Individual differences in developmental trajectory leave

a male polyphenic signature in bulb mite populations

Jacques A. Deere[1*] & Isabel M. Smallegange[2*]

[1] Institute for Biodiversity and Ecosystem Dynamics, University of Amsterdam, PO Box 94240, 1090

GE Amsterdam, the Netherlands

[2] School of Natural and Environmental Sciences, Newcastle University, Newcastle upon Tyne, NE1

7RU, UK

*Equal contribution

Corresponding author: Isabel M. Smallegange – Isabel.smallegange@newcastle.ac.uk

Abstract

Developmental plasticity alters phenotypes and can in that way change the response to selection

and affect, and be affected by, population size-structure when alternative phenotypes show

different life history trajectories, in an eco-evolutionary interaction. Developmental plasticity often

functions to anticipate future conditions but it can also mitigate current stress conditions, raising the

question if these two kinds of developmental plasticity underlie different eco-evolutionary

population dynamics. Here, we tested in a long-term population experiment using the male

polyphenic bulb mite (*Rhizoglyphus robini*), if the selective harvesting of juveniles on different

developmental trajectories elicits eco-evolutionary population responses that are expected if the

male polyphenism is assumed to be anticipatory or mitigating. We found that the frequency of adult

males that expressed costly (fighter) morphology was lowest under the most severe juvenile

harvesting conditions. This response cannot be explained if we assume that adult male phenotype

expression is to anticipate adult (mating) conditions because, in that case, only the manipulation of

adult performance would have an effect. This response can, however, be explained by juveniles

mitigating their increased mortality risk by expediating ontogeny to forego the development of

costly morphology and mature quicker but as a defenceless scrambler. If, like in mammals and birds

where early-life stress effects are extensively studied, we account for such pre-adult viability

selection in coldblooded species, it would allow us to better characterise natural selection on trait

development like male polyphenisms, how it can affect the response to other selections in

adulthood, and how such trait dynamics influence, and are influenced by, population dynamics.

Keywords: alternative male phenotypes, dispersal, eco-evolutionary dynamics, male morph

coexistence, polyphenism

Introduction

Across the animal kingdom, developmental plasticity comprises a switch to alternative developmental pathways that depends on an individual's condition – the resource budget available for the production and maintenance of adaptive traits (Hill 2011; Casasa et al. 2020; Nijhout & McKenna 2018) – and which leads to the expression of alternative phenotypes in adulthood (polyphenism) (West-Eberhard 2003; Nijhout & McKenna 2018). For example, many intraspecific, arthropod male dimorphisms comprise large majors that have weapons for male-male competition, and smaller, weaponless minors that sneak matings (Moczek 2003; Smallegange 2011; Buzatto & Machado 2014). Importantly, these condition-dependent alternative phenotypes typically have different survival, growth and reproduction rates (West-Eberhard 2003; Smallegange 2011; Weir et al. 2016). Differences in individual development will thus affect a population's structure, size and growth. Such population changes can in turn influence individual development through processes like density-dependent competition over resources (food) and density-dependent selection (Travis et al. 2013), creating eco-evolutionary feedbacks at the population level (Smallegange & Deere 2014; Croll et al. 2019; Smallegange 2022). However, accurately predicting the population dynamical responses to perturbation of developmental trajectories is still difficult (Smallegange et al. 2019; Smallegange 2022).

The way in which a perturbation to individual development can impact the eco-evolutionary dynamics of populations depends on what selection pressures are assumed to act on developmental plasticity (Smallegange 2022). In case of the minor and major males described above, whether a male develops into an armed major male or an unarmed minor male depends on whether it reaches a critical resource budget or body size threshold during development (Plaistow et al. 2005; Sasson et al. 2016; Cotton et al. 2004). If developmental plasticity is anticipatory, this condition-cued threshold mechanism is the evolutionary result of disruptive sexual selection on alternative mating tactics (West-Eberhard 2003; Smallegange et al. 2019): good-condition juvenile males maximise fitness by becoming majors, anticipating competitive mating success; bad-condition juveniles anticipate failure

at competitive success and salvage fitness by becoming minors (Shuster & Wade 2003). On the other

hand, the condition-cued threshold mechanism could be part of a heritable developmental system

that adaptively mitigates stressful environmental conditions (Badyaev 2005, Del Giudice et al. 2018;

Ellis & Del Giudice 2019). Such mitigating developmental plasticity would be the result of natural

selection that favours developmental systems that tend to construct phenotypes that are successful,

relative to other variant systems, at surviving and reproducing (Griffiths & Gray 1994). This means

that juvenile males can mitigate stress during maturation by refraining from developing costly

morphology to maintain fitness and mature and survive as an adult minor, as opposed to risk dying

during maturation when investing in costly major morphology (Smallegange et al. 2019). To

summarise, in both kinds of plasticity, the fitness functions of the alternative male phenotypes cross

over a condition gradient, but anticipatory developmental plasticity assumes that the threshold for

alternative male phenotype expression at the cross-section of the fitness functions is evolutionarily

regulated by how adult males perform, whereas mitigating developmental plasticity assumes that it

is regulated by the dynamics of population density, food competition and individual resource

budgets. Together with the fact that the condition distribution itself informs on population size

(Smallegange & Coulson 2013), we have the ingredients to create predictions on how anticipatory or

mitigating developmental plasticity fuel eco-evolutionary population change in response to

perturbations to individual development.

We conducted a long-term population experiment to test hypotheses on how anticipatory

and mitigating developmental plasticity impact eco-evolutionary population responses to the

selective removal of juveniles of different condition in populations of the male polyphenic bulb mite

Rhizoglyphus robini. Adult fighter males of the bulb mite possess a proportionally thickened third leg

pair with dagger-like claws that are used in fights to kill rival males (Radwan et al. 2000). Whereas

fighter males metamorphose from good-condition juveniles that have a large resource budget,

nutrition-deprived juveniles that grow slow or are of worse condition when metamorphosing into an

adult instead express the scrambler phenotype that does not have such leg modifications

(Rhebergen et al. 2022). Further, in response to unfavourable environmental conditions, *R. robini*

expresses a facultative dispersal morph during development, called the deutonymph, which does

not feed and is adapted to phoretic dispersal (Díaz et al. 2000). Deutonymph expression occurs mid-

development, prior to the final instar stage from which juveniles metamorphose into adults (Díaz et

al. 2000), and, thus, prior to when adult male phenotype is determined (which is immediately before

metamorphosis [Rhebergen et al. 2022]). Because deutonymph expression carries developmental

costs, including reduced size at metamorphosis in both sexes and reduced egg production in females

(Deere et al. 2015), our working assumption is that only individuals that are of sufficiently good

condition can develop into one if cued by poor environmental conditions. This assumption is

supported by the fact that in two of our previous studies (Smallegange & Coulson 2011; Deere et al.

2015), male deutonymphs always matured as a fighter, from which we surmise that males always

were in sufficiently good condition to develop costly fighter legs.

In our experiment, we selectively removed individuals of different conditions by removing

deutonymphs, or other juveniles. Assuming that the male polyphenism is anticipatory, we predict

that removing deutonymphs from the population will lower the righthand side of the distribution of

individual conditions (because individuals of relatively good condition are removed), and thus

population size (compare blue dotted distribution size [before harvesting] with red dashed

distribution size [after harvesting] in Fig. 1c). However, we expect that male fitness is unaffected by

juvenile performance and thus that the mean threshold for fighter expression will evolutionarily

track the latter change in the condition distribution because relative, and not absolute condition is

important in determining alternative phenotype fitness (Tomkins et al. 2011) (compare the

proportions of the differently shaded areas under the condition distribution in Fig. 1a [before

harvesting] with Fig. 1c [after harvesting]). Likewise, we predict that the selective removal of

juveniles (bar deutonymphs) of random condition has no effect on fighter expression (compare the

proportions of the differently shaded areas under the condition distribution in Fig. 1a [before

harvesting] with Fig. 1e [after harvesting]), because fitness functions remain unaltered. The

reduction in population size because of the harvesting of juveniles, however, will be reflected in an

overall reduced size of the condition distribution (compare blue dotted distribution size [before

harvesting] with red dashed distribution size [after harvesting] in Fig. 1e).

In contrast, assuming that the male polyphenism is mitigating, the stress mechanism of the

developmental system can fuel a mitigating response to increased deutonymph mortality because

males can mature early as a minor to escape the juvenile stage quickly (Ernande et al. 2004), fuelling

the evolution of scrambler expression. Mean fighter expression is then predicted to decrease in

response to deutonymph harvesting (compare the proportions of the differently shaded areas under

the condition distribution in Fig. 1b [before harvesting] with Fig. 1d [after harvesting]) and in

response to the selective harvesting of other juveniles (compare the proportions of the differently

shaded areas under the condition distribution in Fig. 1b [before harvesting] with Fig. 1f [after

harvesting]). Further, because scramblers mature earlier in life (Smallegange 2011), live longer than

fighters (Radwan & Bogacz 2000) and sire more offspring than fighters (van den Beuken et al. 2019),

an increase in scrambler expression can increase population size (Smallegange & Deere 2014), and,

thus, increase the size of the condition distribution ((compare blue dotted distribution size [before

harvesting] with red dashed distribution size [after harvesting] in Fig. 1d and 1f). But note that the

removal of good-condition juveniles when removing deutonymphs will result in the right-hand side

of the distribution to be at lower condition values (compare blue dotted distribution size [before

harvesting] with red dashed distribution size [after harvesting] in Fig. 1d).

Methods

Study system

The bulb mite has six life stages: egg, larva, protonymph, deutonymph, tritonymph an adult. The

deutonymph stage is a facultative, non-feeding dispersal stage and mites only develop into this stage

during unfavourable conditions (i.e. low food quality and quantity) (Díaz et al 2000). When growing

from one stage to the next, mites moult; this stage is known as the quiescent stage which is

immobile and non-feeding. Source populations were established from mites collected in 2010 from

flower bulbs in storage rooms in North Holland. Mites were kept on an oats diet and maintained as

described in Smallegange (2011). Experimental treatments were established from these source

populations.

Experimental procedure

The experiment ran between June 2016 and April 2017 (302 days) and comprised five treatments: (i)

harvesting of 100% of all deutonymphs (D100), (ii) harvesting of 50% of deutonymphs (D50), (iii)

harvesting of other juveniles (excluding eggs and deutonymphs) at the same percentage at which

deutonymphs where harvested from all juveniles in D100 (J-D100) to keep the demographic impact

of harvesting comparable, (iv) harvesting of other juveniles (excluding eggs and deutonymphs) at the

same percentage at which deutonymphs where harvested from all juveniles in D50 (J-D50), again, to

keep the demographic impact of harvesting comparable, and (v) no harvesting (C; general control

treatment). Two levels of harvesting deutonymphs (D100, D50) and juveniles (J-D100 and J-D50)

were chosen to be able to compare harvesting impacts. Treatments were replicated three times

resulting in fifteen populations. The juvenile harvesting populations were started 8 weeks after the

other treatments, but before harvesting commenced (Fig. 2), for logistical reasons. Populations were

kept in 20 mm diameter, flat-bottomed glass tubes with a plaster of Paris and powdered charcoal

base, which was kept moist to avoid desiccation of the mites. Tubes were sealed by a circle of very

fine mesh (allowing gaseous diffusion), which was held in place by the tubes' standard plastic caps

with ventilation holes cut into them. Populations were kept in an unlit climate chamber at 25℃ and

>70% relative humidity.

Each population was established with 50 randomly selected mites from the source

population with founder mites, which we removed after 15 days to reduce founder effects (founder

mites can be easily distinguished because they are much bigger than the first generation emerging

after 15 days in the experimental population). Each population was fed approximately 1mg of oats

per day (the weight was never less than 0.9g and never exceeded 1.1g). After population

initialization, on each Thursday, mites of each life stage in all population were counted using a hand

counter at 15 X magnification. For three weeks at the start of each time period (day 85-161, day 162-

238 and day 239-302: see statistical analyses below), photos were taken of up to 5 individual

deutonymphs, fighters, scramblers, and females as proxies to assess any shifts in mean condition

across the different life stages in response to experimental conditions. The body length of mites on

each photo was measured to the nearest 0.1 µm using ZEN software blue edition (ZEISS microscopy,

Carl Zeiss, Germany). We measured body length for adult mites as the ventral distance of the

idiosoma (the white bulbous posterior part of the body; from the mouthparts to the furthest part of

the genitalia); for deutonymphs as the dorsal anterior-posterior length (without mouthparts); and

for quiescent tritonymphs as the lateral body length (without mouthparts). Harvesting treatments

started 85 days after populations were initialized, which is a sufficiently long time for dynamics to

stabilise (which typically happens at 40 days after initialisation [Cameron and Benton 2004;

Smallegange and Deere 2014]) (see also Fig. 1). Harvesting was done once a week (on Thursday),

after counting and before feeding. The harvesting treatments were applied for a total of 217 days,

which in a similar experiment was sufficient to observe an evolutionary shift in fighter expression

(Smallegange & Deere 2014). However, even if this period of time is too short to observe an

evolutionary response to harvesting, we can still distinguish between our anticipatory and mitigating

hypotheses. Specifically, under juvenile harvesting (J-100 and J-D50), fighter expression is predicted

to remain unaltered if it is anticipatory (Fig. 2e). However, fighter expression is predicted to decrease

if it is mitigating (Fig. 2f) because juvenile males can respond quickly (within a generation) to

increased juvenile mortality by expediating ontogeny and mature as a scrambler. This plastic

response, in turn, will further fuel the evolution towards developmental systems that produce

scramblers in response to the juvenile harvesting selection pressure.

Life-history assays

Following the end of the long-term population experiment we conducted a common garden life

history assay to assess if any differences in body size and fighter expression between treatments

were plastic or genetic. For each assay we removed ten adult females from each population,

individually isolated them and gave them ad lib access to oats and allowed them to lay eggs. Three of

the ten females (isolating more individuals was logistically not possible) were then randomly

selected and their offspring individually isolated, given ad lib access to oats, and followed until they

reached maturity. Deutonymphs, quiescent tritonymphs and adults (2-5 days after maturation) were

size-measured as above and their sex and morph scored (we measured a total of 420 individuals).

The effects of maternal nutritional conditions are negligible in this species (the effect size of

offspring environment is 15 times larger than that of maternal environment: Smallegange 2011a),

therefore, rearing mites in a common garden environment condition for one generation is sufficient

to eliminate maternal effects. Females and their offspring were kept in 10 mm diameter tubes with a

plaster of Paris and powdered charcoal base, which were kept in an unlit climate chamber at 25°C

and >70% relative humidity.

Statistical analyses

To analyse the results from the population experiment, we only used data from day 85 onwards

(when harvesting commenced) and excluded two population tubes that had fallen over during the

population experiment (one tube in treatment D100 and one in treatment J-D50). We then divided

the experimental period into three periods, day 85-161, day 162-238 and day 239-302 to assess

long-term (instead of transient) temporal changes in the response variables within each treatment

group. To analyse the results of the population experiment, we used a generalised linear mixed

model (GLMM) with Gaussian errors to analyse the effect of the fixed factors harvesting treatment

(C, D100, D50, J-D100, J-D50) and time period (period 1, 2, and 3) and their interaction, including

'population tube' as a random effect (to account for the repeated measures within each

experimental population) on: fighter expression (the number of fighters with the number of adult

males set as an offset so that the model prediction will be proportion of fighter males), and the

mean size of deutonymphs, adult fighters, scramblers and females. The model assumptions of

Gaussian errors and homoscedacity were confirmed by inspecting the probability plots and error

structures. We used a GLMM with Poisson errors to analyse the effects of the fixed factors

harvesting treatment and time period, their interaction, with 'population tube' as a random effect,

on the number of deutonymphs and on total population size.

To analyse the results of the life history assay conducted at the end of the population

experiment, we used a GLMM with Gaussian errors to analyse the effect of the fixed factor

harvesting treatment (C, D100, D50, C-D100, C-D50), including 'population tube' and 'maternal

identity' as random effects on the mean size of adult fighters, scramblers and females. The model

assumptions of Gaussian errors and homoscedacity were again confirmed by inspecting the

probability plots and error structures. We used a GLMM with binomial errors to analyse the effect of

the harvesting treatment on fighter expression ('0' if a male developed into a scrambler, and '1' if in

a fighter), including 'population tube' and 'maternal identity' as random effects.

In each GLMM, the significance of the treatment effects was tested using a model

simplification procedure, whereby the full model was fitted after the least significant term had been

removed (never deleting random terms from the model), if the deletion caused an insignificant

increase in deviance as assessed by a maximum likelihood ratio test (LRT). Pairwise comparisons

among the treatment levels were conducted by creating a reduced model within which two levels

were combined. If the reduced model caused an insignificant increase in deviance as assessed by a

maximum likelihood ratio test (α = 0.05), then the two levels were considered to be not significantly

different. All analyses were conducted in R and for all GLMMs we used the *lme4* package (www-r-

project.org).

Results

Population size, fighter expression and number of deutonymphs

Mean total population size had stabilised across all treatments at the start of harvesting (Fig. 2) but

was significantly affected by harvesting (χ^2_4 = 15.58, p=0.004) and differed between time periods (χ^2_2

= 262.07, p<0.001) (non-significant interaction: χ^2_8 = 7.30, p=0.063). Population size was highest

when 100% of deutonymphs were harvested (D100), lower when 50% of deutonymphs were

harvested (D50), and lowest in all the other treatments (C, J-D50 and J-D100) (treatments C and J-

D50 did not differ significantly (χ^2_1 = 0.35, p=0.553), and neither did C and J-D50 differ significantly

from J-D100 (χ^2_1 = 0.47, p=0.494)) (Fig. 3A). Irrespective of harvesting treatment, total population

size significantly increased over time (Fig. 3B).

The harvesting treatment significantly affected the proportion of fighters but differently at

different time periods (significant interaction harvesting × time period: Σ^2_8 = 29.92, p<0.001). Within

the interaction, the harvesting treatments C and J-D50 did not significantly differ in their effect on

proportion of fighters over time (χ^2_3 = 4.22, p=0.239) and were combined into one level. The next

two similar responses to the harvesting treatments J-D100 and D50 did, however, differ significantly

(χ^2_3 = 11.80, p=0.008) (Fig. 3C). In summary, the proportion of fighters was always low when 100% of

deutonymphs were harvested (D100 treatment) (Fig. 3C), but when other juveniles were harvested

at the same percentage as in D100, (J-D100), and when harvesting 50% of deutonymphs (D50), the

proportion of fighters significantly decreased towards the end of the experiment (Fig. 3C). In

contrast, under the combined harvesting treatments of C and J-D50, the proportion of fighters was

always high over the whole course of the experiment (Fig. 3C).

The harvesting treatment also significantly affected the number of deutonymphs, but, again,

differently at different time periods (significant interaction harvesting $\times$ time period: χ_8^2 = 49.70,

p<0.001). Within the interaction, the harvesting treatment levels J-D100 and C did not significantly

differ (χ_3^2 = 0.40, p=0.940) and were combined into one level. Likewise, the latter combined level did

not significantly differ from the J-D50 harvesting treatment (χ_3^2 = 3.08, p=0.380) and all three control

levels were combined into one level. These three levels, combined, significantly differed from the

D50 harvesting level (χ_3^2 = 12.42, p=0.006) and from the D100 harvesting level (χ_3^2 = 67.71, p<0.001).

We therefore find that, counterintuitively, the number of deutonymphs significantly increased over

time when all deutonymphs were harvested (D100), and, to a lesser extent, when 50% of

deutonymphs were harvested (D50) (Fig. 3D). In contrast, in the other treatments combined, the

number of deutonymphs slightly decreased over the course of the experiment (Fig. 3D).

Furthermore, at any given time period, the number of deutonymphs was always the highest in the

treatment where weekly harvesting removed all deutonymphs (D100) (Fig. 3D).

Deutonymph, fighter, scrambler and adult female mean size

Mean deutonymph size was not significantly affected by harvesting (χ_4^2 = 4.81, p=0.307) but

significantly decreased over the course of the experiment (significant effect of time period: χ_2^2 =

106.31, p<0.001) (Fig. 4A) (non-significant interaction: χ_8^2 = 7.22, p=0.513). Mean fighter size also

decreased over time but size differed significantly between the different harvesting treatments

(significant interaction harvesting $\times$ time period: Σ_8^2 = 19.44, p=0.013) (Fig. 4B). Within the

interaction, fighter size in the harvesting treatment levels J-D50 and J-D100 did not significantly

differ (χ_3^2 = 2.33, p=0.507) and were combined into one level (Fig. 4B). Similarly, fighter size in the

harvesting treatment levels C and D50 did not significantly differ (χ_3^2 = 1.83, p=0.609) and were

combined into one level (Fig. 4B). The overall result was that mean fighter size in the combined

levels J-D50 and J-D100 and in the combined levels C and D50 decreased over time, whereas mean

fighter size in the D100 treatment did not decrease until the last time period (Fig. 4B). Mean

scrambler size was not significantly affected by harvesting (χ_4^2 = 1.17, p=0.883) but significantly

decreased over the course of the experiment (significant effect of time period: χ_2^2 = 96.44, p<0.001)

(Fig. 4C) (non-significant interaction: χ_8^2 = 9.48, p=0.303). Finally, mean adult female size was also not

significantly affected by harvesting (χ_4^2 = 0.62, p=0.961) but significantly decreased over the course

of the experiment (significant effect of time period: χ_2^2 = 67.65, p<0.001) (Fig. 4D) (non-significant

interaction: χ_8^2 = 14.55, p=0.068).

Life history assay

Neither fighter size (Σ_4^2 = 6.58, p=0.160, mean size: 439.76 µm $\pm$ 2.39 SE, n = 190), scrambler size (Σ_4^2

= 2.77, p=0.597, mean size: 401.07 µm $\pm$ 7.33 SE, n = 14) or female adult size (Σ_4^2 = 3.75, p=0.440,

mean size: 334.66 µm $\pm$ 1.65 SE, n = 206) were affected by the harvesting treatments in the common

garden life history assay. Fighter expression was also not significantly affected by harvesting

treatment (Σ_4^2 = 0.51, p=0.973). However, only 14 scramblers emerged in the life history assay (as

opposed to 190 fighters), prohibiting any statistical inference. Nevertheless, it is noteworthy that

mean scrambler size was lowest (Fig. 5A) and mean fighter expression was highest (Fig. 5B) for those

treatments where fighter expression in the long-term population experiment was also highest

(control treatment C and juvenile harvesting treatment J-D50) (Fig. 3C). Similarly, mean scrambler

size was highest (Fig. 5A) and mean fighter expression lowest (Fig. 5B) for those treatments where

fighter expression in the long-term population experiment was also relatively the lowest

(deutonymph harvesting treatments D100 and D50 and juvenile harvesting treatment J-D100) (Fig.

3C).

Discussion

We tested two hypotheses on how developmental plasticity impacts the eco-evolutionary response of male polyphenic bulb mite populations to the selective removal of individuals of different developmental stages. If male alternative male phenotype expression is taken to be anticipatory (anticipating future mating success), we predicted that the selective harvesting of deutonymphs and other juveniles would not impact fighter expression. In contrast, if alternative male phenotype expression is taken to be mitigating, we predicted that the selective harvesting of deutonymphs and other juveniles would decrease fighter expression both plastically, as male juveniles can expediate development and escape the juvenile stage quicker by maturing as a scrambler, and evolutionarily, as the selective removal of juveniles fuels the evolution of developmental systems that produce scramblers. In line with the mitigating hypothesis, we found that fighter expression was overall lowest when 100% of all deutonymphs were harvested (D100), and, at the end of the experiments, had reached similarly low levels in the treatments where 50% of deutonymphs were harvested (D50) and where the highest number of other juveniles were harvested (J-D100). In contrast, fighter expression remained high throughout the experiment in the control treatment and in the treatment where the lowest number of juveniles were harvested (J-D50), perhaps because, there, the harvesting impact was the lowest. Our life history assay at the end of the population experiment did not have sufficient statistical power to tell if these shifts in fighter expression were (in part) evolutionary. However, it is striking that the harvesting treatments where we observed the lowest levels of fighter expression in the population experiment are the same as where we observed the lowest fighter expression levels in the life history assay. But even if this pattern was coincidental and not indicative of an evolutionary change in fighter expression, our findings still point to a role for mitigating developmental plasticity. That is, we expected no shift in fighter expression at all in response to the harvesting of juveniles (bar deutonymphs) under anticipatory developmental plasticity (Fig. 1e), but a reduction in fighter expression under mitigating developmental plasticity (Fig. 1f) because individuals mature earlier as a scrambler to escape the risky juvenile stage. The

latter reduction is indeed what we observed towards the end of the experiment in the treatment

where we harvested the highest number of juveniles other than deutonymphs (J-D100). Our results

therefore suggest that nutrition-deprived male juveniles do not 'make the best of a bad job' by

anticipating a non-fighting mating tactic (Dawkins 1980; Eberhard 1982), but instead develop into

that male adult phenotype because they can adaptively mitigate their early-life stress (Rhebergen

2022). Such adaptive early-life stress responses that have negative effects in adulthood are common

in (warm-blooded) mammals and birds (Lindström 1999; Fisher et al. 2006; Wells 2011; Vickers

2014), likely because their growth during development is physiologically constrained. However, if we

account for such pre-adult viability selection in coldblooded species that typically show flexible

growth, it would allow us to better characterise natural selection on trait development like male

polyphenisms in these species, how it can affect the response to other selections in adulthood (e.g.,

sexual selection: Mojica and Kelly 2010; Lürig & Matthews 2021; Rhebergen 2022), and how such

trait dynamics influence, and are influenced by, population dynamics (Smallegange 2022).

One unexpected finding in our experiment was that the number of deutonymphs was

highest when we harvested all of them, each week. Further, despite being harvested, deutonymph

number increased as the experiment progressed. We can only speculate why this happened.

Perhaps this overcompensatory response was because the per capita food availability increased as

we removed individuals (Verhulst 1838, Schaefer 1954, Hilborn and Walters 1992), although we still

only maximally removed a small percentage of the total population in the deutonymph harvesting

treatments. Furthermore, initiation of the deutonymph stage in this species is due to poorer

resource availability and so our findings are unlikely due to an increase in per capita food availability.

The higher number of deutonymphs we found could also reflect eco-evolutionary dynamics in which

another selection pressure, for example imposed by strong-density dependence (population size was

highest in the 100% deutonymph harvesting treatment), opposed the effects of direct harvesting

(Edeline & Loeuille 2021). Failure to account for either of these eco-evolutionary processes might

fundamentally hamper our ability to understand the expression of dispersal phenotypes. Another

explanation is that it was the actual lack of deutonymphs and their olfactory cues that triggered

deutonymph development. Olfactory cues are common in acarine mites. All acarines have a sensory

site on their forelegs known as the "foretarsal sensory organ" and olfaction is one of this site's

functions (Carr & Roe 2016). If the harvesting of deutonymphs reduced the concentration of

deutonymph olfactory cues, this could trigger more juveniles to develop into a deutonymph

because, with fewer deutonymphs around, competition for attachment sites on insects is lower

(deutonymphs disperse via phoresy on insects [Díaz et al 2000]). We realise, however, that this is

speculation and olfactory cues produced by deutonymphs have yet to be tested in *R. robini*. Finally,

in our previous studies (Smallegange & Coulson 2011; Deere et al. 2015), male deutonymphs always

matured as a fighter, but here, increases in deutonymph numbers did not coincide with increases in

fighter frequency. Perhaps this was because twice as many females emerge from deutonymphs than

males (fighters) (Deere et al. 2015; Stewart & Smallegange in prep) so that our statistical analyses

were unable to pick up significant differences in the few more fighters that emerged. It does

highlight, however, that we still have some way to go to unravel the intricate links between

deutonymph expression, alternative male phenotype expression and sex ratio and how they

together drive population dynamics.

Because a change in the distribution of a trait will almost always also change the size of the

distribution of that trait, it follows that trait change is accompanied by a change in population size. If

a trait is heritable, this constitutes eco-evolutionary change at the population level (Smallegange &

Coulson 2013). This is especially so in the case of alternative phenotypes where we expect a shift in

phenotype expression to be accompanied by a change in population size because the different

phenotypes often have different demographic rates. For example, in the case of bulb mite males,

scramblers mature earlier in life (Smallegange 2011), live longer than fighters (Radwan & Bogacz

2000) and sire more offspring than fighters (van den Beuken et al. 2019) so that an increase (or

decrease) in scrambler (or fighter) expression will increase population size. Indeed, we found the

highest population size in the harvesting treatment with the lowest level of fighter expression (D100:

100% deutonymph harvesting). We suspect that the increase in population size that we observed

over the course of the experiment is associated with the observed decrease in mean body size over

time in response to the strong, density-dependent food conditions, because smaller organisms

typically reproduce faster and can reach higher carrying capacities than larger ones (e.g., Huryn &

Benke 2007). Indeed, Gilbert et al. (2022) recently showed in an experiment with the protist

Tetrahymena pyriformis that mean body size decreased in response to increased density-

dependence and that this rapid change in mean body size had a larger effect on population density

than the reverse. Reductions in mean body size have been observed across a range of taxa in

response to adverse environmental change (Gardner et al. 2011; Sheridan & Bickford 2011). Often,

these reductions are associated with increased mortality rates (e.g., in red knots *Calidris canutus*

[van Gils et al., 2016]). What is more, when populations not only suffer from directional

environmental change but also from selective harvesting, the removal of good condition individuals

(of large size) can lead to population extinction (Knell & Martínez-Ruiz 2017). Thus, unravelling the

intertwined ecological and evolutionary dynamics of population size, body size, condition and

condition-dependent (alternative) phenotypes is integral to understanding eco-evolutionary

population responses to change (Edeline & Loeuille 2021).

Data availability statement

The raw data and the R scripts that we used to analyse the data can be found on Figshare (Deere &

Smallegange 2023).

References

Badyaev AV. 2005. Stress-induced variation in evolution: from behavioural plasticity to genetic assimilation. Proc. R. Soc. B 272: 877–886.

Buzatto BA, Machado G. 2014. Male dimorphism and alternative reproductive tactics in harvestmen (Arachnida: Opiliones). Behav. Processes 109: 2–13.

Carr AL, Roe M. 2016. Acarine attractants: Chemoreception, bioassay, chemistry and control. Pestic Biochem Physiol 131: 60-79.

Casasa S, Zattara EE, Moczek AP. 2020. Nutrition-responsive gene expression and the developmental evolution of insect polyphenism. Nature Ecol. Evol. 4: 970–978.

Cotton S, Fowler K, Pomiankowski A. 2004. Condition-dependence of sexual ornament size and variation in the stalk-eyed fly *Cyrtodiopsis dalmanni* (Diptera: Diopsidae). Evolution 58: 1038–1046.

Dawkins, R. 1980. Good Strategy or Evolutionarily Stable Strategy? In: Sociobiology: Beyond Nature/Nurture? (G. W. Barlow & J. Silverberg, eds), pp. 331–367. Westview Press, Boulder, CO

Deere JA, Coulson T, Smallegange IM. 2015. Life history consequences of the facultative expression of a dispersal life stage in the phoretic bulb mite (*Rhizoglyphus robini*). PLoS ONE 10: e0136872.

Deere JA, Smallegange IM. 2023. Data and analysis of "Individual differences in developmental trajectory leave a male polyphenic signature in bulb mite populations". figshare. Dataset. https://doi.org/10.6084/m9.figshare.22012589.v1

Del Giudice M, Buck CL, Chaby LE, Gormally BM, Taff CC, Thawley CJ, Vitousek MN, Wada H. 2018. What Is Stress? A Systems Perspective. Integr. Comp. Biol. 58: 1019-1032.

Díaz A, Okabe K, Eckenrode CJ, Villani MG, Oconnor BM. 2000. Biology, ecology, and management of the bulb mites of the genus *Rhizoglyphus* (Acari: Acaridae). Exp. Appl. Acarol. 24: 85–113.

Eberhard WG. 1982. Beetle horn dimorphism: making the best of a bad lot. Am. Nat. 119: 420–426.

Edeline E, Loeuille N. 2021. Size-dependent eco-evolutionary feedbacks in harvested systems. Oikos 130: 1636-1649.

Ellis BJ, Del Giudice M. 2019. Developmental Adaptation to Stress: An Evolutionary Perspective. Annu. Rev. Psychol. 70: 111–139.

Ernande B, Dieckmann U, Heino M. 2004. Adaptive changes in harvested populations: plasticity and evolution of age and size at maturation. Proc Biol Sci. 271: 415-423.

Fisher MO, Nager RG, Monaghan P. 2006. Compensatory Growth Impairs Adult Cognitive Performance. PLoS Biology 4: e251.

Gardner JL, Peters A, Kearney MR, Joseph L, Heinsohn R. 2011. Declining body size: A third universal response to warming? Trends Ecol. Evol. 26: 285–291.

Gilbert JP, Han, Z-Y, Wieczynski DJ, Votzke S. 2022. Feedbacks between size and density determine rapid eco-phenotypic dynamics. Funct. Ecol. 36: 1668-1680.

Griffiths PE, Gray RD. 1994. Developmental Systems and Evolutionary Explanation. J. Phil. 91: 277-304.

Hill GE. 2011. Condition-dependent traits as signals of the functionality of vital cellular processes. Ecol. Lett. 14: 625–634.

Hilborn R. Walters C. 1992. Quantitative fisheries stock assessment: choice, dynamics and uncertainty. Springer.

Huryn A, Benke A. 2007. Relationship between biomass turnover and body size for stream communities. In A. Hildrew, D. Raffaelli, & R. Edmonds-Brown (Eds.), Body size: The structure and function of aquatic ecosystems (pp. 55–76). Cambridge University Press.

Knell RJ, Martínez-Ruiz C. 2017. Selective harvesting focused on sexual signal traits can lead to
extinction under directional environmental change. Proc. R. Soc. B 284: 20171788.
Lindström J. 1999 Early development and fitness in birds and mammals. Trends Ecol. Evol. 14: 343–
348.
Lürig MD, Matthews B. 2021. Dietary-based developmental plasticity affects juvenile survival in an
aquatic detritivore. Proc. R. Soc. B 288: 20203136.
Moczek, AP. 2003. The behavioral ecology of threshold evolution in a polyphenic beetle. Behav. Ecol.
14: 841–854.
Mojica JP, Kelly JK. 2010. Viability selection prior to trait expression is an essential component of
natural selection. Proc. R. Soc. B 277: 2945–2950.
Nijhout HF, McKenna KZ. 2018. The distinct roles of insulin signaling in polyphenic development.
Curr. Opinion Insect Sci. 25:58–64.
Plaistow SJ, Tsuchida K, Tsubaki Y, Setsuda K. 2005 The effect of a seasonal time constraint on
development time, body size, condition, and morph determination in the horned beetle
Allomyrina dichotoma L. (Coleoptera: Scarabaeidae). Ecol. Entomol. 30: 692–699.
Radwan J, Bogacz I. 2000. Comparison of life-history traits of the two male morphs of the bulb mite,
Rhizoglyphus robini. Exp. Appl. Acarol. 24: 115–121.
Radwan JW, Czyz M, Konior M, Kołodziejczyk M. 2000. Aggressiveness in two male morphs of the
bulb mite *Rhizoglyphus robini*. Ethology 106: 53-62.
Rhebergen FT. 2022. The ecology of adaptive condition-dependent polyphenism. PhD thesis,
University of Amsterdam, The Netherlands.
Rhebergen FT, Stewart KA, Smallegange IM. 2022. Nutrient-dependent allometric plasticity in a
male-diphenic mite. Ecol. Evol. e9145.
Sasson DA, Munoz PR, Gezan SA, Miller CW. 2016 Resource quality affects weapon and testis size
and the ability of these traits to respond to selection in the leaf-footed cactus bug, *Narnia*
femorata. Ecol. Evol. 6: 2098–2108.
Schaefer MB. 1954. Some aspects of the dynamics of populations important to the management of
commercial marine fisheries. Inter-Am. Trop. Tuna Comm. Bull. 1: 23–56.
Sheridan JA, Bickford D. 2011. Shrinking body size as an ecological response to climate change.
Nature Climate Change 1: 401–406.
Shuster SM, Wade MJ. 2003. Mating Systems and Strategies. Princeton University Press, Princeton,
NJ.
Smallegange IM. 2011. Complex environmental effects on the expression of alternative reproductive
phenotypes in the bulb mite. Evol. Ecol. 25: 857–873.
Smallegange IM. 2022. Integrating developmental plasticity into eco-evolutionary population
dynamics. Trends Ecol. Evol. 37:129-137
Smallegange IM, Coulson T. 2011. The stochastic demography of two coexisting male morphs.
Ecology 92: 755–764.
Smallegange IM, Coulson T. 2013. Towards a general, population-level understanding of eco-
evolutionary change. Trends Ecol. Evol. 28: 143-148.
Smallegange IM, Deere JA. 2014. Eco-evolutionary interactions as a consequence of selection on a
secondary sexual trait. Adv. Ecol. Res. 50: 145–169.
Smallegange IM, Rhebergen FT, Stewart KA. 2019. Cross-level considerations for explaining selection
pressures and the maintenance of genetic variation in condition-dependent male morphs. Curr.
Opinion Insect Sci. 36: 66–73.

Tomkins JL, Hazel WN, Penrose MA, Radwan JW, Lebas NR. 2011. Habitat complexity drives
experimental evolution of a conditionally expressed secondary sexual trait. Curr. Biol. 21: 569–
573.
Travis J, Leips J, Rodd FH. 2013. Evolution in population parameters: density-dependent selection or
density-dependent fitness? Am. Nat. 181: S9-S20.
Van den Beuken TPG, Duinmeijer CC, Smallegange IM. 2019. Costs of weaponry: unarmed males sire
more offspring than armed males in a male-dimorphic mite. J. Evol. Biol. 32: 153-162.
Van Gils JA, Lisovski S, Lok T, Meissner W, Ozarowska A, de Fouw J, … Klaassen M. (2016). Body
shrinkage due to Arctic warming reduces red knot fitness in tropical wintering range. Science
352: 819–821.
Verhulst PF. 1838. Notice sur la loi que la population suit dans son accroissement. – Corresp.
Mathématique Phys. 10: 113–121.
Vickers MH. 2014 Early life nutrition, epigenetics and programming of later life disease. Nutrients 6:
2165–2178.
Weir LK, Kindsvarter HK, Young KA, Reynolds JD. 2016. Sneaker males affect fighter male body size
and sexual size dimorphism in salmon. Am. Nat. 188: 264-271.
Wells JCK. 2011. The Thrifty Phenotype: An Adaptation in Growth or Metabolism? Am. J. Human Biol.
75: 65–75.
West-Eberhard MJ. 2003. Developmental Plasticity and Evolution. Oxford University Press, Oxford.

Figure 1. Eco-evolutionary changes due to the selective harvesting of deutonymphs and other juveniles. Fitness functions of fighters (black lines) and scramblers (grey lines) cross over a condition gradient; the area under each condition distribution curve represents population size. The proportion of individuals adopting alternative phenotypes (contrastingly shaded areas under the curves: light grey for scramblers and dark grey for fighters) is determined by the threshold (t^*) condition value at the intersection of the fitness functions (indicated by a vertical line). Blue dotted lines and red dashed lines represent condition distributions before (**a,b**) and after harvesting (**c-f**), respectively. (**c**) Although the selective harvesting of deutonymphs will reduce the size of the condition distribution, anticipatory developmental plasticity predicts that the fitness functions, and, thus, the threshold (t^*) for fighter expression, will track the change in the condition distribution and remain in the same, position relative to the condition distribution. In this example, the threshold tracks the mean of the condition distribution. (**e**) Assuming that the harvesting of juveniles reduces population size but does not change mean condition, the threshold (t^*) for fighter expression remains unaffected in case of anticipatory developmental plasticity. (**d,f**) In contrast, the stress mechanism of a developmental system can facilitate the mitigating response to increased deutonymph or juvenile mortality, because males can mature early as a minor to escape the juvenile stage quickly. This will increase scrambler fitness, fuelling the evolution of scrambler expression to a higher threshold (t^*) for fighter expression. Additionally, because scramblers mature earlier in life (Smallegange 2011), live longer than fighters (Radwan & Bogacz 2000) and sire more offspring than fighters (van den Beuken et al. 2019), an increase in scrambler expression can increase population size (Smallegange & Deere 2014), and thus increase the size of the condition distribution (**d,f**).

Figure 2. Time series of mean total population size, averaged per harvesting treatment. The
treatments were: harvesting of 100% of all deutonymphs (D100: grey line), harvesting of 50% of
deutonymphs (D50: black line), harvesting of other juveniles (excluding deutonymphs) at the same
percentage at which deutonymphs where harvested from all juveniles in D100 (J-D100: purple line),
harvesting of other juveniles (excluding deutonymphs) at the same percentage at which
deutonymphs where harvested from all juveniles in D50 (J-D50: indigo line), and no harvesting (C;
control treatment: orange line).The harvesting treatments began in week 13 of the experiment. No
data were collected in week 29 of the experiment due to illness.

Figure 3. Population size and structure of the long-term population experiment. Mean total
population size differed between different harvesting treatments **(a)** and between the different time
periods **(b)**. Fighter expression varied over time in most but not all harvesting treatments **(c)**. The
number of deutonymphs also varied over time in some but not all harvesting treatments **(d)**. Errors
in all panels are standard errors. Harvesting treatments were: harvesting of 100% of all
deutonymphs (D100: solid black lines), harvesting of 50% of deutonymphs (D50: dashed black lines),
harvesting of other juveniles (excluding deutonymphs) at the same percentage at which
deutonymphs where harvested from all juveniles in D100 (J-D100: grey dashed line (c) and solid grey
line (d)), harvesting of other juveniles (excluding deutonymphs) at the same percentage at which
deutonymphs where harvested from all juveniles in D50 (J-D50: solid grey lines), and no harvesting
(C; control treatment: solid grey lines).

552
Figure 4. Mean body sizes observed in the long-term population experiment. Mean deutonymph
size (μm) **(a)**, mean fighter size (μm) **(b)**, mean scrambler size (μm) **(c)** and mean female size (μm) **(d)**
all significantly decreased over the course of the experiment, but for fighters, this decrease differed
between harvesting treatments (d). Errors in all panels are standard errors. Harvesting treatments
were: harvesting of 100% of all deutonymphs (D100: solid black line), harvesting of 50% of
deutonymphs (D50: dashed black line), harvesting of other juveniles (excluding deutonymphs) at the
same percentage at which deutonymphs where harvested from all juveniles in D100 (J-D100: grey
solid line), harvesting of other juveniles (excluding deutonymphs) at the same percentage at which
deutonymphs where harvested from all juveniles in D50 (J-D50: grey solid line), and no harvesting (C;
control treatment: dashed black line).

Figure 5. Life history assay results. Mean scrambler size (μm) (a) and mean fighter expression (b) for each harvesting treatment. Errors in all panels are standard errors. Harvesting treatments were: harvesting of 100% of all deutonymphs (D100), harvesting of 50% of deutonymphs (D50), harvesting of other juveniles (excluding deutonymphs) at the same percentage at which deutonymphs where harvested from all juveniles in D100 (J-D100), harvesting of other juveniles (excluding deutonymphs) at the same percentage at which deutonymphs where harvested from all juveniles in D50 (J-D50), and no harvesting (C; control treatment).