Mangrove propagules are limited in their capacity to disperse across long distances

Natasha R. GRANVILLE[1*] and Cristina BANKS-LEITE[1]

[1] Imperial College London, Silwood Park Campus, Ascot, Berkshire SL5 7PY, UK

*Author for correspondence: natasha.granville19@imperial.ac.uk ; tasha.granville@gmail.com

Keywords: Mangroves, isolation-by-distance, dispersal limitation, habitat connectivity, meta-analysis.

Abstract

Mangroves are subject to rapid and large-scale habitat changes which threaten their unique genetic diversity and provision of critically important ecosystem services. Habitat fragmentation reduces connectivity which can impair dispersal and lead to genetic isolation. However, it is unclear whether fragmentation could impact mangrove genetic isolation, as mangrove propagules can disperse long distances. Here, we conducted a meta-analysis of studies reporting a correlation between geographic distance and genetic distance in mangrove plants. From the 22 studies that met the inclusion criteria, we found a significant isolation-by-distance effect; geographic distance was significantly associated with Nei's genetic distance and F_{ST}. Our results show that mangrove propagules may be limited in their capacity to disperse across long distances, which highlights the importance of maintaining close proximity between habitat patches and reducing habitat fragmentation.

1 Introduction

Habitat loss is known to impact dispersal across habitats, which is vital for maintaining genetic diversity and for range shifts in response to environmental changes (Van der Stocken et al. 2019a). Understanding the relationship between spatial and genetic structuring of populations is fundamental for developing effective conservation strategies that maintain habitat connectivity (Durrant et al. 2014, Taylor et al. 2021, Wright et al. 2015), and this understanding is contingent on characterising dispersal capabilities.

Mangrove forests are intertidal wetlands, found along coastlines in tropical, sub-tropical and warm-temperate climates (Bryan-Brown et al. 2020). Globally, there are two main hotspots of mangrove biodiversity; the Indo-West Pacific contains approximately 54 mangrove species, and the Atlantic-East Pacific contains approximately 17 mangrove species (Tomlinson 2016). Between 2000 and 2012, mangrove forests were lost at an average rate of 0.18 % per year in Southeast Asia (Richards & Friess 2016). Mangroves are threatened at the local- and regional- scales by aquaculture, agriculture, urban development and pollution, while simultaneously facing the broader scale threats of sea level rise and climate change. Mangrove forests are of exceptionally high ecological and economic value (estimated $194,000 per hectare per year, Costanza et al. 2014) as they provide unique genetic diversity, and crucial ecosystem services (Bryan-Brown et al. 2020; Mantiquilla et al. 2021). The provision of these services depends on the size and arrangement of forest patterns (Bryan-Brown et al. 2020) Even in areas with low rates of mangrove loss, there is a global trend towards ubiquitous fragmentation, which can pose a threat to mangrove biodiversity (Bryan-Brown et al. 2020). Therefore, there is a need to quantify the impact of fragmentation on mangrove biodiversity.

While it is generally accepted that habitat loss has negative effects on biodiversity, the effects of habitat fragmentation *per se* (independent of habitat loss) are more variable and context-dependent (Andrén 1994, Fahrig 2003). On the one hand, Wilcove et al's 1986 definition of habitat fragmentation as a process whereby *"a large expanse of habitat is transformed into a number of smaller patches of smaller total area, isolated from each other by a matrix of habitats unlike the original"* (Wilcove 1986, as cited in Fahrig 2003) implies that fragmentation leads to increased isolation of patches (Fahrig, 2003). On the other hand, fragmentation *per se*

could theoretically result in many small patches that are less isolated from each other, assuming that dispersal across the matrix is possible (Fahrig 2003, Healey & Hovel 2004). In mangroves, however, the matrix between habitat patches is composed of water, which can be easily traversed by propagules and human-modified land use, which cannot be colonised by mangroves.

It is therefore important to understand the impacts of geographic distance on mangrove propagule dispersal. Buoyant mangrove propagules have the capacity for long-distance dispersal by water because they remain viable for extended periods of time, and can drift in ocean currents (Binks et al. 2019). This could be expected to lead to high connectivity between habitat patches, which could mean that increasing distance between patches would have little effect on genetic isolation. However, field studies have shown that mangrove propagules tend not to disperse far from their release point, leading to patterns of isolation-by-distance (Binks et al. 2019, Clarke 1993, Yan et al. 2016,Van der Stocken et al. 2015). Even in species with highly dispersive life history features, abiotic factors like ocean currents, coastal topography and habitat discontinuities can affect the realised dispersal distance (Cowen & Sponaugle 2009, Selkoe et al. 2016, Binks et al. 2019). Propagules of *Avicennia marina*, the world's most widely distributed mangrove species, can remain viable in water for several months (Rabinowitz 1978). However, their obligate dispersal period, during which they float before developing roots for anchoring, is only around one week (Rabinowitz 1978). Once they land on suitable substrate, they can root and grow rapidly because many taxa exhibit viviparity, where the seed germinates while attached to the parent plant (Clarke 1993, Ng & Sivasothi 2001). These traits, combined with the risks of dispersal, could explain why mangrove propagules might tend to favour establishment near their release point (Clarke 1993, Binks et al. 2019).

We conducted a meta-analysis to quantify the relationship between geographic distance and genetic distance in mangrove plant communities to better understand how far mangrove propagules disperse, with the aim of providing insight into the effect of patch isolation on genetic isolation in mangroves. Measures of genetic distance such as Nei's genetic distance (Nei 1972) and F_{ST} (Wright 1950) provide useful insights into the genetic structure of a habitat (Bohonak 1999). If mangrove patches are more isolated from each other and propagules cannot disperse between them, there will likely be a larger pairwise genetic distance between these patches (Jaquiéry et al. 2011).

Methods

Systematic literature search and inclusion criteria

In May 2021, we conducted an extensive search of the relevant literature, following the PRISMA (Preferred reporting items for systematic review and meta-analysis) statement which provides a standardised framework for reporting systematic reviews and meta-analyses (Moher et al. 2009). The following search terms were used in Web of Science and Scopus:

1. mangrove AND fragment* AND ('genetic diversity' OR 'genetic differentiation')
2. mangrove AND connect* AND ('genetic diversity' OR 'genetic differentiation')
3. mangrove AND isolat* AND ('genetic diversity' OR 'genetic differentiation')

An initial search, after removing duplicates, yielded 199 papers, which we screened for eligibility. This resulted in the exclusion of 148 non-relevant papers and 4 papers that we could not access. From the 47 papers that remained, we selected papers that reported the results of a Mantel test for the correlation between matrices of untransformed Euclidean geographic distance on Nei's genetic distance (11 papers, representing 13 case studies) or untransformed Wright's F_{ST} (8 papers, representing 9 case studies). Nei's genetic distance and F_{ST} were chosen as measures of genetic diversity because they were the most commonly reported so this helped maximise the sample size. We expect that our sample size of 22 studies is large enough to provide sufficient power to make meaningful conclusions (Jackson & Turner 2017). The analysis of the effect of geographic distance on Nei's genetic diversity is henceforth referred to as 'Nei's meta-analysis' and the analysis of the effect of geographic distance on Wright's F_{ST} is henceforth referred to as 'F_{ST} meta-analysis'

Statistical analysis

For the effect size, we extracted Pearson's *r* value from the reported Mantel test in each paper. Where R^2 was reported, it was converted to *r* by taking the square root (this was done in 2 papers for the Nei's meta-analysis and 8 papers for the F_{ST} meta-analysis). Each study was weighted according to the following formula developed by Reed & Frankham (2003) specifically for meta-analyses of genetic diversity: $\sqrt{(A-2) \times N}$, where $A =$ number of populations and $N =$ number of individuals for each paper.

Analyses were conducted using the metafor (Viechtbauer 2010) and robumeta (Fisher et al. 2017) packages in R

version 4.2.1 (R Core Team 2022). We used a random-effects model because studies were drawn from different

populations. For the analysis, Pearson's r was transformed to Fisher's Z to ensure normal distribution. After

performing meta-analytic calculations, Fisher's Z was converted back to Pearson's r for reporting summary

effect sizes (Quintana 2015).

The Q-statistic was calculated to assess heterogeneity among studies. The Q-statistic is the ratio of observed

variation to within-study variance. It evaluates the null hypothesis that all studies are examining the same effect

(Quintana 2015). Different studies used different mangrove plant species and different molecular markers to

assess genetic variation (Table S1). To assess the effect of this, we fitted separate meta-analytic models that

moderated for the effects of species and marker, respectively. To account for effect size dependency resulting

from the same study reporting multiple effect sizes (2 papers in the Nei's meta-analysis and 1 paper in the F_{ST}

meta-analysis), we used robust variance estimation as this is appropriate for meta-analyses with less than 40

studies and does not assume knowledge of within-study correlations (Quintana 2015). We used Egger's

regression test to assess publication bias by testing for funnel plot asymmetry. Publication bias is the

phenomenon whereby studies with larger effect sizes are more likely to be published, and therefore included in

the meta-analysis (Quintana 2015).

Results

Included studies

There were 13 case studies for Nei's genetic distance and 9 case studies for F_{ST} (Table S1, Figure 1). Egger's

regression test for funnel plot asymmetry showed no effect of publication bias (Nei's meta-analysis: $z = 0.158$, p

$= 0.875$. F_{ST} meta-analysis: $z = 1.31$, p $= 0.192$. Figure S1).

Figure 1. PRISMA flowchart (Moher et al. 2009) showing the sequence of selection of papers for meta-analysis

of the effects of geographic distance on genetic distance.

Effect sizes

We found a significant association between geographic distance and Nei's genetic distance (estimated model

coefficient = 0.37, 95 % CI = 0.14 - 0.56. z = 3.07, p = 0.002. Figure 2A), which was not changed when

accounting for effect size dependency by robust variance estimation (estimated model coefficient = 0.39, 95 %

CI = 0.12 – 0.66). We also found a significant association between geographic distance and F_{ST} (estimated model

coefficient = 0.63, 95 % CI = 0.41 – 0.78. z = 4.77, p < 0.0001. Figure 2b). This model coefficient was not

changed significantly when accounting for effect size dependency by robust variance estimation (estimated

model coefficient = 0.75, 95 % CI = 0.38 – 1.11).

Figure 2. Effect of geographic distance on (A) Nei's genetic distance and (B) genetic distance measured by F_{ST}

for the original unadjusted models, with Fisher's *Z* transformed back to Pearson's *r*. The polygon at the bottom

of each plot shows the estimated model coefficient and its bounds represent the 95 % confidence intervals (CIs).

Each squared point corresponds to a different study, as labelled. The size of the square corresponds to the

contribution of the study to the estimated model coefficient.

Heterogeneity and moderator analysis

There was significant heterogeneity among the included studies (Nei's meta-analysis: Q = 63.7, df = 12, p <

0.0001. F_{ST} meta-analysis Q = 91.6, df = 8, p < 0.0001). Li & Chen (2004) contributed disproportionately to the

overall heterogeneity in Nei's meta-analysis. Therefore, for the purpose of comparison, a separate random-effects model was fitted to the same data set but excluding Li & Chen (2004). This reduced the summary effect size from 0.37 (95 % CI = 0.14 - 0.56) to 0.29 (95 % CI = 0.12 - 0.45). The overall heterogeneity was reduced, but there was still significant heterogeneity (Q = 40.4, df = 11, p <0.0001). Since excluding this study did not significantly remove the heterogeneity, all other analyses include this study.

To determine the source of heterogeneity, we conducted mixed-effects moderator analyses with taxon and marker as separate moderator variables. The type of genetic marker significantly moderated the correlation between geographic distance and F_{ST} (QM = 12.0, df = 3, p = 0.0075), but not Nei's genetic distance (QM = 3.63, df = 4, p = 0.46). Whereas differences in the taxon investigated did not significantly moderate either of these correlations (Nei's genetic distance: QM = 12.5, df = 7, p = 0.086. F_{ST}: QM = 7.50, df = 6, p = 0.28).

Discussion

Our global meta-analysis showed a significant correlation between geographic distance and genetic distance in mangrove plant communities. This isolation-by-distance effect (Wright 1943) could suggest that mangrove plants are limited in their capacity to disperse across habitat patches. This is consistent with the conclusions made by Binks et al. (2019) that habitat discontinuities lead to reduced gene flow between patches because mangrove propagules tend not to disperse far from their release point, likely due to a combination of abiotic factors, viviparity and short obligate dispersal period (Clarke 1993, Binks et al. 2019). Maintaining gene flow, which is critical for long-term population persistence (Salm et al. 2000, Wright et al. 2015), will depend on maintaining proximity among habitat patches, especially under conditions of habitat transformation which threaten mangrove biodiversity.

Isolation-by-distance indicates that spatial structure and genetic structure are highly correlated and suggests that dispersal limitation may be important in mangrove communities. Dispersal is essential for enabling sessile organisms, such as plants, to move away from unfavourable conditions if they are unable to adapt to such conditions (Kinlan & Gaines 2003). Isolation-by-distance suggests that these important adaptive responses are constrained by natural dispersal mechanisms (Sexton et al. 2014). Though not explicitly considered in this study,

isolation-by-environment and isolation-by-resistance are relevant to understanding the factors underpinning the genetic structure of mangrove populations. Isolation-by-resistance includes environmental factors, such as land-use changes and biogeographic barriers, that affect the ability of propagules to disperse between patches of suitable habitat. This can affect the genetic structure of populations by modulating the impact of geographic distance on genetic distance (Wang & Bradburd 2014). Furthermore, environmental factors such as habitat heterogeneity can affect the likelihood of gene flow among populations; and these isolation-by-environment factors can act in combination with geographic distance to drive the genetic structure of populations (McRae 2006).

If mangrove propagules are limited in their dispersal capabilities, populations in habitat patches are more likely to become isolated from each other. This could result in a meta-population structure with smaller populations that are more vulnerable to demographic and environmental stochasticity (Lande 1993). Furthermore, the apparent dispersal limitation of mangrove propagules could limit the potential for mangroves to shift their distributional ranges to track changing climatic conditions (Van der Stocken et al. 2022). While we recognise that dispersal depends on several biotic and abiotic factors that affect the release, transport and establishment of propagules (Van der Stocken et al. 2019b), the isolation-by-distance effect shown here highlights the importance of geographic distance in constraining gene flow and suggests that mangrove propagules are limited in their tendency to disperse across long distances. This may be exacerbated by future climatic warming and rising sea levels. Recent analysis by Van der Stocken et al. (2022) indicates that mangrove propagules in fresher and warmer oceans are likely to have increased rates of sinking, which reduces the likelihood of long distance dispersal, especially for widespread mangrove species with dense propagules such as *Avicennia marina* (Van der Stocken et al. 2022). Therefore, when conserving and managing mangroves, the importance of maintaining close proximity between habitat fragments should be considered.

Our results indicate that the genetic structure of mangrove communities is dependent on spatial structure. Existing efforts to restore deforested mangrove forests often involve artificial movement of propagules (Vanderklift et al. 2020) which might be relevant for assisting dispersal if this is needed to maintain connectivity. Furthermore, for protected area networks to successfully maintain landscape connectivity, the size

and arrangement of these networks should reflect the dispersal capabilities of the inhabiting species (Durrant et al. 2014, Shanks et al. 2003). Therefore, optimal design of protected area networks requires knowledge of effective dispersal distances. While the present study does not address exact distances, our results suggest that the realised dispersal capabilities of mangrove propagules depend heavily on the geographic distances across which they are dispersing. This emphasises the need for future studies to quantify effective dispersal distances in mangroves and consider how mangrove dispersal could be affected by habitat change.

Financial support

This research received no specific grant from any funding agency, commercial or not-for-profit sectors.

Competing interest declaration

Competing interests: The authors declare none.

Acknowledgements

We thank two anonymous reviewers for helpful comments on an earlier version of the manuscript.

References

ALBRECHT, M., KNEELAND, K. M., LINDROTH, E. & FOSTER, J. E. 2013. Genetic diversity and relatedness of the mangrove Rhizophora mangle L. (Rhizophoraceae) using amplified fragment polymorphism (AFLP) among locations in Florida, USA and the Caribbean. *Journal of Coastal Conservation* 17:483–491. Springer.

AL-QTHANIN, R. N. & ALHARBI, S. A. 2020. Spatial Structure and Genetic Variation of a Mangrove Species (Avicennia marina (Forssk.) Vierh) in the Farasan Archipelago. *Forests* 11:1287. Multidisciplinary Digital Publishing Institute.

ANDRÉN, H. 1994. Effects of Habitat Fragmentation on Birds and Mammals in Landscapes with Different Proportions of Suitable Habitat: A Review. *Oikos* 71:355–366. [Nordic Society Oikos, Wiley].

ARBELÁEZ-CORTES, E., CASTILLO-CÁRDENAS, M. F., TORO-PEREA, N. & CÁRDENAS-HENAO, H. 2007. Genetic structure of the red mangrove (Rhizophora mangle L.) on the Colombian Pacific detected by microsatellite molecular markers. *Hydrobiologia* 583:321–330.

AZMAN, A., NG, K.-K.-S., NG, C.-H., LEE, C.-T., TNAH, L.-H., ZAKARIA, N.-F., MAHRUJI, S., PERDAN, K., ABDUL-KADIR, M.-Z., CHENG, A. & LEE, S.-L. 2020. Low genetic diversity indicating the threatened status of Rhizophora apiculata (Rhizophoraceae) in Malaysia: declined evolution meets habitat destruction. *Scientific Reports* 10:19112. Nature Publishing Group.

BINKS, R. M., BYRNE, M., MCMAHON, K., PITT, G., MURRAY, K. & EVANS, R. D. 2019. Habitat discontinuities form strong barriers to gene flow among mangrove populations, despite the capacity for long-distance dispersal. *Diversity and Distributions* 25:298–309.

BOHONAK, A. J. 1999. Dispersal, Gene Flow, and Population Structure. *The Quarterly Review of Biology* 74:21–45. The University of Chicago Press.

BRYAN-BROWN, D. N., CONNOLLY, R. M., RICHARDS, D. R., ADAME, F., FRIESS, D. A. & BROWN, C. J. 2020. Global trends in mangrove forest fragmentation. *Scientific Reports* 10:7117. Nature Publishing Group.

CASTILLO-CÁRDENAS, M. F., TORO-PEREA, N. & CÁRDENAS-HENAO, H. 2005. Population Genetic Structure of Neotropical Mangrove Species on the Colombian Pacific Coast: Pelliciera rhizophorae (Pellicieraceae)1. *Biotropica* 37:266–273.

CERÓN-SOUZA, I., TORO-PEREA, N. & CÁRDENAS-HENAO, H. 2005. Population Genetic Structure of Neotropical Mangrove Species on the Colombian Pacific Coast: Avicennia germinans (Avicenniaceae)1. *Biotropica* 37:258–265.

CISNEROS-DE LA CRUZ, D. J., MARTÍNEZ-CASTILLO, J., HERRERA-SILVEIRA, J., YÁÑEZ-ESPINOSA, L., ORTIZ-GARCÍA, M., US-SANTAMARIA, R. & ANDRADE, J. L. 2018. Short-distance barriers affect genetic variability of Rhizophora mangle L. in the Yucatan Peninsula. *Ecology and Evolution* 8:11083–11099.

CLARKE, P. J. 1993. Dispersal of grey mangrove (Avicennia marina) propagules in southeastern Australia. *Aquatic Botany* 45:195–204.

COSTANZA, R., DE GROOT, R., SUTTON, P., VAN DER PLOEG, S., ANDERSON, S. J., KUBISZEWSKI, I., FARBER, S. & TURNER, R. K. 2014. Changes in the global value of ecosystem services. *Global Environmental Change* 26:152–158.

COWEN, R. K. & SPONAUGLE, S. 2009. Larval Dispersal and Marine Population Connectivity. *Annual Review of Marine Science* 1:443–466.

DA SILVA, M. F., CRUZ, M. V., VIDAL JÚNIOR, J. D. D., ZUCCHI, M. I., MORI, G. M. & DE SOUZA, A. P. 2021. Geographical and environmental contributions to genomic divergence in mangrove forests. *Biological Journal of the Linnean Society* 132:573–589.

DENG, S., HUANG, Y., HE, H., TAN, F., NI, X., JAYATISSA, L. P., HETTIARACHI, S. & SHI, S. 2009. Genetic diversity of Aegiceras corniculatum (Myrsinaceae) revealed by amplified fragment length polymorphism (AFLP). *Aquatic Botany* 90:275–281.

DURRANT, H. M. S., BURRIDGE, C. P., KELAHER, B. P., BARRETT, N. S., EDGAR, G. J. & COLEMAN, M. A. 2014. Implications of Macroalgal Isolation by Distance for Networks of Marine Protected Areas. *Conservation Biology* 28:438–445.

FAHRIG, L. 2003. Effects of Habitat Fragmentation on Biodiversity. *Annual Review of Ecology, Evolution, and Systematics* 34:487–515.

FISHER, Z., TIPTON, E. & ZHIPENG, H. 2017, May 29. robumeta: Robust Variance Meta-Regression.

GUO, W., NG, W. L., WU, H., LI, W., ZHANG, L., QIAO, S., YANG, X., SHI, X. & HUANG, Y. 2018. Chloroplast phylogeography of a widely distributed mangrove species, Excoecaria agallocha, in the Indo-West Pacific region. *Hydrobiologia* 807:333–347.

HEALEY, D. & HOVEL, K. A. 2004. Seagrass bed patchiness: effects on epifaunal communities in San Diego Bay, USA. *Journal of Experimental Marine Biology and Ecology* 313:155–174.

JACKSON, D. & TURNER, R. 2017. Power analysis for random-effects meta-analysis. *Research Synthesis Methods* 8:290–302.

JAQUIÉRY, J., BROQUET, T., HIRZEL, A. H., YEARSLEY, J. & PERRIN, N. 2011. Inferring landscape effects on dispersal from genetic distances: how far can we go? *Molecular Ecology* 20:692–705.

JIAN, S. & SHI, S. 2009. Genetic variation in Heritiera littoralis (Malvaceae) from east and south Asia revealed by AFLP markers. *Plant Biosystems - An International Journal Dealing with all Aspects of Plant Biology* 143:S56–S62. Taylor & Francis.

KINLAN, B. P. & GAINES, S. D. 2003. Propagule Dispersal in Marine and Terrestrial Environments: A Community Perspective. *Ecology* 84:2007–2020.

LANDE, R. 1993. Risks of Population Extinction from Demographic and Environmental Stochasticity and Random Catastrophes. *The American Naturalist*. University of Chicago Press.

LI, H.-S. & CHEN, G.-Z. 2004. Genetic Diversity of Sonneratia alba in China Detected by Inter-simple Sequence Repeats (ISSR) Analysis. *Journal of Integrative Plant Biology* 46:515.

MANTIQUILLA, J. A., SHIAO, M.-S., SHIH, H.-C., CHEN, W.-H. & CHIANG, Y.-C. 2021. A review on the genetic structure of ecologically and economically important mangrove species in the Indo-West Pacific. *Ecological Genetics and Genomics* 18:100078.

MCRAE, B. H. 2006. ISOLATION BY RESISTANCE. *Evolution* 60:1551–1561. Oxford Academic.

MOHER, D., LIBERATI, A., TETZLAFF, J., ALTMAN, D. G. & GROUP, T. P. 2009. Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement. *PLOS Medicine* 6:e1000097. Public Library of Science.

NEI, M. 1972. Genetic Distance between Populations. *The American Naturalist* 106:283–292. [University of Chicago Press, American Society of Naturalists].

NG, P. K. L. & SIVASOTHI, N. 2001. A Guide to Mangroves of Singapore. Raffles Museum of Biodiversity Research, National University of Singapore and Singapore Science Centre, Singapore.

NÚÑEZ-FARFÁN, J., DOMÍNGUEZ, * CÉSAR A., EGUIARTE, L. E., CORNEJO, A., QUIJANO, M., VARGAS, J. & DIRZO, R. 2002. Genetic divergence among Mexican populations of red mangrove

(Rhizophora mangle): geographic and historic effects. *Evolutionary Ecology Research* 4:1049–1064. Evolutionary Ecology, Ltd.

QUINTANA, D. S. 2015. From pre-registration to publication: a non-technical primer for conducting a meta-analysis to synthesize correlational data. *Frontiers in Psychology* 6.

R CORE TEAM. 2022. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria.

RABINOWITZ, D. 1978. Dispersal Properties of Mangrove Propagules. *Biotropica* 10:47–57. [Association for Tropical Biology and Conservation, Wiley].

REED, D. H. & FRANKHAM, R. 2003. Correlation between Fitness and Genetic Diversity. *Conservation Biology* 17:230–237.

RICHARDS, D. R. & FRIESS, D. A. 2016. Rates and drivers of mangrove deforestation in Southeast Asia, 2000–2012. *Proceedings of the National Academy of Sciences* 113:344–349. Proceedings of the National Academy of Sciences.

SALM, R. V., SALM, R. V., CLARK, J. R. & SIIRILA, E. 2000. Marine and Coastal Protected Areas: A Guide for Planners and Managers. IUCN. 406 pp.

SELKOE, K. A., D'ALOIA, C. C., CRANDALL, E. D., IACCHEI, M., LIGGINS, L., PURITZ, J. B., HEYDEN, S. VON DER & TOONEN, R. J. 2016. A decade of seascape genetics: contributions to basic and applied marine connectivity. *Marine Ecology Progress Series* 554:1–19.

SEXTON, J. P., HANGARTNER, S. B. & HOFFMANN, A. A. 2014. Genetic Isolation by Environment or Distance: Which Pattern of Gene Flow Is Most Common? *Evolution* 68:1–15.

SHANKS, A. L., GRANTHAM, B. A. & CARR, M. H. 2003. Propagule Dispersal Distance and the Size and Spacing of Marine Reserves. *Ecological Applications* 13:159–169.

STOCKEN, T. V. DER, RYCK, D. J. R. D., VANSCHOENWINKEL, B., DEBOELPAEP, E., BOUMA, T. J., DAHDOUH-GUEBAS, F. & KOEDAM, N. 2015. Impact of landscape structure on propagule dispersal in mangrove forests. *Marine Ecology Progress Series* 524:95–106.

TAYLOR, H. R., ROBERTSON, H., CARTER, A. L. & RAMSTAD, K. M. 2021. The conservation management implications of isolation by distance and high genetic diversity in Great Spotted Kiwi (Apteryx haastii). *Emu - Austral Ornithology* 121:10–22. Taylor & Francis.

TRIEST, L., VAN DER STOCKEN, T., ALLELA AKINYI, A., SIERENS, T., KAIRO, J. & KOEDAM, N. 2020. Channel network structure determines genetic connectivity of landward–seaward Avicennia marina populations in a tropical bay. *Ecology and Evolution* 10:12059–12075.

TRIEST, L., VAN DER STOCKEN, T., SIERENS, T., DEUS, E. K., MANGORA, M. M. & KOEDAM, N. 2021. Connectivity of Avicennia marina populations within a proposed marine transboundary conservation area between Kenya and Tanzania. *Biological Conservation* 256:109040.

TOMLINSON, P. B. 2016. The Botany of Mangroves (2nd edition). Cambridge University Press. 433 pp.

VALENTINE, J. C., PIGOTT, T. D. & ROTHSTEIN, H. R. 2010. How Many Studies Do You Need? A Primer on Statistical Power for Meta-Analysis. *Journal of Educational and Behavioral Statistics* 35:215–247. SAGE Publications. 2455 Teller Road, Thousand Oaks, CA 91320. Tel: 800-818-7243; Tel: 805-499-9774; Fax: 800-583-2665; e-mail: journals@sagepub.com; Web site: http://sagepub.com.

VAN DER STOCKEN, T., CARROLL, D., MENEMENLIS, D., SIMARD, M. & KOEDAM, N. 2019a. Global-scale dispersal and connectivity in mangroves. *Proceedings of the National Academy of Sciences* 116:915–922. Proceedings of the National Academy of Sciences.

VAN DER STOCKEN, T., VANSCHOENWINKEL, B., CARROLL, D., CAVANAUGH, K. C. & KOEDAM, N. 2022. Mangrove dispersal disrupted by projected changes in global seawater density. *Nature Climate Change* 12:685–691. Nature Publishing Group.

VAN DER STOCKEN, T., WEE, A. K. S., DE RYCK, D. J. R., VANSCHOENWINKEL, B., FRIESS, D. A., DAHDOUH-GUEBAS, F., SIMARD, M., KOEDAM, N. & WEBB, E. L. 2019b. A general framework for propagule dispersal in mangroves. *Biological Reviews* 94:1547–1575.

VANDERKLIFT, M. A., DOROPOULOS, C., GORMAN, D., LEAL, I., MINNE, A. J. P., STATTON, J., STEVEN, A. D. L. & WERNBERG, T. 2020. Using Propagules to Restore Coastal Marine Ecosystems. *Frontiers in Marine Science* 7. Frontiers.

VIECHTBAUER, W. 2010. Conducting Meta-Analyses in R with the metafor Package. *Journal of Statistical Software* 36:1–48.

WANG, I. J. & BRADBURD, G. S. 2014. Isolation by environment. *Molecular Ecology* 23:5649–5662.

WEE, A. K. S., TAKAYAMA, K., ASAKAWA, T., THOMPSON, B., ONRIZAL, SUNGKAEW, S., TUNG, N. X., NAZRE, M., SOE, K. K., TAN, H. T. W., WATANO, Y., BABA, S., KAJITA, T. & WEBB, E. L. 2014. Oceanic currents, not land masses, maintain the genetic structure of the mangrove Rhizophora mucronata Lam. (Rhizophoraceae) in Southeast Asia. *Journal of Biogeography* 41:954–964.

WILCOVE, D. S. 1986. Habitat fragmentation in the temperate zone. *Conservation biology: the science of scarcity and diversity*. Sinauer Associates.

WRIGHT, D., BISHOP, J. M., MATTHEE, C. A. & VON DER HEYDEN, S. 2015. Genetic isolation by distance reveals restricted dispersal across a range of life histories: implications for biodiversity conservation planning across highly variable marine environments. *Diversity and Distributions* 21:698–710.

WRIGHT, S. 1943. ISOLATION BY DISTANCE. *Genetics* 28:114–138.

WRIGHT, S. 1950. Genetical Structure of Populations. *Nature* 166:247–249. Nature Publishing Group.

WU, B., GENG, S.-L. & SHU, B. 2012. Genetic variation and the conservation of isolated populations of Derris trifoliata (Leguminosae), a mangrove-associated vine, in southern China. *Biochemical Systematics and Ecology* 40:118–125.

YAHYA, A. F., HYUN, J. O., LEE, J. H., KIM, Y. Y., LEE, K. M., HONG, K. N. & KIM, S.-C. 2014. Genetic variation and population genetic structure of Rhizophora apiculata (Rhizophoraceae) in the greater Sunda Islands, Indonesia using microsatellite markers. *Journal of Plant Research* 127:287–297.

YAN, Y.-B., DUKE, N. C. & SUN, M. 2016. Comparative Analysis of the Pattern of Population Genetic Diversity in Three Indo-West Pacific Rhizophora Mangrove Species. *Frontiers in Plant Science* 7.