Functional Trajectories during innate spinal cord repair

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Abstract

Adult zebrafish are capable of anatomical and functional recovery following severe spinal cord injury. Axon growth, glial bridging and adult neurogenesis are hallmarks of cellular regeneration during spinal cord repair. However, the correlation between these cellular regenerative processes and functional recovery remains to be elucidated. Whereas the majority of established functional regeneration metrics measure swim capacity, we hypothesize that gait quality is more directly related to neurological health. Here, we performed a longitudinal swim tracking study for sixty individual zebrafish spanning eight weeks of spinal cord regeneration. Multiple swim parameters as well as axonal and glial bridging were integrated. We established rostral compensation as a new gait quality metric that highly correlates with functional recovery. Tensor component analysis of longitudinal data supports a correspondence between functional recovery trajectories and neurological outcomes. Moreover, our studies predicted and validated that a subset of functional regeneration parameters measured 1 to 2 weeks post-injury is sufficient to predict the regenerative outcomes of individual animals at 8 weeks post-injury. Our findings established new functional regeneration parameters and generated a comprehensive correlative database between various functional and cellular regeneration outputs.

Introduction

As vertebrates with elevated regenerative potential, adult zebrafish are an excellent model for

innate spinal cord (SC) repair. After complete SC transection, pro-regenerative responses

involving the immune system, neurons, and glia contribute to functional regeneration (1).

Prompted by a host of injury-induced regeneration promoting factors, glial and axonal bridges

reconnect the lesioned tissue, neurons regenerate proximal to the lesion, and swim capacity is

restored, all within 6–8 weeks post-injury (wpi) (2). Due to the remarkable ability of adult zebrafish

to reverse paralysis, recovery of swim function is an ultimate metric and a central readout of SC

repair.

Functional measurements of swim ability may be classified into two broad categories: swim

capacity, which represents the amount or extent of swimming a fish can perform under specific

conditions; and gait quality, which reflects the quality of a swim bout and the healthiness of a fish

during active swimming. One of the earliest assays, in 1997, to evaluate swim function in zebrafish

was a binary, recovered or not recovered, quality scoring system assigned by an observer using

a startle assay (3). In this assay, fish that respond to a stimulus and swim mostly using their tail

instead of their head are labeled as recovered. Based on these and other criteria, swim quality

scoring systems on 1 to 5 and 1 to 6 scales have also been developed (4, 5). Two capacity

measurements were subsequently introduced: time to exhaustion in 1998 (6) and swim distance

in a static tank in 2004 (7). These offered several advantages: they are less subjective, provide

continuous scale measurements, and are scalable for longer assays using computational tools.

Consequently, swim distance or time to exhaustion have been widely applied to SC regeneration

studies between 1997 and 2020. Since 2020, the toolkit to quantify swim function has rapidly

expanded and diversified. Traditional and recently established measurements of swim capacity

include swim endurance as quantified by time to exhaustion against increasing water current

velocities (2, 6, 8, 9, 10), maximum speed either against increasing water current velocities or in

a static tank (11, 12, 13, 14), swim distance (7, 9), time active (9, 10), time swimming against flow

(9), burst frequency (9, 10), and average position of the fish against a water flow axis ("mean y")

(9, 10). Although new quality measurements have been developed for larval fish including body

angle (15), power spectral densities (16), and centerline posture kinematics (17), published quality

measurements for adult fish are limited to manual scoring of perceived swim ability (3, 4, 5), tail

beat frequency and amplitude (12). Moreover, manually assigned scores are subjective and do

not scale well for higher throughput. Thus, behavioral tests to quantify gait quality in adult

zebrafish are less developed, yet much needed for accurate assessment of musculoskeletal
function in general and of neural regeneration in particular.

Gait analysis offers a scalable approach to quantify swim quality. Prior applications of gait
analysis, such as dorsal centerline posture and tail beat kinematics, supported the model that
functional recovery emerges directly from structural recovery among ray-finned fishes (12, 17,
18). These studies revealed stepwise recovery of specific gait features in correlation with axon
regeneration from V2a interneurons (12, 19). These findings suggested that formerly paralyzed
zebrafish may regain the ability to swim in a preinjury-like manner after neural regeneration. Yet,
further analysis of swim gait metrics, including posture and behavior, is required to confirm this
hypothesis.

A first step in gait analysis is the annotation of elemental components of gait in swim videos:
posture and behavior. Automated pose and behavior annotation tools for model organisms are
increasingly accessible. While traditional computational methods for zebrafish pose annotation
include thresholding and skeletonization (20, 21) or template-fitting (22, 23), many newer cross-
species methods prioritize deep learning. Such tools include DeepLabCut (24), DeepPoseKit (25),
OptiFlex (26), and SLEAP (27). For zebrafish, quantifying the dorsal centerline is an appropriate
representation of body posture. Despite its limitations in measuring vertical position, body
deformity in the dorso-ventral axis, or pectoral fin motion, dorsal centerline posture and gait has
been applied to larval and adult locomotion analysis (12, 20, 21) and lateral tail beating is the
source of most swim propulsion (28). Additionally, certain injury-induced gait alterations manifest
in the centerline (12). Similar to recent pose annotation methods, automated behavior annotation
is often performed through machine learning either by clustering in a latent space (20, 21, 29),
such as the pre-packaged tool B-SOiD (30), or through deep learning, including DeepBhvTracking
(31) or DeepEthogram (32). Despite advances in pose and behavior annotation, methods to
compare episodes of the same zebrafish behavior, either between animals or between recovery
timepoints, are yet to be applied. Comparing multiple episodes of the same behavior is a critical
component of gait quality analysis. One viable option includes novelty detection, which classifies
behavior episodes as inliers or outliers relative to a control distribution. Another option is posture
parameter quantification, which has been used in larval zebrafish (17). Examples of posture
parameter quantification from human and mouse studies used behavior and gait features, such
as quantified response to stimulus and range or speed of motion, to measure functional recovery
(33, 34, 35, 36). As swim quality is proposed to be a more direct readout of SC function and

musculoskeletal wellness than swim capacity, recovery metrics based on posture and gait have
the potential to become standards for functional assessment.

Given the recent diversification of swim capacity and gait quality measurements, this study
compares various established and novel functional recovery metrics and describes their
correlations with anatomical regeneration. To this end, we tracked swim function prior to injury
and weekly between 1 and 8 wpi. Fish identities were tracked throughout the experiment, and
cellular regeneration parameters were measured at 8 wpi. Rostral compensation as established
as a new gait feature that highly correlates with functional recovery. For comprehensive analysis
of gait features, we also describe recovery trajectories using tensor decomposition of longitudinal
recovery data. By observing correlations between established swim capacity and new gait
analysis metrics with cellular regeneration features, we propose and validate a strategy for
predicting 8 wpi recovery outcomes using 1 and 2 wpi swim function parameters. This study
establishes a new gait quality measurement of neuromuscular health, provides a comprehensive
correlative analysis of various functional and cellular regeneration outputs, and will help reduce
measurement redundancy in future studies.

Results

A longitudinal study of spinal cord regeneration parameters.
We devised a longitudinal study that allowed us to track cellular and swim recovery metrics in
individual animals for 8 weeks of SC regeneration (Figure 1A). Sixty wild-type fish of the
Ekkwill/AB strains (30 males and 30 females) were housed separately for the duration of the
study. Swim behavior was recorded for each fish prior to injury and weekly between 1 and 8 wpi.
At each time point, fish were placed in an enclosed swim tunnel and a dorsally placed camera
was used to capture swim behavior at 70 frames/s for a total of 15 min at water current velocities
of 0 cm/s (5 min), 10 cm/s (5 min), then 20 cm/s (5 min). At 8 wpi and following swim behavior
recording, axon tracing experiments and glial bridging measurements were performed to assess
cellular regeneration metrics.

To begin to assess SC regeneration parameters in this study, we first calculated previously
established swim behavior measurements from the captured swim videos (Figure 1B-G).
Functional measurements indicated that zebrafish are still capable of recovery from SC
transection while isolated from other fish. Swim distance, centroid time active, time against flow
and mean y-position decreased with acute injury at 1 wpi relative to controls, then significantly

increased between 1 and 8 wpi to approach uninjured control levels (Figure 1B-E). Centroid burst
frequency for male fish decreased at 1 wpi but did not show significant improvement at 8 wpi
(Figure 1F). Similarly, the proportion of fish that swam in a healthy manner, according to
perceptually assessed swim quality, decreased after injury and increased through recovery
(Figure 1G). We noted that male fish exhibited this stereotypical recovery trend more strongly
than females in this experiment (Figure S1A-F). Overall, the female fish in this experiment
regenerated poorly (Figure S1F) and were less energetic both before and after injury, which is
not necessarily typical of zebrafish (14). For this reason, female fish data are presented in the
supplement.

Spinal cord injury disrupts healthy swim gait.
While established measurements of swim behavior captured the recovery of swim capacity after
SCI, our swim videos showed even more pronounced swim gait differences. Comparing one
period of oscillatory forward swim behavior (cruise) before injury and at 1 wpi, we found that
acutely injured fish swim using more motion in the rostral portion of their body (Figure 2A-B). We
thus postulated that a scalable quantification of gait healthiness would provide a markedly
improved functional regeneration parameter compared to established swim capacity metrics and
to manually assigned quality scores. To perform high-throughput gait analysis, we began by
annotating dorsal centerline posture (Figure 2C). After training DeepLabCut (24, 37) to annotate
keypoints on the centerline, we filtered and processed the poses, then trained a classifier to
annotate simplified behaviors including rest, turn, and cruise. For simplicity, we defined a cruise
as a forward swim with at least five measured extrema, or approximately three body oscillations.

Our first approach to probe swim gait was to examine posture variation within healthy and
injured animals without regard to behavior. To this end, we used principal components analysis
(PCA) to decompose all poses where the fish was not near a wall. PCA was performed for all fish
and all assays (Figure S2A), separately for males (Figure S2B) and females (Figure S2C), and at
control and recovery endpoints 1 and 8 wpi per sex group (Figure S2D-E). Exploring the principal
components of control poses, merely three components were required to explain as much as 98%
of the variance in both groups. Using poses from 1 wpi, four components were required to capture
98% of the variance. At the end of 8 weeks in recovery, as many as five components were
required to capture 98% of the variance. These results indicated that centerline posture increased
in complexity after injury and through recovery.

Next, using cruise episodes annotated by the machine learning classifier, we measured body curvature amplitudes and frequencies at each measured angle position. Curvature at each angle position was calculated using the corresponding keypoint position and two adjacent keypoint positions (Figure 2C). We measured center-to-peak (half of peak-to-peak) curvature amplitude at all angle positions during cruise oscillations. Cruise amplitudes along the centerline were plotted for control, 1 and 8 wpi (Figure 2D). At 1 wpi, we observed a significant change in angular amplitude at angle positions 1 to 5 compared to uninjured controls. Cruise amplitudes appeared closer to control values at 8 wpi. These findings indicate that acutely injured fish compensate for paralysis using the rostral portion of their body and that compensation is less pronounced after cellular regeneration. Hypothesizing that fish swim more naturally if they regenerated well, we separately plotted cruise amplitudes for fish that regenerated at least 20% of their glial tissue at the lesion core (Figure 2E). Well-regenerated fish appeared to swim using amplitudes closer to control values. These results were also observed for female fish, although fewer female fish, 5 females versus 16 males, recovered more than 20% of their glial tissue at the lesion (Figure S3B). We quantified maximal rostral compensation and noted that this feature of swim gait does recover toward control levels (Figure 2F). We also measured summed rostral amplitude (Figure S4D). Fish with more regenerated SC tissue recovered healthier gait on average, indicated by lower rostral compensation at 8 wpi (Figure 2G). Individual recoveries were more pronounced: 17 fish at 8 wpi exhibited rostral compensation lower than 3 standard deviations above the control mean, and these fish had high average cellular regeneration: 59% glial bridging, 57% proximal axon regrowth, and 34% distal axon regrowth. All fish with compensation score lower than 0.025 (2.49 standard deviations above the control mean) had at least 20% glial bridge. These results indicate that rostral compensation is most pronounced during earlier stages of SC regeneration and that stronger cellular regeneration is associated with healthier swim quality.

In addition to angular amplitude, we also measured cruise frequency, defined as the number of complete oscillations at each angle position per second. Regardless of injury, the rate of movement during cruises was nearly uniform along the fish's body (Figure S4A). Like amplitude, average body cruise frequency was also disrupted by injury and improved over time (Figure S4B-C). Although functional abilities appeared different between the sexes according to established functional metrics, there was no evidence that rostral compensation differs between males and females at 1 wpi (Student's t-test p-value = 0.6), suggesting that SC transection affects the gait of both sexes equally and independently of swim capacity differences.

Cruising zebrafish produce thrust by ejecting vortices of water behind them (28). Strouhal
number (St) is a unitless value related to vortex shedding mechanics and is defined as tail beat
frequency times peak-to-peak amplitude of the tail tip divided by speed of forward motion. Cross
species research has concluded that St between 0.2 and 0.4 are most physiologically efficient
(38). We calculated tail beat frequency for fish swimming in static water (0 cm/s flow velocity) and
did not observe a significant change after injury (Figure 2H). Using displacement amplitude of the
caudal fin, fin flick frequency, and swim speed during cruises, we calculated St for all cruises
during the first five minutes (0 cm/s flow velocity) of the swim assay. Average St was within or
near the efficiency limits throughout the experiment (Figure 2I, Figure S3F). These results suggest
SCI does not preclude efficient swim thrust, likely due to rostral compensation during periods of
active cruising.

Injury-induced lateral scoliosis is irreversible by 2 weeks post-injury.
Before exploring centerline-derived gait more comprehensively, we had to address injury-induced
scoliosis. Although all fish in the experiment appeared healthy before injury, some fish developed
extreme body curvature after injury. We quantified scoliosis as the summed curvature at eight
angle positions along the fish's dorsal centerline (Figure 2C) averaged over the entire swim.
Representative fish with their scoliosis scores are presented in Figure 3A. Our analysis indicated
that of the 30 male and 30 female fish, 4, 2, and 1 male and 4, 3, and 2 female fish presented
scoliosis scores >1.34, >2.77, and >3.28, respectively. Pairwise comparisons of scoliosis scores
between time points, using Spearman correlation, showed that relative scoliosis rankings
stabilized after 2 wpi (Figure 3B). Fish did not exhibit noticeable scoliosis at 1 wpi, nor did scoliosis
scores at 1 wpi correlate with the following weeks. Scores at pre-injury and 1 wpi weakly
correlated with subsequent time points since the fish were all straight bodied at the beginning of
the experiment and had not developed scoliosis by 1 wpi. The randomness of scoliosis rankings
at 1 wpi and stability of rankings after 2 wpi are presented in Figure 3C using trendlines for each
fish colored by the fish's rank at a specific point in recovery. As an alternative, unsupervised
posture-driven recovery measurement, we used a nearest-neighbors approach to quantify
postural abnormality for each assay. Taken as a ratio compared to representative posture
observed in preinjury assays, we called this measurement "posture novelty". Posture novelty was
closely associated with scoliosis scores. Pairwise rank correlations of scoliosis scores suggest
that injury-induced scoliosis is irreversible by 2 wpi, and these results altogether revealed that

scoliosis is an important factor in the variation of poses observed in the experiment at 8 wpi
(Figure S2D-E).

Gait recovery plateaus around 3 weeks post-injury.
Because our rostral compensation score is just one feature of complex locomotion, we next set
out to comprehensively analyze cruise gate recovery by embedding cruise episodes into a latent
space. To reduce the effects of body shape on gait, we omitted fish with a scoliosis score greater
than 0.35 at 8 wpi. For computational efficiency, we performed embedding on representative
cruise episodes, exemplars, sampled from each assay. To explore the data in posture space, we
decomposed poses using principal components analysis. Cruise traces in posture space
manifested as ellipses, similar to previously described traces for larval zebrafish (20, 21). We
noted that 1 wpi cruise traces are both distinct from preinjury cruises and visually similar for
different animals within the same time point (Figure 4A). Then, we embedded cruise traces using
UMAP for three-dimensional visualization. We plotted cruises from highly regenerative fish, those
that recovered at least 20% glial tissue at the lesion (Figure 4B), separately from cruises of fish
with relatively poor cellular regeneration (Figure 4C). To interpret this embedding, average UMAP
values at each time point for decomposed cruise episode exemplars were plotted for highly
(Figure 4D) and poorly (Figure 4F) regenerating fish. For both fish groups, average UMAP values
exhibited the most change by 3 wpi. Similarly, for both highly and poorly regenerating fish, rostral
compensation recovery slowed around 3 wpi (Figure 2F,G; Figure S4B,D). Kernel density
estimates were calculated for each time point in each UMAP dimension, separated by cellular
regeneration outcome (Figure 4E,G). According to the UMAP kernel density estimates, 8 wpi
cruises did not inhabit the same embedding space as preinjury cruises. Cruises for female fish
were embedded in the same space and the results were visually similar (Figure S5A-G). This
temporal cruise embedding suggested that gait recovers to the greatest extent by 3 wpi and that
injury-induced gait changes are incompletely resolved at 8 wpi.

Selected functional measurements correlate with cellular repair at 8 weeks post-injury.
We next explored relationships between the full array of functional and cellular regeneration
metrics at 8 wpi. We measured Spearman correlations between all 8 wpi measurements, pairwise,
then performed hierarchical clustering on the correlation matrix (Figure 5A). While not all
measurements correlated well with cellular regeneration metrics, some did, including

quantifications of capacity, behavior, and gait. Selected measurements of these classes were
plotted against glial bridging: swim distance, a swim capacity measurement (Figure 5B); burst
frequency quantified using active posture, a behavior measurement (Figure 5C); and rostral
compensation, a gait quality measurement (Figure 5D). Percent glial bridging correlated strongly
($r_s > 0.6$) with both proximal and distal axon regrowth (Figure 5E). Assays could only be included
in the heatmap if the fish exhibited every measurable attribute, and because two fish did not cruise
in the 8 wpi swim assay, their data was omitted from the heatmap in Figure 5A. There were also
three male and three female fish that did not survive to the end of the experiment. Hence
correlations differ slightly between Figure 5A and Figure 5B-E.

Comparing pairwise correlations at 8 wpi, swim capacity and behavior measurements
generally correlated strongly with one another and resided in a swim capacity associated cluster
that we labeled Cluster 2 (Figure 5A). Such measurements include distance, activity, pose burst
frequency, and time swimming against the flow. Mean y-position correlated moderately with time
swimming against the flow, but weakly with the other swim capacity measurements. Most, but not
all, capacity measurements were at least moderately ($r_s > 0.3$) correlated with glial bridging and
axon regrowth. At 8 wpi, tail beat frequency, which did not change significantly after injury or
through recovery (Figure 2H, Figure S3E), did not correlate well with any other functional,
structural, or cellular measurement. A separate cellular regeneration associated cluster, labeled
Cluster 1, contained all cellular regeneration measurements, rostral compensation, perceived
swim quality, scoliosis, and posture novelty. Unlike the swim capacity associated cluster, all
metrics in Cluster 1, were at least moderately, positively correlated with each other. Of all
functional metrics, rostral compensation correlated most strongly with perceived quality and
percent glial bridging, and with axon regrowth to the same extent as posture novelty. Together,
this analysis showed that not all the swim capacity metrics we measured were representative of
the SC's condition at 8 wpi, and that rostral compensation is a neurologically associated
measurement of gait quality.

As previously described, female fish in this experiment regenerated more poorly than male
fish. Nonetheless, a similar clustering of 8 wpi measurements from female fish yielded the same
clusters (Figure S6A). Spearman rank correlation of swim distance, pose burst frequency, rostral
compensation, and axon regrowth with glial bridging did not reach statistical significance for
female fish (Figure S6B-E). Despite their non-significant p-values, rostral compensation and
proximal axon regrowth showed the highest correlation values with glial bridging (Figure S6A).

These results support our model that rostral compensation is the most neurologically relevant of
all metrics in this tracking study.

Tensor decomposition of all assay measurements corresponds with outcomes at 8 wpi.
We applied tensor component analysis (TCA) to explore recovery trajectories. To prepare the
data, we concatenated measurements from all 9 assays of all 60 fish into a rank-3 tensor. The
software that we used for TCA did not allow missing values, and we decided not to use
interpolation, so 16 fish (6 male, 10 female) had to be omitted. We selected parameters for the
model by optimizing for low model error and high (>0.8) model similarity (Figure 6A). After
selecting parameters for the model, we obtained 7 tensor components including factors for each
axis: fish, assay, and functional measurement (Figure 6D). To visualize factors, we sorted fish by
the size of their glial bridge at 8 wpi and sorted assays temporally. By manual inspection, values
for many fish factors showed a directional trend, implying that the trajectories captured in the
factor values represented a functional output relevant to neurological recovery. We explored this
relationship further by clustering fish according to their factor values. We selected a distance
threshold parameter that yielded the most clusters given a high minimum allowed cluster size
(Figure 6B). The five resulting clusters of fish were roughly the same size, including between 7
and 11 fish in each cluster. Cluster averages for measurements taken at 8 wpi were plotted in a
heatmap, with clusters sorted by average glial bridging (Figure 6C). When ordered by average
glial bridging in the cluster, clusters also sorted according to average scoliosis, posture novelty,
perceived quality, and rostral compensation. In the same ordering but omitting cluster 1, clusters
2 to 5 also sorted according to average measurements of axon regrowth (proximal and distal),
swim distance, activity, time against flow, posture burst frequency, and mean y (but in the opposite
direction than expected). The only measurement averages that were not sorted in this ordering
were centroid burst frequency and tail beat frequency. Fish factor values in Figure 6D were
colored by the fish's cluster according to the cluster's average glial bridging. We expected that
assay factors would exhibit a displacement-to-recovery trend from 1 to 8 wpi, mimicking the
recovery trend of most functional metrics (Figure 1B-G, Figure 2F). Although some assay factors
did capture this trend, many diverged away from control through recovery, and the values of TC1
and TC2 did not change dramatically from control until 2 wpi, capturing a trend more similar to
the scoliosis score (Figure 3C). We interpret the diversity of assay factor trendlines to represent
complexities in the differing biological signals of the functional measurements and longitudinal
support for non-uniformity of 8 wpi metric correlation signals shown in Figure 5A. Overall, tensor

component analysis and fish factor clustering indicate that recovery trajectories correspond
strongly with 8 wpi cellular regeneration measurements.

Swim function at 2 weeks post-injury is predictive of cellular regeneration at 8 weeks.
Swim distance and rostral compensation were two functional measurements that most correlated
with SC regeneration at 8 wpi. Swim distance and rostral compensation measure different
functional classes: swim capacity and gait quality, respectively. They also clustered separately in
the pairwise metric analysis and recovered at different rates. Thus, as distinct functional outputs
that correlate well with cellular regeneration, we hypothesized that a combination of swim distance
and rostral compensation assessed during early stages of SC regeneration could predict injury
outcomes at 8 wpi. To test this hypothesis, we averaged 1 and 2 wpi distance and rostral
compensation measurements and plotted them against one another colored by 8 wpi glial bridging
(Figure 7A). As expected from the correlation analysis, fish with high glial bridging generally swam
longer distances and with lower rostral compensation. We ranked fish according to swim distance
and negative rostral compensation, then added the two ranks to create a wellness score for early
recovery. We divided male and female data separately into two equally sized groups across each
sex's wellness score median: a highly regenerative prediction group and a poorly regenerative
one. Both percent glial bridging and proximal axon regrowth were significantly higher in the highly
regenerative group compared to the poorly regenerative group (Figure 7B). Since this analysis
was performed after our tracking experiment was completed and final outcomes were measured,
a new experiment was performed to validate the prediction method.

In the prediction experiment, we predicted regeneration outcomes for sixty (30 male, 30
female) fish using the same wellness score described above (Figure 7C). Swim behavior was
assayed before injury in addition to 1, 2, and 8 wpi. Fish were housed in male-female pairs to
track their identities until swim assays were performed at 2 wpi. Predictions were made separately
for males and females, then fish of both sexes were mixed randomly between tanks within the
prediction groups, 5–7 fish per tank. Predictions were made using the median wellness scores in
the prediction experiment to divide the groups (Figure 7D). At 8 wpi, glial bridging was significantly
elevated in the highly regenerative prediction group compared to the poorly regenerative
prediction group (Figure 7E). The results of this prediction experiment indicated that it is possible
to predict 8 wpi cellular regeneration outcomes from a subset of 2 wpi measurements.

We clustered the 8 wpi functional and regeneration measurements in the same way as we
did for the tracking experiment. In doing so, we discovered that endurance, a measure of swim
capacity that was not evaluated at any other timepoint or in the tracking experiment, was a
member of the neural health-associated cluster, Cluster 1, not the swim capacity cluster (Figure
S7A). Unlike the tracking experiment, swim distance and pose burst frequency were not
associated with bridging to the same extent (Figure S7B-C), and axon regeneration was
measured lower than expected for all fish (Figure S7E). Rostral compensation, however, still
correlated more strongly with glial bridging than any other measurement (Figure S7D). These
findings provided additional evidence that rostral compensation is a neurologically associated gait
quality measurement, and suggested that results derived from gait quality replicate more reliably
than those derived from swim capacity.

Discussion

This study explores connections between functional and cellular regeneration measurements
during SC regeneration in adult zebrafish. Fish were individually housed to enable their
longitudinal tracking for 9 weeks. In comparison with alternative methods of tracking fish identities
such as fin clipping, dye injection, passive integrated transponder tagging or skin pattern
recognition, physical separation was the method of choice to avoid introducing additional injuries
or imaging steps to our experimental pipeline (39). It is important to note that, compared to the
well-known elevated regenerative capacity of zebrafish SC tissues, regeneration efficiency was
reduced in this cohort of 60 fish. Although it remains to be determined whether physical separation
negatively impacts SC regeneration, we propose that our previously established injury and post-
operative standards, including group housing, support optimal regeneration conditions (10, 40).
Even in such conditions where functional recovery is expected to continue to 8 wpi, some fish
experience negligible functional improvement after 6 wpi (3). However, our purpose was not to
demonstrate SC regenerative capacity in zebrafish, but to compare traditional and new functional
measurements and their associations with structural recovery. Our fish exhibited significant
recovery up to 6 wpi, glial and axon bridging were highly correlated, and all three measurements
of cellular regeneration showed similar correlation patterns with examined functional and
structural metrics. Although functional recovery appeared different between the sexes, we saw
no evidence that the cruise gait of males and females are affected differently by SC transection,
comparing male versus female rostral compensation scores at 1 wpi using Student's t-test, (p-

value = 0.6). Thus, irrespective of whether housing conditions for this study were detrimental to
recovery, the broad range of regeneration observed in this study was not only well suited, but also
retrospectively ideally tailored, for our purposes. Our tracking experiment allowed us to perform
comprehensive correlative analyses of recovery metrics over a wide range of regeneration
parameters.

Our study introduces rostral compensation score as a scalable quantification for gait quality.
In both the tracking and prediction experiments, rostral compensation correlated more highly with
glial bridging than any other measurement at 8 wpi. The strong correlations we observed between
rostral compensation and cellular regeneration measurements support a model in which rostral
compensation score reflects the compensatory effort paralyzed fish make to produce sufficient
thrust to move. Whether fish optimize their Strouhal number by learning to adjust the appropriate
amount of rostral compensation or whether the optimal Strouhal number is physiologically
enforced by the natural elasticity of a fish's body remains to be explored (41). The calculation of
rostral compensation was entirely automated after training a classifier to annotate cruise
behaviors from centerline posture. As a recovery metric in the tracking experiment, average
rostral compensation scores showed 50% recovery at 8 wpi. Notable and significant changes in
rostral compensation occurred by 3 wpi, suggesting a threshold of cellular regeneration is required
for healthy functional outcomes and that cellular regeneration beyond 3 wpi has a diminished
effect on functional recovery. Overall, our findings show that rostral compensation is a neural
health-associated measurement that recovers earlier than swim capacity.

To achieve a more comprehensive analysis of cruise gait recovery beyond single gait
features such as rostral compensation, cruise episodes were embedded with UMAP using
dynamic time warping distances. Surprisingly, even after selecting for fish that showed elevated
regeneration metrics, 8 wpi cruises showed a clearly distinct distribution compared to control
cruises prior to injury. These findings suggest that some injury-induced gait feature persists for
many fish after apparently successful recovery. When an abnormal swim gait develops during
recovery, it could be due to mistaken axon connections or suboptimal tissue remodeling. Specific
SC neurons and regions are known to be associated with aspects of zebrafish locomotion (42,
43). After SC transection, as axons reconnect, fast and slow V2a interneurons regrow axons along
the rostro-caudal axis, connecting to neurons of the same type and enabling specific features of
functional recovery (12). In addition to axon regrowth, it is estimated that as much as a third of
SC tissue shows evidence of cell division during SC regeneration (44). Moreover, recent
mammalian studies indicate that certain neurons required to restore walking after SC injury are

not the same as the neurons used for the same behavior before injury (45). Together, these
various modes of cellular regeneration may explain why, given the natural variation in recovery
outcomes, the kernel density estimates of UMAP-embedded cruises did not completely overlap
with those of uninjured control cruises. We propose that differential swim gait development during
the recovery process could be due to misguided axon regrowth or suboptimal tissue remodeling,
and that further cellular and behavioral regeneration studies are needed to better understand the
cellular basis of swim recovery in zebrafish.

We performed a separate experiment to explore the hypothesis that rostral compensation
and swim distance measurements at 1 and 2 wpi can be used to predict 8 wpi cellular regeneration
outcomes. The prediction experiment, while not completely successful at predicting significantly
different outcomes for both glial bridging and axon regrowth, nonetheless provides a proof of
concept for future work. In both the tracking and prediction experiments, glial regeneration
outcomes were significantly different, which indicates that early functional measurements can be
predictive of cellular regeneration measured six weeks later. Moreover, our prediction experiment
provides validation for rostral compensation as a neurologically meaningful functional metric, and
evidence that swim endurance, which was not evaluated in our tracking experiment, is also
associated with neurological health. Importantly, both the tracking and prediction experiments
support our conclusion that rostral compensation provides the strongest correlation with cellular
regeneration than any other functional measurement that we examined. Comparing the two
experiments, we likely overestimated swim distance's predictive power, and knowing that gait
recovers relatively early, rostral compensation alone may be the best metric for outcome
prediction, which remains to be tested.

Conclusion

Our study supports standard functional analysis, demonstrating that recovered swim capacity
measurements correlate with spinal cord regeneration. We introduce rostral compensation as a
gait-driven measurement that correlates more strongly with and likely provides a more accurate
functional proxy for spinal cord regeneration than existing functional regeneration metrics.
Through posture analysis, we discovered that the fish that would develop injury-induced lateral
scoliosis had a set trajectory by two weeks following SC transection. We found that, in still water,
fish swam in a mechanically efficient manner both before and after injury. Finally, we discovered
that grouping fish according to their rank in functional attributes two weeks into recovery can
divide the fish into groups that have statistically divergent cellular regeneration outcomes at eight

weeks post-injury. We propose that developing a gait-driven prediction algorithm could offer powerful benefits to the zebrafish research community such as non-invasively tracking musculoskeletal wellness or reducing the time required to complete SC regeneration experiments.

Acknowledgments

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Figure 1

Figure 1. Longitudinal tracking of swim recovery after spinal cord injury in adult zebrafish.
(A) Schematic representing an overview of the tracking experiment. A total of 60 fish (30 males and 30 females) were housed separately throughout the entire experiment to allow for longitudinal tracking of individual fish throughout the study. Swim behavior was recorded prior to injury, and weekly between 1 and 8 wpi. At each time point, fish were video recorded in a swim tunnel for fifteen minutes. In each 15-min assay, fish were first acclimated in the absence of water current for 5 minutes (0 cm/s), then water current velocity was increased to 10 cm/s at 5 min and to 20 cm/s at 10 min. **(B-G)** A suite of established functional measurements were used to assess swim recovery for male fish following SC transection. The swim recovery outcomes for female fish are shown in Figure S1. Swim distance (B), activity as measured by centroid movement (C), time swimming against the flow (D), mean y-position in the axis of flow (E), and burst frequency as measured by centroid movement (F) represent swim capacity measurements. Perceived swim quality scores are shown in G. Error bars depict SEM and statistical significance was determined by one-way repeated mixed measures ANOVA followed by post-hoc Student's t-tests. p-value markers in black represent comparisons between each time point post-injury relative to control measurements prior to injury. Red horizontal bars and p-values marked in red show significance between 1 wpi and 8 wpi. ***$P<0.001$; **$P<0.01$; *$P<0.05$; ns, $P>0.05$.

Figure 2

Snapshots of Cruise Periods

A Control frames

overlap

B 1 wpi frames

overlap

C Angle Positions

α_1 α_2 α_3 α_4 α_5 α_6 α_7 α_8

0 1 2 3 4 5 6 7 8 9

Keypoint Positions

Caudal Fin Tip

Angular amplitude

Rostral compensation

Tail beat frequency and Strouhal

D All Male Fish

angular amplitude radians

control n=30
1 wpi n=30
8 wpi n=25

rostral   caudal

α_1 α_2 α_3 α_4 α_5 α_6 α_7 α_8
centerline angle position

E Glial Bridge >20%

angular amplitude radians

control n=16
1 wpi n=16
8 wpi n=15

rostral   caudal

α_1 α_2 α_3 α_4 α_5 α_6 α_7 α_8
centerline angle position

F All Male Fish

rostral compensation Δradians

***   ***
***
***   ***   ***   ***   ***   ***

control 1 2 3 4 5 6 7 8
wpi

G Glial Bridge >20%

rostral compensation Δradians

**   **
**
*   **   *   *   *

control 1 2 3 4 5 6 7 8
wpi

H All Male Fish

tail beat frequency 0 cm/s flow, Hz

control 1 2 3 4 5 6 7 8
wpi

I All Male Fish

Strouhal 0 cm/s flow

control 1 2 3 4 5 6 7 8
wpi

Figure 2. Cruise quality quantifications during SC regeneration. (A, B) Cruise periods at a water current velocity of 10 cm/s. Snapshots of swim behavior prior to injury are shown in A and at 1 wpi in B. Overlayed swim forms shown below the separate frames highlight defective caudal movement after SC transection **(C)** Experimental schematic to develop quantifiable gait quality measurements. A diagram showing keypoint and angle positions labeled on a fish's centerline. A DeepLabCut model was trained to identify the keypoints, then the curvature between three adjacent keypoints were calculated at each angle position. The rostral region of the fish corresponds to angle positions α_1 to α_5. **(D)** Cruise curvature profiles for male fish before injury, acutely injured fish at 1 wpi, and recovered fish at 8 wpi. Each cruise curvature profile represents the mean lateral angular amplitude along the dorsal centerline while cruising. The vertical dotted line separates rostral and caudal positions, demonstrating that acutely injured fish swim with markedly elevated curvature in the rostral portion of their body. **(E)** Cruise curvature profiles (control, 1wpi, and 8wpi) for male fish that regenerated more than 20% of the glial tissue at the lesion site. **(F)** Quantification of rostral compensation in male control fish prior to SC injury and at 1 to 8 wpi. Rostral compensation represents the displacement between cruise profiles. This score is the maximum distance, on the vertical axis, between rostral positions of a cruise profile from the control profile. Angle positions α_1 to α_5 from Panel D were used to define the rostral region. **(G)** Rostral compensation scores for male fish that regenerated more than 20% of the glial tissue at the lesion site. **(H)** Tail beat frequency for male fish in still water (0 cm/s), measured at each assayed week. ANOVA was not significant for tail beat frequencies. **(I)** Strouhal numbers for male fish in still water (0 cm/s), measured at each assayed week. Strouhal number is a unitless value related to vortex shedding mechanics and is defined as tail beat frequency times peak-to-peak amplitude of the tail tip divided by speed of forward motion. ANOVA was not significant for Strouhal numbers. Error bars depict SEM and statistical significance was determined by one-way repeated mixed measures ANOVA followed by post-hoc Student's T-tests. p-value markers in black represent comparisons between each time point post-injury relative to control measurements prior to injury. Red horizontal bars and p-values marked in red show significance between 1 wpi and 8 wpi. ***$P<0.001$; **$P<0.01$; *$P<0.05$; ns, $P>0.05$.

Figure 3

A

Representative Scoliosis Scores

B

Week-to-Week Pairwise Spearman Correlations

C

Scoliosis Trend Lines

Figure 3. Assessment of lateral scoliosis among all fish, male and female, in the tracking experiment. (A) Visualization of the scoliosis score. The scoliosis score is the sum of curvature measured at angle positions on the centerline at rest (see methods). To demonstrate how this score relates to body curvature, frames were taken from representative assays identified using k-medoids clustering of scoliosis scores of all assays. **(B)** Pairwise comparison of scoliosis scores between weeks by Spearman's rank correlation. Scoliosis scores correlate weakly between control and 1 wpi to other assays. Scores are most strongly correlated from 2 wpi onward. **(C)** A demonstration that scoliosis ranks stabilize after 2 wpi. Scoliosis trend lines for individual fish are plotted over all assays (x-axis), colored by rank at 1, 2, 3, and 8 wpi (top-left to bottom-right). Fish scoliosis trend lines are colored on a rainbow scale from red (most severe) to violet (least), and colored gray if the fish did not survive to the ranked week.

Figure 4

Non-scoliotic Male Cruise Embedding

Figure 4. UMAP embedding of cruise behaviors during SC regeneration. (A) Cruise poses from control (black) and 1 wpi (red) male zebrafish were decomposed using principal component analysis (PCA). Male fish with scoliosis score < 0.35 at 8 wpi are shown. In this posture space, cruise trajectories from acutely injured fish at 1 wpi are markedly distinct from control fish trajectories. **(B-C)** Cruise poses were embedded using UMAP with precomputed dynamic time warping (DTW) distances. Cruise poses were analyzed for fish that regenerated well (glial bridging > 20%) in B and for fish with compromised cellular regeneration (glial bridging < 20%) in C. **(D)** Gait recovery trendlines in UMAP 1, 2, and 3 (left to right) for fish that regenerated well. Colored bars indicate the mean trend of each UMAP component. Error bars are standard deviations. The vertical dotted black lines align with the average UMAP value at 3 wpi to indicate that most change in each UMAP occurs before 3 wpi. **(E)** Gaussian kernel density estimates for the regions of UMAP space occupied by cruises at each week post-injury for fish that regenerated well. The x-axes are matched between Panels D and E for comparison. **(F)** Gait recovery trendlines in UMAP 1, 2, and 3 (left to right) for fish with compromised cellular regeneration. Colored bars indicate the mean trend of each UMAP component. Error bars are standard deviations. The vertical dotted black lines align with the average UMAP value at 3 wpi to indicate that most change in each UMAP occurs before 3 wpi. **(G)** Gaussian kernel density estimates for the regions of UMAP space occupied by cruises at each week post-injury for fish with compromised cellular regeneration. The x-axes are matched between Panels F and G for comparison.

Figure 5

A

**Pairwise Comparisons of
8 wpi Measurements
25 Male Fish**

Spearman r_s

B

Swim Capacity
vs
Glial Bridging

n = 27
$r_s = 0.61$

C

Behavior
vs
Glial Bridging

n = 27
$r_s = 0.47$

D

Gait
vs
Glial Bridging

n = 25
$r_s = -0.81$

E

Glial Bridging
vs
Axon Regrowth

n = 27
Proximal $r_s = 0.65$
Distal $r_s = 0.69$

Figure 5. Pairwise comparisons of spinal cord regeneration metrics at 8 wpi. (A) Heatmap of Spearman's rank correlations (r_s) between the cellular and functional measurements taken. Regeneration metrics include swim capacity, swim quality, structural, and neurological measurements. A dendrogram representing similarities between correlation patterns of measurements is shown. Male fish at 8 wpi were analyzed. Fish that could not be measured in all attributes were omitted. Correlations are reported regardless of p-value. For the sake of meaningful clustering, a measured attribute was multiplied by -1 if its average was increased at 1 wpi compared to controls. These attributes are marked with the prefix "(-)" on the label. **(B-D)** One metric each of swim capacity (swim distance), behavior (burst frequency), and gait (rostral compensation) were plotted against glial bridging to demonstrate their correlation. Because the Spearman correlation operates on ranked values, we fit monotonic splines to plots where r_s were significant ($p < 0.05$) to visualize possible associations. **(E)** Scatter plot showing a strong association between the size of regenerated glial tissue and axon regrowth, proximal and distal.

Figure 6

TCA of the Tracking Experiment (44 fish)

A TCA Rank Selection

D TCA Factors

Fish (44)   Assays (9)   Functional Measurements (11)

TC1, TC2, TC3, TC4, TC5, TC6, TC7

glial bridging   low   high   control 1 2 3 4 5 6 7 8   wpi

Cluster Colors
c1 c4 c3 c5 c2

B

Measurements Order (left to right)
Swim Distance
Activity
Time Against Flow
Lateral Scoliosis
Posture Novelty
Centroid Burst Frequency
Pose Burst Frequency
Mean Y
Quality
Rostral Compensation
Tail Beat Frequency

C 8 wpi Cluster Averages

	c1	c4	c3	c5	c2
Glial Bridging	6	9	28	36	58
Axon Regrowth (Proximal)	32	19	34	43	73
Axon Regrowth (Distal)	15	6	22	24	47
Swim Distance (x10^5)	1.7	1.5	2.0	2.4	3.7
Activity	49	43	48	61	78
Time Against Flow 10, 20 cm/s	60	59	62	66	70
Lateral Scoliosis	0.8	0.3	0.4	0.2	0.1
Posture Novelty	82	28	27	19	9
Centroid Burst Frequency 0 cm/s	60.46	15.62	6.60	20.51	24.64
Pose Burst Frequency 0 cm/s	39.49	11.51	12.62	43.89	98.42
Mean Y (x10^2) 10, 20 cm/s	9.6	10.2	9.9	9.7	9.0
Quality	3.29	4.09	4.40	4.86	5.00
Rostral Compensation	0.08	0.05	0.04	0.03	0.02
Tail Beat Frequency	12.95	14.06	13.70	13.02	13.83
cluster	c1	c4	c3	c5	c2
N	7	11	10	7	9

Figure 6. Tensor component analysis of swim behavior during SC regeneration. (A) Model selection parameters. Plots on the left show model error various ranks (x-axis). Plots on the right show model similarity for various ranks (x-axis). Unlike PCA, TCA is not deterministic. Thus, multiple models were trained and compared. Three model types are shown. To achieve an optimal balance between low model error and high model similarity, we chose a rank-7 canonical polyadic decomposition by alternating least squares (cp_als). **(B)** Selecting an appropriate distance threshold for clustering. We clustered fish according to their TCA factors using agglomerative clustering. To maximize the number of clusters while keeping a high minimum cluster size, we chose 1.7 as our distance threshold with five clusters identified. **(C)** Recovery outcomes for the five identified fish clusters, sorted by average glial bridging, low to high, from left to right. Each row's color is normalized such that the darkest blue corresponds to the maximum observation among all fish for that measurement. **(D)** Factors from a rank 7 tensor decomposition corresponding to each axis of the full data tensor: fish, assays (wpi), and functional measurements. Fish are sorted on the x-axis by their 8 wpi measured percent glial bridging and colored according to the rank of the cluster with respect to average glial bridging. Assays are sorted temporally. Measurements are listed below the plots in descending order. For panels A-D, 44 out of 60 total fish were analyzed. 16 fish were omitted either due to early death (6 fish) or because cruise waveform statistics were insufficient for at least one assay (10 fish).

Figure 7

Tracking Experiment

A Ranked Metrics

summed rank ○ low ◇ high   % Bridging 0% ▬ 100%

Female   Male

avg rostral comp (Δradians)

avg swim distance (x10⁵ px)

B Cellular Regeneration Outcomes

prediction ▬ low ▬ high

Glial Bridging   Axon Proximal   Axon Distal

regeneration (%)

*** * n.s.

low high   low high   low high

groups chosen by ranked 1 & 2 wpi averages
27 females, 27 males

Prediction Experiment

C Swim Assays

Baseline 1 2   8 wpi

SCI   Predict & sort   Histological validation of predicted groups

▬ housed in pairs male and female
▬ housed in groups by prediction

D Ranked Metrics

summed rank ● low ◆ high

Female   Male

avg rostral comp (Δradians)

avg swim sistance (x10⁵ px)

E Cellular Regeneration Outcomes

prediction ▬ low ▬ high

Glial Bridging   Axon Proximal   Axon Distal

regeneration (%)

** n.s. n.s.

low high   low high   low high

groups chosen by ranked 1 & 2 wpi averages
27 females, 29 males

Figure 7. Predicting 8 wpi regeneration outcomes using measurements taken by 2 wpi. (A) Tracking experiment: swim distance versus rostral compensation was averaged between 1 and 2 wpi. Dots represent individual fish and were colored based on the extent of glial bridging measured at 8 wpi for each fish. Fish that would develop larger glial bridges tended to swim further and exhibit less rostral compensation, in lower right side of each graph. Each fish was assigned a prediction score by adding its rank according to average swim distance to its rank according to average negative rostral compensation, relative to its sex group. **(B)** Tracking experiment: 8 wpi glial bridging and axon regrowth outcomes for groups ranked high in swim distance and low in rostral compensation between 1 and 2 wpi ("high") compared to those ranked low in swim distance and high in rostral compensation between 1 and 2 wpi ("low"). **(C)** Schematic overview of the prediction experiment. A total of 60 adult fish (30 males and 30 females) were housed in male-female pairs to track their identities until low/high regeneration predictions were assigned, then housed in prediction groups. At 8 wpi individuals were separated again to correlate final measurements from the endurance and behavior assays with cellular regeneration. Behavior was assayed before injury, 1 wpi, 2 wpi, and 8 wpi. Predictions were made, as described, after the 2 wpi behavior assay. **(D)** Prediction experiment: swim distance versus rostral compensation was averaged between 1 and 2 wpi. Dots represent individual fish and were colored based on the predicted outcome. **(E)** Prediction experiment: 8 wpi glial bridging and axon regrowth outcomes for each prediction group. Statistical significance in panels B and E was determined by Student's t-test. p-values represent comparisons between prediction groups. ***$P<0.001$; **$P<0.01$; *$P<0.05$; ns, $P>0.05$.

Supplementary Figures

Figure S1

Figure S1. Established functional measurements for female fish in the tracking experiment. **(A-F)** A suite of established functional measurements were used to assess swim recovery for female fish following SCI. Swim distance (A), activity as measured by centroid movement (B), time swimming against the flow (C), mean y-position in the axis of flow (D), and burst frequency as measured by centroid movement (E) represent swim capacity measurements. Perceived swim quality scores are shown in F. Error bars depict SEM and statistical significance was determined by one-way repeated mixed measures ANOVA followed by post-hoc Student's t-tests. p-value markers in black represent comparisons between each time point post-injury relative to control measurements prior to injury. Red horizontal bars and p-values marked in red show significance between 1 wpi and 8 wpi. ***$P<0.001$; **$P<0.01$; *$P<0.05$; ns, $P>0.05$.

Figure S2

A

All Fish All Assays
98% variance in 4 pcs

B

Males All Assays
98% variance in 4 pcs

C

Females All Assays
98% variance in 5 pcs

D

Males Control
98% variance in 3 pcs

Males 1 wpi
98% variance in 4 pcs

Males 8 WPI
98% variance in 4 pcs

E

Females Control
98% variance in 3 pcs

Females 1 wpi
98% variance in 4 pcs

Females 8 wpi
98% variance in 5 pcs

Figure S2. Principal component analysis of angle poses observed in the tracking experiment. On the left side of each PCA is a scree plot showing the percentage of variance captured by each PC. On the right side of each PCA are the first five components plotted as "eigenfish" by adding multiples of each PC to the mean pose. **(A)** A general representation of the posture space, and its complexity, for both healthy and injured fish. PCA of poses from all fish, all assays. **(B)** PCA of poses from all assays, male fish only. **(C)** PCA of poses from all assays, female fish only. **(D)** Posture space for male fish using poses taken from control, 1 wpi, and 8 wpi assays (top to bottom). **(E)** Posture space for female fish using poses taken from control, 1 wpi, and 8 wpi assays (top to bottom). Posture space is similarly complex whether separated by sex or taken altogether.

Figure S3

Figure S3. Cruise gait quality quantifications for female fish in the tracking experiment. (A) Cruise curvature profiles for female fish before injury, acutely injured fish at 1 wpi, and recovered fish at 8 wpi. Each cruise curvature profile represents the mean lateral angular amplitude along the dorsal centerline while cruising. The vertical dotted line separates rostral and caudal positions, demonstrating that acutely injured fish swim with markedly elevated curvature in the rostral portion of their body. **(B)** Cruise curvature profiles (control, 1wpi, and 8wpi) for female fish that regenerated more than 20% of the glial tissue at the lesion site. **(C)** Quantification of rostral compensation in female control fish prior to SC injury and at 1 to 8 wpi. Rostral compensation represents the displacement between cruise profiles. This score is the maximum distance, on the vertical axis, between rostral positions of a cruise profile from the control profile. Angle positions α_1 to α_5 from Panel D were used to define the rostral region. **(D)** Rostral compensation scores for female fish that regenerated more than 20% of the glial tissue at the lesion site. **(E)** Tail beat frequency for female fish in still water (0 cm/s), measured at each assayed week. ANOVA was not significant for tail beat frequencies. **(F)** Strouhal numbers for female fish in still water (0 cm/s), measured at each assayed week. Strouhal number is a unitless value related to vortex shedding mechanics and is defined as tail beat frequency times peak-to-peak amplitude of the tail tip divided by speed of forward motion. ANOVA was not significant for Strouhal numbers. Error bars depict SEM and statistical significance was determined by one-way repeated mixed measures ANOVA followed by post-hoc Student's t-tests. p-value markers in black represent comparisons between each time point post-injury relative to control measurements prior to injury. Red horizontal bars and p-values marked in red show significance between 1 wpi and 8 wpi. ***$P<0.001$; **$P<0.01$; *$P<0.05$; ns, $P>0.05$.

Figure S4

A

Mean Angular Frequencies

B Avg Body Frequency   **C** Body Frequency Displacement

D Avg Rostral Summed Amplitude

Figure S4. Cruise waveform features for male and female fish in the tracking experiment. We measured center-to-peak (half of peak-to-peak) curvature amplitude at all angle positions during cruise oscillations. We also measured cruise frequency as the number of complete oscillations at each angle position per second. The body of the fish corresponds to angle positions α_2 to α_8. The rostral region of the fish corresponds to angle positions α_1 to α_5. **(A)** Mean angle frequencies at each angle position on the centerline (x-axis) measured during cruise episodes, plotted for control, 1 wpi, and 8 wpi. Frequency was stable along the body of the centerline regardless of injury. **(B)** Average angular frequency along body positions for each assay and separated by sex. **(C)** Average body frequency displacement at each assay and separated by sex. Body frequency displacement is calculated similarly to rostral compensation: the maximum distance between a frequency curve and the control frequency curve at body positions. **(D)** The average sum of angular amplitudes measured at rostral positions along the fish, plotted for each assay and separated by sex. Error bars depict SEM and statistical significance was determined by one-way repeated mixed measures ANOVA followed by post-hoc Student's t-tests. p-value markers in black represent comparisons between each time point post-injury relative to control measurements prior to injury. Red horizontal bars and p-values marked in red show significance between 1 wpi and 8 wpi. ***P<0.001; **P<0.01; *P<0.05; ns, P>0.05.

Figure S5

Non-scoliotic Female Cruise Embedding

A Cruise Paths
Posture PCA
15 fish

B High Bridging (> 20%)
UMAP
4 fish

C Low Bridging (≤ 20%)
UMAP
9 fish

D High Bridging (> 20%)

E

F Low Bridging (≤ 20%)

G

Figure S5. UMAP embedding of cruise behaviors during SC regeneration. (A) Cruise poses from control (black) and 1 wpi (red) female zebrafish were decomposed using principal component analysis (PCA). Female fish with scoliosis score < 0.35 at 8 wpi are shown. In this posture space, cruise trajectories from acutely injured fish at 1 wpi are markedly distinct from control fish trajectories. **(B-C)** Cruise poses were embedded using UMAP with precomputed dynamic time warping (DTW) distances. Cruise poses were analyzed for fish that regenerated well (glial bridging > 20%) in B and for fish with compromised cellular regeneration (glial bridging < 20%) in C. **(D)** Gait recovery trendlines in UMAP 1, 2, and 3 (left to right) for fish that regenerated well. Colored bars indicate the mean trend of each UMAP component. Error bars are standard deviations. The vertical dotted black lines align with the average UMAP value at 3 wpi. **(E)** Gaussian kernel density estimates for the regions of UMAP space occupied by cruises at each week post-injury for fish that regenerated well. The x-axes are matched between Panels D and E for comparison. **(F)** Gait recovery trendlines in UMAP 1, 2, and 3 (left to right) for fish with compromised cellular regeneration. Colored bars indicate the mean trend of each UMAP component. Error bars are standard deviations. The vertical dotted black lines align with the average UMAP value at 3 wpi. **(G)** Gaussian kernel density estimates for the regions of UMAP space occupied by cruises at each week post-injury for fish with compromised cellular regeneration. The x-axes are matched between Panels F and G for comparison.

Figure S6

A

Pairwise Comparisons of
8 wpi Measurements
24 Female Fish

Spearman r$_s$

B Swim Capacity vs Glial Bridging
n = 27
p > 0.05

C Behavior vs Glial Bridging
n = 27
p > 0.05

D Gait vs Glial Bridging
n = 24
p > 0.05

E Glial Bridging vs Axon Regrowth
n = 27
Proximal p > 0.05
Distal p > 0.05

Figure S6. Pairwise comparisons of spinal cord regeneration metrics at 8 wpi. (A) Heatmap of Spearman's rank correlations (r_s) between the cellular and functional measurements taken. Regeneration metrics include swim capacity, swim quality, structural, and neurological measurements. A dendrogram representing similarities between correlation patterns of measurements is shown. Female fish at 8 wpi were analyzed. Fish that could not be measured in all attributes were omitted. Correlations are reported regardless of p-value. For the sake of meaningful clustering, a measured attribute was multiplied by -1 if its average was increased at 1 wpi compared to controls. These attributes are marked with the prefix "(-)" on the label. **(B-D)** One metric each of swim capacity (swim distance), behavior (burst frequency), and gait (rostral compensation) were plotted against glial bridging to demonstrate their relationship, though the correlation p-values were not significant. **(E)** Scatter plot showing a non-significant association between the size of regenerated glial tissue and axon regrowth, proximal and distal. The size of regenerated glial tissue plotted against axon regrowth, proximal and distal. r_s were not significant for Panels B-E, likely because most female fish in this experiment exhibited compromised glial regeneration.

Figure S7

A

Prediction Experiment
Pairwise Comparisons of
8 wpi Measurements
54 Fish

B Swim Capacity vs Glial Bridging, n = 56, p > 0.05. glial bridging (%) vs swim distance (px) 0, 10, 20 cm/s

C Behavior vs Glial Bridging, n = 56, $r_s = -0.37$. glial bridging (%) vs pose bursts/min 0 cm/s

D Gait vs Glial Bridging, n = 56, $r_s = -0.59$. glial bridging (%) vs rostral compensation (Δradians) 0, 10, 20 cm/s

E Glial Bridging vs Axon Regrowth, n = 56. Proximal $r_s = 0.36$, Distal p > 0.05. axon regrowth (%) vs glial bridging (%)

Figure S7. Prediction experiment: pairwise comparisons of spinal cord regeneration metrics at 8 wpi. (A) Heatmap of Spearman's rank correlations (r_s) between the cellular and functional measurements taken. Regeneration metrics include swim capacity, swim quality, structural, and neurological measurements. A dendrogram representing similarities between correlation patterns of measurements is shown. Male and female fish at 8 wpi were analyzed. Fish that could not be measured in all attributes were omitted. Correlations are reported regardless of p-value. For the sake of meaningful clustering, a measured attribute was multiplied by -1 if its average was increased at 1 wpi compared to controls. These attributes are marked with the prefix "(-)" on the label. **(B-D)** One metric each of swim capacity (swim distance), behavior (burst frequency), and gait (rostral compensation) were plotted against glial bridging to demonstrate their correlation. Because the Spearman correlation operates on ranked values, we fit monotonic splines to plots where r_s were significant ($p < 0.05$) to visualize possible associations. **(E)** Scatter plot showing a weak association between the size of regenerated glial tissue and axon regrowth, proximal and distal. Axon regrowth was measured lower than expected in this experiment.

Material and Methods

Zebrafish

Zebrafish

Adult zebrafish of the Ekkwill and AB strains were maintained at the Washington University

Zebrafish Core Facility. All animal experiments were performed in compliance with institutional

animal protocols. Male and female animals between 3 and 9 months of ~2–2.5 cm in length were

used.

SC transection

Zebrafish were anaesthetized using MS-222. Fine scissors were used to make a small incision

that transects the spinal cord 4 mm caudal to the brainstem region. Complete transection was

visually confirmed at the time of surgery. Injured animals were also assessed at 2 or 3 dpi to

confirm loss of swim capacity post-surgery.

Swim behavior recording

Two fish at a time were placed in a 5 L swim tunnel device (Loligo, cat# SW100605L, 120V/60Hz).

A customized physical divider was secured in the center of the swim chamber parallel to the flow,

separating the tunnel into 2 chambers to enable individual tracking of each fish. A camera was

placed directly above the tunnel and recorded videos at 70 frames per second. Each 15-minute

assay included three 5-minute periods of increasing water current velocity at 0, 10, and 20 cm/s.

Swim endurance assay

The swim endurance assay performed in the prediction experiment was performed as previously

described (40). Briefly, zebrafish were exercised in groups of 8–12 fish. After 10 minutes of

acclimation inside the enclosed tunnel, water current velocity was increased every two minutes

and fish swam against the current until they reached exhaustion. Exhausted animals were

removed from the chamber without disturbing the remaining fish. Swim time and current velocity

at exhaustion were recorded.

Statistical analysis

Most statistical analysis was performed in Python (https://www.anaconda.com) version 3.7.3, with

metric cluster significance (Figure 5, Figure S6, Figure S7) tested using R (46) version 4.0.4 and

sigclust2 (47, 48) version 1.2.4. One-way repeated measures ANOVA and pairwise t-test post-

hoc tests were performed using the package Pingouin (49) version 0.5.3. To test differences

between outcome prediction groups, we used independent t-test in SciPy (50) version 1.7.3. Plots

were generated using Matplotlib (51) version 3.4.3 and Seaborn (52) version 0.12.1, and were
compiled and stylized using Adobe Illustrator.

Software
All analysis was performed in Python (https://www.anaconda.com) version 3.7.3 and R (46)
version 4.0.4. The primary packages used for our analyses include NumPy (53) version 1.21.5,
pandas (54, 55) version 1.2.4, tensortools (56) version 0.3, scikit-learn (57) version 1.0.2, SciPy
(50) version 1.7.3, Pingouin (49) version 0.5.3, dtw-python (58) version 1.1.12, umap-learn (59)
version 0.4.2, DeepLabCut (24, 37) version 2.1.6.2, Matplotlib (51) version 3.4.3, Seaborn (52)
version 0.12.1, sigclust2 (47, 48) version 1.2.4, and FFmpeg (https://ffmpeg.org) version 4.3.2.

Video processing and pose annotation
We normalized each video and subtracted the background. Because a divider separated two fish
in each assay, we separated the left and right sides of the video leaving one visible fish in each
normalized video. We trained a DeepLabCut model to identify ten points on the dorsal centerline,
from the head to the base of the tail fin, and a point near the tip of the tail fin (Figure 2C) (24, 37).
To smooth the poses, we fit cubic splines to each computer-annotated pose and resampled ten
evenly spaced points from the spline. We filtered away poses that were likely incorrectly
annotated according to the following criteria: DeepLabCut likelihood score (require >0.5 at all
keypoints), total pose length (require within 8 median absolute deviations from the median length),
pose curvature (require z-score < 8 at each angle position), and whether the pose was too far
from annotated poses in adjacent frames (require movement within 45 pixel at all keypoints).

Angles along the centerline were calculated using triplets of adjacent keypoints on the fish's
centerline (see keypoint and angle positions in Figure 2C). We called this sequence of angles an
angle pose. For example, to calculate the angle at Point B from three sequential keypoints A, B,
and C, the angle at B is how far Line AB must rotate to be parallel to Line BC. In this manner,
each ten-keypoint pose was reduced to an angle pose with eight measured angles. Eight angle
positions have been shown to be sufficient to capture ordinary variation in zebrafish posture (12).
Although other research has used different methods to convert keypoints to angle poses, we
chose this representation because the summed angle pose for a resting non-scoliotic fish is close
to zero.

Behavior annotation
We annotated many episodes of simplified behaviors: rest (1355 episodes), cruise (3243), and
clockwise (128) and counterclockwise (165) turns. For the annotation model, we defined "cruise"

as a forward swim with three body oscillations minimum. If the fish performed the behavior in the

assay, we annotated three cruises per flow speed for every fish's uninjured control, 1 week-post

injury (wpi), 3 wpi, and 6 wpi assays. We annotated cruises in the same manner for all nine assays

of 34 fish. Although we focused our analysis on cruise and rest behavior, we annotated and

classified turns so the model would not mistakenly classify every non-rest behavior as cruise.

Annotations included a nearly equal representation by sex (51% female). Representation by wpi

was weighted to include more uninjured behaviors (pre: 26%, 1: 14%, 2: 6%, 3: 15%, 4: 4%, 5:

7%, 6: 14%, 7: 6%, 8: 8%).

To prepare the data for classification, episodes were split into frames. Each annotated frame's

angle pose was concatenated with the six angle poses before and the six angle poses after to

create a larger vector, a temporal "pose window", labeled with the middle frame's annotated

behavior. We withheld 25% of the annotated data for validation. Because cruises were

overrepresented in our annotations, we oversampled the train and validation datasets such that

all behaviors were equally represented with Synthetic Minority Over-sampling Technique

(SMOTE) (60) using 20 nearest neighbors.

We fit a UMAP (59) model (2 components, 50 nearest neighbors) to all pose windows in the

training set, then fit a random forest classifier (57) to the UMAP-transformed windows for final

classification. Finally, because some assays contained unusually high numbers of rest frames

incorrectly classified as active behaviors, particularly for severely scoliotic fish, we applied a

posture change threshold: if an angle pose differed no more than a certain threshold from either

the previous or the next pose, it was labeled as "rest". We optimized this threshold using the same

training set. Classifying and thresholding as described resulted in a high number of correctly

classified cruises (94.8% training, 94.8% validation) and rests (96.5% training, 95.8% validation).

We used this model to assign behavior labels to all frames with high-quality posture as previously

described.

Cruise embedding

Taking all annotated cruise episodes in each assay, we performed a 5-principal component

decomposition on the angle poses, and then created a pairwise distance matrix for these cruise

episodes using dynamic time warping. From each distance matrix, we used affinity propagation

to select exemplars, or cruises that are most representative of the fish's general behavior. If affinity

propagation failed to converge, we randomly sampled at most 16 cruises from the assay. To

calculate the embedding for several assays, we calculated a pairwise DTW distance matrix for all

exemplar cruises from the assays, then fed the distance matrix into UMAP (59) using 20 nearest

neighbors, which is consistent with published work (21). We omitted fish with scoliosis scores
higher than 0.35.

8 wpi measurement correlation clustering
Measurements from 8 wpi assays were scaled to comparable ranges using StandardScaler from
scikit-learn (57). Spearman correlations (r_s) were calculated pairwise between measurements and
a distance matrix of $1 - r_s$ was assembled using SciPy (50). Average linkage hierarchical
clustering was performed, and significance between clusters was tested, using sigclust2, which
was modified to accept $1 - r_s$ as the distance metric.

TCA and fish clustering
A rank-3 tensor was created by concatenating all fish with their measurements in all assays
(sorted temporally) in the tracking experiment. Measurements were scaled to comparable ranges
using StandardScaler from scikit-learn (57). We omitted 6 fish that died before 8 wpi and 10 fish
that did not perform a measurable cruise at every assayed week. Tensor component analysis was
performed with tensortools (56). We compared ranks from 1 to 90 for three varieties of canonical
polyadic (CP) tensor decompositions: unconstrained by alternating least squares (cp_als),
nonnegative by block coordinate descent (ncp_bcd) and nonnegative by hierarchical alternating
least squares (ncp_hals). After comparing model error and replicate similarity for 4 replicates of
each rank and model, our chosen combination was rank-7 CP decomposition by alternating least
squares (Figure 6A). We clustered fish according to their factor values using agglomerative
clustering with distance threshold of 1.7 using scikit-learn. This distance threshold was chosen by
comparing the number of clusters to the minimum cluster size for 1000 threshold values spaced
evenly from 0.001 to 5 (Figure 6B).

Measurements of functional and structural recovery
Classical Function

• Perceived Swim Quality
o This score was assigned by observation on a scale from 1 (low quality) to 5 (high
quality). We first found the best and worst fish, then scored all fish compared to
these. We focused our observation on the first few minutes when the flow started
(5 min) and when the flow accelerated (10 min). Our criteria were as follows:

5: Fish's swimming pattern looks normal. Mostly uses tail movement, not its head,
to swim. No scoliosis.

4: Fish seems like it can swim at the same rate and speed as those categorized as 5. Slight scoliosis. Slightly less tail movement than 5, and slightly more head movement or "wiggling".

3: Fish cannot swim as effectively as 4. Equal head and tail movement. Scoliosis looks "bent" at about a 45-degree angle.

2: Fish have some tail movement, can rotate their body, but cannot swim forward effectively. Severe scoliosis with tail movement. No swimming once flow started.

1: Fish have no tail movement. Severe scoliosis is observed. No swimming once flow started.

- o We treated the above criteria as guidelines because not all fish had all the features as described. For example, a fish could score "3" due to excessive head movement without significant scoliosis.
- Mean y-position
  - o Average centroid position along the water flow axis in the swim tunnel. Higher "mean y" was closer to the flow source. It was assumed that exhausted fish would be pushed backward by the flow and have a lower mean-y. We reported mean y-position only for periods of the behavior assay when water was flowing (10, 20 cm/s).
- Distance
  - o Sum of frame-to-frame centroid displacement measured in pixels. In this study we did not adjust distance according to water flow velocity, and we measured during the entire assay at all flow speeds.
- Activity
  - o Sum of frames where centroid displacement (see Distance) was above a threshold, divided by the total number of frames. In this study we did not adjust activity according to water flow velocity, and we measured during the entire assay at all flow speeds.
- Centroid Burst Frequency
  - o We defined a centroid burst as an uninterrupted period of active frames (see Activity). Centroid burst frequency is the number of centroid bursts per minute. We reported this measurement only for periods of the behavior assay when water was not flowing (0 cm/s).

578 • Time Against Flow

579   ○ Sum of frames where centroid displacement (see Distance) is above a threshold
580   in the direction of the flow, allowing 45-degree deviation in either direction, divided
581   by the total number of frames where water is flowing. Accordingly, time against
582   flow was only calculated for periods of the behavior assay when water was flowing
583   (10, 20 cm/s).

584 • Endurance or Time to Exhaustion

585   ○ The duration of time that a fish can swim against flowing water until exhausted.
586   Fish allow the fish to acclimate in calm water, then increase the flow speed
587   periodically. When the fish can no longer pull away from a barrier at the back of
588   the tunnel, it is exhausted.

589 Gait and Posture

590 • Pose Burst Frequency

591   ○ Same as Centroid Burst Frequency but active frames were defined as those with
592   active posture; that is, for frames not classified as "rest" by our behavior classifier.
593   Although this calculation for activity did not depend on flow velocity, we only
594   reported this value for periods without water flow (0 cm/s) to be comparable with
595   our reported centroid burst frequency.

596 • Lateral Scoliosis

597   ○ For each period of rest, or inactive posture, we calculated the average sum of each
598   angle pose, then took the weighted average of these values over the entire swim
599   where weights are the length of the rest periods. Although the calculation would
600   have been essentially equivalent for non-scoliotic fish, we chose to analyze each
601   rest bout separately and average their results rather than taking the average
602   posture over the entire swim because some fish with severely poor recovery could
603   not control their dorsal-ventral orientation. For example, when a scoliotic fish
604   flipped to its back, its posture inverted; and repeated flipping lead to an artificially
605   lower scoliosis score when posture was averaged over the entire swim. Hence,
606   each period of rest was analyzed separately, then values for all rest periods were
607   averaged as described.

608 • Cruise Angular Frequency

609   ○ The number of posture oscillations per second while swimming forward. Angular
610   frequency was mostly uniform along the centerline (Figure S4A). We computed

611   cruise angular frequency for each of the eight computed angle positions along a

612   fish's centerline, then averaged over the cruise. All cruise values were then

613   averaged to get a single score for each assay. We reported average cruise angular

614   frequency values in Figure S4B and displacement from control average in Figure

615   S4C.

616   •   Cruise Rostral Compensation

617   o   First, we calculated average angular amplitudes at each angle position along the

618   dorsal centerline during episodes of cruise behavior. We called this a cruise

619   curvature profile. We calculated a control profile as the average profile of uninjured

620   fish. We defined rostral compensation as the maximum absolute difference

621   between an assay's cruise curvature profile and the control profile at rostral angle

622   positions α_1 to α_5 (Figure 2C).

623   •   Posture Novelty

624   o   We trained a Local Outlier Factor (LOF) model using sampled uninjured control

625   poses, then used the model to perform novelty detection on all assays. An assay's

626   posture novelty score was calculated as the number of poses identified as "novel"

627   by LOF divided by the total number of poses in the assay.

628   o   We created a representative training set by sampling 1200 poses from each control

629   assay: 100 poses from 12 k-mean clusters. K-means clustering was performed

630   separately for each assay.

631   •   Tail Beat Frequency

632   o   Using an annotated point near the tip of the caudal fin, we measured lateral

633   extrema, then calculated oscillation frequency using those measurements.

634   •   Strouhal Number

635   o   We calculate tail beat frequencies and peak-to-peak caudal fin tip displacement

636   amplitudes for each cruise episode. Strouhal number (St) was defined as St =

637   fA/U, which is tail beat frequency (f) times amplitude (A) divided by forward speed

638   (U) (38). To get average St for one assay, we calculated St for all cruise episodes

639   then took the average of these numbers weighted by the number of periods in each

640   respective cruise.

641 Spinal Cord Structure

642   •   % Bridging

643   ○ GFAP immunohistochemistry was performed on serial transverse sections as
644   previously described(40). The cross-sectional area of the glial bridge and the area
645   of the intact SC rostral to the lesion were measured using ImageJ (Fiji)
646   software(61). Bridging was calculated as a ratio of these measurements.

647   • % Axon Regrowth Proximal and Distal

648   ○ Anterograde axon tracing was performed as previously described(40). Fish were
649   anaesthetized using MS-222 and fine scissors were used to transect the cord
650   4 mm rostral to the lesion site. Biocytin-soaked Gelfoam Gelatin Sponge was
651   applied at the new injury site (Gelfoam, Pfizer, cat# 09-0315-08; Biocytin, saturated
652   solution, Sigma, cat# B4261). Fish were euthanized 6 hours post-treatment and
653   Biocytin-labeled axons were histologically detected using Alexa Fluor 594-
654   conjugated Streptavidin (Thermo Fisher, cat# S-11227). Biocytin-labeled axons
655   were quantified using the "threshold" and "particle analysis" tools in the Fiji
656   software. Four sections per fish at 0.5 (proximal) and 2 (distal) mm caudal to the
657   lesion core, and 2 sections 1 mm rostral to the lesion, were analyzed. Regrowth
658   was calculated as a ratio of these measurements.

Bibliography

1. Cigliola V, Becker CJ, Poss KD. Building bridges, not walls: spinal cord regeneration in zebrafish. Disease Models & Mechanisms. 2020;13(5):dmm044131.

2. Mokalled MH, Patra C, Dickson AL, Endo T, Stainier DYR, Poss KD. Injury-induced ctgfa directs glial bridging and spinal cord regeneration in zebrafish. Science. 2016;354(6312):630-4.

3. Becker T, Wullimann MF, Becker CG, Bernhardt RR, Schachner M. Axonal regrowth after spinal cord transection in adult zebrafish. The Journal of Comparative Neurology. 1997;377(4):577-95.

4. Goldshmit Y, Sztal TE, Jusuf PR, Hall TE, Nguyen-Chi M, Currie PD. Fgf-dependent glial cell bridges facilitate spinal cord regeneration in zebrafish. The Journal of Neuroscience. 2012;32(22):7477-92.

5. Strand NS, Hoi KK, Phan TMT, Ray CA, Berndt JD, Moon RT. Wnt/β-catenin signaling promotes regeneration after adult zebrafish spinal cord injury. Biochemical and biophysical research communications. 2016;477(4):952-6.

6. van Raamsdonk W, Maslam S, de Jong DH, Smit-Onel MJ, Velzing E. Long term effects of spinal cord transection in Zebrafish: swimming performances, and metabolic properties of the neuromuscular system. Acta Histochemica. 1998;100(2):117-31.

7. Becker CG, Lieberoth BC, Morellini F, Feldner J, Becker T, Schachner M. L1.1 is involved in spinal cord regeneration in adult zebrafish. The Journal of Neuroscience. 2004;24(36):7837-42.

8. Klatt Shaw D, Mokalled MH. Efficient CRISPR/Cas9 mutagenesis for neurobehavioral screening in adult zebrafish. G3 (Bethesda, Md). 2021;11(8).

9. Burris B, Jensen N, Mokalled MH. Assessment of swim endurance and swim behavior in adult zebrafish. Journal of Visualized Experiments. 2021(177).

10. Saraswathy VM, Zhou L, Mcadow AR, Burris B, Dogra D, Reischauer S, et al. Myostatin is a negative regulator of adult neurogenesis after spinal cord injury in zebrafish. Cell Reports. 2022;41(8):111705.

11. Huang C-X, Zhao Y, Mao J, Wang Z, Xu L, Cheng J, et al. An injury-induced serotonergic neuron subpopulation contributes to axon regrowth and function restoration after spinal cord injury in zebrafish. Nature Communications. 2021;12(1):7093.

12. Huang C-X, Wang Z, Cheng J, Zhu Z, Guan NN, Song J. De novo establishment of circuit modules restores locomotion after spinal cord injury in adult zebrafish. Cell Reports. 2022;41(4):111535.

13. Chang W, Pedroni A, Bertuzzi M, Kizil C, Simon A, Ampatzis K. Locomotion dependent neuron-glia interactions control neurogenesis and regeneration in the adult zebrafish spinal cord. Nature Communications. 2021;12(1).

14. Hosseini P, Mirsadeghi S, Rahmani S, Izadi A, Rezaei M, Ghodsi Z, et al. Dopamine Receptors Gene Expression Pattern and Locomotor Improvement Differ Between Female and Male Zebrafish During Spinal Cord Auto Repair. Zebrafish. 2022;19(4):137-47.

15. Zeng C-W, Kamei Y, Shigenobu S, Sheu J-C, Tsai H-J. Injury-induced Cavl-expressing cells at lesion rostral side play major roles in spinal cord regeneration. Open biology. 2021;11(2):200304.

16. Vasudevan D, Liu Y-C, Barrios JP, Wheeler MK, Douglass AD, Dorsky RI. Regenerated interneurons integrate into locomotor circuitry following spinal cord injury. Experimental Neurology. 2021;342:113737.

17. Hossainian D, Shao E, Jiao B, Ilin VA, Parris RS, Zhou Y, et al. Quantification of functional recovery in a larval zebrafish model of spinal cord injury. Journal of Neuroscience Research. 2022.

18. Doyle LMF, Roberts BL. Exercise enhances axonal growth and functional recovery in the regenerating spinal cord. Neuroscience. 2006;141(1):321-7.

19.   Ljunggren EE, Haupt S, Ausborn J, Ampatzis K, El Manira A. Optogenetic Activation of Excitatory Premotor Interneurons Is Sufficient to Generate Coordinated Locomotor Activity in Larval Zebrafish. Journal of Neuroscience. 2014;34(1):134-9.

20.   Girdhar K, Gruebele M, Chemla YR. The behavioral space of zebrafish locomotion and its neural network analog. Plos One. 2015;10(7):e0128668.

21.   Mearns DS, Donovan JC, Fernandes AM, Semmelhack JL, Baier H. Deconstructing Hunting Behavior Reveals a Tightly Coupled Stimulus-Response Loop. Current Biology. 2020;30(1):54-69.e9.

22.   Fontaine E, Lentink D, Kranenbarg S, Müller UK, van Leeuwen JL, Barr AH, et al. Automated visual tracking for studying the ontogeny of zebrafish swimming. Journal of Experimental Biology. 2008;211(Pt 8):1305-16.

23.   Thomas LSV, Gehrig J. Multi-template matching: a versatile tool for object-localization in microscopy images. BMC Bioinformatics. 2020;21(1).

24.   Mathis A, Mamidanna P, Cury KM, Abe T, Murthy VN, Mathis MW, et al. DeepLabCut: markerless pose estimation of user-defined body parts with deep learning. Nature Neuroscience. 2018;21(9):1281-9.

25.   Graving JM, Chae D, Naik H, Li L, Koger B, Costelloe BR, et al. DeepPoseKit, a software toolkit for fast and robust animal pose estimation using deep learning. eLife. 2019;8.

26.   Liu X, Yu S-Y, Flierman NA, Loyola S, Kamermans M, Hoogland TM, et al. OptiFlex: Multi-Frame Animal Pose Estimation Combining Deep Learning With Optical Flow. Frontiers in Cellular Neuroscience. 2021;15:621252.

27.   Pereira TD, Tabris N, Matsliah A, Turner DM, Li J, Ravindranath S, et al. SLEAP: A deep learning system for multi-animal pose tracking. Nature Methods. 2022;19(4):486-95.

28.   Mwaffo V, Zhang P, Romero Cruz S, Porfiri M. Zebrafish swimming in the flow: a particle image velocimetry study. PeerJ. 2017;5:e4041.

29.   McCullough MH, Goodhill GJ. Unsupervised quantification of naturalistic animal behaviors for gaining insight into the brain. Current Opinion in Neurobiology. 2021;70:89-100.

30.   Hsu AI, Yttri EA. B-SOiD, an open-source unsupervised algorithm for identification and fast prediction of behaviors. Nature Communications. 2021;12(1).

31.   Sun G, Lyu C, Cai R, Yu C, Sun H, Schriver KE, et al. DeepBhvTracking: A novel behavior tracking method for laboratory animals based on deep learning. Frontiers in Behavioral Neuroscience. 2021;15:750894.

32.   Bohnslav JP, Wimalasena NK, Clausing KJ, Dai YY, Yarmolinsky DA, Cruz T, et al. DeepEthogram, a machine learning pipeline for supervised behavior classification from raw pixels. eLife. 2021;10.

33.   Xu J, Hu C, Jiang Q, Pan H, Shen H, Schachner M. Trimebutine, a small molecule mimetic agonist of adhesion molecule L1, contributes to functional recovery after spinal cord injury in mice. Disease Models & Mechanisms. 2017;10(9):1117-28.

34.   Vrinten DH, Hamers FFT. 'CatWalk' automated quantitative gait analysis as a novel method to assess mechanical allodynia in the rat; a comparison with von Frey testing. Pain. 2003;102(1-2):203-9.

35.   Barbeau H, Ladouceur M, Norman KE, Pépin A, Leroux A. Walking after spinal cord injury: evaluation, treatment, and functional recovery. Archives of Physical Medicine and Rehabilitation. 1999;80(2):225-35.

36.   Maynard FM, Bracken MB, Creasey G, Donovan WH, Ducker TB, Garber SL, et al. International standards for neurological and functional classification of spinal cord injury. Spinal cord. 1997;35(5):266-74.

37.   Nath T, Mathis A, Chen AC, Patel A, Bethge M, Mathis MW. Using DeepLabCut for 3D markerless pose estimation across species and behaviors. Nature Protocols. 2019;14(7):2152-76.

38.   Taylor GK, Nudds RL, Thomas ALR. Flying and swimming animals cruise at a Strouhal number tuned for high power efficiency. Nature. 2003;425(6959):707-11.
39.   Delcourt J, Ovidio M, Denoël M, Muller M, Pendeville H, Deneubourg J-L, et al. Individual identification and marking techniques for zebrafish. Reviews in Fish Biology and Fisheries. 2018;28(4):839-64.
40.   Klatt Shaw D, Saraswathy VM, Zhou L, Mcadow AR, Burris B, Butka E, et al. Localized EMT reprograms glial progenitors to promote spinal cord repair. Developmental Cell. 2021;56(5):613-26.e7.
41.   Beal DN, Hover FS, Triantafyllou MS, Liao JC, Lauder GV. Passive propulsion in vortex wakes. Journal of fluid mechanics. 2006;549(-1):385.
42.   Barrios JP, Wang W-C, England R, Reifenberg E, Douglass AD. Hypothalamic dopamine neurons control sensorimotor behavior by modulating brainstem premotor nuclei in zebrafish. Current Biology. 2020;30(23):4606-18.e4.
43.   Bagnall MW, McLean DL. Modular organization of axial microcircuits in zebrafish. Science. 2014;343(6167):197-200.
44.   Reimer MM, Sörensen I, Kuscha V, Frank RE, Liu C, Becker CG, et al. Motor neuron regeneration in adult zebrafish. The Journal of Neuroscience. 2008;28(34):8510-6.
45.   Kathe C, Skinnider MA, Hutson TH, Regazzi N, Gautier M, Demesmaeker R, et al. The neurons that restore walking after paralysis. Nature. 2022;611(7936):540-7.
46.   Team RC. R: A Language and Environment for Statistical Computing. Vienna, Austria2021.
47.   Kimes PK, Liu Y, Neil Hayes D, Marron JS. Statistical significance for hierarchical clustering. Biometrics. 2017;73(3):811-21.
48.   Kimes P. sigclust2: sigclust2: Statistical Significance of Clustering. 2018.
49.   Vallat R. Pingouin: statistics in Python. Journal of Open Source Software. 2018;3(31):1026.
50.   Virtanen P, Gommers R, Oliphant TE, Haberland M, Reddy T, Cournapeau D, et al. SciPy 1.0: fundamental algorithms for scientific computing in Python. Nature Methods. 2020;17(3):261-72.
51.   Hunter JD. Matplotlib: A 2D Graphics Environment. Computing in Science & Engineering. 2007;9(3):90-5.
52.   Waskom M. seaborn: statistical data visualization. Journal of Open Source Software. 2021;6(60):3021.
53.   Harris CR, Millman KJ, Van Der Walt SJ, Gommers R, Virtanen P, Cournapeau D, et al. Array programming with NumPy. Nature. 2020;585(7825):357-62.
54.   The pandas development t. pandas-dev/pandas: Pandas 1.0.3. Zenodo. 2020.
55.   McKinney W, editor Data structures for statistical computing in python. Python in Science Conference; 2010 2010: SciPy.
56.   Williams AH, Kim TH, Wang F, Vyas S, Ryu SI, Shenoy KV, et al. Unsupervised Discovery of Demixed, Low-Dimensional Neural Dynamics across Multiple Timescales through Tensor Component Analysis. Neuron. 2018;98(6):1099-115.e8.
57.   Pedregosa F, Varoquaux G, Gramfort A, Michel V, Thirion B, Grisel O, et al. Scikit-learn: Machine Learning in Python. Journal of Machine Learning Research. 2011;12:2825-30.
58.   Giorgino T. Computing and Visualizing Dynamic Time Warping Alignments in R: The dtw Package. Journal of Statistical Software. 2009;31(7):1-24.
59.   McInnes L, Healy J, Saul N, Großberger L. UMAP: uniform manifold approximation and projection. The Journal of Open Source Software. 2018;3(29):861.
60.   Lemaitre G, Nogueira F, Aridas CK. Imbalanced-learn: A Python Toolbox to Tackle the Curse of Imbalanced Datasets in Machine Learning. Journal of Machine Learning Research. 2017;18:1-5.

61.   Schindelin J, Arganda-Carreras I, Frise E, Kaynig V, Longair M, Pietzsch T, et al. Fiji: an open-source platform for biological-image analysis. Nature Methods. 2012;9(7):676-82.