Title: Automated Workflows using Quantitative Colour Pattern Analysis (QCPA): A Guide to

Batch Processing and Downstream Data Analysis

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Abstract

Animal and plant colouration presents a striking dimension of phenotypic variation, the study

of which has driven general advances in ecology, evolution, and animal behaviour.

Quantitative Colour Pattern Analysis (QCPA) is a dynamic framework for analysing colour

patterns through the eyes of non-human observers. However, its extensive array of user-

defined image processing and analysis tools means image analysis is often time-consuming.

This hinders the full use of analytical power provided by QCPA and its application to large

datasets. Here, we offer a robust and comprehensive batch script, allowing users to automate

many QCPA workflows. We further provide a set of useful R scripts for downstream data

extraction and analysis. These scripts will empower users to exploit the full analytical power

of QCPA and facilitate the development of customised semi-automated workflows. Such

quantitatively scaled workflows are crucial for exploring colour pattern space and developing

ever-richer models of visual perception in animals other than humans. These advances will,

in turn, facilitate testing hypotheses on the function and evolution of vision and signals at

quantitative and qualitative scales, which are otherwise computationally unfeasible.

1. Introduction

Understanding the perception of visual information by non-human observers is crucial to studying the ecology and evolution of animal and plant colouration. The last two decades have seen the creation and widespread adaptation of tools and methods that allow researchers to simulate or approximate aspects of animal vision, such as colour contrast perception (e.g. Endler and Mielke, 2005; Gawryszewski, 2018; Kemp et al., 2015; Renoult et al., 2017; Vorobyev and Osorio, 1998) and spatial vision (Godfrey et al. 1987; Caves and Johnsen 2018). These advances coincide with the steady development of colour pattern analyses (e.g. Chan et al., 2018; Endler, 1991; Endler, 2012; Stoddard et al., 2014; Van Belleghem et al., 2018; van den Berg et al., 2020) as well as their integration into increasingly comprehensive collections of tools and functions such as Pavo (Maia et al. 2019) or the Multispectral Image Calibration and Analysis toolbox (MICA) (Troscianko and Stevens 2015).

Quantitative Colour Pattern Analysis (QCPA) (van den Berg et al. 2020b) is a recent and powerful addition to this landscape. It provides a dynamic analytical framework for analysing visual scenes through the eyes of ecologically relevant observers and is integrated into the MICA toolbox (Troscianko and Stevens 2015). It allows users to choose preferred tools and analyses uniquely suitable for analysing spatiochromatic information. Briefly, it achieves this by taking full-spectrum images as its input (thereby generating '.mspec' files) before applying a suite of models which can consider the spectral, spatial, and temporal sensation of non-human viewers, to ultimately give image-based and numerical outputs that summarise the structure of visual scenes and stimuli (van den Berg et al. 2020b).

Preparing and processing calibrated images in the framework, however, remains tedious. This is due to the time needed to translate calibrated digital images into cone catch

images, user-guided input for identifying regions of interest (ROIs), and the subsequent application of image processing and analyses. Saving and labelling multiple output files, as well as extracting data for subsequent statistical analysis, also remains a tiresome process. As a result, applying the QCPA at quantitative scales, such as analysing hundreds or thousands of images at multiple viewing distances, is nearly impossible in its original form.

An image currently needs hours of manual, repetitive work to obtain the full numerical output from the various types of analyses in the QCPA framework. This is especially true when considering multiple viewing distances and regions of interest (ROIs). Consequently, most studies using the framework do not consider more than a few dozen individual observations with a limited subset of available image statistics (e.g., Nokelainen et al., 2021; Rodríguez-Morales et al., 2021). This is problematic as quantifying ecologically relevant effects, such as colour pattern variability in natural populations or the analysis of behavioural data, requires sufficiently large sample sizes to provide adequate statistical power. This holds particularly true for a method producing hundreds of image statistics from a single observation, such as the QCPA, where the number of available image statistics easily outnumbers observations (van den Berg et al. 2022). Considering all possibly relevant image statistics and deducing a relevant set of parameters is often the desired approach instead of pre-emptively narrowing down the number of considered image statistics (van den Berg et al. 2022). However, such deductive approaches require adequate statistical solutions and remain an intriguing challenge in using complex colour pattern spaces (Stoddard and Osorio 2019; van den Berg et al. 2020b, 2022).

The QCPA framework is, therefore, in urgent need of automation to facilitate its complete application to large datasets. However, unlike other image analysis tools available

to visual ecologists, such as PAVO (Maia et al. 2019), MICA is an ImageJ (Schneider et al. 2012)

plugin written in Java (Arnold et al. 2005). It is intentionally tailored towards accessibility via

graphical user interfaces (GUIs) rather than command line prompts. Unlike R software (R Core

Team 2021) or Python (van Rossum 1995), frequently taught in undergraduate and HDR

Biology degrees worldwide, Java is not a programming language that biologists are readily

familiar with. Therefore, writing scripts and macros for a complex network of GUI-guided tools

in an unfamiliar language is a hurdle for most users of the QCPA framework seeking to batch-

process large volumes of image data. This creates the potential for unintentional gatekeeping

by researchers with access to programmers familiar with writing scripts in JAVA and an

adequate understanding of the framework's mechanisms. The result is a hidden custom of

informally traded scripts, which undermines fast and equal access to methods in the field.

Here, we present a dynamic batch-processing extension to QCPA, allowing users to

apply almost the entire QCPA framework or selected parts to large datasets via a flexible, GUI-

guided input. We further provide a set of R scripts that efficiently extract numerical data from

the consecutively stored output files for downstream analysis. We describe the functionality

of both—the QCPA batch script and complementary R scripts—and provide detailed worked

examples. While this script does not provide a complete solution for every possible

application of the QCPA, we hope it will offer a viable solution to most. This batch script

should also help researchers customise their automated pipelines, stimulating the open

exchange of programming solutions via platforms such as open-access publications, open-

access platforms such as GitHub or the dedicated user forum for the MICA toolbox

(www.empiricalimaging.com).

2. QCPA batch script

2.1. Intended use

The QCPA batch script is intended to quantify spatiochromatic information of an animal

against its visual background. As such, the script can also be applied to images without an

animal or object of interest or an animal or object itself. The batch script is intended for users

with moderate experience in the manual application of QCPA.

The script allows the user to choose the following desired analytical outputs

individually: 1) Colour Adjacency Analysis (CAA), Visual Contrast Analysis (VCA), Boundary

Strength Analysis (BSA) and particle analysis; 2) Local Edge Intensity Analysis (LEIA); 3. Colour

Maps; and 4) GabRat. For a detailed discussion of these analyses, please see the original

publications and their modifications for QCPA as listed in van den Berg et al. (2020).

Notably, the script requires cone mapping functions derived from calibrated cameras

(several standard profiles included in the MICA toolbox) and currently does not permit the

use of chart-based models. While covering many analyses available in QCPA and the MICA

toolbox, it does not provide a comprehensive library of automation. However, the script is

designed to be easily modified by the user. Such modifications are explicitly invited from the

community, and we encourage their open sharing on community platforms in the interest of

mutual benefit among researchers in the field.

2.2. Data preparation

Image processing in QCPA is specific to a variety of user choices specifying the properties

of the observer's visual system, the light environment in which a picture was taken as well as

the intended target light environment, observer viewing distance, and a variety of parameters

and processing choices that vary depending on the desired analyses. The batch script uses

three approaches to minimise the need to repeat the number of times such input is required:

1. Pre-defined folder structure

2. Auxiliary files

3. Batch processing graphical user interface (Batch GUI)

2.2.1 Folder structure

The batch script requires input data to be organised in a specific way. This refers to

the naming of files and the folder structure. For example, the script expects individual

observations (i.e., animals or objects) to be grouped within an overarching folder (i.e., species

or site) and file naming must be uniform. This allows the script to reliably detect individual

observations while allowing the user to structure their data according to sites or taxa (see the

provided datasets for specific examples).

2.2.2 Auxiliary files

The batch script uses the MICA toolbox's pre-existing approach to associate several

files with each image to be analysed. In addition to the already existing need for a folder with

the specified ROIs and the corresponding .mspec of each image, the batch script requires two

additional text files specifying the rotation of each image and the cone mapping function (see

the manual for details). These allow users to standardise the orientation of all images before

analysis and permit multiple cone mapping functions within the same dataset (e.g., multiple

lighting environments).

2.2.3. Batch GUI

The batch script allows the user to specify which of the available analyses they want

to conduct and what settings they want to use. The Batch GUI consists of a central interface

and a suite of dedicated, tool-specific GUIs activated by user choices. These tool-specific GUIs

provide helpful information and allow running all analyses with independent settings.

Furthermore, the batch script remembers past user input and will pre-fill the previous choices

by the user, helping to test settings and maintain track of chosen settings in case of

incomplete runs. See the manual and worked examples for detailed instructions on how to

use them. Users familiar with QCPA and MICA toolbox will find many GUIs similar or identical

to existing ones.

2.3. Workflow recommendations

The Batch GUI, manual and worked examples contain a suite of recommendations in

addition to those in van den Berg et al. (2020) and the empiricalimaging.com website that

users might find informative. We recommend that the user validate the numerical accuracy

of the batch script output on a small dataset before analysing large datasets. This also allows

the user to confirm that the folder structure and auxiliary files have been arranged correctly.

As processing large datasets can take many hours, days, or even weeks, we recommend that

the user consider running sub-sections of larger datasets on different instances, such as

multiple computers, servers, or cloud-based computing facilities. This will ultimately save

time and prevent data loss while helping to detect faulty files in large datasets faster.

Like all approaches to visual modelling, image analysis with QCPA requires many

different input choices by the user. Keeping a record of these choices is crucial for two

reasons. First, it allows the user to keep track of settings, facilitating record-keeping and

collaboration. Second, it provides for the publication of repeatable research. The latter is

often an issue in visual modelling studies, as modelling choices are often poorly documented

(White et al. 2015). To this end, the QCPA batch script keeps various detailed log files in the

data output. These can easily be added to the supplementary information of any publications

or uploaded/shared as part of the data.

3. R script library

We provide a set of tailored R-scripts enabling the extraction and compilation of batch QCPA

output data into .csv tables that can be used for downstream analysis. The efficient handling

of QCPA batch output is a crucial step in using QCPA at larger scales, as various outputs are

stored in files across a dedicated data set structure. The scripts and functions we provide here

are formulated and commented on in a way that is aimed to facilitate modification by the

user. These scripts and functions are intended to provide guidance for people with limited

experience in managing large and complex datasets. However, these scripts and functions

also aim to provide a starting point for more versed users to tailor their own data analysis

pipelines. A detailed manual for using the R scripts can be found in the electronic Supplement

or here: https://github.com/CaraConradsen/QCPA-r-script.

The R environment provided includes the following functionality:

1. Compiling CAA, VCA, BSA, and particle analysis data.

2. Compiling data from LEIA

3. Compiling data from GabRat

4. Compiling data from Particle Analysis and secondary data output

5. Merging, data cleaning and reshaping various subsets of data

6. Exploratory data visualisation and analysis

7. Exporting compiled files

4. Worked examples

We provide detailed worked examples with test data and corresponding toolbox files, such as

cone mapping models. These aim to highlight the functionality of the batch script and allow

users to familiarise themselves with examples related to their needs. Instructions for the

worked examples can be found in the electronic Supplement or here:

https://github.com/cedricvandenberg/QCPA-batch-script.

4.1 Trichromatic example without UV (sea slugs seen by a triggerfish, *Rhinecanthus aculeatus*)

Here, we showcase the use of the batch script by applying all modules in the script to

investigate how a triggerfish (*R. aculeatus*) perceives two species of cryptic sea slugs

(*Aphelodoris varia, Aplysia sp.*) from the east coast of Australia (Nelson Bay, NSW)

photographed underwater (Fig. 1).

Figure 1 A) *Rhinecanthus aculeatus* (A. Pingstone, public domain). Calibrated images from the test dataset of *Aphelodoris varia* (B) and *Aplysia sp.* (C) by CvdB.

208   4.2 Tetrachromatic example with UV (spiders seen by a bluetit, *Parus major*)

209   This example uses all batch script modules to analyse cryptic spiders (*Tamopsis brisbanensis*)

210   found on eucalypt trees in Sydney, NSW (Australia) (Fig. 2).

Figure 2 A) *Parus major* (© Francis C. Franklin / CC-BY-SA-3.0); B) Calibrated image of *Tamopsis brisbanensis,* taken by A. Aceves (with permission).

5. Discussion

We provide a user-friendly, open-source batch processing extension to QCPA, allowing for the

semi-automated analysis of large datasets within the QCPA framework. This greatly facilitates

the full-scale use of the analytical power of the framework as intended by its original design

(van den Berg et al. 2020b). The open-source code of the script is presented and structured

with the deliberate purpose of enabling users to create customised versions of the batch

script, which might be more suitable to their intended purposes. The batch script comes with

a detailed manual guiding users in preparing and analysing the image data. We further

provide three worked examples and corresponding test data, covering many contexts where

users will likely use the script. Lastly, the user is provided with a set of R-scripts aiding in the

downstream analysis of the data generated by the batch script. These scripts are presented

and structured in a way that facilitates users to customise them to their needs.

The script enables the semi-automated analysis of large image datasets, computing

hundreds of colour pattern descriptors at multiple viewing distances for dozens, hundreds, or

even thousands of observations. This has previously been impossible due to the time-

consuming manual processing of images in QCPA. As QCPA and other colour pattern analyses

using visual modelling approaches (e.g. Maia et al., 2019; Troscianko and Stevens, 2015)

merely relate to approximations of early-stage visual processing (e.g. Endler and Mielke,

2005; Marr, 2010; van den Berg et al., 2020a; van den Berg et al., 2020b; Vorobyev and Osorio,

1998), considering the context and task-dependent relationships with and between a broad

range of descriptors is critical (see van den Berg et al., 2022 for discussion). Thus, the

presented batch script provides a crucial step towards such analytical approaches.

As QCPA provides a conceptual approach towards combining visual modelling and

colour pattern analyses, automating workflows and advancing customisability represent

essential stepping stones towards the continued development and refinement of the

framework and its contribution towards alternative methods. By providing an inviting, well-

annotated, intuitive, open-access and open-source environment, we hope to facilitate the

application of and, importantly, eventual modifications and additions to the framework and

its combined use with other existing methodologies. Cumulatively, this round-table

philosophy will not only advance the development and testing of methods in the field but,

crucially, will facilitate the increased critical understanding of and familiarity with existing

methodology among members of the field. On a more tangible note, this script can easily be

adjusted to enable the analysis of different data structures or include processing steps and

analyses available in QCPA or the MICA toolbox currently not considered in the script. The R

scripts and functions provided here provide useful guidance for users with variable levels of

experience in data wrangling. However, more extensive forms of such customised data

extraction functions will provide an important step towards the dynamic use of large-scale

QCPA output across multiple colour pattern analysis frameworks in the R environment, such

as PAVO.

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255 7. Declarations

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7.2 Conflicts of interest

The authors declare no conflict of interest.

7.3. Ethics approval

Not applicable

7.4 Consent to participate

Not applicable

7.5 Consent for publication

All authors consent to the publication of the manuscript and associated documents.

7.6 Availability of data and material

All data and supplementary materials are available at the respective repositories listed below.

7.7 Code availability

All code is available at the respective repositories listed below.

8. Author's contributions

CvdB – Overall concept (lead), testing, coding, original draft, manuscript writing & review,

Funding, NC –coding batch script (lead), Manuscript review, CC – R scripts (lead), Manuscript

review, TEW – R scripts, Manuscript review, Funding, KLC – Original concept, Funding,

Manuscript review.

9. Supplementary data

Batch script ImageJ plugin, Batch script manual, worked examples guide:

https://github.com/cedricvandenberg/QCPA-batch-script

R scripts and functions:

https://github.com/CaraConradsen/QCPA-r-script

Test data for the worked examples:

https://doi.org/10.48610/3cdcc1f

10. References

10. References

Arnold K, Gosling J, Holmes D (2005) The Java programming language. Addison Wesley

Professional

Caves EM, Johnsen S (2018) AcuityView: An r package for portraying the effects of visual

acuity on scenes observed by an animal. Methods Ecol Evol 9:793–797.

https://doi.org/10.1111/2041-210X.12911

Chan IZW, Stevens M, Todd PA (2018) PAT- GEOM: A Software Package for the Analysis of

Animal Patterns. Methods Ecol Evol 2041–210X.13131. https://doi.org/10.1111/2041-

210X.13131

Endler JA (2012) A framework for analysing colour pattern geometry: Adjacent colours. Biol J

Linn Soc 107:233–253. https://doi.org/10.1111/j.1095-8312.2012.01937.x

Endler JA (1991) Interactions between predators and prey. In: Krebs JR, Davies NB (eds)

Behavioural Ecology. Blackwell Scientific, Oxford, pp 169–196

Endler JA, Mielke PW (2005) Comparing entire colour patterns as birds see them. Biol J Linn

Soc 86:405–431. https://doi.org/10.1111/j.1095-8312.2005.00540.x

Gawryszewski FM (2018) Color vision models: Some simulations, a general n -dimensional

model, and the colourvision R package. Ecol Evol 8:8159–8170.

https://doi.org/10.1002/ece3.4288

Godfrey D, Lythgoe JN, Rumball D a (1987) Zebra stripes and tiger stripes: the spatial

frequency distribution of the pattern compared to that of the background is significant

in display and crypsis. Biol J Linn Soc 32:427–433. https://doi.org/10.1111/j.1095-

8312.1987.tb00442.x

Kemp DJ, Herberstein ME, Fleishman LJ, et al (2015) An integrative framework for the

appraisal of coloration in nature. Am Nat 185:705–724.

https://doi.org/10.1086/681021

Maia R, Gruson H, Endler JA, White TE (2019) pavo 2: New tools for the spectral and spatial

analysis of colour in r. Methods Ecol Evol 0–2. https://doi.org/10.1111/2041-

210X.13174

Marr D (2010) Vision: A Computational Investigation into the Human Representation and

Processing of Visual Information. The MIT Press

Nokelainen O, Scott-Samuel NE, Nie Y, et al (2021) The giant panda is cryptic. Sci Rep 11:1–

10. https://doi.org/10.1038/s41598-021-00742-4

R Core Team (2021) R: A Language and Environment for Statistical Computing

Renoult JP, Kelber A, Schaefer HM (2017) Colour spaces in ecology and evolutionary biology.

Biol Rev 92:292–315. https://doi.org/10.1111/brv.12230

Rodríguez-Morales D, Tapia-McClung H, Robledo-Ospina LE, Rao D (2021) Colour and motion

affect a dune wasp's ability to detect its cryptic spider predators. Sci Rep 11:1–9.

https://doi.org/10.1038/s41598-021-94926-7

Schneider CA, Rasband WS, Eliceiri KW (2012) NIH Image to ImageJ: 25 years of image

analysis. Nat Methods 9:671–675. https://doi.org/10.1038/nmeth.2089

Stoddard MC, Kilner RM, Town C (2014) Pattern recognition algorithm reveals how birds

evolve individual egg pattern signatures. Nat Commun 5:1–10.

https://doi.org/10.1038/ncomms5117

Stoddard MC, Osorio D (2019) Animal Coloration Patterns: Linking Spatial Vision to

Quantitative Analysis. Am Nat 193:000–000. https://doi.org/10.1086/701300

Troscianko J, Stevens M (2015) Image calibration and analysis toolbox – a free software

suite for objectively measuring reflectance, colour and pattern. Methods Ecol Evol

6:1320–1331. https://doi.org/10.1111/2041-210X.12439

Van Belleghem SM, Papa R, Ortiz-Zuazaga H, et al (2018) Patternize : An R package for

quantifying color pattern variation. Methods Ecol Evol 9:390–398.

https://doi.org/10.1111/2041-210X.12853

van den Berg CP, Endler JA, Papinczak DEJ, Cheney KL (2022) Using colour pattern edge

contrast statistics to predict detection speed and success in triggerfish (*Rhinecanthus*

aculeatus). J Exp Biol. https://doi.org/10.1242/jeb.244677

van den Berg CP, Hollenkamp M, Mitchell LJ, et al (2020a) More than noise: Context-

dependant luminance contrast discrimination in a coral reef fish (*Rhinecanthus*

aculeatus). J Exp Biol 223:jeb.232090. https://doi.org/10.1242/jeb.232090

van den Berg CP, Troscianko J, Endler JA, et al (2020b) Quantitative Colour Pattern Analysis

(QCPA): A comprehensive framework for the analysis of colour patterns in nature.

Methods Ecol Evol 11:316–332. https://doi.org/10.1111/2041-210X.13328

van Rossum G (1995) Python tutorial, Technical Report CS-R9526. Cent voor Wiskd en

Inform

Vorobyev M, Osorio D (1998) Receptor noise as a determinant of colour thresholds. Philos

Trans R Soc B Biol Sci 265:351–358. https://doi.org/10.1098/rspb.1998.0302

White TE, Dalrymple RL, Noble DWA, et al (2015) Reproducible research in the study of

biological coloration. Anim Behav 106:51–57.

https://doi.org/10.1016/j.anbehav.2015.05.007