A systematic review of the diversity and virulence correlates of metastrongyle

lungworms in marine mammals.

Jared R. Fischbach[1] and Mauricio Seguel[2,*].

[1]College of Biological Sciences, University of Guelph, Guelph, Ontario, Canada; [2]Department of

Pathobiology, Ontario Veterinary College, University of Guelph, Ontario, Canada.

* **Author for correspondence**: Mauricio Seguel, mseguel@uoguelph.ca

Graphical Abstract:

Abstract

Metastrongyle lungworms could be particularly detrimental for diving animals such as marine mammals, however little is known of the drivers of pathogenic host-parasite relationships in this group. This systematic review analyzed the diversity of metastrongyles in marine mammals and the host and parasite traits associated with virulence. There have been at least 40 species of metastrongyles described in 66 species of marine mammals. After penalization for study biases, *Halocercus hyperoodoni*, *Otostrongylus circumlitus*, *Parafilaroides gymnurus*, *Halocercus brasiliensis*, and *Stenurus minor* were the metastrongyles with the widest host range. Most studies (n=133/166) reported that metastrongyles caused bronchopneumonia, while in the cardiovascular system metastrongyles caused vasculitis in nearly half of the studies (45.5%, 5/11) that assessed these tissues. Metastrongyles were associated with otitis in 23.1% (6/26) of the studies. Metastrongyle infection was considered a potential contributory to mortality in 44.8% (90/201) of the studies while 10.4% (n=21/201) of these studies considered metastrongyles the main cause of death. Metastrongyle species with a wider host range were more likely to induce pathogenic effects. Metastrongyles can cause significant tissue damage and mortality in marine mammals although virulent host-parasite relationships are dominated by a few metastrongyle species with wider host ranges.

Keywords: marine mammals, metastrongyles, lungworms, parasites, pathogenicity, virulence.

1. Introduction

Metastrongyloidea (Nematoda: Strongylida), also known as lungworms or metastrongyles, are

common parasites of the respiratory, cardiovascular, and rarely the auditory system of mammals

(Measures, 2001). The definitive hosts become infected through ingestion of larval stages within

invertebrate intermediate hosts or free on pasture, therefore, metastrongyle infection can be quite

common among free-foraging domestic and wild mammals (Measures, 2001; Regassa *et al.*, 2010;

Niebuhr *et al.*, 2021). Like other helminth infections, the pathogenic effects of

metastrongyloidiasis can be broad with impacts that range from subclinical infection to

bronchopneumonia and death (Measures, 2001; Forbes, 2021). The reasons for these variable

outcomes are not fully understood but site of infection (e.g. upper vs lower airways), burden and

host immune status have been indicated as drivers of pathogenic outcomes (Forbes, 2021).

However, it is unclear if certain host-parasite relationships are more detrimental for the host and

if these more virulent associations obey to ecological factors that could illuminate conserved

drivers of pathogenicity in these nematodes. Interestingly, the virulence of metastrongyles could

be higher for animal species that require optimal respiratory and auditory function for fitness, such

as marine mammals (Bergeron *et al.*, 1997b; Measures, 2001; Seibel *et al.*, 2010). Marine

mammals rely on cardiovascular and lung capacity for foraging dives and the otic apparatus of

cetaceans is critical for communication and echolocation in the water column (Bergeron *et al.*,

1997b; Measures, 2001; Seibel *et al.*, 2010). Therefore, metastrongyles parasitism of these host

compartments could be associated with disease, stranding and death (Bergeron *et al.*, 1997b;

Measures, 2001; Seibel *et al.*, 2010), making this animal group an interesting model to test

conserved correlates of virulence among metastrongyle species. Nevertheless, metastrongyles are

one of the most diagnosed parasites in marine mammals, and in many circumstances are considered

incidental findings of little to no pathologic significance (Geraci and St. Aubin, 1987). This

conflicting evidence could be related to negative metastrongyle effects limited to specific host-

parasite relationships, parasite traits, environmental conditions, or the research methods employed

to assess the health impacts. However, a systematic assessment of the factors that could modulate

the health impacts of metastrongyles has not been reported.

Three families of metastrongyles are prominently found in marine mammals, spanning

seven genera, with a considerable diversity of species within each genus. Despite this diversity,

metastrongyles share similar life histories and host exploitation strategies in marine mammals.

Most aquatic metastrongyle species require an obligate intermediate host, usually an invertebrate,

where several larval stages develop until reaching an infective stage (Dailey, 1970a, 2006a). These

infective larvae are consumed by the definitive host and then migrate from the gastrointestinal

system to the respiratory or cardiovascular system where sexual maturity and reproduction occurs,

releasing embryonated-larvated eggs or larvae (Taylor *et al.*, 2015). Additionally, in some species,

maternal lungworms infect neonates through the placenta and/or milk (Dailey *et al.*, 1991a). In

the airways and respiratory sinuses or cardiovascular system, metastrongyles feed on epithelial-

endothelial cells, host debris and occasionally blood (Dailey, 1970a; Fauquier *et al.*, 2010; Rhyan

et al., 2018a). These processes of migration through tissues, feeding and releasing eggs and larvae

illicit variable immune responses in the host (Baylis and Daubney, 1925; Fauquier *et al.*, 2010;

Groch *et al.*, 2020a). This immune response is usually directed against parasite cuticle, larvae,

eggs or parasite remains and usually consist in the attraction of macrophages, eosinophils and

lymphocytes to the site of infection (Fauquier *et al.*, 2010; Zafra *et al.*, 2015a; Seguel *et al.*, 2020a;

Groch *et al.*, 2020a). The degree of this inflammatory response and its impact on health status can

be quite variable ranging from minimal to severe bronchopneumonia (Fauquier *et al.*, 2010;

Seguel *et al.*, 2020a; Groch *et al.*, 2020a), being the drivers of the variable responses poorly

understood.

There is conflicting evidence regarding the connection between metastrongyle infections

and cetacean strandings (Arbelo *et al.*, 2013). Currently, there is no consensus on whether

inflammation in the otic apparatus, elicited by a presence of metastrongyle parasites, can impair

hearing or echolocation to a significant enough degree to be considered a contributory factor in

cetacean strandings (Geraci and St. Aubin, 1987; Wohlsein *et al.*, 2019a). Similarly, the presence

and feeding behavior of the nematodes in bronchi and lung parenchyma facilitates secondary

bacterial infections with conflicting documented outcomes, ranging from little importance for lung

physiology to fatal bronchopneumonia, especially when parasites are found in the lower

respiratory system (Siebert *et al.*, 2001; Lehnert *et al.*, 2010; Ulrich *et al.*, 2015, 2016; Reckendorf

et al., 2021). This conflicting evidence could be related to several host and parasite traits, although

across biological systems, parasite biomass, usually assessed in terms of burden, is a common

driver of negative health outcomes (Pedersen and Fenton, 2015; Seguel and Gottdenker, 2017).

Similarly, parasite and host sizes play an important role in assessing total parasite biomass and can

sometimes serve as useful proxies of biomass to identify particularly detrimental host-parasite

relationships across studies (Seguel and Gottdenker, 2017). Another potential factor impacting

health outcomes across host-parasite systems is the capacity of certain helminths to parasitize

multiple host species. Parasites with a wider host range can potentially become more virulent

because decreases in host fitness due to infection can be compensated by infecting other

susceptible species (Poulin, 2011; Schmid-Hempel, 2021). However, the role of these host and

parasite traits on the health impact of metastrongyles is not well defined.

The objectives of this systematic review were to i) summarize existing literature concerning

relationships between marine mammals and metastrongyle parasites, ii) assess the relative host

and parasite diversity of infections iii) outline the pathology and health consequences that result

from metastrongyle infections in marine mammals, with assessment of the potential drivers of

these effects.

2. Materials and Methods

2.1 Searching methods and inclusion criteria

A systematic literature review on metastrongyle parasites in marine mammals was

conducted on July 6, 2021. The search was conducted using Google Scholar and PubMed,

following the best practices outlined by Haddaway and Watson (2016) to conduct an aggregative

systematic review in the field of parasitology (Haddaway and Watson, 2016) and the

recommendations outlined in the "Preferred Reporting Items for Systematic Reviews and Meta-

Analyses" (Page *et al.*, 2021).

The search terms used for the literature search on PubMed were as follows: "((Marine) OR

(Cetacean) OR (Delphinidae) OR (Phocidae) OR (Phocoenidae) OR (Otariidae) OR

(Monodontidae) OR (Mustelidae) OR (Trichechidae) OR (Odobenidae) OR (Ursidae) OR

(Odobenidae) OR (Balaenidae) OR (Neobalaenidae) OR (Eschrichtiidae) OR (Balaenopteridae)

OR (Physeteridae) OR (Kogiidae) OR (Ziphiidae) OR (Platanistidae) OR (Iniidae) OR (Lipotidae)

OR (Pontoporiidae) OR (Dugongidae) OR (Seal) OR (Porpoise) OR (Otter) OR (Dolphin) OR

(Sea lion) OR (Whale) OR (Walrus) OR (Manatee)) AND ((Metastrongyle) OR (Metastrongylus)

OR (Metastrongyloidea) OR (Metastrongylidae) OR (Parafilaroides) OR (Stenurus) OR

(Halocercus) OR (Pseudaliidae) OR (Filaroididae) OR (Crenosomatidae) OR (Otostrongylus) OR

(Skrjabingylus))".

The search terms used for the literature search on Google Scholar were reduced due to

limitations in the advanced search software, notably a 256-character limit on searches. The search

terms used for Google Scholar were as follows: "((Metastrongyle) OR (Metastrongylus) OR

(Metastrongyloidea) OR (Metastrongylidae) OR (Parafilaroides) OR (Stenurus) OR (Halocercus)

OR (Pseudaliidae) OR (Filaroididae) OR (Crenosomatidae) OR (Otostrongylus) OR

(Skrjabingylus)) AND ((Marine) OR (Cetacean))".

The search on PubMed yielded 120 results and the search on Google Scholar yielded 2110

results, excluding citations. As Google Scholar can only display 1000 results per search, only the

1000 articles deemed most relevant by the software were manually reviewed for inclusion in the

study, in addition to the 120 articles found via PubMed. To ensure all recently published literature

was consulted, a secondary search on google scholar was conducted with the same search terms,

filtering to only include articles published from 2017 and onward. This secondary search yielded

380 results, excluding citations, which were also manually reviewed for inclusion in the study. All

conference archives (e.g. IAAAM), theses not published in peer-reviewed journals, and posters

were excluded from this study. The full texts for the articles isolated through literature search

results were obtained and manually reviewed by the two authors, being included in the study if

they met the following inclusion criteria: i) The host species was any wild marine mammal, with

captive wild animals only included if the transmission of parasites was stated to have occurred

naturally, prior to their capture. ii) The parasite species belonged to a genus within the

metastrongyle superfamily, including studies that described the presence of parasitic eggs and

larvae provided they could be positively identified to at least the genus level (studies that described

parasites as "lungworms" or "nematodes" without specificity regarding genera were excluded). iii)

The article serves as the primary source describing a relationship between a marine mammal host

and metastrongyle parasite. Duplicate articles, existing in both the Google Scholar and PubMed

searches were manually removed. All non-primary sources which were found through the initial

search terms were screened, and citations were followed, leading to the inclusion of 15 additional

articles which met the inclusion criteria. Later, the World Register of Marine Species (WoRMS,

https://www.marinespecies.org),   The   Society   for   Marine   Mammalogy

(*https://marinemammalscience.org*), and the Natural History Museum Host-Parasite database

(*https://www.nhm.ac.uk/research-curation/scientific-resources/taxonomy-systematics/host-*

parasites/) were consulted to ensure all known unique relationships were reported in this study

(WoRMS Editorial Board, 2022). A total of 35 articles could not be acquired in their full text form

for inclusion in data analysis, but the relationships they report are known through prior citations

in peer-reviewed journals or available abstracts. The articles which met the inclusion criteria were

further evaluated by the two authors, with significant attributes documented on a master

spreadsheet. Many included articles, including the articles not available in full-text form, report

multiple different host-parasite relationships, thus independent rows were created on the master

spreadsheet for each described host-parasite relationship (n=481), allowing for relationship-

specific attributes to be evaluated. The accepted common names and scientific names provided for

host species in this study are taken from the Society for Marine Mammalogy Committee on

Taxonomy (WoRMS Editorial Board, 2022).

A total of 207 full text publications met the inclusion criteria for this study, with 114

appearing only on Google Scholar, 6 appearing only on PubMed, 72 appearing on both Google

Scholar and PubMed, and 15 coming from other sources. Data was collected on all included full-

text publications, including whether they assessed pathogenic effect. An article was deemed to

assess pathogenic effect if it described some biological impact associated with a presence of

metastrongyle parasites in the host, or if it explicitly stated no pathogenic effect was associated

with the parasitism. Further data was collected on whether full-text publications which met the

inclusion criteria mentioned strandings or mortality. For the collection of this data, an article was

deemed to mention strandings if it contained the following terms (not including in the citations):

"stranded" or "stranding" or "strandings". For an article to have been deemed to mention mortality,

it must have contained the following terms (not including in the citations): "mortality" or

"mortalities". The inclusion terms for determining which articles mention stranding or mortality

were isolated through word searching in adobe acrobat, with technologically non-conforming

articles screened manually. Later, the studies mentioning mortality or stranding were assessed to

ensure they discussed these effects in the context of the study's findings. The parasite collection

methodology was also noted for each included full-text publication. Within the articles which met

the inclusion criteria, the geographical location(s) were recorded for 437 documented relationships

and the anatomical location(s) of the parasites were recorded in 371 documented relationships. We

further calculated the geographical dispersion of the 102 unique relationships between marine

mammals and metastrongyles.

For each documented relationship, data was also recorded on any reported pathogenic

effects (n=201). Pathogenic effects can be described as some biological impact on the host

associated with the presence of metastrongyle parasites. In cases where it was explicitly stated that

no pathogenic effect was associated with parasitism, this was recorded as well. Of the 101 articles

that assessed pathogenic effect, there were a total of 201 relationships in which attributes

describing a pathogenic effect or lack thereof were documented.

Between the 207 full-text publications which met our inclusion criteria, and the additional

35 publications we could not retrieve in full-text form, a total of 481 host-parasite relationships

are documented. For each of these relationships, data was collected on their respective species

taxonomy. Next, the frequency at which each metastrongyle parasite species appears in the

literature, in the context of relationships with marine mammal hosts was recorded. For the purposes

of frequency calculations, parasites described only to the genus level were excluded. The

frequency at which marine mammal hosts appear in the literature, in the context of relationships

with metastrongyle parasites was recorded in the same manner. For frequency calculations, hosts

described only to the genus level were excluded. For the purposes of all diversity calculations,

only relationships where both the host and parasite were described to the species level were

considered. Finally, host diversity was calculated as a measure of the diversity of host species a

metastrongyle species can infect, and parasite diversity was calculated as the diversity of

metastrongyle species a marine mammal species can host.

To provide a metric by which the size of a host could be used as a predictor of pathogenic

effects, host body mass was calculated by dividing the median weight in kilograms by the median

length in centimetres for each host species. Median host measurements were compiled as the

median of both genders from a variety of sources based on availability in the following order of

priority: 1) World Register of Marine Species (WoRMS, *https://www.marinespecies.org*); 2)

Society   for   Marine   Mammalogy   (*https://marinemammalscience.org/science-and-*

publications/species-information/facts/); 3) National Oceanic and Atmospheric Administration

(NOAA, *https://www.fisheries.noaa.gov/species-directory*); 4) Animal Diversity Web (ADW,

https://animaldiversity.org). In cases where data on host size was not available through those four

sources, the median measurements of hosts documented during gross necropsy in articles which

met the inclusion criteria of this study were used.

Finally, we produced an up-to-date checklist of all known relationships concerning

metastrongyles in marine mammals, which is included in table 1, complete with information on

anatomical ranges documented for each relationship. The relationships documented are found

within the 207 articles included in this study or within the 35 articles that could not be acquired in

their full text form for inclusion in data analysis, provided the relationships they report are known

through prior citations in peer-reviewed journals.

Table 1: Checklist of all known relationships concerning metastrongyles in marine mammals,

complete with information on anatomical ranges documented for each relationship.

Host Family	Host Species	Common Name	Parasite Family	Parasite species	Anatomical Location	Reference
Balaenopteridae	Megaptera novaeangliae	*Humpback Whale*	Pseudaliidae	*Pharurus alatus*	N/A	(Raga and Balbuena, 1987)
Delphinidae	Cephalorhynchus commersonii	*Commerson's Dolphin*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Greenwood and Taylor, 1979)
				Halocercus sp.	Lower respiratory	(Goodall *et al.*, 1988; Dougnac and Fredes, 2012)
				Stenurus sp.	Lower respiratory	(Duignan, 2000)
	Cephalorhynchus hectori	*Hector's dolphin*	Pseudaliidae	*Halocercus sp.*	Lower respiratory	(McKenzie and Blair, 1983; Duignan, 2000)
	Delphinus delphis	*Short-beaked common dolphin*	Pseudaliidae	*Halocercus cryptocephalus*	N/A	(Delyamure, 1968)
				Halocercus delphini	Lower respiratory	(Abollo *et al.*, 1998; Gibson *et al.*, 1998; Davison *et al.*, 2013; Pool *et al.*, 2020b, 2021)
				Halocercus invaginatus	N/A	(Raga and Balbuena, 1987)
				Halocercus kleinenbergi	N/A	(Harris, 1982)
				Halocercus lagenorhynchi	Lower respiratory	(Tomo *et al.*, 2010)
				Halocercus sp.	Lower respiratory	(Zafra *et al.*, 2015b; Lehnert *et al.*, 2017)

			Stenurus minor	N/A	(Raga, 1994)
			Stenurus ovatus	Lower respiratory	(Pool *et al.*, 2021)
			Stenurus sp.	Lower respiratory	(Díaz-Delgado *et al.*, 2012)
Delphinus sp.	*Common dolphin*	Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Stockin *et al.*, 2009)
Feresa attenuata	*Pygmy killer whale*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Guimarães *et al.*, 2015)
			Halocercus sp.	Lower respiratory	(Mignucci-Giannoni *et al.*, 1998b)
			Stenurus globicephalae	Gastrointestinal, Lower respiratory, Otic apparatus	(Forrester *et al.*, 1980; Mignucci-Giannoni *et al.*, 1998b)
Globicephala macrorhynchus	*Short-finned pilot whale*	Pseudaliidae	*Halocercus kleinenbergi*	Lower respiratory	(Carvalho *et al.*, 2010)
			Stenurus globicephalae	Gastrointestinal, Otic apparatus	(Arnold and Gaskin, 1975; Mignucci-Giannoni *et al.*, 1998b; Carvalho *et al.*, 2010)
			Stenurus minor	N/A	(Morales Vela and Olvera Gómez, 1993)
			Stenurus sp.	Otic apparatus	(Huertas and Lagueux, 2016)
			Torynurus convolutus	N/A	(Dougherty, 1943)
Globicephala melas	*Long-finned pilot whale*	Pseudaliidae	*Stenurus globicephalae*	Lower respiratory, Otic apparatus	(Cowan, 1967; Arnold and Gaskin, 1975; McManus *et al.*, 1984; Abollo *et al.*, 1998; Gibson *et al.*, 1998; Lehnert *et al.*, 2017; Pool *et al.*, 2021)
			Torynurus convolutus	Lower respiratory	(Baylis and Daubney, 1925; Baylis, 1932)
Grampus griseus	*Risso's dolphin*	Pseudaliidae	*Halocercus delphini*	Lower respiratory	(Pool *et al.*, 2021)
			Stenurus globicephalae	Gastrointestinal, Lower respiratory, Otic apparatus	(Arnold and Gaskin, 1975; Fernández *et al.*, 2003; Lehnert *et al.*, 2017; Pool *et al.*, 2021)
			Stenurus minor	N/A	(Tomilin, 1967)
Lagenodelphis hosei	*Fraser's dolphin*	Pseudaliidae	*Stenurus ovatus*	Lower respiratory	(McColl and Obendorf, 1982)
Lagenorhynchus acutus	*White sided dolphin*	Pseudaliidae	*Halocercus lagenorhynchi*	N/A	(Raga and Balbuena, 1987)
			Pseudalius inflexus	N/A	(Evans, 1991)
			Stenurus globicephalae	Gastrointestinal, Otic apparatus	(Arnold and Gaskin, 1975; Beverley-Burton, 1978; Rogan *et al.*, 1997; Schick *et al.*, 2020)
			Torynurus convolutus	N/A	(Gibson and Harris, 1979)

	Lagenorhynchus albirostris	*White-beaked dolphin*	Pseudaliidae	*Halocercus lagenorhynchi*	Lower respiratory	(Baylis and Daubney, 1925; Gibson and Harris, 1979)
	Lagenorhynchus obscurus	*Dusky dolphin*	Pseudaliidae	*Halocercus sp.*	Gastrointestinal	(von Waerebeek *et al.*, 1993)
				Stenurus sp.	Gastrointestinal	(de Castro *et al.*, 2014)
	Lissodelphis borealis	*Northern right whale dolphin*	Pseudaliidae	*Halocercus sp.*	N/A	(Moser and Rhinehart, 1993)
	Orcinus orca	*Killer whale (Orca)*	Pseudaliidae	*Halocercus sp.*	Lower respiratory	(Reckendorf *et al.*, 2018)
	Peponocephala electra	*Melon-headed whale*	Pseudaliidae	*Halocercus sp.*	N/A	(Dailey and Brownell, 1972)
				Stenurus globicephalae	Otic apparatus	(Cannon, 1977; Mignucci-Giannoni *et al.*, 1998a; b; Carvalho *et al.*, 2010)
	Pseudorca crassidens	*False killer whale*	Pseudaliidae	*Stenurus auditivus*	N/A	(Zam *et al.*, 1971)
				Stenurus globicephalae	Otic apparatus	(Zylber *et al.*, 2002)
	Sotalia fluviatilis	*Tucuxi*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Santos *et al.*, 1996; Di Beneditto and Ramos, 2004; Melo *et al.*, 2006; Rodrigues *et al.*, 2018)
	Sotalia guianensis	*Guiana dolphin*	Pseudaliidae	*Halocercus brasiliensis*	Gastrointestinal, Lower respiratory	(Marigo *et al.*, 2010; Carvalho *et al.*, 2010; Guimarães *et al.*, 2013, 2015; Domiciano *et al.*, 2016; Groch *et al.*, 2018, 2020b; Carvalho Demarque *et al.*, 2020)
				Halocercus sp.	Lower respiratory	(Carvalho *et al.*, 2010)
	Sousa chinensis	*Indo-Pacific humpback dolphin*	Pseudaliidae	*Halocercus pingi*	Lower respiratory	(Parsons and Jefferson, 2000; Parsons *et al.*, 2001)
	Sousa plumbea	*Indian Ocean humpback dolphin*	Pseudaliidae	*Halocercus sp.*	Lower respiratory	(Lane *et al.*, 2014)
	Stenella attenuata	*Pantropical spotted dolphin*	Pseudaliidae	*Halocercus delphini*	Lower respiratory	(Dailey and Perrin, 1973)
				Halocercus sp.	Lower respiratory	(Oliveira *et al.*, 2011)
	Stenella clymene	*Clymene dolphin*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Carvalho *et al.*, 2010; Guimarães *et al.*, 2015)
				Halocercus delphini	Lower respiratory, Upper respiratory	(Aguilar-Aguilar *et al.*, 2010; Hernández-Orts *et al.*, 2021)
				Halocercus sp.	Lower respiratory	(Carvalho *et al.*, 2010)
	Stenella coeruleoalba	*Striped dolphin*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Rosas *et al.*, 2002)
				Halocercus delphini	Lower respiratory	(Lehnert *et al.*, 2017; Pool *et al.*, 2020a; b, 2021; Terracciano *et al.*, 2020)

				Halocercus lagenorhynchi	Lower respiratory	(Ross and Bass, 1971; Gibson *et al.*, 1998; Oliveira *et al.*, 2011; Terracciano *et al.*, 2020)
				Halocercus sp.	Lower respiratory	(Abollo *et al.*, 1998; Carvalho *et al.*, 2010; Zafra *et al.*, 2015b; Lehnert *et al.*, 2017; Vargas-Castro *et al.*, 2020; García de los Ríos y Loshuertos *et al.*, 2021)
				Pharurus sp.	N/A	(García de los Ríos y Loshuertos *et al.*, 2021)
				Pseudalius sp.	N/A	(García de los Ríos y Loshuertos *et al.*, 2021)
				Stenurus minor	Gastrointestinal	(Aytemiz *et al.*, 2012)
				Stenurus ovatus	Lower respiratory	(Pool *et al.*, 2020a, 2021)
				Stenurus sp.	N/A	(García de los Ríos y Loshuertos *et al.*, 2021)
	Stenella frontalis	*Spotted dolphin*	Pseudaliidae	*Halocercus delphini*	N/A	(Perrin *et al.*, 1994)
				Halocercus lagenorhynchi	N/A	(Perrin *et al.*, 1987)
				Halocercus sp.	Lower respiratory	(Zafra *et al.*, 2015b)
				Pseudalius sp.	Lower respiratory	(Zafra *et al.*, 2015b)
	Stenella longirostris	*Spinner dolphin*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Carvalho *et al.*, 2010; Domiciano *et al.*, 2016)
				Halocercus delphini	Lower respiratory	(Dailey and Perrin, 1973)
				Halocercus sp.	Lower respiratory	(Carvalho *et al.*, 2010; Oliveira *et al.*, 2011)
	Steno bredanensis	*Rough-toothed dolphin*	Pseudaliidae	*Stenurus sp.*	Otic apparatus	(Ewing *et al.*, 2020)
	Tursiops aduncus	*Indo-Pacific bottlenose dolphin*	Pseudaliidae	*Halocercus lagenorhynchi*	Lower respiratory	(Tomo *et al.*, 2010)
				Halocercus sp.	Lower respiratory	(Stephens *et al.*, 2014; Lane *et al.*, 2014)
				Stenurus ovatus	Lower respiratory	(Tomo *et al.*, 2010)
	Tursiops truncatus	*Common bottlenose dolphin*	Pseudaliidae	*Halocercus cryptocephalus*	Lower respiratory	(Bowie, 1984; Fauquier *et al.*, 2009)
				Halocercus delphini	Lower respiratory	(Abollo *et al.*, 1998; Terracciano *et al.*, 2020; Pool *et al.*, 2021)
				Halocercus invaginatus	N/A	(Birkun and Krivokhizhin, 1996)

Family	Species	Common name	Parasite family	Parasite species	Organ system	References
				Halocercus lagenorhynchi	Lower respiratory	(Woodard *et al.*, 1969; Dailey *et al.*, 1991b; Lipscomb *et al.*, 1996; Mazzariol *et al.*, 2007; McFee and Lipscomb, 2009; Fauquier *et al.*, 2009)
				Halocercus sp.	Lower respiratory	(Sierra *et al.*, 2014; Seguel *et al.*, 2020b)
				Pharurus alatus	Lower respiratory	(Tomo *et al.*, 2010)
				Stenurus minor	Lower respiratory	(Birincioğlu *et al.*, 2017)
				Stenurus ovatus	Lower respiratory	(Bowie, 1984; Kumazawa *et al.*, 2006; Kuwamura *et al.*, 2007; Tomo *et al.*, 2010; Terracciano *et al.*, 2020; Pool *et al.*, 2021)
				Stenurus sp.	Lower respiratory	(Sierra *et al.*, 2014)
Iniidae	Inia geoffrensis	*Amazon river dolphin*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Rodrigues *et al.*, 2018)
				Halocercus sp.	N/A	(Dailey and Brownell, 1972)
Kogiidae	Kogia breviceps	*Pygmy sperm whale*	Pseudaliidae	*Halocercus sp.*	Lower respiratory	(Carvalho *et al.*, 2010)
Monodontidae	Delphinapterus leucas	*Beluga whale*	Pseudaliidae	*Halocercus monoceris*	Cardiovascular, Lower respiratory, Upper respiratory	(Measures *et al.*, 1995)
				Halocercus taurica	Cardiovascular, Lower respiratory, Upper respiratory	(Measures *et al.*, 1995)
				Pharurus pallasii	Central Nervous System, Lower respiratory, Otic apparatus	(Babero and Thomas, 1960; Arnold and Gaskin, 1975; Kenyon and Kenyon, 1977; Wazura *et al.*, 1986; Measures *et al.*, 1995; Houde *et al.*, 2003)
				Stenurus arctomarinus	Cardiovascular, Lower respiratory, Upper respiratory	(Arnold and Gaskin, 1975; Measures *et al.*, 1995; Burek-Huntington *et al.*, 2015)
				Stenurus minor	N/A	(Kühn, 1829)
	Monodon monoceros	*Narwhal*	Pseudaliidae	*Halocercus monoceris*	Lower respiratory	(Webster *et al.*, 1973; MacNeill *et al.*, 1975)
				Pharurus alatus	N/A	(Arnold and Gaskin, 1975)
Otariidae	Arctocephalus australis	*South american fur seal*	Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Jacobus *et al.*, 2016)
				Parafilaroides normani	Lower respiratory	(Echenique *et al.*, 2020)
	Arctocephalus forsteri	*New zealand fur seal*	Filaroididae	*Parafilaroides normani*	Lower respiratory	(Dailey, 2009)
	Arctocephalus gazella	*Antarctic fur seal*	Filaroididae	*Parafilaroides normani*	Lower respiratory	(Jacobus *et al.*, 2016)

	Arctocephalus philippii	*Guadalupe Fur seal*	Filaroididae	*Parafilaroides decorus*	Cardiovascular, Lower respiratory	(Seguel *et al.*, 2018a; Williams *et al.*, 2020)
	Arctocephalus pusillus	*Australian fur seal*	Filaroididae	*Parafilaroides normani*	Lower respiratory	(Dailey, 2009; Jabbar *et al.*, 2014)
	Arctocephalus tropicalis	*Sub-antarctic fur seal*	Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Reisfeld *et al.*, 2019)
				Parafilaroides normani	Lower respiratory	(Dailey, 2009)
	Callorhinus ursinus	*Northern fur seal*	Filaroididae	*Parafilaroides decorus*	N/A	(Gerber *et al.*, 1993)
	Eumetopias jubatus	*Stellar sea lion*	Filaroididae	*Parafilaroides decorus*	Lower respiratory	(Stroud, 1978; Stroud and Roffe, 1979)
				Parafilaroides nanus	N/A	(Dougherty and Herman, 1947)
				Parafilaroides prolificus	N/A	(Dougherty and Herman, 1947)
	Neophoca cinerea	*Australian sea lion*	Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Nicholson and Fanning, 1981)
	Otaria flavescens	*South American sea lion*	Crenosomatidae	*Otostrongylus circumlitus*	N/A	(Ebmer *et al.*, 2020)
				Otostrongylus sp.	N/A	(Hermosilla *et al.*, 2016)
			Filaroididae	*Parafilaroides sp.*	N/A	(Hermosilla *et al.*, 2016)
	Phocarctos hookeri	*New zealand sea lion*	Filaroididae	*Parafilaroides decorus*	N/A	(Duignan, 2000)
	Zalophus californianus	*Californian sea lion*	Crenosomatidae	*Otostrongylus circumlitus*	Cardiovascular, Lower respiratory	(Kelly *et al.*, 2005; Goldstein *et al.*, 2011)
				Parafilaroides sp.	Lower respiratory	(Gulland *et al.*, 2012)
			Filaroididae	*Parafilaroides decorus*	Lower respiratory	(Johnston and Ridgway, 1969; Dailey, 1970b; Fleischman and Squire, 1970; Migaki *et al.*, 1971; Sweeney and Gilmartin, 1974; Stroud, 1978; Gerber *et al.*, 1993; Haulena and Gulland, 2001; Fauquier *et al.*, 2004; Greig *et al.*, 2005; Dennison *et al.*, 2007; Evans, 2011; Kuzmina *et al.*, 2018; Williams *et al.*, 2020; Liu *et al.*, 2021)
	Zalophus wollebaeki	*Galapagos sea lion*	Crenosomatidae	*Otostrongylus sp.*	N/A	(Walden *et al.*, 2018)
			Filaroididae	*Parafilaroides sp.*	N/A	(Walden *et al.*, 2018)
Phocidae	Cystophora cristata	*Hooded seal*	Crenosomatidae	*Otostrongylus circumlitus*	Lower respiratory	(Lucas *et al.*, 2003)
			Filaroididae	*Parafilaroides gymnurus*	Lower respiratory	(Lucas *et al.*, 2003)

			Crenosomatidae	*Otostrongylus circumlitus*	N/A	(Antonelis *et al.*, 1994)
	Erignathus barbatus	*Bearded seal*	Filaroididae	*Parafilaroides sp.*	Cardiovascular, Lower respiratory	(Walden *et al.*, 2020)
				Parafilaroides gymnurus	Lower respiratory	(Gosselin *et al.*, 1998; Walden *et al.*, 2020)
	Halichoerus grypus	*Grey seal*	Crenosomatidae	*Otostrongylus circumlitus*	Cardiovascular, Lower respiratory	(Gosselin *et al.*, 1998; Ulrich *et al.*, 2015; Barnett *et al.*, 2019a, 2021)
				Otostrongylus sp.	Lower respiratory	(Baker, 1987)
			Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Baker, 1987; Barnett *et al.*, 2019a, 2021)
				Parafilaroides gymnurus	Lower respiratory	(Gosselin and Measures, 1997; Gosselin *et al.*, 1998; Ulrich *et al.*, 2015)
				Parafilaroides hispidus	Lower respiratory	(Gosselin and Measures, 1997; Gosselin *et al.*, 1998)
	Histriophoca fasciata	*Ribbon seal*	Crenosomatidae	*Otostrongylus circumlitus*	N/A	(Walden *et al.*, 2020)
				Otostrongylus sp.	N/A	(Walden *et al.*, 2020)
			Filaroididae	*Parafilaroides gymnurus*	Lower respiratory	(Walden *et al.*, 2020)
	Hydrurga leptonyx	*Leopard seal*	Filaroididae	*Parafilaroides hydrurgae*	Lower respiratory	(Mawson, 1953)
	Mirounga angustirostris	*Northern elephant seal*	Crenosomatidae	*Otostrongylus circumlitus*	Cardiovascular, Gastrointestinal, Lower respiratory	(Stroud, 1978; Stroud and Roffe, 1979; Gulland *et al.*, 1997a; Elson-Riggins *et al.*, 2001; Fauquier *et al.*, 2003; Elson-Riggins *et al.*, 2004; Colegrove *et al.*, 2005; Kaye *et al.*, 2016; Sheldon *et al.*, 2017; Kaye *et al.*, 2017; Sheldon *et al.*, 2019)
				Otostrongylus sp.	N/A	(Gerber *et al.*, 1993)
			Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Stroud, 1978; Stroud and Roffe, 1979)
				Parafilaroides decorus	N/A	(Gerber *et al.*, 1993)
				Parafilaroides measuresae	Lower respiratory	(Dailey, 2006b)
	Pagophilus groenlandicus	*Harp seal*	Crenosomatidae	*Otostrongylus circumlitus*	Lower respiratory, Upper respiratory	(Lucas *et al.*, 2003; Piché *et al.*, 2010)
			Filaroididae	*Parafilaroides gymnurus*	Lower respiratory	(Gosselin and Measures, 1997; Gosselin *et al.*, 1998; Lucas *et al.*, 2003)
	Phoca largha	*Spotted seal*	Crenosomatidae	*Otostrongylus circumlitus*	N/A	(Delyamure *et al.*, 1984; Walden *et al.*, 2020)
			Filaroididae	*Parafilaroides gymnurus*	Lower respiratory	(Delyamure *et al.*, 1984; Walden *et al.*, 2020)

	Phoca vitulina	*Harbour seal*	Crenosomatidae	*Otostrongylus circumlitus*	Cardiovascular, Lower respiratory	(Dunn and Wolke, 1976; Kennedy *et al.*, 1989; Claussen *et al.*, 1991; Borgsteede *et al.*, 1991; Gerber *et al.*, 1993; Gosselin *et al.*, 1998; Elson-Riggins *et al.*, 2001, 2004; Fouchier *et al.*, 2001; Vercruysse *et al.*, 2003; Colegrove *et al.*, 2005; McKnight *et al.*, 2005; Lehnert *et al.*, 2007, 2010; Leidenberger and Boström, 2009; Lambourn *et al.*, 2013; Osinga *et al.*, 2015; Ulrich *et al.*, 2015, 2016; Kroese *et al.*, 2018)
				Otostrongylus sp.	N/A	(Herreman *et al.*, 2011)
			Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Garner *et al.*, 1997; Prenger-Berninghoff *et al.*, 2008; Herreman *et al.*, 2011; Lambourn *et al.*, 2013; Rhyan *et al.*, 2018b)
				Parafilaroides decorus	Lower respiratory	(Greig *et al.*, 2014)
				Parafilaroides gullandae	Lower respiratory	(Dailey, 2006b)
				Parafilaroides gymnurus	Lower respiratory, Reproductive	(Schumacher *et al.*, 1990; Claussen *et al.*, 1991; Borgsteede *et al.*, 1991; Gosselin and Measures, 1997; Gosselin *et al.*, 1998; Fouchier *et al.*, 2001; Vercruysse *et al.*, 2003; Lehnert *et al.*, 2007, 2010; Osinga *et al.*, 2015; Ulrich *et al.*, 2015; Elson-Riggins *et al.*, 2020)
	Pusa caspica	*Caspian seal*	Filaroididae	*Parafilaroides caspicus*	N/A	(Kurochkin and Zablotsky, 1958)
	Pusa hispida	*Ringed seal*	Crenosomatidae	*Otostrongylus circumlitus*	Lower respiratory	(Onderka, 1989; Measures and Gosselin, 1994; Bergeron *et al.*, 1997a; b; Walden *et al.*, 2020)
			Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Walden *et al.*, 2020)
				Parafilaroides gymnurus	Gastrointestinal, Lower respiratory	(Measures and Gosselin, 1994; Gosselin and Measures, 1997; Gosselin *et al.*, 1998; Walden *et al.*, 2020)
				Parafilaroides hispidus	Lower respiratory	(Kennedy, 1986; Onderka, 1989; Gosselin and Measures, 1997; Gosselin *et al.*, 1998)
	Pusa sibirica	*Baikal seal*	Crenosomatidae	*Otostrongylus circumlitus*	Lower respiratory	(Suvorova and Prokushina, 2021)
			Filaroididae	*Parafilaroides gymnurus*	Lower respiratory	(Kontrimavichus and Delyamure, 1979; Suvorova and Prokushina, 2021)
Phocoenidae	Neophocaena asiaeorientalis	*Narrow-ridged finless porpoise*	Pseudaliidae	*Halocercus pingi*	Lower respiratory, Reproductive	(Shiozaki and Amano, 2017)
				Halocercus sp.	Lower respiratory	(Wan *et al.*, 2017; Gui *et al.*, 2018)

				Halocercus sunameri	Lower respiratory	(Shiozaki and Amano, 2017)
				Halocercus taurica	Lower respiratory	(Shiozaki and Amano, 2017)
				Pharurus asiaeorientalis	Lower respiratory, Otic apparatus	(Shiozaki and Amano, 2017)
				Pharurus sunameri	Lower respiratory, Otic apparatus	(Shiozaki and Amano, 2017)
				Stenurus nanjingensis	Otic apparatus	(Shiozaki and Amano, 2017)
	Neophocaena phocaenoides	*Indo-Pacific finless porpoise*	Crenosomatidae	*Otostrongylus circumlitus*	N/A	(Yu *et al.*, 2009)
			Pseudaliidae	*Halocercus pingi*	Lower respiratory	(Wu, 1929; Parsons and Jefferson, 2000; Parsons *et al.*, 2001)
				Halocercus sp.	Lower respiratory	(Gui *et al.*, 2018)
				Halocercus sunameri	N/A	(Yamaguti, 1951)
				Halocercus taurica	N/A	(Kuramochi *et al.*, 2000)
				Pharurus asiaeorientalis	N/A	(Petter and Pilleri, 1982)
				Pharurus sunameri	N/A	(Yamaguti, 1951)
				Stenurus auditivus	N/A	(Hsü and Hoeppli, 1933)
				Stenurus nanjingensis	N/A	(Tao, 1983)
	Phocoena dioptrica	*Spectacled porpoise*	Pseudaliidae	*Stenurus minor*	Otic apparatus	(Lehnert *et al.*, 2017)
	Phocoena phocoena	*Harbour Porpoise*	Pseudaliidae	*Halocercus invaginatus*	Cardiovascular, Lower respiratory, Upper respiratory	(Smith and Threlfall, 1973; Andersen, 1974; Arnold and Gaskin, 1975; Dailey and Stroud, 1978; Stroud and Roffe, 1979; Kastelein and Lavaleije, 1992; Moser and Rhinehart, 1993; Balbuena *et al.*, 1994; Abollo *et al.*, 1998; Gibson *et al.*, 1998; Jepson *et al.*, 2000; Rogan *et al.*, 2001; Lehnert *et al.*, 2005, 2010, 2014; Siebert *et al.*, 2006, 2020; Veryeri, 2012; Pekmezci *et al.*, 2013)
				Halocercus kirbyi	N/A	(Dougherty, 1944)
				Halocercus sp.	Lower respiratory	(Smith and Threlfall, 1973; Wunschimann *et al.*, 2001; Jauniaux *et al.*, 2002; Dagleish *et al.*, 2008; Valderrama Vasquez *et al.*, 2008; Fenton *et al.*, 2017; van Elk *et al.*, 2019)

				Halocercus taurica	Lower respiratory	(Arnold and Gaskin, 1975; Gibson *et al.*, 1998; Jepson *et al.*, 2000; Rogan *et al.*, 2001; Pekmezci *et al.*, 2013)
				Pharurus dalli	Lower respiratory, Otic apparatus	(Smith and Threlfall, 1973; Colón-Llavina *et al.*, 2019)
				Pharurus sp.	N/A	(Smith and Threlfall, 1973)
				Pseudalius inflexus	Cardiovascular, Gastrointestinal, Lower respiratory, Otic apparatus	(Andersen, 1974; Arnold and Gaskin, 1975; Kastelein and Lavaleije, 1992; Balbuena *et al.*, 1994; Brosens *et al.*, 1996; Szefer *et al.*, 1998; Gibson *et al.*, 1998; Wünschmann *et al.*, 1999; Jepson *et al.*, 2000; Siebert *et al.*, 2001, 2006, 2013, 2020; Wunschimann *et al.*, 2001; Rogan *et al.*, 2001; Jauniaux *et al.*, 2002; Lehnert *et al.*, 2005, 2010; Dawson *et al.*, 2008; Dagleish *et al.*, 2008; Valderrama Vasquez *et al.*, 2008; Davison *et al.*, 2010a; b; Seibel *et al.*, 2010; Fenton *et al.*, 2017; van Elk *et al.*, 2019; Gabel *et al.*, 2021)
				Stenurus minor	Gastrointestinal, Lower respiratory, Otic apparatus, Upper Respiratory	(Andersen, 1974; Arnold and Gaskin, 1975; Dailey and Stroud, 1978; Stroud and Roffe, 1979; Faulkner, 1995; Brosens *et al.*, 1996; Faulkner *et al.*, 1998; Gibson *et al.*, 1998; Wünschmann *et al.*, 1999; Jepson *et al.*, 2000; Siebert *et al.*, 2001, 2006, 2013, 2020; Wunschimann *et al.*, 2001; Rogan *et al.*, 2001; Jauniaux *et al.*, 2002; Kijewska, 2003; Lehnert *et al.*, 2005, 2010, 2014; Prahl *et al.*, 2008; Valderrama Vasquez *et al.*, 2008; Seibel *et al.*, 2010; Pekmezci *et al.*, 2013; Fenton *et al.*, 2017; Morell *et al.*, 2017; Wohlsein *et al.*, 2019b; van Elk *et al.*, 2019; Colón-Llavina *et al.*, 2019; Gabel *et al.*, 2021)
				Stenurus sp.	Lower respiratory	(Fenton *et al.*, 2017)
				Torynurus convolutus	Cardiovascular, Gastrointestinal, Lower respiratory, Otic apparatus, Upper respiratory	(Arnold and Gaskin, 1975; Dailey and Stroud, 1978; Kastelein and Lavaleije, 1992; Balbuena *et al.*, 1994; Brosens *et al.*, 1996; Faulkner *et al.*, 1998; Gibson *et al.*, 1998; Wünschmann *et al.*, 1999; Jepson *et al.*, 2000; Siebert *et al.*, 2001, 2006, 2013, 2020; Wunschimann *et al.*, 2001; Rogan *et al.*, 2001; Jauniaux *et al.*, 2002; Lehnert *et al.*, 2005, 2010; Valderrama Vasquez *et al.*, 2008; Davison *et al.*, 2010a; Seibel *et al.*, 2010; Fenton *et al.*, 2017; van Elk *et al.*, 2019)
	Phocoena spinipinnis	*Burmeister's porpoise*	Pseudaliidae	*Halocercus sp.*	Lower respiratory	(Reyes and van Waerebeek, 1995)

				Pseudalius inflexus	Lower respiratory	(Corcuera et al., 1995; Reyes and van Waerebeek, 1995; Alvarado-Rybak et al., 2020b)
				Stenurus australis	Lower respiratory, Otic apparatus	(Torres et al., 1994; Alvarado-Rybak et al., 2020b)
				Stenurus minor	Otic apparatus	(Corcuera et al., 1995)
				Stenurus sp.	Otic apparatus	(Reyes and van Waerebeek, 1995)
	Phocoenoides dalli	Dall's porpoise	Pseudaliidae	Halocercus dalli	Lower respiratory	(Conlogue et al., 1985)
				Halocercus kirbyi	N/A	(Dougherty, 1944)
				Halocercus sp.	Lower respiratory	(Migaki et al., 1971)
				Pharurus dalli	Otic apparatus	(Arnold and Gaskin, 1975; Dailey and Walker, 1978; Kuramochi et al., 1990)
				Pharurus sp.	N/A	(Walker, 1975)
				Stenurus minor	Lower respiratory, Otic apparatus	(Johnston and Ridgway, 1969; Migaki et al., 1971; Dailey and Walker, 1978)
				Stenurus truei	Otic apparatus	(Kuramochi et al., 1990)
				Stenurus yamagutii	Otic apparatus	(Kuramochi et al., 1990)
Physteridae	Physeter macrocephalus	Sperm whale	Pseudaliidae	Stenurus ovatus	N/A	(Raga and Balbuena, 1987)
Ziphiidae	Berardius bairdii	Baird's beaked whale	Pseudaliidae	Halocercus hyperoodoni	N/A	(Delyamure, 1968)
	Mesoplodon sp.	Unidentified whale	Pseudaliidae	Halocercus sp.	Lower respiratory	(Moser and Rhinehart, 1993)
	Mesoplodon stejnegeri	Stejneger's beaked whale	Pseudaliidae	Halocercus sp.	Lower respiratory	(Savage et al., 2021)

2.2 Data management and analyses

When assessing the relative host and parasite diversities, corrections were made for literature and intra-study biases which arose from non-uniform sampling efforts. We used a modified approach, based previously published methods (Nunn *et al.*, 2003; Ezenwa *et al.*, 2006; Seguel and Gottdenker, 2017), to create a bias penalization index for each host and parasite species. To

232 do this, we searched scientific names of each host and parasite species individually on Google

233 Scholar and the number of hits was recorded. The number of hits for each species was then

234 multiplied by the number of studies that both met our inclusion criteria and contained mentions of

235 a relationship concerning that respective species. The natural logarithm of these products was then

236 taken, and the result constitutes our penalization index for each species study bias (correction-

237 index). The resulting corrected diversity indexes were plotted using frequency plots constructed in

238 the R package "ggplo2".

239  We explored the predictors of pathogenic effects among different host parasite interactions

240 using binomial generalized linear mixed effects models (GLMMs) with the restricted maximum

241 likelihood fitted using the R package "glmmTMB" (Brooks *et al.*, 2017). Although this method

242 can estimate positively biased odds ratios for binary outcomes in metadata, these are usually more

243 problematic for smaller samples sizes and alternative methods for the analysis of binary metadata

244 present similar issues (Stijnen *et al.*, 2010; Bakbergenuly and Kulinskaya, 2018). We used

245 corrected host diversity, corrected parasite diversity, and host body size as predictors for the

246 presence or absence of pathogenic effects. Study ID was added as a random effect to account for

247 studies documenting multiple host-parasite relationships with their respective outcomes. For each

248 documented host-parasite relationship, with both the host and parasite specified to the species level

249 and for which pathogenic effect was assessed (n=165), binomial GLMMs for the presence or

250 absence of pathogenic effect were fit. A second model was fit to exclusively assess the presence

251 or absence of respiratory specific effect (bronchopneumonia or brochitis). For the respiratory

252 specific effect model, each documented host-parasite relationship was included if the host and

253 parasite were specified to the species level, pathogenic effect was assessed, and the presence of

254 the parasite was documented in either the upper or lower respiratory system (n=134). We attempted

to fit cardiovascular specific effect (DIC, thrombosis, vasculitis, hemorrhage, endocarditis) (n=11)

and otic apparatus specific effect (sinusitis, otitis, otic metaplasia) (n=24) models using the same

method as for respiratory specific effect but were unable because the small sample size affected

model convergence. In both the full and respiratory specific models, we included interaction

effects between the corrected host diversity, corrected parasite diversity, and body size predictors

terms to account for individual permissiveness of host species to parasite species as well as

individual pervasiveness of parasite species to host species.

2.3 Discrepancies in the literature

Hermosilla, C., et al. (2015) reports *Stenurus sp.* in sei whale (*Balaenoptera borealis*), blue

whale (*Balaenoptera musculus*), and fin whale (*Balaenoptera physalus*) as non-primary findings,

however we could not identify a primary source of these relationships (Hermosilla *et al.*, 2015).

Lehnert, K., et al. (2019) reports *Pharurus alatus* as a parasite of short-beaked common dolphin

(*Delphinus delphis*) citing Tomo, I., et al. (2010) (Tomo *et al.*, 2010; Lehnert *et al.*, 2019). Upon

reviewing Tomo, I., et al. (2010), it does not appear to include this relationship (Tomo *et al.*, 2010).

The World Register of Marine Species (WoRMS) documents *Halocercus taurica* in of short-

beaked common dolphin (*Delphinus delphis*) and *Halocercus hyperoodoni* in northern bottlenose

whale (*Hyperoodon ampullatus*), however primary sources could not be identified for either of

these relationships (WoRMS Editorial Board, 2022). Measures, L.N. (2001) documents

Halocercus pingi in dall's porpoise (*Phocoenoides dalli*) as a non-primary finding, however we

could not find a primary source for this relationship (Measures, 2001). Three (n=3) additional

publications document relationships that were not specified to the genus level (Giorda *et al.*, 2017;

Reckendorf *et al.*, 2021; van Wijngaarden *et al.*, 2021).

3. Results

At least 40 species of metastrongyles have been described in 66 marine mammal host species.

Most metastrongyle species parasitize multiple host species, with a total of 213 unique host-

metastrongyle relationships recorded (table 1).

3.1 Characterization of studies

The mean publication year of the 207 articles which met the inclusion criteria was 2003,

with the distribution of publication years skewed towards the most recent decade (Figure 1). For

each of the articles which met the inclusion criteria, 48.8% (101/207) assessed pathogenic effect.

Seventy eight percent of studies (163/207) mentioned stranding and 43.5% (90/207) of articles

mentioned mortality as potentially related to lungworm infection. Gross necropsy is the

predominant parasite collection method for metastrongyles in marine mammals with 92.27%

(191/207) of the studies using this as the only method of collection. The next most common method

of collection was, fecal sampling, in 6.2% (13/207) of the studies (Figure 2).

Figure 1: Distribution of publication year for articles which met the inclusion criteria of this

study. The mean year of publication was 2003.

Figure 2: Parasite collection methodologies documented, with intersections between the

methodologies shown to represent studies that utilize more than one methodology. Articles that

did not report a collection methodology are designated as "Not Reported".

A total of 175 host-metastrongyle relationships were reported in North America (including

the commonly referenced Central America region), and 169 relationships were reported in Europe.

North America and Europe are the most common locations reported for metastrongyle infections

in marine mammals, followed by 54 relationships in South America, 24 in Asia, 23 in Oceania and

11 in Africa (Figure 3). Exactly half of all unique relationships (n=102) are reported in North

America (n=51, 50.00%), with 30 (29.41%) of these relationships being reported exclusively in

North America (Figure 4).

Figure 3: Geographical location(s), denoted by continent, of documented relationships between

metastrongyles and marine mammals. Documented relationships include duplicates of unique

relationships published independently. Intersections are shown for studies which report

individual host species to parasite species relationships occurring in multiple locations.

Figure 4: Geographical location(s), denoted by continent, of all unique relationships between metastrongyles and marine mammals. Intersections are shown for studies which report individual host species to parasite species relationships occurring in multiple locations.

3.2 Frequencies and diversities of metastrongyle infections in marine mammals

Pseudaliidae was the most reported metastrongyle family in marine mammals (n=331), followed by Filaroididae (n=90), and Crenosomatidae (n=58). At the host family level, Delphinidae (n=159), Phocoenidae (n=150), and Phocidae (n=107) are most frequently parasitized

by metastrongyles. Further, metastrongyles from the Pseudaliidae family are reported in hosts from

the Delphinidae (n=158), Phocoenidae (n=148), Monodontidae (n=15), Ziphiidae (n=3), Iniidae

(n=2), Balaenopteridae (n=1), Kogiidae (n=1), and Physeteridae (n=1) families. Metastrongyles

from the Crenosomatidae family are reported to parasitize hosts from the Phocidae (n=52),

Otariidae (n=5), and Phocoenidae (n=1) families. Metastrongyles from the Filaroididae family are

reported to parasitize hosts from the Phocidae (n=54), Otariidae (n=35), and Delphinidae (n=1)

families.

Otostrongylus circumlitus (n=52) is the most reported metastrongyle species followed by

Stenurus minor (n=42) and *Pseudalius inflexus* (n=31) (Supplementary Figure 1). The harbour

porpoise (*Phocoena phocoena*) (n=118) is the most reported host, followed by harbour seal (*Phoca

vitulina*) (n=41) and bottlenose dolphin (*Tursiops truncat*us) (n=23) (Supplementary Figure 2).

Before penalizing for literature and intra-study biases, the host species harbouring the largest

number of metastrongyle species (parasite diversity) were: indo-pacific finless porpoise

(*Neophocaena phocaenoides*) (n=8), short-beaked common dolphin (*Delphinus delphis*) (n=7),

harbour porpoise (*Phocoena phocoena*) (n=7), and bottlenose dolphin (*Tursiops truncatus*) (n=7)

(Supplementary Figure 3). After correction for publication bias, indo-pacific finless porpoise

(*Neophocaena phocaenoides*) remained the most permissive host (Figure 5.). Before penalization,

the metastrongyle species with the broader host diversity were: *Otostrongylus circumlitus* (n=13),

*Stenurus mino*r (n=10), *Parafilaroides gymnurus* (n=9), *Halocercus brasiliensis* (n=8), and

Halocercus delphini (n=8) (Supplementary Figure 4). After correction for publication bias,

Halocercus hyperdooni emerged as the most generalist lungworm species, followed closely by

Otostrongylus circumlitus and *Parafilaroides gymnurus* (Figure 6.).

Figure 5: Parasite diversity for metastrongyles in marine mammals, corrected for publication bias (correction-index).

Figure 6: Host diversity for metastrongyles in marine mammals, corrected for publication bias (correction-index).

3.3 The impact of metastrongyle infections on marine mammals

Of the 481 host parasite relationships documented, 371 reported an anatomical location. Lungworms were found in the lower respiratory system in 82.48% (306/371) of cases, while parasites were found in the otic apparatus in 17.52% (65/371)of cases. In 11.59% of cases

(43/371), a single species of parasite parasitized an individual host in multiple anatomical locations

(Figure 7).

Figure 7: Anatomical location(s) of documented relationships between metastrongyles and

marine mammals. Intersections are shown for studies which report individual host species to

parasite species relationships occurring in multiple locations. Relationships that did not report an

anatomical location are designated as "Not Reported". (CNS= Central nervous system, G.I=

Gastrointestinal tract, C.V= Cardiovascular system).

Out of the 201 unique host-parasite relationships assessed, 44.77% (90/201) mentioned

mortality as "possible" or "potential" contributory cause of death and only 10.44% (21/201)

studies considered metastrongyle infection as the primary cause of death. Of the 201 host-parasite

relationships for which pathogenic effect was assessed, bronchopneumonia was described in

71.64% (144/201) of the relationships, indicating a high prevalence of bronchopneumonia in hosts

infected with metastrongyles (Figure 8). Other reported pathogenic effects included edema

(10.95%, n=22/201), vasculitis (7.46%, n=15/201), thrombosis (5.97%, n=12/201), and

hemorrhage (5.97%, n=12/201). Lungworm parasitism resulted in no discernible pathogenic effect

on the host in 16.42% (33/201) of the relationships.

Figure 8: Pathogenic effect(s) of documented relationships between metastrongyles and marine

mammals. Intersections are shown for studies which report individual host species to parasite

species relationships causing multiple pathogenic effects. Relationships were designated as "No.

P.E." when pathogenic effect was assessed and was determined to be negligible. (DIC=

Disseminated Intravascular Coagulation).

Pathogenic effect was assessed for 166 host-parasite relationships in the respiratory

system, with respiratory specific pathology reported in 80.12% (133/166) of these cases. In

contrast, 26 host-parasite relationships assessed pathogenic effect in the otic apparatus, with otic

specific lesions reported in only 23.08% (6/26) of these relationships.

In the binomial GLMM that assessed pathogenic effect, host body size, corrected host

diversity and corrected parasite diversity were all significant predictors of the presence of

metastrongyle pathogenic effect in marine mammals, with a significant effect of the interaction

between host and parasite diversity indexes (table 2). For respiratory specific effects, corrected

parasite diversity and corrected host diversity main effects and their interaction were significant

predictors of metastrongyle respiratory specific effect in marine mammals, but host body size was

not a significant predictor of respiratory specific effect (table 3).

Table 2: Binomial generalized linear mixed effects model (GLMM) for presence of pathogenic

effect. Presented are predictors estimates, standard error (SE), statistic (Z) and P values. The

model used study ID as a random effect (total observations n=165, study ID n=89).

Predictor	Estimate	SE	Z	P
(Intercept)	-189.93	65.03	-2.921	0.00349
Corrected Parasite Diversity	523.62	165.18	3.170	0.00152
Corrected Host Diversity	232.05	88.44	2.624	0.00869
Host Body Size	166.90	83.04	2.010	0.04444
Corrected Parasite Diversity : Corrected Host Diversity	-625.90	250.44	-2.499	0.01245
Corrected Parasite Diversity : Host Body Size	-435.38	220.87	-1.971	0.04870
Corrected Host Diversity : Host Body Size	-189.10	131.22	-1.441	0.14953
Corrected Parasite Diversity : Corrected Host Diversity : Host Body Size	581.44	438.86	1.325	0.18521

402　Table 3: Binomial generalized linear mixed effects model (GLMM) for respiratory pathogenic

403　effects. Presented are predictors estimates, standard error (SE), statistic (Z) and P values. The

404　model used study ID as a random effect (total observations n=134, study ID n=76).

Predictor	Estimate	SE	Z	P
(Intercept)	0.09403	12.65860	0.007	0.99407
Corrected Parasite Diversity	88.93332	41.43432	2.146	0.03184
Corrected Host Diversity	74.10991	26.73963	2.772	0.00558
Host Body Size	-18.39291	13.61594	-1.351	0.17675
Corrected Parasite Diversity : Corrected Host Diversity	-244.76967	87.16180	-2.808	0.00498
Corrected Parasite Diversity : Host Body Size	8.03858	38.70274	0.208	0.83546
Corrected Host Diversity : Host Body Size	-29.25975	24.54618	-1.192	0.23325
Corrected Parasite Diversity : Corrected Host Diversity : Host Body Size	154.98741	93.04556	1.666	0.09577

4. Discussion and Conclusions

Metastrongyles are common helminth parasites of the respiratory and cardiovascular system

of mammals. Given the importance of these body systems for the survival of diving animals, it

was expected that the health impacts of metastrongyles would be particularly significant in marine

mammals. Our findings confirm that pathogenic effects, mainly bronchopneumonia, are common

in marine mammals infected with metastrongyles, although association of metastrongyle parasites

and mortality is usually not definitive. Importantly, our review suggests that pathogenic host-

parasite relationships are associated to hosts that can harbor a wider range of metastrongyle species

(parasite diversity). From the parasite perspective, metastrongyle species with a broader host

diversity tend to be more pathogenic. These results emphasize the importance of host and parasite

diversity with regards to the virulence of metastrongyles in marine ecosystems.

Most parasitology studies in marine mammals have been conducted through the opportunistic

collection and examination of stranded carcasses. This probably explains in part why in most cases

metastrongyle infections are suggested or considered as a contributory cause of death but rarely

confirmed as the primary cause of mortality. Determining cause of death in stranded marine

mammals is challenging, amid the rapid rate of postmortem changes in these species and little

background information for most cases (Arbelo *et al.*, 2013; Alvarado-Rybak *et al.*, 2020a).

Additionally, access to carcasses is sometimes difficult (*e.g.* tides), limiting proper dissection

techniques and tissue sampling. As with other host-parasite systems, the use of longitudinal

assessments and experimental manipulation of infection could give new perspectives on the impact

of metastrongyles in marine mammal health. Recently, the sampling of living species through the

collection of feces and body fluids has been implemented in cetaceans (Williams *et al.*, 2020).

Further, the development of molecular techniques such as major sperm protein (MSP) ELISAs to

test for serum antibodies to metastrongyles aid in illuminating metastrongyle seroprevalence

(Ulrich *et al.*, 2015, 2016). The continued use of approaches other than opportunistic necropsies

may substantially contribute to the understanding of the drivers and consequences of

metastrongyle infection in marine mammals.

As with many wildlife parasites (Haddaway and Watson, 2016; Seguel and Gottdenker, 2017),

most studies on metastrongyles in marine mammals are conducted in North America and Europe,

leading to overrepresentation of North American and European marine mammal and metastrongyle

species in the literature. Therefore, cosmopolitan coastal marine mammals in these regions such

as the harbor porpoise and the harbor seal along with their parasites were among the most assessed

species. Despite our intended penalization for these study biases, our results should be interpreted

with caution since more studies from different regions of the world and in cryptic and/or pelagic

marine mammal species could influence inferences on the role of host and parasite factors driving

pathogenic effects. For instance, PCBs and other persistent organic pollutants (POPs) are found in

higher concentrations in marine mammals in more industrialized regions of North America and

Europe (Fair *et al.*, 2010; Jepson *et al.*, 2016; Durante *et al.*, 2016; Seguel *et al.*, 2020a). These

immunosuppressive pollutants are thought to increase susceptibility to metastrongyles, facilitating

their pathogenic effects (Jepson *et al.*, 2016). Therefore, more studies from regions of the world

with lower POP concentrations or where new or underrepresented host-parasite relationships occur

could modify our understanding of the health impacts of metastrongyles in marine mammals.

As in domestic and land mammals, bronchopneumonia is the most common pathogenic

outcome of lungworm infection in marine mammals. The presence of lungworms in airways or

pulmonary parenchyma causes tissue damage, eliciting an inflammatory response that also

facilitates colonization by bacteria (Measures, 2001). Therefore, in most of the assessed studies,

verminous bronchopneumonia was usually complicated by secondary bacterial infection. The

impact of metastrongyles in other tissues such as blood vessels, heart or otic apparatus is probably

more direct, since in these tissues bacteria colonization secondary to helminth infection is rare. In

the cardiovascular system, the impact of lungworm infection can be serious with vasculitis,

localized or systemic thrombosis and coagulopathy (Gulland *et al.*, 1997b; Seguel *et al.*, 2018b;

Barnett *et al.*, 2019b). In these cases, there is clear link between metastrongyle infection and

mortality. Interestingly, invasion of the cardiovascular system is usually associated to

metastrongyle species that are not the most common definitive host for that parasite species,

highlighting the role of host specificity in the most pathogenic interactions between metastrongyles

and marine mammals.

Parasite and host diversity increase the chances for a for a pathogenic outcome in

metastrongyle-marine mammal relationships. This finding is supported by general hypotheses on

parasite virulence that suggests that a parasite species with a greater diversity of potential hosts

can evolve to be more pathogenic, as the sustainability of the parasite population is less dependent

on the survival of individual hosts (Poulin, 2011; Schmid-Hempel, 2021). Similarly, a host species

that harbors a greater diversity of parasites may see a greater propensity for pathogenic outcomes

from individual parasite species due to an increase in the stress on the immune and respiratory

systems resulting from co-infections (Seguel *et al.*, 2023). Our results also suggest that larger hosts

are more likely to sustain pathogenic outcomes as a result of metastrongyle parasitism. Since

marine metastrongyles are transmitted through intermediate hosts that represent food items for

marine mammals (Dailey, 1970a), this result could indicate that increased food consumption leads

to a greater larvae intake and subsequently a higher parasite burden. Similarly, a larger host body

size could correlate with access to different prey items, contributing to a greater diversity of

metastrongyle infections.

A comprehensive understanding of the drivers of the negative physiological effects

associated with metastrongyle parasitism in marine mammal hosts is challenged by a presumably

strong bias towards reporting only positive findings. The available literature clearly elucidates a

multitude of host-parasite relationships in which metastrongyles produce a definable pathological

effect, howbeit it is inadequate in clarifying why these effects are observed in some species or

populations and not in others. However, general host and parasite traits associated with pathogenic

outcomes are like those of other helminth and microbial parasites, including host permissiveness

of infection by multiple species and parasite capacity to infect multiple host species. In this context,

the rapidly changing marine environment with generalized loss of biomass could favor parasites

and hosts with a diversified trophic network, potentially selecting for more pathogenic

relationships between metastrongyles and their marine mammal hosts.

5. Supplementary Data

Supplementary figures are submitted as electronic supplementary files. Review protocol not

registered.

6. Financial Support

This work was supported by the University of Guelph President's Research Assistantship, which

was awarded to J.F. as a component of his University of Guelph President's Scholarship funding.

The Author's declare no competing interest.

7. References

Abollo, E., López, A., Gestal, C., Benavente, P. and Pascual, S. (1998). Macroparasites in

cetaceans stranded on the northwestern Spanish Atlantic coast. *Diseases of Aquatic*

Organisms **32**, 227–231. doi: 10.3354/dao032227.

Aguilar-Aguilar, R., Delgado-Estrella, A. and Moreno-Navarrete, R. (2010). New host report

for nematodes in a stranded short-snouted spinner dolphin Stenella clymene (Cetacea:

Delphinidae) from the Mexican Caribbean coast. *Helminthologia* **47**, 136–138. doi:

10.2478/s11687-010-0020-0.

Alvarado-Rybak, M., Toro, F., Abarca, P., Paredes, E., Español-Jiménez, S. and Seguel, M.

(2020a). Pathological Findings in Cetaceans Sporadically Stranded Along the Chilean

Coast. *Frontiers in Marine Science* **7**, 1–10. doi: 10.3389/fmars.2020.00684.

Alvarado-Rybak, M., Toro, F., Abarca, P., Paredes, E., Español-Jiménez, S. and Seguel, M.

(2020b). Pathological Findings in Cetaceans Sporadically Stranded Along the Chilean

Coast. *Frontiers in Marine Science* **7**, 684. doi: 10.3389/fmars.2020.00684.

Andersen, S. H. (1974). A Typical Case History of the Net-Caught Harbour Porpose. *Aquatic*

Mammals **2**, 1–6.

Antonelis, G. A., Melin, S. R. and Bukhtiyarov, Y. A. (1994). Early Spring Feeding Habits of

Bearded Seals (Erignathus Barbatus) in the Central Bering Sea, 1981. *ARCTIC* **47**, 74–

79. doi: 10.14430/arctic1274.

Arbelo, M., De Los Monteros, A. E., Herráez, P., Andrada, M., Sierra, E., Rodríguez, F.,

Jepson, P. D. and Fernández, A. (2013). Pathology and causes of death of stranded

cetaceans in the canary Islands (1999-2005). *Diseases of Aquatic Organisms* **103**, 87–99.

doi: 10.3354/dao02558.

Arnold, P. W. and Gaskin, D. E. (1975). Lungworms (Metastrongyloidea: Pseudaliidae) of

harbor porpoise Phocoena phocoena (L. 1758). *Canadian Journal of Zoology* **53**, 713–

735. doi: 10.1139/z75-087.

Aytemiz, I., Dede, A., Danyer, E. and Tonay, A. M. (2012). Morphological identification of

parasites found in the stomach contents of bycaught striped dolphins (Stenella

coeruleoalba) from Turkish Eastern Mediterranean Sea coast. *Journal of the Black Sea /*

Mediterranean Environment **18**, 238–245.

Babero, B. B. and Thomas, L. J. (1960). A Record of Pharurus oserkaiae (Skrjabin, 1942) in an

Alaskan Whale. *The Journal of Parasitology* **46**, 726. doi: 10.2307/3275519.

Bakbergenuly, I. and Kulinskaya, E. (2018). Meta-analysis of binary outcomes via generalized

linear mixed models: a simulation study. *BMC Medical Research Methodology* **18**, 70.

doi: 10.1186/s12874-018-0531-9.

Baker, J. R. (1987). Causes of mortality and morbidity in wild juvenile and adult grey seals

(Halichoerus grypus). *British Veterinary Journal* **143**, 203–220. doi: 10.1016/0007-

1935(87)90083-2.

Balbuena, J. A., Aspholm, P. E., Andersen, K. I. and Bjørge, A. (1994). Lung-worms

(Nematoda: Pseudaliidae) of harbour porpoises (*Phocoena phocoena*) in Norwegian

waters: patterns of colonization. *Parasitology* **108**, 343–349. doi:

10.1017/S0031182000076186.

Barnett, J. E. F., Bexton, S., Fraija-Fernández, N., Chooneea, D. and Wessels, M. E.

(2019a). Novel Pulmonary Vasculitis with Splendore–Hoeppli Reaction in Grey Seals

(Halichoerus grypus) Associated with Otostrongylus circumlitus Infection. *Journal of*

Comparative Pathology **173**, 83–91. doi: 10.1016/j.jcpa.2019.10.009.

Barnett, J. E. F., Bexton, S., Fraija-Fernández, N., Chooneea, D. and Wessels, M. E.

(2019b). Novel Pulmonary Vasculitis with Splendore–Hoeppli Reaction in Grey Seals

(Halichoerus grypus) Associated with Otostrongylus circumlitus Infection. *Journal of*

Comparative Pathology **173**, 83–91. doi: 10.1016/j.jcpa.2019.10.009.

Barnett, J., Allen, R., Astley, K., Whitehouse, F. and Wessels, M. (2021). Pathology of grey

seals Halichoerus grypus in southwest England including pups in early rehabilitation.

Diseases of Aquatic Organisms **145**, 35–50. doi: 10.3354/dao03600.

Baylis, H. A. (1932). A list of worms parasitic in Cetacea. *Discovery Reports* **6**, 393–418.

Baylis, H. A. and Daubney, R. (1925). A Revision of the Lung-Worms of Cetacea.

Parasitology **17**, 201–215. doi: 10.1017/S0031182000004595.

Bergeron, E., Huot, J. and Measures, L. N. (1997a). Experimental transmission of

Otostrongylus circumlitus (Railliet, 1899) (Metastrongyloidea: Crenosomatidae), a

lungworm of seals in eastern arctic Canada. *Canadian Journal of Zoology* **75**, 1364–

1371. doi: 10.1139/z97-762.

Bergeron, E., Measures, L. N. and Huot, J. (1997b). Lungworm (Otostrongylus circumlitus)

infections in ringed seals (Phoca hispida) from eastern Arctic Canada. *Canadian Journal*

of Fisheries and Aquatic Sciences **54**, 2443–2448. doi: 10.1139/f97-153.

Beverley-Burton, M. (1978). Helminths of the Alimentary Tract from a Stranded Herd of the

Atlantic White-Sided Dolphin, *Lagenorhynchus acutus*. *Journal of the Fisheries*

Research Board of Canada **35**, 1356–1359. doi: 10.1139/f78-211.

Birincioğlu, S. S., Aypak, S., Avci, H., Birincioğlu, B., İPek, E. and Akkoç, A. N. (2017).

Şişe Burunlu Bir Yunusta (Tursiops truncatus) Patolojik ve Parazitolojik İncelemeler.

Kafkas Universitesi Veteriner Fakultesi Dergisi. doi: 10.9775/kvfd.2017.18016.

Birkun, A. A. Jr. and Krivokhizhin, S. V. (1996). Present state and causes of the Black Sea

cetacean populations suppression. Parts I and II. *Vestnik Zoologii* **3**, 36–42.

Borgsteede, F. H. M., Bus, H. G. J., Verplanke, J. A. W. and van Burg, W. P. J. (1991).

Endoparasitic helminths of the harbour seal, Phoca vitulina, in the Netherlands.

Netherlands Journal of Sea Research **28**, 247–250. doi: 10.1016/0077-7579(91)90022-S.

Bowie, J. Y. (1984). Parasites from an Atlantic bottle-nose dolphin (*Tursiops truncatus*), and a

revised checklist of parasites of this host. *New Zealand Journal of Zoology* **11**, 395–398.

doi: 10.1080/03014223.1984.10428253.

Brooks, M. E., Kristensen, K., van Benthem, K. J., Magnusson, A., Berg, C. W., Nielsen, A.,

Skaug, H. J., Mächler, M. and Bolker, B. M. (2017). glmmTMB balances speed and

flexibility among packages for zero-inflated generalized linear mixed modeling. *R*

Journal **9**, 378–400. doi: 10.32614/rj-2017-066.

Brosens, L., Jauniaux, T., Siebert, U., Benke, H. and Coignoul, F. (1996). Observations on

the helminths of harbour porpoises (Phocoena phocoena) and common guillemots (Uria

aalge) from the Belgian and German coasts. *Veterinary Record* **139**, 254–257. doi:

10.1136/vr.139.11.254.

Burek-Huntington, K., Dushane, J., Goertz, C., Measures, L., Romero, C. and Raverty, S.

(2015). Morbidity and mortality in stranded Cook Inlet beluga whales Delphinapterus

leucas. *Diseases of Aquatic Organisms* **114**, 45–60. doi: 10.3354/dao02839.

Cannon, L. R. G. (1977). Some aspects of the biology of Peponocephala electra (Cetacea:

Delphinidae). II. Parasites. *Marine and Freshwater Research* **28**, 717–722.

Carvalho, V. L., Bevilaqua, C. M. L., Iñiguez, A. M., Mathews-Cascon, H., Ribeiro, F. B.,

Pessoa, L. M. B., de Meirelles, A. C. O., Borges, J. C. G., Marigo, J., Soares, L. and

de Lima Silva, F. J. (2010). Metazoan parasites of cetaceans off the northeastern coast of

Brazil. *Veterinary Parasitology* **173**, 116–122. doi: 10.1016/j.vetpar.2010.06.023.

Carvalho Demarque, I. de O., Rodrigues de Oliveira, F. C., da Silveira, L. S., Barbosa, L.

A. and Ederli, N. B. (2020). The Lungworm, Halocercus brasiliensis (Nematoda:

Pseudaliidae), from Guiana Dolphins Sotalia guianensis from Brazil with Pathological

Findings. *Journal of Parasitology* **106**, 254. doi: 10.1645/19-77.

Claussen, D., Strauss, V., Ising, S., Jäger, M., Schnieder, T. and Stoye, M. (1991). The

Helminth Fauna from the Common Seal (Phoca vitulina vitulina, Linné, 1758) of the

Wadden Sea in Lower Saxony*: Part 2: Nematodes. *Journal of Veterinary Medicine,*

Series B **38**, 649–656. doi: 10.1111/j.1439-0450.1991.tb00924.x.

Colegrove, K. M., Greig, D. J. and Gulland, F. M. D. (2005). Causes of Live Strandings of

Northern Elephant Seals (Mirounga angustirostris) and Pacific Harbor Seals (Phoca

vitulina) Along the Central California Coast, 1992-2001. *Aquatic Mammals* **31**, 1–10.

doi: 10.1578/AM.31.1.2005.1.

Colón-Llavina, M. M., Mattiucci, S., Nascetti, G., Harvey, J. T., Williams, E. H. and

Mignucci-Giannoni, A. A. (2019). Some Metazoan Parasites from Marine Mammals

Stranded in California. *Pacific Science* **73**, 461. doi: 10.2984/73.4.3.

Conlogue, G. J., Ogden, J. A. and Foreyt, W. J. (1985). Parasites of the Dall's porpoise

(Phocoenoides dalli True). *Journal of Wildlife Diseases* **21**, 160–166. doi: 10.7589/0090-

3558-21.2.160.

Corcuera, J., Monzon, F., Anguilar, A., Borrell, A. and Raga, J. A. (1995). Life history data,

organochlorine pollutants and parasites from eight Burmeister's porpoises, Phocoena

spinipinnis, caught in northern Argentine waters. *Report of the International Whaling*

Commission.

Cowan, D. F. (1967). Helminth Parasites of the Pilot Whale Globicephala melaena (Traill 1809).

The Journal of Parasitology **53**, 166. doi: 10.2307/3276641.

Dagleish, M. P., Barley, J., Finlayson, J., Reid, R. J. and Foster, G. (2008). Brucella ceti

Associated Pathology in the Testicle of a Harbour Porpoise (Phocoena phocoena).

Journal of Comparative Pathology **139**, 54–59. doi: 10.1016/j.jcpa.2008.03.004.

Dailey, M. D. (1970a). The Transmission of Parafilaroides decorus (Nematoda :

Metastrongyloidea) in the California Sea Lion (Zalophus calif ornianus) 1. *Proc. Helm.*

Soc. Wash. **37**, 215–222.

Dailey, M. D. (1970b). The Transmission of Parafilaroides decorus (Nematoda:

Metastrongyloidea) in the California Sea Lion. *Proceedings of the Helminthological*

Society of Washington **37**, 215–222.

Dailey, M. D. (2006a). Restoration of Parafilaroides (Dougherty, 1946) (Nematoda:

Metastrongyloidea) with description of two new species from pinnipeds of eastern central

Pacific. *The Journal of Parasitology* **92**, 589–594. doi: 10.1645/GE-3525.1.

Dailey, M. D. (2006b). Restoration of Parafilaroides (Dougherty, 1946) (Nematoda:

Metastrongyloidea) with Description of Two New Species from Pinnipeds of Eastern

Central Pacific. *Journal of Parasitology* **92**, 589–594. doi: 10.1645/GE-3525.1.

Dailey, M. D. (2009). A New Species of Parafilaroides (Nematoda: Filaroididae) in Three

Species of Fur Seals (Carnivora: Otariidae) From the Southern Hemisphere. *Journal of*

Parasitology **95**, 156–159. doi: 10.1645/GE-1521.1.

Dailey, M. D. and Brownell, R. L. Jr. (1972). A checklist of marine mammal parasites. In

Mammals of the sea: Biology and medicine, SH Ridgway (ed.). Charles C Thomas,

Springfield, Illinois, pp. 528–589.

Dailey, M. D. and Perrin, W. F. (1973). Helminth parasities of porpoises of the genus Stenella

in the eastern tropical pacific, with descriptions of two new species: Mastigonema

stenellae gen. et. sp. n. (Nematoda: spiruroidea) and Zalophotrema pacificum sp. n.

(Trematoda: digenea). *Fishery Bulletin* **71**, 455–471.

Dailey, M. and Stroud, R. (1978). Parasites and associated pathology observed in cetaceans

stranded along the Oregon coast. *Journal of Wildlife Diseases* **14**, 503–511. doi:

10.7589/0090-3558-14.4.503.

Dailey, M. D. and Walker, W. A. (1978). Parasitism as a Factor (?) in Single Strandings of

Southern California Cetaceans. *The Journal of Parasitology* **64**, 593. doi:

10.2307/3279939.

Dailey, M., Walsh, M., Odell, D. and Campbell, T. (1991a). Evidence of Prenatal Infection in

the Bottlenose Dolphin (Tursiops truncates) with the Lungworm Halocercus

lagenorhynchi (Nematoda: Pseudaliidae). *Journal of Wildlife Diseases* **27**, 164–165. doi:

10.7589/0090-3558-27.1.164.

Dailey, M., Walsh, M., Odell, D. and Campbell, T. (1991b). Evidence of prenatal infection in

the bottlenose dolphin (Tursiops truncatus) with the lungworm Halocercus lagenorhynchi

(Nematoda: Pseudaliidae). *Journal of Wildlife Diseases* **27**, 164–165. doi: 10.7589/0090-

3558-27.1.164.

Davison, N. J., Simpson, V. R., Chappell, S., Monies, R. J., Stubberfield, E. J., Koylass, M.,

Quinney, S., Deaville, R., Whatmore, A. M. and Jepson, P. D. (2010a). Prevalence of

a host-adapted group B Salmonella enterica in harbour porpoises (Phocoena phocoena)

from the south-west coast of England. *Veterinary Record* **167**, 173–176. doi:

10.1136/vr.c3760.

Davison, N., Barnett, J., Rule, B., Chappell, S. and Wise, G. (2010b). Group B Salmonella in

lungworms from a harbour porpoise (Phocoena phocoena). *Veterinary Record* **167**, 351–

352. doi: 10.1136/vr.c4495.

Davison, N. J., Barnett, J. E. F., Perrett, L. L., Dawson, C. E., Perkins, M. W., Deaville, R.

C. and Jepson, P. D. (2013). Meningoencephalitis and Arthritis Associated with

Brucella ceti in a Short-beaked Common Dolphin (Delphinus delphis). *Journal of*

Wildlife Diseases **49**, 632–636. doi: 10.7589/2012-06-165.

Dawson, C. E., Perrett, L. L., Stubberfield, E. J., Stack, J. A., Farrelly, S. S. J., Cooley, W.

A., Davison, N. J. and Quinney, S. (2008). Isolation and characterization of Brucella

from the lungworms of a Harbour porpoise (Phocoena phocoena). *Journal of Wildlife*

Diseases **44**, 237–246. doi: 10.7589/0090-3558-44.2.237.

de Castro, R. L., Vales, D. G., Degrati, M., García, N., Fernández, M. and Crespo, E. A.

(2014). First record of cestode cysts of Phyllobothrium delphini (Phyllobothriidae) from

dusky dolphins (Lagenorhynchus obscurus) off Argentine coast. *Hidrobiológica* **24**, 307–

310.

Delyamure, S. L. (1968). *Helminthofauna of Marine Mammals: Ecology and Phylogeny*. ed.

Skriabin, K. I. Israel Program for Scientific Translations.

Delyamure, S., Yurakhno, M., Popov, V. N., Shults, L. M. and Fay, F. H. (1984).

Helminthological comparison of subpopulations of Bering Sea spotted seals, Phoca

largha Pallas. *Soviet-American cooperative research on marine mammals* **1**, 61–65.

Dennison, S., Gulland, F., Haulena, M., De Morais, H. and Colegrove, K. (2007). Urate

nephrolithiasis in a northern elephant seal (Mirounga angustirostris) and a California sea

lion (Zalophus californianus). *Journal of Zoo and Wildlife Medicine* **38**, 114–120. doi:

10.1638/05-121.1.

Di Beneditto, A. P. M. and Ramos, R. M. A. (2004). Biology of the marine tucuxi dolphin

(Sotalia fluviatilis) in south-eastern Brazil. *Journal of the Marine Biological Association*

of the United Kingdom **84**, 1245–1250. doi: 10.1017/S0025315404010744h.

DÍaz-Delgado, J., Arbelo, M., Sacchini, S., Quesada-Canales, Ó., Andrada, M., Rivero, M.

and Fernández, A. (2012). Pulmonary Angiomatosis and Hemangioma in Common

Dolphins (*Delphinus delphis*) Stranded in Canary Islands. *Journal of Veterinary Medical*

Science **74**, 1063–1066. doi: 10.1292/jvms.11-0573.

Domiciano, I. G., Domit, C., Broadhurst, M. K., Koch, M. S. and Bracarense, A. P. F. R. L.

(2016). Assessing Disease and Mortality among Small Cetaceans Stranded at a World

Heritage Site in Southern Brazil. *PLOS ONE* **11**, e0149295. doi:

10.1371/journal.pone.0149295.

Dougherty, E. C. (1943). Notes on the lungworms of porpoises and their occurrence on the

California coast. *Proceedings of the Helminthological Society of Washington* **10**, 16–22.

Dougherty, E. C. (1944). The lungworms (Nematoda: Pseudalidae) of Odontoceti. Part I.

Parasitology **36**, 80–94.

Dougherty, E. C. and Herman, C. M. (1947). New species of the genus Parafilaroides

Dougherty, 1946 (Nematoda: Metastrongylidae) from sea-lions, with a list of the

lungworms of the Pinnipedia. *Proceedings of the Helminthological Society of*

Washington **14**, 77–87.

Dougnac, C. and Fredes, F. (2012). *Identificación de fauna endoparasitaria en cetáceos de*

Tierra del Fuego: Nuevos registros de helmintos para la zona y para odontocetos de la

región. Editorial Academica Espanola.

Duignan, P. J. (2000). Diseases of cetaceans and pinnipeds. In *Marine wildlife ; the Fabian Fay*

course for veterinarians ; proceedings 335, 4-8 September 2000, pp. 419–447. Univ. of

Sydney, Sydney.

Dunn, J. L. and Wolke, R. E. (1976). Dipetalonema spirocauda infection in the atlantic harbour

seal (Phoca vitulina concolor). *Journal of Wildlife Diseases* **12**, 531–538. doi:

10.7589/0090-3558-12.4.531.

Durante, C. A., Santos-Neto, E. B., Azevedo, A., Crespo, E. A. and Lailson-Brito, J. (2016).

POPs in the South Latin America: Bioaccumulation of DDT, PCB, HCB, HCH and

Mirex in blubber of common dolphin (Delphinus delphis) and Fraser's dolphin

(Lagenodelphis hosei) from Argentina. *Science of The Total Environment* **572**, 352–360.

doi: 10.1016/j.scitotenv.2016.07.176.

Ebmer, D., Navarrete, M. J., Muñoz, P., Flores, L. M., Gärtner, U., Brabec, J., Poppert, S.,

Taubert, A. and Hermosilla, C. (2020). Anthropozoonotic Parasites Circulating in

Synanthropic and Pacific Colonies of South American Sea Lions (Otaria flavescens):

Non-invasive Techniques Data and a Review of the Literature. *Frontiers in Marine*

Science **7**, 543829. doi: 10.3389/fmars.2020.543829.

Echenique, J., Pereira, E., Prado, J., Schild, A. L. and Valente, A. L. (2020). New host and

geographical records for Parafilaroides normani (Nematoda: Filaroididae) Dailey, 2009 in

South American fur seal, Arctocephalus australis, from southern Brazil. *Journal of*

Helminthology **94**, 1–4. doi: 10.1017/S0022149X20000826.

Elson-Riggins, J. G., Al-Banna, L., Platzer, E. G. and Kaloshian, I. (2001). Characterization

of Otostrongylus circumlitis from pacific harbor and northern elephant seals. *Journal of*

Parasitology **87**, 73–78.

Elson-Riggins, J. G., Riggins, S. A., Gulland, F. M. D. and Platzer, E. G. (2004).

Immunoglobulin responses of northern elephant and Pacific harbor seals naturally

infected with Otostrongylus circumlitus. *Journal of Wildlife Diseases* **40**, 466–475. doi:

10.7589/0090-3558-40.3.466.

Elson-Riggins, J. G., Gibbons, L. M., Van Liere, D. W., Zinkstok, E. W., Blake, D. P.,

Alegre, F., Spittle, H., Brakefield, P. M., Udo de Haes, H. A. and Osinga, N. (2020).

Surprisingly long body length of the lungworm Parafilaroides gymnurus from common

seals of the Dutch North Sea. *Parasitology Research* **119**, 1803–1817. doi:

10.1007/s00436-020-06675-7.

Evans, P. (1991). Whales, dolphins and porpoises: order Cetacea. In *The handbook of British*

mammals, pp. 299–350. Blackwell Oxford.

Evans, R. H. (2011). Segniliparus rugosus–associated Bronchiolitis in California Sea Lion.

Emerging Infectious Diseases **17**, 311–312. doi: 10.3201/eid1702.101511.

Ewing, R. Y., Rotstein, D. S., McLellan, W. A., Costidis, A. M., Lovewell, G., Schaefer, A.

M., Romero, C. H. and Bossart, G. D. (2020). Macroscopic and Histopathologic

Findings From a Mass Stranding of Rough-Toothed Dolphins (Steno bredanensis) in

2005 on Marathon Key, Florida, USA. *Frontiers in Veterinary Science* **7**, 572. doi:

10.3389/fvets.2020.00572.

Ezenwa, V. O., Price, S. A., Altizer, S., Vitone, N. D. and Cook, K. C. (2006). Host traits and

parasite species richness in even and odd-toed hoofed mammals, Artiodactyla and

Perissodactyla. *Oikos* **115**, 526–536. doi: 10.1111/j.2006.0030-1299.15186.x.

Fair, P. A., Adams, J., Mitchum, G., Hulsey, T. C., Reif, J. S., Houde, M., Muir, D., Wirth,

E., Wetzel, D., Zolman, E., McFee, W. and Bossart, G. D. (2010). Contaminant

blubber burdens in Atlantic bottlenose dolphins (Tursiops truncatus) from two

southeastern US estuarine areas: Concentrations and patterns of PCBs, pesticides,

PBDEs, PFCs, and PAHs. *Science of the Total Environment* **408**, 1577–1597. doi:

10.1016/j.scitotenv.2009.12.021.

Faulkner, J. (1995). Study of the cranial sinus nematode Stenurus minor, Metastrongyloidea, in

the harbour porpoise, Phocoena phocoena.

Faulkner, J., Measures, L. N. and Whoriskey, F. G. (1998). Stenurus minor

(Metastrongyloidea: Pseudaliidae) infections of the cranial sinuses of the harbour

porpoise, Phocoena phocoena. *Canadian Journal of Zoology* **76**, 1209–1216.

Fauquier, D., Gulland, F., Haulena, M. and Spraker, T. (2003). Biliary Adenocarcinoma in a

Stranded Northern Elephant Seal (Mirounga angustirostris). *Journal of Wildlife Diseases*

39, 723–726. doi: 10.7589/0090-3558-39.3.723.

Fauquier, D., Gulland, F., Haulena, M., Dailey, M., Rietcheck, R. L. and Lipscomb, T. P.

(2004). Meningoencephalitis in Two Stranded California Sea Lions (Zalophus

californianus) Caused by Aberrant Trematode Migration. *Journal of Wildlife Diseases* **40**,

816–819. doi: 10.7589/0090-3558-40.4.816.

Fauquier, D., Kinsel, M., Dailey, M., Sutton, G., Stolen, M., Wells, R. and Gulland, F.

(2009). Prevalence and pathology of lungworm infection in bottlenose dolphins Tursiops

truncatus from southwest Florida. *Diseases of Aquatic Organisms* **88**, 85–90. doi:

10.3354/dao02095.

Fauquier, D. A., Kinsel, M. J., Dailey, M. D., Sutton, G. E., Stolen, M. K., Wells, R. S. and

Gulland, F. M. D. (2010). Prevalence and pathology of lungworm infection in bottlenose

dolphins Tursiops truncatus from southwest Florida. *Diseases of Aquatic Organisms* **88**,

85–90. doi: 10.3354/dao02095.

Fenton, H., Daoust, P., Forzán, M., Vanderstichel, R., Ford, J., Spaven, L., Lair, S. and

Raverty, S. (2017). Causes of mortality of harbor porpoises Phocoena phocoena along

the Atlantic and Pacific coasts of Canada. *Diseases of Aquatic Organisms* **122**, 171–183.

doi: 10.3354/dao03080.

Fernández, M., Agustí, C., Aznar, F. and Raga, J. (2003). Gastrointestinal helminths of

Risso's dolphin Grampus griseus from the Western Mediterranean. *Diseases of Aquatic*

Organisms **55**, 73–76. doi: 10.3354/dao055073.

Fleischman, R. W. and Squire, R. A. (1970). Verminous Pneumonia in the California Sea Lion

(Zalophus californianus). *Pathologia veterinaria* **7**, 89–101. doi:

10.1177/030098587000700201.

Forbes, A. B. (2021). Lungworm infections in cattle. In *Parasites of cattle and sheep: a*

practical guide to their biology and control, pp. 116–138. CAB International, UK doi:

10.1079/9781789245158.0116.

Forrester, D. J., Odell, D. K., Thompson, N. P. and White, J. R. (1980). Morphometrics,

Parasites, and Chlorinated Hydrocarbon Residues of Pygmy Killer Whales from Florida.

Journal of Mammalogy **61**, 356–360. doi: 10.2307/1380067.

Fouchier, R. A. M., Bestebroer, T. M., Martina, B. E. E., Rimmelzwaan, G. F. and

Osterhaus, A. D. M. E. (2001). Infection of grey seals and harbour seals with influenza

B virus. *International Congress Series* **1219**, 225–231. doi: 10.1016/S0531-

5131(01)00647-1.

Gabel, M., Theisen, S., Palm, H. W., Dähne, M. and Unger, P. (2021). Nematode Parasites in

Baltic Sea Mammals, Grey Seal (Halichoerus grypus (Fabricius, 1791)) and Harbour

Porpoise (Phocoena phocoena (L.)), from the German Coast. *Acta Parasitologica* **66**, 26–

33. doi: 10.1007/s11686-020-00246-7.

García de los Ríos y Loshuertos, A., Soler Laguía, M., Arencibia Espinosa, A., López

Fernández, A., Covelo Figueiredo, P., Martínez Gomariz, F., Sánchez Collado, C.,

García Carrillo, N. and Ramírez Zarzosa, G. (2021). Comparative Anatomy of the

Nasal Cavity in the Common Dolphin Delphinus delphis L., Striped Dolphin Stenella

coeruleoalba M. and Pilot Whale Globicephala melas T.: A Developmental Study.

Animals **11**, 441. doi: 10.3390/ani11020441.

Garner, M. M., Lambourn, D. M., Jeffries, S. J., Hall, P. B., Rhyan, J. C., Ewalt, D. R.,

Polzin, L. M. and Cheville, N. F. (1997). Evidence of Brucella Infection in

Parafilaroides Lungworms in a Pacific Harbor Seal (Phoca Vitulina Richardsi). *Journal*

of Veterinary Diagnostic Investigation **9**, 298–303. doi: 10.1177/104063879700900311.

Geraci, J. R. and St. Aubin, D. J. (1987). Effects of parasites on marine mammals.

International Journal for Parasitology **17**, 407–414. doi: 10.1016/0020-7519(87)90116-

0.

Gerber, J. A., Roletto, J., Morgan, L. E., Smith, D. M. and Gage, L. J. (1993). Findings in

pinnipeds stranded along the central and northern California coast, 1984–1990. *Journal*

of Wildlife Diseases **29**, 423–433. doi: 10.7589/0090-3558-29.3.423.

Gibson, D. I. and Harris, E. A. (1979). The helminth-parasites of cetaceans in the collection of

the British Museum (Natural History). *Investigations on Cetacea* **10**, 309–324.

Gibson, D. I., Harris, E. A., Bray, R. A., Jepson, P. D., Kuiken, T., Baker, J. R. and

Simpson, V. R. (1998). A survey of the helminth parasites of cetaceans stranded on the

coast of England and Wales during the period 1990-1994. *Journal of Zoology* **244**, 563–

574. doi: 10.1111/j.1469-7998.1998.tb00061.x.

Giorda, F., Ballardini, M., Di Guardo, G., Pintore, M. D., Grattarola, C., Iulini, B.,

Mignone, W., Goria, M., Serracca, L., Varello, K., Dondo, A., Acutis, P. L.,

Garibaldi, F., Scaglione, F. E., Gustinelli, A., Mazzariol, S., Di Francesco, C. E.,

Tittarelli, C., Casalone, C. and Pautasso, A. (2017). Postmortem findings in cetaceans

found stranded in the pelagos sanctuary, Italy, 2007–14. *Journal of Wildlife Diseases* **53**,

795. doi: 10.7589/2016-07-150.

Goldstein, T., Colegrove, K., Hanson, M. and Gulland, F. (2011). Isolation of a novel

adenovirus from California sea lions Zalophus californianus. *Diseases of Aquatic*

Organisms **94**, 243–248. doi: 10.3354/dao02321.

Goodall, R., Galeazzi, A., Leatherwood, S., Miller, K., Cameron, I., Kastelein, R. and

Sobral, A. (1988). Studies of Commerson's dolphins, Cephalorhynchus commersonii, off

Tierra del Fuego, 1976-1984, with a review of information on the species in the South

Atlantic. *Report of the International Whaling Commission* 3–70.

Gosselin, J.-F. and Measures, L. N. (1997). Redescription of Filaroides (Parafilaroides)

gymnurus(Railliet, 1899) (Nematoda: Metastrongyloidea), with comments on other

species in pinnipeds. *Canadian Journal of Zoology* **75**, 359–370. doi: 10.1139/z97-045.

Gosselin, J.-F., Measures, L. N. and Huot, J. (1998). Lungworm (Nematoda:

Metastrongyloidea) infections in Canadian phocids. *Canadian Journal of Fisheries and*

Aquatic Sciences **55**, 825–834. doi: 10.1139/f97-306.

Greenwood, A. G. and Taylor, D. C. (1979). Odontocete parasites: Some new host records.

Aquatic Mammals **7**, 23–25.

Greig, D. J., Gulland, F. M. D. and Kreuder, C. (2005). A Decade of Live California Sea Lion

(Zalophus californianus) Strandings Along the Central California Coast: Causes and

Trends, 1991-2000. *Aquatic Mammals* **31**, 11–22. doi: 10.1578/AM.31.1.2005.11.

Greig, D., Gulland, F., Smith, W., Conrad, P., Field, C., Fleetwood, M., Harvey, J., Ip, H.,

Jang, S., Packham, A., Wheeler, E. and Hall, A. (2014). Surveillance for zoonotic and

selected pathogens in harbor seals Phoca vitulina from central California. *Diseases of*

Aquatic Organisms **111**, 93–106. doi: 10.3354/dao02762.

Groch, K. R., Santos-Neto, E. B., Díaz-Delgado, J., Ikeda, J. M. P., Carvalho, R. R.,

Oliveira, R. B., Guari, E. B., Bisi, T. L., Azevedo, A. F., Lailson-Brito, J. and Catão-

Dias, J. L. (2018). Guiana Dolphin Unusual Mortality Event and Link to Cetacean

Morbillivirus, Brazil. *Emerging Infectious Diseases* **24**, 1349–1354. doi:

10.3201/eid2407.180139.

Groch, K. R., Díaz-Delgado, J., Santos-Neto, E. B., Ikeda, J. M. P., Carvalho, R. R.,

Oliveira, R. B., Guari, E. B., Flach, L., Sierra, E., Godinho, A. I., Fernández, A.,

Keid, L. B., Soares, R. M., Kanamura, C. T., Favero, C., Ferreira-Machado, E.,

Sacristán, C., Porter, B. F., Bisi, T. L., Azevedo, A. F., Lailson-Brito, J. and Catão-

Dias, J. L. (2020a). The Pathology of Cetacean Morbillivirus Infection and

Comorbidities in Guiana Dolphins During an Unusual Mortality Event (Brazil, 2017–

2018). *Veterinary Pathology* **57**, 845–857. doi: 10.1177/0300985820954550.

Groch, K. R., Díaz-Delgado, J., Santos-Neto, E. B., Ikeda, J. M. P., Carvalho, R. R.,

Oliveira, R. B., Guari, E. B., Flach, L., Sierra, E., Godinho, A. I., Fernández, A.,

Keid, L. B., Soares, R. M., Kanamura, C. T., Favero, C., Ferreira-Machado, E.,

Sacristán, C., Porter, B. F., Bisi, T. L., Azevedo, A. F., Lailson-Brito, J. and Catão-

Dias, J. L. (2020b). The Pathology of Cetacean Morbillivirus Infection and

Comorbidities in Guiana Dolphins During an Unusual Mortality Event (Brazil, 2017–

2018). *Veterinary Pathology* **57**, 845–857. doi: 10.1177/0300985820954550.

Gui, D., He, J., Zhang, X., Tu, Q., Chen, L., Feng, K., Liu, W., Mai, B. and Wu, Y. (2018).

Potential association between exposure to legacy persistent organic pollutants and

parasitic body burdens in Indo-Pacific finless porpoises from the Pearl River Estuary,

China. *Science of The Total Environment* **643**, 785–792. doi:

10.1016/j.scitotenv.2018.06.249.

Guimarães, J. P., Batista, R. L. G., Mariani, D. B. and Vergara-Parente, J. E. (2013).

Ingestion of plastic debris by estuarine dolphin, Sotalia guianensis, off northeastern

Brazil. *Arquivos de Ciências do Mar* **46**, 107–122.

Guimarães, J. P., Febronio, A. M. B., Vergara-Parente, J. E. and Werneck, M. R. (2015).

Lesions Associated with Halocercus brasiliensis Lins de Almeida, 1933 in the Lungs of

Dolphins Stranded in the Northeast of Brazil. *Journal of Parasitology* **101**, 248–251. doi:

10.1645/14-513.1.

Gulland, F. M. D., Beckmen, K., Burek, K., Lowenstine, L., Werner, L., Spraker, T.,

Dailey, M. and Harris, E. (1997a). Nematode (Otostrongylus circumlitus) infestation of

northern elephant seals (Mirounga angustirostris) stranded along the central California

coast. *Marine Mammal Science* **13**, 446–458. doi: 10.1111/j.1748-7692.1997.tb00651.x.

Gulland, F. M. D., Beckmen, K., Burek, K., Lowenstine, L., Werner, L., Spraker, T.,

Dailey, M. and Harris, E. (1997b). NEMATODE (OTOSTRONGYLUS

CIRCUMLITUS) INFESTATION OF NORTHERN ELEPHANT SEALS (MIROUNGA

ANGUSTIROSTRIS) STRANDED ALONG THE CENTRAL CALIFORNIA COAST.

Marine Mammal Science **13**, 446–458. doi: 10.1111/j.1748-7692.1997.tb00651.x.

Gulland, F. M. D., Hall, A. J., Greig, D. J., Frame, E. R., Colegrove, K. M., Booth, R. K. N.,

Wasser, S. K. and Scott-Moncrieff, J. C. R. (2012). Evaluation of circulating

eosinophil count and adrenal gland function in California sea lions naturally exposed to

domoic acid. *Journal of the American Veterinary Medical Association* **241**, 943–949. doi:

10.2460/javma.241.7.943.

Haddaway, N. R. and Watson, M. J. (2016). On the benefits of systematic reviews for wildlife

parasitology. *International Journal for Parasitology. Parasites and Wildlife* **5**, 184–191.

doi: 10.1016/j.ijppaw.2016.05.002.

Harris, E. (1982). The helminth parasites of the Cetacea (or Parasitology with a porpoise).pp.

R71–R71. Cambridge Univ Press.

Haulena, M. and Gulland, F. M. D. (2001). Use of medetomidine-zolazepam-tiletamine with

and without atipamezole reversal to immobilize captive california sea lions. *Journal of*

Wildlife Diseases **37**, 566–573. doi: 10.7589/0090-3558-37.3.566.

Hermosilla, C., Silva, L. M. R., Prieto, R., Kleinertz, S., Taubert, A. and Silva, M. A.

(2015). Endo- and ectoparasites of large whales (Cetartiodactyla: Balaenopteridae,

Physeteridae): Overcoming difficulties in obtaining appropriate samples by non- and

minimally-invasive methods. *International Journal for Parasitology: Parasites and*

Wildlife **4**, 414–420. doi: 10.1016/j.ijppaw.2015.11.002.

Hermosilla, C., Silva, L. M. R., Navarro, M. and Taubert, A. (2016). Anthropozoonotic

Endoparasites in Free-Ranging "Urban" South American Sea Lions (*Otaria flavescens*).

Journal of Veterinary Medicine **2016**, 1–7. doi: 10.1155/2016/7507145.

Hernández-Orts, J. S., Hernández-Mena, D. I., Pantoja, C., Kuchta, R., García, N. A.,

Crespo, E. A. and Loizaga, R. (2021). A Visitor of Tropical Waters: First Record of a

Clymene Dolphin (Stenella clymene) Off the Patagonian Coast of Argentina, With

Comments on Diet and Metazoan Parasites. *Frontiers in Marine Science* **8**, 658975. doi:

10.3389/fmars.2021.658975.

Herreman, J. K., McIntosh, A. D., Dziuba, R. K., Blundell, G. M., Ben-David, M. and

Greiner, E. C. (2011). Parasites of harbor seals (Phoca vitulina) in Glacier Bay and

Prince William Sound, Alaska. *Marine Mammal Science* **27**, 247–253. doi:

10.1111/j.1748-7692.2009.00355.x.

Houde, M., Measures, L. N. and Huot, J. (2003). Lungworm (Pharurus pallasii:

Metastrongyloidea: Pseudaliidae) infection in the endangered St. Lawrence beluga whale

(Delphinapterus leucas). *Canadian Journal of Zoology* **81**, 543–551.

Hsü, H. and Hoeppli, R. (1933). On some parasitic nematodes collected in Amoy. *Peking*

Natural History Bulletin **8**, 155–168.

Huertas, V. and Lagueux, C. J. (2016). First Recorded Mass Stranding of the Short-Finned

Pilot Whale (Globicephala macrorhynchus) on the Caribbean Coast of Nicaragua.

Aquatic Mammals **42**, 27–34. doi: 10.1578/AM.42.1.2016.27.

Jabbar, A., Mohandas, N. and Gasser, R. B. (2014). Characterisation of the mitochondrial

genome of Parafilaroides normani (lungworm) of Arctocephalus pusillus doriferus

(Australian fur seal). *Parasitology Research* **113**, 3049–3055. doi: 10.1007/s00436-014-

3968-8.

Jacobus, K., Marigo, J., Gastal, S. B., Taniwaki, S. A., Ruoppolo, V., Catão-Dias, J. L. and

Tseng, F. (2016). Identification of respiratory and gastrointestinal parasites of three

species of pinnipeds Arctocephalus australis, Arctocephalus gazella, and Otaria

flavescens in southern Brazil. *Journal of Zoo and Wildlife Medicine* **47**, 132–140. doi:

10.1638/2015-0090.1.

Jauniaux, T., Petitjean, D., Brenez, C., Borrens, M., Brosens, L., Haelters, J., Tavernier, T.

and Coignoul, F. (2002). Post-mortem Findings and Causes of Death of Harbour

Porpoises (Phocoena phocoena) Stranded from 1990 to 2000 along the Coastlines of

Belgium and Northern France. *Journal of Comparative Pathology* **126**, 243–253. doi:

10.1053/jcpa.2001.0547.

Jepson, P. D., Kuiken, T., Bennett, P. M., Baker, J. R., Simpson, V. R. and Kennedy, S.

(2000). Pulmonary pathology of harbour porpoises (Phocoena phocoena) stranded in

England and Wales between 1990 and 1996. *Veterinary Record* **146**, 721–728. doi:

10.1136/vr.146.25.721.

Jepson, P. D., Deaville, R., Barber, J. L., Aguilar, À., Borrell, A., Murphy, S., Barry, J.,

Brownlow, A., Barnett, J., Berrow, S., Cunningham, A. A., Davison, N. J., Ten

Doeschate, M., Esteban, R., Ferreira, M., Foote, A. D., Genov, T., Giménez, J.,

Loveridge, J., Llavona, Á., Martin, V., Maxwell, D. L., Papachlimitzou, A., Penrose,

R., Perkins, M. W., Smith, B., De Stephanis, R., Tregenza, N., Verborgh, P.,

Fernandez, A. and Law, R. J. (2016). PCB pollution continues to impact populations of

orcas and other dolphins in European waters. *Scientific Reports* **6**, 1–17. doi:

10.1038/srep18573.

Johnston, D. G. and Ridgway, S. H. (1969). Parasitism in some marine mammals. *Journal of*

the American Veterinary Medical Association **155**, 1064–1072.

Kastelein, R. A. and Lavaleije, M. S. S. (1992). Foreign Bodies in the stomach of female

Habrour porpoises (Phocoena phocoena) from the North Sea. *Aquatic Mammals* **18**, 40–

46.

Kaye, S., Johnson, S., Arnold, R. D., Nie, B., Davis, J. T., Gulland, F., Abou-Madi, N. and

Fletcher, D. J. (2016). Pharamcokinetic study of oral e-aminocaproic acid in the northern

elephant seal (Mirounga Angustirostris). *Journal of Zoo and Wildlife Medicine* **47**, 438–

446. doi: 10.1638/2015-0138.1.

Kaye, S., Johnson, S., Rios, C. and Fletcher, D. J. (2017). Plasmatic coagulation and

fibrinolysis in healthy and Otostrongylus -affected Northern elephant seals (Mirounga

angustirostris). *Veterinary Clinical Pathology* **46**, 589–596. doi: 10.1111/vcp.12540.

Kelly, T. R., Greig, D., Colegrove, K. M., Lowenstine, L. J., Dailey, M., Gulland, F. M. and

Haulena, M. (2005). Metastrongyloid Nematode (Otostrongylus circumlitus) Infection in

a Stranded California Sea Lion (Zalophus californianus)— a New Host-parasite

Association. *Journal of Wildlife Diseases* **41**, 593–598. doi: 10.7589/0090-3558-

41.3.593.

Kennedy, M. J. (1986). Metastrongyloidea) from the lungs of the ringed seal, Phoca hispida

(Phocidae), from the Beaufort Sea, Canada. *Canadian Journal of Zoology* **64**, 1864–

1868. doi: 10.1139/z86-278.

Kennedy, S., Smyth, J. A., Cush, P. F., Duignan, P., Platten, M., McCullough, S. J. and

Allan, G. M. (1989). Histopathologic and Immunocytochemical Studies of Distemper in

Seals. *Veterinary Pathology* **26**, 97–103. doi: 10.1177/030098588902600201.

Kenyon, A. J. and Kenyon, B. J. (1977). Prevalence of Pharurus pallasii in the beluga whale

(Delphinapterus leucas) of Churchill River Basin, Manitoba. *Journal of Wildlife Diseases*

13, 338–340. doi: 10.7589/0090-3558-13.4.338.

Kijewska, A. P., Jankowski, Z., Kuklik, I. and Rokicki, J. (2003). Pathological changes in the

auditory organs of the harbor porpoise (Phocoena phocoena, L.) associated with Stenurus

minor (Kuhn, 1829). *Acta Parasitologica* **48**, 60–63. doi: 10.13140/RG.2.1.4781.7046.

Kontrimavichus, V. and Delyamure, S. (1979). Filaroides of domestic and wild animals.

Fundamentals of nematology **29**, 30–36.

Kroese, M. V., Beckers, L., Bisselink, Y. J. W. M., Brasseur, S., van Tulden, P. W., Koene,

M. G. J., Roest, H. I. J., Ruuls, R. C., Backer, J. A., IJzer, J., van der Giessen, J. W.

B. and Willemsen, P. T. J. (2018). Brucella pinnipedialis in grey seals (Halichoerus

grypus) and harbour seals (Phoca vitulina) in the Netherlands. *Journal of Wildlife*

Diseases **54**, 439. doi: 10.7589/2017-05-097.

Kühn, J. (1829). Description d'un nouvelle espèce de strongyle trouveé dans le marsouin.

Bulletin du Sciences Naturelles et Geologie **17**, 150–153.

Kumazawa, H., Yona, R., Hirai, M. and Hasegawa, H. (2006). A Fatal Case of

Bronchopneumonia Associated with Lungworm lnfection in a Bottle-nosed dolphin,

tursiops truncatus (Cetacea: Dephinidae). *Japanese Journal of Zoo and Wildlife Medicine*

11, 31–34.

Kuramochi, T., Araki, J. and Machida, M. (1990). Pseudaliid nematodes from Dall's

porpoise, Phocoenoides dalli. *Bulletin of the National Science Museum Series A (Zoology*

97–103.

Kuramochi, T., Kikuchi, T., Okamura, H., Tatsukawa, T., Doi, H., Nakamura, K.,

Yamada, T., Koda, Y., Yoshida, Y. and Matsuura, M. (2000). Parasitic helminth and

epizoit fauna of finless porpoise in the Inland Sea of Japan and the western North Pacific

with a preliminary note on faunal difference by host's local population. *Memoirs of the*

National Science Museum, Tokyo **33**, 83–95.

Kurochkin, Y. V. and Zablotsky, V. (1958). On the helminth fauna of the Caspian seal [In

Russian]. *Trudy Astrakhan—skogo Zapovednika 1993* 337–343.

Kuwamura, M., Sawamoto, O., Yamate, J., Aoki, M., Ohnishi, Y. and Kotani, T. (2007).

Pulmonary Vascular Proliferation and Lungworm (Stenurus ovatus) in a Bottlenose

Dolphin (Tursiops turncatus). *Journal of Veterinary Medical Science* **69**, 531–533. doi:

10.1292/jvms.69.531.

Kuzmina, T. A., Spraker, T. R., Kudlai, O., Lisitsyna, O. I., Zabludovskaja, S. O.,

Karbowiak, G., Fontaine, C. and Kuchta, R. (2018). Metazoan parasites of California

sea lions (Zalophus californianus): A new data and review. *International Journal for*

Parasitology: Parasites and Wildlife **7**, 326–334. doi: 10.1016/j.ijppaw.2018.09.001.

Lambourn, D. M., Garner, M., Ewalt, D., Raverty, S., Sidor, I., Jeffries, S. J., Rhyan, J. and

Gaydos, J. K. (2013). Brucella pinnipedialis infections in pacific harbour seals Phoca

vitulina richardsi from Washington State, USA. *Journal of Wildlife Diseases* **49**, 802–

815. doi: 10.7589/2012-05-137.

Lane, E. P., de Wet, M., Thompson, P., Siebert, U., Wohlsein, P. and Plön, S. (2014). A

Systematic Health Assessment of Indian Ocean Bottlenose (Tursiops aduncus) and Indo-

Pacific Humpback (Sousa plumbea) Dolphins Incidentally Caught in Shark Nets off the

KwaZulu-Natal Coast, South Africa. *PLoS ONE* **9**,. doi: 10.1371/journal.pone.0107038.

Lehnert, K., Raga, J. and Siebert, U. (2005). Macroparasites in stranded and bycaught harbour

porpoises from German and Norwegian waters. *Diseases of Aquatic Organisms* **64**, 265–

269. doi: 10.3354/dao064265.

Lehnert, K., Raga, J. A. and Siebert, U. (2007). Parasites in harbour seals (Phoca vitulina)

from the German Wadden Sea between two Phocine Distemper Virus epidemics.

Helgoland Marine Research **61**, 239–245. doi: 10.1007/s10152-007-0072-9.

Lehnert, K., von Samson-Himmelstjerna, G., Schaudien, D., Bleidorn, C., Wohlsein, P. and

Siebert, U. (2010). Transmission of lungworms of harbour porpoises and harbour seals:

Molecular tools determine potential vertebrate intermediate hosts. *International Journal*

for Parasitology **40**, 845–853. doi: 10.1016/j.ijpara.2009.12.008.

Lehnert, K., Seibel, H., Hasselmeier, I., Wohlsein, P., Iversen, M., Nielsen, N. H., Heide-

Jørgensen, M. P., Prenger-Berninghoff, E. and Siebert, U. (2014). Increase in parasite

burden and associated pathology in harbour porpoises (Phocoena phocoena) in West

Greenland. *Polar Biology* **37**, 321–331. doi: 10.1007/s00300-013-1433-2.

Lehnert, K., Randhawa, H. and Poulin, R. (2017). Metazoan parasites from odontocetes off

New Zealand: new records. *Parasitology Research* **116**, 2861–2868. doi:

10.1007/s00436-017-5573-0.

Lehnert, K., Poulin, R. and Presswell, B. (2019). Checklist of marine mammal parasites in

New Zealand and Australian waters. *Journal of Helminthology* **93**, 649–676. doi:

10.1017/S0022149X19000361.

Leidenberger, S. and Boström, S. (2009). Description of the lungworm Otostrongylus

circumlitus (Railliet, 1899) de Bruyn, 1933 (Metastrongyloidea: Crenosomatidae) found

in the heart of harbour seals from Sweden. *Journal of Nematode Morphology and*

Systematics **12**, 169–175.

Lipscomb, T. P., Kennedy, S., Moffett, D., Krafft, A., Klaunberg, B. A., Lichy, J. H., Regan,

G. T., Worthy, G. A. J. and Taubenberger, J. K. (1996). Morbilliviral Epizootic in

Bottlenose Dolphins of the Gulf of Mexico. *Journal of Veterinary Diagnostic*

Investigation **8**, 283–290. doi: 10.1177/104063879600800302.

Liu, H., Plancarte, M., Ball, E. E., Weiss, C. M., Gonzales-Viera, O., Holcomb, K., Ma, Z.-

M., Allen, A. M., Reader, J. R., Duignan, P. J., Halaska, B., Khan, Z., Kriti, D.,

Dutta, J., van Bakel, H., Jackson, K., Pesavento, P. A., Boyce, W. M. and Coffey, L.

L. (2021). Respiratory Tract Explant Infection Dynamics of Influenza A Virus in

California Sea Lions, Northern Elephant Seals, and Rhesus Macaques. *Journal of*

Virology **95**,. doi: 10.1128/JVI.00403-21.

Lucas, Z., Daoust, P.-Y., Conboy, G. and Brimacombe, M. (2003). Health status of harp seals

(Phoca groenlandica) and hooded seals (Cystophora cristata) on Sable Island, Nova

Scotia, Canada, concurrent with their expanding range. *Journal of Wildlife Diseases* **39**,

16–28. doi: 10.7589/0090-3558-39.1.16.

MacNeill, A. C., Neufeld, J. L. and Webster, W. A. (1975). Pulmonary nematodiasis in a

narwhale. *The Canadian Veterinary Journal = La Revue Veterinaire Canadienne* **16**, 53–

55.

Marigo, J., Ruoppolo, V., Rosas, F. C. W., Valente, A. L. S., Oliveira, M. R., Dias, R. A. and

Catão-Dias, J. L. (2010). Helminths of Sotalia guianensis (Cetacea: Delphinidae) from

the South and Southeastern Coasts of Brazil. *Journal of Wildlife Diseases* **46**, 599–602.

doi: 10.7589/0090-3558-46.2.599.

Mawson, P. M. (1953). Parasitic Nematoda collected by the Australian National Antarctic

Research Expedition: Heard Island and Macquarie Island, 1948–1951. *Parasitology* **43**,

291–297. doi: 10.1017/S0031182000018667.

Mazzariol, S., Marruchella, G., Di Guardo, G., Podesta, M., Olivieri, V., Colangelo, P.,

Kennedy, S., Castagnaro, M. and Cozzi, B. (2007). Post-mortem Findings in Cetacean

Stranded along Italian Adriatic Sea coastline (2000-2006).

McColl, K. A. and Obendorf, D. L. (1982). Helminth parasites and associated pathology in

stranded frasers dolphins, Lagenodelohis Hosei (Fraser, 1956). *Aquatic Mammals* **9**, 30–

34.

McFee, W. and Lipscomb, T. P. (2009). Major pathological findings and probable causes of

mortality in bottlenose dolphins stranded in South Carolina from 1993 to 2006. *Journal*

of Wildlife Diseases **45**, 575–593. doi: 10.7589/0090-3558-45.3.575.

McKenzie, J. and Blair, D. (1983). Parasites from Hector's dolphin (Cephalorhynchus

hectori).pp. 126–127. Sir Publishing po box 399, Wellington, New zealand.

McKnight, C. A., Reynolds, T. L., Haulena, M., deLahunta, A. and Gulland, F. M. D.

(2005). Congenital Hemicerebral Anomaly in a Stranded Pacific Harbor Seal (Phoca

vitulina richardsi). *Journal of Wildlife Diseases* **41**, 654–658. doi: 10.7589/0090-3558-

41.3.654.

McManus, T., Wapstra, J., Guiler, E., Munday, B. and Obendorf, D. (1984). Cetacean

strandings in Tasmania from February 1978 to May 1983. *Papers and Proceedings of*

The Royal Society of Tasmania **118**, 117–135. doi: 10.26749/rstpp.118.117.

Measures, L. N. (2001). Lungworms of Marine Mammals. In *Parasitic Diseases of Wild*

Mammals (ed. Samuel, W. M., Pybus, M. J., and Kocan, A. A.), pp. 279–300. Iowa State

University Press, Ames, Iowa, USA doi: 10.1002/9780470377000.ch10.

Measures, L. N. and Gosselin, J. (1994). Helminth parasites of ringed seal, Phoca hispida, from

northern Quebec, Canada. *Journal of the Helminthological Society of Washington* **61**,

240–244.

Measures, L. N., Béland, P., Martineau, D. and Guise, S. D. (1995). Helminths of an

endangered population of belugas, Delphinapterus leucas , in the St. Lawrence estuary,

Canada. *Canadian Journal of Zoology* **73**, 1402–1409. doi: 10.1139/z95-165.

Melo, O. P., Ramos, R. M. A. and Di Beneditto, A. P. M. (2006). Helminths of the marine

tucuxi, Sotalia fluviatilis (Gervais, 1853) (Cetacea: Delphinidae), in northern Rio de

Janeiro State, Brazil. *Brazilian Archives of Biology and Technology* **49**, 145–148. doi:

10.1590/S1516-89132006000100017.

Migaki, G., Van Dyke, D. and Hubbard, R. C. (1971). Some histopathological lesions caused

by helminths in marine mammals. *Journal of Wildlife Diseases* **7**, 281–289.

Mignucci-Giannoni, A. A., Rodríguez-López, M. A., Perez-Zayas, J. J., Montoya-Ospina,

R. A. and Williams, E. H. J. (1998a). First record of the melonhead whale for Puerto

Rico. *Mammalia* **62**, 452–457.

Mignucci-Giannoni, A. A., Hoberg, E. P., Siegel-Causey, D. and Williams, E. H. (1998b).

Metazoan Parasites and Other Symbionts of Cetaceans in the Caribbean. *The Journal of*

Parasitology **84**, 939. doi: 10.2307/3284625.

Morales Vela, B. and Olvera Gómez, L. D. (1993). Varamiento de calderones Globicephala

macrorhynchus (Cetacea: Delphinidae) en la Isla de Cozumel, Quintana Roo, México.

Anales del Instituto de Biología serie Zoología **64**,.

Morell, M., Lehnert, K., IJsseldijk, L., Raverty, S., Wohlsein, P., Gröne, A., André, M.,

Siebert, U. and Shadwick, R. (2017). Parasites in the inner ear of harbour porpoise:

cases from the North and Baltic Seas. *Diseases of Aquatic Organisms* **127**, 57–63. doi:

10.3354/dao03178.

Moser, M. and Rhinehart, H. (1993). The lungworm, Halocercus spp. (Nematoda:

Pseudaliidae) in cetaceans from California. *Journal of Wildlife Diseases* **29**, 507–508.

doi: 10.7589/0090-3558-29.3.507.

Nicholson, A. and Fanning, J. (1981). Parasites and associated pathology of the respiratory

tract of the Australian sea lion: Neophoca cinerea.pp. 178–181.

Niebuhr, C. N., Siers, S. R., Leinbach, I. L., Kaluna, L. M. and Jarvi, S. I. (2021). Variation

in *Angiostrongylus cantonensis* infection in definitive and intermediate hosts in Hawaii, a

global hotspot of rat lungworm disease. *Parasitology* **148**, 133–142. doi:

10.1017/S003118202000164X.

Nunn, C. L., Altizer, S., Jones, K. E. and Sechrest, W. (2003). Comparative Tests of Parasite

Species Richness in Primates. *The American Naturalist* **162**, 597–614. doi:

10.1086/378721.

reduced

Oliveira, J. B., Morales, J. A., González-Barrientos, R. C., Hernández-Gamboa, J. and

Hernández-Mora, G. (2011). Parasites of cetaceans stranded on the Pacific coast of

Costa Rica. *Veterinary Parasitology* **182**, 319–328. doi: 10.1016/j.vetpar.2011.05.014.

Onderka, D. K. (1989). Prevance and pathology of nematode infections in the lungs of ringed

seals (Phoca hispida) of the Western Arctic of Canada. *Journal of Wildlife Diseases* **25**,

218–224. doi: 10.7589/0090-3558-25.2.218.

Osinga, N., Kappe, A. L., Brakefield, P. M., Udo de Haes, H. A. and Elson-Riggins, J. G.

(2015). Comparative biology of common and grey seals along the Dutch coast: stranding,

disease, rehabilitation and conservation. Chapter 8: Observations regarding transmission

of seal nematodes in common seals, Phoca vitulina vitulina from the Wadden Sea.

Page, M. J., McKenzie, J. E., Bossuyt, P. M., Boutron, I., Hoffmann, T. C., Mulrow, C. D.,

Shamseer, L., Tetzlaff, J. M., Akl, E. A., Brennan, S. E., Chou, R., Glanville, J.,

Grimshaw, J. M., Hróbjartsson, A., Lalu, M. M., Li, T., Loder, E. W., Mayo-Wilson,

E., McDonald, S., McGuinness, L. A., Stewart, L. A., Thomas, J., Tricco, A. C.,

Welch, V. A., Whiting, P. and Moher, D. (2021). The PRISMA 2020 statement: An

updated guideline for reporting systematic reviews. *PLOS Medicine* **18**, e1003583. doi:

10.1371/journal.pmed.1003583.

Parsons, E. C. M. and Jefferson, T. A. (2000). Post-mortem investigations on stranded

dolphins and porpoises from Hong Kong waters. *Journal of Wildlife Diseases* **36**, 342–

356. doi: 10.7589/0090-3558-36.2.342.

Parsons, E. C. M., Overstreet, R. M. and Jefferson, T. A. (2001). Parasites from Indo-Pacific

hump-backed dolphins (Sousa chinensis) and finless porpoises (Neophocaena

phocaenoides) stranded in Hong Kong. *Veterinary Record* **148**, 776–780. doi:

10.1136/vr.148.25.776.

Pedersen, A. B. and Fenton, A. (2015). The role of antiparasite treatment experiments in

assessing the impact of parasites on wildlife. *Trends in Parasitology* **31**, 200–211. doi:

10.1016/j.pt.2015.02.004.

Pekmezci, G. Z., Yardimci, B., Gürler, A. T., Bölükbaş, C. S., Açici, M. and Umur, Ş.

(2013). Survey on the Presence of Nematodes and Associated with Pathology in Marine

Mammals from Turkish Waters. *Kafkas Universitesi Veteriner Fakultesi Dergisi* **19**,

1035–1038. doi: 10.9775/kvfd.2013.9409.

Perrin, W. F., Mitchell, E. D., Mead, J. G., Caldwell, D. K., Caldwell, M. C., Bree, P. J. H.

and Dawbin, W. H. (1987). Revision of the spotted dolphins, Stenella spp. *Marine*

Mammal Science **3**, 99–170. doi: 10.1111/j.1748-7692.1987.tb00158.x.

Perrin, W. F., Caldwell, D. K. and Caldwell, M. (1994). Atlantic spotted dolphin Stenella

frontalis (G. Cuvier, 1829). *Handbook of marine mammals* **5**, 173–190.

Petter, A. J. and Pilleri, G. (1982). Pharurus asiaeorientalis new species, metastrongylid

nematode, parasite of Neophocaena asiaeorientalis (Phocoenidae, Cetacea).

Investigations on Cetacea **13**, 141–148.

Piché, C., Measures, L., Bédard, C. and Lair, S. (2010). Bronchoalveolar lavage and

pulmonary histopathology in harp seals (Phoca groenlandica) experimentally infected

with Otostrongylus circumlitis. *Journal of Wildlife Diseases* **46**, 409–421. doi:

10.7589/0090-3558-46.2.409.

Pool, R., Chandradeva, N., Gkafas, G., Raga, J. A., Fernández, M. and Aznar, F. J. (2020a).

Transmission and Predictors of Burden of Lungworms of the Striped Dolphin (Stenella

coeruleoalba) in the Western Mediterranean. *Journal of Wildlife Diseases* **56**, 186. doi:

10.7589/2018-10-242.

Pool, R., Fernández, M., Chandradeva, N., Raga, J. A. and Aznar, F. J. (2020b). The

taxonomic status of Skrjabinalius guevarai Gallego & Selva, 1979 (Nematoda:

Pseudaliidae) and the synonymy of Skrjabinalius Delyamure, 1942 and Halocercus

Baylis & Daubney, 1925. *Systematic Parasitology* **97**, 389–401. doi: 10.1007/s11230-

020-09921-9.

Pool, R., Romero-Rubira, C., Raga, J. A., Fernández, M. and Aznar, F. J. (2021).

Determinants of lungworm specificity in five cetacean species in the western

Mediterranean. *Parasites & Vectors* **14**, 196. doi: 10.1186/s13071-021-04629-1.

Poulin, R. (2011). *Evolutionary Ecology of Parasites: (Second Edition)*. Princeton University

Press doi: doi:10.1515/9781400840809.

Prahl, S., Ketten, D. R. and Siebert, U. (2008). Examinations of ears in Harbour porpoises

Phocoena phocoena from the north and baltic seas. *Bioacoustics* **17**, 85–87. doi:

10.1080/09524622.2008.9753775.

Prenger-Berninghoff, E., Siebert, U., Stede, M., König, A., Weiß, R. and Baljer, G. (2008).

Incidence of Brucella species in marine mammals of the German North Sea. *Diseases of*

Aquatic Organisms **81**, 65–71. doi: 10.3354/dao01920.

Raga, J. A. (1994). Parasitismus bei den Cetacea. In *Robineau D, Duguy R, Klima M (eds.).*

Handbuch der Säugetiere Europas, pp. 132–179.

Raga, J. and Balbuena, J. (1987). Algunas características zoogeográficas de los helmintos de

los cetáceos en el Mediterráneo con especial referencia a la helmintofauna del delfín

listado. *Mamíferos y helmintos. Barcelona: Ketres Editora* **195**, 201.

Reckendorf, A., Ludes-Wehrmeister, E., Wohlsein, P., Tiedemann, R., Siebert, U. and

Lehnert, K. (2018). First record of Halocercus sp. (Pseudaliidae) lungworm infections in

two stranded neonatal orcas (Orcinus orca). *Parasitology* **145**, 1553–1557. doi:

10.1017/S0031182018000586.

Reckendorf, A., Everaarts, E., Bunskoek, P., Haulena, M., Springer, A., Lehnert, K.,

Lakemeyer, J., Siebert, U. and Strube, C. (2021). Lungworm infections in harbour

porpoises (Phocoena phocoena) in the German Wadden Sea between 2006 and 2018, and

serodiagnostic tests. *International Journal for Parasitology: Parasites and Wildlife* **14**,

53–61. doi: 10.1016/j.ijppaw.2021.01.001.

Regassa, A., Toyeb, M., Abebe, R., Megersa, B., Mekibib, B., Mekuria, S., Debela, E. and

Abunna, F. (2010). Lungworm infection in small ruminants: Prevalence and associated

risk factors in Dessie and Kombolcha districts, northeastern Ethiopia. *Veterinary*

Parasitology **169**, 144–148. doi: 10.1016/j.vetpar.2009.12.010.

Reisfeld, L., Sacristán, C., Sánchez-Sarmiento, A. M., Costa-Silva, S., Díaz-Delgado, J.,

Groch, K. R., Marigo, J., Ewbank, A. C., Favero, C. M., Guerra, J. M., Réssio, R.

A., Cremer, M. J., Esperón, F. and Catão-Dias, J. L. (2019). Fatal pulmonary

parafilaroidiasis in a free-ranging subantarctic fur seal (Arctocephalus tropicalis)

coinfected with two gammaherpesviruses and Sarcocystis sp. *Revista Brasileira de*

Parasitologia Veterinária **28**, 499–503. doi: 10.1590/s1984-29612019029.

Reyes, J. C. and van Waerebeek, K. (1995). Aspects of the biology of Burmeister's porpoise

from Peru. *Report of the International Whaling Commission* 349–364.

Rhyan, J., Garner, M., Spraker, T., Lambourn, D. and Cheville, N. (2018a). Brucella

pinnipedialis in lungworms Parafilaroides sp. and Pacific harbor seals Phoca vitulina

richardsi: proposed pathogenesis. *Diseases of Aquatic Organisms* **131**, 87–94. doi:

10.3354/dao03291.

Rhyan, J., Garner, M., Spraker, T., Lambourn, D. and Cheville, N. (2018b). Brucella

pinnipedialis in lungworms Parafilaroides sp. and Pacific harbor seals Phoca vitulina

richardsi: proposed pathogenesis. *Diseases of Aquatic Organisms* **131**, 87–94. doi:

10.3354/dao03291.

Rodrigues, T., Díaz-Delgado, J., Catão-Dias, J., da Luz Carvalho, J. and Marmontel, M.

(2018). Retrospective pathological survey of pulmonary disease in free-ranging Amazon

river dolphin Inia geoffrensis and tucuxi Sotalia fluviatilis. *Diseases of Aquatic*

Organisms **131**, 1–11. doi: 10.3354/dao03280.

Rogan, E., Baker, J. R., Jepson, P. D., Berrow, S. and Kiely, O. (1997). A mass stranding of

white-sided dolphins (Lagenorhynchus acutus) in Ireland: biological and pathological

studies. *Journal of Zoology* **242**, 217–227. doi: 10.1111/j.1469-7998.1997.tb05798.x.

Rogan, E., Penrose, R., Gassner, I., Mackey, M. J. and Clayton, P. (2001). *Marine Mammal*

Strandings: A Collaborative Study for the Irish Sea. The Marine Institute.

Rosas, F. C. W., Monteiro-Filho, E. L. A., Marigo, J., Santos, R. A., Andrade, A. L. V.,

Rautenberg, M., Olivcira, M. R. and Bordignon, M. O. (2002). The striped dolphin,

Stenella coeruleoalba (Cetacea: Delphinidae), on the coast of São Paulo State,

southeastern Brazil. *Aquatic Mammals* **28**, 60–66.

Ross, G. J. B. and Bass, A. J. (1971). Shark attack on an ailing dolphin, Stenella Coeruleoalba

(Meyen). *South African Journal of Science*.

Santos, C., Rohde, K., Ramos, R., Di Beneditto, A. and Capistrano, L. (1996). Helminths of

cetaceans on the Southeastern coast of Brazil. *Journal of the Helminthological Society of*

Washington **63**, 149–152.

Savage, K. N., Burek-Huntington, K., Wright, S. K., Bryan, A. L., Sheffield, G., Webber,

M., Stimmelmayr, R., Tuomi, P., Delaney, M. A. and Walker, W. (2021). Stejneger's

beaked whale strandings in Alaska, 1995–2020. *Marine Mammal Science* **37**, 843–869.

doi: 10.1111/mms.12780.

Schick, L., IJsseldijk, L. L., Grilo, M. L., Lakemeyer, J., Lehnert, K., Wohlsein, P., Ewers,

C., Prenger-Berninghoff, E., Baumgärtner, W., Gröne, A., Kik, M. J. L. and Siebert,

U. (2020). Pathological Findings in White-Beaked Dolphins (Lagenorhynchus albirostris)

and Atlantic White-Sided Dolphins (Lagenorhynchus acutus) From the South-Eastern

North Sea. *Frontiers in Veterinary Science* **7**, 262. doi: 10.3389/fvets.2020.00262.

Schmid-Hempel, P. (2021). Virulence evolution. In *Evolutionary Parasitology*, pp. 353–388.

Oxford University Press doi: 10.1093/oso/9780198832140.003.0013.

Schumacher, U., Horny, H., Heidemann, G., Schultz, W. and Welsch, U. (1990).

Histopathological findings in harbour seals (Phoca vitulina) found dead on the german

north sea coast. *Journal of Comparative Pathology* **102**, 299–309. doi: 10.1016/S0021-

9975(08)80019-9.

Seguel, M. and Gottdenker, N. (2017). The diversity and impact of hookworm infections in

wildlife. *International Journal for Parasitology: Parasites and Wildlife* **6**, 177–194. doi:

10.1016/j.ijppaw.2017.03.007.

Seguel, M., Nadler, S., Field, C. and Duignan, P. (2018a). Vasculitis and Thrombosis due to

the Sea Lion Lungworm, *Parafilaroides decorus* , in a Guadalupe Fur Seal (

Arctocephalus philippii townsendi). *Journal of Wildlife Diseases* **54**, 638–641. doi:

10.7589/2017-12-291.

Seguel, M., Nadler, S., Field, C. and Duignan, P. (2018b). Vasculitis and Thrombosis due to

the Sea Lion Lungworm, *Parafilaroides decorus* , in a Guadalupe Fur Seal (

Arctocephalus philippii townsendi). *Journal of Wildlife Diseases* **54**, 638–641. doi:

10.7589/2017-12-291.

Seguel, M., George, R. C., Maboni, G., Sanchez, S., Page-Karjian, A., Wirth, E., McFee, W.

and Gottdenker, N. L. (2020a). Pathologic findings and causes of death in bottlenose

dolphins Tursiops truncatus stranded along the Georgia coast, USA (2007−2013).

Diseases of Aquatic Organisms **141**, 25–38. doi: 10.3354/dao03509.

Seguel, M., George, R., Maboni, G., Sanchez, S., Page-Karjian, A., Wirth, E., McFee, W.

and Gottdenker, N. (2020b). Pathologic findings and causes of death in bottlenose

dolphins Tursiops truncatus stranded along the Georgia coast, USA (2007-2013).

Diseases of Aquatic Organisms **141**, 25–38. doi: 10.3354/dao03509.

Seguel, M., Budischak, S. A., Jolles, A. E. and Ezenwa, V. O. (2023). Helminth-associated

changes in host immune phenotype connect top-down and bottom-up interactions during

co-infection. *Functional Ecology* **37**, 860–872. doi: 10.1111/1365-2435.14237.

Seibel, H., Beineke, A. and Siebert, U. (2010). Mycotic Otitis Media in a Harbour Porpoise

(Phocoena phocoena). *Journal of Comparative Pathology* **143**, 294–296. doi:

10.1016/j.jcpa.2010.03.002.

Sheldon, J. D., Johnson, S. P., Hernandez, J. A., Cray, C. and Stacy, N. I. (2017). Acute-

phase responses in healthy, malnourished, and Otostrongylus-infected juvenile northern

elephant seals (mirounga angustirostris). *Journal of Zoo and Wildlife Medicine* **48**, 767–

775. doi: 10.1638/2016-0267.1.

Sheldon, J. D., Hernandez, J. A., Johnson, S. P., Field, C., Kaye, S. and Stacy, N. I. (2019).

Diagnostic Performance of Clinicopathological Analytes in Otostrongylus circumlitis-

Infected Rehabilitating Juvenile Northern Elephant Seals (Mirounga angustirostris).

Frontiers in Veterinary Science **6**, 134. doi: 10.3389/fvets.2019.00134.

Shiozaki, A. and Amano, M. (2017). Population- and growth-related differences in helminthic

fauna of finless porpoises (Neophocaena asiaeorientalis) in five Japanese populations.

Journal of Veterinary Medical Science **79**, 534–541. doi: 10.1292/jvms.16-0421.

Siebert, U., Wünschmann, A., Weiss, R., Frank, H., Benke, H. and Frese, K. (2001). Post-

mortem Findings in Harbour Porpoises (Phocoena phocoena) from the German North and

Baltic Seas. *Journal of Comparative Pathology* **124**, 102–114. doi:

10.1053/jcpa.2000.0436.

Siebert, U., Tolley, K., Víkingsson, G. A., Ólafsdottir, D., Lehnert, K., Weiss, R. and

Baumgärtner, W. (2006). Pathological Findings in Harbour Porpoises (Phocoena

phocoena) from Norwegian and Icelandic Waters. *Journal of Comparative Pathology*

134, 134–142. doi: 10.1016/j.jcpa.2005.09.002.

Siebert, U., Jepson, P. D. and Wohlsein, P. (2013). First indication of gas embolism in a

harbour porpoise (Phocoena phocoena) from German waters. *European Journal of*

Wildlife Research **59**, 441–444. doi: 10.1007/s10344-013-0700-4.

Siebert, U., Pawliczka, I., Benke, H., von Vietinghoff, V., Wolf, P., Pilāts, V., Kesselring, T.,

Lehnert, K., Prenger-Berninghoff, E., Galatius, A., Anker Kyhn, L., Teilmann, J.,

Hansen, M. S., Sonne, C. and Wohlsein, P. (2020). Health assessment of harbour

porpoises (PHOCOENA PHOCOENA) from Baltic area of Denmark, Germany, Poland

and Latvia. *Environment International* **143**, 105904. doi: 10.1016/j.envint.2020.105904.

Sierra, E., Zucca, D., Arbelo, M., García-Álvarez, N., Andrada, M., Déniz, S. and

Fernández, A. (2014). Fatal Systemic Morbillivirus Infection in Bottlenose Dolphin,

Canary Islands, Spain. *Emerging Infectious Diseases* **20**, 269–271. doi:

10.3201/eid2002.131463.

Smith, F. R. and Threlfall, W. (1973). Helminths of Some Mammals from Newfoundland.

American Midland Naturalist **90**, 215. doi: 10.2307/2424284.

Stephens, N., Duignan, P. J., Wang, J., Bingham, J., Finn, H., Bejder, L., Patterson, A. P.

and Holyoake, C. (2014). Cetacean Morbillivirus in Coastal Indo-Pacific Bottlenose

Dolphins, Western Australia. *Emerging Infectious Diseases* **20**, 672–676. doi:

10.3201/eid2004.131714.

Stijnen, T., Hamza, T. H. and Özdemir, P. (2010). Random effects meta-analysis of event

outcome in the framework of the generalized linear mixed model with applications in

sparse data. *Statistics in Medicine* **29**, 3046–3067. doi: 10.1002/sim.4040.

Stockin, K. A., Duignan, P. J., Roe, W. A., Meynier, L., Alley, M. and Fettermann, T.

(2009). Causes of mortality in stranded Common Dolphin (Delphinus sp.) from New

Zealand waters between 1998 and 2008. *Pacific Conservation Biology* **15**, 217. doi:

10.1071/PC090217.

Stroud, R. K. (1978). Parasites and associated pathology observed in pinnipeds stranded along

the Oregon coast. *Journal of Wildlife Diseases* **14**, 292–298. doi: 10.7589/0090-3558-

14.3.292.

Stroud, R. K. and Roffe, T. J. (1979). Causes of death in marine mammals stranded along the

oregon coast. *Journal of Wildlife Diseases* **15**, 91–97. doi: 10.7589/0090-3558-15.1.91.

Suvorova, I. V. and Prokushina, K. S. (2021). Pulmonary nematodiasis of the Baikal seal

(Pusa siberica). In *Theory and practice of parasitic disease control: Collection of*

Scientific Articles adapted from the International Scientific Conference., pp. 509–514.

Sweeney, J. C. and Gilmartin, W. G. (1974). Survey of diseases in free-living California sea

lions. *Journal of Wildlife Diseases* **10**, 370–376. doi: 10.7589/0090-3558-10.4.370.

Szefer, P., Rokicki, J., Frelek, K., Skora, K. and Malinga, M. (1998). Bioaccumulation of

selected trace elements in lung nematodes, Pseudalius inflexus, of harbor porpoise

(Phocoena phocoena) in a Polish zone of the Baltic Sea. *The Science of the Total*

Environment **220**, 19–24.

Tao, J.-Y. (1983). A new species and a new chinese record of nematodes from porpoise

neophocaenaphocaenoides. *Acta Zootaxonomica Sinica*.

Taylor, M. A., Coop, R. L. and Wall, R. L. eds. (2015). Veterinary Helminthology. In

Veterinary Parasitology, pp. 1–109. John Wiley & Sons, Inc., Hoboken, NJ, USA doi:

10.1002/9781119073680.ch1.

Terracciano, G., Fichi, G., Comentale, A., Ricci, E., Mancusi, C. and Perrucci, S. (2020).

Dolphins Stranded along the Tuscan Coastline (Central Italy) of the "Pelagos Sanctuary":

A Parasitological Investigation. *Pathogens* **9**, 612. doi: 10.3390/pathogens9080612.

Tomilin, A. (1967). Mammals of the USSR and adjacent countries, Vol. 9, Cetacea. *Israel*

program for scientific translations, Jerusalem **71**,.

Tomo, I., Kemper, C. M. and Lavery, T. J. (2010). Eighteen-year study of south Australian

dolphins shows variation in lung nematodes by season, year, age class, and location.

Journal of Wildlife Diseases **46**, 488–498. doi: 10.7589/0090-3558-46.2.488.

Torres, P., Cortes, P., Oporto, J. A., Brieva, L. and Silva, R. (1994). The occurrence of

stenurus australis Tantalean and Sarmiento, 1991(Nematoda: Mestratongyloidea)in the

Porpoise Phocoena spinipinnis (Burmeister,1865) on the Southern Coast of Chile.

Memórias do Instituto Oswaldo Cruz **89**, 141–143. doi: 10.1590/S0074-

02761994000200004.

Ulrich, S. A., Lehnert, K., Siebert, U. and Strube, C. (2015). A recombinant antigen-based

enzyme-linked immunosorbent assay (ELISA) for lungworm detection in seals. *Parasites*

& Vectors **8**, 443. doi: 10.1186/s13071-015-1054-4.

Ulrich, S. A., Lehnert, K., Rubio-Garcia, A., Sanchez-Contreras, G. J., Strube, C. and

Siebert, U. (2016). Lungworm seroprevalence in free-ranging harbour seals and

molecular characterisation of marine mammal MSP. *International Journal for*

Parasitology: Parasites and Wildlife **5**, 48–55. doi: 10.1016/j.ijppaw.2016.02.001.

Valderrama Vasquez, C. A., Macgregor, S. K., Rowcliffe, J. M. and Jepson, P. D. (2008).

Occurrence of a monophasic strain of Salmonella group B isolated from cetaceans in

England and Wales between 1990 and 2002. *Environmental Microbiology* **10**, 2462–

2468. doi: 10.1111/j.1462-2920.2008.01651.x.

van Elk, C. E., van de Bildt, M. W. G., van Run, P. R. W. A., Bunskoek, P., Meerbeek, J.,

Foster, G., Osterhaus, A. D. M. E. and Kuiken, T. (2019). Clinical, pathological, and

laboratory diagnoses of diseases of harbour porpoises (Phocoena phocoena), live stranded

on the Dutch and adjacent coasts from 2003 to 2016. *Veterinary Research* **50**, 88. doi:

10.1186/s13567-019-0706-3.

van Wijngaarden, M. F. A., Geut, M. I. M., Vernooij, J. C. M., IJsseldijk, L. L. and Tobias,

T. J. (2021). Determinants of mortality of juvenile harbour seals (Phoca vitulina) infected

with lungworm submitted to a Dutch seal rehabilitation centre. *International Journal for*

Parasitology: Parasites and Wildlife **14**, 1–6. doi: 10.1016/j.ijppaw.2020.12.002.

Vargas-Castro, I., Crespo-Picazo, J. L., Rivera-Arroyo, B., Sánchez, R., Marco-Cabedo, V.,

Jiménez-Martínez, M. Á., Fayos, M., Serdio, Á., García-Párraga, D. and Sánchez-

Vizcaíno, J. M. (2020). Alpha- and gammaherpesviruses in stranded striped dolphins

(Stenella coeruleoalba) from Spain: first molecular detection of gammaherpesvirus

infection in central nervous system of odontocetes. *BMC Veterinary Research* **16**, 288.

doi: 10.1186/s12917-020-02511-3.

Vercruysse, J., Salomez, A., Ulloa, A., Osterhaus, A., Kuiken, T. and Alvinerie, M. (2003).

Efficacy of ivermectin and moxidectin against Otostrongylus circumlitus and

Parafilaroides gymnurus in harbour seals (Phoca vitulina). *Veterinary Record* **152**, 130–

134. doi: 10.1136/vr.152.5.130.

Veryeri, N. G. (2012). Postmortem examinations of stranded dolphins found on the Black Sea

coast near Ordu, Turkey (Mammalia: Cetacea). *Zoology in the Middle East* **55**, 129–132.

doi: 10.1080/09397140.2012.10648928.

von Waerebeek, K., Reyes, J. C. and Shigueto, J. A. (1993). Helminth parasites and phoronts

of dusky dolphins Lagenorhynchus obscurus (Gray, 1828) from Peru. *Aquatic Mammals*

19, 159–169.

Walden, H. D. S., Grijalva, C. J., Páez-Rosas, D. and Hernandez, J. A. (2018). Intestinal

Parasites in Galapagos Sea Lions (Zalophus wollebaeki) Sivertsen, 1953 on San Cristóbal

Island, Galapagos, Ecuador. *Journal of Parasitology* **104**, 718–721. doi: 10.1645/17-187.

Walden, H. S., Bryan, A. L., McIntosh, A., Tuomi, P., Hoover-Miller, A., Stimmelmayr, R.

and Quakenbush, L. (2020). Helminth fauna of ice seals in the Alaskan Bering and

Chukchi seas, 2006–15. *Journal of Wildlife Diseases* **56**,. doi: 10.7589/2019-09-228.

Walker, W. A. (1975). Review of the Live-Capture Fishery for Smaller Cetaceans Taken in

Southern California Waters for Public Display, 1966–73. *Journal of the Fisheries*

Research Board of Canada **32**, 1197–1211. doi: 10.1139/f75-139.

Wan, X., Zheng, J., Li, W., Zeng, X., Yang, J., Hao, Y. and Wang, D. (2017). Parasitic

infections in the East Asian finless porpoise Neophocaena asiaeorientalis sunameri living

off the Chinese Yellow/Bohai Sea coast. *Diseases of Aquatic Organisms* **125**, 63–71. doi:

10.3354/dao03131.

Wazura, K. W., Strong, J. T., Glenn, C. L. and Bush, A. O. (1986). Helminths of the Beluga

Whale (Delphinapterus leucas) from the Mackenzie River Delta, Northwest Territories.

Journal of Wildlife Diseases **22**, 440–442. doi: 10.7589/0090-3558-22.3.440.

Webster, W., Neufeld, J. and MacNeill, A. (1973). Halocercus monoceris sp. n. (Nematoda:

Metastrongyloidea) from the Narwhal, Monodon monoceros. *Proceedings of the*

Helminthological Society of Washington **40**, 255–258.

Williams, K. M., Fessler, M. K., Bloomfield, R. A., Sandke, W. D., Malekshahi, C. R.,

Keroack, C. D., Duignan, P. J., Torquato, S. D. and Williams, S. A. (2020). A novel

quantitative real-time PCR diagnostic assay for fecal and nasal swab detection of an

otariid lungworm, Parafilaroides decorus. *International Journal for Parasitology:*

Parasites and Wildlife **12**, 85–92. doi: 10.1016/j.ijppaw.2020.04.012.

Wohlsein, P., Seibel, H., Beineke, A., Baumgärtner, W. and Siebert, U. (2019a).

Morphological and Pathological Findings in the Middle and Inner Ears of Harbour

Porpoises (Phocoena phocoena). *Journal of Comparative Pathology* **172**, 93–106. doi:

10.1016/j.jcpa.2019.09.005.

Wohlsein, P., Seibel, H., Beineke, A., Baumgärtner, W. and Siebert, U. (2019b).

Morphological and Pathological Findings in the Middle and Inner Ears of Harbour

Porpoises (Phocoena phocoena). *Journal of Comparative Pathology* **172**, 93–106. doi:

10.1016/j.jcpa.2019.09.005.

Woodard, J. C., Zam, S. G., Caldwell, D. K. and Caldwell, M. C. (1969). Some Parasitic

Diseases of Dolphins. *Pathologia veterinaria* **6**, 257–272. doi:

10.1177/030098586900600307.

WoRMS Editorial Board (2022). World Register of Marine Species. doi: 10.14284/170.

Wu, H. W. (1929). On Halocercus pingi n. sp. a Lung-Worm from the Porpoise, Neomeris

phocoenoides. *The Journal of Parasitology* **15**, 276. doi: 10.2307/3271983.

Wunschimann, A., Frese, K., Müiller, G., Baumgärtner, W., Siebert, U., Weiss, R.,

Lockyer, C. and Heide-Jørgensen, M. P. (2001). Evidence of infectious diseases in

harbour porpoises (Phocoena phocoena) hunted in the waters of Greenland and by-caught

in the German North Sea and Baltic Sea. *Veterinary Record* **148**, 715–720. doi:

10.1136/vr.148.23.715.

Wünschmann, A., Siebert, U. and Weiss, R. (1999). Rhizopusmycosis in a Harbor Porpoise

from the Baltic Sea. *Journal of Wildlife Diseases* **35**, 569–573. doi: 10.7589/0090-3558-

35.3.569.

Yamaguti, S. (1951). Studies on the helminth fauna of Japan. Part 46. Nematodes of marine

mammals. *Arbeiten aus der Medizinischen Fakultat Okayama* **7**, 295–306.

Yu, J., Sun, Y. and Xia, Z. (2009). The Rescue, Rehabilitation, and Release of a Stranded

Finless Porpoise (Neophocaena phocaenoides sunameri) at Bohai Bay of China. *Aquatic*

Mammals **35**, 220–225. doi: 10.1578/AM.35.2.2009.220.

Zafra, R., Jaber, J. R., Pérez, J., de la Fuente, J., Arbelo, M., Andrada, M. and Fernández,

A. (2015a). Immunohistochemical characterisation of parasitic pneumonias of dolphins

stranded in the Canary Islands. *Research in Veterinary Science* **100**, 207–212. doi:

10.1016/j.rvsc.2015.03.021.

Zafra, R., Jaber, J. R., Pérez, J., de la Fuente, J., Arbelo, M., Andrada, M. and Fernández,

A. (2015b). Immunohistochemical characterisation of parasitic pneumonias of dolphins

stranded in the Canary Islands. *Research in Veterinary Science* **100**, 207–212. doi:

10.1016/j.rvsc.2015.03.021.

Zam, S. G., Caldwell, D. K. and Caldwell, M. C. (1971). Some endoparasites from small

odontocete cetaceans collected in Florida and Georgia. *Cetology* **2**, 1–11.

Zylber, M. I., Failla, G. and Le Bas, A. (2002). Stenurus globicephalae Baylis et Daubney,

1925 (Nematoda: Pseudaliidae) from a False Killer Whale, Pseudorca crassidens

(Cetacea: Delphinidae), Stranded on the Coast of Uruguay. *Memórias do Instituto*

Oswaldo Cruz **97**, 221–225. doi: 10.1590/S0074-02762002000200015.

1459