A systematic review of the diversity and virulence correlates of metastrongyle

lungworms in marine mammals.

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Graphical Abstract:

Abstract

Metastrongyle lungworms could be particularly detrimental for diving animals such as marine

mammals, however little is known of the drivers of pathogenic host-parasite relationships in this

group. This systematic review analyzed the diversity of metastrongyles in marine mammals and

the host and parasite traits associated with virulence. There have been at least 40 species of

metastrongyles described in 66 species of marine mammals. After penalization for study biases,

Halocercus hyperoodoni, *Otostrongylus circumlitus*, *Parafilaroides gymnurus*, *Halocercus*

brasiliensis, and *Stenurus minor* were the metastrongyles with the widest host range. Most studies

(80.12%, n=133/166) reported that metastrongyles caused bronchopneumonia, while in the

cardiovascular system metastrongyles caused vasculitis in nearly half of the studies (45.45%,

n=5/11) that assessed these tissues. Metastrongyles were associated with otitis in 23.08% (n=6/26)

of the studies. Metastrongyle infection was considered a potential contributory to mortality in

44.78% (n=90/201) of the studies while 10.45% (n=21/201) of these studies considered

metastrongyles the main cause of death. Metastrongyle species with a wider host range were more

likely to induce pathogenic effects. Metastrongyles can cause significant tissue damage and

mortality in marine mammals although virulent host-parasite relationships are dominated by a few

metastrongyle species with wider host ranges.

Keywords: marine mammals, metastrongyles, lungworms, parasites, pathogenicity, virulence.

1. Introduction

Metastrongyloidea (Nematoda: Strongylida), also known as lungworms or metastrongyles, are common parasites of the respiratory, cardiovascular, and rarely the auditory system of mammals (Measures, 2001). The definitive hosts become infected through ingestion of larval stages within invertebrate intermediate hosts or free on pasture, therefore, metastrongyle infection can be quite common among free-foraging domestic and wild mammals (Measures, 2001; Regassa *et al.*, 2010; Niebuhr *et al.*, 2021). Like other helminth infections, the pathogenic effects of metastrongyloidiasis can be broad with impacts that range from subclinical infection to bronchopneumonia and death (Measures, 2001; Forbes, 2021). The reasons for these variable outcomes are not fully understood but site of infection (e.g. upper vs lower airways), burden and host immune status have been indicated as drivers of pathogenic outcomes (Forbes, 2021). However, it is unclear if certain host-parasite relationships are more detrimental for the host and if these more virulent associations obey to ecological factors that could illuminate conserved drivers of pathogenicity in these nematodes. Interestingly, the virulence of metastrongyles could be higher for animal species that require optimal respiratory and auditory function for fitness, such as marine mammals (Bergeron *et al.*, 1997b; Measures, 2001; Seibel *et al.*, 2010). Marine mammals rely on cardiovascular and lung capacity for foraging dives and the otic apparatus of cetaceans is critical for communication and echolocation in the water column (Bergeron *et al.*, 1997b; Measures, 2001; Seibel *et al.*, 2010). Therefore, metastrongyles parasitism of these host compartments could be associated with disease, stranding and death (Bergeron *et al.*, 1997b; Measures, 2001; Seibel *et al.*, 2010), making this animal group an interesting model to test conserved correlates of virulence among metastrongyle species. Nevertheless, metastrongyles are one of the most diagnosed parasites in marine mammals, and in many circumstances are considered

incidental findings of little to no pathologic significance (Geraci and St. Aubin, 1987). This

conflicting evidence could be related to negative metastrongyle effects limited to specific host-

parasite relationships, parasite traits, environmental conditions, or the research methods employed

to assess the health impacts. However, a systematic assessment of the factors that could modulate

the health impacts of metastrongyles has not been reported.

Three families of metastrongyles are prominently found in marine mammals, spanning

seven genera, with a considerable diversity of species within each genus. Despite this diversity,

metastrongyles share similar life histories and host exploitation strategies in marine mammals.

Most aquatic metastrongyle species require an obligate intermediate host, usually an invertebrate,

where several larval stages develop until reaching an infective stage (Dailey, 1970a, 2006a). These

infective larvae are consumed by the definitive host and then migrate from the gastrointestinal

system to the respiratory or cardiovascular system where sexual maturity and reproduction occurs,

releasing embryonated-larvated eggs or larvae (Taylor *et al.*, 2015). Additionally, in some species,

maternal lungworms infect neonates through the placenta and/or milk (Dailey *et al.*, 1991a). In

the airways and respiratory sinuses or cardiovascular system, metastrongyles feed on epithelial-

endothelial cells, host debris and occasionally blood (Dailey, 1970a; Fauquier *et al.*, 2010; Rhyan

et al., 2018a). These processes of migration through tissues, feeding and releasing eggs and larvae

illicit variable immune responses in the host (Baylis and Daubney, 1925; Fauquier *et al.*, 2010;

Groch *et al.*, 2020a). This immune response is usually directed against parasite cuticle, larvae,

eggs or parasite remains and usually consist in the attraction of macrophages, eosinophils and

lymphocytes to the site of infection (Fauquier *et al.*, 2010; Zafra *et al.*, 2015a; Seguel *et al.*, 2020a;

Groch *et al.*, 2020a). The degree of this inflammatory response and its impact on health status can

be quite variable ranging from minimal to severe bronchopneumonia (Fauquier *et al.*, 2010;

81 Seguel *et al.*, 2020a; Groch *et al.*, 2020a), being the drivers of the variable responses poorly

82 understood.

83   There is conflicting evidence regarding the connection between metastrongyle infections

84 and cetacean strandings (Arbelo *et al.*, 2013). Currently, there is no consensus on whether

85 inflammation in the otic apparatus, elicited by a presence of metastrongyle parasites, can impair

86 hearing or echolocation to a significant enough degree to be considered a contributory factor in

87 cetacean strandings (Geraci and St. Aubin, 1987; Wohlsein *et al.*, 2019a). Similarly, the presence

88 and feeding behavior of the nematodes in bronchi and lung parenchyma facilitates secondary

89 bacterial infections with conflicting documented outcomes, ranging from little importance for lung

90 physiology to fatal bronchopneumonia, especially when parasites are found in the lower

91 respiratory system (Siebert *et al.*, 2001; Lehnert *et al.*, 2010; Ulrich *et al.*, 2015, 2016; Reckendorf

92 *et al.*, 2021). This conflicting evidence could be related to several host and parasite traits, although

93 across biological systems, parasite biomass, usually assessed in terms of burden, is a common

94 driver of negative health outcomes (Pedersen and Fenton, 2015; Seguel and Gottdenker, 2017).

95 Similarly, parasite and host sizes play an important role in assessing total parasite biomass and can

96 sometimes serve as useful proxies of biomass to identify particularly detrimental host-parasite

97 relationships across studies (Seguel and Gottdenker, 2017). Another potential factor impacting

98 health outcomes across host-parasite systems is the capacity of certain helminths to parasitize

99 multiple host species. Parasites with a wider host range can potentially become more virulent

100 because decreases in host fitness due to infection can be compensated by infecting other

101 susceptible species (Poulin, 2011; Schmid-Hempel, 2021). However, the role of these host and

102 parasite traits on the health impact of metastrongyles is not well defined.

The objectives of this systematic review were to i) summarize existing literature concerning

relationships between marine mammals and metastrongyle parasites, ii) assess the relative host

and parasite diversity of infections iii) outline the pathology and health consequences that result

from metastrongyle infections in marine mammals, with assessment of the potential drivers of

these effects.

2. Materials and Methods

2.1 Searching methods and inclusion criteria

A systematic literature review on metastrongyle parasites in marine mammals was

conducted on July 6, 2021. The search was conducted using Google Scholar and PubMed,

following the best practices outlined by Haddaway and Watson (2016) to conduct an aggregative

systematic review in the field of parasitology (Haddaway and Watson, 2016) and the

recommendations outlined in the "Preferred Reporting Items for Systematic Reviews and Meta-

Analyses" (Page *et al.*, 2021).

The search terms used for the literature search on PubMed were as follows: "((Marine) OR

(Cetacean) OR (Delphinidae) OR (Phocidae) OR (Phocoenidae) OR (Otariidae) OR

(Monodontidae) OR (Mustelidae) OR (Trichechidae) OR (Odobenidae) OR (Ursidae) OR

(Odobenidae) OR (Balaenidae) OR (Neobalaenidae) OR (Eschrichtiidae) OR (Balaenopteridae)

OR (Physeteridae) OR (Kogiidae) OR (Ziphiidae) OR (Platanistidae) OR (Iniidae) OR (Lipotidae)

OR (Pontoporiidae) OR (Dugongidae) OR (Seal) OR (Porpoise) OR (Otter) OR (Dolphin) OR

(Sea lion) OR (Whale) OR (Walrus) OR (Manatee)) AND ((Metastrongyle) OR (Metastrongylus)

OR (Metastrongyloidea) OR (Metastrongylidae) OR (Parafilaroides) OR (Stenurus) OR

(Halocercus) OR (Pseudaliidae) OR (Filaroididae) OR (Crenosomatidae) OR (Otostrongylus) OR

(Skrjabingylus))".

The search terms used for the literature search on Google Scholar were reduced due to

limitations in the advanced search software, notably a 256-character limit on searches. The search

terms used for Google Scholar were as follows: "((Metastrongyle) OR (Metastrongylus) OR

(Metastrongyloidea) OR (Metastrongylidae) OR (Parafilaroides) OR (Stenurus) OR (Halocercus)

OR (Pseudaliidae) OR (Filaroididae) OR (Crenosomatidae) OR (Otostrongylus) OR

(Skrjabingylus)) AND ((Marine) OR (Cetacean))".

The search on PubMed yielded 120 results and the search on Google Scholar yielded 2110

results, excluding citations. As Google Scholar can only display 1000 results per search, only the

1000 articles deemed most relevant by the software were manually reviewed for inclusion in the

study, in addition to the 120 articles found via PubMed. To ensure all recently published literature

was consulted, a secondary search on google scholar was conducted with the same search terms,

filtering to only include articles published from 2017 and onward. This secondary search yielded

380 results, excluding citations, which were also manually reviewed for inclusion in the study. All

conference archives (e.g. IAAAM), theses not published in peer-reviewed journals, and posters

were excluded from this study. The full texts for the articles isolated through literature search

results were obtained and manually reviewed by the two authors, being included in the study if

they met the following inclusion criteria: i) The host species was any wild marine mammal, with

captive wild animals only included if the transmission of parasites was stated to have occurred

naturally, prior to their capture. ii) The parasite species belonged to a genus within the

metastrongyle superfamily, including studies that described the presence of parasitic eggs and

larvae provided they could be positively identified to at least the genus level (studies that described

148 parasites as "lungworms" or "nematodes" without specificity regarding genera were excluded). iii)

149 The article serves as the primary source describing a relationship between a marine mammal host

150 and metastrongyle parasite. Duplicate articles, existing in both the Google Scholar and PubMed

151 searches were manually removed. All non-primary sources which were found through the initial

152 search terms were screened, and citations were followed, leading to the inclusion of 15 additional

153 articles which met the inclusion criteria. Later, the World Register of Marine Species (WoRMS,

154 *https://www.marinespecies.org*), The Society for Marine Mammalogy

155 (*https://marinemammalscience.org*), and the Natural History Museum Host-Parasite database

156 (*https://www.nhm.ac.uk/research-curation/scientific-resources/taxonomy-systematics/host-*

157 *parasites/)* were consulted to ensure all known unique relationships were reported in this study

158 (WoRMS Editorial Board, 2022). A total of 35 articles could not be acquired in their full text form

159 for inclusion in data analysis, but the relationships they report are known through prior citations

160 in peer-reviewed journals or available abstracts. The articles which met the inclusion criteria were

161 further evaluated by the two authors, with significant attributes documented on a master

162 spreadsheet. Many included articles, including the articles not available in full-text form, report

163 multiple different host-parasite relationships, thus independent rows were created on the master

164 spreadsheet for each described host-parasite relationship (n=481), allowing for relationship-

165 specific attributes to be evaluated. The accepted common names and scientific names provided for

166 host species in this study are taken from the Society for Marine Mammalogy Committee on

167 Taxonomy (WoRMS Editorial Board, 2022).

168  A total of 207 full text publications met the inclusion criteria for this study, with 114

169 appearing only on Google Scholar, 6 appearing only on PubMed, 72 appearing on both Google

170 Scholar and PubMed, and 15 coming from other sources. Data was collected on all included full-

text publications, including whether they assessed pathogenic effect. An article was deemed to

assess pathogenic effect if it described some biological impact associated with a presence of

metastrongyle parasites in the host, or if it explicitly stated no pathogenic effect was associated

with the parasitism. Further data was collected on whether full-text publications which met the

inclusion criteria mentioned strandings or mortality. For the collection of this data, an article was

deemed to mention strandings if it contained the following terms (not including in the citations):

"stranded" or "stranding" or "strandings". For an article to have been deemed to mention mortality,

it must have contained the following terms (not including in the citations): "mortality" or

"mortalities". The inclusion terms for determining which articles mention stranding or mortality

were isolated through word searching in adobe acrobat, with technologically non-conforming

articles screened manually. Later, the studies mentioning mortality or stranding were assessed to

ensure they discussed these effects in the context of the study's findings. The parasite collection

methodology was also noted for each included full-text publication. Within the articles which met

the inclusion criteria, the geographical location(s) were recorded for 437 documented relationships

and the anatomical location(s) of the parasites were recorded in 371 documented relationships. We

further calculated the geographical dispersion of the 102 unique relationships between marine

mammals and metastrongyles.

For each documented relationship, data was also recorded on any reported pathogenic

effects (n=201). Pathogenic effects can be described as some biological impact on the host

associated with the presence of metastrongyle parasites. In cases where it was explicitly stated that

no pathogenic effect was associated with parasitism, this was recorded as well. Of the 101 articles

that assessed pathogenic effect, there were a total of 201 relationships in which attributes

describing a pathogenic effect or lack thereof were documented.

Between the 207 full-text publications which met our inclusion criteria, and the additional

35 publications we could not retrieve in full-text form, a total of 481 host-parasite relationships

are documented. For each of these relationships, data was collected on their respective species

taxonomy. Next, the frequency at which each metastrongyle parasite species appears in the

literature, in the context of relationships with marine mammal hosts was recorded. For the purposes

of frequency calculations, parasites described only to the genus level were excluded. The

frequency at which marine mammal hosts appear in the literature, in the context of relationships

with metastrongyle parasites was recorded in the same manner. For frequency calculations, hosts

described only to the genus level were excluded. For the purposes of all diversity calculations,

only relationships where both the host and parasite were described to the species level were

considered. Finally, host diversity was calculated as a measure of the diversity of host species a

metastrongyle species can infect, and parasite diversity was calculated as the diversity of

metastrongyle species a marine mammal species can host.

To provide a metric by which the size of a host could be used as a predictor of pathogenic

effects, host body mass was calculated by dividing the median weight in kilograms by the median

length in centimetres for each host species. Median host measurements were compiled as the

median of both genders from a variety of sources based on availability in the following order of

priority: 1) World Register of Marine Species (WoRMS, *https://www.marinespecies.org*); 2)

Society for Marine Mammalogy (*https://marinemammalscience.org/science-and-*

publications/species-information/facts/); 3) National Oceanic and Atmospheric Administration

(NOAA, *https://www.fisheries.noaa.gov/species-directory*); 4) Animal Diversity Web (ADW,

https://animaldiversity.org). In cases where data on host size was not available through those four

sources, the median measurements of hosts documented during gross necropsy in articles which

met the inclusion criteria of this study were used.

Finally, we produced an up-to-date checklist of all known relationships concerning

metastrongyles in marine mammals, which is included in table 1, complete with information on

anatomical ranges documented for each relationship. The relationships documented are found

within the 207 articles included in this study or within the 35 articles that could not be acquired in

their full text form for inclusion in data analysis, provided the relationships they report are known

through prior citations in peer-reviewed journals.

Table 1: Checklist of all known relationships concerning metastrongyles in marine mammals,

complete with information on anatomical ranges documented for each relationship.

Host Family	Host Species	Common Name	Parasite Family	Parasite species	Anatomical Location	Reference
Balaenopteridae	Megaptera novaeangliae	*Humpback Whale*	Pseudaliidae	*Pharurus alatus*	N/A	(Raga and Balbuena, 1987)
Delphinidae	Cephalorhynchus commersonii	*Commerson's Dolphin*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Greenwood and Taylor, 1979)
				Halocercus sp.	Lower respiratory	(Goodall *et al.*, 1988; Dougnac and Fredes, 2012)
				Stenurus sp.	Lower respiratory	(Duignan, 2000)
	Cephalorhynchus hectori	*Hector's dolphin*	Pseudaliidae	*Halocercus sp.*	Lower respiratory	(McKenzie and Blair, 1983; Duignan, 2000)
	Delphinus delphis	*Short-beaked common dolphin*	Pseudaliidae	*Halocercus cryptocephalus*	N/A	(Delyamure, 1968)
				Halocercus delphini	Lower respiratory	(Abollo *et al.*, 1998; Gibson *et al.*, 1998; Davison *et al.*, 2013; Pool *et al.*, 2020b, 2021)
				Halocercus invaginatus	N/A	(Raga and Balbuena, 1987)
				Halocercus kleinenbergi	N/A	(Harris, 1982)
				Halocercus lagenorhynchi	Lower respiratory	(Tomo *et al.*, 2010)
				Halocercus sp.	Lower respiratory	(Zafra *et al.*, 2015b; Lehnert *et al.*, 2017)

				Stenurus minor	N/A	(Raga, 1994)
				Stenurus ovatus	Lower respiratory	(Pool *et al.*, 2021)
				Stenurus sp.	Lower respiratory	(Díaz-Delgado *et al.*, 2012)
Delphinus sp.	*Common dolphin*	Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Stockin *et al.*, 2009)	
Feresa attenuata	*Pygmy killer whale*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Guimarães *et al.*, 2015)	
			Halocercus sp.	Lower respiratory	(Mignucci-Giannoni *et al.*, 1998b)	
			Stenurus globicephalae	Gastrointestinal, Lower respiratory, Otic apparatus	(Forrester *et al.*, 1980; Mignucci-Giannoni *et al.*, 1998b)	
Globicephala macrorhynchus	*Short-finned pilot whale*	Pseudaliidae	*Halocercus kleinenbergi*	Lower respiratory	(Carvalho *et al.*, 2010)	
			Stenurus globicephalae	Gastrointestinal, Otic apparatus	(Arnold and Gaskin, 1975; Mignucci-Giannoni *et al.*, 1998b; Carvalho *et al.*, 2010)	
			Stenurus minor	N/A	(Morales Vela and Olvera Gómez, 1993)	
			Stenurus sp.	Otic apparatus	(Huertas and Lagueux, 2016)	
			Torynurus convolutus	N/A	(Dougherty, 1943)	
Globicephala melas	*Long-finned pilot whale*	Pseudaliidae	*Stenurus globicephalae*	Lower respiratory, Otic apparatus	(Cowan, 1967; Arnold and Gaskin, 1975; McManus *et al.*, 1984; Abollo *et al.*, 1998; Gibson *et al.*, 1998; Lehnert *et al.*, 2017; Pool *et al.*, 2021)	
			Torynurus convolutus	Lower respiratory	(Baylis and Daubney, 1925; Baylis, 1932)	
Grampus griseus	*Risso's dolphin*	Pseudaliidae	*Halocercus delphini*	Lower respiratory	(Pool *et al.*, 2021)	
			Stenurus globicephalae	Gastrointestinal, Lower respiratory, Otic apparatus	(Arnold and Gaskin, 1975; Fernández *et al.*, 2003; Lehnert *et al.*, 2017; Pool *et al.*, 2021)	
			Stenurus minor	N/A	(Tomilin, 1967)	
Lagenodelphis hosei	*Fraser's dolphin*	Pseudaliidae	*Stenurus ovatus*	Lower respiratory	(McColl and Obendorf, 1982)	
Lagenorhynchus acutus	*White sided dolphin*	Pseudaliidae	*Halocercus lagenorhynchi*	N/A	(Raga and Balbuena, 1987)	
			Pseudalius inflexus	N/A	(Evans, 1991)	
			Stenurus globicephalae	Gastrointestinal, Otic apparatus	(Arnold and Gaskin, 1975; Beverley-Burton, 1978; Rogan *et al.*, 1997; Schick *et al.*, 2020)	
			Torynurus convolutus	N/A	(Gibson and Harris, 1979)	

	Lagenorhynchus albirostris	*White-beaked dolphin*	Pseudaliidae	*Halocercus lagenorhynchi*	Lower respiratory	(Baylis and Daubney, 1925; Gibson and Harris, 1979)
	Lagenorhynchus obscurus	*Dusky dolphin*	Pseudaliidae	*Halocercus sp.*	Gastrointestinal	(von Waerebeek *et al.*, 1993)
				Stenurus sp.	Gastrointestinal	(de Castro *et al.*, 2014)
	Lissodelphis borealis	*Northern right whale dolphin*	Pseudaliidae	*Halocercus sp.*	N/A	(Moser and Rhinehart, 1993)
	Orcinus orca	*Killer whale (Orca)*	Pseudaliidae	*Halocercus sp.*	Lower respiratory	(Reckendorf *et al.*, 2018)
	Peponocephala electra	*Melon-headed whale*	Pseudaliidae	*Halocercus sp.*	N/A	(Dailey and Brownell, 1972)
				Stenurus globicephalae	Otic apparatus	(Cannon, 1977; Mignucci-Giannoni *et al.*, 1998a; b; Carvalho *et al.*, 2010)
	Pseudorca crassidens	*False killer whale*	Pseudaliidae	*Stenurus auditivus*	N/A	(Zam *et al.*, 1971)
				Stenurus globicephalae	Otic apparatus	(Zylber *et al.*, 2002)
	Sotalia fluviatilis	*Tucuxi*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Santos *et al.*, 1996; Di Beneditto and Ramos, 2004; Melo *et al.*, 2006; Rodrigues *et al.*, 2018)
	Sotalia guianensis	*Guiana dolphin*	Pseudaliidae	*Halocercus brasiliensis*	Gastrointestinal, Lower respiratory	(Marigo *et al.*, 2010; Carvalho *et al.*, 2010; Guimarães *et al.*, 2013, 2015; Domiciano *et al.*, 2016; Groch *et al.*, 2018, 2020b; Carvalho Demarque *et al.*, 2020)
				Halocercus sp.	Lower respiratory	(Carvalho *et al.*, 2010)
	Sousa chinensis	*Indo-Pacific humpback dolphin*	Pseudaliidae	*Halocercus pingi*	Lower respiratory	(Parsons and Jefferson, 2000; Parsons *et al.*, 2001)
	Sousa plumbea	*Indian Ocean humpback dolphin*	Pseudaliidae	*Halocercus sp.*	Lower respiratory	(Lane *et al.*, 2014)
	Stenella attenuata	*Pantropical spotted dolphin*	Pseudaliidae	*Halocercus delphini*	Lower respiratory	(Dailey and Perrin, 1973)
				Halocercus sp.	Lower respiratory	(Oliveira *et al.*, 2011)
	Stenella clymene	*Clymene dolphin*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Carvalho *et al.*, 2010; Guimarães *et al.*, 2015)
				Halocercus delphini	Lower respiratory, Upper respiratory	(Aguilar-Aguilar *et al.*, 2010; Hernández-Orts *et al.*, 2021)
				Halocercus sp.	Lower respiratory	(Carvalho *et al.*, 2010)
	Stenella coeruleoalba	*Striped dolphin*	Pseudaliidae	*Halocercus brasiliensis*	Lower respiratory	(Rosas *et al.*, 2002)
				Halocercus delphini	Lower respiratory	(Lehnert *et al.*, 2017; Pool *et al.*, 2020a; b, 2021; Terracciano *et al.*, 2020)

				Halocercus lagenorhynchi	Lower respiratory	(Ross and Bass, 1971; Gibson et al., 1998; Oliveira et al., 2011; Terracciano et al., 2020)
				Halocercus sp.	Lower respiratory	(Abollo et al., 1998; Carvalho et al., 2010; Zafra et al., 2015b; Lehnert et al., 2017; Vargas-Castro et al., 2020; García de los Ríos y Loshuertos et al., 2021)
				Pharurus sp.	N/A	(García de los Ríos y Loshuertos et al., 2021)
				Pseudalius sp.	N/A	(García de los Ríos y Loshuertos et al., 2021)
				Stenurus minor	Gastrointestinal	(Aytemiz et al., 2012)
				Stenurus ovatus	Lower respiratory	(Pool et al., 2020a, 2021)
				Stenurus sp.	N/A	(García de los Ríos y Loshuertos et al., 2021)
	Stenella frontalis	Spotted dolphin	Pseudaliidae	Halocercus delphini	N/A	(Perrin et al., 1994)
				Halocercus lagenorhynchi	N/A	(Perrin et al., 1987)
				Halocercus sp.	Lower respiratory	(Zafra et al., 2015b)
				Pseudalius sp.	Lower respiratory	(Zafra et al., 2015b)
	Stenella longirostris	Spinner dolphin	Pseudaliidae	Halocercus brasiliensis	Lower respiratory	(Carvalho et al., 2010; Domiciano et al., 2016)
				Halocercus delphini	Lower respiratory	(Dailey and Perrin, 1973)
				Halocercus sp.	Lower respiratory	(Carvalho et al., 2010; Oliveira et al., 2011)
	Steno bredanensis	Rough-toothed dolphin	Pseudaliidae	Stenurus sp.	Otic apparatus	(Ewing et al., 2020)
	Tursiops aduncus	Indo-Pacific bottlenose dolphin	Pseudaliidae	Halocercus lagenorhynchi	Lower respiratory	(Tomo et al., 2010)
				Halocercus sp.	Lower respiratory	(Stephens et al., 2014; Lane et al., 2014)
				Stenurus ovatus	Lower respiratory	(Tomo et al., 2010)
	Tursiops truncatus	Common bottlenose dolphin	Pseudaliidae	Halocercus cryptocephalus	Lower respiratory	(Bowie, 1984; Fauquier et al., 2009)
				Halocercus delphini	Lower respiratory	(Abollo et al., 1998; Terracciano et al., 2020; Pool et al., 2021)
				Halocercus invaginatus	N/A	(Birkun and Krivokhizhin, 1996)

				Halocercus lagenorhynchi	Lower respiratory	(Woodard et al., 1969; Dailey et al., 1991b; Lipscomb et al., 1996; Mazzariol et al., 2007; McFee and Lipscomb, 2009; Fauquier et al., 2009)
				Halocercus sp.	Lower respiratory	(Sierra et al., 2014; Seguel et al., 2020b)
				Pharurus alatus	Lower respiratory	(Tomo et al., 2010)
				Stenurus minor	Lower respiratory	(Birincioğlu et al., 2017)
				Stenurus ovatus	Lower respiratory	(Bowie, 1984; Kumazawa et al., 2006; Kuwamura et al., 2007; Tomo et al., 2010; Terracciano et al., 2020; Pool et al., 2021)
				Stenurus sp.	Lower respiratory	(Sierra et al., 2014)
Iniidae	Inia geoffrensis	Amazon river dolphin	Pseudaliidae	Halocercus brasiliensis	Lower respiratory	(Rodrigues et al., 2018)
				Halocercus sp.	N/A	(Dailey and Brownell, 1972)
Kogiidae	Kogia breviceps	Pygmy sperm whale	Pseudaliidae	Halocercus sp.	Lower respiratory	(Carvalho et al., 2010)
Monodontidae	Delphinapterus leucas	Beluga whale	Pseudaliidae	Halocercus monoceris	Cardiovascular, Lower respiratory, Upper respiratory	(Measures et al., 1995)
				Halocercus taurica	Cardiovascular, Lower respiratory, Upper respiratory	(Measures et al., 1995)
				Pharurus pallasii	Central Nervous System, Lower respiratory, Otic apparatus	(Babero and Thomas, 1960; Arnold and Gaskin, 1975; Kenyon and Kenyon, 1977; Wazura et al., 1986; Measures et al., 1995; Houde et al., 2003)
				Stenurus arctomarinus	Cardiovascular, Lower respiratory, Upper respiratory	(Arnold and Gaskin, 1975; Measures et al., 1995; Burek-Huntington et al., 2015)
				Stenurus minor	N/A	(Kühn, 1829)
	Monodon monoceros	Narwhal	Pseudaliidae	Halocercus monoceris	Lower respiratory	(Webster et al., 1973; MacNeill et al., 1975)
				Pharurus alatus	N/A	(Arnold and Gaskin, 1975)
Otariidae	Arctocephalus australis	South american fur seal	Filaroididae	Parafilaroides sp.	Lower respiratory	(Jacobus et al., 2016)
				Parafilaroides normani	Lower respiratory	(Echenique et al., 2020)
	Arctocephalus forsteri	New zealand fur seal	Filaroididae	Parafilaroides normani	Lower respiratory	(Dailey, 2009)
	Arctocephalus gazella	Antarctic fur seal	Filaroididae	Parafilaroides normani	Lower respiratory	(Jacobus et al., 2016)

	Arctocephalus philippii	*Guadalupe Fur seal*	Filaroididae	*Parafilaroides decorus*	Cardiovascular, Lower respiratory	(Seguel *et al.*, 2018a; Williams *et al.*, 2020)
	Arctocephalus pusillus	*Australian fur seal*	Filaroididae	*Parafilaroides normani*	Lower respiratory	(Dailey, 2009; Jabbar *et al.*, 2014)
	Arctocephalus tropicalis	*Sub-antarctic fur seal*	Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Reisfeld *et al.*, 2019)
				Parafilaroides normani	Lower respiratory	(Dailey, 2009)
	Callorhinus ursinus	*Northern fur seal*	Filaroididae	*Parafilaroides decorus*	N/A	(Gerber *et al.*, 1993)
	Eumetopias jubatus	*Stellar sea lion*	Filaroididae	*Parafilaroides decorus*	Lower respiratory	(Stroud, 1978; Stroud and Roffe, 1979)
				Parafilaroides nanus	N/A	(Dougherty and Herman, 1947)
				Parafilaroides prolificus	N/A	(Dougherty and Herman, 1947)
	Neophoca cinerea	*Australian sea lion*	Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Nicholson and Fanning, 1981)
	Otaria flavescens	*South American sea lion*	Crenosomatidae	*Otostrongylus circumlitus*	N/A	(Ebmer *et al.*, 2020)
				Otostrongylus sp.	N/A	(Hermosilla *et al.*, 2016)
			Filaroididae	*Parafilaroides sp.*	N/A	(Hermosilla *et al.*, 2016)
	Phocarctos hookeri	*New zealand sea lion*	Filaroididae	*Parafilaroides decorus*	N/A	(Duignan, 2000)
	Zalophus californianus	*Californian sea lion*	Crenosomatidae	*Otostrongylus circumlitus*	Cardiovascular, Lower respiratory	(Kelly *et al.*, 2005; Goldstein *et al.*, 2011)
			Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Gulland *et al.*, 2012)
				Parafilaroides decorus	Lower respiratory	(Johnston and Ridgway, 1969; Dailey, 1970b; Fleischman and Squire, 1970; Migaki *et al.*, 1971; Sweeney and Gilmartin, 1974; Stroud, 1978; Gerber *et al.*, 1993; Haulena and Gulland, 2001; Fauquier *et al.*, 2004; Greig *et al.*, 2005; Dennison *et al.*, 2007; Evans, 2011; Kuzmina *et al.*, 2018; Williams *et al.*, 2020; Liu *et al.*, 2021)
	Zalophus wollebaeki	*Galapagos sea lion*	Crenosomatidae	*Otostrongylus sp.*	N/A	(Walden *et al.*, 2018)
			Filaroididae	*Parafilaroides sp.*	N/A	(Walden *et al.*, 2018)
Phocidae	Cystophora cristata	*Hooded seal*	Crenosomatidae	*Otostrongylus circumlitus*	Lower respiratory	(Lucas *et al.*, 2003)
			Filaroididae	*Parafilaroides gymnurus*	Lower respiratory	(Lucas *et al.*, 2003)

		Crenosomatidae	*Otostrongylus circumlitus*	N/A	(Antonelis *et al.*, 1994)
Erignathus barbatus	*Bearded seal*	Filaroididae	*Parafilaroides sp.*	Cardiovascular, Lower respiratory	(Walden *et al.*, 2020)
			Parafilaroides gymnurus	Lower respiratory	(Gosselin *et al.*, 1998; Walden *et al.*, 2020)
Halichoerus grypus	*Grey seal*	Crenosomatidae	*Otostrongylus circumlitus*	Cardiovascular, Lower respiratory	(Gosselin *et al.*, 1998; Ulrich *et al.*, 2015; Barnett *et al.*, 2019a, 2021)
			Otostrongylus sp.	Lower respiratory	(Baker, 1987)
		Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Baker, 1987; Barnett *et al.*, 2019a, 2021)
			Parafilaroides gymnurus	Lower respiratory	(Gosselin and Measures, 1997; Gosselin *et al.*, 1998; Ulrich *et al.*, 2015)
			Parafilaroides hispidus	Lower respiratory	(Gosselin and Measures, 1997; Gosselin *et al.*, 1998)
Histriophoca fasciata	*Ribbon seal*	Crenosomatidae	*Otostrongylus circumlitus*	N/A	(Walden *et al.*, 2020)
			Otostrongylus sp.	N/A	(Walden *et al.*, 2020)
		Filaroididae	*Parafilaroides gymnurus*	Lower respiratory	(Walden *et al.*, 2020)
Hydrurga leptonyx	*Leopard seal*	Filaroididae	*Parafilaroides hydrurgae*	Lower respiratory	(Mawson, 1953)
Mirounga angustirostris	*Northern elephant seal*	Crenosomatidae	*Otostrongylus circumlitus*	Cardiovascular, Gastrointestinal, Lower respiratory	(Stroud, 1978; Stroud and Roffe, 1979; Gulland *et al.*, 1997a; Elson-Riggins *et al.*, 2001; Fauquier *et al.*, 2003; Elson-Riggins *et al.*, 2004; Colegrove *et al.*, 2005; Kaye *et al.*, 2016; Sheldon *et al.*, 2017; Kaye *et al.*, 2017; Sheldon *et al.*, 2019)
			Otostrongylus sp.	N/A	(Gerber *et al.*, 1993)
		Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Stroud, 1978; Stroud and Roffe, 1979)
			Parafilaroides decorus	N/A	(Gerber *et al.*, 1993)
			Parafilaroides measuresae	Lower respiratory	(Dailey, 2006b)
Pagophilus groenlandicus	*Harp seal*	Crenosomatidae	*Otostrongylus circumlitus*	Lower respiratory, Upper respiratory	(Lucas *et al.*, 2003; Piché *et al.*, 2010)
		Filaroididae	*Parafilaroides gymnurus*	Lower respiratory	(Gosselin and Measures, 1997; Gosselin *et al.*, 1998; Lucas *et al.*, 2003)
Phoca largha	*Spotted seal*	Crenosomatidae	*Otostrongylus circumlitus*	N/A	(Delyamure *et al.*, 1984; Walden *et al.*, 2020)
		Filaroididae	*Parafilaroides gymnurus*	Lower respiratory	(Delyamure *et al.*, 1984; Walden *et al.*, 2020)

	Phoca vitulina	*Harbour seal*	Crenosomatidae	*Otostrongylus circumlitus*	Cardiovascular, Lower respiratory	(Dunn and Wolke, 1976; Kennedy *et al.*, 1989; Claussen *et al.*, 1991; Borgsteede *et al.*, 1991; Gerber *et al.*, 1993; Gosselin *et al.*, 1998; Elson-Riggins *et al.*, 2001, 2004; Fouchier *et al.*, 2001; Vercruysse *et al.*, 2003; Colegrove *et al.*, 2005; McKnight *et al.*, 2005; Lehnert *et al.*, 2007, 2010; Leidenberger and Boström, 2009; Lambourn *et al.*, 2013; Osinga *et al.*, 2015; Ulrich *et al.*, 2015, 2016; Kroese *et al.*, 2018)
				Otostrongylus sp.	N/A	(Herreman *et al.*, 2011)
			Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Garner *et al.*, 1997; Prenger-Berninghoff *et al.*, 2008; Herreman *et al.*, 2011; Lambourn *et al.*, 2013; Rhyan *et al.*, 2018b)
				Parafilaroides decorus	Lower respiratory	(Greig *et al.*, 2014)
				Parafilaroides gullandae	Lower respiratory	(Dailey, 2006b)
				Parafilaroides gymnurus	Lower respiratory, Reproductive	(Schumacher *et al.*, 1990; Claussen *et al.*, 1991; Borgsteede *et al.*, 1991; Gosselin and Measures, 1997; Gosselin *et al.*, 1998; Fouchier *et al.*, 2001; Vercruysse *et al.*, 2003; Lehnert *et al.*, 2007, 2010; Osinga *et al.*, 2015; Ulrich *et al.*, 2015; Elson-Riggins *et al.*, 2020)
	Pusa caspica	*Caspian seal*	Filaroididae	*Parafilaroides caspicus*	N/A	(Kurochkin and Zablotsky, 1958)
	Pusa hispida	*Ringed seal*	Crenosomatidae	*Otostrongylus circumlitus*	Lower respiratory	(Onderka, 1989; Measures and Gosselin, 1994; Bergeron *et al.*, 1997a; b; Walden *et al.*, 2020)
			Filaroididae	*Parafilaroides sp.*	Lower respiratory	(Walden *et al.*, 2020)
				Parafilaroides gymnurus	Gastrointestinal, Lower respiratory	(Measures and Gosselin, 1994; Gosselin and Measures, 1997; Gosselin *et al.*, 1998; Walden *et al.*, 2020)
				Parafilaroides hispidus	Lower respiratory	(Kennedy, 1986; Onderka, 1989; Gosselin and Measures, 1997; Gosselin *et al.*, 1998)
	Pusa sibirica	*Baikal seal*	Crenosomatidae	*Otostrongylus circumlitus*	Lower respiratory	(Suvorova and Prokushina, 2021)
			Filaroididae	*Parafilaroides gymnurus*	Lower respiratory	(Kontrimavichus and Delyamure, 1979; Suvorova and Prokushina, 2021)
Phocoenidae	Neophocaena asiaeorientalis	*Narrow-ridged finless porpoise*	Pseudaliidae	*Halocercus pingi*	Lower respiratory, Reproductive	(Shiozaki and Amano, 2017)
				Halocercus sp.	Lower respiratory	(Wan *et al.*, 2017; Gui *et al.*, 2018)

				Halocercus sunameri	Lower respiratory	(Shiozaki and Amano, 2017)
				Halocercus taurica	Lower respiratory	(Shiozaki and Amano, 2017)
				Pharurus asiaeorientalis	Lower respiratory, Otic apparatus	(Shiozaki and Amano, 2017)
				Pharurus sunameri	Lower respiratory, Otic apparatus	(Shiozaki and Amano, 2017)
				Stenurus nanjingensis	Otic apparatus	(Shiozaki and Amano, 2017)
	Neophocaena phocaenoides	*Indo-Pacific finless porpoise*	Crenosomatidae	*Otostrongylus circumlitus*	N/A	(Yu *et al.*, 2009)
			Pseudaliidae	*Halocercus pingi*	Lower respiratory	(Wu, 1929; Parsons and Jefferson, 2000; Parsons *et al.*, 2001)
				Halocercus sp.	Lower respiratory	(Gui *et al.*, 2018)
				Halocercus sunameri	N/A	(Yamaguti, 1951)
				Halocercus taurica	N/A	(Kuramochi *et al.*, 2000)
				Pharurus asiaeorientalis	N/A	(Petter and Pilleri, 1982)
				Pharurus sunameri	N/A	(Yamaguti, 1951)
				Stenurus auditivus	N/A	(Hsü and Hoeppli, 1933)
				Stenurus nanjingensis	N/A	(Tao, 1983)
	Phocoena dioptrica	*Spectacled porpoise*	Pseudaliidae	*Stenurus minor*	Otic apparatus	(Lehnert *et al.*, 2017)
	Phocoena phocoena	*Harbour Porpoise*	Pseudaliidae	*Halocercus invaginatus*	Cardiovascular, Lower respiratory, Upper respiratory	(Smith and Threlfall, 1973; Andersen, 1974; Arnold and Gaskin, 1975; Dailey and Stroud, 1978; Stroud and Roffe, 1979; Kastelein and Lavaleije, 1992; Moser and Rhinehart, 1993; Balbuena *et al.*, 1994; Abollo *et al.*, 1998; Gibson *et al.*, 1998; Jepson *et al.*, 2000; Rogan *et al.*, 2001; Lehnert *et al.*, 2005, 2010, 2014; Siebert *et al.*, 2006, 2020; Veryeri, 2012; Pekmezci *et al.*, 2013)
				Halocercus kirbyi	N/A	(Dougherty, 1944)
				Halocercus sp.	Lower respiratory	(Smith and Threlfall, 1973; Wunschimann *et al.*, 2001; Jauniaux *et al.*, 2002; Dagleish *et al.*, 2008; Valderrama Vasquez *et al.*, 2008; Fenton *et al.*, 2017; van Elk *et al.*, 2019)

				Halocercus taurica	Lower respiratory	(Arnold and Gaskin, 1975; Gibson et al., 1998; Jepson et al., 2000; Rogan et al., 2001; Pekmezci et al., 2013)
				Pharurus dalli	Lower respiratory, Otic apparatus	(Smith and Threlfall, 1973; Colón-Llavina et al., 2019)
				Pharurus sp.	N/A	(Smith and Threlfall, 1973)
				Pseudalius inflexus	Cardiovascular, Gastrointestinal, Lower respiratory, Otic apparatus	(Andersen, 1974; Arnold and Gaskin, 1975; Kastelein and Lavaleije, 1992; Balbuena et al., 1994; Brosens et al., 1996; Szefer et al., 1998; Gibson et al., 1998; Wünschmann et al., 1999; Jepson et al., 2000; Siebert et al., 2001, 2006, 2013, 2020; Wunschimann et al., 2001; Rogan et al., 2001; Jauniaux et al., 2002; Lehnert et al., 2005, 2010; Dawson et al., 2008; Dagleish et al., 2008; Valderrama Vasquez et al., 2008; Davison et al., 2010a; b; Seibel et al., 2010; Fenton et al., 2017; van Elk et al., 2019; Gabel et al., 2021)
				Stenurus minor	Gastrointestinal, Lower respiratory, Otic apparatus, Upper Respiratory	(Andersen, 1974; Arnold and Gaskin, 1975; Dailey and Stroud, 1978; Stroud and Roffe, 1979; Faulkner, 1995; Brosens et al., 1996; Faulkner et al., 1998; Gibson et al., 1998; Wünschmann et al., 1999; Jepson et al., 2000; Siebert et al., 2001, 2006, 2013, 2020; Wunschimann et al., 2001; Rogan et al., 2001; Jauniaux et al., 2002; Kijewska, 2003; Lehnert et al., 2005, 2010, 2014; Prahl et al., 2008; Valderrama Vasquez et al., 2008; Seibel et al., 2010; Pekmezci et al., 2013; Fenton et al., 2017; Morell et al., 2017; Wohlsein et al., 2019b; van Elk et al., 2019; Colón-Llavina et al., 2019; Gabel et al., 2021)
				Stenurus sp.	Lower respiratory	(Fenton et al., 2017)
				Torynurus convolutus	Cardiovascular, Gastrointestinal, Lower respiratory, Otic apparatus, Upper respiratory	(Arnold and Gaskin, 1975; Dailey and Stroud, 1978; Kastelein and Lavaleije, 1992; Balbuena et al., 1994; Brosens et al., 1996; Faulkner et al., 1998; Gibson et al., 1998; Wünschmann et al., 1999; Jepson et al., 2000; Siebert et al., 2001, 2006, 2013, 2020; Wunschimann et al., 2001; Rogan et al., 2001; Jauniaux et al., 2002; Lehnert et al., 2005, 2010; Valderrama Vasquez et al., 2008; Davison et al., 2010a; Seibel et al., 2010; Fenton et al., 2017; van Elk et al., 2019)
	Phocoena spinipinnis	*Burmeister's porpoise*	Pseudaliidae	*Halocercus sp.*	Lower respiratory	(Reyes and van Waerebeek, 1995)

				Pseudalius inflexus	Lower respiratory	(Corcuera *et al.*, 1995; Reyes and van Waerebeek, 1995; Alvarado-Rybak *et al.*, 2020b)
				Stenurus australis	Lower respiratory, Otic apparatus	(Torres *et al.*, 1994; Alvarado-Rybak *et al.*, 2020b)
				Stenurus minor	Otic apparatus	(Corcuera *et al.*, 1995)
				Stenurus sp.	Otic apparatus	(Reyes and van Waerebeek, 1995)
	Phocoenoides dalli	*Dall's porpoise*	Pseudaliidae	*Halocercus dalli*	Lower respiratory	(Conlogue *et al.*, 1985)
				Halocercus kirbyi	N/A	(Dougherty, 1944)
				Halocercus sp.	Lower respiratory	(Migaki *et al.*, 1971)
				Pharurus dalli	Otic apparatus	(Arnold and Gaskin, 1975; Dailey and Walker, 1978; Kuramochi *et al.*, 1990)
				Pharurus sp.	N/A	(Walker, 1975)
				Stenurus minor	Lower respiratory, Otic apparatus	(Johnston and Ridgway, 1969; Migaki *et al.*, 1971; Dailey and Walker, 1978)
				Stenurus truei	Otic apparatus	(Kuramochi *et al.*, 1990)
				Stenurus yamagutii	Otic apparatus	(Kuramochi *et al.*, 1990)
Physteridae	Physeter macrocephalus	*Sperm whale*	Pseudaliidae	*Stenurus ovatus*	N/A	(Raga and Balbuena, 1987)
Ziphiidae	Berardius bairdii	*Baird's beaked whale*	Pseudaliidae	*Halocercus hyperoodoni*	N/A	(Delyamure, 1968)
	Mesoplodon sp.	*Unidentified whale*	Pseudaliidae	*Halocercus sp.*	Lower respiratory	(Moser and Rhinehart, 1993)
	Mesoplodon stejnegeri	*Stejneger's beaked whale*	Pseudaliidae	*Halocercus sp.*	Lower respiratory	(Savage *et al.*, 2021)

2.2 Data management and analyses

When assessing the relative host and parasite diversities, corrections were made for literature and intra-study biases which arose from non-uniform sampling efforts. We used a modified approach, based previously published methods (Nunn *et al.*, 2003; Ezenwa *et al.*, 2006; Seguel and Gottdenker, 2017), to create a bias penalization index for each host and parasite species. To

do this, we searched scientific names of each host and parasite species individually on Google

Scholar and the number of hits was recorded. The number of hits for each species was then

multiplied by the number of studies that both met our inclusion criteria and contained mentions of

a relationship concerning that respective species. The natural logarithm of these products was then

taken, and the result constitutes our penalization index for each species study bias (correction-

index). The resulting corrected diversity indexes were plotted using frequency plots constructed in

the R package "ggplo2".

We explored the predictors of pathogenic effects among different host parasite interactions

using binomial generalized linear mixed effects models (GLMMs) with the restricted maximum

likelihood fitted using the R package "glmmTMB" (Brooks *et al.*, 2017). Although this method

can estimate positively biased odds ratios for binary outcomes in metadata, these are usually more

problematic for smaller samples sizes and alternative methods for the analysis of binary metadata

present similar issues (Stijnen *et al.*, 2010; Bakbergenuly and Kulinskaya, 2018). We used

corrected host diversity, corrected parasite diversity, and host body size as predictors for the

presence or absence of pathogenic effects. Study ID was added as a random effect to account for

studies documenting multiple host-parasite relationships with their respective outcomes. For each

documented host-parasite relationship, with both the host and parasite specified to the species level

and for which pathogenic effect was assessed (n=165), binomial GLMMs for the presence or

absence of pathogenic effect were fit. A second model was fit to exclusively assess the presence

or absence of respiratory specific effect (bronchopneumonia or bronchitis). For the respiratory

specific effect model, each documented host-parasite relationship was included if the host and

parasite were specified to the species level, pathogenic effect was assessed, and the presence of

the parasite was documented in either the upper or lower respiratory system (n=134). We attempted

to fit cardiovascular specific effect (DIC, thrombosis, vasculitis, hemorrhage, endocarditis) (n=11)

and otic apparatus specific effect (sinusitis, otitis, otic metaplasia) (n=24) models using the same

method as for respiratory specific effect but were unable because the small sample size affected

model convergence. In both the full and respiratory specific models, we included interaction

effects between the corrected host diversity, corrected parasite diversity, and body size predictors

terms to account for individual permissiveness of host species to parasite species as well as

individual pervasiveness of parasite species to host species.

2.3 Discrepancies in the literature

Hermosilla, C., et al. (2015) reports *Stenurus sp*. in sei whale (*Balaenoptera borealis*), blue

whale (*Balaenoptera musculus*), and fin whale (*Balaenoptera physalus*) as non-primary findings,

however we could not identify a primary source of these relationships (Hermosilla *et al.*, 2015).

Lehnert, K., et al. (2019) reports *Pharurus alatus* as a parasite of short-beaked common dolphin

(*Delphinus delphis*) citing Tomo, I., et al. (2010) (Tomo *et al.*, 2010; Lehnert *et al.*, 2019). Upon

reviewing Tomo, I., et al. (2010), it does not appear to include this relationship (Tomo *et al.*, 2010).

The World Register of Marine Species (WoRMS) documents *Halocercus taurica* in of short-

beaked common dolphin (*Delphinus delphis*) and *Halocercus hyperoodoni* in northern bottlenose

whale (*Hyperoodon ampullatus*), however primary sources could not be identified for either of

these relationships (WoRMS Editorial Board, 2022). Measures, L.N. (2001) documents

Halocercus pingi in Dall's porpoise (*Phocoenoides dalli*) as a non-primary finding, however we

could not find a primary source for this relationship (Measures, 2001). Three (n=3) additional

publications document relationships that were not specified to the genus level (Giorda *et al.*, 2017;

Reckendorf *et al.*, 2021; van Wijngaarden *et al.*, 2021).

3. Results

At least 40 species of metastrongyles have been described in 66 marine mammal host species.

Most metastrongyle species parasitize multiple host species, with a total of 213 unique host-

metastrongyle relationships recorded (table 1).

3.1 Characterization of studies

The mean publication year of the 207 articles which met the inclusion criteria was 2003,

with the distribution of publication years skewed towards the most recent decade (Figure 1). For

each of the articles which met the inclusion criteria, 48.88% (n=101/207) assessed pathogenic

effect. Additionally, 78.74% (n=163/207) of studies mentioned stranding and 43.48% (n=90/207)

of articles mentioned mortality as potentially related to lungworm infection. Gross necropsy is the

predominant parasite collection method for metastrongyles in marine mammals with 92.27%

(n=191/207) of the studies using this as the only method of collection. The next most common

method of collection was, fecal sampling, in 6.28% (n=13/207) of the studies (Figure 2).

Figure 1: Distribution of publication year for articles which met the inclusion criteria of this

study. The mean year of publication was 2003.

Figure 2: Parasite collection methodologies documented, with intersections between the

methodologies shown to represent studies that utilize more than one methodology. Articles that

did not report a collection methodology are designated as "Not Reported".

A total of 175 host-metastrongyle relationships were reported in North America (including

the commonly referenced Central America region), and 169 relationships were reported in Europe.

North America and Europe are the most common locations reported for metastrongyle infections

in marine mammals, followed by 54 relationships in South America, 24 in Asia, 23 in Oceania and

11 in Africa (Figure 3). Exactly half (50.00%, n=51/102) of all unique relationships are reported

in North America, with 29.41% (n=30/102) of these relationships being reported exclusively in

North America (Figure 4).

Figure 3: Geographical location(s), denoted by continent, of documented relationships between

metastrongyles and marine mammals. Documented relationships include duplicates of unique

relationships published independently. Intersections are shown for studies which report

individual host species to parasite species relationships occurring in multiple locations.

Figure 4: Geographical location(s), denoted by continent, of all unique relationships between

metastrongyles and marine mammals. Intersections are shown for studies which report individual

host species to parasite species relationships occurring in multiple locations.

3.2 Frequencies and diversities of metastrongyle infections in marine mammals

Pseudaliidae was the most reported metastrongyle family in marine mammals (n=331),

followed by Filaroididae (n=90), and Crenosomatidae (n=58). At the host family level,

Delphinidae (n=159), Phocoenidae (n=150), and Phocidae (n=107) are most frequently parasitized

by metastrongyles. Further, metastrongyles from the Pseudaliidae family are reported in hosts from

the Delphinidae (n=158), Phocoenidae (n=148), Monodontidae (n=15), Ziphiidae (n=3), Iniidae

(n=2), Balaenopteridae (n=1), Kogiidae (n=1), and Physeteridae (n=1) families. Metastrongyles

from the Crenosomatidae family are reported to parasitize hosts from the Phocidae (n=52),

Otariidae (n=5), and Phocoenidae (n=1) families. Metastrongyles from the Filaroididae family are

reported to parasitize hosts from the Phocidae (n=54), Otariidae (n=35), and Delphinidae (n=1)

families.

Otostrongylus circumlitus (n=52) is the most reported metastrongyle species followed by

Stenurus minor (n=42) and *Pseudalius inflexus* (n=31) (Supplementary Figure 1). The harbour

porpoise (*Phocoena phocoena*) (n=118) is the most reported host, followed by harbour seal (*Phoca*

vitulina) (n=41) and bottlenose dolphin (*Tursiops truncat*us) (n=23) (Supplementary Figure 2).

Before penalizing for literature and intra-study biases, the host species harbouring the largest

number of metastrongyle species (parasite diversity) were: indo-pacific finless porpoise

(*Neophocaena phocaenoides*) (n=8), short-beaked common dolphin (*Delphinus delphis*) (n=7),

harbour porpoise (*Phocoena phocoena*) (n=7), and bottlenose dolphin (*Tursiops truncatus*) (n=7)

(Supplementary Figure 3). After correction for publication bias, indo-pacific finless porpoise

(*Neophocaena phocaenoides*) remained the most permissive host (Figure 5.). Before penalization,

the metastrongyle species with the broader host diversity were: *Otostrongylus circumlitus* (n=13),

*Stenurus mino*r (n=10), *Parafilaroides gymnurus* (n=9), *Halocercus brasiliensis* (n=8), and

Halocercus delphini (n=8) (Supplementary Figure 4). After correction for publication bias,

Halocercus hyperdooni emerged as the most generalist lungworm species, followed closely by

Otostrongylus circumlitus and *Parafilaroides gymnurus* (Figure 6.).

Figure 5: Parasite diversity for metastrongyles in marine mammals, corrected for publication bias (correction-index).

Figure 6: Host diversity for metastrongyles in marine mammals, corrected for publication bias
(correction-index).

3.3 The impact of metastrongyle infections on marine mammals

Of the 481 host parasite relationships documented, 371 (77.13%) reported an anatomical
location. Lungworms were found in the lower respiratory system in 82.48% (n=306/371) of cases,
while parasites were found in the otic apparatus in 17.52% (n=65/371) of cases. In 11.59%

(n=43/371) of cases, a single species of parasite parasitized an individual host in multiple

anatomical locations (Figure 7).

Figure 7: Anatomical location(s) of documented relationships between metastrongyles and

marine mammals. Intersections are shown for studies which report individual host species to

parasite species relationships occurring in multiple locations. Relationships that did not report an

anatomical location are designated as "Not Reported". (CNS= Central nervous system, G.I=

Gastrointestinal tract, C.V= Cardiovascular system).

Out of the 201 unique host-parasite relationships assessed, 44.77% (n=90/201) mentioned

mortality as "possible" or "potential" contributory cause of death and only 10.44% (n=21/201)

studies considered metastrongyle infection as the primary cause of death. Of the 201 host-parasite

relationships for which pathogenic effect was assessed, bronchopneumonia was described in

71.64% (n=144/201) of the relationships, indicating a high prevalence of bronchopneumonia in

hosts infected with metastrongyles (Figure 8). Other reported pathogenic effects included edema

(10.95%, n=22/201), vasculitis (7.46%, n=15/201), thrombosis (5.97%, n=12/201), and

hemorrhage (5.97%, n=12/201). Lungworm parasitism resulted in no discernible pathogenic effect

on the host in 16.42% (n=33/201) of the relationships.

Figure 8: Pathogenic effect(s) of documented relationships between metastrongyles and marine

mammals. Intersections are shown for studies which report individual host species to parasite

species relationships causing multiple pathogenic effects. Relationships were designated as "No.

P.E." when pathogenic effect was assessed and was determined to be negligible. (DIC=

Disseminated Intravascular Coagulation).

Pathogenic effect was assessed for 166 host-parasite relationships in the respiratory system,

with respiratory specific pathology reported in 80.12% (n=133/166) of these cases. In contrast, 26

host-parasite relationships assessed pathogenic effect in the otic apparatus, with otic specific

lesions reported in only 23.08% (n=6/26) of these relationships.

In the binomial GLMM that assessed pathogenic effect, host body size, corrected host

diversity and corrected parasite diversity were all significant predictors of the presence of

metastrongyle pathogenic effect in marine mammals, with a significant effect of the interaction

between host and parasite diversity indexes (table 2). For respiratory specific effects, corrected

parasite diversity and corrected host diversity main effects and their interaction were significant

predictors of metastrongyle respiratory specific effect in marine mammals, but host body size was

not a significant predictor of respiratory specific effect (table 3).

Table 2: Binomial generalized linear mixed effects model (GLMM) for presence of pathogenic

effect. Presented are predictors estimates, standard error (SE), statistic (Z) and P values. The

model used study ID as a random effect (total observations n=165, study ID n=89).

Predictor	Estimate	SE	Z	P
(Intercept)	-189.93	65.03	-2.921	0.00349
Corrected Parasite Diversity	523.62	165.18	3.170	0.00152
Corrected Host Diversity	232.05	88.44	2.624	0.00869
Host Body Size	166.90	83.04	2.010	0.04444
Corrected Parasite Diversity : Corrected Host Diversity	-625.90	250.44	-2.499	0.01245
Corrected Parasite Diversity : Host Body Size	-435.38	220.87	-1.971	0.04870
Corrected Host Diversity : Host Body Size	-189.10	131.22	-1.441	0.14953
Corrected Parasite Diversity : Corrected Host Diversity : Host Body Size	581.44	438.86	1.325	0.18521

402 Table 3: Binomial generalized linear mixed effects model (GLMM) for respiratory pathogenic

403 effects. Presented are predictors estimates, standard error (SE), statistic (Z) and P values. The

404 model used study ID as a random effect (total observations n=134, study ID n=76).

Predictor	Estimate	SE	Z	P
(Intercept)	0.09403	12.65860	0.007	0.99407
Corrected Parasite Diversity	88.93332	41.43432	2.146	0.03184
Corrected Host Diversity	74.10991	26.73963	2.772	0.00558
Host Body Size	-18.39291	13.61594	-1.351	0.17675
Corrected Parasite Diversity : Corrected Host Diversity	-244.76967	87.16180	-2.808	0.00498
Corrected Parasite Diversity : Host Body Size	8.03858	38.70274	0.208	0.83546
Corrected Host Diversity : Host Body Size	-29.25975	24.54618	-1.192	0.23325
Corrected Parasite Diversity : Corrected Host Diversity : Host Body Size	154.98741	93.04556	1.666	0.09577

4. Discussion and Conclusions

Metastrongyles are common helminth parasites of the respiratory and cardiovascular system

of mammals. Given the importance of these body systems for the survival of diving animals, it

was expected that the health impacts of metastrongyles would be particularly significant in marine

mammals. Our findings confirm that pathogenic effects, mainly bronchopneumonia, are common

in marine mammals infected with metastrongyles, although association of metastrongyle parasites

and mortality is usually not definitive. Importantly, our review suggests that pathogenic host-

parasite relationships are associated to hosts that can harbor a wider range of metastrongyle species

(parasite diversity). From the parasite perspective, metastrongyle species with a broader host

diversity tend to be more pathogenic. These results emphasize the importance of host and parasite

diversity with regards to the virulence of metastrongyles in marine ecosystems.

Most parasitology studies in marine mammals have been conducted through the opportunistic

collection and examination of stranded carcasses. This probably explains in part why in most cases

metastrongyle infections are suggested or considered as a contributory cause of death but rarely

confirmed as the primary cause of mortality. Determining cause of death in stranded marine

mammals is challenging, amid the rapid rate of postmortem changes in these species and little

background information for most cases (Arbelo *et al.*, 2013; Alvarado-Rybak *et al.*, 2020a).

Additionally, access to carcasses is sometimes difficult (*e.g.*, tides), limiting proper dissection

techniques and tissue sampling. As with other host-parasite systems, the use of longitudinal

assessments and experimental manipulation of infection could give new perspectives on the impact

of metastrongyles in marine mammal health. Recently, the sampling of living species through the

collection of feces and body fluids has been implemented in cetaceans (Williams *et al.*, 2020).

Further, the development of molecular techniques such as major sperm protein (MSP) ELISAs to

test for serum antibodies to metastrongyles aid in illuminating metastrongyle seroprevalence

(Ulrich *et al.*, 2015, 2016). The continued use of approaches other than opportunistic necropsies

may substantially contribute to the understanding of the drivers and consequences of

metastrongyle infection in marine mammals.

As with many wildlife parasites (Haddaway and Watson, 2016; Seguel and Gottdenker, 2017),

most studies on metastrongyles in marine mammals are conducted in North America and Europe,

leading to overrepresentation of North American and European marine mammal and metastrongyle

species in the literature. Therefore, cosmopolitan coastal marine mammals in these regions such

as the harbor porpoise and the harbor seal along with their parasites were among the most assessed

species. Despite our intended penalization for these study biases, our results should be interpreted

with caution since more studies from different regions of the world and in cryptic and/or pelagic

marine mammal species could influence inferences on the role of host and parasite factors driving

pathogenic effects. For instance, PCBs and other persistent organic pollutants (POPs) are found in

higher concentrations in marine mammals in more industrialized regions of North America and

Europe (Fair *et al.*, 2010; Jepson *et al.*, 2016; Durante *et al.*, 2016; Seguel *et al.*, 2020a). These

immunosuppressive pollutants are thought to increase susceptibility to metastrongyles, facilitating

their pathogenic effects (Jepson *et al.*, 2016). Therefore, more studies from regions of the world

with lower POP concentrations or where new or underrepresented host-parasite relationships occur

could modify our understanding of the health impacts of metastrongyles in marine mammals.

As in domestic and land mammals, bronchopneumonia is the most common pathogenic

outcome of lungworm infection in marine mammals. The presence of lungworms in airways or

pulmonary parenchyma causes tissue damage, eliciting an inflammatory response that also

facilitates colonization by bacteria (Measures, 2001). Therefore, in most of the assessed studies,

verminous bronchopneumonia was usually complicated by secondary bacterial infection. The

impact of metastrongyles in other tissues such as blood vessels, heart or otic apparatus is probably

more direct, since in these tissues bacteria colonization secondary to helminth infection is rare. In

the cardiovascular system, the impact of lungworm infection can be serious with vasculitis,

localized or systemic thrombosis and coagulopathy (Gulland *et al.*, 1997b; Seguel *et al.*, 2018b;

Barnett *et al.*, 2019b). In these cases, there is clear link between metastrongyle infection and

mortality. Interestingly, invasion of the cardiovascular system is usually associated to

metastrongyle species that are not the most common definitive host for that parasite species,

highlighting the role of host specificity in the most pathogenic interactions between metastrongyles

and marine mammals.

Parasite and host diversity increase the chances for a for a pathogenic outcome in

metastrongyle-marine mammal relationships. This finding is supported by general hypotheses on

parasite virulence that suggests that a parasite species with a greater diversity of potential hosts

can evolve to be more pathogenic, as the sustainability of the parasite population is less dependent

on the survival of individual hosts (Poulin, 2011; Schmid-Hempel, 2021). Similarly, a host species

that harbors a greater diversity of parasites may see a greater propensity for pathogenic outcomes

from individual parasite species due to an increase in the stress on the immune and respiratory

systems resulting from co-infections (Seguel *et al.*, 2023). Our results also suggest that larger hosts

are more likely to sustain pathogenic outcomes because of metastrongyle parasitism. Since marine

metastrongyles are transmitted through intermediate hosts that represent food items for marine

mammals (Dailey, 1970a), this result could indicate that increased food consumption leads to a

greater larvae intake and subsequently a higher parasite burden. Similarly, a larger host body size

could correlate with access to different prey items, contributing to a greater diversity of

metastrongyle infections.

A comprehensive understanding of the drivers of the negative physiological effects

associated with metastrongyle parasitism in marine mammal hosts is challenged by a presumably

strong bias towards reporting only positive findings. The available literature clearly elucidates a

multitude of host-parasite relationships in which metastrongyles produce a definable pathological

effect, howbeit it is inadequate in clarifying why these effects are observed in some species or

populations and not in others. However, general host and parasite traits associated with pathogenic

outcomes are like those of other helminth and microbial parasites, including host permissiveness

of infection by multiple species and parasite capacity to infect multiple host species. In this context,

the rapidly changing marine environment with generalized loss of biomass could favor parasites

and hosts with a diversified trophic network, potentially selecting for more pathogenic

relationships between metastrongyles and their marine mammal hosts.

5. Supplementary Data

Supplementary figures are submitted as electronic supplementary files. Review protocol not

registered.

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