: 1177–1187. doi: 10.1093/molbev/msw003.
- Shido K, Kojima K, Yamasaki K, Hozawa A, Tamiya G, Ogishima S, et al.
  Susceptibility loci for tanning ability in the Japanese population identified by a
  genome-wide association study from the Tohoku Medical Megabank Project Cohort
  Study. J Invest Dermatol. 2019;139: 1605-1608.e13. doi: 10.1016/j.jid.2019.01.015.
- Mathias RA, Fu W, Akey JM, Ainsworth HC, Torgerson DG, Ruczinski I, et al.
  Adaptive evolution of the FADS gene cluster within Africa. PLoS One. 2012;7:
  e44926. doi: 10.1371/journal.pone.0044926.
- Mathieson I, Lazaridis I, Rohland N, Mallick S, Patterson N, Roodenberg SA, et al.
  Genome-wide patterns of selection in 230 ancient Eurasians. Nature. 2015;528: 499–
  503. doi: 10.1038/nature16152.
- 742 24. Kothapalli KSD, Ye K, Gadgil MS, Carlson SE, O'Brien KO, Zhang JY, et al.
  743 Positive selection on a regulatory insertion-deletion polymorphism in FADS2
  744 influences apparent endogenous synthesis of arachidonic acid. Mol Biol Evol.
  745 2016;33: 1726–1739. doi: 10.1093/molbev/msw049.
- Buckley MT, Racimo F, Allentoft ME, Jensen MK, Jonsson A, Huang H, et al.
  Selection in Europeans on fatty acid desaturases associated with dietary changes.
  Mol Biol Evol. 2017;34: 1307–1318. doi: 10.1093/molbev/msx103.
- 749 26. Mathieson S, Mathieson I. FADS1 and the timing of human adaptation to agriculture. Mol Biol Evol. 2018;35: 2957–2970. doi: 10.1093/molbev/msy180.
- 751 27. Stern AJ, Wilton PR, Nielsen R. An approximate full-likelihood method for inferring
  752 selection and allele frequency trajectories from DNA sequence data. PLoS Genet.
  753 2019;15: e1008384. doi: 10.1371/journal.pgen.1008384.
- Nagasaki M, Yasuda J, Katsuoka F, Nariai N, Kojima K, Kawai Y, et al. Rare
  variant discovery by deep whole-genome sequencing of 1,070 Japanese individuals.
  Nat Commun. 2015;6: 8018. doi: 10.1038/ncomms9018.

- Gudbjartsson DF, Helgason H, Gudjonsson SA, Zink F, Oddson A, Gylfason A, et
  al. Large-scale whole-genome sequencing of the Icelandic population. Nat Genet.
  2015;47: 435–444. doi: 10.1038/ng.3247.
- 30. Suzuki H. Discoveries of the fossil man from Okinawa Island. Anthropol Sci.
  1975;83: 113–124. doi: 10.1537/ase1911.83.113.
- Nakagawa R, Doi N, Nishioka Y, Nunami S, Yamauchi H, Fujita M, et al.
  Pleistocene human remains from Shiraho-Saonetabaru Cave on Ishigaki Island,
  Okinawa, Japan, and their radiocarbon dating. Anthropol Sci. 2010;118: 173–183.
  doi: 10.1537/ase.091214.
- MacArthur DG, Balasubramanian S, Frankish A, Huang N, Morris J, Walter K, et al.
  A systematic survey of loss-of-function variants in human protein-coding genes.
  Science. 2012;335: 823–828. doi: 10.1126/science.1215040.
- Xarczewski KJ, Francioli LC, Tiao G, Cummings BB, Alföldi J, Wang Q, et al. The mutational constraint spectrum quantified from variation in 141,456 humans. Nature.
  2020;581: 434–443. doi: 10.1038/s41586-020-2308-7.
- 772 34. Cui R, Kamatani Y, Takahashi A, Usami M, Hosono N, Kawaguchi T, et al.
  773 Functional variants in ADH1B and ALDH2 coupled with alcohol and smoking
  774 synergistically enhance esophageal cancer risk. Gastroenterology. 2009;137: 1768–
  775 1775. doi: 10.1053/j.gastro.2009.07.070.
- Matoba N, Akiyama M, Ishigaki K, Kanai M, Takahashi A, Momozawa Y, et al.
  GWAS of 165,084 Japanese individuals identified nine loci associated with dietary
  habits. Nat Hum Behav. 2020;4: 308–316. doi: 10.1038/s41562-019-0805-1.
- 36. Oota H, Pakstis AJ, Bonne-Tamir B, Goldman D, Grigorenko E, Kajuna SLB, et al.
  The evolution and population genetics of the ALDH2 locus: random genetic
  drift, selection, and low levels of recombination. Ann Hum Genet. 2004;68: 93–109.
  doi: 10.1046/j.1529-8817.2003.00060.x.
- 37. Han Y, Gu S, Oota H, Osier M V, Pakstis AJ, Speed WC, et al. Evidence of positive selection on a class I ADH locus. Am J Hum Genet. 2007;80: 441–456. doi: 10.1086/512485.
- 38. Luo H-R, Wu G-S, Pakstis AJ, Tong L, Oota H, Kidd KK, et al. Origin and dispersal of atypical aldehyde dehydrogenase ALDH2\*487Lys. Gene. 2009;435: 96–103. doi: 10.1016/j.gene.2008.12.021.
- 39. Koganebuchi K, Haneji K, Toma T, Joh K, Soejima H, Fujimoto K, et al. The allele
  frequency of ALDH2\*Glu504Lys and ADH1B\*Arg47His for the Ryukyu islanders
  and their history of expansion among East Asians. Am J Hum Biol. 2017;29:
  e22933. doi: 10.1002/ajhb.22933.

- Fumagalli M, Moltke I, Grarup N, Racimo F, Bjerregaard P, Jørgensen ME, et al.
  Greenlandic Inuit show genetic signatures of diet and climate adaptation. Science.
  2015;349: 1343–1347. doi: 10.1126/science.aab2319.
- Mallick S, Li H, Lipson M, Mathieson I, Gymrek M, Racimo F, et al. The Simons
  Genome Diversity Project: 300 genomes from 142 diverse populations. Nature.
  2016;538: 201–206. doi: 10.1038/nature18964.
- Franke KR, Crowgey EL. Accelerating next generation sequencing data analysis: an
  evaluation of optimized best practices for Genome Analysis Toolkit algorithms.
  Genomics Inform. 2020;18: e10. doi: 10.5808/GI.2020.18.1.e10.
- 43. Li H, Durbin R. Fast and accurate short read alignment with Burrows–Wheeler transform. Bioinformatics. 2009;25: 1754–1760. doi: 10.1093/bioinformatics/btp324.
- 44. Danecek P, Bonfield JK, Liddle J, Marshall J, Ohan V, Pollard MO, et al. Twelve
  years of SAMtools and BCFtools. GigaScience. 2021;10: giab008. doi:
  10.1093/gigascience/giab008.
- Freed D, Aldana R, Weber JA, Edwards JS. The Sentieon Genomics Tools A fast
  and accurate solution to variant calling from next-generation sequence data. bioRxiv.
  2017; 115717. doi: 10.1101/115717.
- 46. McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GRS, Thormann A, et al. The
  ensembl variant effect predictor. Genome Biol. 2016;17: 122. doi: 10.1186/s13059016-0974-4.
- 47. Liu X, Li C, Mou C, Dong Y, Tu Y. dbNSFP v4: a comprehensive database of
  transcript-specific functional predictions and annotations for human nonsynonymous
  and splice-site SNVs. Genome Med. 2020;12: 103. doi: 10.1186/s13073-020-008039.
- Kawai Y, Mimori T, Kojima K, Nariai N, Danjoh I, Saito R, et al. Japonica array:
  improved genotype imputation by designing a population-specific SNP array with
  1070 Japanese individuals. J Hum Genet. 2015;60: 581–587. doi:
  10.1038/jhg.2015.68.
- 49. Manichaikul A, Mychaleckyj JC, Rich SS, Daly K, Sale M, Chen W-M. Robust
  relationship inference in genome-wide association studies. Bioinformatics. 2010;26:
  2867–2873. doi: 10.1093/bioinformatics/btq559.
- So. Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation PLINK: rising to the challenge of larger and richer datasets. GigaScience.
  2015;4: 7. doi: 10.1186/s13742-015-0047-8.

Kawaguchi S, Higasa K, Shimizu M, Yamada R, Matsuda F. HLA-HD: An accurate
HLA typing algorithm for next-generation sequencing data. Hum Mutat. 2017;38:
788–797. doi: 10.1002/humu.23230.

- 831 52. Robinson J, Barker DJ, Georgiou X, Cooper MA, Flicek P, Marsh SGE. IPD832 IMGT/HLA database. Nucleic Acids Res. 2020;48: D948–D955. doi:
  833 10.1093/nar/gkz950.
- 53. Delaneau O, Zagury J-F, Robinson MR, Marchini JL, Dermitzakis ET. Accurate,
  scalable and integrative haplotype estimation. Nature Commun. 2019;10: 5436. doi:
  10.1038/s41467-019-13225-y.
- 54. Zhou Y, Browning SR, Browning BL. A fast and simple method for detecting
  identity-by-descent segments in large-scale data. Am J Hum Genet. 2020;106: 426–
  437. doi: 10.1016/j.ajhg.2020.02.010.
- Speidel L, Forest M, Shi S, Myers SR. A method for genome-wide genealogy
  estimation for thousands of samples. Nat Genet. 2019;51: 1321–1329. doi:
  10.1038/s41588-019-0484-x.
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#### 853 **Figure legends**

854 Fig 1. Quality control metrics of whole-genome sequencing. Quality control metrics for 855 each sample are plotted against the sample in the horizontal axis direction; QC indices are 856 (A) average coverage of reads in autosomal loci after excluding duplicated reads, (B) 857 mapping rate, and (C) average insert length. Saliva-derived samples are colored by yellow. 858 Fig 2. Genetic structure of NCBN samples. (A) The first and second principal 859 components are plotted. The continental population of the international 1000 genomes and 860 NCBN are plotted in different colors and shapes. (B) PCA plots of the East Asian 861 population of the International 1000 Genomes and NCBN samples are shown. JPT: 862 Japanese in Tokyo, Japan, CHB: Han Chinese in Beijing, China, CHS: Han Chinese South, 863 KHV: Kinh in Ho Chi Minh City, Vietnam, CDX: Chinese Dai in Xishuangbanna, China 864 Fig 3. Comparison of allele frequency between different populations. (A) The non-865 reference allele frequencies of the Hondo population of NCBN samples (X-axis) and the 866 corresponding variants of GEM Japan (Y-axis) were counted and then the numbers were 867 plotted as density. (B) Same plot for Hondo population (X-axis) and Ryukyu population 868 (Y-axis). 869 Fig 4. Analysis of loss-of-function (LoF) variants. (A) The allele frequency distribution 870 of newly detected HC LoF SNPs in the Hondo population. (B) The number of LoF alleles 871 and (C) the number of homozygous of LoF alleles per individual for Hondo Ryukyu, and

populations of the International 1000 Genomes. (D) The number of homozygous of LoF

873 a	alleles per	individual	by allele	frequency	for Hondo,	Ryukyu,	and the po	pulations of	of the
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874 International 1000 Genomes.

875	Fig 5. Estimation of	past po	pulation size	from IBD	sharing. (A	) Short-term effective
				-		/

- 876 population size change in Hondo and Ryukyu populations by IBDNe. (B) Distribution of
- 877 IBD segment length in Hondo. (C) Distribution of IBD segment length in Ryukyu.
- 878 Fig 6. Trajectories of allele frequency of genes. Allele frequency trajectories of (A)
- ADH1B rs1229984, (B) ALDH2 rs671, (C) OCA2 rs1800414, and (D) FADS1 rs174599
- are shown.

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#### 890 Supporting information

- 891 S1 Fig. Genetic structure of East Asian populations. The clusters consisting of the
- 892 NCBN samples in Figure 2 are classified into Hondo (black), Ryukyu (orange), and others

893 (blue).

- 894 S2 Fig. Analysis of Loss-of-function (LoF) variants. The numbers of LoF sites per
- individual by category are presented: (A) and (B) stop gained SNV; (C) and (D) splice site
- 896 SNV; (E) and (F) frameshift INDELs.
- 897 S3 Fig. HLA alleles frequencies (%) between NCBN vs HLA Foundation Laboratory,
- 898 Kyoto, Japan. Comparison for class I HLA genes (HLA-A, -C, -B) (left). Comparison for
- class II HLA genes (HLA-DRB1, -DQA1, -DQB1, -DPA1, -DPB1) (right). Only common
- 900 HLA alleles (HLA frequencies > 1%) are included in this analysis.
- 901 S4 Fig. Long-term effective population size change of Hondo, Ryukyu, and Han
- 902 Chinese. The changes in population size were estimated from the gene genealogy across903 the genome.
- 904 S5 Fig. Manhattan plot of the selection scan result of the whole-genome SNPs by
- 905 **Relate.** The red line represents the genome-wide significance level  $(5 \times 10^{-8})$ .
- 906 S6 Fig. QQ plot of the selection scan result of the whole-genome SNPs by Relate. The
- 907 red line denotes y=x.

- 908 S7 Fig. Gene genealogy estimated by Relate. Genealogy of (a) ALDH2 rs671, (b)
- ADH1B rs1229984 (c) OCA2 rs1800414 (d) FADS1 rs174599 are presented. The vertical
- 910 axis represents the age (years before present). Derived allele carriers are shown in red.
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## Figure 1.

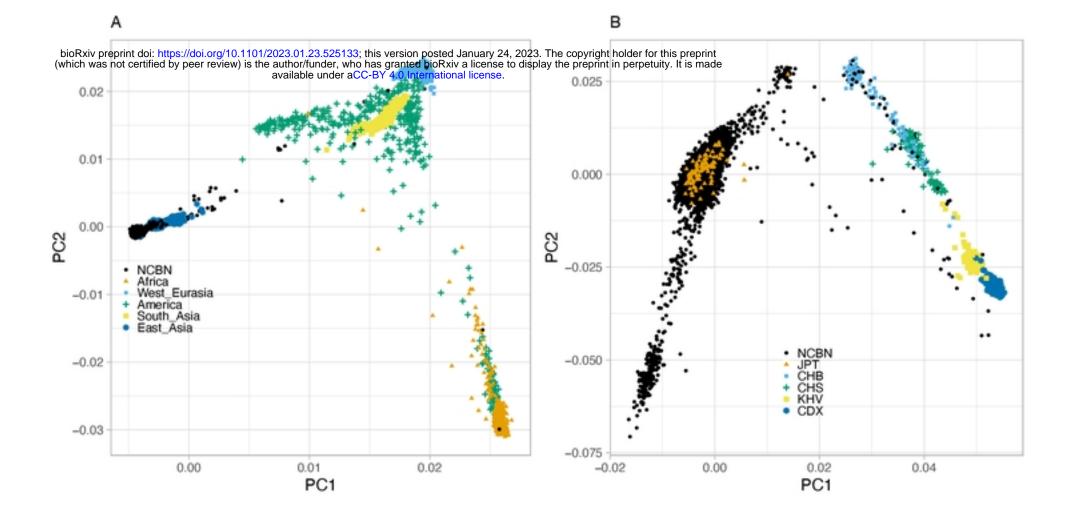
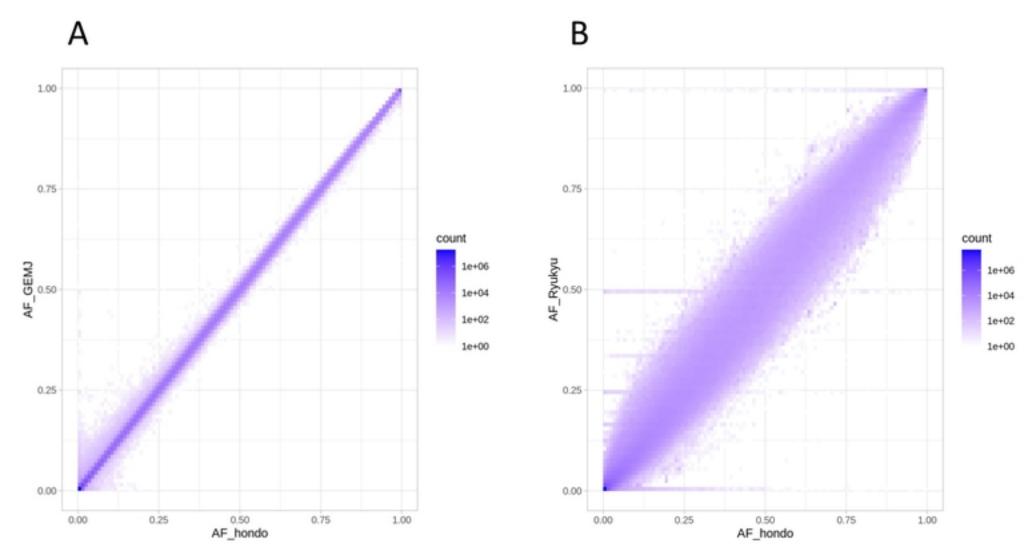
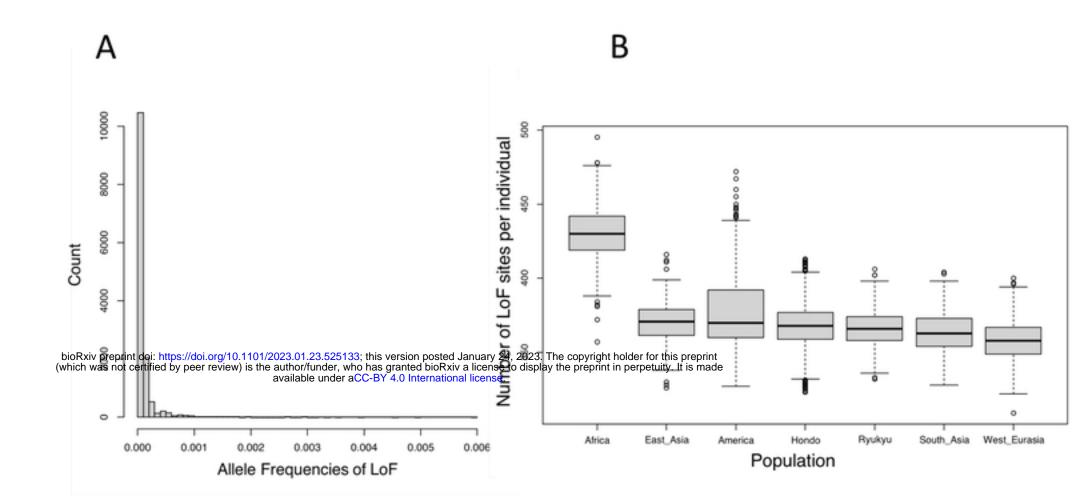
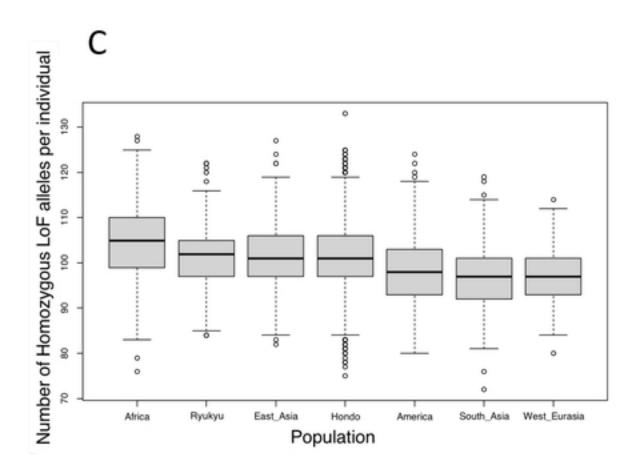


Figure 2.

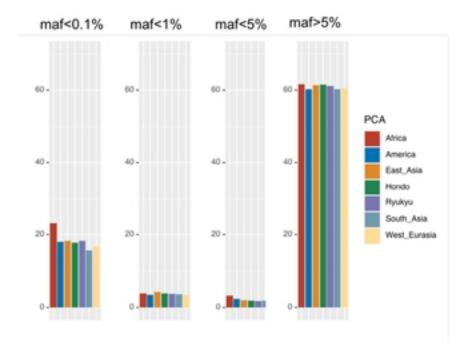


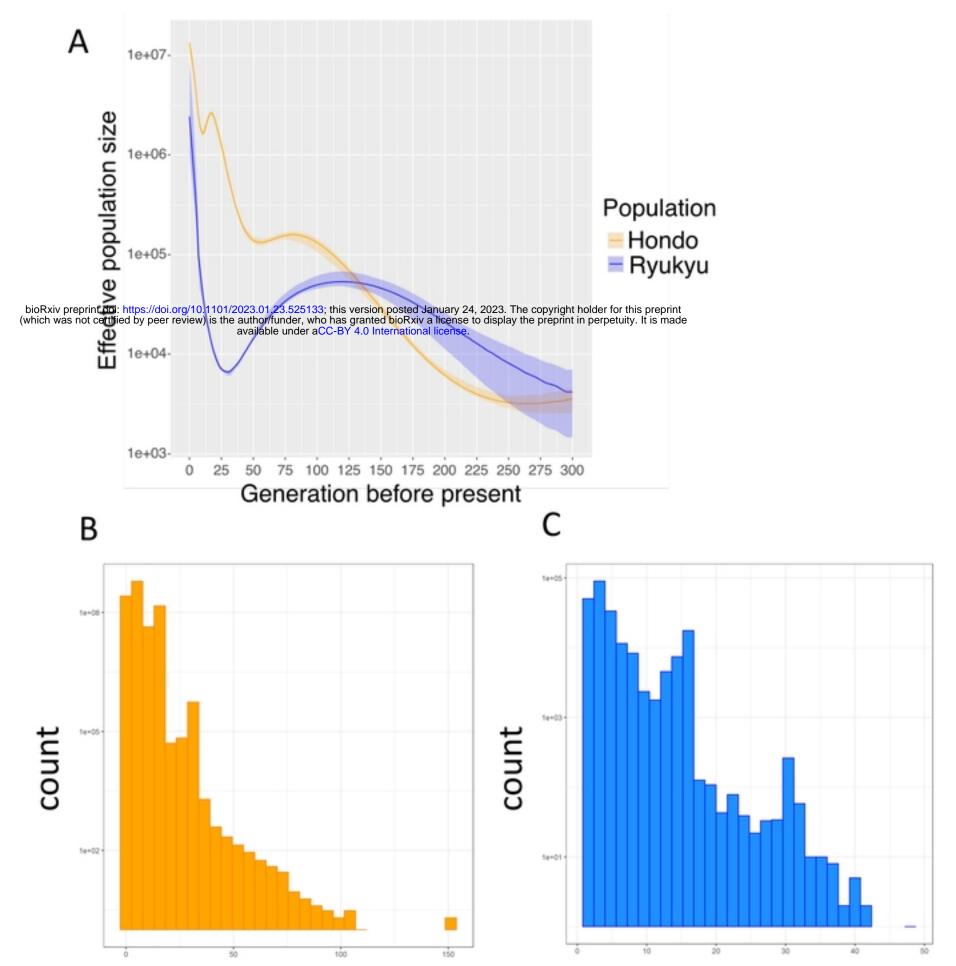
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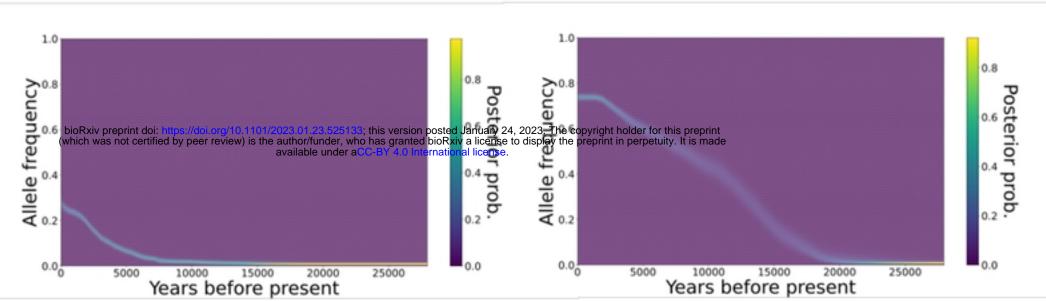
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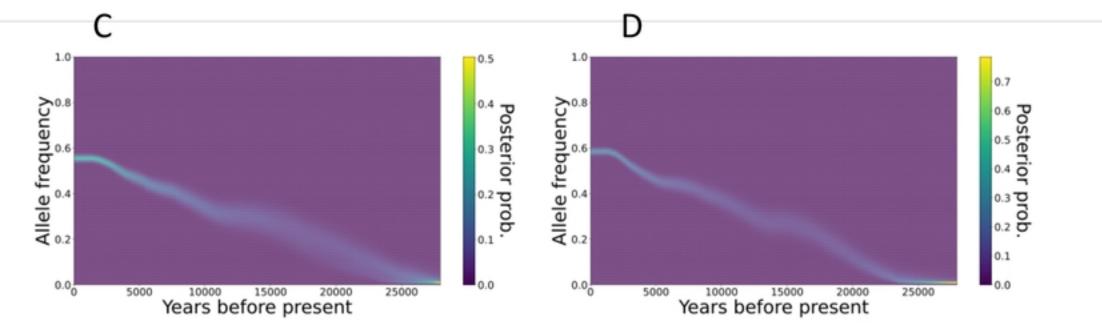




## IBD sharing (cM)

## IBD sharing (cM)





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