A selfish genetic element and its suppressor causes gross damage to testes in a fly

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Abstract

Selfish genetic elements (SGEs), specifically X-chromosome meiotic drive (XCMD), create huge

conflicts within a host's genome and can have profound effects on fertility. Suppressors are a

common evolutionary response to XCMD to negate its costs. However, whether suppressors

themselves can cause negative non-target effects remains understudied. Here, we examine whether

the intragenomic conflicts created by XCMD and its suppressor affects gonad morphology in

Drosophila subobscura. We found significant differences in testes, seminal vesicle, and accessory

gland size depending on whether a male carried a non-driving X chromosome, an XCMD, and if the

XCMD was suppressed. We also found the first evidence of extreme whole-organ damage to testes

that is specifically associated with a suppressor of XCMD. Unlike other studies, our evidence suggests

that XCMD in *D. subobscura* inflicts extreme damage on male gonads. This damage is most severe if

both XCMD and its suppressor are both present. While costs of suppression have importance in

theoretical models, they have largely been ignored in empirical XCMD systems. Overall, this study

highlights that genetic conflict, created by SGEs and their suppressors, is a potent evolutionary force

that can have major impacts on gonad development and gametogenesis.

Introduction

Gamete production, and with this testes and ovary development, are shaped by both natural and sexual selection. For instance, egg size is strongly affected by natural selection (1,2) whereas testes size and sperm number is heavily influenced by sexual selection (3,4), with trade-offs between investment in testes and somatic function. Thus, testes are classic examples of organs that have to trade-off between sexual and natural selection (5). However, gonads are also the site of an additional set of trade-offs; the need to defend against selfish genetic elements (SGEs) (6–8).

Challenging the laws of 'fair' Mendelian inheritance, SGEs can bias their own transmission into the next generation without increasing the fitness of the carrier and often at the cost of the rest of the genome (9). For example, B chromosomes are additional to an organisms normal chromosome complement, that carry superfluous DNA (10,11). They exhibit super non-Mendelian inheritance, allowing them to selfishly accumulate in the germline (10). As a result, they are widely reported in nature and can cause phenotypic differences and affect fertility (10,12,13). Co-evolutionary responses to intragenomic conflicts created by SGEs in gonads are likely to shape their evolution, alongside natural and sexual selection. Thus, the role of SGEs on gonad evolution should not be overlooked, and could be significant and universal because SGEs are both diverse and near ubiquitous in nature (9).

Conflicts created by SGEs can have enormous consequences on the host's fertility. For instance, *P* element dysgenesis is a property of transposable elements active in the germline (14). Strong evidence for the role of SGEs in evolution comes from observation of the consequences of SGE presence in a naïve host that it has not coevolved with. The *P* element in its native host is not associated with any gross defects (15). Yet when this element is horizontally transmitted into naïve hosts, it becomes associated with a number of abnormalities, including sterility and dysgenic ovaries (16). Another example is X chromosome meiotic drive (XCMD), where male carriers have their Y-sperm killed or damaged thus ensuring the transmission of only the selfish X chromosome to the

next generation (17). This reduces a males fertility and sperm competitive ability (18–20). Thus, a

range of SGEs can negatively impact their hosts and are capable of damaging the gonads and fertility

of both sexes. When faced with a multitude of conflicts caused by SGEs, how do organisms and in

particular gonads, respond to deal with these challenges?

Here we investigate how the development of the testes respond when faced with XCMD and

a suppressor of this system. XCMD is a type of SGE that creates huge conflicts within the testes (21).

XCMD manipulates the process of meiosis in males, where selfish X chromosomes kill Y-bearing

sperm during spermatogenesis, resulting in the biased production of daughters (17). The

transmission advantage gained by XCMD can allow it to rapidly spread through populations,

distorting sex ratios and even leading to population extinction owing to a lack of males (22–24).

Thus, testes in XCMD carrying males are a good model to investigate how SGEs impact the evolution

of gonads. Additionally, testes are particularly vulnerable to SGEs due to the asymmetry of gamete

production. Eggs are large and very costly for a female to produce, so anything that damages eggs

will be extremely costly and very quickly selected against. On the other hand, because male gametes

are generally massively over-produced, damage to sperm may be more readily be tolerated. In cases

where males only produce a reduced amount of sperm, they can still be successful, making sperm

and testes a good target for the activity of SGEs.

In XCMD, male carriers lose up to half of all the sperm that they produce (25). Despite the

lack of sperm produced, drive carrying males do not often show fertility costs in single matings when

compared to wildtype males (18,26–29). Indeed, XCMD in stalk eyed flies even appears to have

resulted in the response to the loss of sperm created by drive through compensation (30–32). Here,

males carrying XCMD have an increased testes size that allows them to produce an equivalent

amount of sperm to standard males. These studies, that look at how the testes respond to drive,

raises the question as to whether drive systems in general promote changes in the testes.

Meiotic drive is deleterious to male function, and selection commonly favours unlinked

suppressor factors that restore Mendelian segregation (21,33). Suppressors, either Y-linked or

autosomal, must have the ability to prevent some killing of Y-bearing sperm when paired with a

driving X chromosome (34). Once a suppressor has evolved, it is expected to rapidly spread to

fixation (35,36). Despite this, the impact of suppressors of XCMD on the testes themselves has been

rarely considered in empirical studies, despite being recognised as potentially important in

theoretical studies (37). Ongoing co-evolution between drive and its suppressor acting within the

testes could have different outcomes depending on how a suppressor works, whether it has

associated costs, and if it has non-target effects. Thus, the question remains does the emergence of

suppressors negate any testes phenotype (and selection) produced by SGEs. Alternatively, the need

for testes to evolve could be enhanced if they have to deal with both XCMD and suppression, which

are locked in a spiralling co-evolutionary arms race (37–40).

In this paper, we use the XCMD system in *Drosophila subobscura* to look for any effects on

testes morphology. *D. subobscura* harbours an XCMD system called 'SR$_{sub}$' (41). The strength of drive

is strong with SR$_{sub}$ carrying males producing broods that are 85-100% female (41,42). Given this

strong drive, a simple expectation is that SR$_{sub}$ should rapidly spread to fixation everywhere.

However, it does not. Instead, in North Africa, SR$_{sub}$ has remained at a stable frequency of

approximately 20% for at least 50 years (41–43). Additionally, *D. subobscura* are reported to be

strictly monandrous (28,44) so effects of sperm competition are irrelevant in this species (18,45). We

recently discovered suppressors of the SR$_{sub}$ system (43) that are capable of fully restoring a normal

50:50 sex ratio to SR$_{sub}$ carrying males. Despite the prediction that suppressors against drive should

rapidly fix within populations, this suppressor remains rare at a frequency of 1-5% (46). The lack of

spread of this suppressor may indicate that it is associated with its own costs and creates its own

burden upon its host.

We determine whether testes morphology is affected when males carry the selfish SR_{sub} X

chromosome (henceforth referred to as SR_{sub} males) in comparison to standard (ST) males that do

not carry SR_{sub}. We also examine if SR_{sub} males that also carry the suppressor genes ($^{Supp}SR_{sub}$ males)

can restore intact testes or whether the suppressor itself can have its own influence on testes

morphology. We used independent SR_{sub} from multiple locations in Morocco to investigate whether

drive in populations have evolved and responded in the same way when it is paired with suppressors

that originated from populations in Tunisia.

Materials and Methods

Origin and maintenance of fly stocks

Isofemale lines (or "isolines") are a method for capturing genetic variation from a wild

population by inbreeding the offspring of single wild caught females to produce a genetically

invariant line (47). Isolines of *Drosophila subobscura* were originally collected from Tabarka, Tunisia

(36.95°N 8.74°E) in 2013 (48), Ras el Oud, Morocco (34.15°N 4.00°W), Demnat, Morocco (31.73°N

7.01°W) and Ouzoud, Morocco (32.02°N 6.72°W). Moroccan collections took place in 2019 and all

flies were caught using banana, yeast and beer baited traps (49). All isolines were kept in standard

Drosophila vials on an ASG double yeast medium (10g agar, 85g sucrose, 40g yeast extract, 60g

maize, 1000ml H_2O, 25ml 10% nipagin) at 18°C on a 12:12 light/dark cycle. Experimental flies were

generated by collecting unmated flies from isolines from each population. All eclosed flies were

removed from populations 24 hours prior to ensure flies had not mated. Fly collection was done

using CO_2 anaesthesia to separate flies by sex. Flies were stored in new vials on a sugar yeast

medium (100g sugar, 100g yeast extract, 20g agar, 1000ml H_2O, 25ml 10% nipagin, 5ml propionic

acid) with no more than 10 flies per vial.

Isolation and introgression of SR_{sub} X chromosomes

We isolated an SR_{sub} chromosome from a wild caught fly from each of Ras el Oud, Morocco,

Demnat, Morocco and Ouzoud, Morocco, where SR_{sub} occurs at a frequency of approximately 20%

(43). SR_{sub} males were initially identified though a phenotypic scan by counting their offspring sex

ratio. Males that produced more than 85% females were deemed SR_{sub} males and were used to

isolate their driving X chromosomes. In order to produce the SR_{sub} males (that carry an SR_{sub}

chromosome), suppressed SR_{sub} males ($^{Supp}SR_{sub}$), that carry a suppressed version of the SR_{sub}

chromosome, and standard (ST) males (that do not carry the SR_{sub} chromosome) that were used in

this investigation, we introgressed both vulnerable and suppressing ST isolines onto the isolated

SR_{sub} chromosomes. This introgression process meant we were able to produce flies that only differ

genetically in carrying drive (males either carried an SR_{sub} X chromosome or an ST X chromosome)

and either carried the suppression genes or genes susceptible to drive. Preliminary experiments

have shown that the Moroccan isolines show no suppression against SR_{sub} (43), thus these isolines

were used to produce the SR_{sub} males that produce 100% daughters. Additionally, the ST isoline from

Tunisia contains genes that are capable of suppressing SR_{sub}. This Tunisian ST isoline was therefore

used to produce the $^{Supp}SR_{sub}$ males that produce a normal 50:50 sex ratio despite the male carrying

SR_{sub}.

We introgressed the SR_{sub} X chromosomes onto either the vulnerable Moroccan ST isolines or

the suppressing Tunisian ST isoline for 14 to 17 generations. The introgression process was done in

two stages. First, females homozygous for the SR_{sub} X chromosome ('SRSR' females) were crossed to

a standard ('ST') male from the target isoline. This cross produces heterozygous females ('SRST'

females) that are discarded, and 'SR_{sub}' males that carry the SR_{sub} X chromosome and the target

isoline autosomes. These SR_{sub} males are then crossed to SRSR females to produce the next

generation of introgressed SRSR females (Supplementary figure 1).

Dissection of male testes

We examined the size of male testes and seminal vesicles (SV's) of SR_{sub} males, $^{Supp}SR_{sub}$ males

and ST males. To look for changes to male testes that may develop over time as they mature or

alternatively be present when just sexually mature and disappear with age, we dissected males

when they were either just sexually mature (at 3 days old) or when they had reached full sexual

maturity (at 7 days old) (50). When at either 3- or 7-days-old, males were anesthetised using FlyNap

(Carolina Biological Supply Co., Burlington, USA) and the testes were dissected out of the males in

300µl of phosphate buffered saline (PBS). Once dissected, the testes were transferred onto a

microscope slide containing 80µl of PBS. A photograph of the testes was taken under constant light

conditions with a microscope mounted Nikon D5100 camera at x40 magnification. In total, between

30 and 50 males were dissected for each treatment (see supplementary Table 1). To control for body

size effects, the left wing of each male was removed using forceps under a microscope (alternatively,

the right wing was taken if the left was damaged) and mounted onto a glass microscope slide. A

photograph of each wing was taken using the microscope mounted camera.

162   Measuring damage to testes

We first examined photographs for evidence of visible damage to the testes. Only damage to the

testes themselves was considered. Damage was scored as a binary variable by the presence of

physical holes in the orange sheath that coats the testes (Figure 1a). An intact orange sheath that

showed thinning was not classed as damage. To avoid bias and ensure consistency, this process was

double blinded and each photo was independently evaluated by three people. Here, the proportion

of males that experienced damage to their testes differed by observer. However, when repeating

the analysis just using the data of each observer, we found the same effect that $^{Supp}SR_{sub}$ males

experienced more damage than both SR_{sub} and ST males (Analysis 1: GLM: $F_{8,210}$ = 11.227, p <0.001;

Analysis 2: GLM: $F_{8,210}$ = 9.821, p <0.001; Analysis 3: GLM: $F_{8,210}$ = 8.818, p <0.001) (Supplementary

Figure 2). Thus, a composite of the three analyses was used to determine a final testes damage score

for each male in which testes were assigned as damaged if two or more evaluators scored them as

damaged.

175   Measuring testes size, SV size, accessory gland size and wing length

Testes and SV size was measured from images using ImageJ 1.48 (51). A subset of images with

visible accessory glands were also analysed in ImageJ to measure accessory gland size. The area of

each individual testis was measured (Figure 1b) and a mean of the two taken for each male. If only

one testis could be measured for size, it was this single measurement that represented a male's

mean testes size. The same procedure was performed for the SV's and accessory glands. Wing

lengths were also measured in ImageJ to control for body size (Figure 1b), using the method in

Gilchrist *et al.,* 2001 (52). The mean testes, SV and accessory gland size for each male was divided by

wing length to generate testes, SV or accessory gland size relative to wing length. These relative

measurements were used in all statistical analyses.

Figure 1: Analysis and measurement of male reproductive organs and wing size. a) Analysing testes images for evidence of damage where testes (T), seminal vesicles (SV), accessory glands (AG) and damage to testes (D) are all visible. ai) Image of intact testes with no damage to the orange sheath. aii) Damaged testes defined by the presence of physical holes in the orange sheath. bi) Measurement of testes (yellow), seminal vesicles (green) and accessory gland (blue) area using ImageJ. bii) Measurement of wing length in ImageJ as defined in Gilchrist *et al.,* 2001 (52).

Statistical analysis

All statistical analysis was performed using the statistical software R, version 3.6.0 (R Core Team, 2019). A binomial generalised linear model (GLM) was used to investigate differences in damage to male testes. Testes size, SV size and accessory gland size were analysed by fitting a GLM with a gaussian error distribution. All models considered the effects of the type of male (SR_{sub}, $^{Supp}SR_{sub}$ or ST males), and the age of the male when it was dissected (3- or 7 days old). For the damage, testes

size and SV size analysis, the original location of origin of the SR_{sub} X chromosome was included in

the model as this provided a better model fit. In these analyses, the effect of SR_{sub} X chromosome

location was significant but idiosyncratic and does not affect the main effects of drive (for individual

line effects, see supplementary figures 5-8). For all GLMs, a fully saturated model was produced and

was systematically reduced to the minimum adequate model through a step-wise simplification

process using Akaike information criterion (AIC). Difference in responses to all GLMs under different

treatments were assessed by analysis of variance (ANOVA) followed by a TukeyHSD comparison.

Results

Wing size

Wing size (a proxy for body size), was analysed using a GLM. The model considered the type of male and the age of the male and their interaction. The interaction was not significant and was dropped from the model. Wing size was affected by age (GLM: $F_{1,219} = 6.412$, $p = 0.012$). Whether a male was SR_{sub}, $^{Supp}SR_{sub}$ or was ST also had a significant effect on wing size (Supplementary figure 4: GLM: $F_{2,219} = 25.471$, $p < 0.001$). SR_{sub} males were bigger than both $^{Supp}SR_{sub}$ males (TukeyHSD, $P < 0.001$) and ST males (TukeyHSD, $p < 0.001$). However, there was no difference in wing length between ST and $^{Supp}SR_{sub}$ males (TukeyHSD, $p = 0.615$).

Testes size

Mean testes size relative to wing length was analysed using a GLM. The model considered the type of male and the age of the male. A significant age by type of male interaction was found (Figure 2a: GLM: $F_{2,215} = 5.210$, $p = 0.00617$). Here, ST males have significantly reduced testes size as they age from 3-days-old to 7-days-old (TukeyHSD, $p < 0.001$). This reduction in testes size was not seen in SR_{sub} and $^{Supp}SR_{sub}$ males as they aged (TukeyHSD, $p = 0.919$, TukeyHSD, $p = 0.630$, respectively).

Seminal vesicle size

Mean SV size relative to wing length was analysed using a GLM. The model considered the type of male and the age of the male and their interaction. The interaction was not significant, so was dropped from the model. Overall, SV's were bigger in 7-day-old males than 3-day-old males (Figure 2b: GLM: $F_{1,214} = 22.762$, $p < 0.001$). Whether a male carried SR_{sub}, $^{Supp}SR_{sub}$ or was ST also had a significant effect on mean SV size (Figure 3b: GLM: $F_{2,214} = 5.145$, $p = 0.00657$). Here, ST males have larger SV's than SR_{sub} males (TukeyHSD, $p = 0.00950$). However, there was no difference in SV size between $^{Supp}SR_{sub}$ and SR_{sub} males (TukeyHSD, $p = 0.066$) or between $^{Supp}SR_{sub}$ and ST males (TukeyHSD, $p = 0.797$).

Figure 2: The average size of a male's two testes (a) or seminal vesicles (b) of *Drosophila subobscura* males that were either SR_{sub}, ST or $^{Supp}SR_{sub}$ relative to body size. The boxplots display the upper and lower quartiles, the median and the range. Individual points represent each measurement obtained.

Damage to testes

Differences in damage to testes was analysed using a binomial GLM. The model considered

the type of male (ST; SR_{sub}; $^{Supp}SR_{sub}$) and the age of the male (3 or 7 days old) and their interaction.

Neither age nor the interaction age by male type interaction had an effect on the amount of damage

to a male's testes (GLM: $F_{1,\,209}$ = 0.0669, p = 0.791) so they were dropped from the model. Whether

the male was SR_{sub}, $^{Supp}SR_{sub}$ or ST had a significant effect on the amount of damage that occurred to

the testes (Figure 3: GLM: $F_{2,\,210}$ = 8.837, p < 0.001). $^{Supp}SR_{sub}$ males had a significantly higher

proportion of damaged testes than SR_{sub} and ST males (where by our measure, 49% of $^{Supp}SR_{sub}$ males

had visibly damaged testes in comparison to 23% of ST males and 26% of SR_{sub} males) (Figure 3:

TukeyHSD, p < 0.001, TukeyHSD, p < 0.001, respectively). There was no difference in the amount of

damage to testes between ST and SR_{sub} males (Figure 3: TukeyHSD, p = 0.905). An example of both

damaged and undamaged testes can be seen in Figure 1a.

Figure 3: The mean proportion of males that have visible damage to their testes that were either SR_{sub}
males, ST males or $^{Supp}SR_{sub}$ males, with 95% confidence intervals.

Accessory gland size

270   Accessory glands were measured in any images where they were clearly viable. Mean

accessory gland size relative to wing length was analysed using a GLM. The model considered the

type of male and the age of the male. Models analysing accessory gland size did not consider the

individual location the SR_{sub} X chromosome originated from due to the smaller sample size and

having little power to detect differences. A significant age by type of male interaction was found

(Figure 4: GLM: $F_{2, 145}$ = 4.950, p = 0.00833). Here, $^{Supp}SR_{sub}$ males' accessory glands increase in size as

they age (TukeyHSD, p = 0.00207). This change in accessory gland size was not seen in SR_{sub} males

(TukeyHSD, p = 0.999) or ST males (TukeyHSD, p = 0.999). Whether the male carried SR_{sub}, $^{Supp}SR_{sub}$ or

was ST also had an effect on accessory gland size (GLM: $F_{2, 145}$ = 13.570, p <0.001). Overall, $^{Supp}SR_{sub}$

males had larger accessory glands than both SR_{sub} and ST males (TukeyHSD, p = 0.045, TukeyHSD, p <

0.001, respectively). SR_{sub} males also had larger accessory glands than ST males (TukeyHSD, p =

0.0021). Finally, the age of the male had an effect on accessory gland size (GLM: $F_{1, 145}$ = 5282, p =

0.022). Overall, 7-day-old males had larger accessory glands than 3-day-old males (TukeyHSD, p =

0.023).

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285 **Figure 4:** The mean size of accessory glands of male *Drosophila subobscura* that were either SR_{sub}, ST
286 or $^{Supp}SR_{sub}$ relative to body size. A significant age by interaction term was found where $^{Supp}SR_{sub}$ males
287 accessory glands increase in size as they age. This effect was not seen in SR_{sub} or ST males.

Discussion

XCMD and their suppressors generate potent genetic conflict that may also be a force

influencing gonad development, morphology and evolution (31,32). In this study, we examined

whether the intragenomic conflicts created by SR_{sub} and its suppressor affects gonad morphology in

D. subobscura. We found significant differences in testes, seminal vesicle, and accessory gland size

depending on whether a male carried XCMD or not, and also if XCMD was suppressed. We also

found the first evidence of extreme whole-organ damage to testes associated with carrying a

suppressor of XCMD. While males with SR_{sub} from independent origins had differences in the

morphological traits observed, this variation was idiosyncratic and overall the main effect of SR_{sub}

and its suppressor were the same, suggesting these effects are robust across populations. In

conclusion, we found strong evidence that XCMD and its suppressor can have a major impact on

gonad development and morphology of its carriers.

Testes and Seminal Vesicle size

We found evidence that the testes of ST males, but not those of SR_{sub} and $^{Supp}SR_{sub}$ males,

reduce in size as they age. *Drosophila subobscura* are strictly monandrous (28,44), and males may

not benefit from high investment in continually producing large quantities of sperm (53–55). In

particular, if *D. subobscura* can partition their ejaculates, like in numerous species across a wide

array of taxa (56,57), they may only transfer to a female the sperm she will need, rather than their

whole ejaculate. This results in reduced pressure on the testes to continually produce sperm (and

the costs of spermatogenesis this entails). SVs are sperm storage organs in *Drosophila* males, where

mature sperm that develop in the testes move into the SV for storage until mating (58,59). Thus, in

unmated males, SV size is generally related to the amount of sperm stored (60). The combination of

ST males' testes reducing in size and having larger SVs suggest that these males initially produce

more sperm that fill their SVs, before subsequent investment in sperm production is reduced. In

contrast to the reduction in testes size observed in ST males, it appears that SR_{sub} and $^{Supp}SR_{sub}$ testes

remain the same size as males age. We propose three mutually non-exclusive reasons for this: (1)

compensation, (2) damage or (3) a delay in sperm production.

In stalk-eyed flies, males that carry XCMD have enlarged testes to compensate drive induced

sperm mortality, and XCMD males produce an equivalent number of sperm to normal males (31).

The compensation of male testes also appears to occur from an early development stage (32).

Compensation may also be the effect that we have observed in *D. subobscura*. Indeed, a previous

study has suggested that the loss of Y-bearing sperm in SR_{sub} males is compensated through cyst

doubling in the testes (61,62). Additionally, SR_{sub} males are capable of producing an equivalent

number of offspring to ST males in single matings, although this could be attributed to other factors,

such as increased mating duration or females not being sperm limited in this context (28). However,

the smaller SVs observed in SR_{sub} suggests that these males may actually be producing and storing

fewer sperm, which is not consistent with the compensation of sperm production.

Although bigger than SR_{sub} males, SVs of $^{Supp}SR_{sub}$ were still smaller than ST males. A key

feature of suppression is that it must prevent some of the killing of Y sperm (33). As $^{Supp}SR_{sub}$ males

have at least some of their Y sperm protected, this result suggests they produce more sperm than

SR_{sub}. However, the suppressor may not fully rescue all Y sperm or it may be associated with its own

damaging effects on sperm production. The smaller SVs of both types of XCMD carrying males in

comparison to ST males indicates fewer sperm are made and stored by these males, again

challenging the idea of compensation in this system. In addition, as there is no sperm competition in

D. subobscura (28,44), loss of sperm number may not need to be compensated.

The above discussion presumes testes size reflects functional capacity. Alternately, damage

to testes may be causing cellular damage, and this is preventing the reduction in testes size as males

age. Any damage that is occurring within the testes, either through the killing of Y-bearing sperm or

by carrying the suppressor, could result in inflammation or the accumulation of stress proteins at the

site of damage as males age. Necrotic cell death may additionally contribute to the testes continued

enlargement via a loss of plasma membrane integrity followed by cytoplasmic leakage (63,64). This

could lead to the cells swelling, producing inflamed tissues as a consequence. This is in contrast to

programmed apoptotic cell death that is characterised by cell shrinkage and membrane blebbing

(64,65). Thus, the lack of reduction in size may be more likely to be a combination of factors

associated with damage that is occurring in the testes.

Delays in sperm production could also affect the differences in testes sizes and SV sizes we

observe in our male types. If testes are largest simply when the process of spermatogenesis is at its

peak, then sperm production, and the reaching of a full sperm complement, may simply be delayed

in SR_{sub} and $^{Supp}SR_{sub}$ males. As a result, testes would remain larger for a longer period of time, while

they continue full sperm-production to an older age. Currently, little is known about the interaction

of XCMD and genes generally involved in gonad development (66,67). SR_{sub} males having the

smallest SVs, followed by $^{Supp}SR_{sub}$ males, and ST males having the largest SVs would also be

consistent with this development delay, with XCMD males that are suppressed potentially showing

an intermediate timing to reach full ejaculate complement. A closer examination of the fitness of

males of different ages, under repeated mating treatments, might reveal if developmental delays in

sperm production could explain the observed difference in testes morphology and whether these

delays influence male fitness.

Whole testes damage

Remarkably, we found evidence that, when the suppressor is paired with a driving X SR_{sub}

chromosome, it causes extreme physical damage to testes. Overall, this damage is less in XCMD

males that lack the suppressor and comparable to standard males. Once evolved, suppressors

against drive are usually expected to remove or minimise the negative impacts of drive (33,68,69).

However, we demonstrate for the first time that suppression, while mitigating some negative effects

of XCMD, can generate its own damage (and presumably, costs). In addition, given the scope of the

morphological differences we see, suppression genes may influence general testes development

that, when expressed and paired with SR_{sub}, results in their damage or developmental failures. To

what effect, if any, this damage then has on a male's fertility remains an open question.

XCMD is known and expected to impose fitness costs upon its carriers (18,45,70). Despite

this, less is known about how suppressors evolve to respond and if suppression is costly (33). Thus,

there is currently an imbalance in our knowledge, where we have a deeper understanding of the

costs of drive, but less so of the suppressors themselves and the balance between drive and

suppression. If costs of the suppressor are strong enough, suppressors may not spread. Hence, the

suppressor remains rare in populations where SR_{sub} is present (46). In spite of this, the suppressor

has not been lost from populations. This could be because the suppressor genes only appear costly

when paired with XCMD, and our data is consistent with this. Thus, partial suppression may allow

XCMD to persist for long periods of time (41,43). To fully understand the extent of this damage to

gonads, we need to; (1) determine if this damage is altering the internal structure of the male

gonads; (2) see the extent of cell death within XCMD carriers that are both suppressed and

unsuppressed; and (3) use an omics approach to investigate difference in the proteomes (71).

It is critical this damage is understood as it could affect the balance between drivers and

suppressor that may provide insight into the long-term stability of drive that we see in many natural

systems (43,72–74). Additionally, if suppressors spread despite deleterious effects, they have the

potential to fix negative mutations within a population, which is rarely considered in empirical

studies. There are many cases of cryptic (or muted) drive systems that are fixed within populations

(68,69,75). Here, drive systems are only revealed through outcrosses with other populations or

species that do not have the suppressor, suggesting that suppression has become fixed. These

factors are especially important to consider in relation to artificial drive systems (76). What would

happen to artificial drive if a suppressor unexpectedly evolved and fixed deleterious mutations

within the target population? The evidence we have found that major damage is associated with

suppression against XCMD suggests that not only can SGEs carry huge costs, but in addition

suppressors and their interactions with SGEs must be accounted for.

Accessory gland size

The extent of changes to male gonads as a result of XCMD seem not to just be limited to the

testes. We observed changes to the whole of the male gonads, including the accessory glands.

Accessory glands are organs that are responsible for the production of seminal fluid that contains

accessory gland proteins (77). In *Drosophila*, accessory gland proteins influence female behaviour

and a males sperm competitive ability (78,79). In some systems, accessory gland size is also

correlated with male mating rate and success (80,81). In this study, we found that $^{Supp}SR_{sub}$ accessory

glands are the largest, followed by SR_{sub} which were larger than ST males. Carrying SR_{sub} may be

associated with compensatory increases in investment in accessory gland proteins to ensure their

mating success rather than transferring an equivalent amount of sperm to ST males. Additionally,

the highest investment in accessory glands was from $^{Supp}SR_{sub}$. When SR_{sub} males are suppressed they

also endured the highest amount of damage to their testes, which may indicate males are ensuring

their mating success as they endure fitness costs in other ways. Despite this, it remains unclear as to

what the role of accessory glands is in a monogamous species where the importance of them

becomes largely unknown.

Conclusion

Here, we demonstrate that the presence of XCMD and its suppressor can cause significant

morphological changes to male gonads. This is the second system, that we are aware of, where

XCMD strongly influences male gonad development (31,32). Unlike the XCMD system in stalk-eyed

flies, where morphological changes were deemed a compensation mechanism against drive, our

evidence suggests that XCMD in *D. subobscura* inflicts extreme damage on male gonads that

becomes even more severe in the presence of its suppressor. This highlights that SGEs are a strong

evolutionary force that can broadly impact gonad development and gametogenesis. Additionally,

considering drive alone may only reveal half of the story. Suppressors can also have their own severe

effects on their carriers. Overall, the fitness effects of either an SGE or its suppressors may be severe

for individuals that carry them.

Acknowledgements

417   The authors would like to thank Jola Tanianis for her help with the isolation of the SR_{sub}

418   chromosomes, and Kieran Romain for measuring the accessory glands. This work was funded by the

419   National Environment Research Council ACCE DTP and the grant no. NE/S001050/1.

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